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Microwave-assisted synthesis of 3-aminobenzo[*b*]-thiophene scaffolds for the preparation of kinase inhibitors†

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Microwave irradiation of 2-halobenzonitriles and methyl thioglycolate in the presence of triethylamine in DMSO at 130 °C provides rapid access to 3-aminobenzo[*b*]thiophenes in 58–96% yield. This transformation has been applied in the synthesis of the thieno[2,3-*b*]pyridine core motif of LIMK1 inhibitors, the benzo[4,5]thieno[3,2-*e*][1,4]diazepin-5(2H)-one scaffold of MK2 inhibitors and a benzo[4,5]thieno[3,2-*d*]-pyrimidin-4-one inhibitor of the PIM kinases.

Introduction

Benzothiophenes are naturally-occurring heterocycles, found in petroleum deposits in their simplest form but also discovered recently as a motif in more complex glycosides isolated from the roots of *E. griffissii*.¹ Benzothiophenes are important components of organic semiconductors due to their potential for elongated and highly delocalised electronic structures.^{2,3} Substituted benzothiophenes have also found application in drug discovery as highly-privileged structures and valuable building blocks in medicinal chemistry, being incorporated into tubulin polymerisation inhibitors,^{4,5} acetyl-CoA carboxylase inhibitors,⁶ antidepressants,⁷ and as estrogen receptor modulators.^{8,9} Benzothiophenes are present in a number of clinical agents, including Raloxifene,¹⁰ a selective estrogen receptor modulator, Zileuton,¹¹ an inhibitor of 5-lipoxygenase and leukotriene biosynthesis used for the treatment of asthma, and the antifungal agent Sertaconazole, which inhibits the synthesis of ergosterol.¹²

Scaffolds based upon 2- or 3-aminobenzo[*b*]thiophenes have enormous potential for further derivatization and have shown great promise in fragment-based drug discovery and in hit identification or lead development, including approaches towards antimitotic agents^{5,13} and in the development of inhibitors of kinase targets, such as the LIMK protein family,¹⁴ PIM-kinases¹⁵ and MAPK-2 kinase (MK2) (Fig. 1).^{16,17} A number of 3-aminothieno[2,3-*b*]pyridine-2-carboxamide hits,

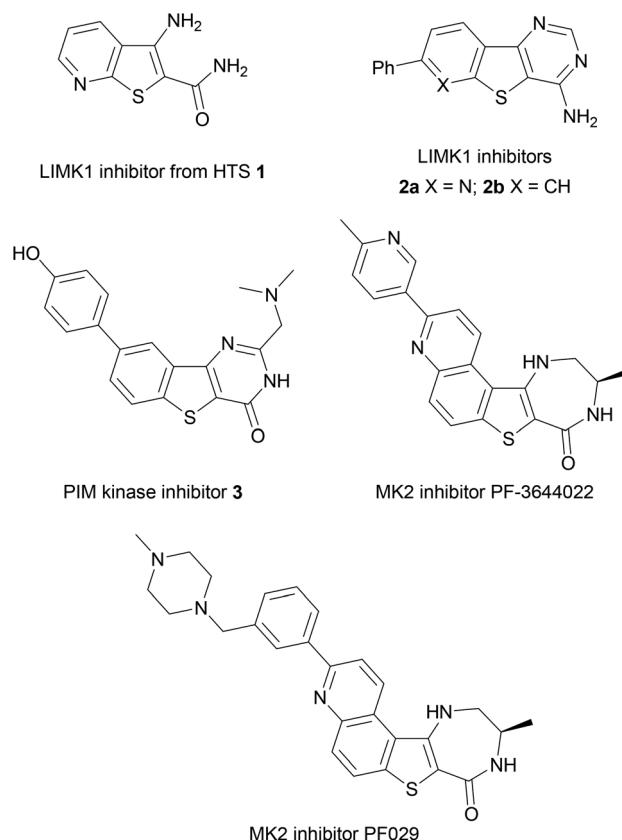


Fig. 1 Aminobenzothiophene scaffolds in drug discovery.

