



Cite this: *Polym. Chem.*, 2015, **6**, 5827

Received 15th June 2015,
Accepted 15th June 2015

DOI: 10.1039/c5py00924c

www.rsc.org/polymers

Engineering thermoresponsive polyether-based nanogels for temperature dependent skin penetration†

M. Asadian-Birjand,^a J. Bergueiro,^a F. Rancan,^b J. C. Cuggino,^a R.-C. Mutihac,^a K. Achazi,^a J. Dervedde,^c U. Blume-Peytayi,^b A. Vogt^b and M. Calderón^{a*}

Highly biocompatible thermoresponsive nanogels (tNGs) based on oligo ethylene glycol (OEG) as thermoresponsive unit and dendritic polyglycerol (dPG) as cross-linker, were precisely engineered in terms of size and volume phase transition temperature (VPTT). Preliminary uptake studies into human skin were realized to show the temperature-dependent internalization behavior of these systems.

Transdermal delivery has many advantages in comparison to intravenous or oral administration like patient amenability, reduced side effects, and enhanced activity by avoiding hydrolytic enzymes or hard environmental/metabolic conditions.^{1–5} Nonetheless efficient transdermal administration of certain hydrophilic molecules of biomedical interest (peptides, proteins, hydrophilic drugs, *etc.*) is still a challenge due to the protecting barrier function performed by the highly organized stratum corneum (SC) in the skin.^{6–9} Several techniques based on the disruption of the SC's ordered structure have been developed, of which microneedles,¹⁰ laser ablation¹¹ or ultrasound¹² are the most commonly used. But all of these techniques present limitations and disadvantages highlighting the need for a more sophisticated way of action in transdermal delivery.

Numerous studies suggest that selective skin penetration and specific targeting effects can be achieved by nanoparticles with controlled size, lipophilicity, surface charge, and stability.¹³ Recently new soft synthetic, polymer based nanometric architectures have emerged that can potentially solve the transdermal delivery challenge.^{14,15} New developments have highlighted polymeric nanoparticles as possible drug delivery devices for transdermal applications by accumulation, *e.g.*, by

improving drug penetration, taking advantage of the reservoir function in hair follicle (HF) openings, and by performing controlled release.^{16,17} Of all the polymeric nanoparticles, nanogels (NGs) have gained the most attention. NGs are aqueous dispersions of hydrogel particles in the nanometer range and are formed with physically or chemically crosslinked polymer network chains.^{18,19} NGs have already shown many interesting intrinsic properties, such as high water content, soft nature, flexibility, cell and tissue compatibility, and excellent water dispersion/solubility.

When responsive polymers are used as building blocks for NGs, these architectures can largely respond to external factors like temperature,²⁰ pH,²¹ light,²² electrical fields,⁹ *etc.*, and change their properties as a result.²³ Particularly, thermoresponsive polymers react to environmental changes, *i.e.*, temperature can change NG properties like hydrophobic/hydrophilic balance, softness, size, and aggregation behavior, which are all important for biomedical applications.^{24–26} Because of these changes upon environmental triggers, the drug encapsulation and release profiles of these systems have been well studied as drug delivery devices.^{18,27,28} The same concept has already been explored for transdermal drug delivery systems,^{21,29–32} but, to the best of our knowledge, no study of a temperature dependent dermal uptake of thermoresponsive NGs has been reported yet. Nevertheless, optimization of synthetic methodologies that allow a reproducible synthetic control over size, shape, and physicochemical properties is still a milestone in the synthesis of NGs.^{22,33,34}

More precisely, the development of NGs for transdermal applications calls for an optimization of synthetic routes that enable control over the nanoparticle properties like size, flexibility, deformability, and drug loading capacity. Among the typical methodologies for preparing NGs, free radical dispersion/precipitation polymerization is particularly attractive because it yields particles with sizes between 0.1 to 15 μm in a single reaction step.^{25,35,36} Therefore this methodology was utilized in this work to synthesize tNGs through a free radical polymerization of acrylated dendritic polyglycerol (dPG-Ac) and ethylene glycol methacrylates, as shown in Fig. 1a and Table S1 (ESI†).

