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Assigning regioisomeric or diastereoisomeric relations of problematic trisubstituted double-bonds through heteronuclear 2D selective *J*-resolved NMR spectroscopy†

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Although one of the first 2D NMR methods, but so far neglected, selective *J*-resolved NMR spectroscopy offers a unique opportunity to help organic chemists in structure elucidation, avoiding natural and non-natural product misassignments. This NMR method indeed allowed us to unambiguously and simply assign (natural) product structures exhibiting trisubstituted unsaturations. For example, as demonstrated here, the isomeric aurone, flavone, coumarin and isocoumarin structures could easily be distinguished; regioisomer assignments in furans could be solved, as well as isomerisms in compounds containing trisubstituted double-bonds (aurones, lactones, divinyl ketones).

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Introduction

In natural product research, structural and/or stereochemical assignments are obviously the key step, providing the foundation for a whole set of disciplines, from synthesis to biology, biochemistry or ecology, with often the production of new therapeutic agents as the most practical consequences. Despite the current sophisticated techniques available, assignments often prove delicate and sometimes cumbersome. The number of natural product structural misassignments reported each year is surprisingly high (Fig. 1).¹ These numbers are probably underestimated, since those encountered misassignments are frequent and only revealed through total synthesis enterprises.²

Furthermore, the efforts and associated costs required to correct structures and appropriately reassign them could be quite significant and higher than those associated with the initial structure, especially in total synthesis endeavors.³ Besides the increased enrolments for total syntheses, wrong assignments might have dramatic effects due to the different biological activities the compounds could exhibit.

What was observed for natural products is also true in any synthetic efforts addressing compounds in which stereo- and/or regioisomers could be obtained, especially when only one is formed. In this area, the lack of re-synthesis minimizes the

discovery of misassignments. Therefore, it is clear that, since the number of incorrectly determined structures of new products, natural or not, is quite large, reducing errors is clearly an important and useful challenge. Several routes are currently explored, among which are computer-assisted structure elucidation (CASE) method,⁴ NMR shift calculations combined with probability analyses⁵ or artificial neural network pattern recognition,⁶ combination of calculations with various spectroscopies, notably circular dichroism,⁷ and even atomic force microscopy imaging at atomic resolution.⁸ Despite these recent new developments, structure elucidation of new compounds, including natural products, heavily relies on two-dimensional NMR experiments. Correlation spectroscopy (COSY) and heteronuclear multiple and single bond correlation (HMBC/HSQC) detect interactions (couplings) of protons, respectively, with adjacent protons (from 2 to 4 bonds) or with neighbor carbon atoms (from 2 to 3 bonds). Both techniques provide connectivity between atoms. Stereochemistry and conformation of molecules are usually deduced from two-dimensional nuclear Overhauser enhancement spectroscopy (NOESY). Based on spin relaxation processes *via* dipolar coupling interacting through space, this method is able to detect protons that are in close proximity.

Facing various structural problems either in the total synthesis of natural products⁹ or in the development of new methods,¹⁰ we recently applied a neglected but simple NMR method, which successfully helped us to solve connectivity and/or stereochemistry problems. Being very efficient in our hands, we share here our results gained by applying the selective *J*-resolved NMR method during the process of structure elucidation with some typical examples.

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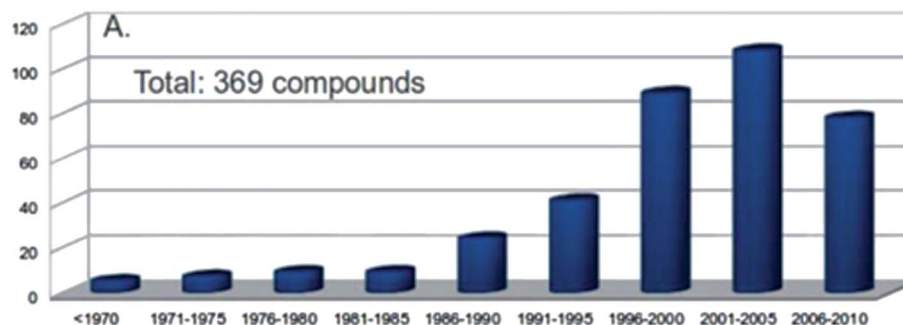


Fig. 1 Numbers of known natural product misassignments per 5 year period. From ref. 1.

Results and discussion

The (selective) *J*-resolved NMR method

Introduced in 1976 by Ernst *et al.*,¹¹ the 2D *J*-resolved NMR experiments (JRES) are used to separate chemical shifts and *J*-couplings into two independent dimensions.^{12,13} The spin coupling may be homocoupling, usually between protons, or heterocoupling, essentially between ¹H and ¹³C, although ¹H and ³¹P have also been reported.¹⁴ The 2D spectra exhibit in one dimension the equivalent of a broad range decoupled proton or carbon spectrum and in the other the coupling, either ¹H–¹H or ¹³C–¹H. Although one of the first 2D NMR experiments, JRES has been supplanted by the more recent COSY and HMBC methods in chemistry. These NMR techniques could also provide information about such coupling constants, but in a less practical way (see below).

Interestingly, the JRES method is currently re-gaining interest for the study of complex mixtures,¹⁵ mostly in biology and medicine as a tool for metabolic screening¹⁶ and metabolomics¹⁷ as well as for biomarker identification, especially for cancer monitoring.¹⁸

Stereochemical double-bond assignment is still a problem for the trisubstituted ones,¹⁹ unfortunately quite common in natural products. Classically solved for disubstituted double-bonds with the *E* or *Z* ³*J* coupling constants (¹H NMR), the case of double-bonds carrying a single hydrogen is far from obvious. Coupling between vinylic and allylic hydrogens (⁴*J*) can be measured, but the corresponding *E* and *Z* values being low and close to each other do not allow unambiguous assignments.²⁰ Due to the parallelism between ¹H–¹H and ¹³C–¹H coupling constants, the *E* or *Z* geometry could also be reflected by the ³*J* or ²*J* (¹³C–¹H) coupling constants.²¹ The usual way to measure them is to record ¹³C coupled ¹H NMR.^{22,23} However, this technique leads to very complex spectra, due to overlapping of signals and to the complexity of multiplets, which may require computational method to solve out. Furthermore, rather long measurement times are needed and typically, signal-to-noise ratio is poor for such spectra.

Some of these aspects could also occur for the classic JRES method, but to a lower extent as all ^{*n*}*J* (¹³C–¹H) coupling constants are expressed in 2D. However, the heteronuclear selective *J*-resolved NMR experiment (SELJRES)^{12b} elegantly solves this problem by inverting the resonance of the considered

proton allowing to remove from the 2D map all other coupling constants excepted those of the selectively inverted proton:²⁴ the required coupling can thus be easily available in one of the two dimensions.

Such stereochemical double-bond problems are often encountered in various families of natural products or in synthesis of unsaturated compounds, but only four typical examples will be detailed here.

Regioisomer assignment of flavonoids

Among flavonoids,²⁵ coumarins, isocoumarins, aurones, and flavones constitute a sub-family contributing to the pigmentation of flowers and fruits, but also exhibiting interesting pharmaceutical activities.²⁶ These four subclasses are structurally related, each one being isomeric to the others around a trisubstituted double-bond (Scheme 1). Due to their similarity, the structural elucidation of these natural products remains a challenge, and already led to numerous misassignments, from which only a few were detected and induced structural revision.^{9a,27}

Facing such problem, we – as others – relied on total synthesis of each possible isomer,^{9a} but this gave us the opportunity to apply heteronuclear SELJRES and to find a useful and quick distinction between these isomers. Indeed, the couplings between the vinylic proton and the carbonyl carbon (red C in Scheme 1), the vinylic carbon (blue C) and the *ortho* carbon on the phenyl substituent (green C) in each series proved typical.²⁸

Coumarin 1 showed a weak ²*J*_{Cvinylic–H} coupling constant of 1.1 Hz compared to the three other isomers 2–4 (²*J*_{Cvinylic–H} = 5.1 ± 0.4 Hz), probably reflecting the lack of oxygenated substituent on the double-bond. Isocoumarin 2 could easily be distinguished from the three other possible isomers by the lack of coupling between the vinylic proton and the carbonyl carbon. Flavone 3 and aurone 4 usually cannot be distinguished between themselves by their ¹H NMR spectra due to a very similar shift of their vinylic proton (6.8–6.9 ppm) and other chemical shifts. However, they could be unambiguously distinguished by the presence or not of a coupling between the vinylic proton and the *ortho* carbon of the aromatic substituent: aurone 4 exhibits a ³*J* coupling (6.1 Hz) while flavone 3 can only have a ⁴*J* coupling, almost not detectable (Scheme 1).



	coumarin 1	isocoumarin 2	flavone 3	aurone 4	
$J_{C(O)-H}$ (Hz) =	4.1	.. ^a	2.2	3.6	} coupling constants
$^2J_{C-H}$ (Hz) =	1.1	5.3	5.5	4.7	
$J_{C(Ar)-H}$ (Hz) =	0.9	.. ^a	.. ^a	6.1	

^aCoupling < 0.5 Hz

Scheme 1 Relation structure-SELJRES coupling constants for flavonoid derivatives.

Regioisomer assignment of trisubstituted furans

Furans constitute an important class of aromatic compounds, found in many natural products.²⁹ Therefore, a large number of synthetic methods have been developed to build such five membered rings.³⁰ Our group recently contributed to this area, achieving mild and convenient access to trisubstituted furans.³¹ During this study, as for furan natural product structural assignments, the regioselectivity has to be determined.

With a single proton, such isomeric furans may be difficult to distinguish. When substituents in positions 2 and 5 do not carry hydrogen, there is no possibility to measure the $^3J(^1H-^1H)$ coupling with the furanic proton. Since the situation of the furanic proton can be compared to those described in the previous section, we envisaged to solve this question by applying the SELJRES method. Indeed, the single proton should exhibit 2J coupling with the furanic carbon (red or blue C in Scheme 2), reflecting the proximity between the two atoms.

