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Highly efficient synthesis of polyfluorinated dendrons suitable for click chemistry†

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A highly efficient convergent synthesis of C_{2v} -symmetric dendrons is presented. The generation build-up was achieved by copper(I)-catalyzed azide-alkyne cycloadditions. The resulting G2-dendron consists of 72 magnetically equivalent fluorine atoms and displays a single sharp resonance in its ^{19}F NMR spectrum.

Nowadays, magnetic resonance imaging (MRI) is an established and very versatile, non-invasive, non-destructive imaging technique for diagnosis in human medicine.¹ MRI provides high resolution between soft tissues (few millimeters for human scanners and up to 50 μm for ultra high-field animal scanners), but without the need for ionizing and thus damaging radiation.² Besides ^1H , other NMR active heteronuclei could be used as well for MRI, which allows for adding a second “color” to anatomical scans obtained by classical ^1H MRI.³ However, most nuclei are unfit for MRI applications, because of their inherent physical, chemical and biological properties. But ^{19}F has been shown to be exceptionally suitable for MRI as it is the second most sensitive, stable nucleus for MR spectroscopy (directly after ^1H) and has 100% natural abundance.^{4,5} Furthermore, the resonance frequency of ^{19}F differs by only 6% from the frequency of ^1H , which allows creating ^{19}F MR images on common ^1H MRI devices.³

The conventional ^1H MRI makes use of the nuclear spin of the ubiquitous water molecules within organic tissues to enable visualization. On the contrary, fluorine is virtually absent in the biosphere.⁵ The external addition of a suitable, fluorinated

compound (called probe or tracer) is thus mandatory for ^{19}F MRI.⁶ Only teeth and the bone matrix contain considerable amounts of endogenous fluorine. However, immobilized fluorine exhibits a very short spin-spin relaxation time and is therefore not detectable by ^{19}F MRI.⁵ Consequently, ^{19}F MR images of biological samples do not have an intrinsic background signal, which results in an extremely high contrast.⁵ In summary, the information obtained from ^1H and ^{19}F MRI is complementary: ^1H MRI collects morphological data (anatomy) and ^{19}F MRI enables tracking of a fluorinated, exogenous compound (e.g. drugs).⁷

Suitable probes for ^{19}F MRI should preferably fulfill several requirements. Of course, the probe must be chemically inert and biologically compatible.³ Most organofluorine compounds easily match these requirements because of the extraordinary strength of the C–F bond.⁸ The probe should also have a large number of fluorine atoms to provide a high spin density.³ This is often accomplished by using perfluorinated compounds, like perfluorooctyl bromide (PFOB).⁹ But perfluorinated molecules typically have the drawback of displaying multiple, split ^{19}F signals, which reduces signal intensity and leads to image artefacts.¹⁰ This problem can be overcome by applying ^{19}F MRI probes with high symmetry, like perfluoro[15]crown-5 ether (PF15C5).¹¹ Finally, it would be advantageous, if the probe is water-soluble to allow for easy application without the need for complex formulation.¹² This is challenging since highly fluorinated molecules are in general very hydrophobic.¹³ As a consequence, many probes have to be applied as stable emulsions, e.g. by mixing with a surfactant like lecithin.^{9,11} In some cases, amphiphilic polyfluorinated polymers have also been shown to self-assemble into micelles in aqueous solution, which allowed for direct application without the need for stabilizing additives.¹⁴ Nevertheless, a more elegant approach would be to acquire the requested water solubility *via* conjugation with hydrophilic compounds.¹² Hence, ^{19}F MRI probes that follow a rational design, should also provide a conjugation site for tuning their water solubility.

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Inspired by the spherically symmetric ^{19}F MRI tracers from Yu *et al.*^{12,15} and owing our own interest in fluorine chemistry,¹⁶ we set out to find an easy access to dendrimeric molecules, which can meet the aforementioned requirements. Herein, we present an efficient three-step procedure for the preparation of C_{2v} -symmetric polyfluorinated dendrons by applying the copper(I)-catalyzed azide-alkyne cycloaddition (CuAAC),¹⁷ which is a common reaction for the preparation of various dendrimers and MRI probes,¹⁸ as a key reaction.

