Flaws in foldamers: conformational uniformity and signal decay in achiral helical peptide oligomers†

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Although foldamers, by definition, are extended molecular structures with a well-defined conformation, minor conformers must be populated at least to some extent in solution. We present a quantitative analysis of these minor conformers for a series of helical oligomers built from achiral but helicogenic α-amino acids. By measuring the chain length dependence or chain position dependence of NMR or CD quantities that measure screw-sense preference in a helical oligomer, we quantify values for the decay constant of a conformational signal as it passes through the molecular structure. This conformational signal is a perturbation of the racemic mixture of M and P helices that such oligomers typically adopt by the inclusion of an N or C terminal chiral inducer. We show that decay constants may be very low (<1% signal loss per residue) in non-polar solvents, and we evaluate the increase in decay constant that results in polar solvents, at higher temperatures, and with more conformationally flexible residues such as Gly. Decay constants are independent of whether the signal originates from the N or the C terminus. By interpreting the decay constant in terms of the probability with which conformations containing a screw-sense reversal are populated, we quantify the populations of these alternative minor conformers within the overall ensemble of secondary structures adopted by the foldamer. We deduce helical persistence lengths for Aib polymers that allow us to show that in a non-polar solvent a peptide helix, even in the absence of chiral residues, may continue with the same screw sense for approximately 200 residues.

Introduction

A foldamer is ‘a polymer [or oligomer] with a strong tendency to adopt a specific compact conformation’, and foldamers4-6 can typically be viewed as synthetic analogues of biopolymers – peptides, proteins and nucleic acids.7 The three-dimensional structure of such biopolymers is in many cases characterised by the existence of one particularly favourable conformation, with this gross conformational uniformity being intimately linked with function. Examples include the helical conformation of DNA under normal conditions and structural proteins such as collagen.9 However, many other biopolymers exhibit more complex conformational behaviour in which alternative conformers may be populated. Classic examples10 include allosterically switchable molecules such as haemoglobin11 or phosphorylase enzymes,12 and conformationally switchable proteins involved in signal transduction.13,14 Yet other biopolymers appear to be almost fully unstructured in a three-dimensional sense, with many energetically similar alternative conformational states being populated.15 Synthetic analogues of these less conformationally uniform biomolecules should form their own group within the general class of foldamers, comprising structures to which more than one well defined ‘specific compact conformation’ is available. Indeed, entropic considerations make it reasonable to expect that even the most conformationally well defined foldamers must spend at least some of their time in minor, less favourable conformations. Studies of the relative tendency towards helicity of alternative monomer units have been made in some foldamer classes by exploring their ability to communicate a screw-sense preference between two helical domains.16,17 More detailed exploration by classic spectroscopic techniques of the multiple conformers populated by some peptidomimetic structures poses well-established difficulties.18-20 For this reason, the intrusion of alternative conformers into foldamer structures of uniform primary sequence has rarely been quantified, explored, or exploited.21 Nonetheless, such studies are essential if the broader field of foldamer chemistry is to deliver biomimetic function as well as biomimetic structure.

Oligomers of achiral quaternary amino acids, of which 2-aminoisobutyric acid (Aib) is the archetype, but which also include 1-aminocyclohexanecarboxylic acid (Ac6c, Fig. 1) along with 1-aminocyclopentanecarboxylic acid (Ac5c), diethylglycine...
We may consider a conformational perturbation such as the screw-sense preference induced by the chiral N-terminal \(\alpha\)-MeVal residues of 1 as a signal that propagates through the polymeric molecule without erosion only if that molecule truly adopts one single ‘specific compact conformation’. Any random intrusion of alternative conformations must attenuate the signal, allowing the local conformational preference within the helix to decay towards an equal population of left- and right-handed helices the further the polymer is extended away from the chiral terminus.

It is the occurrence of these alternative minor foldamer conformations in the solution state that we explore in this paper. We use NMR and other spectroscopic methods to quantify the attenuation of a conformational preference with increasing distance from a source of screw-sense induction located at either terminus of a series of Aib-containing foldamers. We show that a conformational signal is in many cases remarkably persistent, but its decrease in intensity on moving in either direction through the polymer may be characterized as a solvent-dependent exponential decay. We interpret the spatial decay constant of the signal in terms of the stochastic intrusion of screw-sense reversals into the three-dimensional structure of the oligomers, and use the results to quantify the population of minor contributors to the overall ensemble of conformations that describe the dynamic three-dimensional structure of the foldamer.

