



Molecular architecture control in synthesis of spherical Ln-containing nanoparticles^{†‡}

Wuyuan Zhang,^a Jonathan Martinelli,^a Florian Mayer,^a Célia S. Bonnet,^b Frédéric Szeremeta^b and Kristina Djanashvili*^a

Among the procedures to prepare lanthanide-containing nanoparticles a gap exists in the range between 5 and 40 nm. The miniemulsion technique presented here is intended to fill this discontinuity and offers a facile method that can be applied for the preparation of nanoparticles for various applications, e.g. medical imaging, optics and catalysis. We demonstrate that formation of nanodroplets under emulsion conditions is the key step in the size control of the nanoparticles. The type of surfactant and the nature of the dispersed and continuous phases strongly influence the interfacial activity and, consequently, the size of the final solid particles that result from the subsequent thermal decomposition. Moreover, the choice of the surfactant determines the final elemental composition of the particles, leading to either lanthanide oxides or oxysulfates when using Brij® 35 or sodium dodecyl sulfate, respectively. Nanoparticles of holmium and gadolinium were prepared and their applicability as magnetic resonance imaging contrast agents is shown.

Received 19th May 2015
Accepted 10th August 2015

DOI: 10.1039/c5ra09374k
www.rsc.org/advances

Introduction

Lanthanide-containing nanoparticles (Ln-NPs) have attracted considerable interest in recent years, and their properties have been thoroughly investigated in relation to the various applications such as optics,¹ catalysis² or biomedicine,^{3–6} including cell labelling, diagnostics, therapy and combinations thereof. For instance, due to their high Ln³⁺-loading, targeting Ln-NPs can deliver high payloads of Ln-ions to the site of interest,⁷ which allows the detection of biomarkers expressed at low levels on a cell surface.⁸ Size and morphology control plays an essential role in application of NPs. For example, it has been demonstrated that non-spherical shapes translate into cytotoxicity due to a higher chance to damage the cell membranes.⁹ Furthermore, the cellular as well as tissue and organ distribution of NPs is highly dependent on the particle size.^{10,11} Particles less than 100 nm in diameter can easily pass through the vasculature, and the smallest ones can even spread into organs.^{12,13} In case of application of NPs as contrast agents (CAs) for magnetic resonance imaging (MRI), the size becomes particularly essential as it determines their usefulness for either

T_1 - or T_2 -weighted imaging. Since the efficacy of positive (T_1) nanoparticulate CAs depends on the interaction of water molecules with paramagnetic ions at the surface of NPs, a high surface-to-volume ratio is required and thus NPs with a diameter smaller than 10 nm are preferred. On the other hand, negative (T_2) contrast depends on magnetic susceptibility and the magnitude of magnetic moment, which are in direct relation with the number of paramagnetic ions per particle and hence the size of the applied NP.

A large number of publications reports on Ln-based NPs such as oxides,^{14,15} carbonates,¹⁶ oxysulfides,¹⁷ polysiloxane networks,¹⁸ hybrids,¹⁹ zeolites, and porous silica supporters.⁶ The size of these particles varies widely in the range of 1–200 nm depending on the preparation methodologies applied. The polyol method is particularly useful to prepare ultrasmall Gd-oxide NPs (1–3 nm).²⁰ However, for larger particles this procedure is time-consuming due to slow nucleation and the need for repetition of seed growth, and poorly reproducible as there is no control over the yield and uniformity of the resultant NPs. The polyol method has also raised concerns about inhibitory effects on neutrophil oxidative burst that might be caused by the presence of diethylene glycol on the surface of particles prepared in such solvent.²¹ We have previously reported that Ln carbonate NPs (turning into oxides upon calcination) can be readily prepared *via* controlled microwave-assisted seed growth.¹⁶ The smallest size achievable with reasonable yield and good homogeneity by this method is 40 nm, as demonstrated by dynamic light scattering. Finally, the solvothermal method is dominating in the synthesis of upconversion NPs with highly controlled size and morphological uniformity. The resulting

^aDepartment of Biotechnology, Delft University of Technology, Julianalaan 136, 2628 BL, Delft, The Netherlands. E-mail: k.djanashvili@tudelft.nl

^bCentre de Biophysique Moléculaire, UPR4301, CNRS, Université d'Orléans, Rue Charles Sadron, 45071 Orléans Cedex 2, France

† This paper is dedicated to Professor Imre Tóth from the University of Debrecen, Hungary on the occasion of his 65th birthday.

‡ Electronic supplementary information (ESI) available: XRD, EDS, FT-IR, TGA, and TEM measurements of the dried Ho- and Gd-containing droplets and corresponding nanoparticles. See DOI: 10.1039/c5ra09374k



particles have promising applicability in optical imaging, but their usefulness with respect to MRI is limited.²² Recently, Eu-doped Gd_2O_3 particles in a 5–200 nm range for multimodal/sequential CAs prepared by chemical vapor synthesis at high temperature have been reported.²³ Alternatively, a laser ablation method performed by focusing the laser beam onto a Gd target has been described,²⁴ resulting in spherical Gd_2O_3 particles with an average diameter of 7 nm.

The miniemulsion (or nanoemulsion) technique, widely applied in polymer chemistry,^{25,26} is however uncommon for the preparation of inorganic NPs. Considering the great interest towards NPs with defined size, we describe herein a novel miniemulsion method for the facile size-controlled preparation of spherical Ln-based NPs in the otherwise not readily accessible range of 5–40 nm. A miniemulsion consists of dispersed nanosized droplets formed from two phases of different polarity upon exposure to high power ultrasound. The resulting nanodroplets usually have a narrow size distribution and can be well stabilized in emulsion by an appropriate surfactant. The formation of the droplets highly depends on tunable factors such as composition, phase or surfactant,²⁵ and the miniemulsion technique can be extended to the generation of composite materials, in which different metals as well as surface coatings can be incorporated.^{27–29}

The method reported herein involves two steps: an easy and efficient preparation of miniemulsions consisting of Ln nanodroplets, and a subsequent thermal decomposition to obtain Ln-containing solid particles. It is noteworthy that the nature of the product obtained through thermal decomposition is highly dependent on the surfactant in the continuous phase: for example Ln_2O_3 is obtained using Brij® 35, while $\text{Ln}_2\text{O}_2\text{SO}_4$ is produced in the presence of sodium dodecyl sulfate (SDS). This simple miniemulsion method opens new and interesting alternatives to prepare NPs for a variety of applications, including imaging and therapy.