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such as **1**, were identified from high throughput screening (HTS) as inhibitors of LIMK1, leading to the development of tricyclic derivatives such as **2a** and the benzothieno[3,2-*d*]pyri-

midine **2b** as a LIMK1 inhibitor lead candidate, to disrupt actin polymerisation and thus prevent the metastatic potential of tumour cells where LIMK is over-expressed.¹⁴

Benzothienopyrimidinones have been investigated as PIM kinase inhibitors.¹⁵ The PIM kinases (PIM1, PIM2 and PIM3) have been implicated in tumourigenesis and simultaneous targeting of all three isoforms has presented itself as a promising approach in cancer therapy, with PIM triple knockout mice found to be viable and fertile.¹⁸ The benzothiophene scaffold was again identified from an initial HTS hit,¹⁵ leading to the development of a range of potent and selective benzo[b]thiophene-derived inhibitors such as **3** with nM activity (K_i values of 2, 3 and 0.5 nM against PIM1, PIM2 and PIM3, respectively) with oral bioavailability in mouse models.

Examples of aminobenzothiophene derivatives are also found amongst inhibitors of the mitogen activated protein kinase (MAPK) family of enzymes. These enzymes are essential for inflammatory cell signalling events and contain historically popular drug targets for inflammatory diseases, including rheumatoid arthritis and Crohn's disease because of their involvement in the production of pro-inflammatory cytokines,¹⁹ as well as being implicated in accelerated cellular ageing in Werner syndrome (WS) *via* p38.^{20–23} MAPK-activated protein kinase (MK2) is a rate-limiting kinase downstream of p38 in the MAPK pathway and has been the subject of many studies in recent years,²⁴ as MK2 knock-out mice possess normal healthy phenotypes whereas p38 knock-out mice are lethal.²⁵ The aminobenzo[b]thiophene derivative PF-3644022 shows excellent kinase selectivity for MK2, *in vivo* potency on a nanomolar scale and projected ADME characteristics that suggested it was suitable for oral human dosing.^{17,26,27} However, PF-3644022 was found to result in hepatotoxicity in dogs²⁷ and so the analogue PF029 was developed and exhibited an improved toxicological profile with no loss of cellular potency. This was rationalized through installation of a metabolic shunt onto the reactive diazepinone ring and extension of the biaryl ring section to increase the compound's cationic character, thus reducing its molecular affinity for transporter proteins.

As part of our interest in the synthesis of MAPK inhibitors for the study of cellular ageing in Werner syndrome,^{20,28–33} the benzothiophene scaffold, and its selectivity and cellular activity profile for MK2 exemplified by PF-3644022, made it an attractive target for synthesis. We have shown that treating young WS cell cultures with p38 MAPK inhibitors can bring about a complete reversal of the ageing phenotype, giving increased replicative life-span, growth rates comparable to normal young cells and a reduction in levels of F-actin stress fibres.^{20,23} These findings suggested that WS could be amenable to therapeutic intervention, but with high toxicity and poor kinase selectivity exhibited *in vivo* by many p38 inhibitors,^{34–36} an inhibitor scaffold that targeted the downstream kinase MK2 would offer a promising alternative target.^{20,37}

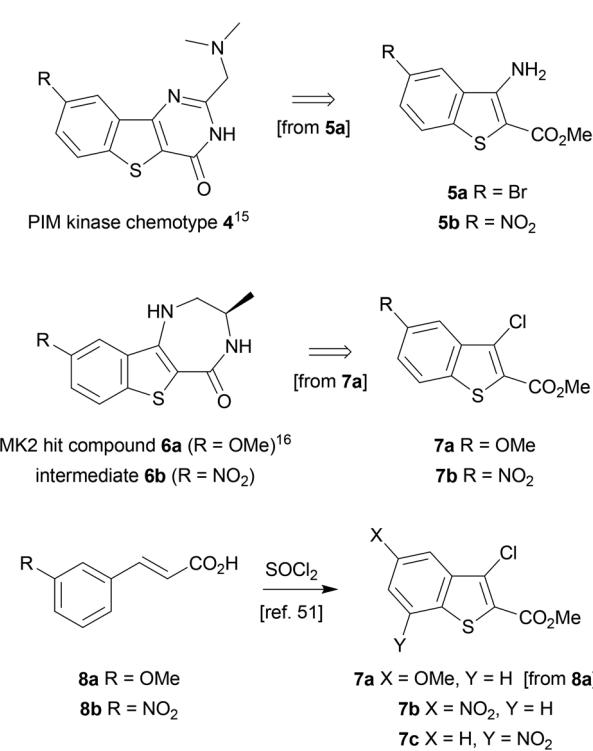
With such a range of biological properties, there is a continuing interest in the search for new methods to access substituted benzothiophenes.³⁸ One approach, with the potential to incorporate diversity into a target library, would be to employ

