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# Pushing the limits for enzyme-based membrane-less hydrogen fuel cells – achieving useful power and stability†

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The performance characteristics of simple enzyme-based membrane-less hydrogen fuel cells running on non-explosive H<sub>2</sub>-rich air mixtures have been established using an adjustable test bed that allows multiple unit cells to operate in series or parallel. Recent advances with '3D' electrodes constructed from compacted porous carbon loaded with hydrogenase (anode) and bilirubin oxidase (cathode) have been extended in order to scale up fuel cell power to useful levels. One result is an appealing 'classroom' demonstration of a model house containing small electronic devices powered by H<sub>2</sub> mixed with a small amount of air. The 3D electrodes work by greatly increasing catalyst loading (at both the anode and cathode) and selectively restricting the access of O<sub>2</sub> (relative to H<sub>2</sub>) to enzymes embedded in pores at the anode. The latter property raises the possibility of using standard hydrogenases that are not O<sub>2</sub>-tolerant: however, experiments with such an enzyme reveal good short-term performance due to restricted O<sub>2</sub> access, but low long-term stability because the root cause of O<sub>2</sub> sensitivity has not been addressed. Hydrogenases that are truly O<sub>2</sub> tolerant must therefore remain the major focus of any future enzyme-based hydrogen fuel cell technology.

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## Introduction

Isolation and characterization of O<sub>2</sub>-tolerant [NiFe]-hydrogenases that are able to catalyze H<sub>2</sub> oxidation in the presence of O<sub>2</sub> has stimulated new concepts in hydrogen fuel cells.<sup>1–7</sup> Not only do enzymes prove the feasibility of replacing platinum metal catalysts with ones derived from abundant elements (indeed the active sites of enzymes are more active than Pt<sup>8–11</sup>) but their high selectivity lends itself to simple, membrane-less fuel cells that could be miniaturized, thus compensating for the large footprint of the catalyst.<sup>12–15</sup> Our focus is therefore on membrane-less cells. Although commercial development is, even optimistically, a long way ahead (depending on achieving high levels of miniaturization, long-term stability and identification of a niche use) orders-of-magnitude improvements have been made since the demonstrations, in 2005 and 2006, of an enzyme-based membrane-less H<sub>2</sub> fuel cell running in the presence of CO<sup>16</sup> or on a non-explosive H<sub>2</sub>-weak-air-rich mixture (3% H<sub>2</sub> in air).<sup>17</sup> The H<sub>2</sub>-weak-air-rich mixture proved problematic because even an O<sub>2</sub>-tolerant hydrogenase is overwhelmed by O<sub>2</sub> under low-load conditions,<sup>18</sup> but no such problem was encountered when using a non-explosive H<sub>2</sub>-rich-air-weak mixture (80% H<sub>2</sub> in air).

Since 2010, power densities and lifetimes of enzyme-based, membrane-less H<sub>2</sub> fuel cells have advanced in two stages. It was demonstrated that a reasonably stable power density of 0.1 mW cm<sup>–2</sup> (based on the anode area) could be achieved by covalently attaching a hydrogenase and a 'blue' Cu oxidase to graphite electrodes modified with multi-walled carbon nanotubes.<sup>19</sup> Then, in a subsequent development, Xu and Armstrong used 3D compacted mesoporous carbon electrodes with re-proportioned anode/cathode areas to increase the power density up to 1.7 mW cm<sup>–2</sup> (based on geometric area of anode) with an extended half-life of approximately one week.<sup>20</sup> Porous 3D electrodes (we use 'CPC' as a general abbreviation for different types of compacted porous carbon) increase the number of reaction sites per unit geometric electrode area, thus substantially raising power density.<sup>21–27</sup> The enzyme molecules permeate deeply, leading to a greatly enhanced catalyst loading, while the porous channels are interconnected and open to the surface.<sup>20</sup> Varying the relative sizes of anode and cathode further increases power output which is otherwise limited by the low O<sub>2</sub> level.

Oxygen tolerance of the hydrogenase that is used as the anodic electrocatalyst is an essential requirement for all enzyme-based H<sub>2</sub> fuel cells, even if a membrane is used (due to unavoidable O<sub>2</sub> crossover). All hydrogenases characterized to date are inactivated or permanently damaged by O<sub>2</sub> and/or high potentials,<sup>28</sup> and even robustly O<sub>2</sub>-tolerant [NiFe]-hydrogenases such as Hydrogenase-1 (Hyd-1) from *Escherichia coli*, have a lower, yet stable, activity when O<sub>2</sub> is present, provided the

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potential is not taken too high.<sup>3</sup> The O<sub>2</sub>-tolerance of [NiFe]-hydrogenases originates from the following features:<sup>29</sup> (i) availability of additional electron sources close to the active site that can be immediately recruited to ensure the complete reduction of an attacking O<sub>2</sub> molecule to harmless water, resulting in formation of an oxidized Ni(III)-OH species (a 'resting state') known as Ni-B, (ii) fast recovery of activity by rapid one-electron reduction of Ni-B, upon which the enzyme re-joins the catalytic cycle, (iii) limiting the access of O<sub>2</sub> to the active site (*via* restrictions in internal gas channels). The fractional current level  $f$  (measured relative to the current for 100% H<sub>2</sub>) that can be maintained under O<sub>2</sub> is given by the expression  $f = \text{rate}(\text{act}) / [\text{rate}(\text{act}) + \text{rate}(\text{inact})]$  where  $\text{rate}(\text{inact})$  is the rate of inactivation by O<sub>2</sub> and  $\text{rate}(\text{act})$  is the rate of reactivation.<sup>30</sup> The term  $\text{rate}(\text{inact})$  increases with increasing O<sub>2</sub> concentration whereas  $\text{rate}(\text{act})$  increases as the potential is lowered, and both  $\text{rate}(\text{inact})$  and  $\text{rate}(\text{act})$  are affected in a more complex way, by the concentration of H<sub>2</sub>. Hence  $f$  is increased by impeding O<sub>2</sub> access to the active site and increasing the electron availability for reactivation.