Experimental section

Materials and methods

All chemicals were used as supplied from commercial sources: analytical grade solvents, sodium dodecyl sulfate, octadecane (Sigma-Aldrich), polyethylene glycol dodecyl ether (Brij® 35, $M_n \sim 1198$), acetylacetone (acac) (Fluka). “ H_2O ” refers to high purity water with conductivity of 0.04 $\mu\text{S cm}^{-1}$, obtained from a Milli-Q purification system. Lanthanide chlorides ($\text{LnCl}_3 \cdot 6\text{H}_2\text{O}$, Strem Chemicals) were dissolved in 10 ml water, the obtained solutions were filtered over a 200 μm nylon syringe filter and evaporated to dryness under high vacuum before use. $\text{Ln}(\text{acac})_3$ were synthesized according to published procedures:³⁰ typically, acetylacetone (0.220 mol) was dissolved in H_2O (135 ml) and the pH of this solution was adjusted to 8.5 by portion-wise addition of 25% ammonium hydroxide; LnCl_3 (10 mmol; Ln = Gd, Ho) was dissolved in H_2O (5 ml) and slowly added to the above-mentioned solution under stirring. The mixture was stirred overnight and the pink ($\text{Ho}(\text{acac})_3$) or white ($\text{Gd}(\text{acac})_3$) crystals were filtered, washed three times with water and air-dried.

Preparation of Ln-containing nanodroplets by a miniemulsion method

In a typical procedure to prepare the miniemulsion, the dispersed hydrophobic phase was obtained by mixing $\text{Ln}(\text{acac})_3$ (50 mg) in CHCl_3 or dichloromethane (DCM) (2.5 ml). This suspension was then added dropwise to a continuous phase consisting of SDS or Brij® 35 dissolved in H_2O (8 ml). The mixture was vigorously stirred at room temperature for 1 h (pre-emulsion) and then ultrasonicated (Qsonica Sonicator, 500 W, 1/2 inch tip, 80% amplitude, time interval 0.5 s) for 5 min under ice cooling, yielding milky miniemulsions (pink for Ho and white for Gd). The hydrophobic solvents were removed by careful rotary evaporation at 30 °C to avoid aggregation, until a turbid dispersion appeared. These dispersions were used to measure the hydrodynamic radius of the nanodroplets, and they were then freeze-dried yielding Ln-containing powders.

Phase transformation

Thermal decomposition of the previously obtained powders was performed by calcination at 800 °C under air atmosphere for 1 h applying a heating rate of 2.5 °C min⁻¹. The average yield of this procedure was 82% with respect to $\text{Ln}(\text{acac})_3$.

Characterization

The size of the nanodroplets in the miniemulsion was measured by dynamic light scattering (DLS). The experiments were carried out by using a Malvern Zetasizer NanoZS operating in a particle size range from 0.6 nm to 6 μm and equipped by a He-Ne laser with $\lambda = 633$ nm. The measurements were performed at 25 °C, and the average diameter was based on three individual measurements, 20 scans for each measurement. To evaluate the process of conversion of the freeze-dried Ln-containing nanodroplets into solid particles, thermogravimetric analysis (TGA) was conducted by using a Perkin-Elmer Thermogravimetric Analyzer TGA7 equipped with a Thermal Analysis Controller TAC 7/DX, from 25 to 900 °C (10 °C min⁻¹) under air atmosphere. X-ray diffractometry (XRD) measurements were performed on a Bruker D8 Advance X-ray diffractometer using Co K α radiation (1.789 Å) at 35 kV and 40 mA. The data were collected from 5° to 80° 2 θ with a step size of 0.020° 2 θ and a counting time of 0.5 s per step. Lattice constants were calculated and corrected using MDI JADE software. Transmission electron microscopy (TEM) analyses were performed on a JOEL JEM-2100 transmission electron microscope. Samples of nanoparticles from dilute aqueous dispersions were evaporated over a 400-mesh copper measurement grid. The instrument operated at an accelerating voltage of 200 kV for the acquisition of section images. The surface composition of formed NPs was characterized by Infrared (IR) spectroscopy performed on a Perkin-Elmer Spectrum One FT-IR spectrometer using KBr tablet. The chemical composition of NPs was analyzed by energy dispersive X-ray spectroscopy (EDS) on a JEOL-JMS 6010 scanning electron microscope with an acceleration voltage of 20 kV. The longitudinal (T_1) and transversal (T_2) water proton relaxation times were measured on a Varian-INOVA 300 NMR



spectrometer applying inversion recovery and CPMG sequences, respectively. For the latter sequence the half interval time (τ_{cp}) between the successive 180° pulses was fixed at 1.0 ms. The samples for these measurements were prepared by suspending defined amounts of particles in 0.5% xanthan gum solution. The same equipment was exploited to determine the concentrations of Ln^{3+} ($\text{Ln} = \text{Ho}$ and Gd) ions in the aqueous solutions *via* bulk magnetic susceptibility (BMS) method.³¹ MRI experiments were conducted on a PharmaScan 7 T horizontal magnet (B-C 70/16 US, Bruker BioSpin, Wissembourg, France) equipped with B-GA09 gradient system (120 mT m⁻¹ maximal strength and 90 mm inner diameter) and Paravision 4.0 software (Bruker BioSpin). T_1 -weighted and T_2 -weighted MR images were acquired with spin-echo sequence (RARE sequence with one echo to get a small echo time (TE) and to make the effective-TE equal to TE) at 25 °C. T_1 -weighted images were acquired with 10.6 ms TE and 250 ms TR (repetition time) and 200 × 200 μm^2 resolution with a matrix 128 × 128 in 32 s. T_2 -weighted images were acquired with 90 ms TE and 3000 ms TR and 400 × 400 μm^2 resolution with a matrix 64 × 64 in 3 min. All images have 2.0 mm thickness.

Results and discussion

Formation of the nanodroplets

In order to obtain a miniemulsion containing homogeneous and stable nanodroplets, a hydrophobic dispersed phase and a hydrophilic continuous phase need to be mixed and sonicated. In general, once homogeneously distributed nanodroplets are obtained, no decomposition or aggregation takes place and the nanodroplets can be converted into a solid material by removal of the solvents. This strategy was successfully exploited for the synthesis of lanthanide-containing NPs (Scheme 1).

