



Investigating pyridazine and phthalazine exchange in a series of iridium complexes in order to define their role in the catalytic transfer of magnetisation from *para*-hydrogen[†]

Kate M. Appleby,^{ab} Ryan E. Mewis,^{ab} Alexandra M. Olaru,^{ab} Gary G. R. Green,^{ab} Ian J. S. Fairlamb^b and Simon B. Duckett^{*ab}

The reaction of $[\text{Ir}(\text{IMes})(\text{COD})\text{Cl}]$, [$\text{IMes} = 1,3\text{-bis}(2,4,6\text{-trimethylphenyl})\text{imidazol-2-ylidene}$, COD = 1,5-cyclooctadiene] with pyridazine (pdz) and phthalazine (phth) results in the formation of $[\text{Ir}(\text{COD})(\text{IMes})(\text{pdz})]\text{Cl}$ and $[\text{Ir}(\text{COD})(\text{IMes})(\text{phth})]\text{Cl}$. These two complexes are shown by nuclear magnetic resonance (NMR) studies to undergo a haptotropic shift which interchanges pairs of protons within the bound ligands. When these complexes are exposed to hydrogen, they react to form $[\text{Ir}(\text{H})_2(\text{COD})(\text{IMes})(\text{pdz})]\text{Cl}$ and $[\text{Ir}(\text{H})_2(\text{COD})(\text{IMes})(\text{phth})]\text{Cl}$, respectively, which ultimately convert to $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$, as the COD is hydrogenated to form cyclooctane. These two dihydride complexes are shown, by NMR, to undergo both full N-heterocycle dissociation and a haptotropic shift, the rates of which are affected by both steric interactions and free ligand $\text{p}K_{\text{a}}$ values. The use of these complexes as catalysts in the transfer of polarisation from *para*-hydrogen to pyridazine and phthalazine via signal amplification by reversible exchange (SABRE) is explored. The possible future use of drugs which contain pyridazine and phthalazine motifs as *in vivo* or clinical magnetic resonance imaging probes is demonstrated; a range of NMR and phantom-based MRI measurements are reported.

Received 2nd March 2015
Accepted 27th April 2015

DOI: 10.1039/c5sc00756a
www.rsc.org/chemicalscience

1. Introduction

Pyridazine and phthalazine are nitrogen-containing heterocycles with derivatives that have proven to be biologically active as anti-asthmatic,¹ antifungal,² anti-HIV-1,³ anti-cancer^{4,5} and anti-inflammatory drugs^{6–8} in addition to acting as treatments for Alzheimer's disease.^{9,10} A wide range of pyridazine and phthalazine coordination complexes are known where ligation takes place through one or both of the nitrogen centres.^{11–17} When these ligands bind through a single nitrogen centre, the resulting complexes undergo both dissociative ligand loss and 1,2-metallotropic shifts through which the contiguous nitrogen donor atoms interchange positions.¹² Such shifts were first described in 1972 for a series of ruthenium porphine complexes that contained an axially-bound methylated pyridazine ligand.¹² Rates for ligand shift and dissociation were obtained by nuclear magnetic resonance spectroscopy *via* line-shape analysis and subsequently the activation parameters for these processes were determined. The authors suggested that the sizeable, and

positive, activation entropies that were observed are consistent with a dissociative pathway, although the precise mechanism for the 1,2-metallotropic shift was not discussed.¹²

More recent studies have focused on related octahedral systems where full ligand dissociation was more difficult and the 1,2-metallotropic shift proceeded readily. Examples of such complexes can be found for ruthenium,¹³ chromium,¹⁴ tungsten, platinum^{11,15,16} and rhenium¹⁷ metals. In the more recent reports, the 1,2-metallotropic shift has been referred to as a 'haptotropic shift' because of the potential to involve a π -bonding interaction between the two nitrogen centres during exchange. A theoretical study on an octahedral chromium carbonyl complex suggests that this shift can proceed within the solvent cage that surrounds a 16-electron intermediate.¹⁸ These processes are illustrated in Scheme 1. The tendency for ligand dissociation has been predicted to increase as the metal becomes less electron rich or the ligand *trans* to the exchanging site becomes more basic.¹⁹

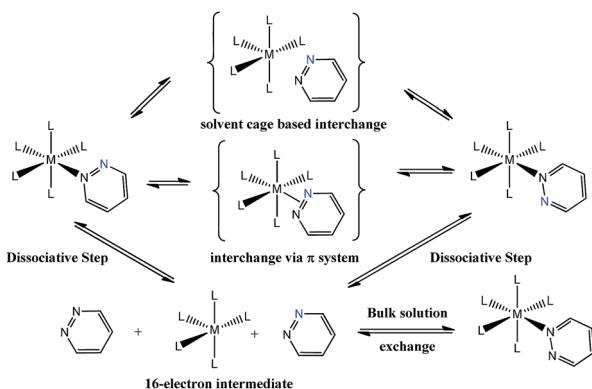
Ligand exchange processes are readily probed by Nuclear Magnetic Resonance (NMR) spectroscopy,²⁰ despite its low sensitivity.²¹ When slow, such exchange pathways are often followed by 1-D and 2-D nuclear Overhauser effect spectroscopy (NOESY).²⁰ When fast, line-shape analysis allows exchange to be probed.^{22,23} It has been shown that magnetisation transfer from *para*-hydrogen ($p\text{-H}_2$) can help to overcome the sensitivity

^aCentre for Hyperpolarization in Magnetic Resonance, University of York, York Science Park, York, YO10 5NY, UK. E-mail: simon.duckett@york.ac.uk

^bDepartment of Chemistry, University of York, Heslington, York, YO10 5DD, UK

[†] Electronic supplementary information (ESI) available: Synthesis, Characterisation, kinetic and hyperpolarisation data with the kinetic model is available here. See DOI: 10.1039/c5sc00756a





Scheme 1 Proposed ligand exchange pathways of pyridazine, the blue nitrogen label is a visualization aid.

problem associated with NMR.^{24,25} This is because, whilst *p*-H₂ itself has no net spin angular momentum and is NMR silent, when used as a reagent in a reaction, products can be formed that possess non-Boltzmann nuclear spin distributions.^{26–29} The resulting signals that are detected in such NMR experiments can be many thousands of times stronger than normal. *Para*-hydrogen induced polarisation (PHIP)³⁰ that is created in this way has been shown to enable the study of reaction intermediates that cannot otherwise be detected by NMR spectroscopy because they exist in low concentration.³¹ In this traditional approach, the reactants must accept H₂ and thus be both unsaturated and highly reactive.³¹

p-H₂ has also been used in a non-hydrogenative manner,³² where molecules are hyperpolarised through the establishment of *J*-coupling with *p*-H₂, whilst they are simultaneously bound to a metal centre in low field. This has also been achieved by radio frequency excitation.^{33–35} The non-hydrogenative process requires the substrate and *p*-H₂ to exchange freely with their ligated forms in order to build-up a concentration of hyperpolarised substrate in solution. This effect has been termed signal amplification by reversible exchange (SABRE).³⁶ Polarisation transfer now proceeds over the short time period that the sample is stored in a small pre-determined magnetic field.²⁵ This process has attracted interest because, unlike the traditional PHIP approach, it does not result in any chemical change to the detected species and reflects a novel catalytic process.^{37–40} A growing range of materials, including pyridine, nicotinamide, isoniazid, pyrazinamide and acetonitrile have been polarised in this manner.^{41–43}

Here, we examine the substrates pyridazine (pdz) and phthalazine (phth) using the SABRE approach, whilst taking the opportunity to simultaneously explore their coordination chemistry. [Ir(IMes)(COD)Cl] (**1**) (where IMes = 1,3-bis(mesilyl)imidazol-2-ylidene and COD = 1,5-cyclooctadiene) is used as the synthetic precursor in this study.⁴⁴ We show that upon reaction of **1** with pyridazine or phthalazine, [Ir(COD)(IMes)-(pdz)]Cl (**2a**) and [Ir(COD)(IMes)(phth)]Cl (**2b**) form, respectively. We find that these complexes add H₂ reversibly to yield highly reactive [Ir(H)₂(COD)(IMes)(pdz)]Cl (**3a**) and [Ir(H)₂(COD)(IMes)(phth)]Cl (**3b**). They subsequently undergo COD

hydrogenation to form [Ir(H)₂(IMes)(pdz)]Cl (**4a**) and [Ir(H)₂(IMes)(phth)]Cl (**4b**). The mechanism of the ligand dissociation process exhibited by these complexes is probed by NMR spectroscopy and the roles of **4a** and **4b** as polarisation transfer catalysts in the SABRE of pyridazine and phthalazine are explored. Both NMR and phantom-based MRI measurements are reported to illustrate the wider benefits of hyperpolarisation.

2. Experimental procedures

All the experimental procedures associated with this work were carried out under nitrogen using standard Schlenk techniques or in an MBraun Unilab glovebox. The solvents used in the synthetic chemistry were dried using an Innovative Technology anhydrous solvent system, or distilled from an appropriate drying agent under nitrogen. Methanol-*d*₄, pyridazine and phthalazine were obtained from Sigma-Aldrich and used as supplied. [Ir(IMes)(COD)Cl] (**1**) was prepared according to literature methods.⁴⁵

2.1. Instrumentation and procedures

All NMR measurements were recorded on Bruker Avance III series 400 MHz or 500 MHz systems. NMR samples were prepared in 5 mm NMR tubes fitted with Young's valves. Samples were degassed prior to *p*-H₂ (3 bar) addition. Typical procedures for reactions with pyridazine and phthalazine are described. NMR characterisation data was collected using a range of 1 and 2 D methods that include nOe, COSY and HMQC procedures.^{46–50} The slow dynamic processes exhibited by the complexes studied here were monitored by EXSY methods.⁵¹

[Ir(H)₂(IMes)(pdz)]Cl (4a). In a typical experiment, **1** (2 mg, 3.12 μ mol) and five equivalents of pyridazine (1.13 μ L, 15.6 μ mol) were dissolved in methanol-*d*₄ (0.5 mL) in a 5 mm NMR tube and the solution was degassed. H₂, at a pressure of 3 bar, was added to the sample prior to it being heated to 348 K for 15 minutes to fully activate it thereby forming **4a** in solution.