It is thus timely to consider further interesting implications of developing membrane-less H<sub>2</sub> fuel cells with CPC electrodes. We have therefore investigated the minimum configuration, using multiples of simple unit cells arranged in parallel or series, that is required to produce useful power (*i.e.* sufficient to run small electronic devices). A test bed has been constructed, in which parallel and series connections of sandwich-like electrode stacks can be varied. As explained below, powers are reported in mW per total cell volume as well as in mW per area (geometric electrode area), although no attempt has been made to optimize volume power density (see below) as we focused on having sufficient space to manipulate the cells easily. Secondly, we have addressed the issue of whether CPC electrodes can usefully improve the O<sub>2</sub> tolerance of O<sub>2</sub>-sensitive (standard) hydrogenases (which usually have higher activity) an idea that complements, in a timely way, a recent paper by Plumeré *et al.*, who found that embedding an O<sub>2</sub>-sensitive hydrogenase into a viologen-based redox hydrogel significantly improved its O<sub>2</sub> tolerance.<sup>31</sup>

## Results and discussion

### Test bed structure

The test bed shown in Fig. 1A(i) consists of 16 rectangular stainless steel plates (1.5 cm × 5 cm) separated by 0.5 cm, with CPC electrode plates (thickness 0.3 mm) attached to the side of each plate by conductive adhesive. Conceptually, there are three components – unit cell, parallel multi-cell, and series multi-cell. The unit cell (denoted 1 × 1; total volume 3.75 cm<sup>3</sup>) contains one anodic plate and one cathodic plate (Fig. 1A(ii)). Enzymes were applied to anode and cathode as described in Methods. To balance the anodic and cathodic activities in the non-explosive H<sub>2</sub>-air mixture, the area ratio of anode to cathode was re-proportioned in each unit cell: thus the cathodes (modified with bilirubin oxidase (BOD) from *Myrothecium verrucaria*) were 4 × 1.5 cm while the anodes (modified with Hyd-1) were smaller, at 0.8 × 1.5 cm. The additional 1 cm of exposed

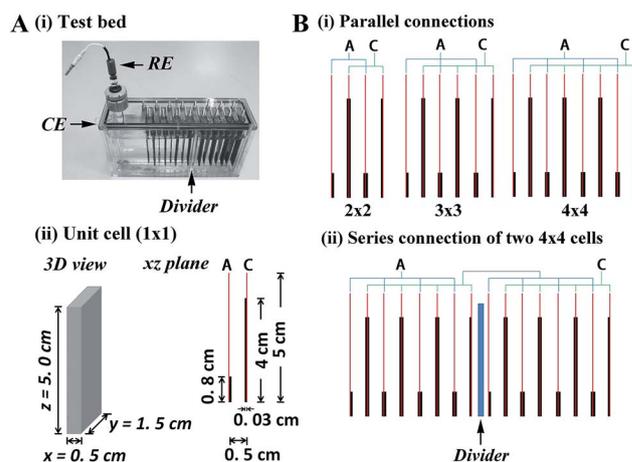


Fig. 1 A. (i) Photograph of the test bed structure (RE = reference electrode, CE = counter electrode). (ii) The scheme for the unit cell (A = anode, C = cathode). B. Schemes for (i) the parallel multi-cells (2 × 2, 3 × 3, 4 × 4) and (ii) the series multi-cell (A = anode, C = cathode).

stainless steel plate (electrochemically inert) above the cathode was required so that the H<sub>2</sub>-air mixture could be bubbled into each cell without losing electrolyte. In the parallel multi-cells, 2, 3 or 4 anodic plates and 2, 3 or 4 cathodic plates were connected (denoted 2 × 2, 3 × 3, 4 × 4) to contain 3, 5 or 7 unit cells, respectively (Fig. 1B(i)). For a series multi-cell, a divider was used to separate the electrolyte solutions (Fig. 1B(ii)). Reductive activation of Hyd-1 was carried out electrochemically, as described previously,<sup>19</sup> by connecting the anodes to a potentiostat and inserting reference and counter electrodes at one side of the divider (Fig. 1A(i)). Before activating Hyd-1, the solution was raised above the central divider to connect all the cells; then after activation was complete, the level was lowered to isolate the cells series-wise. Volume power density was calculated by dividing total power by the total volume of the cells (3.75 cm<sup>3</sup> for each unit cell).

### 3D CPC electrodes

The 3D CPC electrodes were made by compacting graphite (G), multi-walled carbon nanotubes (MCNT), or mixtures of both (G/MCNT). Fig. 2(i) shows typical N<sub>2</sub> adsorption-desorption isotherms of compacted 100% G, 60/40 G/MCNT, and 100% MCNT electrodes. The three N<sub>2</sub> adsorption-desorption isotherms each show a characteristic Type IV curve, that is, a hysteresis loop related to capillary condensation in mesopores (pore diameter: 2–50 nm).<sup>32</sup> Using the Brunauer-Emmett-Teller (BET) method,<sup>33</sup> the specific surface areas of compacted 100% G, 60/40 G/MCNT and 100% MCNT electrodes were calculated to be 15.6 m<sup>2</sup> g<sup>-1</sup>, 154 m<sup>2</sup> g<sup>-1</sup> and 256 m<sup>2</sup> g<sup>-1</sup>, respectively. Fig. 2(ii and iii) shows the relative and cumulative pore size distributions of the three 3D CPC electrodes based on the Barrett-Joyner-Halenda (BJH) model.<sup>34</sup> The pore volumes of compacted 100% G, 60/40 G/MCNT and 100% MCNT electrodes are 0.052 cm<sup>3</sup> g<sup>-1</sup>, 0.56 cm<sup>3</sup> g<sup>-1</sup> and 1.2 cm<sup>3</sup> g<sup>-1</sup>, respectively. The volumes of those pores having diameters larger than 10 nm are 0.034 cm<sup>3</sup> g<sup>-1</sup>, 0.42 cm<sup>3</sup> g<sup>-1</sup> and 1.0 cm<sup>3</sup> g<sup>-1</sup>, representing 65%,