transition metal-mediated processes from the corresponding 3-halobenzo[b]thiophenes.³⁹ However, methods for the synthesis of 3-halobenzo[b]thiophenes are currently fairly limited. In particular, routes can be problematic when using ring halogenation due to the low reactivity of the heteroaryl unit and its functional group compatibility.^{40–43} The 5-*endo*-dig halocyclisation of *ortho*-alkynylaryl thiophenol derivatives offers an alternative approach,^{44–48} but this requires installation of an alkyne by metal-catalyzed cross-coupling followed by cyclisation, mediated by a halogen-containing electrophile, so can exhibit a number of inherent disadvantages.

Herein, we present an annulation-based method for the rapid preparation of 3-halo and 3-amino-2-substituted benzo[b]thiophenes suitable for elaboration to a range of kinase inhibitors.⁴⁹ It employs microwave irradiation as a convenient platform for fast reaction kinetics, and to improve reaction efficiency, and avoids the need for metal-catalyzed processes to establish the parent heterocycle. This method is shown to be suitable to access the pharmacophore of a range of biologically-active scaffolds for application in medicinal chemistry and drug discovery.

Results and discussion

The benzothiophene-containing chemotypes appearing in recent drug discovery programmes (Fig. 1) feature, or could in principle be derived from, electron-poor aminobenzothiophene intermediates or their 7-aza analogues (Scheme 1). For



Scheme 1 Benzothiophene precursors to kinase inhibitors.



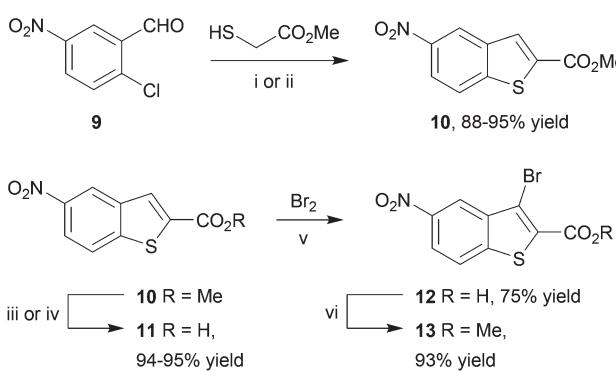
example, the PIM kinase inhibitor scaffold **4** has been accessed from 5-bromobenzothiophene **5a**, using the halogen as a handle for library diversification in a late-stage Suzuki coupling.¹⁵ Similarly, it could be hypothesized that inhibitors of MK2 for study in WS cells, such as PF-3644022 (Fig. 1), could be prepared from the same core motif **5**, using 5-nitrobenzothiophene **5b**, rather than by the functionalization of 6-nitroquinoline as reported by Anderson *et al.*¹⁷ This approach would enable the synthesis of a range of diverse chemical tools from a single common template. The original route to MK2 hit compound **6a**, prior to the development of PF-3644022,¹⁶ employed cinnamic acid **8a** in reaction with thionyl chloride in chlorobenzene at 120 °C to establish the 3-chlorobenzothiophene scaffold **7a** (Scheme 1).⁵⁰ Unfortunately this route would be wholly inappropriate for the synthesis of benzothiophene **7b** for elaboration to the desired intermediate **6b** on route to PF-3644022, as altering the substituent-directing effects to a nitro group results in poor yields and inseparable mixtures of **7b** and **7c** in the benzothiophene synthesis, as well as giving other side products, as reported by Higa.⁵¹ Hence an alternative route had to be sought.

Our first approach towards scaffold **6b** used an alternative and established method to access 3-halobenzothiophenes by halogenation of the corresponding benzothiophene.⁴⁰ The condensation of methyl thioglycolate with 2-chloro-5-nitrobenzaldehyde (**9**) under basic conditions gave methyl 5-nitrobenzo[b]-thiophene-2-carboxylate (**10**) in high yield (Scheme 2). We have shown in previous work how microwave dielectric heating can be used to dramatically reduce reaction times in the synthesis of inhibitor scaffolds.^{30,32,33,37,52} Given that elevated temperature has promoted this,^{53,54} and a closely-related process for the synthesis of 2-acetylbenzothiophenes,⁵⁵ we carried out this transformation under microwave irradiation at 90 °C to give the benzothiophene **10** in good yield, whilst shortening the reaction time from 17 h to 15 min. Selective halogenation at C-3 of **10** was facilitated in a convoluted sequence of reactions *via* the carboxylic acid **11** to overcome poor heterocycle reactivity.⁴⁰ Saponification, again conducted by microwave dielectric