[Ir(H)₂(IMes)(phth)]Cl (4b). **1** (2 mg, 3.12 μ mol) and five equivalents of phthalazine (2.03 mg, 15.6 μ mol) were dissolved in methanol-*d*₄ (0.5 mL) in a 5 mm NMR tube and the solution was degassed. H₂ (3 bar) was added, and the sample was heated to 348 K for 15 minutes to form **4b**.

2.2. SABRE analysis

NMR samples were prepared as above. Arrays of NMR measurements were collected using a range of substrate concentrations, as revealed in the ESI.† ¹H NMR spectra were recorded with either $\pi/4$ or $\pi/2$ excitation pulses in order to differentiate PHIP and SABRE effects. Each single-scan NMR spectrum was collected immediately after shaking the sample in a magnetic field of 65 G (unless otherwise specified), whilst under 3 bar of *p*-H₂. Enhancement factors were calculated by comparing the peak areas in the resulting NMR spectra with those of analogous traces collected under normal H₂ and Boltzmann equilibrium conditions. In order to do this, the peak areas of individual resonances in the corresponding normal



NMR spectra were calibrated to 1 and enhancements calculated by comparing these values to those in the polarised spectra. Total signal enhancements were calculated by multiplying this enhancement value by the number of protons in the group and summing across the molecule.

2.3. Polarisation transfer field measurements

Polarisation transfer field measurements were made using a dedicated hyperpolarisation apparatus.⁴⁴ Samples were first prepared in 3 mL of methanol-*d*₄ in an ampoule containing 10 mg of **1** and 5 equivalents of pyridazine or phthalazine. The solution was degassed, and the ampoule was filled with 3 bar pressure of H₂ and heated to 348 K overnight to form **4a** or **4b**. At this point, the resulting solutions were introduced into the flow system and the SABRE effect monitored for polarisation transfer fields that ranged from 0.5 to 140.5 G, in steps of 10 G. This involved the collection of a series of single-scan spectra using either $\pi/2$ pulses or shaped pulses^{50,52} for the multiple quantum filtered experiments. The peak integrals in the resulting NMR spectra were compared to those of a thermally polarised spectrum to determine signal enhancement factors.

2.4. EXSY measurements

A series of exchange spectroscopy (EXSY) measurements were made to probe the dynamic behaviour of these systems.⁵³ This process involved the selective excitation of a single resonance and the subsequent measurement of a ¹H NMR spectrum at time, *t*, after the initial pulse. The resulting measurements consisted of a series of data arrays such that *t* was varied over 10 values, typically from 0.1 to 1.0 s. The precise values were varied with temperature to suit the speed of the process. Data was collected for a range of temperatures and sample concentrations as tabulated in the ESI.[†]

2.5. Kinetic analysis

Integrals for the interchanging peaks in the associated ¹H EXSY spectra were obtained and converted into a percentage of the total detected signal. These data were then analysed as a function of the mixing time according to a differential kinetic model.⁵⁴ Rates of exchange were determined by employing the solver add-on in Microsoft EXCEL to minimize the sum of the residuals in the associated least mean squares analysis. The rate constants obtained in this way were doubled when calculating barrier heights to account for step reversibility.⁵⁵ Errors associated with the calculated rate constant were determined by the Jackknife procedure of Harris.⁵⁶

2.6. Image acquisition and processing

For the imaging experiments, samples containing **4a** and **4b** were prepared as described earlier. All data was then acquired on a 400 MHz Bruker spectrometer equipped with a micro imaging gradient system (1 T m⁻¹) and a standard 30 mm ¹H-¹³C double resonance birdcage. For the SABRE experiments the sample tubes were filled with 3 bar of *p*-H₂ and shaken in the stray field of the MRI magnet (*ca.* 70 G). As a reference,

images of the samples with Boltzmann-equilibrated magnetisations were recorded in conjunction with a recycle delay of 100 s and 512 averages.

A standard rapid acquisition with refocused echoes (RARE)⁵⁷ sequence composed of a train of 64 π pulses separated by an echo time (TE) of 8.15 ms (effective echo time, TEeff = 114 ms) was used to collect the images. According to Hennig *et al.*, by using a relatively small number of echoes, combined with a short echo time, the effect of transverse relaxation is minimised and the image contrast is dictated by spin density.⁵⁷ All images used a field of view (FOV) of 30 × 30 mm, a slice thickness of 5 mm and a 64 × 64 matrix size. This led to a raw resolution of 0.47 × 0.47 × 5 mm³. Data were processed by FT and zero filled to 128 × 128 (resulting digital resolution: 0.23 × 0.23 × 5 mm³) using a sine bell noise filter. Signal-to-noise (SNR) values were calculated using an algorithm that assumed a Rice-type noise distribution.

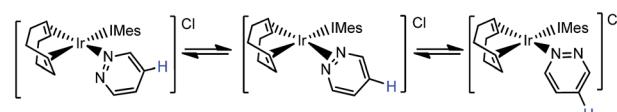
3. Experimental results

3.1. Reaction of [Ir(IMes)(COD)Cl] (1) with pyridazine and phthalazine

It has already been reported that when **1** reacts with pyridine in methanol-*d*₄ it forms [Ir(COD)(IMes)(py)Cl].⁴⁴ When pyridine is replaced with either pyridazine or phthalazine the corresponding solutions immediately change colour from yellow to orange due to the formation of the analogous complexes [Ir(COD)(IMes)(pdz)]Cl (**2a**) and [Ir(COD)(IMes)(phth)]Cl (**2b**). Both of these complexes have been characterized by NMR spectroscopy and mass spectrometry. Attempts to follow this process by stop-flow UV-Vis spectroscopy, with a dead-time of 0.6 ms, failed because the reaction was too rapid.

3.2. Ligand exchange exhibited by [Ir(COD)(IMes)(pdz)]Cl (**2a**) and [Ir(COD)(IMes)(phth)]Cl (**2b**)

When these products were analysed by ¹H NMR spectroscopy, signals for the bound N-heterocycles could not be resolved at room temperature. To overcome this, a series of NMR spectra were collected at 253 K. The ESI[†] presents the corresponding ¹H, ¹³C and ¹⁵N resonance assignments for these species. A series of 2-D nOe spectra were also collected at 253 K. These confirmed that the NCHCH and NCH protons of the pyridazine ligand in **2a** and the phthalazine ligand in **2b** switch their nitrogen binding sites on a timescale of 0.5 seconds (see Scheme 2). The broadening of the NMR signals for these ligands, seen at room temperature, is due to this rapid exchange. NMR signals for the free ligand were also present in these spectra, although no exchange into them was observed at



Scheme 2 Ligand exchange pathway exhibited by **2a**.



this temperature, or upon warming to 298 K. This is therefore an intramolecular process.

The intramolecular nature of this process means that the 1,2-metallotropic shift is haptotropic. It occurs *via* an intermediate with an $\eta^2\text{-N}=\text{N}$ coordination mode according to Eaton *et al.*,¹² rather than dissociation which would lead to 14-electron $[\text{Ir}(\text{COD})(\text{IMes})]^+$ and the observation of exchange with the free ligand pool.

The rate of the haptotropic shift was determined over the temperature range 240 to 260 K by EXSY spectroscopy. Exchange in **2a**, between the proton signal at δ 7.69 (corresponding to IrNCHCH) and that at δ 7.51 (corresponding to IrNNCHCH), was probed. The analogous process in **2b** was followed between 235 and 255 K by selecting the proton signal at δ 9.55 (corresponding to IrNCH) and observing its exchange into the signal at δ 8.37 (corresponding to IrNNCH). The associated exchange rates were quantified by data fitting (see ESI†). Fig. 1 shows a typical exchange profile, while Table 1 lists the corresponding rate data. The activation parameters for these processes were calculated by Eyring analysis and are given in Table 2. They are similar to those reported.^{11,14–17}

The enthalpy of activation for the haptotropic shift of **2a** is larger than that of **2b** by 5 kJ mol^{−1}. This difference suggests that the Ir-pdz bond is stronger than the Ir-phth bond and yet the reported pK_a values of pyridazine and phthalazine are 2.33 and 3.50, respectively.⁵⁸ We suggest that steric interactions result in the smaller activation entropy gain for binding site interchange of the larger and more basic phthalazine ligand.

3.3. Addition of H₂ to $[\text{Ir}(\text{COD})(\text{IMes})(\text{pdz})]\text{Cl}$ (**2a**) and $[\text{Ir}(\text{COD})(\text{IMes})(\text{phth})]\text{Cl}$ (**2b**)

Two separate methanol-*d*₄ solutions of **2a** and **2b** were exposed to a 3 bar pressure of hydrogen at 240 K. In both cases the solutions changed colour from orange to colourless due to the formation of $[\text{Ir}(\text{H})_2(\text{COD})(\text{IMes})(\text{pdz})]\text{Cl}$ (**3a**) and $[\text{Ir}(\text{H})_2(\text{COD})(\text{IMes})(\text{phth})]\text{Cl}$ (**3b**), respectively. These complexes are easily identified by ¹H NMR spectroscopy as they yield two coupled hydride resonances, which in the case of **3a** lie at δ −13.84 and −17.69, and for **3b** lie at δ −13.87 and −17.55. As the hydride ligands of both these products are chemically inequivalent, they must adopt one of two possible product

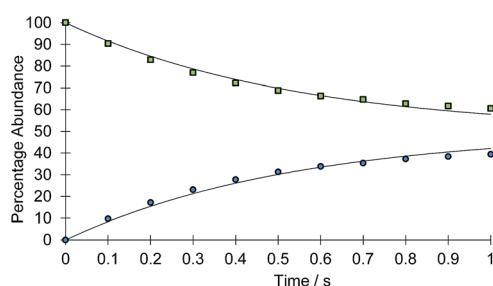


Fig. 1 Signal intensity profile change associated with the interchange of the IrNCHCH (green squares) and IrNNCHCH (blue circles) resonances of **2a**, according to Scheme 2 above, as a function of reaction time (*t*). Simulated data is presented as the solid line.