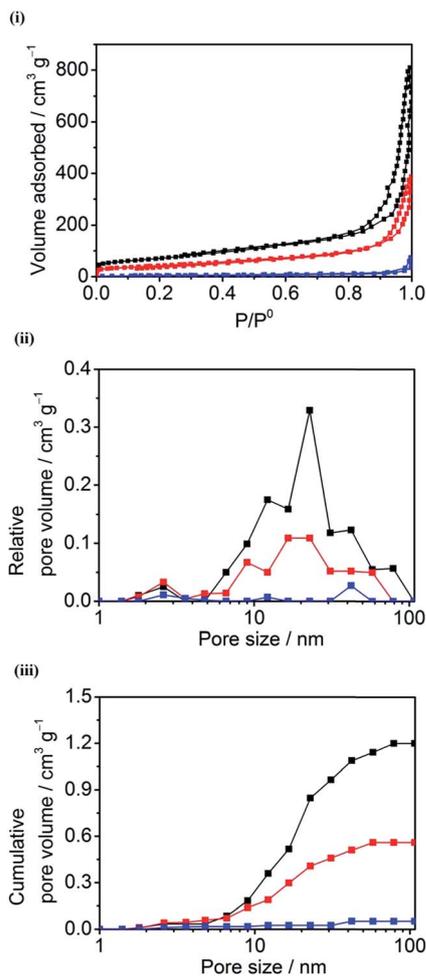


Fig. 2 (i)  $N_2$  adsorption–desorption isotherms and (ii & iii) the corresponding pore size distribution curves of the compacted 100% MCNT (black), 60/40 G/MCNT (red), and 100% G (blue) electrodes.

75% and 85% of the total pore volumes of compacted 100% G, 60/40 G/MCNT and 100% MCNT electrodes, respectively. Based upon maximum molecular diameters of Hyd-1 (12 nm) and BOD (8 nm),<sup>35,36</sup> it follows that most pores of the CPC electrodes are large enough to be permeable to both enzymes. Electrodes with greater pore volumes will therefore have a higher density of electrocatalytic sites.

Assuming complete uptake of enzymes into the porous channels of CPC electrodes, the total amounts of hydrogenase (RMM approximately 100 kDa) and BOD (RMM approximately 50 kDa) deposited on respective electrodes are 1.2 nmol and 18 nmol, respectively (see Methods). A calculation indicates that an atomically flat surface occupied by a monolayer of Hyd-1 (assuming a footprint  $12 \times 12$  nm) and BOD (assuming a footprint  $8 \times 8$  nm) would extend to approximately  $1000 \text{ cm}^2$  and  $7000 \text{ cm}^2$  respectively, to be compared with geometric anode and cathode areas of  $1.2 \text{ cm}^2$  and  $6 \text{ cm}^2$ . The enzyme loading is therefore equivalent to that of about  $10^3$  monolayers in both cases, the vast excess being drawn into the pores of the electrode.

### Parallel and series connections

Fig. 3(i) shows power curves of unit cell and parallel multi-cells constructed with Hyd-1-modified compacted 60/40 G/MCNT anodes and BOD-modified compacted 60/40 G/MCNT cathodes. For most of the following experiments, 60/40 G/MCNT was used instead of 100% MCNT – the reason being that this composition worked sufficiently well for Hyd-1 and is a cheaper option. For both Hyd-1 and BOD, catalytic current densities measured by cyclic voltammetry scaled in the order 100% MCNT > 60/40 G/MCNT  $\gg$  100% G. The maximum power outputs of the  $1 \times 1$ ,  $2 \times 2$ ,  $3 \times 3$  and  $4 \times 4$  cells are  $0.61 \pm 0.03$  mW,  $1.80 \pm 0.04$  mW,  $2.98 \pm 0.08$  mW, and  $4.14 \pm 0.09$  mW, respectively. Fig. 3(ii) shows the dependences of power and volume power density on the cell voltage of the series multi-cell (two  $4 \times 4$  cells connected in series, denoted  $2[4 \times 4]$ ) and its individual unit (a single  $4 \times 4$  cell). Volume power density has been used to give an idea of the power/size ratio that is achieved, noting that the test bed has not been designed to optimise this factor but, rather, to have easy access to the components: it is likely that specific attention to engineering could still result in large increases in volume power density. The open-circuit voltage of this series stack is 2.09 V, almost twice as large as that of a single  $4 \times 4$  cell (1.05 V). The maximum power and volume power density of the  $2[4 \times 4]$  series multi-cell are  $7.84 \pm 0.11$  mW and  $0.149 \pm 0.002 \text{ mW cm}^{-3}$ , respectively, at a cell voltage of 1.22 V.

An exhibit was constructed to demonstrate that the test bed with series  $2[4 \times 4]$  configuration, is capable of powering electronics from a non-explosive 78%  $H_2$ –22% air mixture at  $20^\circ \text{C}$ .

