Effective magnetic moment in cyclodextrin–polynitroxides: potential supramolecular vectors for magnetic resonance imaging†

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Nitroxides have great potential as contrast agents for Magnetic Resonance Imaging (MRI). Two β-cyclodextrin (βCD) derivatives bearing one or seven (2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO) units on the small rim of βCD (CD3 and CD6 respectively) were synthesized. Their effective magnetic moments were measured by DC-SQUID magnetometry obtaining the values \( \mu_{\text{eff}}/\mu_B \approx 1.7 \) and \( \mu_{\text{eff}}/\mu_B \approx 4.2 \) for CD3 and CD6 respectively. Interestingly, while isothermal magnetization data of CD3 were well described by a Brillouin function for a \( S = 1/2 \) single spin system, those associated with CD6 could not be explained in the framework of a non-interacting spins model. For this reason, four different configurations for the seven interacting nitroxides were considered and modeled. The numerical results evidenced that only the configurations with a privileged central spin could take into account the experimental observations, thus justifying the reduced effective magnetic moment of CD6. The water relaxivity \( r_1 \) in DMSO-d_6–water (9 : 1 v : v) solutions was also measured for both the derivatives obtaining the values \( r_1 = 0.323 \, \text{mM}^{-1} \, \text{s}^{-1} \) and \( r_1 = 1.596 \, \text{mM}^{-1} \, \text{s}^{-1} \) for CD3 and CD6 respectively.

1. Introduction

Persistent nitroxyl radicals (nitroxides) have found increasing applications in recent years in different important fields such as catalysts for oxidative processes, for the development of materials for energy storage, in molecular magnetism and spintronics. Moreover, nitroxides have a promising future in the biomedical field. Indeed, they have been used for the synthesis of bio-compatible materials capable of protecting biological tissues from the damage caused by reactive oxygen species (ROS). Due to their magnet-sensitive character, good cell membrane permeability and reduced toxicity, nitroxides are suitable spin-labels for in vivo Electron Spin Resonance (ESR) and Dynamic Nuclear Polarization (DNP) biological studies. A very remarkable property of the nitroxides is their capability of decreasing the relaxation times (longitudinal, \( T_1 \), and transversal, \( T_2 \)) of water in biological tissues. For this reason they are currently investigated as MRI contrast agents (CA), suggesting their use as a potential alternative to the clinically used, but more toxic, gadolinium(III) complexes. In general, the ability of a CA to enhance the proton relaxation rate \( (1/T_1, 2, \, \text{s}^{-1}) \) is defined in terms of its relaxivity \( r_{1,2,} \, \text{mM}^{-1} \, \text{s}^{-1} \) according to the eqn (1) where [CA] (mM) is the concentration of the CA and \( 1/T_{1,2,} \) is the relaxation rate in absence of CA:

\[
1/T_{1,2} = 1/T_{1,2}^{0} + r_{1,2}[\text{CA}].
\]  

Without going into the details of the Solomon–Bloembergen–Morgan theory of the paramagnetic relaxation, the relaxivity of a CA is proportional to the square of its effective magnetic moment, \( \mu_{\text{eff}} \):

\[
r_{1,2} = k\mu_{\text{eff}}^2
\]  

However, several factors, mainly the solvent and the molecular structure of the CA, can significantly affect the proportionality constant \( (k) \), and thus the relaxivity value. In order to be appealing in terms of imaging resolution, newly designed CA should be competitive with the present state-of-the-art
represented by Gd(III) complexes. With its seven unpaired electrons, a Gd(III) ion has an effective magnetic moment of 7.94 μB (where μB is the Bohr magneton). Therefore the synthesis of new derivatives with an effective magnetic moment comparable or even higher than Gd(III) ion but less harmful to the human body is of high applicative interest. With regard to this aspect, the adoption of poly-nitroxides could be a valuable approach and significant contributions in this direction have been given by different investigators in the last years.9