^aInstitut für Chemie und Biochemie, Freie Universität Berlin, Takustrasse 3, 14195 Berlin, Germany. E-mail: marcelo.calderon@fu-berlin.de; Fax: +49-30-838-459368

^bClinical Research Center of Hair and Skin Science, Department of Dermatology and Allergy, Charité-Universitätsmedizin Berlin, Charitéplatz 1, 10117 Berlin, Germany

^cCharité – Universitätsmedizin Berlin, Institut für Laboratoriumsmedizin, Klinische Chemie und Pathobiochemie, CVK, Augustenburger Platz 1, 13353 Berlin, Germany

†Electronic supplementary information (ESI) available. See DOI: 10.1039/c5py00924c



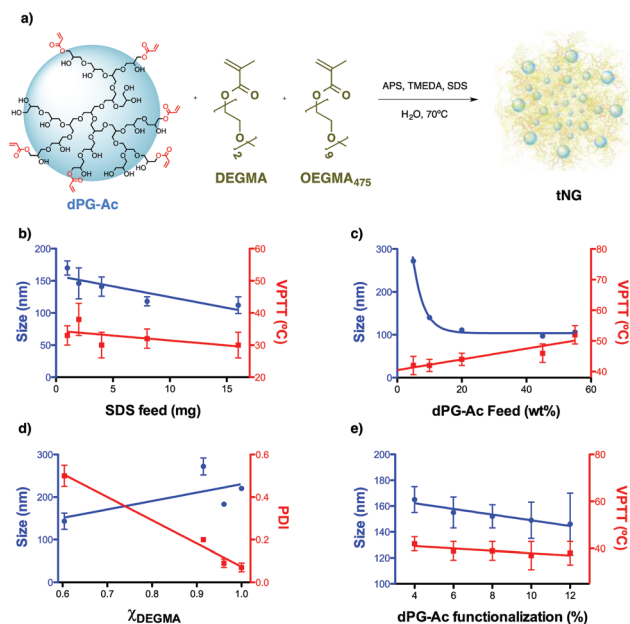


Fig. 1 (a) General scheme of the nanogelification reaction and parameters screened for size and VPTT tuning: (b) SDS feed, (c) dPG-Ac feed, (d) monomer ratio, and (e) dPG-Ac functionalization degree. All data are shown in detail in Table S1 (ESI[†]).

dPG-Ac as a water soluble crosslinker and di(ethylene glycol) methyl ether methacrylate (DEGMA) and oligo ethylene glycol methacrylate (OEGMA475) as the thermoresponsive monomers, were chosen, respectively, as building blocks to combine their inherent multifunctionality, responsiveness, and biocompatibility.^{37–41}

It is difficult to predict the size at which a particle will pass the skin barrier. It is known from the literature that the biggest particle size is 10 nm for not easily deformable materials and higher (above 50 nm) for soft and flexible particles.⁴² Therefore we aimed for tNGs in the size range of 50–200 nm and with a VPTT of 36–40 °C, slightly higher than the temperature of healthy skin.⁴³ It is expected that such tNGs would not easily penetrate the lipophilic SC barrier below their transition temperature due to their high hydrophilicity. But once the tNGs surpass their VPTT, for instance in inflamed skin areas, their lipophilicity will change from a hydrophilic to a hydrophobic state. This change is expected to improve the interaction with the hydrophobic structures of the SC that would result in a better penetration in the skin. To achieve tNGs with these requirements, we fine tuned their size and VPTT by varying the synthetic parameters like sodium dodecyl sulphate (SDS) feed, dPG-Ac feed, monomer ratio, and dPG-Ac functionalization degree (Fig. 1b–e, Table S1, ESI[†]).

The size of the tNGs was tuned first by modifying the SDS feed prior to polymerization. As expected,⁴⁴ increasing the SDS feed from 1 to 16 mg decreased the tNG's size from 170 to 112 nm, respectively. VPTT showed a slightly decreasing tendency with the SDS feed increasing. With a size close to

150 nm and a transition temperature above 30 °C, a value of 2 mg SDS feed was therefore set as the first fixed parameter (Fig. 1b).