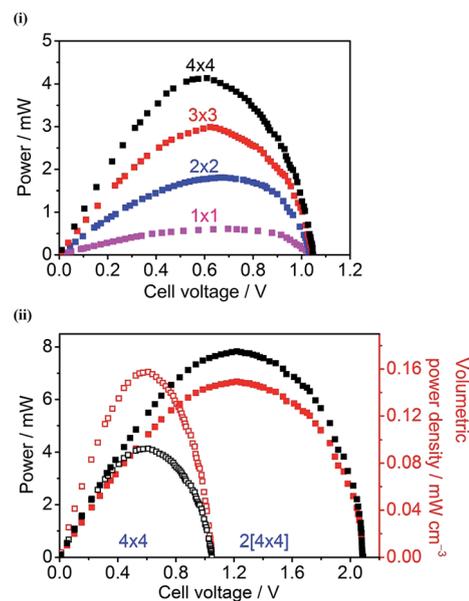


Fig. 3 (i) Power curves of the unit cell ( $1 \times 1$ ) and the parallel multi-cells ( $2 \times 2$ ,  $3 \times 3$ ,  $4 \times 4$ ). (ii) Power curves of the series multi-cell  $2[4 \times 4]$  (■) and one  $4 \times 4$  cell (□); dependence of volume power density on the cell voltage of the  $2[4 \times 4]$  (■) and one  $4 \times 4$  (□). All the curves were recorded in the test bed using the compacted 60/40 G/MCNT electrodes under a 78%  $H_2$ –22% air atmosphere at  $20^\circ \text{C}$ . Other conditions: 0.1 M sodium phosphate buffer, pH 6.0.



The festive 'Hydrogen House' shown in Fig. 4 contains five red LED tree lights and a miniature electronic clock. A light dependent resistor probe (LDR)<sup>37</sup> was used to monitor changes in LED light intensity with time, in order to establish the working stability of the fuel cell under continuous power output. The light intensity showed no decrease over the course of 8 hours (Fig. S1†).

### Improving the O<sub>2</sub> tolerance of hydrogenases

The following experiments were carried out to determine if the H<sub>2</sub> oxidation activity of hydrogenases under aerobic conditions is improved if the enzyme is embedded in a 3D CPC electrode as compared to adsorption at a conventional pyrolytic graphite edge (PGE) electrode. Fig. 5 compares the current–time profiles for H<sub>2</sub> oxidation by Hyd-1, previously fully activated, at a CPC electrode (60/40 G/MCNT) and at a PGE electrode. In each case, the electrode potential was held at +0.206 V vs. SHE (a more oxidizing value, and thus more demanding conditions, than used in earlier experiments<sup>3,30</sup>) and the H<sub>2</sub> level in non-explosive H<sub>2</sub>–air mixtures was varied periodically.

The Hyd-1/CPC electrode showed no loss in H<sub>2</sub> oxidation current over the initial period of 1200 s under 100% H<sub>2</sub>. Then, upon switching the gas supply to 89% H<sub>2</sub>–11% air, the current dropped rapidly to 58% of its original value ( $f = 0.58$ ) and remained constant at that value for a further 1200 s. Switching back to 100% H<sub>2</sub> resulted in a rapid and complete recovery of the original current. After a further 1200 s, during which the same stability was observed, the gas supply was switched to 78% H<sub>2</sub>–22% air and the current dropped rapidly to 42% of the

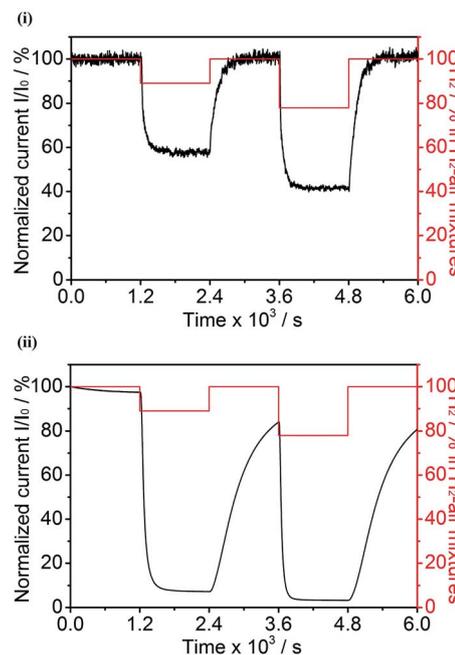


Fig. 5 Chronoamperograms of Hyd-1 adsorbed on 3D and '2D' carbon electrodes: (i) compacted 60/40 G/MCNT electrode and (ii) conventional PGE electrode. Electrode potential in each case is +0.206 V vs. SHE. The H<sub>2</sub>–air mixture was varied at 1200 s periods, as indicated in red (100% H<sub>2</sub> → 89% H<sub>2</sub>–11% air → 100% H<sub>2</sub> → 78% H<sub>2</sub>–22% air → 100% H<sub>2</sub>). Other conditions: 0.1 M sodium phosphate buffer at pH 6.0 and 20 °C.

original value ( $f = 0.42$ ), remaining stable at this value for 1200 s. Finally, the gas mixture was switched back to 100% H<sub>2</sub> and complete recovery of the original current was observed. Exactly the same procedures were used to measure the performance of the conventional Hyd-1/PGE electrode. The slow decrease in current measured in the 100% H<sub>2</sub> atmosphere is usually described as 'film loss'. After 1200 s, switching the gas mixture from 100% H<sub>2</sub> to 89% H<sub>2</sub>–11% air resulted in a rapid drop in current to approximately 7.5% of the original value ( $f = 0.075$ ). Switching the gas supply back to 100% H<sub>2</sub> then produced an increase in current that was much slower than observed for the Hyd-1/CPC electrode; in fact, recovery was incomplete even after 1200 s, when the gas supply was switched to 78% H<sub>2</sub>–22%, resulting in the current dropping rapidly to approximately 3.5% of the original value ( $f = 0.035$ ). Finally, restoring the gas supply to 100% H<sub>2</sub> resulted again in a slow increase in activity.