Among the different compounds suitable for the creation of molecular architectures bearing poly-nitroxides, cyclodextrins (CDs) could represent a valid choice. Indeed, CDs are macrocyclic oligosaccharides composed by 6, 7, or 8 glucosidic units (α-, β- and γ-CD, respectively). These molecules are characterized by a truncated cone-shaped three-dimensional structure with an empty inner cavity having hydrophobic character. The hydroxyl groups of the anhydroglucose units are located both on the large rim (OH in position 2 and 3) and the small rim of the truncated cone (OH in position 6) (Fig. 1). Depending on their specific position, CDs’ hydroxyl groups have different reactivity and a large number of methods for their selective chemical functionalization have been developed.10 For this reason, CDs can be fruitfully applied for the synthesis of materials bearing multi-radical moieties.11 Moreover, CDs have found relevant applications in the biomedical field as drug delivery systems due to their capability to form “host-guest” inclusion complexes both in solution and in the solid state with small molecules or portions of larger molecules.12 Hence, in order to verify how CDs bearing nitroxyl radicals could be potentially attractive for possible application as CA for MRI, a detailed characterization of their magnetic properties is mandatory.

For this very reason, in this paper we present a detailed study on the magnetic properties of spin labelled CDs obtained by introducing TEMPO radical moieties onto βCD small rim via a click-chemistry approach (see Fig. 1). Both the mono-TEMPO (CD3) and the hepta-TEMPO (CD6) derivative have been synthesized and characterized. Their paramagnetic properties have been investigated by ESR spectroscopy and DC-SQUID magnetometry. The experimental results have been analysed and discussed on the basis of a standard quantum-statistical model. The longitudinal relaxivity of water, in presence of both CD3 and CD6, and using DMSO as solvent has been measured and discussed as well.

2. Experimental

2.1. General

β-Cyclodextrin (βCD) and other reagents and solvents were commercially available and used as received unless otherwise stated. The synthesis of 6A-O-p-toluenesulfonyl-βCD (CD1) was performed following a published procedure with minor modifications as detailed in the ESI.13 The corresponding azide derivative (CD2) was obtained reacting CD1 with an excess of NaN₃ in DMSO at 90 °C overnight (see ESI for details). Heptakis-6-iodo-6-deoxy-βCD (CD4) and the corresponding azide (CD5) were synthesized as reported in literature.14 Propargyl-TEMPO was obtained following the paper of Bogdan and McQuade.22

2.2. Synthesis of CD3 and CD6

CD3 was prepared by dissolving 1.160 g (1 mmol) of CD2 in 5 mL of DMF containing 48 mg (0.25 mmol) of CuI and few drops of Et₃N in a two-necked flask (10 mL). The mixture was stirred under nitrogen atmosphere for 15 min at 50 °C. Then, 315 mg of propargyl-TEMPO (1.5 mmol) were added. After 24 h under the same conditions, the liquid mixture was concentrated under reduced pressure (1-1.5 mL final volume) and transferred into a chromatography column containing a silica-gel bed packed using DMF with a 30% ammonia aqueous solution (DMF : NH₄OH = 100 : 1, v : v) in order to remove the copper salts. The eluted CD3-containing DMF solution was finally concentrated under reduced pressure (10 mbar – thermal bath at 30 °C) until almost complete evaporation of the DMF. The addition of acetone (100 mL) allowed the precipitation of the product that was recovered by filtration on paper and washed with acetone (4 × 50 mL). An orange colored powder, soluble in DMSO and DMF, scarcely soluble in water, and insoluble in acetone, was obtained in quantitative yield. The TLC analysis (dissolving the product in DMF) was performed using silica-gel 60 F254 aluminium sheets (Merck) and 2-propanol : water : EtOAc : NH₄OH (5 : 3 : 1 : 0.5) solution as eluent (Rf 0.56).