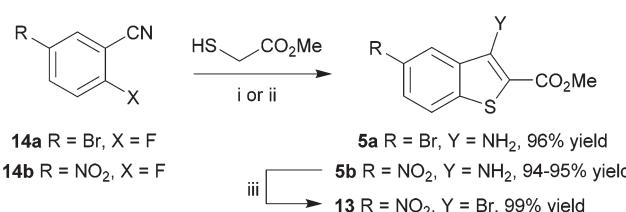
heating, under basic conditions was complete in 3 min at 100 °C and gave the carboxylic acid **11** in excellent yield. Subsequent heating with excess bromine and sodium acetate in glacial acetic acid did give 3-bromobenzothiophene **13** in good yield after esterification using methyl iodide on a number of occasions. However, the bromination was found to be highly variable and efforts to develop an alternative process using microwave heating were constantly frustrated⁵⁶ and so a more reliable and efficient route was sought. The poor yield of this bromination reaction is catalogued in a recent report.⁵⁷

In an alternative approach, our success in the microwave-assisted synthesis of **10** was adapted to incorporate an amino group at C-3, amenable by diazotization chemistry to provide efficient access to 3-bromobenzothiophene **13**. The cyclocondensation of methyl thioglycolate with 2-nitrobenzonitriles under basic conditions has been established by Beck,^{58–60} and adapted methods with halide displacement have also been reported.^{61–64} By switching the base from NaOMe⁶¹ to Et₃N⁶³ and heating either 5-bromo-2-fluorobenzonitrile (**14a**) or 2-fluoro-5-nitrobenzonitrile (**14b**) and methyl thioglycolate gave the corresponding 3-aminobenzothiophene **5a,b** in very high yield (Scheme 3), *e.g.* for **5b** either at 100 °C in DMSO for 2 h using conductive heating (95% yield) or under microwave irradiation at 130 °C for 11 min (94% yield), after simply pouring the reaction mixture into ice-water and collecting the product by filtration. Subsequent deaminative bromination⁶⁰ of aminobenzothiophene **5b** using *tert*-butyl nitrite in acetonitrile in the presence of copper(II) bromide gave the target bromobenzothiophene **13** in excellent yield by this much more direct route.

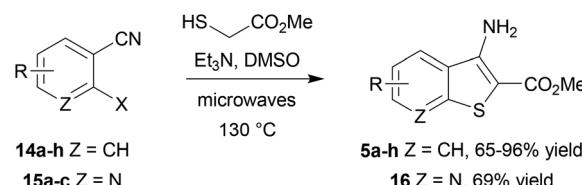
The scope of this method was further explored by investigating a number of substrates (Scheme 4, Table 1), suitable for



Scheme 2 Reagents and conditions: (i) K₂CO₃, DMF, RT, 17 h; (ii) K₂CO₃, DMF, microwaves, 90 °C, 15 min; (iii) NaOH, MeOH–H₂O, reflux, 3 h; (iv) NaOH, MeOH–H₂O, microwaves, 100 °C, 3 min; (v) Br₂, AcOH, NaOAc, 55 °C, 48 h; (vi) MeI, K₂CO₃, DMF, RT, 3 h.



Scheme 3 Reagents and conditions: (i) Et₃N, DMSO, 100 °C, 2 h; H₂O; (ii) Et₃N, DMSO, microwaves, 130 °C, 11 min; H₂O; (iii) *tert*-BuONO, CuBr₂, MeCN, 0 °C; RT, 2 h; HCl (aq.).



Scheme 4 Synthesis of benzothiophenes **5a–h** and **7-aza-16**.