It was found that increasing the dPG-Ac feed effectively decreased the tNG diameter within the range of 272–97 nm (Fig. 1c). This was attributed to the higher crosslinker density within the network, which consequentially decreased the primary chain length. Moreover, the greater degree of crosslinking may have provoked an increase in the polymer network's elastic tension, which conspicuously reduced the swelling of the NGs and, as a result, their hydrodynamic diameter. The dPG-Ac feed ratio within the tNGs was confirmed by ¹H-NMR spectroscopy, which showed an increase in the ethylene glycol chain proton signals for the tNGs with a lower dPG-Ac feed ratio (Fig. S1, ESI[†]). This result indicated a positive correlation of the crosslinker composition inside the tNG with the dPG-Ac feed. Moreover, VPTT was also affected by the dPG-Ac feed. Since VPTT depends on the balance between hydrophobicity and hydrophilicity within the tNG, increasing the dPG-Ac composition also increased the network's hydrophilicity. This caused the phase transition temperature to rise until a dPG-Ac feed ratio of 55 wt%, at which point the VPTT was no longer measurable (Fig. 1c). A compromise between a size below 200 nm and a low transition temperature around 40 °C was found at a dPG-Ac feed ratio of 10 wt%.

Varying the monomer mole fraction (χ_{DEGMA}) had a big influence on the polydispersity of the yielded tNGs. Copolymerization with dPG-Ac (7% acrylation degree) only showed an acceptable polydispersity value when monomer mol fractions near 1 were used ($\chi_{\text{DEGMA}} = 1$, PDI = 0.07) (Fig. 1d). As a result, the synthetic control over tNG's VPTT could be investigated by tailoring the hydrophobic/hydrophilic balance on the macromolecular crosslinker with its acrylation degree.

As expected,³⁵ when the functionalization degree of the dPG-Ac (4–12%) increased, the size of the particles tended to decrease from 165 to 146 nm. Moreover, when the crosslinker's degree of functionalization rose, the VPTT moderately descended from 42 to 37 °C. A higher dPG-Ac functionalization degree increased the network's hydrophobicity and caused the VPTT to slightly drop (Fig. 1e). In summary, a dPG acrylation degree of around 10% was found to be ideal for temperature-dependant skin penetration purposes and yielded tNGs with sizes below 200 nm and convenient transition temperatures between 37–42 °C.

The four screened parameters were used to tune the size and transition temperatures of the synthesized thermoresponsive system in order to allow the design of tNGs that would facilitate the shuttling of active compounds across the skin barrier. Temperature differences between healthy and diseased skin or external heating were expected to improve skin penetration of tNGs due to the change in physicochemical properties at different temperatures.

Consequently, rhodamine B (Rhd) labelled tNGs were synthesized to be trackable in *ex vivo* human skin experiments. The above-mentioned parameters were applied for the Rhd labelled tNGs synthesis: 2 mg as SDS feed, $\chi_{\text{DEGMA}} = 1$, 10 wt%



DPG-Ac feed, and 9% DPG-Ac acrylation degree. A post-synthetic labelling of tNG with Rhodamine (Rh) through ester formation coupling reaction only yielded low conjugation efficiencies and NGs with dye ratios in the sub $\mu\text{mol}_{\text{Rh}}/\text{mg}_{\text{NG}}$ range (data not shown). Therefore, a 2/8 mixture of dye labelled DPG-Ac (Rh-DPG-Ac) and unlabelled DPG-Ac was employed to introduce the dye with a total crosslinker feed value of 10 wt%. Adding an extra crosslinking unit like Rhodamine-DPG-Ac, the system's hydrophilicity was increased. Consequently, the size for Rhodamine-tNG (NG20) changed from the expected 150 to 80 nm. The VPTT, however, did not significantly differ (36 °C) and remained in the desired range when only 2 wt% feed of the hydrophilic crosslinker Rhodamine-DPG-Ac was used (ESI†). Moreover reversible thermoresponsive behavior was demonstrated when NG20 was analysed *via* turbidimetry (Fig. S2†). As a negative control for skin internalization experiments, a non-thermo-responsive NG was synthesized by mainly modifying the cross-linking content (ESI†). By employing only Rhodamine-DPG-Ac as the crosslinker and a monomer ratio of $\chi_{\text{DEGMA}} = 0.91$, NG21 was obtained with a size of 78 nm, which was comparable to NG20 (Table S1, ESI†), but did not show a phase transition below 75 °C (Fig. 2c). Both NGs were intensively characterized (Fig. 2 and ESI†). The characteristic monomer and crosslinker peaks as well as the lack of vinyl signals in the $^1\text{H-NMR}$ spectra revealed a complete