The reduced form of CD3 (CD3-H) was obtained dissolving CD3 (140 mg, 0.1 mmol) in 1 mL of DMSO and then adding an excess of phenyldiazene. The mixture was stirred at r.t. for 4 h. The solution was finally poured in acetone. The white precipitate was filtered on paper and extensively washed with acetone. 

1H-NMR (DMSO-d₆ + D₂O, 500 MHz): δ = 7.98 (s, 1H, triazole ring), 5.10–4.79 (m, 7H, H-1), 4.77 (m, 1H, H-6′a), 4.66–4.56 (m, 1H, H-6′b), 4.49 (s, 2H, CH₂), 3.90 (t, 1H, H-5′), 1.89 (m, 2H, CH-εq), 1.27 (m, 2H, CH-ax), 1.08 (s, 6H, CH₃-εq), 1.04 (s, 6H, CH₃-ax).

The product CD6 was obtained similarly to CD3 (50 °C under N₂ for 24 h) starting from 350 mg (0.27 mmol) of CD5 and 526 mg (2.50 mmol) of propargyl-TEMPO in DMF (2 mL), in presence of Cu(30 mg, 0.16 mmol) and Et₃N (3 drops). Purification was performed by column chromatography using silica-gel, first eluting with Hex : EtOAc (8 : 2) in order to remove the...
excess of propargyl-TEMPO and then with methanol for the recovery of the product. The evaporation of the solvent provided in quantitative yield an orange colored powder, soluble in CHCl₃, methanol, acetonitrile. TLC (silica-gel 60 F254, CHCl₃ : MeOH 5 : 3) revealed one spot with Rᵣ 0.70. ¹H-NMR (DMSO-d₆ + phenylhydrazine + D₂O, 500 MHz): δ = 7.86 (s, 1H, triazole ring), 5.08 (s, 7H, H-1), 1.78 (m, 14H, CH-eq), 1.18 (m, 14H, CH-ax), 1.04 (s, 42H, CH₃-eq), 1.00 (s, 42H, CH₃-ax).

2.3. Characterization

¹H, ¹³C NMR, and ¹H-¹H TOCSY spectra of the products were recorded at 305 K with a Bruker Avance-500 MHz NMR spectrometer. The FT-IR solid phase spectra of the powdered sample with infrared grade KBr were recorded using a Varian 640-IR spectrometer. ESI-MS mass spectra were collected on a Bruker Esquire 3000+ with electrospray ionization source and ion-trap detector. The samples were analysed by direct infusion of suitable solutions in the spectrometer source. The peak assignment was supported by the simulation of the isotopic cluster of the corresponding molecular formula (see ESIf). EPR spectra were recorded at room temperature using an ELEXYS E500 spectrometer equipped with a NMR gaussmeter for the calibration of the magnetic field and a frequency counter for the determination of g-factors. Nitroxide concentrations were measured with respect to a solution of 2,2-diphenyl-1-picrylhydrazyl (DPPH) of known concentration. Concentrations were measured with respect to the sodium cationized residual [BCD] and the CD3 analogous with two nitroxide units attached. The NMR characterization was performed considering CD3-H obtained after reducing with phenylhydrazine the nitroxide radical of CD3 to the corresponding hydroxylamine. The ¹H-NMR spectrum of CD3-H is reported in Fig. 2. The peaks assignment has been obtained from the corresponding ¹H-¹H TOCSY NMR spectrum reported in Fig. S16.†