To look at this hypothesis, five different trisubstituted furans **5a-e** were submitted to JRES by selecting the frequency of the furanic proton for acquisition. The analysis showed a 2J coupling constant around 7 to 8 Hz between the furanic proton and its adjacent β -carbon (red or blue C on Scheme 2). Interestingly, no 3J coupling was expressed with the γ -furanic carbon. This combination thus allowed for the unambiguous determination of the substitution pattern of furans. Furthermore, the analysis can be done on a mixture of two isomers as it was shown with **5a** and **5b**.

Diastereomer assignment of aurones and alkylidenelactones

As for the previous examples, the *E* or *Z* stereochemical assignment of trisubstituted double-bonds may not be

obvious. This could also be true for methylenic systems. Aurones are typical of the former case, and our synthetic endeavor³¹ again provided us the opportunity to look at the JRES potential. Naturally occurring aurones usually exhibit a *Z* configuration of the double-bond, but such type of compounds can isomerize under irradiation or on silica gel.³² Alkylidene lactones also offer useful examples,^{9a,10b} with the methylene lactones being typical of the latter case. Both allowed interesting comparisons.

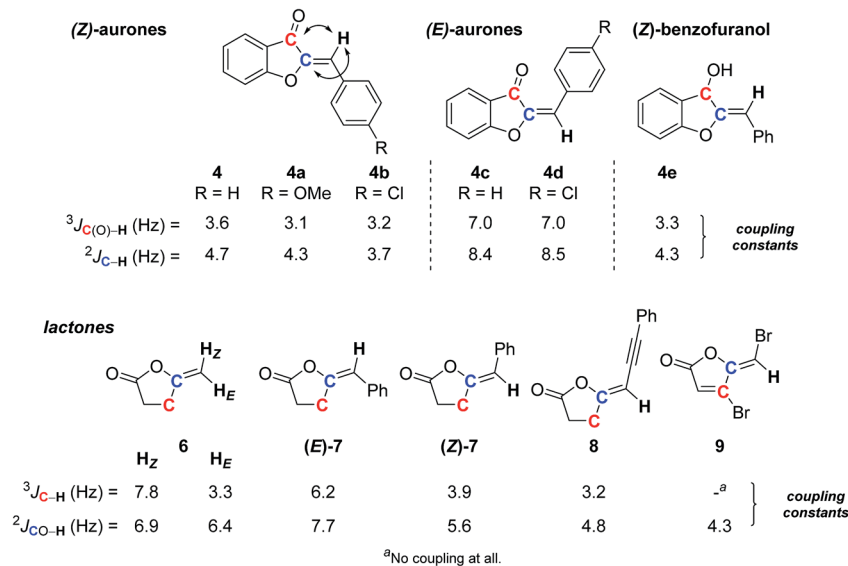
As before, SELJRES allowed us to get 2J or $^3J(^{13}C-^1H)$ coupling constants, from which those between the vinylic proton and the carbonyl or vinylic carbons simply proved distinctive for each stereoisomer. The data collected for *E* or *Z* aurones **4** and **4a-d** and related compound **4e** clearly showed a correlation between such coupling constants and the stereochemistry of the double-bond (Scheme 3). The *Z* always exhibited 3J and/or $^2J(^{13}C-^1H)$ values of 3–4 Hz, while the *E* isomer exhibited values of 7–8 Hz. Such clear-cut differences could obviously be fruitful in assigning stereochemistry and would have predictive value. This was confirmed with the alkylidene lactone series, the stereochemistry of which being already assigned by other techniques. Indeed, even in the simple methylene lactone **6**, the same set of values was observed for the *E* and *Z* protons for the $^3J(^{13}C-^1H)$ coupling (3.3 and 7.8 Hz, respectively) (Scheme 3). In other alkylidene lactones (*Z*)-**7** and **8**, similar trends could be observed with $^3J(^{13}C-^1H)$ values of 3.5 ± 0.4 Hz noticed in *Z* isomers and 6.2 Hz for the *E* isomer (*E*)-**7**. In some specific cases, no 3J value was detected as for the bromomethylene furanone **9**, structurally very close to the fimbrolide natural product family.³³ However, a 2J coupling constant of 4.3 Hz was expressed in SELJRES, which could be

	5a	5b	5c	5d	5e	
$J_{C(Alkyl)-H}$ (Hz) =	.. ^a	7.7 ^b	.. ^a	7.9	7.8	} coupling constants
$J_{C(Ar)-H}$ (Hz) =	7.6	.. ^a	7.6	.. ^a	.. ^a	

^aNo coupling at all. ^bDetermined on a mixture 5:1 of **5a** and **5b**

Scheme 2 Relation structure-SELJRES coupling constants for furan derivatives.





Scheme 3 Simple *E/Z* assignment of aurones and lactone derivatives by SELJRES spectral analysis.

attributed to the *Z* isomer by comparison with lactones (*E*)-7, (*Z*)-7 and 8.

It is worth mentioning that correlations from ^1H -coupled ^{13}C spectra with double-bond diastereoisomers have already been observed and compiled from around forty examples in the seventies (Table 1).²¹ The listed values are surprisingly large and similar to classical ^1H - ^1H coupling constants, the *E* 3J (^{13}C - ^1H) coupling constants being always larger than the *Z* ones, but with a strong influence upon hybridization of the selected carbon. Although the general tendency is the same, with larger coupling constants for *E* relative to *Z* proton, the 3J SELJRES values we collected are far different from those already reported: between 3–4 Hz vs. 4–10 Hz for *Z* and 7–8 Hz vs. 9–17 Hz for *E* (Scheme 3 vs. Table 1) and, in our study, no influence of the hybridization has been noticed (see compound 4 vs. 4e). More surprisingly, all our measured SELJRES values should correspond to *cis* proton/*Z* systems according to Table 1 data.

Diastereomer assignment of divinyl ketones

For a program devoted to gold catalysis and to the synthesis of cyclopentenone derivatives *via* the Nazarov reaction,³⁴ we obtained various divinyl ketones,³⁵ for which we had to

assign the double-bond stereochemistry. Indeed, the reaction we developed led to divinyl ketones in which one double-bond stereochemistry was not defined by the synthetic pathway. In a first attempt, 2D NOESY was used, leading to some conclusions. However, based on the absence of correlation and due to the flexibility of such structures, assignments were suspicious. Fortunately, we were able to obtain crystal structures allowing us to unambiguously determine the stereochemistry. This helped us to look again at the correlations between ^{13}C - ^1H SELJRES values and double-bond stereochemistry. Among the divinyl ketones we prepared, different examples were selected and submitted to SELJRES NMR experiments.

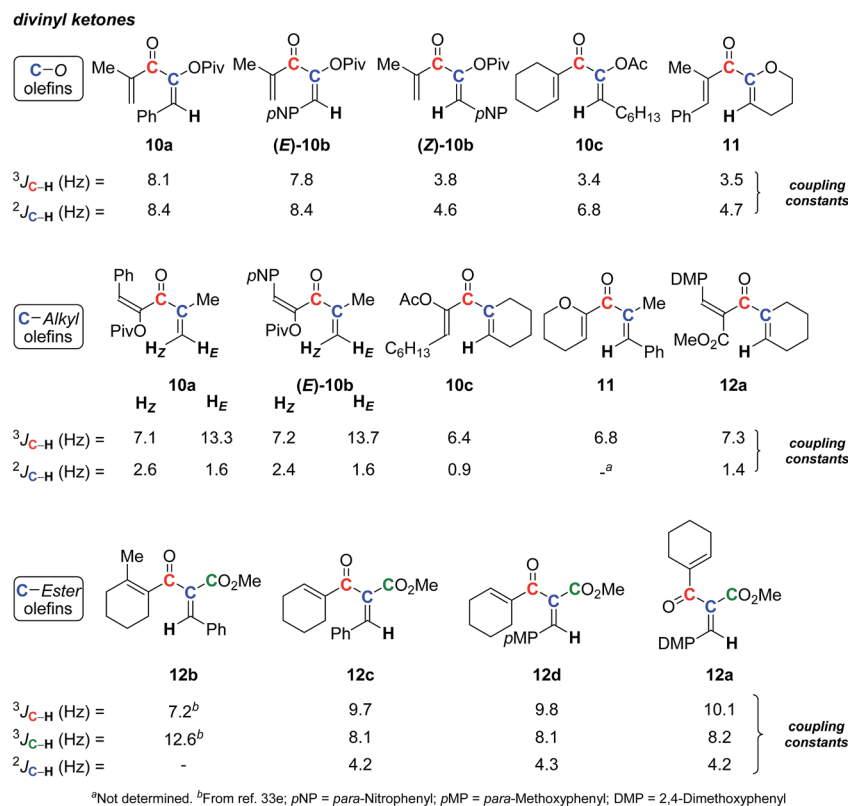
A series of *O*-substituted divinyl ketones (**10a–c** and **11**) was first evaluated, including one (**10a**) with X-ray crystallography structure (see ESI[†]) and one ((*E*)-**10b** and (*Z*)-**10b**) with both diastereomers. Their 3J and 2J (^{13}C - ^1H) coupling constants, respectively, between the vinylic proton and the carbonyl carbon or the adjacent sp^2 carbon were extracted (Scheme 4). Values similar to those already gained (see preceding section and Scheme 3) were obtained, with 3J and 2J (^{13}C - ^1H) coupling constants of 8 ± 0.2 Hz for *E* isomers (**10a** and (*E*)-**10b**) and 3.6 ± 0.2 Hz for *Z* compounds ((*Z*)-**10b**, **10c** and **11**). These two sets of constants clearly depend on the geometry of the system and can easily be exploited to unambiguously assign double-bond geometry.

We also looked at the coupling constants of the double-bonds having C-substituents (**10a–c**, **11**, and **12a**). Once again, the geometry of each diastereomer could be easily determined by SELJRES with 3J ($^{13}\text{C}(\text{O})$ - ^1H) values around 13 Hz for *trans* isomers (H_E) and between 6.4 and 7.3 Hz for *cis* isomers (H_Z). However, in the case of C-substituted divinyl ketones, the 2J (^{13}C - ^1H) coupling constants did not follow the tendency of 3J (^{13}C - ^1H) values: only weak values were measured (1.8 ± 0.8 Hz) without direct correlation with the *E* and *Z* nature of olefins.