These results show that the great advantage afforded by embedding a hydrogenase in a 3D carbon electrode extends to an improved ability to function when the gas supply contains O<sub>2</sub>. Hyd-1 is inherently O<sub>2</sub>-tolerant, which means that if O<sub>2</sub> is added to the H<sub>2</sub> gas supply, only the oxidized 'resting' Ni–B state is formed:<sup>30</sup> Ni–B is reactivated in a potential-dependent process, although the rate is slow at +206 mV – accounting generally for why the  $f$  values are quite low. Qualitatively, the behaviour and levels of activity obtained at different O<sub>2</sub> levels for the Hyd-1/PGE electrode are fully consistent with previous observations. For example, in recent experiments in which the

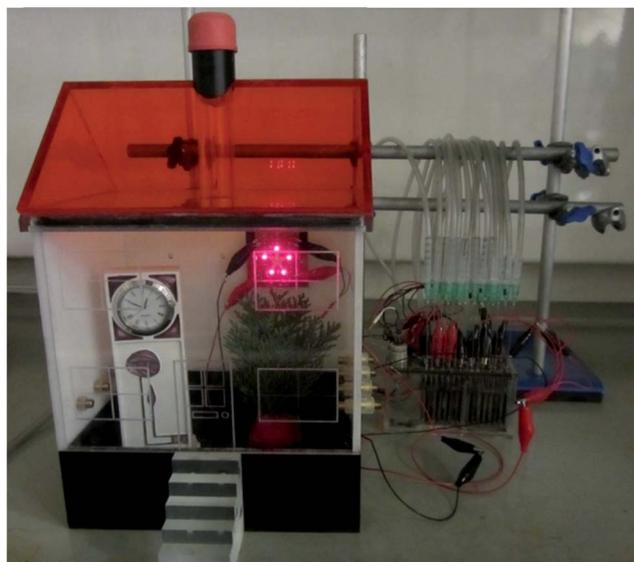


Fig. 4 Photograph of the working test bed powering the 'Hydrogen House' containing five red LEDs (the forward voltage is 1.6 V) and a 'Charles Rennie Macintosh' miniature clock (the LEDs and clock are all connected in parallel to the test bed). Working conditions: test bed uses the parallel and series connections, in which the two parallel stacks (4 × 4) cells are connected in series; compacted 60/40 G/MCNT electrodes, 78% H<sub>2</sub>–22% air mixture at 20 °C, electrolyte is 0.1 M sodium phosphate buffer at pH 6.0.



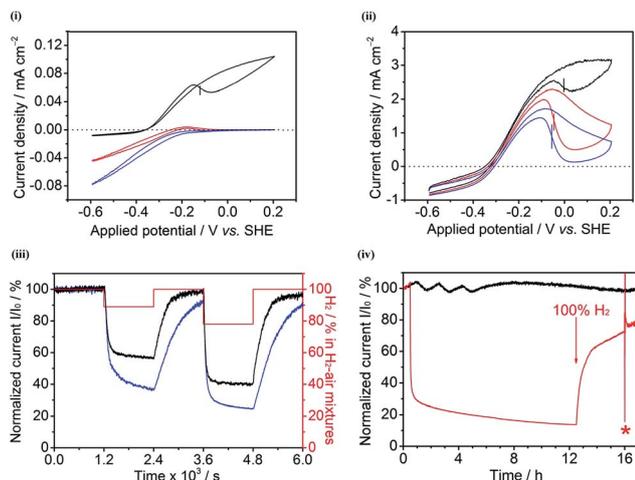
electrode potential was held at 0 mV vs. SHE at 30 °C, upon changing from 100% H<sub>2</sub> to 10% H<sub>2</sub>-10% O<sub>2</sub>-80% N<sub>2</sub>, the current decrease corresponded to  $f = 0.35$ ; the effect of the less favourable H<sub>2</sub>/O<sub>2</sub> ratio (which increases rate(inact)) being offset by the far more reducing potential (which increases rate(act)).<sup>30</sup>

Regardless of whether a porous CPC electrode improves the performance of hydrogenase or BOD electrochemistry in the absence or presence of O<sub>2</sub>, an obvious rationale so far is that the electrode contains so much enzyme that losses, whatever their origin, have little effect on current or stability. We therefore examined whether a 3D CPC electrode could allow an O<sub>2</sub>-sensitive [NiFe]-hydrogenase, Hydrogenase-2 (Hyd-2) from *Escherichia coli*, to function under aerobic conditions. Fig. 6(i and ii) show cyclic voltammograms of Hyd-2 adsorbed at a conventional PGE electrode and at a 3D CPC electrode under conditions of 100% H<sub>2</sub>, 89% H<sub>2</sub>-11% air and 78% H<sub>2</sub>-22% air mixtures. For reasons that will become evident, 100% MCNT was used for the 3D CPC electrode. Under aerobic conditions, Hyd-2 adsorbed at a PGE electrode (Fig. 6(i)) shows no H<sub>2</sub>-oxidation activity under 78% H<sub>2</sub>-22% air and just a tiny window of activity is evident under 89% H<sub>2</sub>-11% air. The negative current at potentials < -0.2 V which completely dominates the voltammetry and almost doubles between 11% air and 22% air, is due to direct reduction of O<sub>2</sub> at the electrode: a

similar observation is made with or without hydrogenase present. In contrast, Hyd-2 embedded in the 3D CPC electrode maintains substantial H<sub>2</sub> oxidation activity Fig. 6(ii) provided the potential is not taken above -0.1 V, where reversible inactivation takes place. The negative current at potentials < -0.3 V increases only by a very small degree as the O<sub>2</sub> level is raised, first as 11% air, then as 22% air. The profound contrast in behaviour, from voltammetry completely dominated by direct O<sub>2</sub>-reduction (PGE) to voltammetry completely dominated by catalytic H<sub>2</sub> oxidation (3D CPC), must reflect not only the increased density of enzyme on the 3D CPC electrode but also the inability of O<sub>2</sub> to permeate to sites at which it would be detected electrochemically. Viewed in a different way, the hydrogenase serves as a probe by which to gauge how effectively the porous structure filters out O<sub>2</sub> relative to H<sub>2</sub>.