In order to obtain CD6 we first synthesized the hepta-iodo derivative CD4 following a classic procedure. The substitution of the seven iodine atoms with azide afforded quantitatively the product CD5. Finally, propargyl-TEMPO units were attached to the derivative CD5 providing the product CD6 in quantitative yield. Also in this case, FT-IR analysis (Fig. S13†) confirmed the complete disappearance of the azide peak. The ESI-MS analysis in positive mode and using methanol as solvent showed the presence of a small peak with m/z 2084.1 corresponding to [CD6 + Na]⁺, a very intense peak at m/z 1414.0 attributed to [CD6 + 2Na]²⁻ and close to a smaller peak associated to [CD6 + H + Na]³⁺ at m/z 1403.5. A small cluster at m/z 950.8 was also observed and attributed to [CD6 + 3Na]³⁺ (from Fig. S18–12†). The NMR characterization of CD6 was obtained by reducing the nitroxyl radicals with phenylhydrazine directly in the NMR tube and using DMSO-d₆ as solvent (see Fig. S17†). The resolved spectrum showed the presence of a broad singlet at 5.08 ppm corresponding to the seven anemic protons, and a multiplet in the region 6.0–5.8 ppm, assigned to the fourteen OH protons at the C2 and C3 positions of the anhydroglucose.
3.2. ESR spectroscopy

Fig. 3a shows the ESR spectrum of CD3 (0.05 mM) recorded in water at room temperature. It is a three lines pattern spectrum similar to the one described by Lucarini et al. for an analogous derivative obtained from α-CD.\textsuperscript{14} ESR spectrum of CD6 (Fig. 3b) was recorded in acetonitrile at room temperature. It shows mainly one broad signal (\( g = 2.0058 \)) whose integrated intensity corresponds quantitatively to the theoretical concentration of nitroxide units. The broadening (peak to peak line width 11.5 G) of the signal is independent of concentration, and thus inter-assembly interactions can be discounted. This spectrum is reminiscent of those obtained from very concentrated nitroxide solutions. Since it was obtained at 0.5 mM concentration, the signal broadening is ascribed to the proximity of spin centers of nitroxide units. The broadening (peak to peak line width 11.5 G) corresponds quantitatively to the theoretical concentration of CD6 within the framework of the cyclodextrin rim. This is consistent with exchange-coupled nitroxides with a coupling constant significantly larger than the \( ^{14} \text{N} \) hyperfine coupling, \( |J|/g_B \mu_B \approx 2.0 \text{ mK} \). The lack of resolved ESR multiplets should be attributed to the presence of many different transitions from different spin states arising from multiple interactions between the seven radical units. The spectrum shows also a very weak sharp satellite signals presumably due to some non-exchanging nitroxide units. However, theoretical simulation of the experimental spectrum indicated that these correspond to less than 0.5% of the total amount of nitroxide.\textsuperscript{15}

3.3. Magnetic behavior of TEMPO-CD

The temperature dependence of the molar magnetic susceptibility for the CD3 (a) and CD6 (b) samples both measured under the form of grinded powders are shown in Fig. 4. Both the curves show a dominant paramagnetic behavior. In Fig. 5 the isothermal magnetization as a function of \( H/T \) at several temperatures from 2 up to 30 K is reported for CD3 (a) and CD6 (b), respectively. It is worth noticing that in the case of CD3 all the experimental points collapse on the same line as expected for a system of non-interacting magnetic moments. Instead, this is not the case for CD6 whose behavior will be explained by taking into account antiferromagnetic (AFM) intra-molecular interactions (\textit{vide infra}). The temperature dependent magnetic susceptibility was analyzed in the framework of a modified Curie–Weiss model (eqn (3)) where \( C \) is the Curie constant, \( \theta \) the Curie–Weiss temperature and \( \chi_0 \) is a temperature independent contribution.

\[
\chi = \frac{C}{T - \theta} + \chi_0 \tag{3}
\]

The obtained parameters for CD3 and CD6 are reported in Table 1. First, the \( T \)-independent term \( \chi_0 \) is zero in the case of CD6 while the positive value obtained for CD3 was ascribed to an almost negligible ferromagnetic contribution. Since this spurious effect was indeed almost fully removed in a reference CD3 sample precipitated from hot water, we ascribed it to a change of the molecular packing (data not shown).

A more relevant feature that emerges in both the samples is the existence of small and negative \( \theta \) which indicates the presence of very weak AFM-like interactions. It is worth to notice that in case of CD3 these interactions have inter-molecular origin while in CD6 they are mainly due to intra-molecular exchange interactions as we will detail here below.