Table 1 Range of ^{13}C - ^1H 3J coupling constants values depending on carbon hybridization from ref. 21

C Hybridization	3J (C-H) coupling constant		
	H_Z (Hz)	H_E (Hz)	
	sp (CN, CC)	8.2–9.0	13.8–15.0
	sp^2 (CO)	4.3–10.0	9.5–16.9
	sp^3 (CH_2X)	5.7–8.0	7.7–11.0





Scheme 4 3J and 2J ($^{13}C-^1H$) SELJRES values of variously substituted divinyl ketones 10–12.

Being well represented in the literature due to their good reactivity in the Nazarov reaction,³⁶ divinyl ketones carrying an ester group (12a–d) were also investigated. In our case, only *E* isomers were available. SELJRES spectra showed that their 3J ($^{13}C(O)-^1H$) (*trans* values) and 3J ($^{13}CO_2Me-^1H$) (*cis* values) coupling constants lay, respectively, between 9 to 10 Hz and around 8 Hz for compounds 12a and 12c–d. Working on the Nazarov reaction, Frontier *et al.* have also been interested in assigning the stereochemistry of divinyl ketones, and they have reported the 3J ($^{13}C-^1H$) extracted from ^{13}C non-decoupled 1H NMR spectra, for *Z* and *E* isomers analogs.^{36e} In this report, the coupling constants for *E* isomers correlated perfectly with the range of values obtained for 12a and 12c–d (9–10 Hz). However, the 3J ($^{13}CO-^1H$) (*cis* values, 4.6–7.2 Hz) and 3J ($^{13}CO_2Me-^1H$) (*trans* values, 10.7–12.8 Hz) for *Z* isomers are far different, such as in 12b, which confirmed the *Z* nature of the considered olefin.

Conclusion

In this work, we have demonstrated that SELJRES NMR spectra gave direct and unambiguous access to the regioisomerism of various types of molecules, such as flavonoids and furans. SELJRES NMR thus offers a simple and easy method to rapidly determine the *E* and *Z* diastereochemistry of trisubstituted double-bonds in aurone, alkylidene lactone or divinyl ketone derivatives. Indeed, the 3J ($^{13}C-^1H$) coupling constants extracted from SELJRES are always larger for the *E* isomers, between 7–13 Hz, than for the *Z* isomers (3–8 Hz) depending on the substrate nature.

Experimental section

General informations

Proton (1H NMR) and carbon (^{13}C NMR) nuclear magnetic resonance spectra were recorded on a 500- or 600 MHz Bruker Avance II spectrometer equipped with a 5 mm DCH dual cryoprobe, with a z -gradient and operating at 500.130 MHz for 1H and 125.758 MHz for ^{13}C . The solvent peak was used as reference value, for 1H NMR: $CHCl_3 = 7.26$ ppm or benzene-*d* = 7.16 ppm and for ^{13}C NMR: $CHCl_3 = 77.16$ ppm or benzene-*d* = 128.06 ppm. For 1H NMR, data are presented as follows: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, b = broad), coupling constants (J /Hz), integration, and attribution. For ^{13}C NMR, data are presented as follows: chemical shift (ppm) and attribution. For $^{13}C\{-^1H\}$ selective J -resolved NMR, data are presented as follows: (chosen proton), ^{13}C chemical shift (ppm) and coupling constant (J /Hz) or folded 1J ($^{13}C-^1H$), actually the acquisition method prevented the direct detection of the 1J ($^{13}C-^1H$) constant that is folded. Remark: despite all discussions about trisubstituted double-bonds, during the characterization it was observed that, for a chosen proton, sometimes the 2J ($^{13}C-^1H$) is smaller than its 3J ($^{13}C-^1H$).³⁷ Assignments were determined either on the basis of unambiguous chemical shifts or coupling patterns, COSY, HSQC, HMBC, and NOESY experiments, to fully interpret spectra for related compounds. The pulse sequence used for measuring nJ ($^{13}C-^1H$) coupling constants is similar to the *seljresqfsp* sequence of the Bruker



library. Experiments were acquired at 293 K with a relaxation delay of 3 s. Acquisition parameters were adjusted when necessary, but typically spectral windows were set to 25 kHz (200 ppm) for ^{13}C and 50 Hz for ^1H ($^{13}\text{C}-^1\text{H}$). Data size was at least 16k points in the direct dimension and 32 points in the indirect dimension. Broadband decoupling during acquisition was achieved by a composite pulse sequence Waltz16. Refocusing of ^{13}C magnetization during evolution period was performed with an adiabatic shape pulse Crp60comp.4 of 2 ms. Simultaneously, a selective 180° pulse focused on the chosen proton frequency was achieved with a refocusing band-selective pulse (REBURP) of 80 ms. Melting points were recorded on a Melting Point Apparatus SMP3 and are uncorrected. Infrared spectra were recorded neat with Hewlett-Packard spectrometer. Wavelengths of maximum absorbance (ν_{max}) are quoted in wave numbers (cm^{-1}). High resolution mass spectra (HR-MS) data were recorded on a microTOF spectrometer equipped with orthogonal electrospray interface (ESI). The parent ions $[\text{M} + \text{Na}]^+ [\text{M} + \text{H}]^+$ is quoted. Analytical thin-layer chromatography (TLC) was carried out over silica gel 60 F254 plates or basic alumina (63–200 μm) with visualization by ultraviolet light, cerium ammonium molybdate (CAM) or potassium permanganate dip (KMnO₄). Flash-column chromatography was carried out using silica gel 60 (40–63 μm) and the procedure included the subsequent evaporation of solvents *in vacuo*. Solvents were purified using standard means. 1,2-Dichloroethane (DCE), benzene and triethylamine (NEt₃) were distilled from CaH₂; pyridine (py) was distilled from potassium carbonate; diisopropylamine (DIPA) and *N,N*-diisopropylethylamine (DIPEA) were distilled from KOH; all under an argon atmosphere. Tetrahydrofuran (THF), acetonitrile (CH₃CN) and dichloromethane (DCM) were dried using Glasstechnology GT S100 device. AuCl (Premion grade, 99.99%) and NaAuCl₄·2H₂O (Premion grade, 99.99%) were purchased from Alfa Aesar. AgSbF₆ (98%) and PdCl₂(dppf)·DCM were purchased from STREM Chemicals. IPrAuCl, PdCl₂(PPh₃)₂ (98%), HNTf₂ and CuI (98%) were purchased from Aldrich. AgNTf₂ was prepared from HNTf₂ (Aldrich) and Ag₂CO₃.³⁸ All phosphinegold(i) chloride precatalysts were prepared by reduction of NaAuCl₄·2H₂O with thiodiethanol and subsequent addition of the appropriate phosphine.³⁹ PPh₃AuNTf₂ catalyst was prepared from the corresponding triphenylphosphinegold(i) chloride and AgNTf₂.⁴⁰ All other chemicals were used as received.

4-Phenyl-2H-chromen-2-one (1)

4-Phenyl-2H-chromen-2-one (1) was obtained in 91% yield over 2 steps from chromane-2,4-dione, using reported procedures (see ESI†).⁴¹ White solid; TLC *R_f* 0.23 (cyclohexane/EtOAc 20%); ^1H NMR (500 MHz, CDCl₃) δ 6.38 (s, 1H, H₃), 7.23 (dd, *J* = 7.9, 7.1 Hz, 1H, H₆), 7.41 (d, *J* = 8.1 Hz, 1H, H₈), 7.44–7.47 (m, 2H, H₂), 7.50 (d, *J* = 7.9 Hz, 1H, H₅), 7.52–7.55 (m, 3H, H₃/4), 7.55 (dd, *J* = 8.1, 7.1 Hz, 1H, H₇); ^{13}C NMR (125.8 MHz, CDCl₃) δ 115.3 (C₃), 117.5 (C₈), 119.1 (C_{4a}), 124.3 (C₆), 127.1 (C₅), 128.6 (C₂), 129.0 (C₃'), 129.8 (C₄'), 132.1 (C₇), 135.3 (C₁'), 154.3 (C_{8a}), 155.8 (C₄), 160.9 (C₂); $^{13}\text{C}-\{^1\text{H}\}$ selective *J*-resolved NMR (125.8 MHz, CDCl₃, chosen proton: H₃ 6.38 ppm) δ 115.3 (folded $^1J_{\text{CH}}$),

119.1 ($^3J_{\text{CH}}$ = 7.2 Hz), 128.6 ($^4J_{\text{CH}}$ = 0.9 Hz), 135.3 ($^3J_{\text{CH}}$ = 5.0 Hz), 155.8 ($^2J_{\text{CH}}$ = 1.1 Hz), 160.9 ($^2J_{\text{CH}}$ = 4.1 Hz).

3-Phenyl-1H-isochromen-1-one (2) (commercially available)

White solid; ^1H NMR (500 MHz, CDCl₃) δ 6.96 (s, 1H, H₄), 7.41–7.53 (m, 5H, H₃/4', H₈ and H₆), 7.72 (dd, *J* = 8.0, 6.9 Hz, 1H, H₇), 7.89 (d, *J* = 8.1 Hz, 2H, H₂), 8.31 (d, *J* = 8.2 Hz, 1H, H₅); ^{13}C NMR (125.8 MHz, CDCl₃) δ 102.0 (C₄), 120.7 (C_{4a}), 125.4 (C₂), 126.1 (C₆), 128.3 (C₈), 129.0 (C₃'), 129.8 (C₅), 130.1 (C₄'), 132.1 (C₁'), 135.0 (C₇), 137.6 (C_{8a}), 153.7 (C₃), 162.5 (C₁); $^{13}\text{C}-\{^1\text{H}\}$ selective *J*-resolved NMR (125.8 MHz, CDCl₃, chosen proton: H₄ 6.96 ppm) δ 102.0 (folded $^1J_{\text{CH}}$), 120.7 ($^2J_{\text{CH}}$ = 5.4 Hz), 153.7 ($^2J_{\text{CH}}$ = 5.3 Hz).