Table 1 Scope of the microwave-assisted synthesis of 3-amino benzothiophenes **5a–h** ($Y = \text{NH}_2$) and the 7-aza analogue **16**

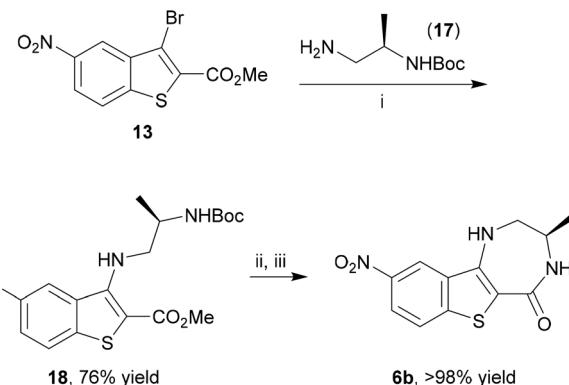
Entry	Substrate	R	X	Z	Time/ min ^a	Product	Yield ^b (%)
1	14a	5-Br	F	CH	11	5a	96
2	14b	5-NO ₂	F	CH	11	5b	94
3	14c	5-Cl	F	CH	11	5c	92
4	14d	4-CF ₃	F	CH	18	5d ($R = 6\text{-CF}_3$)	80
5	14e	4-NO ₂	F	CH	35	5e ($R = 6\text{-NO}_2$)	67
6 ^c	14f	5-Ph	F	CH	15	5f	85
7	14g	H	F	CH	15	5g	65
8	14g	H	Br	CH	15	5g	23
9	14g	H	I	CH	15	5g	47
10	14h	5-CF ₃	F	CH	20	5h	88
11	15a	H	F	N	15	16	66
12	15b	H	Cl	N	15	16	69
13	15c	H	Br	N	15	16	51

^a Hold time at the given temperature, as measured by the in-built IR sensor, by modulation of the initial microwave power. ^b Isolated yield of product **5** or **16** after reaction according to Scheme 4, cooling in a stream of compressed air and pouring the reaction mixture into iced water. ^c Product was isolated by aqueous work up, followed by purification by column chromatography on silica gel.

elaboration to a range of benzothiophene-containing scaffolds found in drug discovery (Fig. 1).

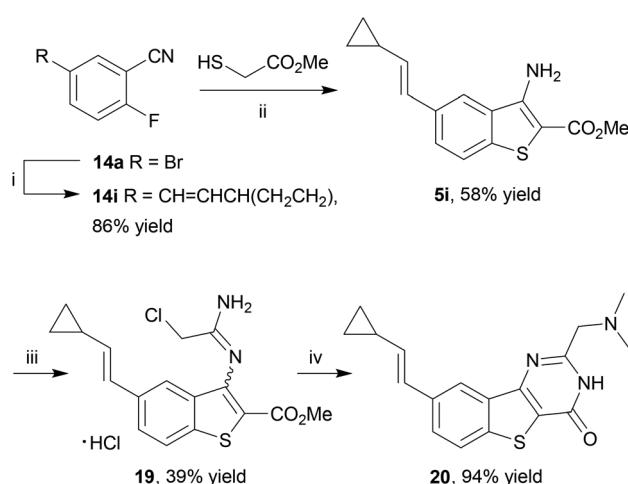
It was found that the process was most efficient for highly electron-poor precursors, such as **14a–d** (entries 1–4), and was generally most effective with 2-fluorides, but could also accommodate bromides and iodides albeit with reduced efficiency (Table 1, entries 8 and 9). In most cases a very simple work up procedure was effective, providing benzothiophenes **5a–h** in 65–96% yield in reaction times varying between 11 and 35 min, depending upon substrate. The efficiency compared well to other available methods (*cf.* Table 1, entry 7, with Beck synthesis of **5g**,⁵⁸ 52% yield after 20 h), and so this method was adopted as the route of choice to access the benzothiophene scaffold. Furthermore, it was possible to apply the procedure to the synthesis of 7-azabenzothiophene **16** using 2-halonicotinonitriles **15a–c**. Interestingly, the choice of halogen as substrate did not cause much variation in the yield of azabenzothiophene product **16** (entries 11–13), which represents the core heterocyclic motif of the 3-aminothieno[2,3-*b*]pyridine-2-carboxamide inhibitors¹⁴ of LIMK1.

Having developed this rapid microwave-assisted method to prepare benzothiophenes, the 5-nitro analogue **13** was further elaborated to the benzo[4,5]thieno[3,2-*e*][1,4]diazepin-5(2*H*)-one MK2 inhibitor scaffold **6b**. Buchwald–Hartwig coupling of (*R*)-*tert*-butyl (1-aminopropan-2-yl)carbamate (**17**) and bromobenzothiophene **13** gave a good yield of the *N*-arylated product **18** under microwave irradiation at 150 °C after 75 min (Scheme 5). Subsequent Boc-deprotection using TFA and lactamization by treatment with NaOMe using a modified procedure of Boschelli⁶⁵ under conductive heating gave the MK2 inhibitor scaffold **6b**, bearing suitable functionality for further elaboration, in essentially quantitative yield.