The 2 K CD3 isothermal magnetization data reported in Fig. 6 was well fitted by a Brillouin function leaving \( S \) as free parameter.\textsuperscript{16} We obtained \( S = 0.452 \pm 0.001 \) which confirms...
that CD3 mono radical fully behaves as an ensemble of non-interacting spin $S = 1/2$. Indeed, the very small difference from the expected value $S = 1/2$ can be ascribed to the presence of tiny AFM inter-molecular interactions ($\theta < 0$ as reported in Table 1). The same figure also reports the experimental magnetization data of CD6 at 2 K and the theoretical calculation for the case of a seven-non-interacting-spin-1/2 model. It is interesting to note that the rate $TdM/dH|_{H=0}$ is remarkably lower than the one expected for the non-interacting case. We ascribe this effect to the existence of non-negligible intra-molecular spin-interactions as we will detail hereafter. Firstly, we can safely exclude the presence of relevant inter-molecular interactions since the same result was obtained for a DMSO diluted sample (not shown).

### Table 1

<table>
<thead>
<tr>
<th>Sample</th>
<th>$C$ [10$^{-6}$ m$^3$ K mol$^{-1}$]</th>
<th>$\theta$ [K]</th>
<th>$\chi_0$ [10$^{-6}$ m$^3$ mol$^{-1}$]</th>
<th>$\mu_{\text{eff}}/\mu_B$</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD3</td>
<td>4.471(1)</td>
<td>-0.240(4)</td>
<td>0.030(1)</td>
<td>1.686(1)</td>
</tr>
<tr>
<td>CD6</td>
<td>27.646(1)</td>
<td>-0.713(7)</td>
<td>0</td>
<td>4.193(1)</td>
</tr>
</tbody>
</table>

$^a C = \mu_B N_A k_B^2 / 3 k_B$ where $N_A$ is the Avogadro’s number, and $k_B$ is the Boltzmann constant.

### 3.4. The spin interacting model

The failure of the non-interacting model in reproducing CD6 experimental magnetization curve points out the necessity of including spin exchange interactions. In particular, we used a standard statistical approach to describe the case of heptagonal radicals with intra-molecular interactions. All the theoretical calculations and fits have been implemented by home-made Mathematica 9.0 routines. Since the radicals can be imagined as equally spaced on CD rim, at first we considered a configuration of seven spins at the vertices of a regular heptagon (heptagonal-configuration) as shown in Fig. 7a with periodic boundary conditions $(S_{N+1} = S_1)$ described by a Heisenberg-like Hamiltonian:

$$H = -g\mu_B \sum_i \mu_i \mathbf{H} \cdot \mathbf{S}_i - 2J_1 \sum_{\substack{i \neq j \in \{\text{f.n.}\}}} \mathbf{S}_i \cdot \mathbf{S}_j - 2J_2 \sum_{\substack{i \neq j \in \{\text{s.n.}\}}} \mathbf{S}_i \cdot \mathbf{S}_j$$  \hspace{1cm} (4)

The first term represents the Zeeman contribution, the second and the third terms describes the coupling between first-neighbors (f.n.) and second-neighbors (s.n.) spins respectively with $J_1$ and $J_2$ representing the exchange integrals. Unfortunately, no reasonable choice of $J_1$ and $J_2$ parameters could successfully take into account our experimental data (see Fig. SI14†).

**Fig. 6** Comparison between CD3 and CD6 magnetization data. The continuous lines represent a Brillouin function for a $S = 1/2$ non-interacting spin ensemble and a model function for a system of seven non-interacting $S = 1/2$ spins, for CD3 and CD6 respectively.