The ability of Hyd-2 to operate in the presence of O<sub>2</sub> also varies between the different 3D CPC electrodes. Fig. 6(iii) shows chronoamperograms of Hyd-2 at compacted 100% MCNT and 60/40 G/MCNT electrodes, recorded at an electrode potential of +6 mV vs. SHE. It should be noted that a more negative potential was used for the Hyd-2 experiments (compared to Hyd-1) because the characteristic potential ( $E_{\text{switch}}$ ) at which Hyd-2 is reactivated from the Ni-B state is >0.2 V more negative than for Hyd-1.<sup>3</sup> Switching the gas supply from 100% H<sub>2</sub> to an 89% H<sub>2</sub>-11% air mixture results in an immediate drop in current that is significantly more pronounced for the 60/40 G/MCNT CPC electrode than for the 100% MCNT CPC electrode: furthermore, the resulting suppressed current measured with the 60/40 G/MCNT CPC electrode continues to drop at a faster rate than for the 100% MCNT CPC electrode. When, after 1200 s under 89% H<sub>2</sub>-11% air, the gas supply was switched back to 100% H<sub>2</sub>, recovery was faster and more complete for the 100% MCNT CPC electrode. A second exposure to O<sub>2</sub>, this time using 78% H<sub>2</sub>-22% air for 1200 s, revealed the same pattern but with lower currents, as expected. Finally, Fig. 6(iv) shows the result of an experiment to compare the current produced by a Hyd-2-60/40 G/MCNT CPC electrode over a continuous period of 17 h under 100% H<sub>2</sub> with the current recorded over a 12 h period under a 78% H<sub>2</sub>-22% air mixture, after which the gas supply was restored to 100% H<sub>2</sub>. The current under 100% H<sub>2</sub> remains steady, whereas the switch to 78% H<sub>2</sub>-22% air causes an immediate drop in current that is followed by a continual slow decrease. Upon restoring a 100% H<sub>2</sub> supply, only partial recovery is observed, which is barely affected by switching the electrode potential to -0.194 V for 1 minute: since this procedure should drive the activation of any remaining Ni-B, the observation shows that O<sub>2</sub> causes a significant degree of unrecoverable damage to Hyd-2.

The results shown in Fig. 6(i and ii) show clearly that the electrochemical contribution of enzyme-catalyzed H<sub>2</sub> oxidation relative to O<sub>2</sub>-derived inactivation and direct O<sub>2</sub> reduction is massively enhanced at the 100% MCNT CPC electrode compared to that at a PGE electrode. The observation provides firm evidence that the 100% MCNT CPC electrode restricts O<sub>2</sub> transport, and therefore that the improved O<sub>2</sub> tolerance in CPC electrodes is not attributable only to the buffering effect of high enzyme loading or to electrochemical removal of O<sub>2</sub>. In the



**Fig. 6** Cyclic voltammograms of Hyd-2 at (i) a PGE electrode and (ii) a compacted 100% MCNT electrode in 100% H<sub>2</sub> (black), 89% H<sub>2</sub>-11% air (red) and 78% H<sub>2</sub>-22% air (blue) mixtures. The  $E_{\text{switch}}$  values are indicated by vertical bars.<sup>38</sup> Other conditions: 0.1 M sodium phosphate buffer at pH 6.0 and 20 °C, scan rate: 1 mV s<sup>-1</sup>. (iii) Chronoamperograms of Hyd-2 at compacted 100% MCNT (black) and 60/40 G/MCNT (blue) electrodes recorded at +0.006 V vs. SHE. The H<sub>2</sub>-air mixtures with different ratios are periodically applied (every 1200 s), as indicated in red (100% H<sub>2</sub> → 89% H<sub>2</sub>-11% air → 100% H<sub>2</sub> → 78% H<sub>2</sub>-22% air → 100% H<sub>2</sub>). (iv) Chronoamperograms of Hyd-2 at compacted 60/40 G/MCNT electrodes exposed to (black) 100% H<sub>2</sub> for 17 h, recorded at +0.006 V vs. SHE, and (red) first exposed to 100% H<sub>2</sub> (0.5 h) → 78% H<sub>2</sub>-22% air (12 h) → 100% H<sub>2</sub> (3.5 h) at +0.006 V vs. SHE, then after the potential was stepped to -0.194 V vs. SHE for 1 min (\*) under 100% H<sub>2</sub>, followed by returning the potential to +0.006 V vs. SHE for 1 h under 100% H<sub>2</sub>. Other conditions: 0.1 M sodium phosphate buffer at pH 6.0 and 20 °C.