polymerization. The sizes were investigated with dynamic light scattering (DLS), transmission electron microscopy (TEM), and liquid/fluid atomic force microscopy (AFM) techniques, which gave correlative sizes as well as a low degree of dispersion (Fig. 2d–g, Fig. S3, ESI†). Dye concentration was determined by UV-Vis spectroscopy which yielded values of 8.26 and 68.3 $\text{mmol}_{\text{Rh}}/\text{mg}_{\text{NG}}$ for NG20 and NG21, respectively. The bigger dye amount per NG in the non-thermo-responsive NG was obtained due to the use of only Rhodamine-DPG-Ac as crosslinker.

To test the tNGs' biocompatibility, the cytotoxicity in a human keratinocyte cell line (HaCaT) and fibroblast cell line (NIH3T3) was analyzed. Metabolic activity (MTS assay) and real time cell analysis (RTCA) showed IC_{50} values between 10 and 33 mg mL^{-1} , respectively (Fig. S4, ESI†). These high tolerable doses demonstrate tNGs' great cytocompatibility and potential for topical applications. Cell uptake experiments in A549 cell line revealed an endosomal uptake pathway on cancer cells (Fig. S5, ESI†).

Rhodamine-tNG (NG20) was employed to study temperature-dependent transdermal penetration into full-thickness excised human skin after topical application. Non-thermo-responsive Rhodamine-NG (NG21) was used as the negative control. Both NG samples were applied on the surface of the excised human skin at the same dye concentration of 41.3 mM. The skin penetration profile was analyzed on skin sections after 4 hours incubation at two different temperatures, below (4 °C) and above (37 °C) the VPTT of NG20. Dye fluorescence intensity was observed on the skin surface and in SC of all analyzed cryosections, regardless of the type of NGs and the incubation temperature (Fig. 3). Similar skin penetration profiles were also found in our previous studies with more rigid nanoparticles like polystyrene and silica oxide.^{45,46} However, fluorescence intensity was also detected in the epidermis, that indicated the presence of tNGs in viable skin (Fig. 3b). Mean fluorescence intensity (MFI) analysis confirmed that there were no significant differences observed in the analyzed SC sections between the thermo-responsive and non-thermo-responsive NGs, regardless of the incubation temperature. On the contrary, higher fluorescence intensities were measured in viable skin for the NG20 samples incubated at 37 °C in comparison to the same NGs at 4 °C incubation temperature and NG21 samples at 37 °C (Fig. 3). Even though diffusion processes might be slowed down at 4 °C, there were no remarkable differences observed between MFI values of NG20 and NG21 in the viable skin regions. These observations reflect the temperature-dependent conformational changes in the tNGs and the resultant alteration of their physicochemical properties like increased hydrophobicity, which allowed them to penetrate the SC and translocate to the epidermis. Accumulation of NGs in the HF canal was also investigated. Higher fluorescence intensity was observed in the HF canal for NG20 incubated at 37 °C than for the same sample incubated at 4 °C (Fig. S6, ESI†). No statistical analysis could be performed because of the low amount of HF in the analyzed skin sample.