**Results**

**Solvent-dependent signal decay moving away from the N terminus**

The reactions shown in Fig. 1 (ref. 48) exhibit remote stereochemical control as a result of quantitative induction by the N-terminal chiral dimer (shown in green) of a screw-sense preference in the achiral helical chain (shown in blue and black). As a result the reaction site (shown in red) finds itself in a nonracemic environment, and reacts diastereoselectively by face-selective attack on the arene on an acyliminium ion generated *in situ*. The diastereoselectivity (measured as diastereoisomeric excess, de) of the reaction is in each case the product of the induced helical excess at the N-terminal dimer (he), the fidelity with which this helical excess is communicated to the C terminus, and the diastereofacial selectivity (ds) with which a pure M or P helix would induce attack on the iminium ion intermediate in the acid-catalysed reaction. As the first and last of these factors are constant in all four reactions, the decay in diastereoselectivity in the reactions of the oligomers 1a–d in Fig. 1, from 93 : 7 dr (86% de) for a 1.16 relationship to 88 : 12 dr (76% de) for a 1.61 relationship, allows us to make an initial estimate of the effect of oligomer length on the ability of an Aib/Ac6c-containing helix to communicate a screw-sense preference. Fig. 2a shows diastereoselectivity of the reaction 1a–d \(\to\) 2a–d plotted against the distance in residues from the inducer of helical preference at residue \(t\) to the reaction site. On the assumption that Aib and Ac6c have equivalent conformational preferences, we fitted exponential decays to these curves, using the formula

\[
\frac{d}{dt} = -k \cdot \theta
\]

where \(\theta\) is the concentration of the inducer Aib and \(k\) is a rate constant. The fit to this equation gives an initial estimate of the decrease in de on moving away from the reaction centre of 1a–d. Since the concentration of the inducer is constant in these experiments, this decrease in de reflects the random intrusion of alternative conformations that can adopt a form of switchability analogous to that of achiral foldamers.

In summary, the results in this paper demonstrate that even artificial oligomers can communicate stereochemical information in a manner that reflects the properties of their natural counterparts. The ability of these oligomers to adopt a specific conformation, the resilience of this property, and the mechanism by which this property is transmitted through the polymer, are all the subject of future work.
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Diastereoselectivity of the reaction corresponds to a ds of 95% to 5% for seven to 26 residues, where de is the diastereoselectivity induced in a reaction, and ds is the diastereomeric excess of the reaction.

We deduce that de decays by a factor of 0.8% per residue for (L-Meval)2 of 99% (ref. 31) to deduce that the inherent diastereoselectivity is 95%.

Fig. 1 shows how the separation $\Delta \delta$ of the CHN signals of the diastereomers at 23 °C in deuterated THF and at +23 °C or 40 °C in a more polar solvent, deuterated ethanol, and Fig. 2b plots anisochronicity against characteristic chain length.

The same arguments must hold for pairs of signals arising from corresponding groups in diastereomeric structures. The detailed factors governing their anisochronicity will be more complicated to analyse, but these detailed conformational considerations are irrelevant to the extraction of an exponential decay factor. A decrease in the fast exchange regime anisochronicity of peaks arising from two diastereoisomers on increasing the length of the achiral segment separating the stereogenic centres that give rise to their diastereomeric relationship may be interpreted simply as a measure of that segment’s decreasing ability to communicate stereochemical information between the relevant stereogenic centres.

The detailed factors governing their anisochronicity of peaks arising from two diastereoisomers are located at residue $i + n$, $D_{ij}$ is the anisochronicity of the CHN signals when they are located at residue $i + n$, and $\Delta \delta_i$ simply a limiting value to which we assign no chemical significance. Decay constants $f$ (the per-residue decrease in $D_{ij}$) can be extracted of 0.1% for THF at 23 °C, 1.9% for ethanol at 23 °C, and 2.5% for ethanol at 40 °C. The near-zero value of the decay constant in THF confirms the observation from Fig. 2b that the screw-sense preference is highly persistent in this solvent. A more polar solvent and a higher temperature lead to a greater rate of decay, presumably by facilitating the breaking of intramolecular...
Fig. 3 Three Gly residues as NMR reporters of helical excess embedded in an Aib-rich oligomer.

hydrogen bonds and by increasing the population of alternative minor conformers.