Since perfluoro-*tert*-butanol already has nine magnetically equivalent fluorine atoms and displays neither interfering ^1H - ^{19}F nor ^{19}F - ^{19}F coupling,^{15a} this molecule was chosen as a fluorinated precursor. Furthermore, Horváth *et al.* have shown that the sodium salt of this perfluorinated alcohol reacts with various benzyl bromides *via* nucleophilic substitution in moderate to good yields.¹⁹ After a twofold substitution of dibromide **1-Br**²⁰ the polyfluorinated aryl iodide **1-OC(CF₃)₃** was accessible in excellent yield (Scheme 1). Next, silylalkyne **2-SiMe₃** was synthesized *via* Sonogashira coupling²¹ and upon subsequent deprotection with K_2CO_3 in methanol terminal alkyne **2-H** was obtained. Notably, both silane **2-SiMe₃** and alkyne **2-H** were poorly soluble in the reaction mixture, although methanol is known to be a good fluorophilic solvent.¹³ However, the deprotection proceeded quantitatively and alkyne **2-H** could be isolated by simple filtration without the need for further purification. The first-generation dendron (**G1-dendron**) **3** was built up by twofold CuAAC of alkyne **2-H** with diazide **1-N₃**.²⁰ The three-step generation build-up (Sonogashira reaction, deprotection, CuAAC) for the **G1-dendron** **3** had a total yield of 82% and could be done easily on a gram scale.

During this sequence the ^{19}F NMR spectrum of every compound showed just one sharp singlet. This proved that all fluorine atoms are indeed magnetically equivalent. Moreover, iodide **1-OC(CF₃)₃** and alkyne **2-H** could be crystallized from perfluorohexane or cyclohexane, respectively, to obtain single crystals suitable for X-ray crystallography. The molecular structures of those perfluoro-*tert*-butyl ethers are depicted in Fig. 1.

The generation build-up sequence was repeated once more to afford the second-generation dendron **5** (Scheme 2). Expectedly, the overall yield for this second cycle decreased slightly (67%), which can be explained by increased steric hindrance.^{15b} Nevertheless the yields for each step were still quite high (77–94%). The 72 fluorine atoms of dendron **5** resulted in just one sharp singlet in the respective ^{19}F NMR spectrum. Hence,

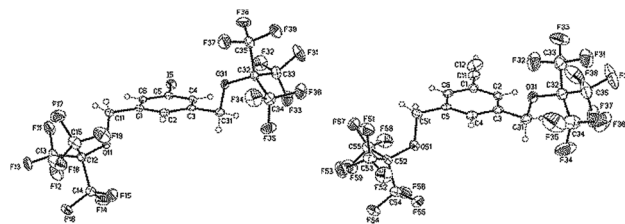
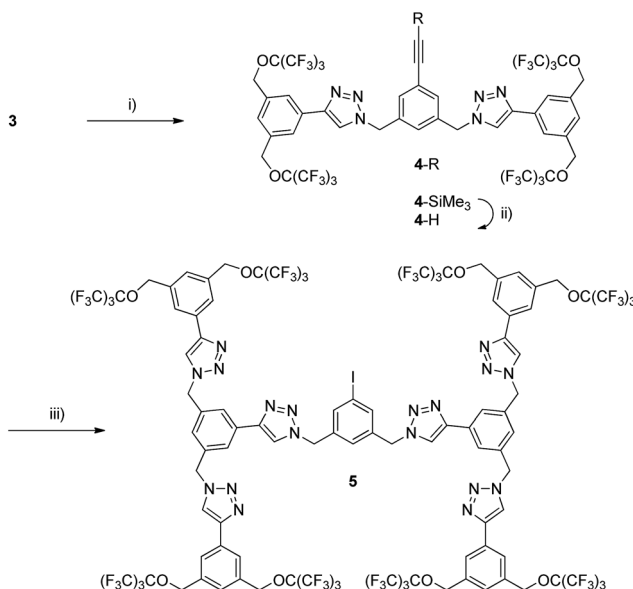


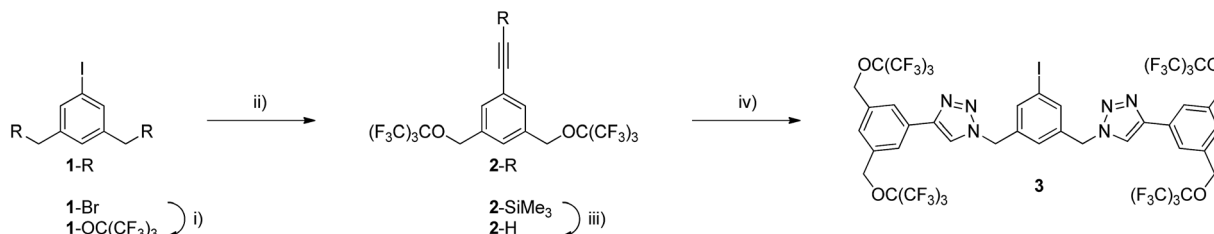
Fig. 1 Molecular structure of iodide **1-OC(CF₃)₃** (left side) and alkyne **2-H** (right side) with displacement ellipsoids drawn at 50% probability level.²²

dendron **5** has considerably more magnetically equivalent fluorine atoms than typical, commercially available ^{19}F MRI probes (e.g. PF15C5).