Table 1 Ligand exchange rate constants of **2a** and **2b** in methanol-*d*₄ solution

Temperature/K	Rate of haptotropic shift/s ^{−1}	
	2a	2b
235	—	0.923 ± 0.001
240	0.729 ± 0.001	1.873 ± 0.003
245	1.501 ± 0.003	3.955 ± 0.003
250	3.155 ± 0.002	7.318 ± 0.004
255	5.80 ± 0.01	13.426 ± 0.004
260	12.74 ± 0.02	—

geometries, assuming that hydrogen addition follows the expected concerted route, as shown in Scheme 3.⁵⁹ In one of these product geometries, the hydride ligands lie *trans* to the N-heterocycle and COD (**3a**) while in the second they lie *trans* to IMes and COD (**3a**).⁶⁰

One route to differentiate these two orientations is to recognise that a *trans* H–Ir–¹⁵N coupling is significantly larger than a *cis* coupling. When a ¹⁵N-optimized HMQC NMR spectrum was recorded to probe this, strong cross-peaks were found between the hydride signals at δ −17.55 (of **3b**) and −17.69 (of **3a**) and the nitrogen signals of the bound N-heterocycle at δ 294.17 and 266.53 respectively. This confirms their mutual *trans* orientation and that **3a** and **3b** form by H_2 addition over the COD–Ir–N axis of **2a** and **2b** respectively. This is consistent with the nature of the ligands determining the direction of addition, which in this case favours addition over the axis containing the weak nitrogen donor rather than the strong IMes ligand.⁶¹

Furthermore, the corresponding 2D-¹H COSY NMR spectrum contains cross-peaks between the hydride ligand signals at δ −13.84 (of **3a**) and −13.87 (of **3b**) and the inequivalent proton signals that arise from the two CH environments in the bound COD that are *trans* to hydride. In the case of **3a** these COD signals appear at δ 5.09 and 4.45. Both species **3a** and **3b** have been fully characterised by NMR spectroscopy.

We note that when solutions of **3a** and **3b** are degassed, reductive elimination of H_2 occurs and **2a** and **2b** reform. H_2 addition is therefore a reversible process. These complexes are examples of a relatively rare class of alkene dihydride species that are typically active in alkene hydrogenation. A related iridium pyridylpyrrolide COD complex has been shown by Searles *et al.* to reversibly bind hydrogen in a similar way.⁶²

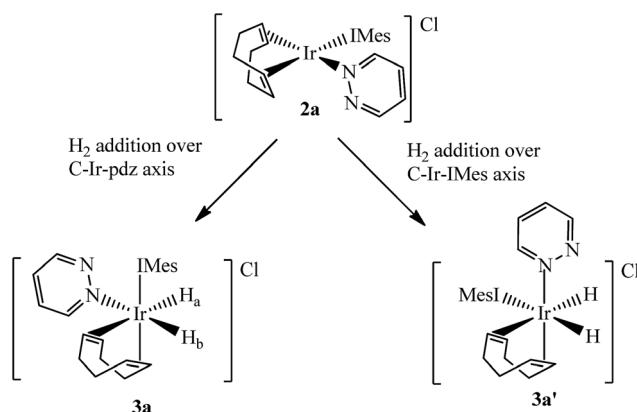
3.4. Reaction of $[\text{Ir}(\text{COD})(\text{H})_2(\text{IMes})(\text{pdz})]\text{Cl}$ (**3a**) and $[\text{Ir}(\text{COD})(\text{H})_2(\text{IMes})(\text{phth})]\text{Cl}$ (**3b**) with *p*-H₂

One route to probe the concerted nature of this process is to employ *p*-H₂. This is because PHIP requires the pair-wise

Table 2 Activation parameters for the haptotropic shift exhibited by **2a** and **2b** in methanol-*d*₄ solution

Complex	$\Delta H^\ddagger/\text{kJ mol}^{-1}$	$\Delta S^\ddagger/\text{J mol}^{-1} \text{K}^{-1}$	$\Delta G^\ddagger/\text{kJ mol}^{-1} (300 \text{ K})$
2a	71.3 ± 0.3	57 ± 1	54.2 ± 0.4
2b	66.0 ± 0.1	43.0 ± 0.6	53.1 ± 0.2



Scheme 3 Products of H_2 addition to **2a**.

transfer of H_2 . When the H_2 atmosphere of these samples is replaced with *p*- H_2 , and the NMR tube shaken immediately prior to ^1H NMR signal detection, dramatic hydride signal enhancements result. These signal enhancements are illustrated in Fig. 2 for **3a**. The experimentally observed signal enhancement was 90-fold for the sharper signal at δ -13.84 due to H_a and 66-fold for the broader signal at δ -17.69 due to H_b . For **3b**, smaller 52-fold and 35-fold signal enhancements are observed for the corresponding resonances.

In order to compare the reactivity of these complexes we prepared a sample that contained approximately equal amounts of pyridazine and phthalazine (6.73 equivalents of phth and 6.16 equivalents of pdz relative to **1** as measured by NMR). Upon NMR examination, the binding of phthalazine proved to dominate over that of pyridazine, with the ratio of **2b** : **2a** being $3.7 : 1$. When corrected for these slightly unequal starting ligand concentrations, the equilibrium ratio is $3.40 : 1$ and $\Delta\Delta G_{2a-2b}^0$ at 240 K is approximately -2.44 kJ mol^{-1} with **2b** being more stable than **2a**.

The normalized PHIP enhancements that are exhibited by the hydride signals of species **3a** and **3b** upon *p*- H_2 addition at

240 K are 56.8-fold and 38.2-fold, respectively. As the *p*- H_2 concentration in solution is constant and its purity identical at the point of measurement, these numbers can be compared. If we assume that both complexes retain the same level of *p*- H_2 derived magnetisation at the point of their formation then their relative signal intensities will be proportional to the product of their rates of reaction and concentration. The greater abundance of **2b** ($\times 3.7$), in combination with the hydride signal enhancement levels, suggests that the rate of H_2 addition to the more stable **2b** is a factor of *ca.* 4.7 slower than that to **2a**.

The PHIP effect that is seen in these hydride signals vanishes rapidly upon the introduction of the sample into the NMR probe at 240 K . This is because the hydrides do not undergo H_2 elimination on the NMR timescale at this temperature. However, if the sample is removed from the magnet and shaken to dissolve fresh *p*- H_2 , the PHIP effect is re-established in the associated hydride ligand signals. This is due to the transiently higher temperatures that the sample experiences whilst shaken. Under these conditions, however, **3a** and **3b** are present and it is their exchange rates that control the level of PHIP. In subsequent NMR spectra the levels of observed signal enhancement are almost equal, at 27.6-fold and 20.1-fold. However, in the corresponding thermally equilibrated NMR spectra, at 240 K , the phthalazine dihydride complex **3b** still dominates, now by a factor of 3.2. In fact, these data suggest that ΔG_{3a-3b}^0 is -2.33 kJ mol^{-1} at 240 K . Based on these data we can conclude that less stable **3a** undergoes more rapid H_2 loss than **3b**.

Limited evidence for the formation of the isomers **3a'** and **3b'** of Scheme 3 was observed. This is in agreement with the IMes-Ir-COD axis being the most electron rich and the most sterically hindered, and therefore the least able to promote H_2 addition across it.^{59,63} Hydride signals for these species were seen at δ -9.12 and -13.48 for **3a'** and at δ -9.11 and -13.48 for **3b'**, with very low intensity. The values of the hydride chemical shifts in this pair of complexes reflect their *trans*-orientations to soft carbene and COD ligands.⁶⁴ The signals did not exhibit PHIP, despite being coupled. They are also not visible in the low temperature ^1H NMR spectra but can be observed at 298 K . Upon cooling from this temperature, in an effort to trap the species, the corresponding hydride signals vanish and only those for **3a** and **3b** remain.

We conclude from these data that stability increases in the order **2a** $<$ **2b** $<$ **3a** $<$ **3b** and that the addition of H_2 proceeds more rapidly to **3a**.

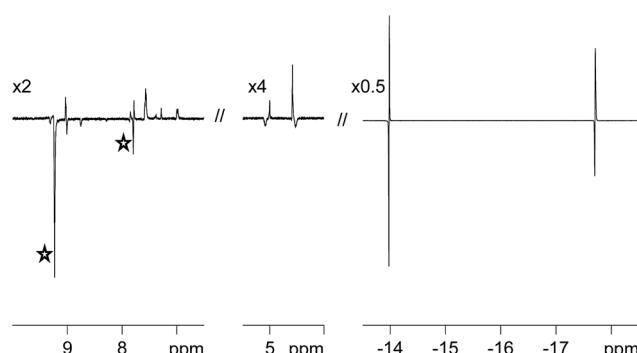


Fig. 2 Selected regions of a *p*- H_2 -enhanced ^1H NMR spectrum of **3a**, showing free (stars) and bound pyridazine (left) and $\eta^2\text{-CH}$ COD (middle) signal enhancements resulting from low-field polarisation transfer in addition to the corresponding PHIP enhanced hydride signals for H_a and H_b (right). The vertical expansion scales, relative to the original spectrum, are indicated.

3.5. Observation of the SABRE effect in the NMR signals of $[\text{Ir}(\text{COD})(\text{H})_2(\text{IMes})(\text{pdz})]\text{Cl}$ (**3a**) and $[\text{Ir}(\text{COD})(\text{H})_2(\text{IMes})(\text{phth})]\text{Cl}$ (**3b**)

The SABRE effect is evident in the first scan of these PHIP enhanced ^1H NMR spectra. This is reflected in the observation of enhanced pyridazine resonances for **3a** at δ 9.06 , 9.03 , 7.91 and 7.62 , as shown in Fig. 2. Additional enhanced signals corresponding to free pyridazine at δ 9.27 and 7.85 (the NCH and NCHCH environments respectively) are also observed. The ratio of the NCH signal for the free and bound ligand is $4.29 : 1$.

These data confirm that **3a** undergoes pyridazine loss on a shorter timescale than NMR relaxation.

One surprising feature of these chemical systems is the polarisation of bound CH proton environments of the η^4 -COD ligand, at δ 5.09 and 4.45 in the case of **3a**, although no free COD, COE or COA signals are evident. We note that there is no evidence in the corresponding EXSY spectra to suggest that the bound COD reversibly accepts a hydride ligand.⁶⁵ SABRE magnetisation transfer proceeds readily from the hydride ligands of **3a** and **3b** into the ligands that are *trans* to them.