An alternative, simpler measure of signal decay may be obtained by inserting several NMR reporters, each containing a pair of diastereotopic groups giving rise to potentially anisochronous signals, at successive positions along a helical oligomer. Provided that the reporter groups are sufficiently far from the ends of the chain for their anisochronicity to be dictated solely by their local helical environment, the decrease in their anisochronicity along the chain gives a measure of signal decay. The challenge in such an approach is identifying which of several identical reporters is which. This is less difficult if the rate at which anisochronicity decreases is relatively high, because the signals from successive reporters are not superimposed upon one another, and a simplifying assumption can be made that anisochronicity can only decrease along the chain. We applied this principle to oligomer 3 (Fig. 3) which contains three glycine residues. These serve the dual purpose of firstly introducing greater flexibility than Aib, and therefore a steeper rate of decay of screw-sense preference, and secondly of providing pairs of diastereotopic protons as spectroscopic reporters of screw-sense preference. In ethanol and methanol, the three glycine ABX systems were resolved by Lorentz–Gauss transformation, while in CDCl3 and in CD2Cl2 poor dispersion prevented reliable assignment. In THF, however, the A and B coupled signals of the three ABX spin systems were identified using an absorption-mode 2D J-resolved experiment based on the PSYCHE method.66,67 Fig. 4 plots the fast exchange anisochronicity $\Delta\delta$ between the A and B components of the AB systems against the position $n$ of the relevant Gly residue relative to the N terminal Cbz-L-Phe at position $i$, and in each case a value for $f$ was obtained using eqn (3).

This time, the additional conformational flexibility complicates the analysis of the decay constants. Gly introduces much more flexibility than Aib into a peptide chain,65,68 so we expect the values of $f$ for Aib ($f_{\text{Aib}}$) and for Gly ($f_{\text{Gly}}$) to differ significantly. There are four Aib residues for every Gly in the chain, so the value $f$ obtained from fitting eqn (3) to Fig. 4 is therefore:

$$f = 0.8f_{\text{Aib}} + 0.2f_{\text{Gly}} \quad (4)$$

The values of $f$ obtained exhibit a solvent dependence consistent with that noted for 2, with the more polar methanol allowing greater conformational flexibility than the less polar THF, and ethanol lying somewhere between. Using our previous estimates of values for $f_{\text{Aib}}$ at $+23 \, ^\circ\text{C}$ in THF, in ethanol (Fig. 2) and in methanol,31,34 we can further deduce values of $f_{\text{Gly}}$ from eqn (4) indicating that glycine residues introduce a fall in $\Delta\delta$ at $+23 \, ^\circ\text{C}$ of 13% in THF, 31% in C2D5OD or 49% in CD3OD, confirming that the Gly residues introduce significantly more conformational flexibility than the Aib residues.

**Solvent-dependent signal decay moving away from the C terminus**

Identification of individual residues in an all-Aib oligomer is more challenging, because of the poor dispersion of the Aib methyl groups, which all lie within a <0.35 ppm range in the $^1\text{H}$ NMR spectrum. Dispersion in the $^{13}\text{C}$ NMR spectrum is better, and individual residues may be labelled by means of $^{13}\text{C}$-enriched methyl groups.44 However, difficulties of signal assignment remain, firstly because signals are likely to overlap when different parts of the oligomer exhibit a similar helical excess (in other words, when decay constants are low), and secondly because diastereotopic pairs of signals are harder to identify because they are not related to each other by coupling as they are in the $^1\text{H}$ NMR spectra of oligomers containing geminal CH2 groups. We have previously shown, however, that successive members of a series of $^{13}$C labels may themselves each be labelled by changing their $^{13}$C abundance through successive dilutions with $^{12}$C, allowing paired diastereotopic labels to be identified by relative integrals.44

We used this approach to explore the communication of conformational information through an Aib oligomer from the C terminus to the N terminus. The multiply labelled
compounds 7 were synthesized using the strategy illustrated in Fig. 5. Labelled and unlabelled Aib trimers 4** and 4 were built by standard methods from a starting monomer that was either 100% doubly (4**) or 1.1% singly (4, i.e. natural abundance) $^{13}$C labelled in its Aib methyl groups. The two samples of trimer 4** and 4 were then mixed in ratios of 50:50 or 25:75 and these two samples sequentially ligated to the methyl ester of 4** to make a nonamer 5, three of whose residues are labelled with different abundances of $^{13}$C, as shown in Fig. 5. 5 was extended by ligation with an unlabelled Aib dimer, trimer or tetramer to make the three oligomers 6a–c, which were each coupled at the C terminus to the powerful chiral controlling residue L-AlaNH$_3^+$-Bu$^+$ to provide 12-mer 7a, 13-mer 7b and 14-mer 7c.