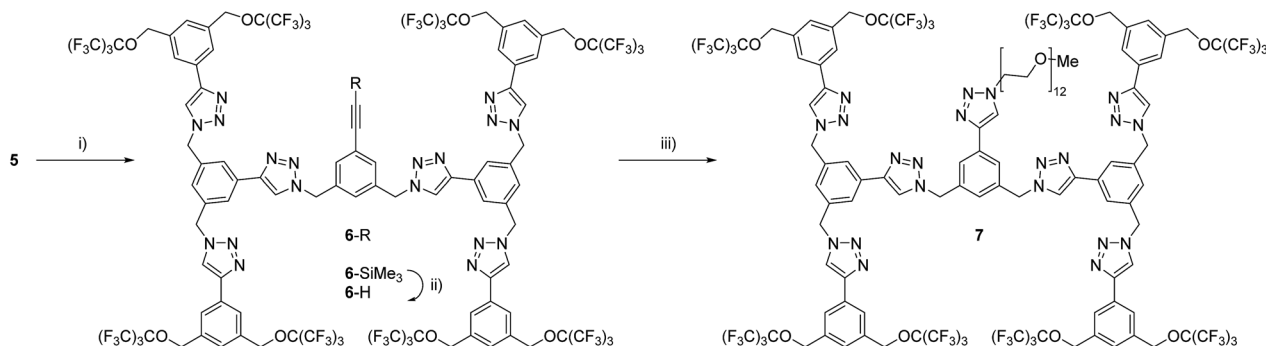
To provide dendron **5** with an appropriate conjugation site, the sequence of Sonogashira coupling with trimethylsilylacetylene and subsequent desilylation with K_2CO_3 was repeated one last time (Scheme 3). Alkyne **6-H** was obtained in 37% overall yield after 9 steps from dibromide **1-Br**. Finally, we tried to



Scheme 2 Synthesis of **G2-dendron 5**. Reagents and conditions: (i) trimethylsilylacetylene, $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$, CuI , NEt_3 , THF, 45 °C, overnight, 94%; (ii) K_2CO_3 , MeOH, rt, 1.5 day, 93%; (iii) **1-N₃**, $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$, 2,6-lutidine, THF, 40 °C, 2 days, 77%.



Scheme 1 Synthesis of **G1-dendron 3**. Reagents and conditions: (i) $\text{NaOC(CF}_3)_3$, DMF, rt, 18 h, 88%; (ii) trimethylsilylacetylene, $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$, CuI , NEt_3 , THF, 45 °C, 4 h, 89%; (iii) K_2CO_3 , MeOH, rt, 1.5 day, >99%; (iv) **1-N₃**, $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$, 2,6-lutidine, THF, 40 °C, 2 days, 92%.



Scheme 3 Synthesis of conjugate **7**. Reagents and conditions: (i) trimethylsilylacetylene, $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$, CuI , NEt_3 , THF, 45°C , overnight, 82%; (ii) K_2CO_3 , MeOH, rt, 1.5 day, 92%; (iii) *O*-(2-azidoethyl)-*O'*-methylundecaethylene glycol, $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$, 2,6-lutidine, THF, rt, 2.5 days, 31%.