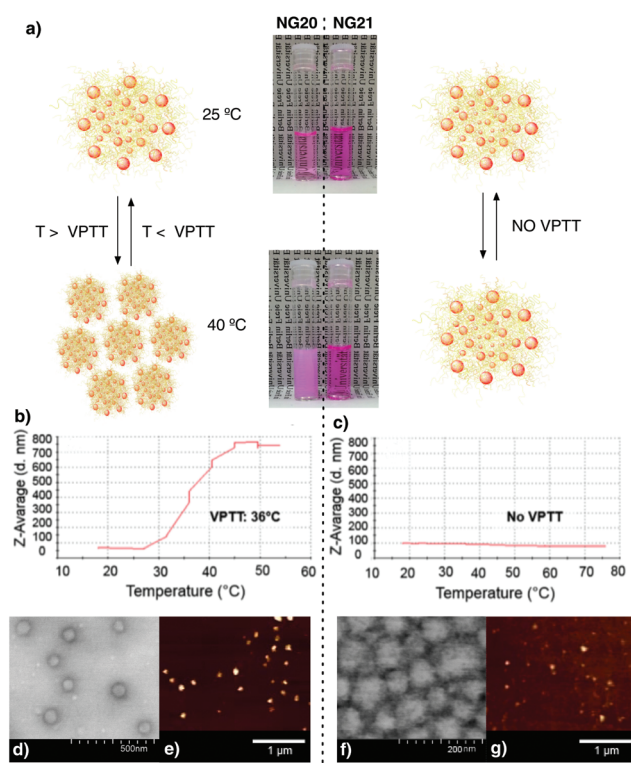


Fig. 2 (a) Images of thermo-responsive (NG20, left) and non thermo-responsive (NG21, right) NGs at two temperatures, below (25 °C) and above (40 °C) the VPTT of NG20. DLS temperature-trend measurements of NG20 (b) and NG21 (c). TEM (d and f) and AFM (e and g) images of NG20 and NG21.



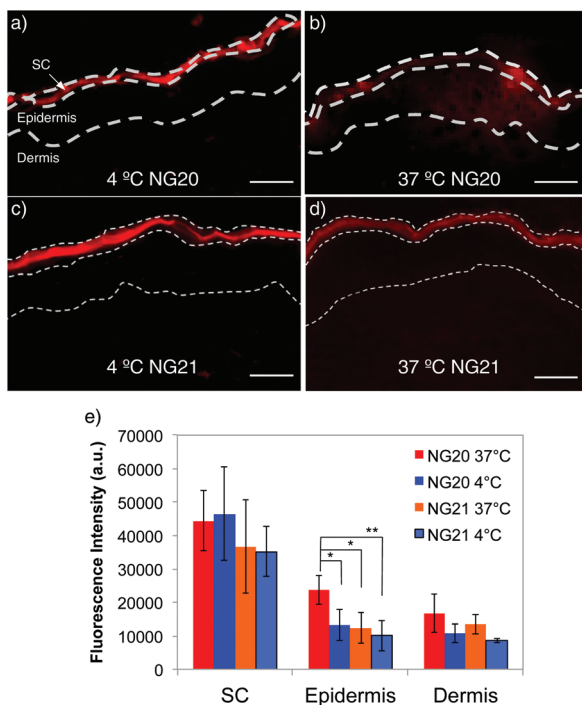


Fig. 3 Fluorescence images of skin internalization at 4 and 37 °C of NG20 (a and b) and NG21 (c and d) scale bars = 50 μ m. At least 20 sections per sample were analyzed. In approximately 25% of sections fluorescence intensity was detected also in the epidermis. Control images with non treated skin and free Rhd are shown in Fig. S11 (ESI[†]). (e) Average MFI value of skin sections where fluorescence intensity was detected in both SC and epidermis (ESI[†]) (* p < 0.05, ** p < 0.01).

Conclusions

The dispersion/precipitation polymerization methodology proved to be robust for the synthesis of tNGs. Sizes and VPTTs were modified by changing the surfactant concentration (SDS feed), crosslinker (dPG-Ac) acrylation degree and feed, and OEGMA feed ratio. By varying the studied synthetic parameters, Rhd labeled NGs were synthesized with required sizes and VPTT. Application of tNGs on human skin explants showed, for the first time, a temperature-dependent interaction of soft nanoparticles with the skin barrier and HF canals. Dye labeled NGs easily penetrated into SC and occasionally in the epidermis. The thermoresponsive nanogel NG20 had a better penetration in the epidermis than the non-thermoresponsive one when incubation temperatures were above the VPTT. These first experiments strengthen the rationale for further development of tNGs for applications in dermatotherapy and transdermal drug delivery. While in this study we investigated the influence of temperature on the skin penetration profile of tNGs, in ongoing studies, we are investigating tNGs loaded with non-covalently bound dyes and drugs in order to evaluate the influence of thermal triggers on the tNGs release properties.