**Fig. 7** Four different configurations for the seven radicals: (a) highly symmetric heptagonal configuration: the blue and red lines represent the $J_1$ and the $J_2$ coupling constants respectively. (b–d) In this case $J_2$ represent the exchange interaction between the central spin and the others ones whereas $J_2$ takes into account all other possible interactions excluding the central spin (see text for details).
As second step we considered a less symmetric spin configuration with a fixed (central) spin with exchange interactions $J_1$ with its six neighbors (blue lines in Fig. 7b–d) and $J_2$ coupling between the other spin (red lines). In principle, this could be realized if one of the seven radicals is located into CD cavity and/or the whole molecule could be somehow distorted. In Fig. 7 we represent three different possible configurations, namely (b) 313-, (c) 2221- and (d) 61 as derived by following ref. 17.

The results of the analysis in the framework of our model are shown in Fig. 8 and summarized in Table 2. We note the following: (i) all the fitting functions are in very good agreement with experimental data. (ii) The values of the exchange integral $J_1$ and $J_2$ are both negative in all the configurations pointing to the AFM nature of the supposed intra-molecular interactions. (iii) They are of the same order of $k_B T$ confirming the regime of weak coupling in which the Brillouin functions are not applicable. (iv) $J_1$, $J_2$ values are in good agreement with the case of other multiradicals complex like calix[4]arene nitroxide derivatives, the $1/r_{AB}$

3.5. Relaxivity measurements

In view of the possible application for MRI of the proposed derivatives, the $1/T_1$ relaxation rates of water in CD3 and CD6 solutions in DMSO-d$_6$–H$_2$O (9 : 1) solutions at 305 K at different concentrations were measured. The associated water relaxivities ($r_1$) were calculated by fitting the data reported in Fig. 9 with eqn (1). For comparison, the same measurements were carried out considering also the derivative 4-OH-TEMPO as model molecule. DMSO was chosen as solvent because both CD3 and CD6 were scarcely soluble in water. The relaxation rate increased in a linear fashion for all the compounds. The linear regression fitting provided the relaxivity values reported in Table 3. The relaxivity associated to 4-OH-TEMPO is consistent with the value reported in literature ($0.28$ mM$^{-1}$ s$^{-1}$, in phosphate buffer at 25 °C). A slight increment of the relaxivity is observed with regard to the compound CD3. Probably, this effect could be associated to the higher molecular weight of CD3 compared to 4-OH-TEMPO. Indeed, it is reported that the water relaxivity increases with the molecular weight of CD3 compared to 4-OH-TEMPO. According to the eqn (2), the relaxivity is proportional to the square of the effective magnetic moment. As the two derivatives CD3 and CD6 differ only for the number of TEMPO units onto the βCD upper rim, it is interesting to verify how this aspect affects the proportionality constant between $r_1$ and $μ_{eff}^2$. The ratio $r_1$(CD6)/$r_1$(CD3) = 4.94 is quite close to the ratio of the squared magnetic moments, $μ_{eff}^2$(CD6)/$μ_{eff}^2$(CD3) ≈ 6.19, but these values are not coincident. Thus, the inequality $r_1$(CD6)/$r_1$(CD3) < $μ_{eff}^2$(CD6)/$μ_{eff}^2$(CD3) is equivalent to $k$(CD6) < $k$(CD3) (see eqn (2)).

![Fig. 8 CD6 magnetization data vs. 313 (blue line), 2221 (red line) and 61 (green line) configuration model calculations.](image1)

![Fig. 9 Relaxation rates for 4-OH-TEMPO (▲), CD3 (■) and CD6 (◆) in DMSO-d$_6$–H$_2$O (9 : 1) at 305 K.](image2)