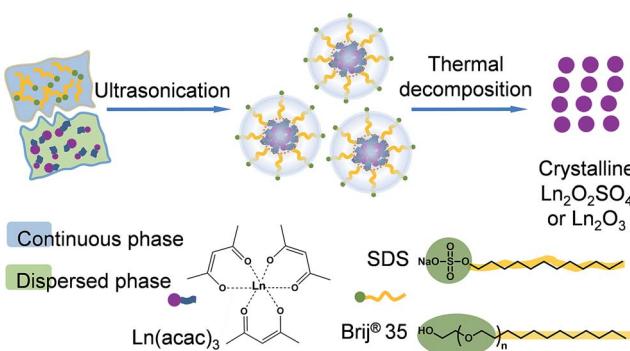
Homogeneous nanodroplets in miniemulsion were prepared by powerful sonication of a mixture of $\text{Ln}(\text{acac})_3$ in the dispersed phase and surfactant in the continuous phase. The conditions of sonication in terms of mechanical energy and duration were chosen to allow the nanodroplets in emulsion to reach a steady state.³⁰

The presence of a surfactant facilitates the nucleation by forming a region of hydrophobic tails where nanosized chloroform or dichloromethane cores containing $\text{Ln}(\text{acac})_3$ are trapped and act as nanoreactors; the hydrophilic moieties extend into a polar continuous phase of water. Such configuration provides discrete particle formation. The resulting nanodroplets were then isolated and thermally decomposed into solid nanoparticles.

Different synthetic conditions, such as combinations and amounts of solvents and surfactants, were investigated (Table 1). The various products obtained were preliminary characterized *via* DLS size measurements. At first, preparations were carried out with $\text{Ho}(\text{acac})_3$ in chloroform as dispersed phase, while SDS was used as surfactant in the aqueous phase. It is a common strategy in miniemulsion techniques to add a hydrophobic co-stabilizing agent to stabilize the droplets against Ostwald ripening.³² Here, we selected octadecane to adjust the osmotic pressure of the miniemulsion. It appeared that the presence of octadecane in the dispersed phase had a positive effect on reducing the size of the Ho-containing droplets, but decreasing its concentration led to higher polydispersity index (PDI) values (Table 1, entries 1–3). TEM images of the particles obtained after thermal decomposition of these droplets showed that the presence of octadecane leads to less homogenous solid particles (Fig. S1†). Therefore, a hydrophobic agent was omitted in the next preparations in order to achieve a better homogeneity of the resultant solid particles.

Although it has been demonstrated that in general the concentration of surfactant plays a crucial role in controlling the size of the droplets,³³ this effect was observed only to a limited extent in the present case: varying the concentration of SDS in the continuous phase from 10 to 30 mM (Table 1, entries 3 and 4) did not change the droplet size significantly (as measured by DLS, 45 vs. 51 nm). Furthermore, increasing the SDS concentration over 30 mM resulted in unstable nanodroplets precipitating within 3 days.

The diffusion of the dispersed phase through the water phase has been shown to limit the growth of nanodroplets.²⁵ With this in mind, we investigated the use of more polar DCM as the solvent of the dispersed phase. When keeping the other conditions unmodified (Table 1, entries 3 and 5) the diameter of the droplets in the miniemulsion was only 26 nm as compared to 45 nm in CHCl_3 . This can be explained in terms of a different interface activity, where the interfacial tension is 32.80 mN m⁻¹ for $\text{H}_2\text{O}/\text{CHCl}_3$ and 28.31 mN m⁻¹ for $\text{H}_2\text{O}/\text{DCM}$.³⁴ Thus, a relatively minor change in the polarity of the dispersed phase has a strong impact on the formation of the droplets in the miniemulsion. To further investigate this effect, 12.5 wt% of ethanol was added to the continuous phase to improve the diffusion at the interface between H_2O and DCM;³⁵ this led to an increase in the size of the nanodroplets from 26 to 37 nm (Table 1, entries 5 and 6), in accordance with the improved solubility of $\text{Ln}(\text{acac})_3$ and the higher miscibility of DCM towards the continuous phase. Doubling the concentration of SDS for the $\text{H}_2\text{O} + \text{EtOH}/\text{DCM}$ resulted in notably bigger nanodroplets (from 37 to 57 nm; Table 1, entries 6 and 7). It was also noticed that with anionic SDS as surfactant, the PDIs of the



Scheme 1 Two-step preparation of Ln-NPs. Small and homogeneous nanodroplets in miniemulsion were first achieved by powerful sonication followed by evaporation of the hydrophobic solvent; freeze-drying and subsequent calcination led to the aimed systems.



Table 1 Experimental data for the preparation of Ln-containing nanodroplets and NPs under different conditions

Entry	Metal ^a	Dispersed phase ^b	Octadecane (mg)	Continuous phase	Surfactant ^c (mM)	DLS sized ^d (nm)	PDI	Product after calcination ^e	TEM size (nm)
1	Ho	CHCl ₃	30	H ₂ O	SDS (10)	37 ± 6	0.25	Ho ₂ O ₂ SO ₄	15 ± 3.1
2	Ho	CHCl ₃	5	H ₂ O	SDS (10)	43 ± 4	0.34	Ho ₂ O ₂ SO ₄	18 ± 3.8
3	Ho	CHCl ₃	0	H ₂ O	SDS (10)	45 ± 4	0.37	Ho ₂ O ₂ SO ₄	24 ± 5.7
4	Ho	CHCl ₃	0	H ₂ O	SDS (30)	51 ± 7	0.44	Ho ₂ O ₂ SO ₄	— ^f
5	Ho	DCM	0	H ₂ O	SDS (10)	26 ± 3	0.39	Ho ₂ O ₂ SO ₄	8 ± 1.8
6	Ho	DCM	0	H ₂ O/EtOH	SDS (10)	37 ± 9	0.31	Ho ₂ O ₂ SO ₄	18 ± 2.0
7	Ho	DCM	0	H ₂ O/EtOH	SDS (20)	57 ± 6	0.39	Ho ₂ O ₂ SO ₄	— ^f
8	Ho	DCM	0	H ₂ O	Brij® 35 (5.2)	109 ± 9	0.087	Ho ₂ O ₃	12 ± 2.0
9	Gd	DCM	0	H ₂ O	SDS (10)	33 ± 7	0.40	Gd ₂ O ₂ SO ₄	— ^f
10	Gd	DCM	0	H ₂ O	Brij® 35 (5.2)	98 ± 11	0.10	Gd ₂ O ₃	7 ± 2.1

^a 50 mg of Ln(acac)₃ for each batch. ^b 2.5 ml for each batch. ^c With respect to the continuous phase. ^d Average diameter. ^e As determined by XRD.

^f Not measured.

obtained nanodroplets were relatively high (generally between 0.3 and 0.4). This may be explained by the effect of collisions between nanodroplets: the smaller the droplets, the shorter the distance and the higher their number, thus the collision rate increases; eventually, this leads to destabilization and a broader PDI.³⁶ However, when SDS was replaced with Brij® 35, a purely organic and non-ionic surfactant, the PDI dropped dramatically (e.g. 0.09 vs. 0.39; Table 1, entries 5 and 8). At the same time the size of the nanodroplets increased significantly (109 vs. 26 nm). The latter phenomenon is the consequence of the good affinity between water and the long hydrophilic polyethylene glycol chain of Brij® 35.