3.6. Formation of the efficient magnetisation transfer catalysts $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (**4a**) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (**4b**)

Both **3a** and **3b** were stable at temperatures up to 250 K, with their hyperpolarisation effects disappearing in solution after several seconds. Above this temperature, however, the COD ligand hydrogenates and forms COA; there is no NMR evidence for COE.⁶⁵ The vacant sites that are created by the hydrogenation of COD then become occupied by two further pyridazine or phthalazine ligands. This leads to the formation of $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (**4a**) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (**4b**) respectively. These reactions take approximately one hour to go to completion at 298 K. Both **4a** and **4b** have been characterised by NMR spectroscopy and mass spectrometry. The reaction pathway for the conversion of **1** into **4a** is illustrated in Scheme 4. A weak unassigned hydride signal is seen in these ^1H NMR spectra at δ –22.87. This may correspond to an intermediate with a similar structure to that of $[\text{IrH}(\text{1-}\kappa\text{-C}_8\text{H}_{13})(\text{PMe}_3)(\text{NCCH}_3)_2]\text{BF}_4$, which provides a diagnostic hydride resonance at δ –20.35.⁶⁵

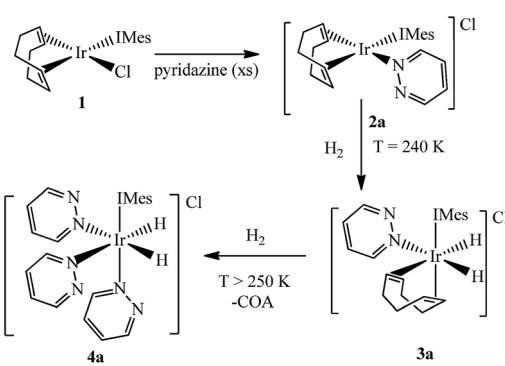
The organic region of a ^1H NMR spectrum of **4a** is shown in Fig. 3. The corresponding hydride region, which is not displayed, contains a single peak, at δ –21.47, due to the two chemically equivalent, but magnetically distinct, hydride ligands of this complex. The organic region of this NMR spectrum is complicated as the symmetry of pyridazine is broken upon binding to the metal centre. Seven new aromatic ^1H resonances are visible, four of which appear as simple doublets, at δ 9.62, 9.31, 8.75 and 8.29 with integral ratios of 1 : 2 : 2 : 1 respectively, relative to the 2-proton hydride ligand signal. Their doublet nature means that they arise from pyridazine NCH environments, and their relative areas confirm the presence of a

single axial ligand (δ 9.62 and 8.29) and two equivalent equatorial ligands (δ 9.31 and 8.75). The remaining three multiplets, at δ 7.62, 7.48 and 7.40, have integral ratios of 2 : 1 : 3 respectively and arise from the remaining NCHCH environments. The arrangement of these groups around the metal centre was confirmed by nOe measurements at 300 K with mixing times of ≥ 0.6 s. These data revealed that both the equatorial IrNCH and IrNNCH environments interact with methyl-IMes protons that resonate at δ 2.22 and 2.04. These connections confirm that the equatorial pyridazine ligands lie in close proximity to the IMes ligand. Complex **4a** was further characterised by extensive COSY, ^{13}C -optimised HMQC and ^{15}N -optimised HMQC measurements.

The phthalazine analogue of **4a**, $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (**4b**) was characterised in a similar way. Part of its ^1H NMR spectrum is shown in Fig. 3b. A single hydride peak is also observed for this product, at δ –21.0. Peaks at δ 9.63, 8.26 and 8.15 correspond to the NCH, NCHCCH and NCHCCCH environments of free phthalazine respectively. Four singlets are observed at δ 10.28, 10.16, 9.27 and 8.60 with integral ratios of 1 : 2 : 2 : 1. These resonances correspond to the bound NCH environments of **4b** and can be broken down into axial and equatorial contributions. NMR data for **4a** and **4b** is given in the ESI.†

3.7. Using ^{15}N chemical shifts to support the characterisation of $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (**4a**) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (**4b**)

Modern NMR methods, specifically inverse sequences, allow insensitive nuclei, such as ^{15}N , to be detected when they are present at natural abundance.⁶⁶ ^{15}N NMR data, collected by inverse HMQC methods for **4a** and **4b** are detailed in Table 3. In the case of **4a**, pyridazine binding to iridium moves the ^{15}N



Scheme 4 Products of H_2 and pyridazine addition to **1**.

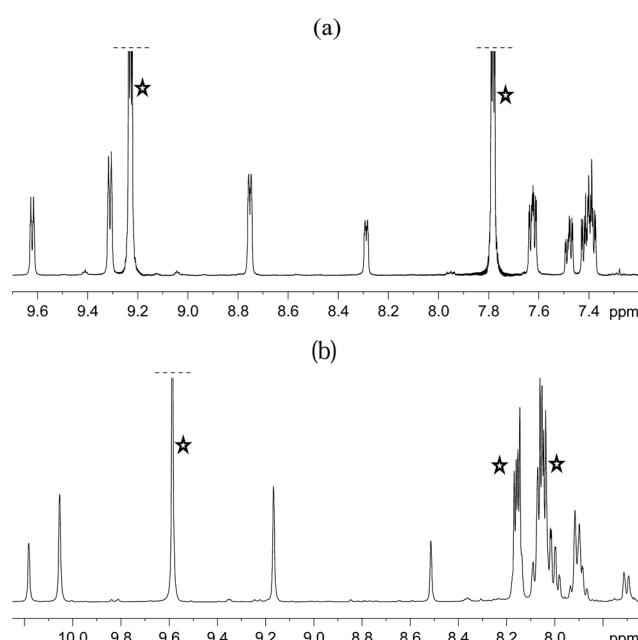


Fig. 3 ^1H NMR spectra of **4a** (a) and **4b** (b) showing the organic region with key equatorial and axial IrNCH and IrNNCH resonances. Free substrate resonances are also shown (stars).



chemical shift of the free ligand from δ 383.2, to δ 326.2, when bound *trans* to hydride, and to δ 305.8 when bound *trans* to carbene. The value of the change in chemical shift on binding $\Delta\delta$ (free-bound) has been suggested to relate to the extent of interaction with the metal.⁶⁷ As revealed in Table 3, the largest $\Delta\delta$ value is observed for the axial N-heterocycle arrangement with the in-plane change being 30% smaller. This suggests that there is a weaker in-plane ligand interaction as a consequence of the greater *trans*-influence of hydride relative to IMes.⁶⁷ The corresponding $\Delta\delta$ values for the remote nitrogen centres in these ligands are much smaller and positive in sign thereby illustrating that electron density surrounding them is significantly less perturbed by the introduction of a metal-ligand interaction.

3.8. Hydride and N-heterocycle ligand exchange dynamics of $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (4a) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (4b)

The ligand exchange pathways of **4a** and **4b** have been explored by NMR spectroscopy. The samples examined here employ d_4 -methanol as the solvent. A series of 1D EXSY measurements were undertaken, at 10 different temperatures, and both ligand exchange and H_2 loss pathways were revealed. These processes were too slow to be detected at temperatures below 280 K and the upper limit of our measurements was 325 K, due to the boiling point of methanol. At this temperature, the exchange rates were too slow to observe any line-shape changes.

In the first of our EXSY experiments, peaks corresponding to the NCH environment of the axial ligands in **4a** and **4b** were probed. No magnetisation transfer was observed from these sites, at 320 K, and we conclude that the axial ligands in both **4a** and **4b** remain bound to the metal on this timescale and that their involvement in exchange can be neglected. This observation matches the higher strength of interaction predicted through the larger $^{15}\text{N}-\Delta\delta$ value detailed earlier. In contrast, when the equatorial ligands of **4a** and **4b** were probed, magnetisation transfer indicative of ligand exchange and the 'shifting' of the equatorial ligand's nitrogen binding centre was observed by a 1,2-metallotropic shift. A typical EXSY trace is shown in Fig. 4. Both of these processes are common in such systems.^{11–17} Shifting coordination from one nitrogen centre to the other is likely to proceed *via* a trigonal bipyramidal transition state, rather than a Berry-pseudorotation, in accordance with the molecular orbital studies by Alvarez *et al.*¹⁹ When the hydride ligand signals of **4a** and **4b** were examined, in a similar way, exchange into the signal for H_2 was observed.

The rate constants for these processes were calculated by analysing an array of EXSY spectra that were recorded with a series of magnetisation transfer times. ESI Scheme 2 \dagger illustrates

Table 3 ^{15}N NMR chemical shift (δ) and $\Delta\delta$ data for complexes **4a** and **4b** (where $\Delta\delta = \delta_{\text{free}} - \delta_{\text{bound}}$)

	4a	$\Delta\delta$	4b	$\Delta\delta$
^{15}N equatorial $\text{Ir}-\text{N}=\text{N}-$	326.2	-57.0	299.7	-49.9
^{15}N equatorial $\text{Ir}-\text{N}=\text{N}-$	390.0	6.8	360.9	11.3
^{15}N axial $\text{Ir}-\text{N}=\text{N}-$	305.8	-77.4	278.8	-70.8
^{15}N axial $\text{Ir}-\text{N}=\text{N}-$	392.2	9.0	362.8	13.2

pictorially the exchange processes that occur with pyridazine, and the extracted rate constants as a function of temperature are also available. Eyring analysis of these data provided the activation parameters that are shown in Table 4. The barrier to Ir-N bond rupture (dissociation) is twice the sum of the full ligand loss rate if upon reaching the transition state there is an equal probability of a null-reaction. The haptotropic shift was considered as a separate intramolecular pathway in this analysis.

3.9. Mechanism of pyridazine and phthalazine ligand dissociation and binding site shift in $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (4a) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (4b)

Exchange behaviour was probed as a function of substrate concentration at 285 K (see ESI Tables 6–10 \dagger). A plot of ligand exchange rate *versus* ligand concentration, for a constant concentration of **1**, is presented in Fig. 5b for **4a** and Fig. 5c for **4b**. It can be seen from these data that the experimental rate constants for pyridazine and phthalazine ligand dissociation are unaffected by an increase in the free ligand concentration (Fig. 5b and c). This zero-order rate behaviour is consistent with a dissociative process and a haptotropic shift.¹⁸

The ΔH^\ddagger values in Table 4 reflect the associated Ir-N bond energies and the barriers to ligand exchange. For each pair of complexes, the enthalpy change on reaching the transition states for the haptotropic shift is lower than that of dissociation. Bulkier phthalazine has a weaker Ir-N bond and hence smaller ΔH^\ddagger term. Consequently, the Ir-N bond is longer and the gain in entropic freedom on reaching the transition state for both phthalazine dissociation and shifting falls. The two effects act to compensate each other such that their $\Delta G^\ddagger(300\text{ K})$ values are similar. This is particularly noticeable in the haptotropic shift, where the ΔH^\ddagger and ΔS^\ddagger terms of phthalazine are lower by 16.2 kJ mol $^{-1}$ and 55 J K $^{-1}$ mol $^{-1}$ respectively.