<table>
<thead>
<tr>
<th>Configuration</th>
<th>$J_1$/($k_B T$)</th>
<th>$J_2$/($k_B T$)</th>
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<tbody>
<tr>
<td>313</td>
<td>−1.07</td>
<td>−0.47</td>
</tr>
<tr>
<td>2221</td>
<td>−1.06</td>
<td>−0.81</td>
</tr>
<tr>
<td>61</td>
<td>−1.07</td>
<td>−0.53</td>
</tr>
</tbody>
</table>
result is probably ascribed to an enhanced hydrophobicity of the derivative CD6 compared to CD3, due to the reduced number of hydroxyl groups and the increased number of TEMPO units in CD6. The structural differences between CD3 and CD6 are expected, in turn, to affect parameters such as the number of inner-sphere bound water molecules and the correlation time, factors concurring to the overall values of $k^\text{eff}$.\footnote{In conclusion, if from one side the introduction of more TEMPO units on the βCD molecule positively affects the relaxivity by increasing the effective magnetic moment of the CA, from another side it leads to a modification of the interactions between the water molecules and the CA, causing a small, but not negligible, reduction of the water relaxivity values. Finally, we would like to add a note about the reduced solubility in water of the proposed derivatives CD3 and CD6. Despite water soluble molecules are required for the applications, this aspect is not crucial in the present work. Instead, it is aimed mainly at investigating how the number of TEMPO radical units onto the βCD molecule affects the magnetic behavior of the corresponding derivative. The interesting results obtained so far strengthen the validity of the methodological approach followed in this work that does not depend on the effective solubility in water of the compounds. With regard to this point, CD3 and CD6 should be considered as model molecules rather than final derivatives for a direct application. However, a future development of the work is certainly oriented towards the synthesis and characterization of cyclodextrin-polynitroxides with increased solubility in biological media.

4. Conclusions

In summary, two βCD derivatives bearing one or seven TEMPO radical units (CD3 and CD6 respectively) on the small rim of the βCD molecule have been synthesized and characterized. Their effective magnetic moments have been measured by DC-SQUID magnetometry. While isothermal magnetization data of CD3 are well fitted by a Brillouin function for $S = 1/2$ single spin system, those associated to CD6 cannot be simply described in the framework of a non-interacting spins model. In order to fully understand CD6 experimental data, four different configurations for the seven interacting free-radicals, with or without the presence of a central spin, have been considered and modeled. The numerical results evidences that only the configurations with a privileged central spin fully takes into account the experimental behavior justifying a reduced effective magnetic moment. The effective magnetic moments $\mu_{\text{eff}}$ of both the derivatives were determined from dc susceptibility measurements obtaining the values $\mu_{\text{eff}}/\mu_B = 1.686$ and $\mu_{\text{eff}}/\mu_B = 4.193$ for CD3 and CD6 respectively.

Considering the envisaged application of the proposed βCD-TEMPO derivatives as macromolecular building blocks for nitroxides-based MRI contrast agents, we studied the NMR relaxivities ($r_1$) in DMSO-d$_6$–water (9 : 1 v : v) solutions. The values $r_1 = 0.323$ mM$^{-1}$ s$^{-1}$ and $r_1 = 1.596$ mM$^{-1}$ s$^{-1}$ for CD3 and CD6 respectively were obtained. Interestingly, the values associated to CD6 is not too far from Gd-based CA relaxivity.\footnote{Taking into account the well-known reduced toxicity of TEMPO radicals, βCD bearing multiple TEMPO moieties can be surely considered as promising MRI and ESR CA for in vivo experiment. For this reason, future efforts will be devoted to the synthesis of analogous water-soluble derivatives.}

Contributions

L. Me. contributed to the synthetic part. F. Cg., F. Cn. and G. L. carried out the magnetic measurements, and analyzed the data by a quantum-statistical model. F. C., M. F. and A. M. performed the relaxivity measurements. P. F. and M. L. performed the ESR spectroscopy measurements. All the coauthors contributed to the analysis and the interpretation of the data. L. Me., F. Cg. and G. L. wrote the manuscript, with the support of all the coauthors.

Acknowledgements

The Ministry for Education, University and Research (MIUR) is kindly acknowledged through the following Research Projects: PRIN 2010-2011-NANOMED, PRIN-2010-2011-PROXi project 2010PFLRJR_005, and PRIN MULTINANOITA.

Notes and references