These three ‘frame-shifted’ oligomers between them contain a labelled probe at every one of the nine positions in the chain from $i = 3$ to $i = 11$. Only 7a was soluble in pure methanol, but all were soluble in a 9:1 mixture of methanol and chloroform. The relevant portions of the three $^{13}$C spectra are shown in Fig. 7. The abundance-labelled labels are evident in the successively smaller anisochronicities of pairs of peaks with successively smaller integrals (100:50:25). Similar spectra in THF and in CDCl$_3$ showed significant peak overlap due to the much lower decay of anisochronicity in these.

![Fig. 5 Synthesis of oligomers 7a–c with residues identified by abundance-labelled $^{13}$C labels.](image)

![Fig. 6 Multi-labelled oligomers 7a–c allow measurement of screw sense at each of the nine positions in the oligomer from $i = 3$ to $i = 11$.](image)

![Fig. 7 Illustrative $^{13}$C NMR spectra of 7a (top), 7b (middle) and 7c (bottom) in 9:1 methanol:chloroform, with peaks arising from the 100%, 50% and 25% labels indicated by colours used in Fig. 6.](image)

![Fig. 8 Anisochronicity in the labelled Aib** residues of 7a–c in THF, ethanol (0 °C, 25 °C and 40 °C) and methanol.](image)
solvents, but individual peaks were still evident in THF. In CDCl$_3$, Lorentz–Gauss transformation assisted assignment of the 50% labelled signals, but the 25% labelled signals could not be located reliably.

Fig. 8 shows a plot of anisochronicity at each of the positions in the chain against the position (−$n$) of the label in the chain relative to the C-terminal alaninamide (residue i). Fitting an exponential decay (eqn (3), using positive values of n) to each plot gives signal decay constants $f$ (the per-residue decrease in $\Delta\theta$) of 6.1% in methanol, 5.4% in 9:1 methanol:chloroform, and 0.5% in THF. Earlier work had shown that aggregation effects may become important in solvents such as chloroform,\(^8\) but these were shown to be negligible in this case as the anisochronicity was independent of concentration over a 4-fold concentration range (4.6–18 μM). Changes in temperature induced a small variation in the value of $f$ for the oligomers in methanol, which was 5.5% at 0 °C; 6.1% at 23 °C and 6.5% at 40 °C. These values correspond to an enthalpy difference of about 3 kJ mol$^{-1}$ and an entropy difference of −13 J mol$^{-1}$ K$^{-1}$.

A further group of compounds 8a–d (Fig. 9), developed as part of a related project,\(^9\) had a better solubility profile than 7, and allowed comparisons of induced helical excess through four, eight or twelve achiral residues in methanol, chloroform and THF, as well as correlation with circular dichroism data.\(^1\)\(^H\) NMR chemical shift separations $\Delta\delta$ between $H_a$ and $H_b$, at the two equivalent methylene groups of the azepine rings in 8a–d compounds at +23 °C are shown plotted against −$n$ in Fig. 10, and an exponential decay (eqn (3)) fitted to these separations gave signal decay constants $f$ of 1.3% in THF and 7.7% in MeOH. In CDCl$_3$ the signals unfortunately showed no anisochronicity.

In the circular dichroism spectra of 8a–d the chromophore associated with the azepine biaryl ($\theta_{\text{max}}$ at 249 nm) is well separated from those associated with the carbonyl groups in the chain ($\theta_{\text{max}} < 220$ nm), and thus we may expect molar ellipticity at 249 nm to be proportional\(^13\) to the induced helical excess at the N terminus of the chain. Fig. 10b shows a plot of this value against chain length. From a modification of eqn (3)

$$\theta = \theta_i \times (1 - f)^n$$

we deduce signal decay constants $f$ of 2.5% in THF and 7.6% in MeOH, at +23 °C.