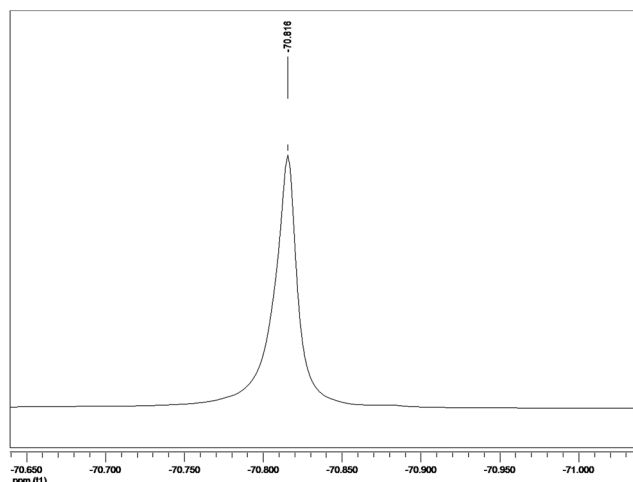


Fig. 2 ^{19}F NMR spectrum (377 MHz) of dendron **7** in CDCl_3 (trifluoroacetic acid was used as internal standard).

convert dendron **6-H** into a more hydrophilic form *via* conjugation with an azide-functionalized undecaethylene glycol derivative. This strategy of tethering unpolar molecules to hydrophilic ethylene glycols is commonly used in medicine to improve the water solubility.²³ In addition, poly(ethylene glycol) moieties have also been used to improve the biocompatibility of highly fluorinated hyperbranched polymers.²⁴ However, conjugate **7** was still too hydrophobic to dissolve in water. But overall this is just a minor drawback as ^{19}F MRI probes can also be applied in emulsified form.^{9,11,14}

The ^{19}F NMR spectrum of conjugate **7** expectedly showed a sharp singlet (Fig. 2). At a magnetic flux density of 9.4 T, the resonance signal had a narrow full width at half maximum of 7 Hz. This clearly demonstrates the potential applicability of the synthesized dendron as ^{19}F MRI probe.

In summary, conjugate **7** could overcome most of the challenges that should be met by appropriate ^{19}F MRI probes. It exhibits 72 fluorine atoms and has just one single, sharp resonance in the ^{19}F NMR spectrum. As a result conjugate **7** features a high spin density without the disadvantage of causing chemical shift artefacts. The only remaining challenge would be the lack of water solubility. To address this issue, dendron **6-H**

could be conjugated with other more hydrophilic (bio)molecules.²⁵ Hence, alkyne **6-H** is a versatile precursor for the generation of various ^{19}F MRI agents.

Conclusions

We described a facile and highly efficient procedure for the preparation of C_{2v} -symmetric dendrons that are well suited for ^{19}F MRI. This dendron synthesis is comprised of three steps that are repeated iteratively: Sonogashira coupling, alkyne deprotection and CuAAC for the generation build-up. After 9 steps the second-generation dendron **6-H** was obtained in excellent overall yield. This molecule displays 72 magnetically equivalent fluorine atoms and exhibits a terminal alkyne moiety, which serves as a tether. The latter is an important feature, because it could be used for adjusting the water solubility of a potential ^{19}F MRI probe *via* conjugation with hydrophilic molecules. Since conjugate **7** is still too unpolar to be water-soluble, other hydrophilic compounds will be tested in the near future to create polyfluorinated probes that are even more suitable for ^{19}F MRI.

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Notes and references

- (a) A. M. Blamire, *Br. J. Radiol.*, 2008, **81**, 601; (b) N. Mistry, A. Rosenkrantz, S. Roys, J. Papadimitriou, K. Siddiqui, M. Naslund, J. Borin, W. D'Souza and R. P. Gullapalli, *Mol. Cell. Pharmacol.*, 2013, **5**, 5; (c) D. Bartusik, D. Aebischer and B. Tomanek, *J. Mol. Imaging Dyn.*, 2013, **2**, 112.
- J. C. Knight, P. G. Edwards and S. J. Paisley, *RSC Adv.*, 2011, **1**, 1415.
- M. Srinivas, A. Heerschap, E. T. Ahrens, C. G. Figdor and I. J. M. de Vries, *Trends Biotechnol.*, 2010, **27**, 363.
- (a) Y. B. Yu, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2013, **5**, 646; (b) Z. X. Jiang, Y. Feng and Y. B. Yu, *Chem. Commun.*, 2011, **47**, 7233.