Once the factors playing a role in the preparation of Ho-nanodroplets were assessed, their applicability and reproducibility were checked by preparing analogous Gd-based systems. With Gd(acac)₃ under the same conditions (Table 1, entries 5 and 9) the obtained Gd-nanodroplets had a size of 33 nm, consistent with the value previously measured for the corresponding Ho-based products (26 nm). Also the replacement of SDS with Brij® 35 as surfactant had similar effects (Table 1, entries 9 and 10); a remarkable increase in size (98 vs. 33 nm) and a significant lowering of the PDI (from 0.40 to 0.10). These results show that the formation of nanodroplets in a miniemulsion process can be reliably tuned not only by adjusting the polarity of the two phases, but also by playing with the other components in general and the surfactant in particular.

Formation of the nanoparticles

The second step in the preparation of Ln-NPs consists of the transformation of the nanodroplets from the miniemulsion into solid particles through calcination (Scheme 1). Both the Ho- and Gd-containing nanodroplets were dried and then decomposed under aerobic conditions at 800 °C for 1 h. The nature of the product after thermal decomposition turned out to depend on the surfactant added during the synthesis. When SDS was used, the calcination yielded oxysulfates (Ln₂O₂SO₄) nanocrystals, the surfactant being the source of sulfur. The evolution in the crystal structure can be observed by X-ray powder diffraction measurements (Fig. 1): the XRD pattern of

the dried Ho-nanodroplets is mainly featureless, indicating a basically amorphous nature (Fig. S2‡); after calcination, Ho₂O₂SO₄ is obtained instead in a crystalline form (Fig. 1a, top). All the XRD reflections revealed a crystal structure of Ho₂O₂SO₄ with calculated lattice constants $a = 4.045 \text{ \AA}$, $b = 4.185 \text{ \AA}$ and $c = 12.963 \text{ \AA}$. In addition, a diffraction peak of Ho₂O₃ was observed (Fig. 1a), the intensity of which was mainly dependent on the applied Ho(acac)₃ : SDS ratio.

The thermal decomposition of Gd-containing nanodroplets showed analogous effects (Fig. 1a, bottom). Noteworthy, it is possible to obtain Ln₂O₂SO₄ NPs with higher purity by adjusting the molar ratio of the starting materials, as shown for Gd(acac)₃ and SDS during the preparation of the corresponding nanodroplets. The XRD pattern of the obtained crystalline Gd₂O₂SO₄ reveals calculated lattice constants $a = 4.051 \text{ \AA}$, $b = 4.174 \text{ \AA}$ and $c = 12.973 \text{ \AA}$.

When non-ionic surfactant Brij® 35 was used in the miniemulsion, the subsequent thermal decomposition yielded crystalline oxide (Ln₂O₃) nanocrystals (Fig. 1b). All reflections in the XRD spectra can be indexed to the cubic structure of Ln₂O₃ (space group: $Ia\bar{3}$ (no. 206)) with calculated lattice constants $a = 10.606 \text{ \AA}$ for Ho₂O₃ and $a = 10.812 \text{ \AA}$ for Gd₂O₃, in perfect agreement with those reported (JCPDS 44-1268, $a = 10.610 \text{ \AA}$ and JCPDS 65-3181, $a = 10.818 \text{ \AA}$ respectively). The observed red shift of the patterns is attributed to the use of Co K α radiation (1.789 \AA) instead of usual Cu (1.541 \AA).

TGA profiles of the thermal decompositions were recorded (Fig. 2). The starting material Ho(acac)₃ has a major multistep weight loss in the temperature range 30–550 °C that includes the dehydration from structurally bound water between 30 and 200 °C and decomposition of the organic chelate acetylacetone up to 550 °C; holmium oxide is finally obtained. The thermogravogram between 30 and 850 °C of a sample of Ho-containing nanodroplets obtained from Ho(acac)₃ in the presence of SDS as surfactant shows a dominant loss (45%) between 170 and 350 °C, that can be attributed to the major decomposition of SDS and acetylacetone. A total 62% loss was measured until 760 °C, in agreement with the value calculated when considering the reaction (Scheme 1) with 35 wt% retained taking S, Ho and O into account.



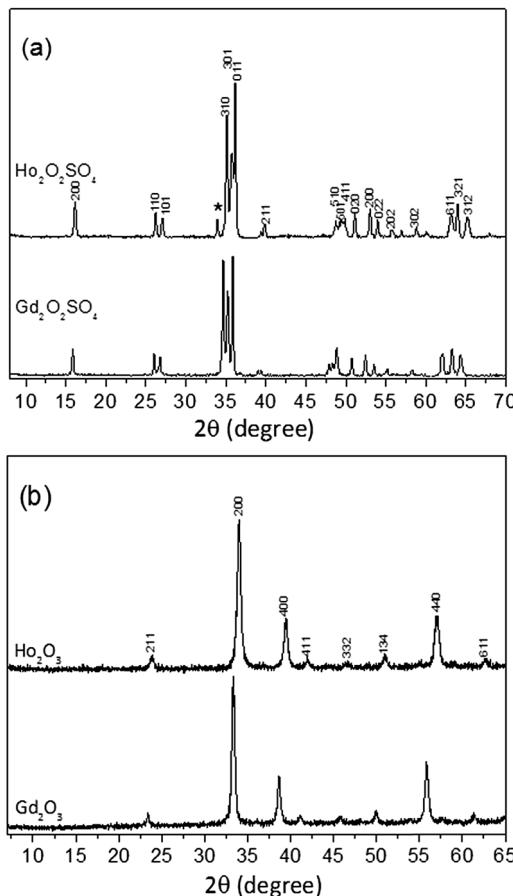


Fig. 1 XRD patterns of the solid Ln-containing NPs obtained by miniemulsion and subsequent open-air thermal decomposition at 800 °C for 1 h. (a) $\text{Ln}_2\text{O}_2\text{SO}_4$ NPs, SDS was used as surfactant; the peak marked with an asterisk is due to Ho_2O_3 . (b) Ln_2O_3 NPs, Brij® 35 was used as surfactant.