Scheme 5 Reagents and conditions: (i) $\text{Pd}(\text{OAc})_2$ (5 mol%), Cs_2CO_3 , (\pm) -BINAP (13 mol%), PhMe, microwaves, 150 °C, 75 min; (ii) TFA, CH_2Cl_2 , RT, 4.5 h; (iii) NaOMe, MeOH, 50 °C, 2 h; reflux, 2 h; HCl (aq.).

Finally, an application of our microwave-assisted method to access a pre-functionalized benzothiophene **5i** for direct transformation to a benzo[4,5]thieno[3,2-*d*]pyrimidin-4-one scaffold **21** as a chemical tool for PIM kinase inhibition^{15,66} was investigated. This inhibitor exhibits subnanomolar to single-digit nanomolar K_i values against all three PIM kinases and has been co-crystallised with PIM1, guiding subsequent SAR studies. It was postulated that rather than introducing the cyclopropylvinyl group by a late-stage Suzuki coupling, in accordance with the original diversification study, this group could be incorporated from the start in order to access **20** directly. To that end, 2-fluorobenzonitrile **14i** was prepared by the Pd-catalyzed Suzuki–Miyaura coupling of 5-bromo-2-fluorobenzonitrile (**14a**) and the corresponding boronate ester at 80 °C,¹⁵ and heated with methyl thioglycolate in the presence



Scheme 6 Reagents and conditions: (i) *trans*-2-cyclopropylvinylboronic acid pinacol ester, $\text{Pd}(\text{Ph}_3)_2\text{Cl}_2$ (10 mol%), 1 M Na_2CO_3 , DME/EtOH/H₂O, 80 °C, 16 h; (ii) Et_3N , DMSO, microwaves, 130 °C, 35 min; (iii) 2-chloroacetonitrile, HCl, dioxane, RT, 16 h; (iv) Me_2NH , EtOH, RT, 16 h; SCX column, $\text{MeOH}/\text{CH}_2\text{Cl}_2$, NH_3 , MeOH.



of Et_3N in DMSO at 130 °C for 35 min under microwave irradiation to give the corresponding benzothiophene **5i** in reasonable yield after purification by column chromatography (Scheme 6). The cyclopropylvinyl group was found to be compatible with the subsequent chemistry: reaction of **5i** with chloroacetonitrile in 4 N HCl in dioxane gave chloromethyl derivative **19** which, on reaction with dimethylamine, underwent further cyclization to give the thienopyrimidinone scaffold **20** after purification by immobilization on an acidic resin. Thus the method delivered this known PIM kinase inhibitor by an extremely rapid route, suitable for biological study.

Experimental

Materials and methods

Commercially available reagents were used without further purification; solvents were dried by standard procedures. Light petroleum refers to the fraction with bp 40–60 °C and ether refers to diethyl ether. Unless otherwise stated, reactions were performed under an atmosphere of air. Flash chromatography was carried out using Merck Kieselgel 60 H silica or Matrix silica 60. Analytical thin layer chromatography was carried out using aluminium-backed plates coated with Merck Kieselgel 60 GF₂₅₄ that were visualised under UV light (at 254 and/or 360 nm). Microwave irradiation experiments were performed in a sealed Pyrex tube using a self-tunable CEM Discover, CEM Explorer or Biotage Initiator 2.5 EXP EU focused monomodal microwave synthesizer at the given temperature using the instrument's in-built IR temperature measuring device, by varying the irradiation power (initial power given in parentheses).