- 5 (a) J. Ruiz-Cabello, B. P. Barnett, P. A. Bottomley and J. W. M. Bulte, *NMR Biomed.*, 2011, **24**, 114; (b) K. K. J. Chan and D. O'Hagan, *Methods Enzymol.*, 2012, **516**, 219.
- 6 M. Srinivas, P. Boehm-Sturm, C. G. Figdor, I. J. de Vries and M. Hoehn, *Biomaterials*, 2012, **33**, 8830.
- 7 (a) M. Higuchi, N. Iwata, Y. Matsuba, K. Sato, K. Sasamoto and T. C. Saido, *Nat. Neurosci.*, 2008, **8**, 527; (b) M. Srinivas, P. A. Morel, L. A. Ernst, D. H. Laidlaw and E. T. Ahrens, *Magn. Reson. Med.*, 2007, **58**, 725; (c) Y. B. Yu and Z.-X. Jiang, *J. Pharm. Drug Deliv. Res.*, 2012, **1**, 1.
- 8 (a) D. O'Hagan, *Chem. Soc. Rev.*, 2008, **37**, 308; (b) S. F. Flaim, *Artif. Cells, Blood Substitutes, Immobilization Biotechnol.*, 1994, **22**, 1043.
- 9 (a) D. M. Freeman, H. H. Muller, R. E. Hurd and S. W. Young, *Magn. Reson. Imaging*, 1988, **6**, 61; (b) C. Giraudeau, B. Djemaï, M. A. Ghaly, F. Boumezbeur, S. Mériaux, P. Robert, M. Port, C. Robic, D. L. Bihan, F. Lethimonnier and J. Valette, *NMR Biomed.*, 2012, **25**, 654; (c) O. Diou, N. Tsapis, C. Giraudeau, J. Valette, C. Gueutin, F. Bourasset, S. Zanna, C. Vauthier and E. Fattal, *Biomaterials*, 2012, **33**, 5593.
- 10 (a) H. K. Lee, O. Nalcioğlu and R. B. Buxton, *Magn. Reson. Med.*, 1991, **21**, 21; (b) H. K. Lee, O. Nalcioğlu and R. B. Buxton, *Magn. Reson. Med.*, 1992, **23**, 254.
- 11 (a) J. M. Janjic and E. T. Ahrens, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2009, **1**, 492; (b) R. B. Heeswijk, Y. Pilloud, U. Flögel, J. Schwitter and M. Stuber, *PLoS One*, 2012, **7**, e42236; (c) M. Srinivas, L. J. Cruz, F. Bonetto, A. Heerschap, C. G. Figdor and I. J. M. de Vries, *Biomaterials*, 2010, **31**, 7070.
- 12 (a) Z.-X. Jiang, X. Liu, E.-K. Jeong and Y. B. Yu, *Angew. Chem.*, 2009, **121**, 4849; *Angew. Chem., Int. Ed.*, 2009, **48**, 4755; (b) M. B. Taraban, L. Yu, Y. Feng, E. V. Jouravleva, M. A. Anisimov, Z.-X. Jiang and Y. B. Yu, *RSC Adv.*, 2014, **4**, 54565.
- 13 I. T. Horváth, D. P. Curran and J. A. Gladysz, *Handbook of Fluorous Chemistry*, Wiley-VCH, Weinheim, 2004.
- 14 W. Du, A. M. Nyström, L. Zhang, K. T. Powell, Y. Li, C. Cheng, S. A. Wickline and K. L. Wooley, *Biomacromolecules*, 2008, **9**, 2826.
- 15 (a) Z.-X. Jiang and Y. B. Yu, *Tetrahedron*, 2007, **63**, 3982; (b) X. Yue, M. B. Taraban, L. L. Hyland and Y. B. Yu, *J. Org. Chem.*, 2012, **77**, 8879.
- 16 (a) M. S. Wiehn, S. D. Lindell and S. Bräse, *J. Comb. Chem.*, 2009, **11**, 960; (b) M. S. Wiehn, D. Fűrnis and S. Bräse, *J. Comb. Chem.*, 2009, **11**, 982; (c) M. Döbele, S. Vanderheiden, N. Jung and S. Bräse, *Angew. Chem.*, 2010, **122**, 6122; *Angew. Chem., Int. Ed.*, 2010, **49**, 5986; (d) M. Döbele, M. S. Wiehn and S. Bräse, *Angew. Chem.*, 2011, **123**, 11737; *Angew. Chem., Int. Ed.*, 2011, **50**, 11533; (e) A. Hafner and S. Bräse, *Adv. Synth. Catal.*, 2011, **353**, 3044; (f) A. Hafner and S. Bräse, *Angew. Chem.*, 2012, **124**, 3773; *Angew. Chem., Int. Ed.