It has been reported that an alternative stacking of $\text{La}_2\text{O}_2^{2+}$ and SO_4^{2-} layers takes place under treatment at high temperature of hydrous nitrates ($\text{Ln}(\text{NO}_3)_3 \cdot n\text{H}_2\text{O}$) intercalated with dodecyl sulfate ions.^{37,38} The gradual decomposition observed in the range 300–700 °C can thus be ascribed to the stacking mechanism leading to the formation of $\text{Ln}_2\text{O}_2\text{SO}_4$. Such gradual process proceeds until 760 °C to yield Ln-oxysulfates, in agreement with the XRD evolution (Fig. S3†). The formation of $\text{Gd}_2\text{O}_2\text{SO}_4$ follows a totally analogous pathway (Fig. 2b).

Different TGA profiles were recorded for dried nanodroplets obtained in the presence of Brij® 35 as surfactant (Fig. 2). The dominant weight loss (~78%) between 200 and 500 °C corresponds to the decomposition of organic Brij® 35 and acetylacetone. Above 500 °C, the thermogravogram reaches a plateau, indicating that Ho_2O_3 or Gd_2O_3 NPs are obtained.

The surface chemical structure of the obtained various NPs was characterized by Fourier transform infrared spectroscopy (Fig. S4†). The main starting material $\text{Ln}(\text{acac})_3$ ($\text{Ln} = \text{Ho}$ or Gd)

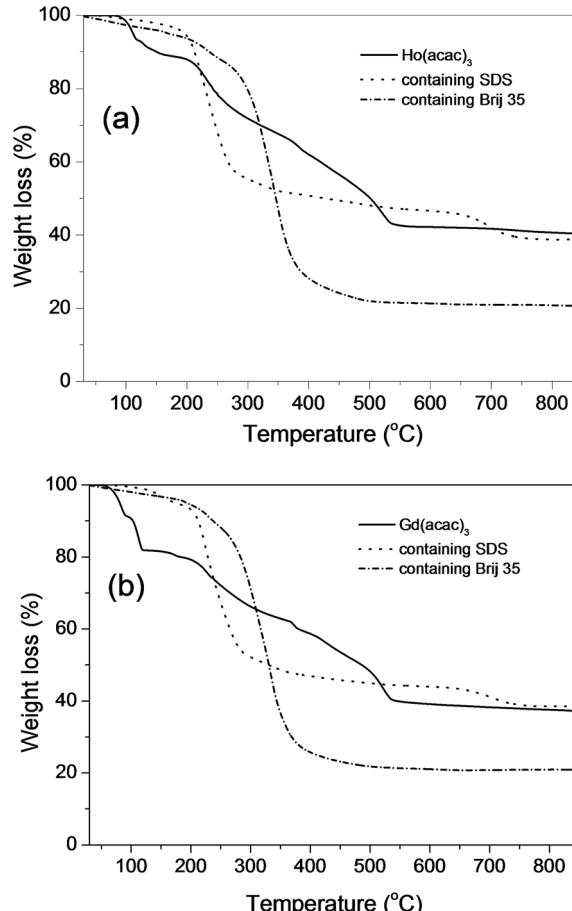


Fig. 2 TGA profiles of $\text{Ln}(\text{acac})_3$ and the corresponding Ln-containing nanodroplets prepared by using SDS or Brij® 35 as surfactants. $\text{Ln} = \text{Ho}$ (a) or Gd (b).

shows the characteristic bands of enolized β -diketones at about 1610, 1520 and 1400 cm^{-1} . After thermal decomposition, the resulting $\text{Ln}_2\text{O}_2\text{SO}_4$ exhibited characteristic bands of SO_4^{2-} ion coordinated to Ln^{3+} ion at 1210, 1130, 997, 664 and 619 cm^{-1} , with an indication of low site symmetry in the oxysulfate.³⁹ In contrast, the spectra of materials prepared by using Brij® 35 exhibited a strong peak at 550 cm^{-1} , typical for cubic-type Ln_2O_3 NPs.⁴⁰

Energy dispersive X-ray spectroscopy (EDS) was further applied to determine the compositions of various Ln-based NPs. Fig. 3 reports the profiles for $\text{Ho}_2\text{O}_2\text{SO}_4$ and Ho_2O_3 NPs, while the data corresponding to the Gd-analogues are given in the ESI (Fig. S5†). It is clear that the obtained NPs are composed of $\text{Ln}_2\text{O}_2\text{SO}_4$ and Ln_2O_3 when SDS and Brij® 35 were used, respectively. The solids obtained after thermal decomposition consisted of fairly uniform spherical particles, as determined by TEM images (Fig. 4 and S1†).

As mentioned above, with this technique it was also possible to observe how the use of octadecane as a co-stabilizing agent in miniemulsions affects the final size of the particles: under the same reaction conditions (Table 1, entries 2 and 3) the addition of octadecane (10 wt% with respect to Ln-acetylacetone) led to

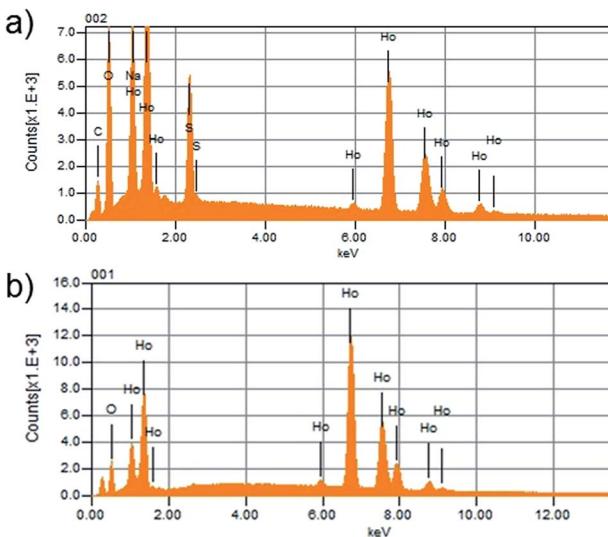


Fig. 3 EDS spectrum of (a) $\text{Ho}_2\text{O}_2\text{SO}_4$ and (b) Ho_2O_3 NPs obtained after calcination.