Fully characterized compounds were chromatographically homogeneous. Melting points were determined on a Kofler hot stage apparatus or Stanford Research Systems Optimelt and are uncorrected. Specific rotations were measured at the indicated temperature (in °C) using a ADP440 polarimeter (Bellingham + Stanley) at the sodium D line and are given in deg $\text{cm}^{-3} \text{ g}^{-1} \text{ dm}^{-1}$ with concentration c in $10^{-2} \text{ g cm}^{-3}$. Infra-red spectra were recorded in the range 4000–600 cm^{-1} on a Perkin-Elmer 1600 series FTIR spectrometer using an ATR probe or a Shimadzu IRAffinity-1 equipped with an ATR accessory and are reported in cm^{-1} . NMR spectra were recorded using a Varian VNMRS instrument operating at 400 or 500 MHz or a Bruker Avance III 400 MHz or Bruker Avance DRX 500 MHz for ^1H spectra and 100 or 126 MHz for ^{13}C spectra; J values were recorded in Hz and multiplicities were expressed by the usual conventions. Low resolution mass spectra were determined using a Waters Q-TOF Ultima using electrospray positive ionization, A Waters LCT premier XE using atmospheric pressure chemical ionization (APCI), an Agilent 6130 single quadrupole with an APCI/electrospray dual source, a Fisons Instrument VG Autospec using electron ionization at 70 eV (low resolution) or a ThermoQuest Finnigan LCQ DUO electrospray, unless otherwise stated. TOFMS refers to time-of-flight mass spectrometry, ES refers to electrospray ionization, CI refers to chemical ionization (ammonia), FTMS refers to Fourier trans-

form mass spectrometry, NSI refers to nano-electrospray ionization and EI refers to electron ionization. A number of high resolution mass spectra were obtained courtesy of the EPSRC Mass Spectrometry Service at University College of Wales, Swansea, UK using the ionization methods specified.

Synthetic procedures

General procedure for synthesis of 3-aminobenzo[*b*]thiophenes 5 from benzonitriles 14. A mixture of the benzonitrile **14** (1.0 equiv.), methyl thioglycolate (1.05 equiv.) and triethylamine (3.1 equiv.) in dry DMSO (2 M) was irradiated in a Biotage Initiator 2.5 EXP EU or CEM Discover microwave synthesizer at 130 °C for the time specified (hold time) by modulating the initial microwave power. After cooling to room temperature in a stream of compressed air, the reaction mixture was poured into ice-water and the resulting solid collected, washed with water and dried *in vacuo* to give the desired product.

Methyl 3-amino-5-bromobenzo[*b*]thiophene-2-carboxylate (5a). 5-Bromo-2-fluorobenzonitrile (**14a**) (0.50 g, 2.50 mmol), methyl thioglycolate (0.22 mL, 2.60 mmol) and triethylamine (1.10 mL, 7.75 mmol) were reacted according to the above general procedure for 11 min to give the *title compound* (700 mg, 96%) as a brown solid, mp 170–171 °C (Found [ES⁺]: 285.9535. $\text{C}_{10}\text{H}_9\text{BrNO}_2\text{S}$ [MH] requires 285.9532); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 3477 (N–H), 3363 (N–H), 2954 (C–H), 1681 (C=O); ^1H NMR (400 MHz, d_6 -DMSO) $\delta_{\text{H}}/\text{ppm}$ 8.44 (1H, d, J = 1.9 Hz, 4-CH), 7.82 (1H, d, J = 8.6 Hz, 7-CH), 7.64 (1H, dd, J = 8.6, 1.9 Hz, 6-CH), 7.17 (2H, bs, NH₂), 3.79 (3H, s, Me); ^{13}C NMR (101 MHz, d_6 -DMSO) $\delta_{\text{C}}/\text{ppm}$ 164.6 (C), 148.6 (C), 137.7 (C), 133.2 (C), 131.0 (C), 125.7 (CH), 125.2 (CH), 117.1 (CH), 96.1 (C), 51.4 (Me); m/z (ES) 286 ($\text{M}^{+}79\text{Br}\text{H}^+$, 100%).

Methyl 3-amino-5-nitrobenzo[*b*]thiophene-2-carboxylate (5b). 2-Fluoro-5-nitrobenzonitrile (**14b**) (0.50 g, 3.0 mmol), methyl thioglycolate (0.30 mL, 3.3 mmol) and triethylamine (1.3 mL, 9.3 mmol) were reacted according to the above general procedure for 11 min to give the *title compound* (0.71 g, 94%) as an orange solid, mp 243.4–244.1 °C (acetone) (lit.⁶⁷ mp 244–246 °C) (Found [FTMS]: 253.0280. $\text{C}_{10}\text{H}_9\text{N}_2\text{O}_4\text{S}$ [MH] requires 253.0278); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 3445 (N–H), 3342 (N–H), 1681 (C=O), 1572 (NO₂), 1432 (C–C), 1328 (NO₂), 1276 (C–O), 1093 (C–O); ^1H NMR (500 MHz, d_6 -DMSO) $\delta_{\text{H}}/\text{ppm}$ 9.24 (1H, d, J = 2 Hz, 4-CH), 8.29 (1H, dd, J = 9, 2 Hz, 6-CH), 8.12 (1H, d, J = 9 Hz, 7-CH), 7.47 (2H, bs, NH₂), 3.82 (3H, s, Me); ^{13}C NMR (100 MHz, d_6 -DMSO) $\delta_{\text{C}}/\text{ppm}$ 164.2 (C), 149.6 (C), 144.9 (C), 144.5 (C), 131.4 (C), 124.3 (CH), 122.0 (CH), 119.4 (CH), 96.8 (C), 51.4 (Me); m/z (EI) 252 (M^+ , 100%), 219 (72). ^1H and ^{13}C NMR spectroscopic analyses were in good agreement with literature data.⁶⁷