2-Phenyl-4H-chromen-4-one (3)⁴²

2-Phenyl-4H-chromen-4-one (3)⁴² was obtained in 41% yield over 3 steps from salicylaldehyde, using reported procedures (see ESI†).^{9a,43} White solid; TLC *R_f* 0.30 (cyclohexane/EtOAc 30%); ^1H NMR (500 MHz, CDCl₃) δ 6.83 (s, 1H, H₃), 7.42 (dd, *J* = 8.1, 7.4 Hz, 1H, H₆), 7.49–7.55 (m, 3H, H₃/4'), 7.57 (d, *J* = 8.1 Hz, 1H, H₈), 7.70 (dd, *J* = 8.1, 7.4 Hz, 1H, H₇), 7.93 (d, *J* = 7.7 Hz, 2H, H₂), 8.23 (d, *J* = 8.1 Hz, 1H, H₅); ^{13}C NMR (125.8 MHz, CDCl₃) δ 107.7 (C₃), 118.2 (C₈), 124.1 (C_{4a}), 125.4 (C₆), 125.8 (C₅), 126.4 (C₂'), 129.2 (C₃'), 131.7 (C₄'), 131.9 (C₁'), 133.9 (C₇), 156.4 (C_{8a}), 163.5 (C₂), 178.6 (C₄); $^{13}\text{C}-\{^1\text{H}\}$ selective *J*-resolved NMR (125.8 MHz, CDCl₃, chosen proton: H₃ 6.83 ppm) δ 107.7 (folded $^1J_{\text{CH}}$), 124.1 ($^3J_{\text{CH}}$ = 3.8 Hz), 163.5 ($^2J_{\text{CH}}$ = 5.5 Hz), 178.6 ($^2J_{\text{CH}}$ = 2.2 Hz).

Compounds 4, 4a, 4b and 4e were prepared from salicylaldehyde, using reported procedures (see ESI†).^{9a}

(Z)-2-Benzylidenebenzofuran-3-one (4)^{9a}

(Z)-2-Benzylidenebenzofuran-3-one (4)^{9a} was obtained in 68% yield over 3 steps. Yellow solid; TLC *R_f* 0.16 (pentane/Et₂O 10%); ^1H NMR (500 MHz, CDCl₃) δ 6.89 (s, 1H, H₁'), 7.21 (dd, *J* = 7.8, 7.4 Hz, 1H, H₅), 7.32 (d, *J* = 8.3 Hz, 1H, H₇), 7.39 (t, *J* = 7.3 Hz, 1H, H₅'), 7.45 (dd, *J* = 8.1, 7.3 Hz, 2H, H₄'), 7.64 (dd, *J* = 8.3, 7.4 Hz, 1H, H₆), 7.80 (d, *J* = 7.8 Hz, 1H, H₄), 7.92 (d, *J* = 8.1 Hz, 2H, H₃); ^{13}C NMR (125.8 MHz, CDCl₃) δ 113.0 (C₇), 113.1 (C₁'), 121.7 (C_{3a}), 123.6 (C₅), 124.8 (C₄), 129.0 (C₄'), 129.995 (C₅'), 131.6 (C₃'), 132.4 (C₂'), 137.0 (C₆), 147.0 (C₂), 166.2 (C_{7a}), 184.9 (C₃); $^{13}\text{C}-\{^1\text{H}\}$ selective *J*-resolved NMR (125.8 MHz, CDCl₃, chosen proton: H₁' 6.89 ppm) δ 113.1 (folded $^1J_{\text{CH}}$), 131.6 ($^3J_{\text{CH}}$ = 6.1 Hz), 147.0 ($^2J_{\text{CH}}$ = 4.7 Hz), 184.9 ($^3J_{\text{CH}}$ = 3.6 Hz).

(Z)-2-(4-Methoxybenzylidene)benzofuran-3-one (4a)^{9a}

(Z)-2-(4-Methoxybenzylidene)benzofuran-3-one (4a)^{9a} was obtained in 17% yield over 3 steps. Yellow solid; TLC *R_f* 0.38 (cyclohexane/EtOAc 30%); ^1H NMR (500 MHz, CDCl₃) δ 3.87 (s, 3H, H_{P-OMe}), 6.89 (s, 1H, H₁'), 6.98 (d, *J* = 8.9 Hz, 2H, H₄'), 7.21 (dd, *J* = 7.9, 7.2 Hz, 1H, H₅), 7.32 (d, *J* = 8.3 Hz, 1H, H₇), 7.64 (dd, *J* = 8.3, 7.2 Hz, 1H, H₆), 7.80 (d, *J* = 7.9 Hz, 1H, H₄), 7.92 (d, *J* = 8.9 Hz, 2H, H₃); ^{13}C NMR (125.8 MHz, CDCl₃) δ 55.5 (C_{P-OMe}), 113.0 (C₇), 113.6 (C₁'), 114.6 (C₄'), 122.0 (C_{3a}), 123.4 (C₅), 124.7 (C₄), 125.1 (C₂'), 133.6 (C₃'), 136.7 (C₆), 146.0 (C₂), 161.2 (C₄'), 165.9 (C_{7a}), 184.7 (C₃); $^{13}\text{C}-\{^1\text{H}\}$ selective *J*-resolved NMR (125.8



MHz, CDCl₃, chosen proton: $H_{1'}$ 6.89 ppm) δ 113.6 (folded $^1J_{CH}$), 133.6 ($^3J = 6.2$ Hz), 146.0 ($^2J_{CH} = 4.3$ Hz), 184.7 ($^3J_{CH} = 3.1$ Hz).

(Z)-2-(4-Chlorobenzylidene)benzofuran-3-one (4b)^{9a}

(Z)-2-(4-Chlorobenzylidene)benzofuran-3-one (4b)^{9a} was obtained in 39% yield over 3 steps. Yellow solid; TLC R_f 0.38 (cyclohexane/EtOAc 30%); 1H NMR (500 MHz, CDCl₃) δ 6.82 (s, 1H, $H_{1'}$), 7.23 (dd, $J = 7.7, 7.0$ Hz, 1H, H_5), 7.33 (d, $J = 8.4$ Hz, 1H, H_7), 7.42 (d, $J = 8.9$ Hz, 2H, $H_{4'}$), 7.67 (dd, $J = 8.4, 7.0$ Hz, 1H, H_6), 7.80 (d, $J = 7.7$ Hz, 1H, H_4), 7.84 (d, $J = 8.9$ Hz, 2H, H_3); ^{13}C NMR (125.8 MHz, CDCl₃) δ 111.7 ($C_{1'}$), 113.1 (C_7), 121.6 (C_{3a}), 123.8 (C_5), 124.9 (C_4), 129.3 ($C_{4'}$), 130.9 ($C_{5'}$), 132.7 ($C_{3'}$), 136.0 ($C_{2'}$), 137.2 (C_6), 147.1 (C_2), 166.193 (C_{7a}), 184.8 (C_3); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: $H_{1'}$ 6.82 ppm) δ 111.7 (folded $^1J_{CH}$), 132.7 ($^3J = 6.1$ Hz), 147.1 ($^2J_{CH} = 3.7$ Hz), 184.8 ($^3J_{CH} = 3.2$ Hz).

(Z)-2-Benzylidene-2,3-dihydrobenzofuran-3-ol (4e)^{9a}

(Z)-2-Benzylidene-2,3-dihydrobenzofuran-3-ol (4e)^{9a} was obtained in 61% yield over 2 steps. White solid; TLC R_f 0.22 (cyclohexane/EtOAc 20%); 1H NMR (500 MHz, C₆D₆) δ 1.52 (br, 1H, HO), 5.32 (dd, $J = 5.4, 1.6$ Hz, 1H, H_3), 5.83 (d, $J = 1.6$ Hz, 1H, $H_{1'}$), 6.76 (dd, $J = 7.6, 7.3$ Hz, 1H, H_5), 6.82 (d, $J = 7.9$ Hz, 1H, H_7), 6.96 (dd, $J = 7.9, 7.6$ Hz, 1H, H_6), 7.09 (t, $J = 7.4$ Hz, 1H, $H_{5'}$), 7.18 (d, $J = 7.3$ Hz, 1H, H_4), 7.28 (dd, $J = 7.9, 7.4$ Hz, 2H, $H_{4'}$), 7.82 (d, $J = 7.9$ Hz, H_3); ^{13}C NMR (125.8 MHz, CDCl₃) δ 72.5 (C_3), 106.1 ($C_{1'}$), 110.6 (C_7), 122.9 (C_5), 125.9 (C_4), 127.0 ($C_{5'}$), 127.8 (C_{3a}), 128.8 ($C_{4'}$), 129.2 ($C_{3'}$), 130.4 (C_6), 135.4 ($C_{2'}$), 157.6 (C_2), 158.1 (C_{7a}); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: $H_{1'}$ 5.83 ppm) δ 72.5 ($^3J_{CH} = 3.3$ Hz), 106.1 (folded $^1J_{CH}$), 129.2 ($^3J_{CH} = 5.8$ Hz), 157.6 ($^2J_{CH} = 4.3$ Hz).

