Biosynthesis of fosfazinomycin is a convergent process†

Zedu Huang,abc Kwo-Kwang A. Wang,abc Jaeheon Leea and Wilfred A. van der Donk*abc

Fosfazinomycin A is a phosphonate natural product in which the C-terminal carboxylate of a Val–Arg dipeptide is connected to methyl 2-hydroxy-2-phosphono-acetate (Me-HPnA) via a unique hydrazide linkage. We report here that Me-HPnA is generated from phosphonoacetaldehyde (PnAA) in three biosynthetic steps through the combined action of an O-methyltransferase (FzmB) and an α-ketoglutarate (α-KG) dependent non-heme iron dioxygenase (FzmG). Unexpectedly, the latter enzyme is involved in two different steps, oxidation of the PnAA to phosphonoacetic acid as well as hydroxylation of methyl 2-phosphonoacetate. The N-methyltransferase (FzmH) was able to methylate Arg-NHNH₂ (3) to give Arg-NHNHMe (4), constituting the second segment of the fosfazinomycin molecule. Methylation of other putative intermediates such as desmethyl fosfazinomycin B was not observed. Collectively, our current data support a convergent biosynthetic pathway to fosfazinomycin.

Introduction

Phosphonate natural products are a class of compounds featuring a C–P bond.1 Many of these natural products show interesting and potent bioactivity. For instance, fosfomycin is an antibiotic in clinical use, while phosphonothricin tripeptide (PTT) is a widely used herbicide (Fig. 1).1 The bioactivities of phosphonates are generally attributed to their C–P bonds, which mimic, but are much more stable than, the corresponding O–P bond of phosphate esters.1 In recent years phosphonate natural products have drawn attention not only because of their bioactivity, but also because of the plethora of unusual and interesting chemistries associated with the biosynthesis of these molecules.2,3

Originally isolated from Streptomyces lavendofolius 630, fosfazinomycins (Fig. 1) are natural phosphonates with antifungal activity.4,5 Compared to fosfazinomycin B (fosB), fosfazinomycin A (fosA) contains an extra Val residue at the N-terminus. The most interesting structural feature of fosfazinomycins is the unusual hydrazide linkage between the carboxylic acid of Arg and the phosphonic acid. Although the structures of these antibiotics were elucidated three decades ago, information on their biosynthesis was unavailable until our very recent description of the biosynthetic gene cluster.6

Very little is known about how nitrogen–nitrogen bonds are fashioned in nature. In principle, three different pathways can
be envisioned in the case of fosfazinomycin (Fig. 2). The
hydrazide moiety could be initially attached to the phosphonate
and then coupled to the Arg or the Val–Arg dipeptide (retro-
synthetic disconnection a, Fig. 2). Alternatively, the hydrazide
could be formed on the Arg or the Val–Arg dipeptide and then
be condensed with the phosphonic acid (disconnection b), or
the hydrazide could be formed from an amide and phospho-
namide (disconnection c). Herein we report the first in vitro
biochemical investigation of fosfazinomycin biosynthesis.

Our studies assign the function of the enzymes involved in
biosynthetic studies of fosfazinomycin. We have shown,
 challengingly, that fosfazinomycins are synthesized through a convergent pathway that is consistent
with disconnection b (Fig. 2).

Results

The fzmB and fzmG genes were amplified by the polymerase chain reaction (PCR) from the fosfomycin gene cluster.

The amplified genes were inserted into the expression vector pET-15b. Both proteins were expressed in Escherichia coli as N-terminal hexahistidine-tagged constructs and purified by immobilized metal affinity chromatography (IMAC) to near homogeneity (>90% purity) according to SDS/PAGE analysis
(Fig. S1 in the ESI†).

With the soluble proteins in hand, we first tested the activity of His6-FzmB with two putative substrates, phosphonoacetate
(PnA) and rac-2-hydroxy-2-phosphono-acetate (rac-HpNA)
(Fig. 1). When PnA was used as substrate in the presence of S-
adenosyl methionine (SAM), ca. 90% conversion to the methyl ester was achieved according to analysis by 31P NMR
spectroscopy. The product was confirmed to be Me-PnA by spiking with an authentic synthetic standard (Fig. S2†). In the absence of either the enzyme or SAM, only the starting material was recovered. In addition, complete conversion of rac-HpNA to Me-HpNA was observed in the presence of His6-FzmB and SAM
(Fig. S3†). Therefore His6-FzmB was able to methylate both enantiomers of HpNA. The high conversions in both reactions
made it difficult to conclude whether PnA or HpNA would be the physiological substrate of FzmB. To further study its substrate
tolerance, methylenbis(phosphonic acid) (Fig. 1) and malonic
acid were tested as close analogs of PnA, but product was not detected in either reaction.