**Discussion**

There are numerous polymeric structures that are helical even though the individual monomers are achiral.\(^2,7,21\) Examples include not only oligomers of Aib,\(^7,7,29\) but also polyacetylenes,\(^8,8,84\) polyisocyanides,\(^8,8,86\) polyisocyanates,\(^8,8,88\) polyaromatic structures of various types,\(^80,7,83,89,91\) and even polyacetics.\(^8,9,90\) The conformations of such helices consist of domains of identical screw-sense which may span the whole length of the oligomer, or alternatively may be interrupted by ‘helical perversions’:\(^9,93\) locations where the screw sense reverses.\(^9,93\) The lengths of the resulting domains of screw-sense uniformity have in some cases been quantified, and in one well studied case, the polyisocyanates,
this length can be some 600 monomers at 20 °C.7 In other words, the average number of monomers separating one screw-sense reversal from the next is 600.

The decay of screw-sense preference on lengthening a helical domain interposed between a screw-sense controller and reporter may be viewed in terms of the chance of helix reversals occurring in that domain. A domain containing a single reversal, or indeed any odd number of reversals, will give rise to opposite screw-sense preferences at the two termini of the domain, while zero or an even number of reversals will lead to identical screw senses at the termini. Thus a helix having $P$ screw sense at one terminus will have $P$ screw sense at the other if it contains zero or an even number of reversals and $M$ screw sense if it contains an odd number of reversals. The likelihood of the two ends of the domain having the same screw sense may be calculated by considering the excess probability of an odd number over an even number of screw sense reversals, assuming an equal probability of screw-sense reversal at each location in the domain.

For a domain of $N$ elements, the difference in probability between having an even number of elements in state $x$ and an odd number in state $\bar{x}$ is

$$\left(2p - 1\right)^N$$

where the probability of state $x$ at each element is $(1 - p)$. Since helical excess $(he) = \left((P) - (M)\right)/\left((P) + (M)\right)$, interpreting state $x$ as a helix reversal allows the inference that $he$ at position $i$ in a helix built from achiral monomers is related to $he$ at position $i + n$ by this same expression

$$he_i = he_i \times \left(2p - 1\right)^n$$

where $p$ may be interpreted as the ‘fidelity’ with which a screw sense preference is transmitted from one monomer to the next ($p = 1$ implies complete communication of screw sense; $p = 0.5$ implies complete loss of screw sense control, and $p = 0$ implies complete inversion of screw sense).

Thus the empirical values for the signal decay constant $f$ derived from eqn (3)-(5) may be interpreted structurally as a measure of the probability of screw-sense reversal, with $f = 2 - 2(1 - p)$ in other words with $p$ as the screw-sense fidelity, $f/2$ may be interpreted as the probability of a helix reversal occurring, and $f$, the signal decay constant, as the fractional decrease in helical excess as the screw-sense preference passes from one monomer to the next. Values for decay constant $f$ and hence fidelity $p$ obtained from Fig. 2, 4, 8 and 10 are assembled into Table 1.

This interpretation of the decay of helical preference along a chain in terms of the likelihood of the two termini of a helical domain having the same or opposite screw sense makes it clear that the decay constant $f$ for any monomer must be independent of whether the ‘signal’ is passing from the N to the C (as in 2 and 3) or the C to the N (as in 7 and 8) terminus. The values of $p$ obtained for Aib in MeOH from 2 or 7 or 8 for a signal travelling from the C terminus to the N terminus ($p = 0.970, 0.962, 0.962$) are close to values previously calculated for a signal travelling from the N terminus to the C terminus (0.970; ref. 31, 34, 96), 0.965 (ref. 65)). It is also notable that the value of $p$ obtained for Gly from 3 corresponds closely to a previous observation7,67 that Gly conducts a helical preference 59% as efficiently as Aib (i.e. $(1 - f_{Aib})/(1 - f_{Gly}) = 0.59$; the corresponding value calculated from the data in Table 1 is 0.51/0.939 = 0.54).