(E)-2-Benzylidenebenzofuran-3-one (4c)

A solution of (Z)-2-benzylidenebenzofuran-3-one (4) (93 mg, 0.42 mmol) in CDCl₃ (20 mL) was irradiated by the sunlight during 6 days. Solvent was then evaporated and the residue was purified by flash chromatography over silica gel (pentane/Et₂O 5 to 10%) to yield the desired (E)-2-benzylidenebenzofuran-3-one in 42% yield (39 mg, 0.18 mmol). Starting material was also partially recovered (29%, 27 mg, 0.12 mmol). Yellow solid; mp 178 °C; TLC R_f 0.28 (pentane/Et₂O 10%); IR (neat) ν_{max} 481, 552, 598, 666, 696, 752, 857, 884, 1023, 1037, 1085, 1144, 1186, 1239, 1297, 1323, 1459, 1475, 1495, 1596, 1656, 1712, 1727 cm⁻¹; 1H NMR (500 MHz, CDCl₃) δ 6.97 (s, 1H, $H_{1'}$), 7.18 (dd, $J = 8.2, 6.8$ Hz, 1H, H_5), 7.21 (d, $J = 8.2$ Hz, 1H, H_7), 7.39–7.48 (m, 3H, $H_{4'}/5'$), 7.63 (dd, $J = 7.5, 6.8$ Hz, 1H, H_6), 7.79 (d, $J = 7.5$ Hz, 1H, H_4), 8.17 (d, $J = 7.9$ Hz, 2H, H_3); ^{13}C NMR (125.8 MHz, CDCl₃) δ 112.7 (C_7), 122.8 ($C_{1'}$), 122.9 (C_5), 123.5 (C_{3a}), 124.8 (C_4), 128.6 ($C_{4'}$), 130.5 ($C_{5'}$), 131.0 ($C_{3'}$), 131.9 ($C_{2'}$), 136.9 (C_6), 148.2 (C_2), 165.4 (C_{7a}), 182.9 (C_3); HR-MS 245.0606 [C₁₅H₁₀O₂ + Na calcd 245.0573]; ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: $H_{1'}$ 6.97 ppm) δ 122.8 (folded $^1J_{CH}$), 131.0 ($^3J_{CH} = 6.3$ Hz), 131.9 ($^2J_{CH} = 1.7$ Hz), 148.2 ($^2J_{CH} = 8.4$ Hz), 182.9 ($^3J_{CH} = 7.0$ Hz).

(E)-2-(4-Chlorobenzylidene)benzofuran-3-one (4d)

(E)-2-(4-Chlorobenzylidene)benzofuran-3-one (4d) was obtained as a 1 : 6 mixture of 4b : 4d over 1 step from the corresponding (Z) isomer (4b), after irradiation at 350 nm in methanol for 30 h.^{27a} SELJRES NMR Analysis was directly performed on the crude mixture. 1H NMR (500 MHz, CDCl₃) δ 7.25 (s, 1H, $H_{1'}$), 7.53 (dd, $J = 7.7, 7.6$ Hz, 1H, H_5), 7.55 (d, $J = 8.2$ Hz, 1H, H_7), 7.74 (d, $J = 8.6$ Hz, 1H, $H_{4'}$), 7.98 (dd, $J = 8.2, 7.6$ Hz, 1H, H_6), 8.13 (d, $J = 7.7$ Hz, 1H, H_4), 8.47 (d, $J = 8.6$ Hz, 2H, H_3); ^{13}C NMR (125.8 MHz, CDCl₃) δ 112.7 (C_7), 121.4 ($C_{1'}$), 123.0 (C_5), 123.3 (C_{3a}), 124.8 (C_4), 128.8 ($C_{4'}$), 130.4 ($C_{2'}$), 132.3 ($C_{3'}$), 136.3 ($C_{5'}$), 137.1 (C_6), 148.4 (C_2), 165.4 (C_{7a}), 183.0 (C_3); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: $H_{1'}$ 7.25 ppm) δ 121.4 (folded $^1J_{CH}$), 130.4 ($^2J_{CH} = 1.6$ Hz), 132.3 ($^3J_{CH} = 6.4$ Hz), 148.4 ($^2J_{CH} = 8.5$ Hz), 183.0 ($^3J_{CH} = 7.0$ Hz).

Compounds 5a, 5b and 5c were prepared using reported procedures (see ESI†).³¹

2-(tert-Butyl)-5-phenylfuran-3-yl pivalate (5a)³¹

2-(tert-Butyl)-5-phenylfuran-3-yl pivalate (5a)³¹ was obtained in 43% yield over 2 steps, from 4,4-dimethylpent-1-yn-3-yl pivalate. Colorless oil; TLC R_f 0.64 (pentane/Et₂O 10%); 1H NMR (500 MHz, CDCl₃) δ 1.35 (s, 9H, $H_{3''}$), 1.37 (s, 9H, $H_{2''}$), 6.59 (s, 1H, H_4), 7.23 (t, $J = 7.3$ Hz, 1H, $H_{4''}$), 7.35 (dd, $J = 8.0, 7.3$ Hz, 2H, $H_{3''}$), 7.59 (d, $J = 8.0$ Hz, 2H, $H_{2''}$); ^{13}C NMR (125.8 MHz, CDCl₃) δ 27.3 ($C_{3''}$), 29.0 ($C_{2''}$), 33.2 ($C_{1'}$), 39.1 ($C_{2''}$), 102.7 (C_4), 123.4 ($C_{3''}$), 127.3 ($C_{4''}$), 128.7 ($C_{2''}$), 131.0 ($C_{1''}$), 134.0 (C_2), 148.9 (C_5), 149.1 (C_3), 176.4 ($C_{1''}$); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: H_4 6.59 ppm) δ 102.7 (folded $^1J_{CH}$), 131.0 ($^3J_{CH} = 1.0$ Hz), 148.9 ($^2J_{CH} = 7.6$ Hz), 149.1 ($^2J_{CH} = 4.9$ Hz).

5-(tert-Butyl)-2-phenylfuran-3-yl pivalate (5b)³¹

5-(tert-Butyl)-2-phenylfuran-3-yl pivalate (5b)³¹ was obtained in 5% yield over 2 steps, from 4,4-dimethylpent-1-yn-3-yl pivalate, and analyzed as a 5a : 5b 5.1 : 1 mixture of regioisomers. TLC R_f 0.71 (pentane/Et₂O 10%); 1H NMR (500 MHz, CDCl₃) δ 1.32 (s, 9H, $H_{2''}$), 1.39 (s, 9H, $H_{3''}$), 6.24 (s, 1H, H_4), 7.21 (t, $J = 7.3$ Hz, 1H, $H_{4'}$), 7.38 (dd, $J = 8.0, 7.3$ Hz, 2H, $H_{3'}$), 7.68 (d, $J = 8.0$ Hz, 2H, $H_{2'}$); ^{13}C NMR (125.8 MHz, CDCl₃) δ 27.4 ($C_{3''}$), 29.0 ($C_{2''}$), 33.1 ($C_{1''}$), 39.3 ($C_{2''}$), 101.2 (C_4), 123.9 ($C_{3'}$), 126.7 ($C_{4'}$), 128.6 ($C_{2'}$), 130.3 ($C_{1'}$), 135.7 (C_2), 138.4 (C_3), 161.6 (C_5), 176.0 ($C_{1''}$); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: H_4 6.24 ppm) δ 101.2 (folded $^1J_{CH}$), 138.4 ($^2J_{CH} = 5.0$ Hz), 161.6 ($^2J_{CH} = 7.7$ Hz).

2-Methyl-5-phenylfuran-3-yl pivalate (5c)⁴⁴

2-Methyl-5-phenylfuran-3-yl pivalate (5c)⁴⁴ was obtained in 22% yield over 2 steps, from but-3-yn-2-yl pivalate. White solid; TLC R_f 0.60 (pentane/Et₂O 5%); 1H NMR (500 MHz, CDCl₃) δ 1.35 (s, 9H, $H_{3''}$), 2.26 (s, 3H, $H_{1'}$), 6.64 (s, 1H, H_4), 7.23 (t, $J = 7.3$ Hz, 1H, $H_{4''}$), 7.36 (dd, $J = 7.8, 7.3$ Hz, 2H, $H_{3''}$), 7.59 (d, $J = 7.8$ Hz, 2H, $H_{2''}$); ^{13}C NMR (125.8 MHz, CDCl₃) δ 10.8 ($C_{1'}$), 27.3 ($C_{3''}$), 39.2 ($C_{2''}$), 102.1 (C_4), 123.4 ($C_{2''}$), 127.3 ($C_{4''}$), 128.7 ($C_{3''}$), 131.0 ($C_{1''}$), 135.9 (C_2), 139.8 (C_3), 150.1 (C_5), 176.2 ($C_{1''}$); ^{13}C - $\{^1H\}$ selective J -resolved NMR (125.8 MHz, CDCl₃, chosen proton: H_4



6.64 ppm) δ 102.1 (folded $^1J_{\text{CH}}$), 131.0 ($^3J_{\text{CH}} = 1.1$ Hz), 139.8 ($^2J_{\text{CH}} = 5.4$ Hz), 150.1 ($^2J_{\text{CH}} = 7.6$ Hz).

Compounds **5d** and **5e** were prepared using reported procedures (see ESI†).^{31,45}

Diethyl (5-methyl-2-phenylfuran-3-yl) phosphate (**5d**)⁴⁵

Diethyl (5-methyl-2-phenylfuran-3-yl) phosphate (**5d**)⁴⁵ was obtained in 10% yield over 5 steps, from but-1-yn-3-ol. Colorless oil; TLC R_f 0.30 (cyclohexane/EtOAc 20%); IR (neat) ν_{max} 490, 578, 659, 693, 761, 800, 905, 1011, 1147, 1277, 1445, 1630, 2913, 2985 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.33 (td, $J = 7.1$ Hz, $J_{\text{HP}} = 0.9$ Hz, 6H, $H_{2''}$), 2.32 (s, 3H, $H_{2''}$), 4.15–4.27 (m, 4H, $H_{1''}$), 6.28 (s, 1H, H_4), 7.21 (t, $J = 7.5$ Hz, 1H, $H_{4'}$), 7.37 (dd, $J = 8.6$, 7.5 Hz, 2H, $H_{3'}$), 7.73 (d, $J = 8.6$ Hz, 2H, $H_{2'}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 14.3 ($C_{2''}$), 16.2 (d, $J_{\text{CP}} = 6.9$ Hz, $C_{2''}$), 64.9 (d, $J_{\text{CP}} = 6.4$ Hz, $C_{1''}$), 103.5 (C_4), 123.6 ($C_{2'}$), 126.6 ($C_{4'}$), 128.6 ($C_{3'}$), 129.9 ($C_{1'}$), 135.6 (d, $J_{\text{CP}} = 6.9$ Hz, C_2), 138.1 (d, $J_{\text{CP}} = 6.9$ Hz, C_3), 150.0 (C_5); ^{31}P NMR (162.0 MHz, CDCl_3) δ -5.32; HR-MS 333.084 ($\text{C}_{15}\text{H}_{19}\text{O}_5\text{P} + \text{Na}$ calcd 333.086); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_4 6.28 ppm) δ 14.3 ($^3J_{\text{CH}} = 1.2$ Hz), 103.5 (folded $^1J_{\text{CH}}$), 150.0 ($^2J_{\text{CH}} = 7.9$ Hz).