As mentioned previously, the literature values for the $\text{p}K_a$ of pyridazine and phthalazine are 2.33 and 3.50 respectively. Our observed values of $^{15}\text{N}-\Delta\delta$ upon ligand binding were -57.0 and -49.9 respectively. Less basic pyridazine exhibits a stronger bonding interaction than phthalazine; ground state steric effects must therefore control the Ir-N bond strength.

3.10. Mechanism of H_2 loss from $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (4a) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (4b)

The rate of H_2 loss from **4a** depends on the pressure of H_2 for a constant substrate excess, as shown in Fig. 5a. However, when

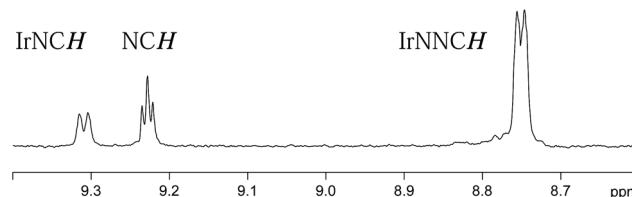


Fig. 4 1D EXSY ^1H NMR spectrum, illustrating the extent of magnetisation transfer from the δ 8.75 equatorial IrNNCH resonance of **4a** into the δ 9.23 free pyridazine signal and the δ 9.31 equatorial IrNCH resonance of **4a** after a magnetisation transfer time of 1 second.



Table 4 Activation parameters for the indicated ligand exchange processes in **4a** and **4b** in methanol-*d*₄

Process	$\Delta H^\ddagger/\text{kJ mol}^{-1}$		$\Delta S^\ddagger/\text{J K}^{-1} \text{mol}^{-1}$		$\Delta G_{(300 \text{ K})}^\ddagger/\text{kJ mol}^{-1}$	
	4a	4b	4a	4b	4a	4b
complex						
Substrate dissociation	97.9 ± 0.3	92.0 ± 0.3	85 ± 1	62 ± 1	72.5 ± 0.3	73.3 ± 0.3
Haptotropic shift	94.3 ± 0.4	78.13 ± 0.06	72 ± 1	17.4 ± 0.2	72.6 ± 0.3	72.93 ± 0.06
H ₂ loss	95.6 ± 0.2	83.6 ± 0.1	82.4 ± 0.8	39.8 ± 0.4	70.9 ± 0.3	71.6 ± 0.2

the H₂ loss pathway was monitored at constant H₂ pressure as a function of substrate concentration, inhibition was observed (Fig. 5b and c). The pseudo-rate constant for H₂ loss is inversely proportional to the substrate concentration (Fig. 5d). This behaviour matches with that predicted by DFT for the related pyridine system in which the dihydrogen-dihydride intermediate [Ir(H)₂(H₂)(IMes)(py)]Cl is involved in H₂ exchange.⁴⁴ The inhibition by substrate occurs as it reacts with 16-electron [Ir(H)₂(IMes)(sub)]Cl rather than H₂.

3.11. SABRE ¹H NMR signal enhancement by [Ir(H)₂(IMes)(pdz)₃]Cl (**4a**) and [Ir(H)₂(IMes)(phth)₃]Cl (**4b**)

Both **4a** and **4b** undergo ligand exchange on the NMR timescale. Such exchange is necessary for the SABRE effect to successfully hyperpolarise a material. We investigated the effect of shaking a sample of **1** (5.2 mM), containing 5 equivalents of pyridazine and 3 bar of *p*-H₂, in the Earth's magnetic field for 10 seconds at 298 K, prior to collecting a ¹H NMR spectrum. ¹H NMR signal enhancements are observed in all of the free and bound equatorial substrate resonances, and the hydride resonance of these complexes. No evidence for the formation of any dinuclear complexes was evident, and the phase of the observed hyperpolarised signals for the corresponding free and bound sites were identical.

Upon integration, the free pyridazine NCH resonance was 180-times larger than the corresponding signal recorded when the magnetisation was probed under thermal equilibrium conditions. The total signal enhancement of the two resonances in this substrate was 410-fold. When a similar phthalazine sample was probed in an identical way, the hydride signal for **4b**, and all of the free and equatorially bound substrate resonances were again enhanced. In this case, the corresponding free NCH resonance was 85-fold enhanced and the total molecular enhancement was 195-fold. The net enhancement factor therefore drops significantly on moving from pyridazine to phthalazine, despite their comparable ligand loss rates. Furthermore, the remote ring of phthalazine, which is connected by a 5-bond coupling of 0.43 Hz into the first ring is only weakly polarised by comparison to the first ring (14%). The underlying difference of 2.1-fold between the polarisation levels observed for these two molecules cannot be accounted for by a spin dilution effect (the consequence of sharing the magnetisation of *p*-H₂ with 6 rather than 4 receptor protons). By implication, the *J*-coupling that exists between the hydride and Ir-NCH protons of pyridazine must be larger than that to phthalazine, hence better polarisation transfer results; this fits with the predicted shorter bond length.

The concentration dependence on the level of SABRE seen in a series of ¹H NMR spectra recorded with $\pi/2$ pulses is illustrated in Fig. 6a for pyridazine and Fig. 6b for phthalazine. It is

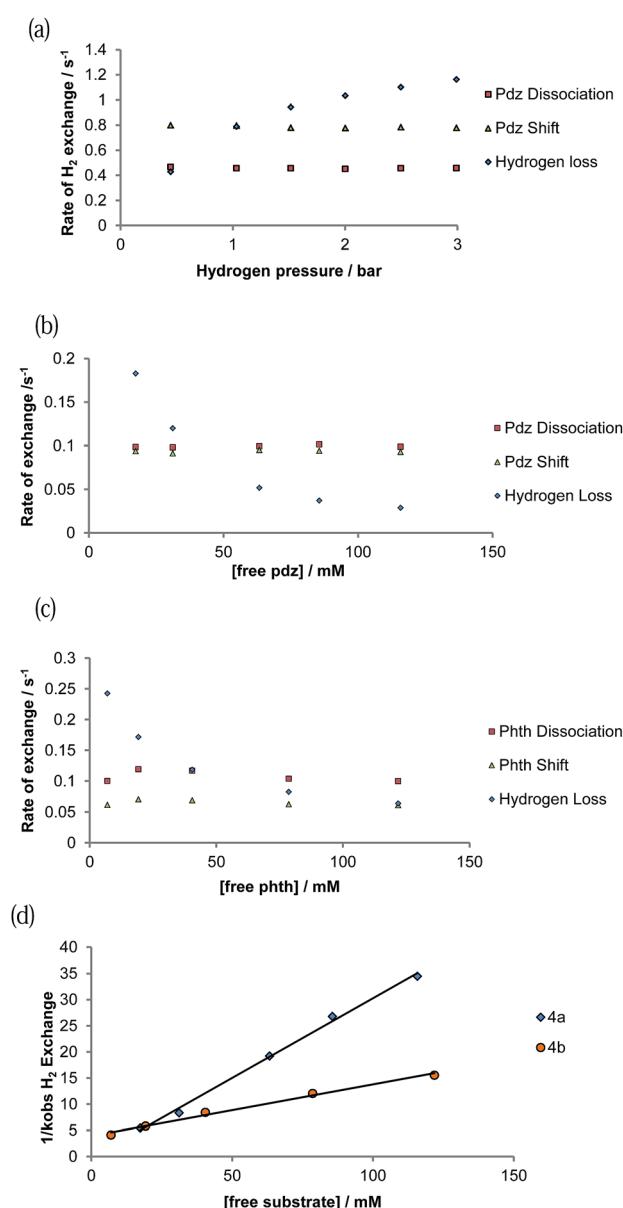


Fig. 5 Plot showing how the rate of ligand exchange varies as a function of ligand concentration for **4a** and **4b**: (a) [H₂] dependence, (b) [pyridazine] dependence (c) [phthalazine] dependence, (d) plot of the inverse of the H₂ loss rate versus ligand concentration.



clear, in these data, that the proton resonance which corresponds to the site next to nitrogen is most strongly enhanced with **4a** delivering a 380-fold NCH intensity gain when a 17 mM concentration of pyridazine is employed in conjunction with a metal concentration of 6.24 mM. The signal enhancement level falls as the substrate concentration increases, when a constant metal concentration is maintained. In addition to reflecting spin-dilution, this change also results in a progressive reduction in the rate of H_2 exchange. Both of these effects reduce the levels of signal gain seen for all of the protons of pyridazine and phthalazine. Surprisingly, when the inverse of the sum of the enhancement levels is plotted against equivalents of free substrate, a negative linear relationship can be seen (Fig. 6c).

The signal intensity gains that were observed in these 1H NMR spectra also varied with temperature as shown in Fig. 7. A continuous growth in the hyperpolarisation level with increasing temperature is observed between 273 K and 323 K, the highest temperature used. In the first of these samples, the concentration of free pyridazine was 12.3 mM while in the second, the concentration of free phthalazine was 14.1 mM. The

corresponding rates of H_2 loss at 323 K were estimated to be 29.7 s^{-1} and 13.49 s^{-1} respectively. This suggests that an optimal ligand exchange rate has not been reached at 323 K, despite the short lifetimes of these complexes.

The gain in signal strength appears to be linear with temperature, although the gradient exhibited by the 5 resonance intensities differs substantially, falling with distance from the ligation site. This change mirrors the efficiency of their polarisation. The gradients shown for the pyridazine signals are larger than those for phthalazine in accordance with its greater exchange rate sensitivity to temperature. Furthermore, the sensitivity to temperature falls as the substrate excess increases (Fig. 7c). This observation can be rationalised by the fall in H_2 exchange rates with increase in ligand concentration.