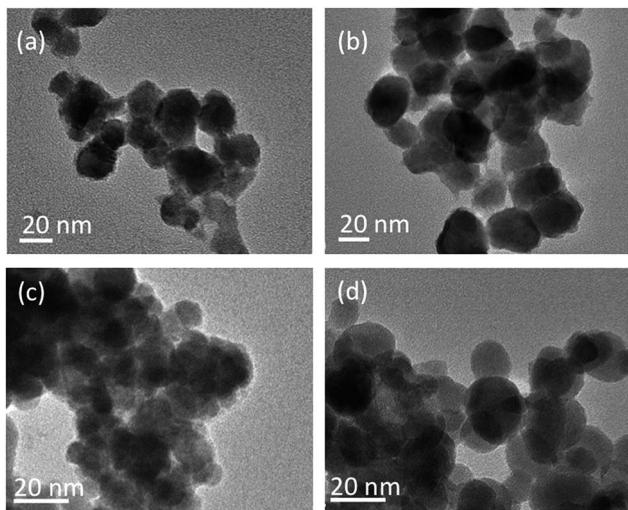


Fig. 4 TEM images of solid $\text{Ho}_2\text{O}_2\text{SO}_4$ NPs prepared using SDS in the continuous phase: (a) and (b) correspond to entries 2 and 3 in Table 1 with 10 and 0 wt% of octadecane in the dispersed phase, respectively; (c) and (d) correspond to entry 5 and 6 with 0 and 12.5 wt% of ethanol added in the continuous phase, respectively.

particles (Fig. 4a) with a diameter (18 nm) smaller than that (24 nm) obtained in its absence (Fig. 4b). This effect is consistent with the reported claim that increasing the osmotic pressure leads to a decrease in the particles size.³⁰ However, one of the advantages of the method discussed herein is that a hydrophobic co-reagent is not decisive in the preparation of nanodroplets.

As described above, changing the solvent of the dispersed phase from CHCl_3 to DCM (Table 1, entries 3 and 5) resulted in Ho-loaded nanodroplets with a significantly reduced diameter (from 45 to 26 nm). This is confirmed by the TEM images of the corresponding final $\text{Ho}_2\text{O}_2\text{SO}_4$ particles (Fig. 4b and c) with

diameters 24 and 8 nm, respectively. This further highlights the importance of the interfacial activity on the formation of the nanodroplets and, consequently, of the solid particles. The increase in size of the nanodroplets prepared with the addition of ethanol (Table 1, entry 6) is reflected in the TEM images of the corresponding final NPs (Fig. 4d), for which an average diameter of 8 nm was measured. Upon replacement of SDS with Brij® 35 as surfactant in the miniemulsion, the TEM images of Ho_2O_3 and Gd_2O_3 obtained upon calcination showed spherical particles with a diameter of 12 and 7 nm, respectively (Fig. S6†). Although the hydrodynamic diameters of the corresponding nanodroplets were by far larger (109 and 98 nm, respectively) due to the long polymer chain of Brij® 35 (Table 1, entries 9 and 10, and Fig. S7†), the solid Ln_2O_3 particles showed the expected small sizes.

Relaxivity and MRI studies of $\text{Ln}_2\text{O}_2\text{SO}_4$ and Ln_2O_3 NPs

As an example of their applicability, the Ln-NPs prepared as described above were evaluated in terms of their performance as magnetic resonance imaging contrast agents. Because of its seven unpaired 4f-electrons Gd is the most paramagnetic stable metal ion, and therefore Gd^{3+} -complexes are nowadays the most common longitudinal or T_1 CAs in medical MRI.^{41,42} On the other hand, Ho (together with dysprosium) is the most efficient transversal or T_2 CA for its highest magnetic moment.⁴³ In view of their possible application as MRI CAs, the final Ho- or Gd-based NPs were evaluated for transverse (r_2) and longitudinal (r_1) proton relaxivities. The water proton relaxation rates ($R_n = 1/T_n$, $n = 1$ or 2) of four samples of each lanthanide were measured at 25 °C and 7 T, and then plotted against the related concentration of paramagnetic Ln^{3+} ion in order to attain the corresponding relaxivity value as the slope of the obtained straight line (Fig. 5a). For $\text{Gd}_2\text{O}_2\text{SO}_4$ (12 nm) and Gd_2O_3 (7 nm) particles r_1 is 1.0 and $3.1 \text{ mM}^{-1} \text{ s}^{-1}$, respectively and is expected to be more impressive at the clinically used magnetic fields (1.5–3 T), according to the typical ^1H nuclear magnetic relaxation dispersion profiles of solid NPs.^{20,42} The effect of particle size on the MRI contrast enhancement is demonstrated by the acquisition of T_1 - and T_2 -weighted images of phantoms containing various concentrations of the particles at 25 °C and 7 T. The brightness of T_1 -weighted images acquired with Gd_2O_3 or $\text{Gd}_2\text{O}_2\text{SO}_4$ particles of 7 nm increases with increasing concentrations between 0.1 and 0.3 mM (Fig. 5b). This effect is in agreement with the obtained r_2/r_1 ratio of 10, which is known to be favorable for T_1 contrast.⁶ Increasing the particle size to 12 nm has a clear negative (dark) effect on the T_1 -weighted intensity, due to higher r_2/r_1 ratio (up to 36, Table 2) and a consequent dominance of T_2 on the intensity (Fig. 5b).

The r_2 values obtained for Ho_2O_3 (12 nm) and $\text{Ho}_2\text{O}_2\text{SO}_4$ (18 nm) are 31.1 and $43.9 \text{ mM}^{-1} \text{ s}^{-1}$, respectively. The difference in diameter between the two systems must be taken into account, due to the strong linear dependence of r_2 on the particle size.⁴⁴ Based on the low r_1 values, and consequently, high r_2/r_1 ratio, these particles are suitable for T_2 -weighted imaging, as demonstrated in Fig. 5c.



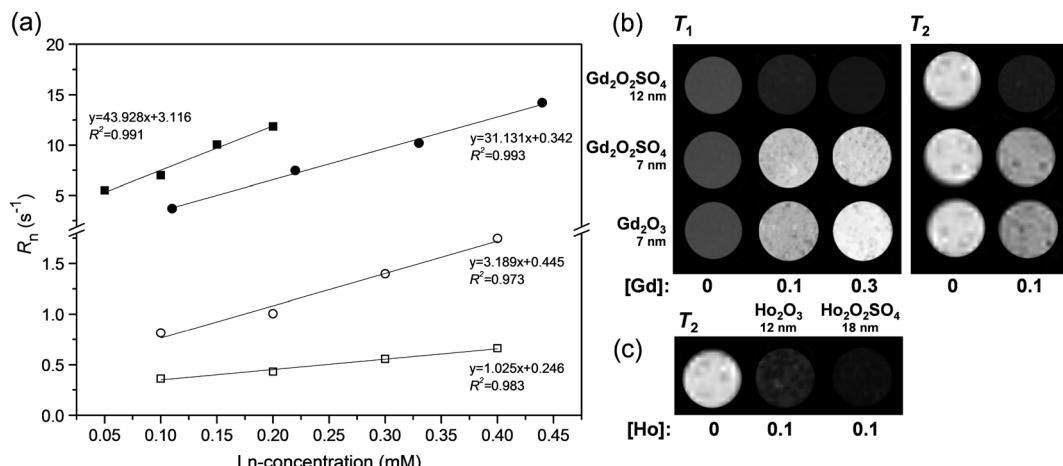


Fig. 5 MR relaxation rates and MR images of Gd- and Ho-based NPs dispersed in 0.5% solution of xanthan: (a) transversal (Ho, solid symbols) and longitudinal (Gd, open symbols) relaxation rates of oxysulfates (squares) and oxides (circles) NPs (7 T and 25 °C), (b) MR phantoms containing 0.1 and 0.3 mM Gd₂O₃ and Gd₂O₂SO₄ NPs (7 T and 25 °C), left: T_1 -weighted spin-echo MR images, TE = 10.6 ms and TR = 250 ms; right: T_2 -weighted MR images, TE = 90 ms and TR = 3 s, (c) T_2 -weighted images of xanthan phantoms containing 0.1 mM Ho₂O₃ and Ho₂O₂SO₄.