Diethyl (5-pentyl-2-phenylfuran-3-yl) phosphate (**5e**)⁴⁵

Diethyl (5-pentyl-2-phenylfuran-3-yl) phosphate (**5e**)⁴⁵ was obtained in 24% yield over 5 steps, from oct-1-yn-2-ol (see ESI†). Colorless oil; TLC R_f 0.24 (cyclohexane/EtOAc 40%); ^1H NMR (600 MHz, CDCl_3) δ 0.90 (t, $J = 7.1$ Hz, 3H, $H_{5''}$), 1.33 (td, $J = 7.2$ Hz, $J_{\text{HP}} = 1.1$ Hz, 6H, $H_{2''}$), 1.33–1.37 (m, 4H, $H_{4''/3''}$), 1.64–1.70 (m, 2H, $H_{2''}$), 2.61 (t, $J = 7.7$ Hz, 2H, $H_{1''}$), 4.16–4.26 (m, 4H, $H_{1''}$), 6.29 (s, 1H, H_4), 7.20 (t, $J = 7.4$ Hz, 1H, $H_{4'}$), 7.37 (dd, $J = 8.6$, 7.4 Hz, 2H, $H_{3'}$), 7.73 (d, $J = 8.6$ Hz, 2H, $H_{2'}$); ^{13}C NMR (150.9 MHz, CDCl_3) δ 14.1 ($C_{5''}$), 16.2 (d, $J_{\text{CP}} = 6.8$ Hz, $C_{2''}$), 22.5 ($C_{4''}$), 27.5 ($C_{2''}$), 28.6 ($C_{1''}$), 31.4 ($C_{3''}$), 64.9 (d, $J_{\text{CP}} = 6.1$ Hz, $C_{1''}$), 102.7 (C_4), 123.7 ($C_{2'}$), 126.6 ($C_{4'}$), 128.6 ($C_{3'}$), 130.0 ($C_{1'}$), 135.6 (d, $J_{\text{CP}} = 6.2$ Hz, C_2), 137.9 (d, $J_{\text{CP}} = 10.5$ Hz, C_3), 154.4 (C_5); ^{31}P NMR (121.5 MHz, CDCl_3) δ -5.18; ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_4 6.29 ppm) δ 28.6 ($^3J_{\text{CH}} = 1.0$ Hz), 102.7 (folded $^1J_{\text{CH}}$), 154.4 ($^2J_{\text{CH}} = 7.8$ Hz).

Compounds **6**, (**Z**)-**7** and **8** were prepared using reported procedures (see ESI†).^{9b}

5-Methylenedihydrofuran-2-one (**6**)^{9b}

5-Methylenedihydrofuran-2-one (**6**)^{9b} was obtained in 96% yield over 1 step, from pent-4-ynoic acid. Colorless oil; TLC R_f 0.18 (pentane/Et₂O 10%); ^1H NMR (500 MHz, CDCl_3) δ 2.64–2.70 (m, 2H, H_3), 2.85–2.91 (m, 2H, H_4), 4.31 (dt, $J = 2.2$, 2.0 Hz, 1H, H_E), 4.72 (td, $J = 2.3$, 2.2 Hz, 1H, H_2); ^{13}C NMR (125.8 MHz, CDCl_3) δ 25.2 (C_4), 28.1 (C_3), 88.9 ($C_{1'}$), 155.7 (C_5), 175.0 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_Z 4.72 ppm) δ 25.2 ($^3J_{\text{CH}} = 7.8$ Hz), 28.1 ($^4J_{\text{CH}} = 1.0$ Hz), 88.9 (folded $^1J_{\text{CH}}$), 155.7 ($^2J_{\text{CH}} = 6.9$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_E 4.31 ppm) δ 25.2 ($^3J_{\text{CH}} = 3.3$ Hz), 88.9 (folded $^1J_{\text{CH}}$), 155.7 ($^2J_{\text{CH}} = 6.4$ Hz).

(Z)-5-Benzylidenedihydrofuran-2-one ((Z)-7)^{9b}

(Z)-5-Benzylidenedihydrofuran-2-one ((Z)-7)^{9b} was obtained in 49% yield over 1 step, from 5-phenylpent-4-ynoic acid. Beige solid; TLC R_f 0.17 (cyclohexane/EtOAc 10%); ^1H NMR (500 MHz, CDCl_3) δ 2.68–2.74 (m, 2H, H_3), 3.03 (td, $J = 8.6$, 1.8 Hz, 2H, H_4), 5.55 (t, $J = 1.8$ Hz, 1H, $H_{1'}$), 7.21 (t, $J = 7.4$ Hz, 1H, $H_{5'}$), 7.33 (dd, $J = 7.8$, 7.4 Hz, 2H, $H_{4'}$), 7.55 (d, $J = 7.8$ Hz, 2H, $H_{3'}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 26.5 (C_4), 27.1 (C_3), 105.0 ($C_{1'}$), 126.9 ($C_{5'}$), 128.4 ($C_{3'}$), 128.6 ($C_{4'}$), 134.0 ($C_{2'}$), 148.2 (C_5), 175.1 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: $H_{1'}$ 5.55 ppm) δ 26.5 ($^3J_{\text{CH}} = 3.9$ Hz), 105.0 (folded $^1J_{\text{CH}}$), 128.4 ($^3J_{\text{CH}} = 5.6$ Hz), 148.2 ($^2J_{\text{CH}} = 5.6$ Hz).

(Z)-5-(3-Phenylprop-2-yn-1-ylidene)dihydrofuran-2-one (**8**)^{9b}

(Z)-5-(3-Phenylprop-2-yn-1-ylidene)dihydrofuran-2-one (**8**)^{9b} was obtained in 75% yield over 1 step, from 7-phenylheptan-4,6-diyonic acid. Yellow solid; TLC R_f 0.47 (cyclohexane/EtOAc 10%); ^1H NMR (500 MHz, CDCl_3) δ 2.72–2.77 (m, 2H, H_3), 2.99 (td, $J = 8.5$, 1.9 Hz, 2H, H_4), 5.04 (t, $J = 1.9$ Hz, 1H, $H_{1'}$), 7.27–7.24 (m, 3H, $H_{6'/7'}$), 7.42–7.48 (m, 2H, $H_{5'}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 25.6 (C_4), 27.5 (C_3), 82.3 ($C_{2'}$ or $C_{3'}$), 85.6 ($C_{1'}$), 93.8 ($C_{2'}$ or $C_{3'}$), 123.4 ($C_{4'}$), 128.3 ($C_{7'}$), 128.4 ($C_{6'}$), 131.6 ($C_{5'}$), 158.3 (C_5), 173.9 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: $H_{1'}$ 5.04 ppm) δ 25.6 ($^3J_{\text{CH}} = 3.2$ Hz), 82.3 ($^{2/3}J_{\text{CH}} = 2.3$ Hz), 85.6 (folded $^1J_{\text{CH}}$), 93.8 ($^{2/3}J_{\text{CH}} = 4.6$ Hz), 123.4 ($^4J_{\text{CH}} = 1.7$ Hz), 131.6 ($^5J_{\text{CH}} = 1.7$ Hz), 158.3 ($^2J_{\text{CH}} = 4.8$ Hz).

(E)-5-Benzylidenedihydrofuran-2-one ((E)-7)

(E)-5-Benzylidenedihydrofuran-2-one ((E)-7) was obtained in 68% yield over 1 step from pent-4-ynoic acid, using reported procedure (see ESI†).⁴⁶ Beige solid; TLC R_f 0.20 (pentane/Et₂O 10%); ^1H NMR (500 MHz, CDCl_3) δ 2.73–2.79 (m, 2H, H_3), 3.17 (td, $J = 8.6$, 2.2 Hz, 2H, H_4), 6.33 (t, $J = 2.2$ Hz, 1H, $H_{1'}$), 7.20–7.25 (m, 3H, $H_{3'/5'}$), 7.35 (dd, $J = 7.8$, 7.8 Hz, 2H, $H_{4'}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 25.3 (C_4), 27.9 (C_3), 107.2 ($C_{1'}$), 126.8 ($C_{5'}$), 127.9 ($C_{3'}$), 128.8 ($C_{4'}$), 134.5 ($C_{2'}$), 151.3 (C_5), 174.3 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: $H_{1'}$ 6.33 ppm) δ 25.3 ($^3J_{\text{CH}} = 6.2$ Hz), 107.2 (folded $^1J_{\text{CH}}$), 127.9 ($^3J_{\text{CH}} = 5.3$ Hz), 151.3 ($^2J_{\text{CH}} = 7.7$ Hz).

(Z)-4-Bromo-5-(bromomethylene)-2-furanone (**9**) (commercially available)

White solid; ^1H NMR (500 MHz, CDCl_3) δ 6.42 (s, 1H, $H_{1'}$), 6.50 (s, 1H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 93.9 ($C_{1'}$), 121.2 (C_3), 135.4 (C_5), 151.2 (C_4), 165.6 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: $H_{1'}$ 6.42 ppm) δ 93.9 (folded $^1J_{\text{CH}}$), 135.4 ($^2J_{\text{CH}} = 4.3$ Hz), 151.2 ($^3J_{\text{CH}} = 10.6$ Hz).