3.12. Probing the effect of sample dilution on the level of SABRE enhancement

In the case of pyridazine, the effect of the concentration of **4a** (with the excess of free pyridazine kept constant) on the level of SABRE enhancement has been rigorously studied. The smallest ligand excess delivered the largest level of polarisation,

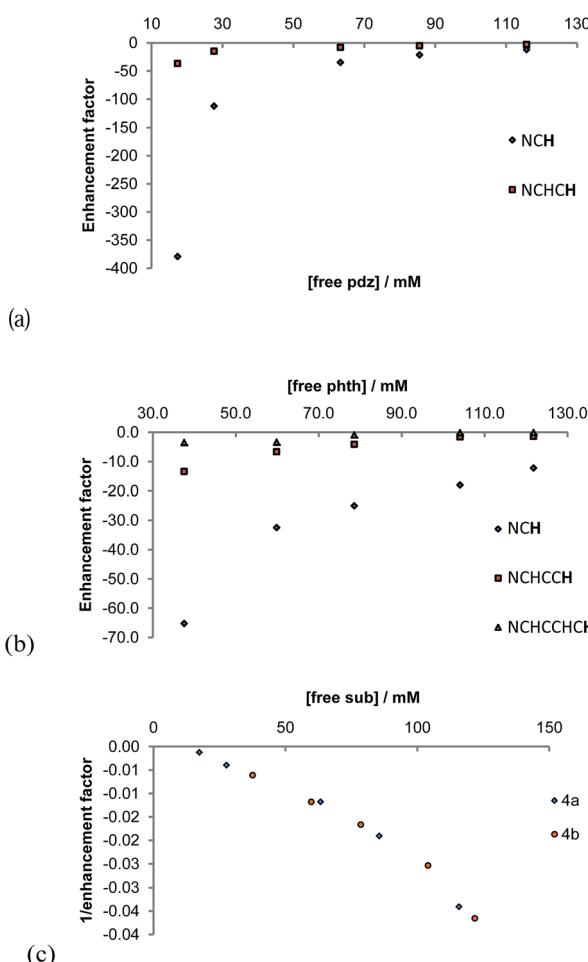


Fig. 6 Plots showing the effect of substrate concentration, for a constant metal concentration, on the level of pyridazine and phthalazine hyperpolarisation achieved by **4a** (a) and **4b** (b) with plot (c) revealing how the reciprocal of the total enhancement level relates to substrate concentration.

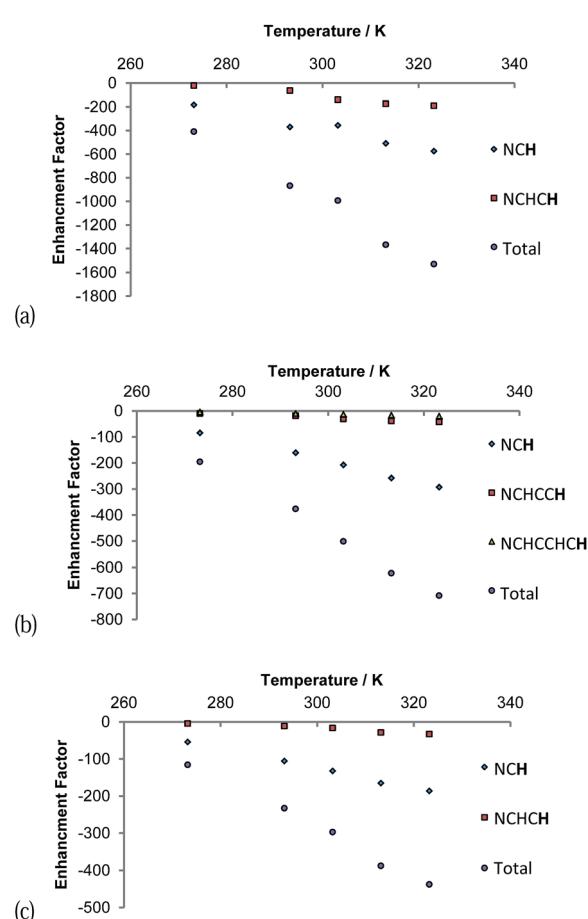


Fig. 7 Plots of reaction temperature versus (a) pyridazine (12.3 mM free pdz, 2-fold excess) and (b) phthalazine (14.1 mM free pth, 2.3-fold excess) proton hyperpolarisation level under SABRE. Plot (c) presents analogous data when the pyridazine excess is 12.1-fold (75.1 mM free pdz).

alongside a metal concentration of 2.8 mM. The maximum total signal enhancement observed in the ^1H NMR spectra collected using pyridazine was 1278-fold with a $\pi/2$ pulse. Normally, the S/N value associated with a given resonance is proportional to concentration. Here, however, the S/N value is linked to the product of the concentration and the hyperpolarisation level. An approximately linear dependence on the detected signal strength with concentration was observed experimentally for samples in which the substrate concentration lay between 2.8 mM and 6.2 mM, as shown in Fig. 8. We conclude from these observations that there is no benefit in working at higher concentration, even if there is an abundance of the available substrate, since the fall in SABRE level acts to offset the signal gain resulting from an increase in concentration.^{68,69}

3.13. Probing the effect of the polarisation transfer field on the efficiency of SABRE

The variation in the level of SABRE observed with the magnetic field experienced by the sample at the point of polarisation transfer is substantial (see ESI †). Both substrates behave in a similar way, each yielding their largest free ligand single spin enhancement values when the polarisation transfer field is 65 G. Consequently, this field should be used for detection. SABRE, however, also creates higher order magnetic terms which are optimally enhanced at different field values (see ESI †).⁷⁰ It is possible to selectively observe the signals from these substrates without the need for a deuterated solvent by employing the OPSY protocol as shown in Fig. 9.^{71,72}

3.14. SABRE enhancements delivered by $[\text{Ir}(\text{H})_2(\text{IMes})(\text{pdz})_3]\text{Cl}$ (4a) and $[\text{Ir}(\text{H})_2(\text{IMes})(\text{phth})_3]\text{Cl}$ (4b) as a function of the polarisation transfer time

Increasing the time that these samples are exposed to $p\text{-H}_2$ during the SABRE step further increases the level of signal enhancement. The plots in ESI Fig. 13–23 † show how the resonance intensities vary with the polarisation transfer time.

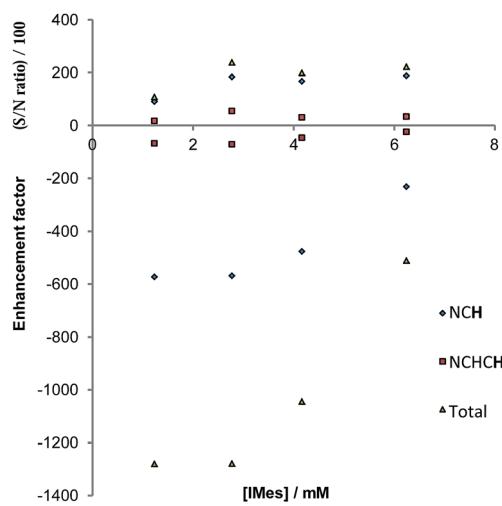


Fig. 8 Plots of S/N (upper), and the indicated hyperpolarised ^1H NMR signal intensities (lower) of pyridazine as a function of catalyst concentration.

Ultimately, the maximum polarisation level reaches a plateau as $T_{1(\text{effective})}$ of the probed state counteracts the rate of SABRE build-up. The polarisation builds up most quickly in the single spin NCH protons, and ultimately reaches a higher polarisation level than any other spin state. While the observed rate of two-spin order build up is comparable to that of a single spin their ultimate amplitudes are lower as a consequence of their more rapid relaxation (see ESI †).

3.15. Using SABRE to collect MRI data

We recorded a series of SABRE images and compared them to their thermal counterparts (Fig. 10). As expected, these images exhibited better signal to noise, improved contrast, and signal intensity. Furthermore, the samples of pyridazine were superior to those of phthalazine in terms of signal intensity (one order of magnitude higher) for the same concentration of substrate.

In order to more precisely quantify the improvement in image quality, five one-shot images of the hyperpolarised samples were acquired and the resulting image SNR's calculated. The average result was divided by the SNR of a reference image that was recorded under Boltzmann equilibrium conditions using 512 averages. The underlying improvement in contrast and image quality is revealed in Table 5. A considerable increase in the SNR values was observed, and the acquisition times required for SABRE images were 6 orders of magnitude smaller than the scan times necessary under thermal equilibrium conditions (260 ms *versus* several hours).

The optimum conditions used to hyperpolarise both pyridazine and phthalazine in methanol, at 298 K, have been applied to 5-aminophthalazine, which has antifungal properties.⁷³ Preliminary results are very promising – with total signal enhancements of *ca.* 500-fold being observed. The applicability of SABRE, in hyperpolarising pyridazine and phthalazine-derived drug molecules for use as contrast agents, is dependent ultimately on performance in biologically-compatible solvents. Although water would seem an obvious solvent choice, SABRE

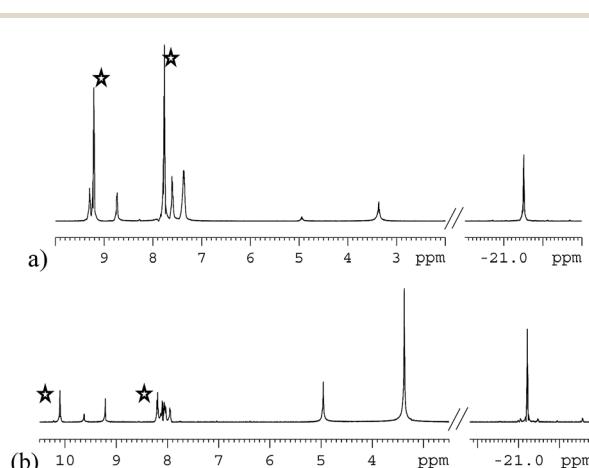


Fig. 9 Double quantum selected ^1H NMR spectra of protio-methanol solutions of **1** (5.2 mM) containing 5 equivalents of pyridazine (a), and phthalazine (b) after SABRE. Free substrate resonances are indicated (stars).