H<sub>2</sub>-air mixtures, the main constituents are H<sub>2</sub>, N<sub>2</sub> and O<sub>2</sub>. The mean free paths ( $\lambda$ ) for H<sub>2</sub>, N<sub>2</sub> and O<sub>2</sub> at 293 K and 101.325 kPa are calculated to be 122 nm, 66 nm and 71 nm, respectively, according to the expression:<sup>39</sup>

$$\lambda = \frac{RT}{\sqrt{2}\pi d^2 LP}$$

where  $R$  is the universal gas constant,  $T$  is temperature,  $d$  is the collision diameter of the gas molecule (H<sub>2</sub>: 2.71 Å, N<sub>2</sub>: 3.70 Å, O<sub>2</sub>: 3.55 Å),<sup>40</sup>  $L$  is the Avogadro constant, and  $P$  is pressure. As shown in Fig. 2(iii), the volumes of pores having diameters smaller than 70 nm make up >95% and 100% of the total pore volumes of compacted 100% MCNT and 60/40 G/MCNT electrodes, respectively. Here, N<sub>2</sub> is assumed to be inert, so only H<sub>2</sub> and O<sub>2</sub> are considered. Effusion occurs when the pore size is smaller than the mean free path of a gas molecule,<sup>39</sup> so the pore size distributions of 3D CPC electrodes fulfil the effusion condition for H<sub>2</sub> and O<sub>2</sub>. According to Graham's law of effusion, the rate of effusion of a gas is inversely proportional to the square root of its molecular mass.<sup>39</sup> Therefore, H<sub>2</sub> effuses 4 times as fast as O<sub>2</sub>, which results in more H<sub>2</sub> molecules passing through effusion orifices per unit time. As a H<sub>2</sub>-air mixture effuses through many pores, the proportion of H<sub>2</sub> will gradually increase. The 3D structure allows for effusion in depth and permits the H<sub>2</sub>-air mixture to undergo such effusions with sufficient frequency to separate H<sub>2</sub> from O<sub>2</sub>, thus Hyd-1 and Hyd-2 molecules that are sufficiently buried are effectively exposed only to H<sub>2</sub>. In other words, the 3D porous structure acts as an O<sub>2</sub> filter, protecting deeply buried hydrogenase molecules against both rapid, easily-reversed inactivation (to Ni-B) and the progressive, irreversible inactivation that is normally observed for standard, O<sub>2</sub>-sensitive hydrogenases.

Volumes of those pores having diameters between 10 nm and 70 nm are 0.959 cm<sup>3</sup> g<sup>-1</sup> and 0.422 cm<sup>3</sup> g<sup>-1</sup>, respectively, for the compacted 100% MCNT and 60/40 G/MCNT electrodes. Within the range of pore sizes of 10–70 nm, the 3D CPC electrode with a larger pore volume also has more effusion pores through which the H<sub>2</sub>-air mixture can pass, thus resulting in a higher level of separation of H<sub>2</sub> and O<sub>2</sub>. The compacted 100% MCNT electrode therefore allows Hyd-2 to maintain a higher level of apparent O<sub>2</sub> tolerance than the compacted 60/40 G/MCNT electrode (Fig. 6(iii)). Similarly, increasing the thickness of the 3D porous electrode can increase effusion pores and thus increase the apparent O<sub>2</sub> tolerance of hydrogenases. Hyd-2 embedded in the compacted 100% MCNT electrode with a thickness of 0.3 mm maintains a higher H<sub>2</sub> oxidation current under aerobic conditions than in a 100% MCNT electrode of thickness 0.2 mm (Fig. S2†). The 100% MCNT electrodes having a thickness greater than 0.3 mm were prone to crumbling, so the thickness was restricted to 0.3 mm.

### O<sub>2</sub>-sensitive hydrogenase as fuel cell catalyst

Finally, a full cell experiment was carried out in which O<sub>2</sub>-sensitive Hyd-2 was used as the anodic catalyst on 100% MCNT, a combination that our investigations now suggested might work. The experiment was conducted in an 89% H<sub>2</sub>-11% air

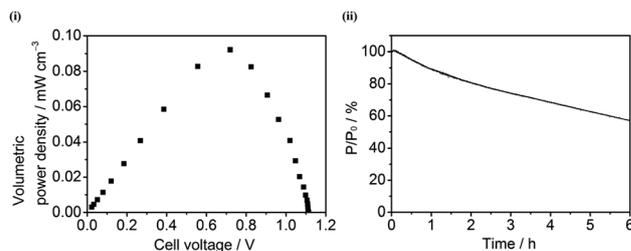


Fig. 7 (i) Dependence of volume power density on the cell voltage and (ii) dependence of stability of power (%) with time at an applied constant potential of 0.75 V in one unit cell composed of a Hyd-2-modified compacted 100% MCNT anode and a BOD-modified compacted 100% MCNT cathode under an 89% H<sub>2</sub>-11% air mixture at 20 °C. Other conditions: 0.1 M sodium phosphate buffer, pH 6.0.

mixture at 20 °C. Fig. 7(i) shows the dependence of volume power density on the cell voltage of the unit cell composed of a Hyd-2-modified compacted 100% MCNT anode and a BOD-modified 100% MCNT cathode. The open-circuit voltage of the fuel cell is 1.114 V, a value somewhat higher than that measured for the similar unit cell constructed with Hyd-1 in 60/40 G/MCNT. The improved voltage observed for the Hyd-2 fuel cell is consistent with it behaving as a reversible catalyst, in contrast to Hyd-1 and other O<sub>2</sub>-tolerant [NiFe]-hydrogenases that display a marked onset overpotential for H<sub>2</sub> oxidation at neutral pH.<sup>16–20</sup> The maximum volume power density is 0.09 mW cm<sup>-3</sup> at a cell voltage of 0.72 V. Fig. 7(ii) shows the time dependence of power stability for the Hyd-2/100% MCNT fuel cell at an applied constant potential of 0.75 V under a 89% H<sub>2</sub>-11% air mixture at 20 °C: only about 55% of the power was retained after 6 h. The long-term performance under aerobic conditions is therefore far lower than achieved with Hyd-1, both as measured previously (54% of activity remaining after 7 days (ref. 20)) and as evident from the working experiment with Hyd-1 shown in Fig. S1† which shows 100% retention of LED intensity after 8 h.