Next we investigated the function of the putative α-KG
dependent dioxygenase FzmG. Initially Me-PnA was tested,
resulting in complete conversion of Me-PnA to Me-HpNA
(Fig. 3A and B). The reaction was enzyme-catalysed and α-KG
was essential for catalysis (Fig. 3C and E). In addition, the
common reductant l-ascorbate accelerated the reaction (Fig. 3A
and D). Recently Hammerschmidt and co-workers have deter-
mined the absolute configuration of Me-HpNA in fosfazinomy-
cins to be S by comparison of chemically synthesized standards
to the fragments generated from acid-mediated hydrolysis of fosfazinomycins. In addition, they found that Me-HpNA was configurational stable in water at slightly acidic pH but completely racemized at pH 7–8. Given its ease to racemize at the pH we used for the enzymatic assays, we did not attempt to determine the stereochemistry of our enzymatic product. PnA
was also incubated with His6–FzmG. Under the same conditions
that resulted in complete consumption of Me-PnA, the conver-
sion of PnA was only 30%; the product was confirmed to be HpNA by spiking with authentic synthetic standard (Fig. S4†).

![Scheme 1](image1)

Scheme 1 Previously proposed initial steps of the biosynthetic pathway of fosfazinomycin A.
DhpA is another α-KG dependent non-heme iron dioxygenase that hydroxylates 2-hydroxyethyl-phosphonate (2-HEP) to generate 1,2-dihydroxyethylphosphonate (1,2-DHEP) (Fig. 1) during the biosynthesis of dehydrophos. A typical 31P NMR spectrum of the reaction mixture spiked with authentic standards for assignments. Monitoring the O2 consumption rate using a Hansatech O2 electrode, the apparent steady-state kinetic parameters for several substrates were determined (Table 1). Clearly, Me-PnA is a much better substrate than PnA, suggesting that methylation occurs before hydroxylation. Furthermore, PnAA is also a reasonably good substrate (Fig. S7†), consistent with FzmG catalysing two steps in the biosynthesis of fosfazinomycin.

To further test this hypothesis, we determined the kinetic parameters of His6–FzmG towards the substrates. By measuring the O2 consumption rate using a Hansatech O2 electrode, the apparent steady-state kinetic parameters for several substrates were determined (Table 1). Clearly, Me-PnA is a much better substrate than PnA, suggesting that methylation occurs before hydroxylation. Furthermore, PnAA is also a reasonably good substrate (Fig. S7†), consistent with FzmG catalysing two steps in the biosynthesis of fosfazinomycin.

Table 1. Apparent steady-state kinetic parameters of His6–FzmG towards various substrates

<table>
<thead>
<tr>
<th>Substrate</th>
<th>kcat [s⁻¹]</th>
<th>Km [mM]</th>
<th>kcat/Km [M⁻¹ s⁻¹]</th>
</tr>
</thead>
<tbody>
<tr>
<td>Me-PnA</td>
<td>0.27 ± 0.01</td>
<td>0.08 ± 0.02</td>
<td>3 x 10⁴</td>
</tr>
<tr>
<td>PnA</td>
<td>0.17 ± 0.01</td>
<td>8 ± 1</td>
<td>2 x 10⁴</td>
</tr>
<tr>
<td>2-HEP</td>
<td>0.35 ± 0.02</td>
<td>0.21 ± 0.04</td>
<td>1.7 x 10³</td>
</tr>
<tr>
<td>PnAA</td>
<td>0.46 ± 0.08</td>
<td>0.8 ± 0.3</td>
<td>6 x 10²</td>
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</table>

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new methyl peak was observed in the $^1$H NMR spectrum when Arg-NHNH$_2$ (3) was incubated with His$_6$-FzmH in the presence of SAM. The product was confirmed to be Arg-NHNHMe (4) by spiking with a synthetic authentic standard (Fig. 5B and C), and this assignment was also supported by high resolution Fourier transform MS (Fig. S14†). In the absence of His$_6$-FzmH, no methylation occurred (Fig. 5A). These results are consistent with a convergent pathway (route b) and not with a linear pathway and provide important insights into the potential identity of substrates for N-N bond formation (e.g., this process does not occur on phosphonate substrates). To further study the substrate specificity of FzmH, Ac-Arg-NHNH$_2$ (5) and Val-Arg-NHNH$_2$ (6) (Scheme 2) were synthesized. The enzyme methylated both of these compounds on the terminal nitrogen and the product identity was again confirmed by spiking with synthetic authentic standards (Fig. S15 and S16†) and FT-MS (Fig. S17 and S18†).

**Discussion**

The O-methyltransferase His$_6$-FzmB methylated both PnA and HPnA with high efficiency in vitro, preventing assignment of the physiological substrate. On the other hand, His$_6$-FzmG hydroxylated Me-PnA and PnA with very different catalytic efficiencies. The $K_m$ of Me-PnA was 0.08 ± 0.02 mM, 100-fold lower than that of PnA (8 ± 1 mM). Combined with the similar $k_{cat}$ values (0.27 ± 0.01 s$^{-1}$ versus 0.17 ± 0.01 s$^{-1}$), these results strongly suggest Me-PnA is the physiological substrate of FzmG. Therefore, methylation of PnA by FzmB likely occurs before hydroxylation. This order of methylation and hydroxylation also matches well with the fact that Me-PnA is one of the observed metabolites produced by fosfazinomycin producing strains.