Acknowledgements

We gratefully acknowledge financial support from the Bundesministerium für Bildung und Forschung (BMBF) through the NanoMatFutur award (13N12561). Moreover, we acknowledge financial support from the Sonderforschungsbereich 1112 (<http://www.sfb1112.de/>), Projects A04 and C04. Dr. Julian Bergueiro acknowledges Dahlem Research Center for a Dahlem International Network PostDocs fellowship. Furthermore, we thank Prof. Dr. Rainer Haag for fruitful discussion, Pamela Winchester for proof reading the manuscript, and Dr. Maria Molina, Katrin Michel, and Dr. Christoph Böttcher for performing AFM and TEM measurements.

Notes and references

- 1 Y. Tahara, S. Honda, N. Kamiya, H. Piao, A. Hirata, E. Hayakawa, T. Fujii and M. Goto, *J. Controlled Release*, 2008, **131**, 14–18.
- 2 B. J. Thomas and B. C. Finnin, *Drug Discovery Today*, 2004, **9**, 697–703.
- 3 L. B. Lopes, C. M. Brophy, E. Furnish, C. R. Flynn, O. Sparks, P. Komalavilas, L. Joshi, A. Panitch and M. V. L. B. Bentley, *Pharm. Res.*, 2005, **22**, 750–757.
- 4 M. R. Prausnitz, S. Mitragotri and R. Langer, *Nat. Rev. Drug Discovery*, 2004, **3**, 115–124.
- 5 K. Fukushima, A. Ise, H. Morita, R. Hasegawa, Y. Ito, N. Sugioka and K. Takada, *Pharm. Res.*, 2010, **28**, 7–21.
- 6 K. C. Madison, *J. Invest. Dermatol.*, 2003, **121**, 231–241.
- 7 B. W. Barry, *Nat. Biotechnol.*, 2004, **22**, 165–167.
- 8 D. Lee, J. N. Ashcraft, E. Verploegen, E. Pashkovski and D. A. Weitz, *Langmuir*, 2009, **25**, 5762–5766.
- 9 G. Cevc and U. Vierl, *J. Controlled Release*, 2010, **141**, 277–299.
- 10 M. R. Prausnitz, *Adv. Drug Delivery Rev.*, 2004, **56**, 581–587.
- 11 W.-R. Lee, S.-C. Shen, K.-H. Wang, C.-H. Hu and J.-Y. Fang, *J. Pharm. Sci.*, 2002, **91**, 1613–1626.
- 12 A. Azagury, L. Khoury, G. Enden and J. Kost, *Adv. Drug Delivery Rev.*, 2014, **72**, 127–143.
- 13 F. Rancan, A. Todorova, S. Hadam, D. Papakostas, E. Luciani, C. Graf, U. Gernert, E. Rühl, B. Verrier, W. Sterry, U. Blume-Peytavi and A. Vogt, *Eur. J. Pharm. Biopharm.*, 2012, **80**, 76–84.
- 14 J. Khandare, M. Calderón, N. M. Dagia and R. Haag, *Chem. Soc. Rev.*, 2012, **41**, 2824–2848.
- 15 R. Haag and F. Kratz, *Angew. Chem., Int. Ed.*, 2006, **45**, 1198–1215.
- 16 J. Lademann, F. Knorr, H. Richter and S. Jung, *J. Innov. Opt. Health Sci.*, 2014, **8**, 1530004–1530012.
- 17 F. Rancan, D. Papakostas, S. Hadam, S. Hackbarth, T. Delair, C. Primard, B. Verrier, W. Sterry, U. Blume-Peytavi and A. Vogt, *Pharm. Res.*, 2009, **26**, 2027–2036.
- 18 M. Asadian-Birjand, A. Sousa-Herves, D. Steinhilber, J. C. Cuggino and M. Calderón, *Curr. Med. Chem.*, 2012, **19**, 5029–5043.