The values in Table 1 clearly show that helical reversals are more likely at higher temperatures, and in more polar solvents. It may be deduced that whatever disruption of the hydrogen-bonding network is required to allow a helix reversal is more favourable in polar than non-polar solvents. The question of what structures these helix reversals adopt is still under investigation, but one important observation suggests that helix reversal does not require the formation of an unstructured random coil conformation. In such a conformation, oligomer

### Table 1: Values for signal decay constant $f$, screw-sense fidelity $p$, and helix persistence length $L$ for compounds in the paper in various solvents

<table>
<thead>
<tr>
<th>Compound series</th>
<th>Monomer</th>
<th>Solvent</th>
<th>$T^\circ C$</th>
<th>Per-residue decrease in hel $f/%$</th>
<th>Screw-sense fidelity $p$</th>
<th>Average length of uniform screw sense $L$</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>Aib/Ac6c</td>
<td>THF</td>
<td>0</td>
<td>0.8</td>
<td>0.996</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Aib/Ac6c</td>
<td>THF</td>
<td>23</td>
<td>0.1</td>
<td>0.9996</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>Aib</td>
<td>THF</td>
<td>23</td>
<td>0.5</td>
<td>0.9975</td>
<td>200</td>
</tr>
<tr>
<td>8</td>
<td>Aib</td>
<td>THF</td>
<td>23</td>
<td>1.3</td>
<td>0.993</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Aib</td>
<td>THF</td>
<td>23</td>
<td>2.5</td>
<td>0.988</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Aib/Ac6c</td>
<td>EtOH</td>
<td>23</td>
<td>1.9</td>
<td>0.991</td>
<td>50</td>
</tr>
<tr>
<td>2</td>
<td>Aib/Ac6c</td>
<td>EtOH</td>
<td>40</td>
<td>2.5</td>
<td>0.987</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>Aib</td>
<td>9:1 MeOH:CDCl$_3$</td>
<td>23</td>
<td>5.4</td>
<td>0.973</td>
<td>18</td>
</tr>
<tr>
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<td>5.5</td>
<td>0.972</td>
<td>18</td>
</tr>
<tr>
<td>7</td>
<td>Aib</td>
<td>MeOH</td>
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<td>6.1</td>
<td>0.970</td>
<td>16</td>
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<tr>
<td>8</td>
<td>Aib</td>
<td>MeOH</td>
<td>40</td>
<td>6.5</td>
<td>0.967</td>
<td>15</td>
</tr>
<tr>
<td>8</td>
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<td>MeOH</td>
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<td>7.7</td>
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<tr>
<td>3</td>
<td>Gly</td>
<td>THF</td>
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<td>13</td>
<td>0.935</td>
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<tr>
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<td>MeOH</td>
<td>23</td>
<td>49</td>
<td>0.76</td>
<td></td>
</tr>
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</table>

* Determined by NMR unless otherwise stated. * Determined by circular dichroism.
NH groups would be exposed to the deuterated solvent and would therefore undergo hydrogen–deuterium exchange.79,99 Yet both the observed rate at which the NH signals of the oligomer exchange with D from the deuterated solvent, and the rate at which protonated signals returned on dissolution of deuterated oligomer in protonated solvent, are extremely slow. H/D exchange in CD3OD occurs on a time scale of hours to days at ambient temperature, while helical inversion occurs on the millisecond time scale,79,146–49 suggesting that random coil conformations make a negligible contribution to the ensemble of structures adopted by Aib oligomers. Thus we propose that the helical reversals that lead to signal decay are well-defined, local structural features which entail only a small perturbation of the 310 helical structure, and retain a well-defined intramolecularly hydrogen-bonded network. We are currently exploring experimentally the possible detailed structures for these helix reversals. As suggested by Kubasik,80 migration of a well-structured helical reversal along a chain, sequentially breaking and reforming only one or maybe two hydrogen bonds at a time, could provide a mechanism for the remarkably rapid inversion of a 310 helix.

The values also allow for the first time an estimate to be made, using the one-dimensional Ising model,95,100 of the average length over which hypothetical extended Aib homopolymers might be expected to adopt a uniform screw sense. The Ising model, first used to describe ferromagnetic and antiferromagnetic arrays of atoms, assumes that the energetic cost of a state reversal (here a change in screw sense) in a linear array is determined solely by nearest neighbours. If the chance of a helix reversal at any one residue is \((1 - p)\), the Boltzmann relation gives

\[
(1 - p)p = e^{-2J/kT}
\]

(8)

where \(2J\) is the energetic penalty for introducing a helix reversal.

According to the one-dimensional Ising model, the helical excess will decay exponentially over \(n\) segments as the \(n\)th power of \(\tanh(J/kT)\). This factor is equivalent to our empirical decay factor \((1 - f)\) (eqn (3)), giving as expected \(f = 2 \times (1 - p)\). The persistence length \(L\) (the average distance between reversals) may be obtained from the Ising model by95

\[
L = -1/\ln[\tanh(J/kT)] = -1/\ln(2p - 1) = -1/\ln(1 - f)
\]

(9)

Values for \(L\) are shown in Table 1 for selected data. A sufficiently soluble Aib polymer, in a non-polar solvent, can evidently be expected to maintain a preferred screw sense over distances measurable in hundreds of monomers.