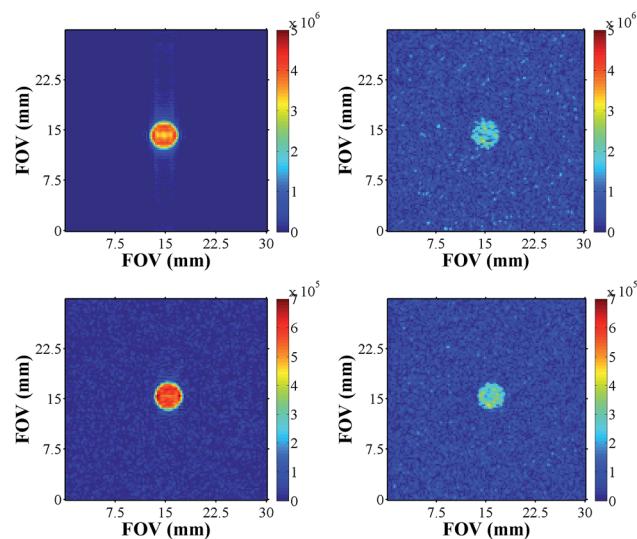


Fig. 10 Hyperpolarised images of pyridazine (upper, left) and phthalazine (lower left) compared to their respective thermally equilibrated images (right).

Table 5 Average, and maximum values, of the absolute SNR observed in the processed images of hyperpolarised pyridazine and phthalazine and the ratios of hyperpolarised to reference image SNR respectively

Catalyst	SNR _{hyp}	SNR _{hyp} (max. value)	SNR _{hyp} /SNR _B	SNR _{hyp} /SNR _B (max. value)
4a	185	234	1100	1392
4b	72	96	229	306

has not yet been shown to work efficiently in this medium.^{37,39} An ethanol–water mixture is, however, biocompatible, hence the use of ethanol, followed by dilution has been predicted to be viable.^{70,74} We have therefore examined pyridazine and phthalazine in ethanol-*d*₆. Our preliminary results reveal superior SABRE in this solvent, with the pyridazine signal enhancement of *ca.* 400-fold (9.62-fold excess, 60 mM free pyridazine) being 33% better than in methanol. For phthalazine the total signal enhancement was 480-fold (2.12-fold excess, 13.2 mM free phthalazine) or 20% better. Our studies on 5-aminophthalazine confirm that further functionalisation of the remote ring will not prevent SABRE activity and an array of such materials could be probed.⁷⁵

4. Conclusions

We have established that [Ir(IMes)(COD)Cl] (**1**) reacts rapidly with pyridazine and phthalazine to form the ligand substitution products [Ir(IMes)(COD)(pdz)]Cl, **2a** and [Ir(IMes)(COD)(phth)]Cl **2b**, respectively. Phthalazine is more basic than pyridazine, and **2b** is more stable than **2a**.⁷⁶ These complexes undergo a haptotropic shift on the NMR timescale which is more rapid for **2b**.

Complexes **2a** and **2b** add H₂ to form the corresponding Ir(III) complexes [Ir(H)₂(COD)(IMes)(pdz)]Cl (**3a**) and [Ir(H)₂(COD)(IMes)(phth)]Cl (**3b**), where **3b** is more stable than **3a**. When the H₂ addition reaction was monitored with *p*-H₂, PHIP was observed in the hydride NMR signals of these two products and **2a** is more reactive to H₂ addition than **2b**. **3a** and **3b** also exhibit SABRE in specific ligand resonances that lie *trans* to the hydrides. This demonstrates that hyperpolarisation can be used to characterise such complexes, even if they have a transient existence.

3a and **3b** are highly reactive and upon warming they rapidly convert to the dihydrides [Ir(H)₂(IMes)(pdz)]Cl (**4a**) and [Ir(H)₂(IMes)(phth)]Cl (**4b**). These two complexes also undergo a haptotropic ligand shift in addition to full ligand loss on the NMR timescale. The iridium–nitrogen bonds of **4a** are stronger than those of **4b**. The hyperpolarisation of both pyridazine and phthalazine by **4a** and **4b** respectively, under SABRE, proved to be facile. NMR enhancements increase with temperature, due to faster ligand exchange, and are optimised with 5 equivalents of substrate, and a polarisation transfer field of 65 G. An optimised enhancement level, at 298 K, of 1278-fold was achieved for pyridazine.

The total signal enhancement of pyridazine was consistently larger than that of phthalazine even though the ligand loss rates of **4a** and **4b** were comparable. This difference in behaviour cannot be accounted for by the sharing of magnetisation from *p*-H₂ with 4 rather than 6 receptor protons, but instead suggests that the *J*-coupling between the hydride and Ir–NCH protons of pyridazine is larger than that to phthalazine. This deduction agrees with the predicted shorter Ir–N bond length in **4a**. A series of more complex magnetic states can also be created which allow the observation of these substrates in protic solvents through the OPSY protocol.

Finally, we have shown that it is possible to collect optimised MRI images for these substrates on phantoms. Whilst neither pyridazine nor phthalazine are used clinically, their derivatives, in the form of drugs such as Hydralazine,⁷⁷ play an important role in medicine; we have polarised 5-aminophthalazine which has antifungal properties and achieved total enhancement levels of 500-fold. In the future, we plan to use the results of this study to aid in the optimisation of SABRE for the analysis of similar heterocycles that are used pharmaceutically.

Conflict of interest

The authors have no competing financial interest.

Abbreviations

COA	Cyclooctane
COD	1,5-Cyclooctadiene
COE	Cyclooctene
DFT	Density functional theory
η	Eta coordination
EXSY	Exchange spectroscopy
IMes	1,3-Bis(2,4,6-trimethylphenyl)imidazol-2-ylidene
MRI	Magnetic resonance imaging
nOesy	Nuclear overhauser effect spectroscopy



NMR	Nuclear magnetic resonance
pdz	Pyridazine
<i>p</i> -H ₂	para-Hydrogen
PHIP	para-Hydrogen-induced polarisation
phth	Phthalazine
SABRE	Signal amplification by reversible exchange
σ-CAM	Sigma-complex assisted metathesis
<i>S/N</i>	Signal to noise ratio

- 13 Y. Chen and R. E. Shepherd, *Inorg. Chim. Acta*, 1998, **279**, 85–94.
- 14 K. R. Dixon, D. T. Eadie and S. R. Stobart, *Inorg. Chem.*, 1982, **21**, 4318–4321.
- 15 E. W. Abel, P. J. Heard, K. G. Orrell and V. Sik, *Polyhedron*, 1994, **13**, 2907–2913.
- 16 E. W. Abel, P. J. Heard, K. G. Orrell, M. B. Hursthouse and K. M. A. Malik, *J. Chem. Soc., Dalton Trans.*, 1995, 3165–3171.
- 17 E. W. Abel, P. J. Heard and K. G. Orrell, *Inorg. Chim. Acta*, 1997, **255**, 65–71.
- 18 S. K. Kang, T. A. Albright and C. Mealli, *Inorg. Chem.*, 1987, **26**, 3158–3166.
- 19 S. Alvarez, M. J. Bermejo and J. Vinaixa, *J. Am. Chem. Soc.*, 1987, **109**, 5316–5323.
- 20 J. Jeener, B. H. Meier, P. Bachmann and R. R. Ernst, *J. Chem. Phys.*, 1979, **71**, 4546–4553.
- 21 K. Golman, R. in't Zandt and M. Thaning, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 11270–11275.
- 22 J. Feeney, J. G. Batchelor, J. P. Albrand and G. C. K. Roberts, *J. Magn. Reson.*, 1979, **33**, 519–529.
- 23 G. Binsch, *J. Am. Chem. Soc.*, 1969, **91**, 1304–1309.
- 24 J. Natterer and J. Bargon, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1997, **31**, 293–315.
- 25 R. A. Green, R. W. Adams, S. B. Duckett, R. E. Mewis, D. C. Williamson and G. G. R. Green, *Prog. Nucl. Magn. Reson. Spectrosc.*, 2012, **67**, 1–48.
- 26 C. R. Bowers and D. P. Weitekamp, *Phys. Rev. Lett.*, 1986, **57**, 2645–2648.
- 27 S. B. Duckett and C. J. Sleigh, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1999, **34**, 71–92.
- 28 D. Blazina, S. B. Duckett, J. P. Dunne and C. Godard, *Dalton Trans.*, 2004, 2601–2609.
- 29 S. Gloggler, J. Colell and S. Appelt, *J. Magn. Reson.*, 2013, **235**, 130–142.
- 30 T. C. Eisenschmid, R. U. Kirss, P. P. Deutsch, S. I. Hommeltoft and R. Eisenberg, *J. Am. Chem. Soc.*, 1987, **109**, 8089–8091.
- 31 S. B. Duckett and R. E. Mewis, *Acc. Chem. Res.*, 2012, **45**, 1247–1257.
- 32 R. W. Adams, J. A. Aguilar, K. D. Atkinson, M. J. Cowley, P. I. P. Elliott, S. B. Duckett, G. G. R. Green, I. G. Khazal, J. Lopez-Serrano and D. C. Williamson, *Science*, 2009, **323**, 1708–1711.
- 33 K. D. Atkinson, M. J. Cowley, S. B. Duckett, P. I. P. Elliott, G. G. R. Green, J. Lopez-Serrano, I. G. Khazal and A. C. Whitwood, *Inorg. Chem.*, 2009, **48**, 663–670.
- 34 T. Theis, M. Truong, A. M. Coffey, E. Y. Chekmenev and W. S. Warren, *J. Magn. Reson.*, 2014, **248**, 23–26.
- 35 A. N. Pravdivtsev, A. V. Yurkovskaya, H. M. Vieth and K. L. Ivanov, *Phys. Chem. Chem. Phys.*, 2014, **16**, 24672–24675.
- 36 K. D. Atkinson, M. J. Cowley, P. I. P. Elliott, S. B. Duckett, G. G. R. Green, J. Lopez-Serrano and A. C. Whitwood, *J. Am. Chem. Soc.*, 2009, **131**, 13362–13368.
- 37 H. F. Zeng, J. D. Xu, M. T. McMahon, J. A. B. Lohman and P. C. M. van Zijl, *J. Magn. Reson.*, 2014, **246**, 119–121.
- 38 J. B. Hovener, S. Knecht, N. Schwaderlapp, J. Hennig and D. von Elverfeldt, *ChemPhysChem*, 2014, **15**, 2451–2457.

Acknowledgements

Funding Sources. We are grateful for financial support from the EPSRC (grant no. EP/G009546/1), the MRC (studentship KDA) and Bruker BioSpin (CASE studentship KMA, and equipment). We also acknowledge helpful discussions with Dr J. Lohman. SBD, IJSF and GGRG would like to thank the Wellcome Trust and Wolfson Foundation (grants 092506 and 098335).