## Conclusions

These studies have shown how the power provided by a membrane-less enzyme fuel cell acting on a mixed H<sub>2</sub>-air feed is easily scaled up to useful levels. Stable power, readily gained by exploiting the high enzyme loading possible with '3D' porous carbon electrodes, is multiplied in terms of current or voltage using a simple stack design. There is considerable room for improvement in volume power density through miniaturization *via* careful attention to engineering. In support of a recent paper by Plumeré *et al.*, we find that the ability of hydrogenases to operate effectively in the presence of O<sub>2</sub> is greatly increased by incorporation into a matrix, although our experiments show that the major effect in the case of a 3D porous electrode is the lower effusion rate of O<sub>2</sub> relative to H<sub>2</sub>, rather than electrochemical removal of O<sub>2</sub> or enhanced rates of reactivation. Although a lower effusion rate of O<sub>2</sub> would also impact on the current limiting electrocatalysis at the BOD-modified porous cathode, this aspect is addressed, empirically, by appropriate



scaling of anode and cathode sizes. Despite the advantage afforded by the porous electrode, an O<sub>2</sub>-sensitive hydrogenase is still eventually overwhelmed, because the root cause of long-term inactivation is not access of O<sub>2</sub> to the active site but the inability of the enzyme to deal with each attack with 100% fidelity.<sup>30</sup>

## Methods

### Materials

Graphite and multi-walled carbon nanotubes were purchased from Sigma-Aldrich and used without further purification. The membrane-bound hydrogenases from *E. coli*, Hyd-1 and Hyd-2, were isolated and purified as described previously.<sup>3</sup> Bilirubin oxidase (BOD) was purchased from Amano Enzyme Inc. and further purified by hydrophobic interaction chromatography with a HiTrap Phenyl HP hydrophobic column (GE Healthcare). Sodium dihydrogen orthophosphate and disodium hydrogen orthophosphate were purchased from Fisher Scientific and used to prepare the sodium phosphate buffer solution. The PGE electrodes were constructed from pyrolytic graphite blocks (Momentive Performance Materials Ltd). Conductive silver paint (Agar Scientific) was used to glue the CPC electrodes to the stainless steel plates of the test bed. All aqueous solutions were prepared using deionized water from a Milli-Q water system (18 MΩ cm).

### Construction of the test bed

The fuel cell test bed consisted of one compartment constructed from polycarbonate with a partial divider also made of polycarbonate. A reference electrode (Ag/AgCl) and a Pt mesh counter electrode were positioned at one end to use for electrochemical measurements under potential control. The main part of the test bed consisted of 16 stainless-steel plates as supports for the 3D CPC electrodes. The CPC electrodes were attached to both sides of each plate using conductive silver adhesive. The exposed metal plate areas not covered by the CPC electrodes were painted to insulate against the electrolyte solution. The top edges of the plates were bent and fixed to the ceiling of the test bed by screws and nuts, which also served as electrical connectors. Such an assembly also allowed for convenient dismantling of the plates. The anodic and cathodic plates were fixed in alternating mode. Gas supply into each unit cell was achieved through gas inlets positioned close to the bottom of the cell and positioned midway between anode and cathode.

### Fabrication of 3D CPC electrodes

A certain quantity of G, MCNT, or mixtures of the two carbon allotropes was placed in a pellet die (Specac) and subjected to a pressure of 0.5 tonnes using a hydraulic press (Specac). The quantities of G, MCNT, or G/MCNT mixtures that were used depended on what thickness of the 3D CPC electrode was required. For a typical thickness of 0.3 mm, (i) the compacted 100% G electrode required 47 mg of G; (ii) the compacted 60/40 G/MCNT electrode required 12 mg of G and 8 mg of MCNT; (iii)

the compacted 100% MCNT electrode required 11 mg of MCNT. For a 0.2 mm thickness, the compacted 100% MCNT electrode required 7 mg of MCNT. The CPC discs were then cut to size for the required geometric area and attached to the stainless steel plates as described above. After each plate had been constructed, 30 μL of Hyd-1 (or Hyd-2) (4 mg mL<sup>-1</sup>) was applied to the anode side and 150 μL of BOD (6 mg mL<sup>-1</sup>) was applied to the cathode side. The enzyme concentrations and quantities were chosen after evaluating how oxidation and reduction current densities varied with amounts of Hyd-1 and BOD respectively (Fig. S3†). The enzyme-modified CPC electrodes were subsequently placed in a cold room (4 °C) for 1 h to allow the enzymes to permeate into the electrodes before starting the electrochemical characterization and fuel cell tests. The molecular quantities of Hyd-1 (RMM = 100 kDa) and BOD (RMM = 50 kDa) used to modify the CPC electrodes are therefore 1.2 nmol and 18 nmol, respectively. For experiments conducted with conventional PGE electrodes, 5 μL of the enzyme solution (Hyd-1/Hyd-2: 4 mg mL<sup>-1</sup>, BOD: 6 mg mL<sup>-1</sup>) was pipetted onto the surface of the freshly polished electrode, which was subsequently placed in a cold room (4 °C) for 1 h and rinsed with deionized water before the electrochemical experiments.

### Instrumentation

Electrochemical measurements were performed using an Ivium potentiostat (CompactStat, Ivium Technologies). All potentials were adjusted to the standard hydrogen electrode (SHE) using the relationship  $E_{\text{SHE}} = E_{\text{Ag/AgCl}} + 0.206 \text{ V}$  (25 °C).<sup>41</sup> Power measurements were obtained by varying the applied load and recording the potential in each case (Keithley 195A digital voltmeter). The ratios of H<sub>2</sub> to air were controlled by two mass flow controllers (Sierra Instruments). A Sorptomatic 1990 instrument (CE Instruments) was used to acquire N<sub>2</sub> adsorption-desorption isotherms at 77 K, and BET surface areas were calculated from the linear part of BET plots. Pore size distribution plots were obtained by using the BJH model.<sup>34</sup> The light intensity of the red LED was monitored by a cadmium sulphide (CdS) light dependent resistor connected to a digital multimeter.

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