Compounds **10a** and (**E**)-**10b** were prepared using reported procedures (see ESI†).^{35a}

(E)-4-Methyl-1-phenyl-2-pivaloxy-penta-1,4-dien-3-one (**10a**)^{35a}

(E)-4-Methyl-1-phenyl-2-pivaloxy-penta-1,4-dien-3-one (**10a**)^{35a} was obtained in 59% yield over 3 steps, from 2-methylbut-1-en-3-yne. Yellow solid; mp 35 °C; TLC R_f 0.51 (cyclohexane/EtOAc 20%); ^1H NMR (500 MHz, CDCl_3) δ 1.30 (s, 9H, $H_{3''}$), 1.82 (dd,



$J = 1.4, 1.2$ Hz, 3H, $H_{1''}$), 5.53 (dq, $J = 1.6, 1.4$ Hz, 1H, H_{5E}), 5.94 (dq, $J = 1.6, 1.2$ Hz, 1H, H_{5Z}), 6.84 (s, 1H, H_1), 7.10–7.14 (m, 2H, H_2), 7.25–7.29 (m, 3H, $H_{3'/4'}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 17.3 ($C_{1''}$), 27.0 ($C_{3''}$), 38.7 ($C_{2''}$), 124.2 (C_1), 127.8 (C_5), 128.5 (C_3), 128.5 (C_4), 129.2 ($C_{2'}$), 133.3 ($C_{1'}$), 142.7 (C_4), 144.0 (C_2), 177.0 ($C_{1''}$), 193.3 (C_3); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_1 6.84 ppm) δ 124.2 (folded $^1J_{\text{CH}}$), 129.2 ($^3J_{\text{CH}} = 6.4$ Hz), 144.0 ($^2J_{\text{CH}} = 8.4$ Hz), 193.3 ($^3J_{\text{CH}} = 8.1$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_{5Z} 5.94 ppm) δ 17.3 ($^3J_{\text{CH}} = 10.3$ Hz), 127.8 (folded $^1J_{\text{CH}}$), 142.7 ($^2J_{\text{CH}} = 2.6$ Hz), 193.3 ($^3J_{\text{CH}} = 7.1$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_{5E} 5.53 ppm) δ 17.3 ($^3J_{\text{CH}} = 6.1$ Hz), 127.8 (folded $^1J_{\text{CH}}$), 142.7 ($^2J_{\text{CH}} = 1.6$ Hz), 193.3 ($^3J_{\text{CH}} = 13.3$ Hz).

(*E*)-4-Methyl-1-(4-nitrophenyl)-2-pivaloxypenta-1,4-dien-3-one ((*E*)-10b)

(*E*)-4-Methyl-1-(4-nitrophenyl)-2-pivaloxypenta-1,4-dien-3-one ((*E*)-10b) was obtained in 18% yield over 3 steps, from 2-methylbut-1-en-3-yne (see ESI†). Yellowish solid; mp 78 °C; TLC R_f 0.42 (cyclohexane/EtOAc 20%); IR (neat) ν_{max} 695, 738, 821, 858, 1037, 1103, 1152, 1202, 1273, 1342, 1513, 1592, 1667, 1720, 1745, 2874, 2935, 2976, 3079 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.29 (s, 9H, $H_{3''}$), 1.84 (dd, $J = 1.7, 1.0$ Hz, 3H, $H_{1''}$), 5.67 (dq, $J = 1.7, 1.2$ Hz, 1H, H_{5E}), 5.94 (dq, $J = 1.2, 1.0$ Hz, 1H, H_{5Z}), 6.80 (s, 1H, H_1), 7.30 (d, $J = 8.9$ Hz, 2H, H_2), 8.14 (d, $J = 8.9$ Hz, 2H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 17.2 ($C_{1''}$), 26.9 ($C_{3''}$), 38.7 ($C_{2''}$), 121.2 (C_1), 123.8 (C_3), 128.3 (C_5), 129.8 ($C_{2'}$), 139.8 ($C_{4'}$), 142.9 (C_4), 146.5 (C_2), 147.4 ($C_{1'}$), 176.5 ($C_{1''}$), 192.1 (C_3); HR-MS 340.1175 ($\text{C}_{17}\text{H}_{19}\text{NO}_5$ + Na calcd 340.1155); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_1 6.80 ppm) δ 121.2 (folded $^1J_{\text{CH}}$), 129.8 ($^3J_{\text{CH}} = 4.7$ Hz), 146.5 ($^2J_{\text{CH}} = 8.4$ Hz), 147.4 ($^2J_{\text{CH}} = 1.0$ Hz), 192.1 ($^3J_{\text{CH}} = 7.8$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_{5Z} 5.94 ppm) δ 17.2 ($^3J_{\text{CH}} = 10.2$ Hz), 128.3 (folded $^1J_{\text{CH}}$), 142.9 ($^2J_{\text{CH}} = 2.4$ Hz), 192.1 ($^3J_{\text{CH}} = 7.2$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_{5E} 5.67 ppm) δ 17.2 ($^3J_{\text{CH}} = 6.0$ Hz), 128.3 (folded $^1J_{\text{CH}}$), 142.9 ($^2J_{\text{CH}} = 1.6$ Hz), 192.1 ($^3J_{\text{CH}} = 13.7$ Hz).

Compounds (*Z*)-10b and 10c were prepared using reported procedures (see ESI†).^{35a,47}

(*Z*)-4-Methyl-1-(4-nitrophenyl)-2-pivaloxypenta-1,4-dien-3-one ((*Z*)-10b)

(*Z*)-4-Methyl-1-(4-nitrophenyl)-2-pivaloxypenta-1,4-dien-3-one ((*Z*)-10b) was obtained in 45% yield over 2 steps, from 2-methylbut-1-en-3-yne. Yellowish solid; mp 68 °C; TLC R_f 0.46 (pentane/Et₂O 10%); IR (neat) ν_{max} 691, 748, 778, 850, 892, 1031, 1092, 1164, 1270, 1340, 1514, 1596, 1656, 1716, 1751, 2874, 2934, 2977 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.35 (s, 9H, $H_{3''}$), 2.01 (dd, $J = 1.4, 1.0$ Hz, 3H, $H_{1''}$), 5.86 (dq, $J = 1.4, 1.2$ Hz, 1H, H_5), 5.90 (dq, $J = 1.2, 1.0$ Hz, 1H, H_5), 6.88 (s, 1H, H_1), 7.75 (d, $J = 8.9$ Hz, 2H, H_2), 8.23 (d, $J = 8.9$ Hz, 2H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 18.4 ($C_{1''}$), 27.0 ($C_{3''}$), 39.0 ($C_{2''}$), 123.7 (C_1), 123.9 (C_3), 125.9 (C_5), 130.7 ($C_{2'}$), 138.9 ($C_{4'}$), 142.6 (C_4), 147.0 (C_2), 147.7 ($C_{1'}$), 176.0 ($C_{1''}$), 191.5 (C_3); HR-MS 340.1175 ($\text{C}_{17}\text{H}_{19}\text{NO}_5$

+ Na calcd 340.1155); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_1 6.88 ppm) δ 123.7 (folded $^1J_{\text{CH}}$), 130.7 ($^3J_{\text{CH}} = 5.4$ Hz), 147.0 ($^2J_{\text{CH}} = 4.6$ Hz), 191.5 ($^3J_{\text{CH}} = 3.8$ Hz).

(*Z*)-2-Acetoxy-1-(cyclohex-1-enyl)non-2-en-1-one (10c)

(*Z*)-2-Acetoxy-1-(cyclohex-1-enyl)non-2-en-1-one (10c) was obtained in 90% yield over 2 steps, from ethynylcyclohex-1-ene. Colorless oil; TLC R_f 0.22 (pentane/Et₂O 20%); IR (neat) ν_{max} 590, 701, 745, 835, 883, 975, 1014, 1092, 1136, 1204, 1250, 1276, 1307, 1435, 1647, 1759, 2857, 2927 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.88 (t, $J = 6.9$ Hz, 3H, H_9), 1.23–1.25 (m, 6H, $H_{8/7/6}$), 1.43 (tt, $J = 7.4, 7.1$ Hz, 2H, H_5), 1.59–1.69 (m, 4H, $H_{4''/5''}$), 2.02 (dt, $J = 7.6, 7.4$ Hz, 2H, H_4), 2.21 (s, 3H, H_2), 2.20–2.30 (m, 4H, $H_{3''/6''}$), 6.05 (t, $J = 7.6$ Hz, 1H, H_3), 6.69–6.73 (m, 1H, $H_{2''}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 14.2 (C_9), 20.5 ($C_{2'}$), 21.7 ($C_{4''}$), 22.0 ($C_{5''}$), 22.6 (C_8), 24.0 ($C_{6''}$), 25.9 ($C_{3''}$), 26.1 (C_4), 28.5 (C_5), 29.1 (C_6), 31.6 (C_7), 131.9 (C_3), 137.7 ($C_{1''}$), 140.8 ($C_{2''}$), 145.3 (C_2), 169.0 ($C_{1'}$), 191.1 (C_1); HR-MS 301.175 ($\text{C}_{17}\text{H}_{26}\text{O}_3$ + Na calcd 301.177); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_3 6.05 ppm) δ 26.1 ($^2J_{\text{CH}} = 1.8$ Hz), 28.5 ($^3J_{\text{CH}} = 3.2$ Hz), 131.9 (folded $^1J_{\text{CH}}$), 145.3 ($^2J_{\text{CH}} = 6.8$ Hz), 191.1 ($^3J_{\text{CH}} = 3.4$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: $H_{2''}$ 6.69–6.73 ppm) δ 21.7 ($^3J_{\text{CH}} = 5.5$ Hz), 24.0 ($^3J_{\text{CH}} = 7.2$ Hz), 25.9 ($^2J_{\text{CH}} = 3.7$ Hz), 137.7 ($^2J_{\text{CH}} = 0.9$ Hz), 140.8 (folded $^1J_{\text{CH}}$), 191.1 ($^3J_{\text{CH}} = 6.4$ Hz).