- 19 Q. Wang, Y. Zhao, Y. Yang, H. Xu and X. Yang, *Colloid Polym. Sci.*, 2007, **285**, 515–521.
- 20 J. Ramos, A. Imaz, J. Callejas-Fernández, L. Barbosa-Barros, J. Estelrich, M. Quesada-Pérez and J. Forcada, *Soft Matter*, 2011, **7**, 5067–5082.
- 21 S. Mangalathillam, N. S. Rejinold, A. Nair, V.-K. Lakshmanan, S. V. Nair and R. Jayakumar, *Nanoscale*, 2012, **4**, 239–250.
- 22 A. V. Kabanov and S. V. Vinogradov, *Angew. Chem., Int. Ed.*, 2009, **48**, 5418–5429.
- 23 G. Chen and A. S. Hoffman, *Nature*, 1995, **373**, 49–52.
- 24 M. Giubudagian, M. Asadian-Birjand, D. Steinhilber, K. Achazi, M. Molina and M. Calderón, *Polym. Chem.*, 2014, **5**, 6909–6913.
- 25 M. Molina, M. Giubudagian and M. Calderón, *Macromol. Chem. Phys.*, 2014, **215**, 2414–2419.
- 26 M. Molina, M. Asadian-Birjand, J. Balach, J. Bergueiro, E. Miceli and M. Calderón, *Chem. Soc. Rev.*, 2015, DOI: 10.1039/C5CS00199D.
- 27 L. Zha, B. Banik and F. Alexis, *Soft Matter*, 2011, **7**, 5908–5916.
- 28 J. Bergueiro and M. Calderón, *Macromol. Biosci.*, 2015, **15**, 183–199.
- 29 M. Kim, B. Jung and J.-H. Park, *Biomaterials*, 2011, **33**, 668–678.
- 30 P. P. Shah, P. R. Desai, A. R. Patel and M. S. Singh, *Biomaterials*, 2012, **33**, 1607–1617.
- 31 W. Il Choi, J. H. Lee, J.-Y. Kim, J.-C. Kim, Y. H. Kim and G. Tae, *J. Controlled Release*, 2012, **157**, 272–278.
- 32 N. H. A. Samah and C. M. Heard, *Int. J. Pharm.*, 2013, **453**, 630–640.
- 33 Y. Sasaki and K. Akiyoshi, *Chem. Rec.*, 2010, **10**, 366–376.
- 34 A. L. Sisson and R. Haag, *Soft Matter*, 2010, **6**, 4968–4975.
- 35 J. C. Cuggino, C. I. Alvarez I, M. C. Strumia, P. Welker, K. Licha, D. Steinhilber, R.-C. Mutihac and M. Calderón, *Soft Matter*, 2011, **7**, 11259–11266.
- 36 C. K. Ober and K. P. Lok, *Macromolecules*, 1987, **20**, 268–273.
- 37 R. K. Kainthan, S. R. Hester, E. Levin, D. V. Devine and D. E. Brooks, *Biomaterials*, 2007, **28**, 4581–4590.
- 38 M. Calderón, M. A. Quadir, S. K. Sharma and R. Haag, *Adv. Mater.*, 2010, **22**, 190–218.
- 39 J.-F. Lutz, A. Özgür Akdemir and A. Hoth, *J. Am. Chem. Soc.*, 2006, **128**, 13046–13047.
- 40 H. Zhang and M. W. Grinstaff, *Macromol. Rapid Commun.*, 2014, **35**, 1906–1924.
- 41 A. Thomas, S. S. Mueller and H. Frey, *Biomacromolecules*, 2014, **15**, 1935–1954.
- 42 M. Schneider, F. Stracke, S. Hansen and U. F. Schaefer, *Derm.-Endocrinol.*, 2009, **1**, 197–206.
- 43 E. F. J. Ring and K. Ammer, *Physiol. Meas.*, 2012, **33**, R33–R46.
- 44 N. Sanson and J. Rieger, *Polym. Chem.*, 2010, **1**, 965–977.
- 45 A. Vogt, B. Combadiere, S. Hadam, K. M. Stieler, J. Lademann, H. Schaefer, B. Autran, W. Sterry and U. Blume-Peytavi, *J. Invest. Dermatol.*, 2006, **126**, 1316–1322.
- 46 F. Rancan, Q. Gao, C. Graf, S. Troppens, S. Hadam, S. Hackbarth, C. Kembuan, U. Blume-Peytavi, E. Rühl, J. Lademann and A. Vogt, *ACS Nano*, 2012, **6**, 6829–6842.