Notes and references

- 1 M. Kuwahara, Y. Kawano, M. Kajino, Y. Ashida and A. Miyake, *Chem. Pharm. Bull.*, 1997, **45**, 1447–1457.
- 2 C. Lamberth, S. Trah, S. Wendeborn, R. Dumeunier, M. Courbot, J. Godwin and P. Schneiter, *Bioorg. Med. Chem.*, 2012, **20**, 2803–2810.
- 3 D. G. H. Livermore, R. C. Bethell, N. Cammack, A. P. Hancock, M. M. Hann, D. V. S. Green, R. B. Lamont, S. A. Noble, D. C. Orr, J. J. Payne, M. V. J. Ramsay, A. H. Shingler, C. Smith, R. Storer, C. Williamson and T. Willson, *J. Med. Chem.*, 1993, **36**, 3784–3794.
- 4 L. S. Chen, S. Redkar, D. Bearss, W. G. Wierda and V. Gandhi, *Blood*, 2009, **114**, 4150–4157.
- 5 V. Pogacic, A. N. Bullock, O. Fedorov, P. Filippakopoulos, C. Gasser, A. Biondi, S. Meyer-Monard, S. Knapp and J. Schwaller, *Cancer Res.*, 2007, **67**, 6916–6924.
- 6 S. Bingham, P. J. Beswick, C. Bountra, T. Brown, I. B. Campbell, I. P. Chessell, N. Clayton, S. D. Collins, P. T. Davey, H. Goodland, N. Gray, C. Haslam, J. P. Hatcher, A. J. Hunter, F. Lucas, G. Murkitt, A. Naylor, E. Pickup, B. Sargent, S. G. Summerfield, A. Stevens, S. C. Stratton and J. Wiseman, *J. Pharmacol. Exp. Ther.*, 2005, **312**, 1161–1169.
- 7 J. M. Craft, D. M. Watterson and L. J. Van Eldik, *Expert Opin. Ther. Targets*, 2005, **9**, 887–900.
- 8 E. Abignente, F. Arena, E. Luraschi, C. Saturnino, E. Marmo, L. Berrino and D. Donnoli, *Farmaco*, 1990, **45**, 1075–1087.
- 9 T. M. Bare, J. F. Resch and P. F. Jackson, *US Pat.*, US 5652239, 1997.
- 10 W. R. Carling, A. M. MacLeod, R. McKernan, A. J. Reeve, F. Sternfeld and L. J. Street, *US Pat.*, US 6465462, 2002.
- 11 E. W. Abel, E. S. Blackwall, P. J. Heard, K. G. Orrell, V. Sik, M. B. Hursthouse, M. A. Mazid and K. M. A. Malik, *J. Chem. Soc., Dalton Trans.*, 1994, 445–455.
- 12 S. S. Eaton, G. R. Eaton and R. H. Holm, *J. Organomet. Chem.*, 1972, **39**, 179–195.



39 M. L. Truong, F. Shi, P. He, B. X. Yuan, K. N. Plunkett, A. M. Coffey, R. V. Shchepin, D. A. Barskiy, K. V. Kovtunov, I. V. Koptyug, K. W. Waddell, B. M. Goodson and E. Y. Chekmenev, *J. Phys. Chem. B*, 2014, **118**, 13882–13889.

40 D. A. Barskiy, K. V. Kovtunov, I. V. Koptyug, P. He, K. A. Groome, Q. A. Best, F. Shi, B. M. Goodson, R. V. Shchepin, A. M. Coffey, K. W. Waddell and E. Y. Chekmenev, *J. Am. Chem. Soc.*, 2014, **136**, 3322–3325.

41 H. Zeng, J. Xu, J. Gillen, M. T. McMahon, D. Artemov, J.-M. Tyburn, J. A. B. Lohman, R. E. Mewis, K. D. Atkinson, G. G. R. Green, S. B. Duckett and P. C. M. van Zijl, *J. Magn. Reson.*, 2013, **237**, 73–78.

42 E. B. Duecker, L. T. Kuhn, K. Muennemann and C. Griesinger, *J. Magn. Reson.*, 2012, **214**, 159–165.

43 M. Fekete, O. Bayfield, S. B. Duckett, S. Hart, R. E. Mewis, N. Pridmore, P. J. Rayner and A. Whitwood, *Inorg. Chem.*, 2013, **52**, 13453–13461.

44 M. J. Cowley, R. W. Adams, K. D. Atkinson, M. C. R. Cockett, S. B. Duckett, G. G. R. Green, J. A. B. Lohman, R. Kerssebaum, D. Kilgour and R. E. Mewis, *J. Am. Chem. Soc.*, 2011, **133**, 6134–6137.

45 O. Torres, M. Martin and E. Sola, *Organometallics*, 2009, **28**, 863–870.

46 K. Stott, J. Stonehouse, J. Keeler, T. L. Hwang and A. J. Shaka, *J. Am. Chem. Soc.*, 1995, **117**, 4199–4200.

47 J. Stonehouse, P. Adell, J. Keeler and A. J. Shaka, *J. Am. Chem. Soc.*, 1994, **116**, 6037–6038.

48 A. A. Shaw, C. Salaun, J. F. Dauphin and B. Ancian, *J. Magn. Reson.*, 1996, **120**, 110–115.

49 B. Ancian, I. Bourgeois, J. F. Dauphin and A. A. Shaw, *J. Magn. Reson.*, 1997, **125**, 348–354.

50 H. Kessler, H. Oschkinat and C. Griesinger, *J. Magn. Reson.*, 1986, **70**, 106–133.

51 N. G. Vassilev and V. S. Dimitrov, *Magn. Reson. Chem.*, 2001, **39**, 607–614.

52 C. Bauer, R. Freeman, T. Frenkiel, J. Keeler and A. J. Shaka, *J. Magn. Reson.*, 1984, **58**, 442–457.

53 K. Stott, J. Keeler, Q. N. Van and A. J. Shaka, *J. Magn. Reson.*, 1997, **125**, 302–324.

54 W. D. Jones, G. P. Rosini and J. A. Maguire, *Organometallics*, 1999, **18**, 1754–1760.

55 M. L. H. Green, L. L. Wong and A. Sella, *Organometallics*, 1992, **11**, 2660–2668.

56 D. C. Harris, *J. Chem. Educ.*, 1998, **75**, 119–121.

57 J. Hennig, A. Nauerth and H. Friedburg, *Magn. Reson. Med.*, 1986, **3**, 823–833.

58 H. J. S. Machado and A. Hinchliffe, *J. Mol. Struct.: THEOCHEM*, 1995, **339**, 255–258.

59 C. E. Johnson and R. Eisenberg, *J. Am. Chem. Soc.*, 1985, **107**, 3148–3160.

60 A. L. Sargent, M. B. Hall and M. F. Guest, *J. Am. Chem. Soc.*, 1992, **114**, 517–522.

61 A. L. Sargent and M. B. Hall, *Inorg. Chem.*, 1992, **31**, 317–321.

62 K. Searles, M. Pink, K. G. Caulton and D. J. Mindiola, *Dalton Trans.*, 2012, 9619–9622.

63 C. E. Johnson, B. J. Fisher and R. Eisenberg, *J. Am. Chem. Soc.*, 1983, **105**, 7772–7774.

64 R. F. R. Jazzaar, S. A. Macgregor, M. F. Mahon, S. P. Richards and M. K. Whittlesey, *J. Am. Chem. Soc.*, 2002, **124**, 4944–4945.

65 M. Martin, E. Sola, O. Torres, P. Plou and L. A. Oro, *Organometallics*, 2003, **22**, 5406–5417.

66 A. Bax, S. W. Sparks and D. A. Torchia, *Methods Enzymol.*, 1989, **176**, 134–150.

67 L. Pazderski, *Magn. Reson. Chem.*, 2008, **46**, S3–S15.

68 N. Eshuis, N. Hermkens, B. J. A. van Weerdenburg, M. C. Feiters, F. Rutjes, S. S. Wijmenga and M. Tessari, *J. Am. Chem. Soc.*, 2014, **136**, 2695–2698.

69 S. Gloggler, M. Emondts, J. Colell, R. Muller, B. Blumich and S. Appelt, *Analyst*, 2011, **136**, 1566–1568.

70 R. E. Mewis, K. D. Atkinson, M. J. Cowley, S. B. Duckett, G. G. R. Green, R. A. Green, L. A. R. Highton, D. Kilgour, L. S. Lloyd, J. A. B. Lohman and D. C. Williamson, *Magn. Reson. Chem.*, 2014, **52**, 358–369.

71 J. A. Aguilar, P. I. P. Elliott, J. Lopez-Serrano, R. W. Adams and S. B. Duckett, *Chem. Commun.*, 2007, 1183–1185.

72 J. A. Aguilar, R. W. Adams, S. B. Duckett, G. G. R. Green and R. Kandiah, *J. Magn. Reson.*, 2011, **208**, 49–57.

73 C.-K. Ryu, R.-E. Park, M.-Y. Ma and J.-H. Nho, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 2577–2580.

74 J. B. Hovener, N. Schwaderlapp, R. Borowiak, T. Lickert, S. B. Duckett, R. E. Mewis, R. W. Adams, M. J. Burns, L. A. R. Highton, G. G. R. Green, A. Olaru, J. Hennig and D. von Elverfeldt, *Anal. Chem.*, 2014, **86**, 1767–1774.

75 A. Gomtsyan, E. K. Bayburt, R. G. Schmidt, G. Z. Zheng, R. J. Perner, S. Didomenico, J. R. Koenig, S. Turner, T. Jinkerson, I. Drizin, S. M. Hannick, B. S. Macri, H. A. McDonald, P. Honore, C. T. Wismer, K. C. Marsh, J. Wetter, K. D. Stewart, T. Oie, M. F. Jarvis, C. S. Surowy, C. R. Faltynek and C.-H. Lee, *J. Med. Chem.*, 2005, **48**, 744–752.

76 M. Brorson, M. R. Dyxenburg, F. Galsbol and K. Simonsen, *Acta Chem. Scand.*, 1996, **50**, 289–293.

77 J. N. Cohn, G. Johnson, S. Ziesche, F. Cobb, G. Francis, F. Tristani, R. Smith, W. B. Dunkman, H. Loeb, M. L. Wong, G. Bhat, S. Goldman, R. D. Fletcher, J. Doherty, C. V. Hughes, P. Carson, G. Cintron, R. Shabetai and C. Haakenson, *N. Engl. J. Med.*, 1991, **325**, 303–310.