(*E*)-1-(4,5-Dihydro-6H-pyran-2-yl)-2-methyl-3-phenylprop-2-en-1-one (11)⁴⁸

(*E*)-1-(4,5-Dihydro-6H-pyran-2-yl)-2-methyl-3-phenylprop-2-en-1-one (11)⁴⁸ was obtained in 26% yield over 2 steps from dihydropyran, using reported procedure (see ESI†).^{35a} Colorless oil; TLC R_f 0.40 (cyclohexane/EtOAc 30%); ^1H NMR (500 MHz, CDCl_3) δ 1.91 (tdd, $J = 6.4, 6.0, 4.3$ Hz, 2H, H_5), 2.13 (d, $J = 1.4$ Hz, 3H, $H_{1''}$), 2.26 (td, $J = 6.4, 4.2$ Hz, 2H, $H_{4'}$), 4.17 (dd, $J = 6.0, 4.3$ Hz, 2H, H_6), 5.82 (t, $J = 4.2$ Hz, 1H, H_3), 7.24 (q, $J = 1.4$ Hz, 1H, H_3), 7.29–7.36 (m, 1H, $H_{4''}$), 7.35–7.42 (m, 4H, $H_{2''/3''}$); ^{13}C NMR (125.8 MHz, CDCl_3) δ 14.9 ($C_{1''}$), 21.0 ($C_{4'}$), 21.7 ($C_{5'}$), 66.5 (C_6), 113.4 (C_3), 128.4 ($C_{4''}$), 128.5 ($C_{3''}$), 129.7 ($C_{2''}$), 135.9 ($C_{1''}$ or C_2), 136.0 ($C_{1''}$ or C_2), 138.9 (C_3), 151.4 ($C_{2'}$), 193.9 (C_1); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_3 5.82 ppm) δ 14.9 ($^3J_{\text{CH}} = 8.3$ Hz), 21.0 ($^2J_{\text{CH}} = 2.6$ Hz), 21.7 ($^3J_{\text{CH}} = 5.5$ Hz), 113.4 (folded $^1J_{\text{CH}}$), 151.4 ($^2J_{\text{CH}} = 4.7$ Hz), 193.9 ($^3J_{\text{CH}} = 3.5$ Hz); ^{13}C - $\{^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_3 7.24 ppm) δ 138.9 (folded $^1J_{\text{CH}}$), 193.9 ($^3J_{\text{CH}} = 6.8$ Hz).

Compounds 12a, 12c and 12d were prepared using reported procedures (see ESI†).^{36e,49}

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-(2,4-dimethoxyphenyl)prop-2-enoate (12a)

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-(2,4-dimethoxyphenyl)prop-2-enoate (12a) was obtained in 64% yield over 2 steps, from cyclohex-1-en-1-carboxylic acid. Yellowish solid; mp 99 °C; TLC R_f 0.09 (pentane/Et₂O 30%); IR (neat) ν_{max} 401, 461, 530, 768, 802, 826, 927, 995, 1030, 1044, 1072, 1124, 1162, 1192, 1207, 1237, 1301, 1313, 1422, 1434, 1459, 1501, 1593, 1632,



1646, 1706, 2836, 2929, 2945 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.50–1.57 (m, 2H, H_4^m), 1.58–1.64 (m, 2H, H_5^m), 2.07–2.14 (m, 2H, H_3^m), 2.29–2.35 (m, 2H, H_6^m), 3.77 (s, 3H, H_{OMe}), 3.79 (s, 3H, H_{o-} or $p\text{-OMe}$), 3.82 (s, 3H, H_{o-} or $p\text{-OMe}$), 6.35 (dd, $J = 8.8, 2.5$ Hz, 1H, $H_{3'}$ or $5'$), 6.39 (d, $J = 2.5$ Hz, 1H, $H_{3'}$ or $5'$), 6.80–6.84 (m, 1H, H_2^m), 7.12 (d, $J = 8.8$ Hz, 1H, H_6'), 8.14 (s, 1H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 21.6 (C_4^m), 21.9 (C_5^m), 22.8 (C_6^m), 26.3 (C_3^m), 52.4 (C_{OMe}), 55.6 (C_{o-} or $p\text{-OMe}$), 55.6 (C_{o-} or $p\text{-OMe}$), 98.3 ($C_{3'}$ or $5'$), 105.0 ($C_{3'}$ or $5'$), 115.6 (C_1), 128.1 (C_2), 131.2 (C_6'), 136.9 (C_3), 139.2 (C_1^m), 144.8 (C_2^m), 159.8 (C_2), 162.9 (C_4'), 166.5 (C_1), 197.8 (C_1^m); HR-MS 353.1389 ($\text{C}_{19}\text{H}_{22}\text{O}_5 + \text{Na}$ calcd 353.1359); $^{13}\text{C}\{-^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_3 8.14 ppm) δ 128.1 ($^2J_{\text{CH}} = 4.2$ Hz), 131.2 ($^3J_{\text{CH}} = 6.3$ Hz), 136.9 (folded $^1J_{\text{CH}}$), 159.8 ($^3J_{\text{CH}} = 3.5$ Hz), 166.5 ($^3J_{\text{CH}} = 8.2$ Hz), 197.8 ($^3J_{\text{CH}} = 10.1$ Hz); $^{13}\text{C}\{-^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_2^m 6.80–6.84 ppm) δ 21.6 ($^3J_{\text{CH}} = 5.3$ Hz), 22.8 ($^3J_{\text{CH}} = 6.8$ Hz), 26.3 ($^2J_{\text{CH}} = 3.7$ Hz), 139.2 ($^2J_{\text{CH}} = 1.4$ Hz), 144.8 (folded $^1J_{\text{CH}}$), 197.8 ($^3J_{\text{CH}} = 7.3$ Hz).

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-phenylprop-2-enoate (12c)

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-phenylprop-2-enoate (12c) was obtained in 54% yield over 2 steps, from cyclohex-1-en-1-carboxylic acid. Yellowish solid; mp 43 °C; TLC R_f 0.19 (cyclohexane/EtOAc 10%); IR (neat) ν_{max} 401, 492, 551, 580, 680, 770, 830, 929, 992, 1068, 1102, 1185, 1212, 1391, 1421, 1497, 1619, 1647, 1713, 2861, 2929 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.52–1.58 (m, 2H, H_4^m), 1.60–1.66 (m, 2H, H_5^m), 2.09–2.14 (m, 2H, H_3^m), 2.32–2.38 (m, 2H, H_6^m), 3.79 (s, 3H, H_{OMe}), 6.83 (t, $J = 7.6$ Hz, 1H, H_2^m), 7.29–7.35 (m, 5H, $H_{2'/3'/4'}$), 7.81 (s, 1H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 21.5 (C_4^m or C_5^m), 21.8 (C_4^m or C_5^m), 22.8 (C_6^m), 26.4 (C_3^m), 52.7 (C_{OMe}), 128.8 ($C_{3'}$), 130.1 (C_2), 130.3 (C_4'), 131.4 (C_2), 133.4 (C_1'), 139.3 (C_1^m), 142.1 (C_3), 145.6 (C_2^m), 165.9 (C_1), 196.9 (C_1^m); HR-MS 293.114 ($\text{C}_{17}\text{H}_{18}\text{O}_3 + \text{Na}$ calcd 293.115); $^{13}\text{C}\{-^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 , chosen proton: H_3 7.81 ppm) δ 130.1 ($^3J_{\text{CH}} = 5.6$ Hz), 131.4 ($^2J_{\text{CH}} = 4.2$ Hz), 133.4 ($^2J_{\text{CH}} = 1.2$ Hz), 142.1 (folded $^1J_{\text{CH}}$), 165.9 ($^3J_{\text{CH}} = 8.1$ Hz), 196.9 ($^3J_{\text{CH}} = 9.7$ Hz).

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-(4-methoxyphenyl)prop-2-enoate (12d)

Methyl (*E*)-2-(cyclohex-1-enylcarbonyl)-3-(4-methoxyphenyl)prop-2-enoate (12d) was obtained in 56% yield over 2 steps, from cyclohex-1-en-1-carboxylic acid. Colorless oil; TLC R_f 0.16 (pentane/Et₂O 30%); IR (neat) ν_{max} 520, 540, 830, 924, 989, 1027, 1070, 1119, 1171, 1200, 1250, 1305, 1433, 1511, 1599, 1632, 1651, 1704, 2839, 2859, 2933 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.54–1.61 (m, 2H, H_4^m), 1.63–1.69 (m, 2H, H_5^m), 2.11–2.16 (m, 2H, H_3^m), 2.35–2.40 (m, 2H, H_6^m), 3.78 (s, 3H, H_{OMe}), 3.81 (s, 3H, $H_{p\text{-OMe}}$), 6.83 (d, $J = 8.9$ Hz, 2H, $H_{3'}$), 6.85–6.88 (m, 1H, H_2^m), 7.27 (d, $J = 8.9$ Hz, 2H, H_2), 7.75 (s, 1H, H_3); ^{13}C NMR (125.8 MHz, CDCl_3) δ 21.6 (C_4^m or C_5^m), 21.8 (C_4^m or C_5^m), 22.8 (C_6^m), 26.4 (C_3^m), 52.5 (C_{OMe}), 55.5 ($C_{p\text{-OMe}}$), 114.3 ($C_{3'}$), 125.9 (C_1), 128.7 (C_2), 132.2 (C_2), 139.3 (C_1^m), 141.8 (C_3), 145.4 (C_2^m), 161.3 (C_4'), 166.2 (C_1), 197.5 (C_1^m); HR-MS 323.125 ($\text{C}_{18}\text{H}_{20}\text{O}_4 + \text{Na}$ calcd 323.125); $^{13}\text{C}\{-^1\text{H}\}$ selective J -resolved NMR (125.8 MHz, CDCl_3 ,

chosen proton: H_3 7.75 ppm) δ 125.9 ($^2J_{\text{CH}} = 1.1$ Hz), 128.7 ($^2J_{\text{CH}} = 4.3$ Hz), 132.2 ($^3J_{\text{CH}} = 5.9$ Hz), 141.8 (folded $^1J_{\text{CH}}$), 166.2 ($^3J_{\text{CH}} = 8.1$ Hz), 197.5 ($^3J_{\text{CH}} = 9.8$ Hz).

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