One advantage of the Ising model is that it allows a simple estimate, from eqn (8), of the energetic cost \(2J\) of a helix reversal. For example, in the case of 2a–d in THF at \(-50^\circ\text{C}\) [Fig. 2a], \(J = 10\ \text{kJ mol}^{-1}\); for 7a–c in methanol at \(+40^\circ\text{C}\) it is \(9\ \text{kJ mol}^{-1}\), both considerably less than the typical strength of a hydrogen bond. This is consistent with the observation noted earlier that hydrogen–deuterium exchange is very slow in such systems, providing strong evidence that helix inversion simply involves a rearrangement of intramolecular hydrogen bonds rather than any reduction in their number. The Ising model assumes that the driving force for the retention of helicity is purely energetic; as the analysis of the temperature dependence of \(f\) for 7a–c in methanol in the preceding section shows, a substantial part of the driving force is in fact entropic.

**Conclusion**

A screw-sense preference induced by a chiral residue at the terminus of an achiral helical oligomer may be considered as a signal whose strength decays as with decreasing distance from the chiral inducer. The spatial decay rate may be quantified by means of NMR or CD probes of helical excess located at successive positions in an oligomeric structure, or located at the terminus of oligomers of varying lengths. The decay rate depends on solvent and temperature (signal decay is faster at higher temperature and in more polar solvents) and on the nature of the achiral oligomers in the chain (greater flexibility gives rise to faster signal decay). Typical values are of the order of 5% for Aib in MeOH, and 1% for Aib in THF, leading to helix persistence lengths in non-polar solvents of around 200 monomers. The decay rate may be interpreted as the result of a helix reversal occurring randomly at any position in the oligomer. Helix reversals appear to be localized, well-ordered structures, and ongoing work seeks to establish their detailed geometry.

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**Notes and references**

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A previous study indicated that the Aib and Ac6c have very similar abilities to communicate a screw-sense preference led to a rapid stereoselectivity of the reactions in Fig. 1 is due to intramolecular conformational effects. We define helical excess in a manner analogous to enantiomeric excess, but with regard to helix screw sense: he is the difference in population of P and M conformers as a percentage of the total. Sugino has employed the term ‘screw sense excess’ (se), generally quantified by CD, for the same quantity: see ref. 55 and 56. Several control experiments established unequivocally that the stereoselectivity of the reactions in Fig. 1 is due to intramolecular conformational effects. We define helical excess in a manner analogous to enantiomeric excess, but with regard to helix screw sense: he is the difference in population of P and M conformers as a percentage of the total. Sugino has employed the term ‘screw sense excess’ (se), generally quantified by CD, for the same quantity: see ref. 55 and 56.
68 The anisochronicity \( \Delta \alpha \) is given by \( \rho_0 \Delta \alpha = \left[ ( f_1 - f_3 )^2 - J_{AB} \right]^{1/2} \left[ ( f_2 - f_4 )^2 - J_{AB} \right]^{1/2} \left[ ( f_1 - f_4 )^2 - ( f_2 - f_3 )^2 \right]^{1/2} \) where \( f_{1,2,3,4} \) are the observed resonance frequencies in order of the four lines comprising the AB multiplet, \( J_{AB} \) is the coupling constant, and \( \rho_0 \) is the spectrometer frequency.
70 V. Diemer, S. J. Webb and J. Clayden, manuscript in preparation.
96 This value was recalculated to higher precision using the data reported in ref. 34. This more precise value was the one used to extrapolate values for control of helical excess in ref. 31.
97 Ref. 17 uses the term ‘helix conductivity’, which may be interpreted as the value \( 1 – f \).
98 M. A. Kubasik, A. Blom and E. Daly, Biophys. J., 2005, 88, 558A.
99 M. A. Kubasik, E. Daly and A. Blom, Biopolymers, 2005, 80, 585.
101 It may be noted that the model will also accommodate the case where the more energetically favourable arrangement is one with alternating helix sense, corresponding to an antiferromagnetic array, in which case \( L = -1 \ln(1 - 2p) \).