Table 2 Longitudinal and transversal relaxivities of Gd- and Ho-NPs as function of their size at 7 T and 25 °C

Material	Diameter (nm)	r_1 ($\text{s}^{-1} \text{mM}^{-1}$)	r_2 ($\text{s}^{-1} \text{mM}^{-1}$)	Preparation method ^a	r_2/r_1	Ref.
Gd ₂ O ₃	2.2	8.8	11.4	PL	1.3	20
Gd ₂ O ₃	3.8	8.8	28.8	PL	3.3	20
Gd ₂ O ₃	4.6	4.4	28.9	PL	6.6	20
Gd ₂ O ₃	7.0	3.1	30.0	ME	9.7	^b
Gd ₂ O ₂ SO ₄	12.0	1.0	35.7	ME	35.7	^b
Gd ₂ O ₂ S	25.0	0.2	55.0	ST	275.0	17
Ho ₂ O ₃	12.0	0.2	31.1	ME	155.5	^b
Ho ₂ O ₂ SO ₄	18.0	0.4	43.9	ME	109.8	^b
Ho ₂ (CO ₃) ₃	40.0	—	105.0	MW	—	16

^a PL = polyol; ME = miniemulsion; ST = solvothermal; MW = microwave. ^b This work.

Besides the size and chemical composition, several experimental parameters, such as temperature, pH, strength of the applied magnetic field *etc.* determine the r_1 and r_2 of NPs. Therefore, the relaxivities of even similar types of NPs reported in the literature often cannot be compared directly. Taking this into account, in Table 2 a small selection of literature data is compiled to demonstrate the consistency of the relaxivities of Gd- and Ho-NPs obtained by different preparation methods, and measured under the same conditions (7 T, 25 °C). The data clearly show a decrease of r_1 for Gd-NPs upon increasing their particle size as the consequence of the decrease in the surface-to-volume ratio for oxides as well as for oxysulfates and oxysulfides.

Ho-containing NPs show a strong size-dependence of r_2 . The chemical composition of NPs seems to be of minor importance, as the values for oxide, oxysulfate and carbonate follow the same linear increase of r_2 upon increase in the size. Finally, the relaxivities of Gd- and Ho-NPs obtained from the miniemulsion method fit perfectly in the r_n vs. diameter trend and are in

agreement with those previously reported for analogous Ln-containing NPs.⁴¹

Conclusions

We have demonstrated a facile miniemulsion technique and subsequent thermal decomposition that can be advantageously applied for the preparation of Ln-containing NPs. Such methodology is particularly useful for the preparation of NPs in the size range of 5 to 40 nm as it is far less time-consuming, more reproducible and results in a higher productivity than the polyol strategies suitable for ultrasmall NPs. Therefore, the present miniemulsion method is a valid and convenient alternative strategy with attractive advantages for the synthesis of Ln-NPs.

The formation of nanodroplets under emulsion conditions is highly influenced by the selection of the surfactant and the nature of the dispersed and continuous phases. In the present study, the interfacial activity and the type of surfactant revealed to be the key parameters determining not only the size of the nanodroplets and hence the final solid particles,





but also their chemical composition. In particular, depending on the surfactant used, the main products are either lanthanide oxysulfates or oxides. By adjusting these parameters, spherical NPs with a defined diameter can be readily obtained through further thermal decomposition. Both types of particles exhibited the relaxivities that are in accordance with the values reported in the literature. The r_2 relaxivities measured on smaller Ho-NPs (<20 nm) confirm the linear size-dependence reported previously for larger Ho-NPs (>40 nm).¹⁶ The MR images indicate a good T_2 -weighted contrast, and hence, suitability of these NPs prepared by the mini-emulsion method as T_2 CAs after a proper surface functionalization. On the contrary, the r_1 relaxivities of Gd-containing NPs decrease with the particle size, and as the result of T_2 -effect becoming dominant, the brightness of the T_1 -weighted MR images acquired with bigger Gd-NPs (>7 nm) is lower. These observations demonstrate the importance of size-control for the applications of Ln-based nanoparticles as MRI CAs.

Considering the similarity of chemical properties of the f-elements, the present technique can be predictably expanded for the preparation of NPs of the whole series of lanthanides to be exploited in a variety of fields such as catalysis (oxysulfates for oxygen storage/release),² optics (oxides with luminescence properties),¹ and diagnostics (paramagnetic nanoparticles).⁴³

Acknowledgements

This research was performed in the framework of the EU COST Action TD1004, "Theranostics Imaging and Therapy: an Action to Develop Novel Nanosized Systems for Imaging-Guided Drug Delivery" and supported by China Scholarship Council (W. Z.) and the Netherlands Organization for Scientific Research (K. D. and J. M., NWO Veni grant-722.012.009). The authors thank Ms. Wan Gengping (Hainan University, China) for TEM and Mr Ben Norder (Delft University of Technology) for the XRD measurements.