PnAA is a common intermediate in phosphonate biosynthesis, and it is processed in three different ways in various characterized pathways. PnAA can be converted to 2-aminoethylphosphonate (2-AEP) by a transaminase or reduced to 2-HEP by an alcohol dehydrogenase. Finally it can also condense with oxaloacetate to give 2-keto-4-hydroxy-5-phosphonopentanoic acid in rhizoctin biosynthesis. To our knowledge, the fosfazinomycin pathway is the first example in which PnAA is transformed to PnA in a phosphonate biosynthetic pathway. We note that the same reaction catalysed by an aldehyde dehydrogenase PhnY was reported in phosphonate catabolic pathways. Our current findings that FzmG was able to generate PnA from PnAA with only 6-fold lower catalytic efficiency compared to its oxidation of Me-PnA ($k_{cat}/K_m$ for Me-PnA = 5 × 10$^3$ M$^{-1}$ s$^{-1}$ versus $k_{cat}/K_m$ for PnA = 3 × 10$^2$ M$^{-1}$ s$^{-1}$) suggests that FzmG possesses a relaxed substrate specificity. Hence, it is possibly the missing enzyme in our previously proposed fosfazinomycin biosynthetic pathway (Scheme 1). However, we cannot rule out the possibility that a housekeeping aldehyde dehydrogenase outside the gene cluster might oxidize PnAA to PnA. There are other examples of α-KG dependent non-heme iron dioxygenases catalysing multiple reactions in a single biosynthetic pathway. One such example is the clavamin synthase, which catalyses three separate oxidative steps during clavulanic acid biosynthesis, involving hydroxylation, cyclization and desaturation reactions.

Although most secondary metabolites are biosynthesized in a linear fashion, some small molecules, mostly polyketides and non-ribosomal peptides, are generated via a convergent manner. For example, the chromophore of maduropeptin, an endoeine antitumor compound, was installed from four components using a convergent strategy. In contrast, until this study, all known phosphonates with characterized biosynthetic
pathways were shown to be generated in a linear pathway. Our observation that FzmH can convert Arg-NH₂ and related compounds 5 and 6 to their N-methylated products, whereas other potential substrates were not accepted, strongly supports methylation of an acylhydrazine during the biosynthesis of fosfizinomycins. In turn, these results indicate that fosfizinomycin B is biosynthesized from two advanced intermediates, Arg-NH₂Me₃ (4) and (S)-Me-HpNA following a convergent manner. The enzymes involved in the formation of 3 and in the condensation reaction remain to be identified among the proteins encoded in the cluster whose functions have not yet been assigned. Such studies can now commence with the knowledge gained about potential substrates in this study.

Methods

General methods

For reagents, molecular biology procedures, enzyme purifications, synthetic procedures for the preparation of substrates and standards, and their spectroscopic characterization, see the ESL.†

FzmB activity assays

The reaction mixture (500 μL) contained 5 mM of the appropriate phosphonic acid, 6 mM S-adenosyl methionine (SAM), 0.9 mg mL⁻¹ AdoHcy nucleosidase, and 74 μM FzmB in 50 mM sodium phosphate (NaPi) buffer, pH 7.7. After 5.5 h incubation at ambient temperature, the proteins were removed by passing the reaction mixture through an Amicon spin column of 10 kDa molecular weight cut-off (MWCO), and 150 μL D₂O was added into the sample before NMR analysis.

FzmG activity assays

The reaction mixture (500 μL) contained 5 mM of the appropriate phosphonic acid, 10 mM α-ketoglutarate, 1 mM l-ascorbic acid, 0.2 mM (NH₄)₂Fe(SO₄)₃, and 37 μM FzmG in 50 mM NaPi, pH 7.7. Typically the enzyme had been reconstituted in an anaerobic glove box (Coy, Grass Lake, MI) with 1.2 equivalent of NaPi, pH 7.7. After 7.5 h incubation at ambient temperature, the proteins were removed by passing the reaction mixture through an Amicon spin column of 10 kDa MWCO, and 150 μL D₂O was added into the sample before NMR analysis.

Kinetic assays with FzmG

A Hansatech O₂ electrode was used to generate the Michaelis–Menten curves by varying the concentration of the appropriate phosphonic acid while keeping α-KG at a saturating concentration (1 mM) and at a constant FzmG concentration (3.7 μM) for Me-PnA, PnAA and 2-HEP assays, and 9.25 μM for PnA assay). All reactions were initiated by the addition of FzmG, and the initial rate of O₂ consumption was measured in triplicate. Nonlinear regressions were calculated using Igor Pro version 6.1.

FzmH activity assays

The reaction mixture (500 μL) contained 2.5 mM of the appropriate substrate, 3.5 mM S-adenosyl methionine (SAM), 0.9 mg mL⁻¹ AdoHcy nucleosidase, and 26 μM FzmH in 50 mM NaPi, pH 7.7. After 6.5 h incubation at ambient temperature, the proteins were removed by passing the reaction mixture through an Amicon spin column (10 kDa MWCO), and 150 μL D₂O was added into the sample before NMR analysis.

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Notes and references