*, 2012, **51**, 3713; (g) A. Hafner and S. Bräse, *Adv. Synth. Catal.*, 2013, **355**, 996; (h) A. Hafner, A. Bihlmeier, M. Nieger, W. Kloppe and S. Bräse, *J. Org. Chem.*, 2013, **78**, 7938; (i) A. Hafner, T. J. Feuerstein and S. Bräse, *Org. Lett.*, 2013, **15**, 3468; (j) A. Hafner, T. S. Fischer and S. Bräse, *Eur. J. Org. Chem.*, 2013, 7996; (k) D. K. Kölmel, B. Rudat, D. M. Braun, C. Bednarek, U. Schepers and S. Bräse, *Org. Biomol. Chem.*, 2013, **11**, 3954; (l) S. Zhong, A. Hafner, C. Hussal, M. Nieger and S. Bräse, *RSC Adv.*, 2015, **5**, 6255.
- 17 For a review: (a) E. F. V. Scriven and K. Turnbull, *Chem. Rev.*, 1988, **88**, 297; (b) S. Bräse, C. Gil, K. Knepper and V. Zimmermann, *Angew. Chem.*, 2005, **117**, 5320; *Angew. Chem., Int. Ed.*, 2005, **44**, 5188; (c) S. Bräse and K. Banert, *Organic Azides – Syntheses and Applications*, Wiley, Chichester, 2009; (d) J. E. Hein and V. V. Fokin, *Chem. Soc. Rev.*, 2010, **39**, 1302; (e) B. T. Worrell, J. A. Malik and V. V. Fokin, *Science*, 2013, **340**, 457; (f) R. Berg and B. F. Straub, *Beilstein J. Org. Chem.*, 2013, **9**, 2715; (g) N. V. Sokolova and V. G. Nenajdenko, *RSC Adv.*, 2013, **3**, 16212; (h) D. K. Kölmel, N. Jung and S. Bräse, *Aust. J. Chem.*, 2014, **67**, 328.
- 18 (a) G. Franc and A. Kakkar, *Chem. Commun.*, 2008, 5267; (b) M. Suchý, R. Bartha and R. H. E. Hudson, *RSC Adv.*, 2103, **3**, 3249; (c) A. Thirunarayanan, S. Raja, G. Mohanraj and P. Rajakumar, *RSC Adv.*, 2014, **4**, 41778.
- 19 X. Zhao, W. Y. Ng, K.-C. Lau, A. E. C. Collis and I. T. Horváth, *Phys. Chem. Chem. Phys.*, 2012, **14**, 3909.
- 20 B. Sookcharoenpinyo, E. Klein, Y. Ferrand, D. B. Walker, P. R. Brotherhood, C. Ke, M. P. Crump and A. P. Davis, *Angew. Chem.*, 2012, **124**, 4664; *Angew. Chem., Int. Ed.*, 2012, **51**, 4586.
- 21 For a review: (a) R. Chinchilla and C. Nájera, *Chem. Soc. Rev.*, 2011, **40**, 5084; (b) M. Schilz and H. Plenio, *J. Org. Chem.*, 2012, **77**, 2798; (c) S. Bräse, in *Organometallics in Synthesis, Third Manual*, ed. M. Schlosser, Wiley, Hoboken, 2013, p. 777.
- 22 ESI†
- 23 G. Pasut and F. M. Veronese, *Adv. Drug Delivery Rev.*, 2009, **61**, 1177.
- 24 (a) K. J. Thurecht, I. Blakey, H. Peng, O. Squires, S. Hsu, C. Alexander and A. K. Whittaker, *J. Am. Chem. Soc.*, 2010, **132**, 5336; (b) B. E. Rolfe, I. Blakey, O. Squires, H. Peng, N. R. B. Boase, C. Alexander, P. G. Parsons, G. M. Boyle, A. K. Whittaker and K. J. Thurecht, *J. Am. Chem. Soc.*, 2014, **136**, 2413.
- 25 (a) S. B. L. Vollrath, D. Fűrnis, U. Schepers and S. Bräse, *Org. Biomol. Chem.*, 2013, **11**, 8197; (b) D. Fűrnis, T. Mack, F. Hahn, S. B. L. Vollrath, K. Koroniak, U. Schepers and S. Bräse, *Beilstein J. Org. Chem.*, 2013, **9**, 56; (c) D. K. Kölmel, A. Hörner, F. Rönicke, M. Nieger, U. Schepers and S. Bräse, *Eur. J. Med. Chem.*, 2014, **79**, 231; (d) D. Althunon, F. Rönicke, D. Fűrnis, J. Quan, I. Wellhöfer, N. Jung, U. Schepers and S. Bräse, *Org. Biomol. Chem.*, 2015, **13**, 4226.