Notes and references

- 1 S. V. Eliseeva and J.-C. G. Bunzli, *Chem. Soc. Rev.*, 2010, **39**, 189–227.
- 2 M. Machida, K. Kawamura, K. Ito and K. Ikeue, *Chem. Mater.*, 2005, **17**, 1487–1492.
- 3 S. Comby, E. M. Surender, O. Kotova, L. K. Truman, J. K. Molloy and T. Gunnlaugsson, *Inorg. Chem.*, 2013, **53**, 1867–1879.
- 4 Y. Liu and N. Zhang, *Biomaterials*, 2012, **33**, 5363–5375.
- 5 K. Nicolay, G. Strijkers and H. Grüll, in *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, John Wiley & Sons, Ltd, 2013, pp. 449–487.
- 6 J. A. Peters and K. Djanashvili, *Eur. J. Inorg. Chem.*, 2012, **2012**, 1961–1974.
- 7 M. A. Bruckman, X. Yu and N. F. Steinmetz, *Nanotechnology*, 2013, **24**, 20.
- 8 W. Lin, Y. Hou, Y. Lu, A. I. Abdelrahman, P. Cao, G. Zhao, L. Tong, J. Qian, V. Baranov, M. Nitz and M. A. Winnik, *Langmuir*, 2014, **30**, 3142–3153.
- 9 T. Kihara, Y. Zhang, Y. Hu, Q. Mao, Y. Tang and J. Miyake, *J. Biosci. Bioeng.*, 2011, **111**, 725–730.
- 10 S.-D. Li and L. Huang, *Mol. Pharm.*, 2008, **5**, 496–504.
- 11 F. Lu, S.-H. Wu, Y. Hung and C.-Y. Mou, *Small*, 2009, **5**, 1408–1413.
- 12 C. He, Y. Hu, L. Yin, C. Tang and C. Yin, *Biomaterials*, 2010, **31**, 3657–3666.
- 13 G. Sonavane, K. Tomoda and K. Makino, *Colloids Surf., B*, 2008, **66**, 274–280.
- 14 N. Lu, X. Song and J. Zhang, *Mater. Lett.*, 2009, **63**, 1089–1092.
- 15 J. A. Nelson, L. H. Bennett and M. J. Wagner, *J. Am. Chem. Soc.*, 2002, **124**, 2979–2983.
- 16 F. Mayer, J. A. Peters and K. Djanashvili, *Chem.-Eur. J.*, 2012, **18**, 8004–8007.
- 17 S. A. Osseni, S. Lechevallier, M. Verelst, P. Perriat, J. Dexpert-Ghys, D. Neumeyer, R. Garcia, F. Mayer, K. Djanashvili, J. A. Peters, E. Magdeleine, H. Gros-Dagnac, P. Celsis and R. Mauricot, *Nanoscale*, 2014, **6**, 555–564.
- 18 C. Truillet, F. Lux, J. Moreau, M. Four, L. Sancey, S. Chevreux, G. Boeuf, P. Perriat, C. Frochot, R. Antoine, P. Dugourd, C. Portefaix, C. Hoeffel, M. Barberi-Heyob, C. Terryn, L. van Gulick, G. Lemercier and O. Tillement, *Dalton Trans.*, 2013, 12410–12420.
- 19 B. Zhang, H. Jin, Y. Li, B. Chen, S. Liu and D. Shi, *J. Mater. Chem.*, 2012, **22**, 14494–14501.
- 20 J.-L. Bridot, A.-C. Faure, S. Laurent, C. Rivière, C. Billotey, B. Hiba, M. Janier, V. Josserand, J.-L. Coll, L. Vander Elst, R. Muller, S. Roux, P. Perriat and O. Tillement, *J. Am. Chem. Soc.*, 2007, **129**, 5076–5084.
- 21 N. Abrikossova, C. Skoglund, M. Ahrén, T. Bengtsson and K. Uvdal, *Nanotechnology*, 2012, **23**, 275101.
- 22 G. Chen, H. Qiu, P. N. Prasad and X. Chen, *Chem. Rev.*, 2014, **114**, 5161–5214.
- 23 J. G. Assouline and S. K. Sweeney, WO Pat., WO2014035620A1, 2014.
- 24 N. Luo, X. Tian, C. Yang, J. Xiao, W. Hu, D. Chen and L. Li, *Phys. Chem. Chem. Phys.*, 2013, **15**, 12235–12240.
- 25 K. Landfester, in *Colloid Chemistry II*, ed. M. Antonietti, Springer, Berlin, Heidelberg, 2003, vol. 227, ch. 4, pp. 75–123.
- 26 T. G. Mason, J. N. Wilking, K. Meleson, C. B. Chang and S. M. Graves, *J. Phys.: Condens. Matter*, 2006, **18**, R635.
- 27 S. B. Barrios, J. F. Petry, C. K. Weiss, C. L. Petzhold and K. Landfester, *J. Appl. Polym. Sci.*, 2014, **131**, 40569.
- 28 E. Schreiber, U. Ziener, A. Manzke, A. Plettl, P. Ziemann and K. Landfester, *Chem. Mater.*, 2009, **21**, 1750–1760.
- 29 Y. Yang, E. S. Daniels and A. Klein, *J. Appl. Polym. Sci.*, 2015, **132**, 41933–41940.
- 30 K. Landfester, N. Bechthold, F. Tiarks and M. Antonietti, *Macromolecules*, 1999, **32**, 5222–5228.
- 31 D. M. Corsi, C. Platas-Iglesias, H. v. Bekkum and J. A. Peters, *Magn. Reson. Chem.*, 2001, **39**, 723–726.
- 32 M. Antonietti and K. Landfester, *Prog. Polym. Sci.*, 2002, **27**, 689–757.

33 L. L. Hecht, C. Wagner, K. Landfester and H. P. Schuchmann, *Langmuir*, 2011, **27**, 2279–2285.

34 A. H. Demond and A. S. Lindner, *Environ. Sci. Technol.*, 1993, **27**, 2318–2331.

35 J. Zhang, Z. Chen, Z. Wang, W. Zhang and N. Ming, *Mater. Lett.*, 2003, **57**, 4466–4470.

36 A. K. Chesters, *Chem. Eng. Res. Des.*, 1991, **69**, 259–270.

37 M. Machida, K. Kawamura, T. Kawano, D. Zhang and K. Ikeue, *J. Mater. Chem.*, 2006, **16**, 3084–3090.

38 D. Zhang, F. Yoshioka, K. Ikeue and M. Machida, *Chem. Mater.*, 2008, **20**, 6697–6703.

39 M. Machida, T. Kawano, M. Eto, D. Zhang and K. Ikeue, *Chem. Mater.*, 2007, **19**, 954–960.

40 N. T. McDevitt and W. L. Baun, *Spectrochim. Acta*, 1964, **20**, 799–808.

41 T. J. Kim, K. S. Chae, Y. Chang and G. H. Lee, *Curr. Top. Med. Chem.*, 2013, **13**, 422–433.

42 É. Tóth, L. Helm and A. Merbach, in *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, John Wiley & Sons, Ltd, 2013, pp. 25–81.

43 M. Norek and J. A. Peters, *Prog. Nucl. Magn. Reson. Spectrosc.*, 2011, **59**, 64–82.

44 M. Norek, E. Kampert, U. Zeitler and J. A. Peters, *J. Am. Chem. Soc.*, 2008, **130**, 5335–5340.