Methyl 3-amino-5-chlorobenzo[*b*]thiophene-2-carboxylate (5c). 5-Chloro-2-fluorobenzonitrile (**14c**) (0.50 g, 3.2 mmol), methyl thioglycolate (0.30 mL, 3.36 mmol) and triethylamine (1.38 mL, 9.92 mmol) were reacted according to the above general procedure for 11 min to give the *title compound* (710 mg, 92%) as a colourless solid, mp 170–172 °C (Found [ES⁺]: 242.0039. $\text{C}_{10}\text{H}_9^{35}\text{ClNO}_2\text{S}$ [MH] requires 242.0037); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 3477 (N–H), 3365 (N–H), 1681 (C=O), 1278

(C–O); ^1H NMR (500 MHz, d_6 -DMSO) δ_{H} /ppm 8.30 (1H, d, J = 2.1 Hz, 4-CH), 7.88 (1H, d, J = 8.6 Hz, 7-CH), 7.53 (1H, dd, J = 8.6, 2.1 Hz, 6-CH), 7.17 (2H, bs, NH₂), 3.79 (3H, s, Me); ^{13}C NMR (126 MHz, d_6 -DMSO) δ_{C} /ppm 164.6 (C), 148.7 (C), 137.3 (C), 132.7 (C), 129.1 (C), 128.4 (CH), 125.0 (CH), 122.7 (CH), 96.2 (C), 51.4 (Me); m/z (ES) 242 ($\text{M}^{[35\text{Cl}]}\text{H}^+$, 100%). ^1H NMR spectroscopic analyses were in good agreement with literature data.¹⁵

Methyl 3-amino-6-(trifluoromethyl)benzo[b]thiophene-2-carboxylate (5d). 2-Fluoro-4-(trifluoromethyl)benzonitrile (14d) (0.50 g, 2.64 mmol), methyl thioglycolate (0.24 mL, 2.73 mmol) and triethylamine (1.14 mL, 8.20 mmol) were reacted according to the above general procedure for 20 min to give the *title compound* (580 mg, 80%) as a colourless solid, mp 159–161 °C (Found [ES $^+$]: 298.0120. $\text{C}_{11}\text{H}_8\text{F}_3\text{NNaO}_2\text{S}$ [MNa] requires 298.0124); IR (neat) ν_{max} /cm $^{-1}$ 3471 (N–H), 3344 (N–H), 2964 (C–H), 1664 (C=O); ^1H NMR (400 MHz, d_6 -DMSO) δ_{H} /ppm 8.38–8.32 (2H, 5-CH and 7-CH), 7.72 (1H, d, J = 8.7 Hz, 4-CH), 7.26 (2H, bs, NH₂), 3.82 (3H, s, Me); ^{13}C NMR (126 MHz, d_6 -DMSO) δ_{C} /ppm 164.5 (C), 148.8 (C), 138.9 (C), 134.1 (C), 128.4 (q, $^2J_{\text{C}-\text{F}} = 31.8$ Hz, C), 124.2 (q, $^1J_{\text{C}-\text{F}} = 272.7$ Hz, C), 124.1 (CH), 121.0–120.7 (m, CH), 120.3–120.0 (m, CH), 97.4 (C), 51.5 (Me); m/z (ES) 276 (MH $^+$, 100%).

Methyl 3-amino-6-nitrobenzo[b]thiophene-2-carboxylate (5e). 2-Fluoro-4-nitrobenzonitrile (14e) (0.50 g, 3.00 mmol), methyl thioglycolate (0.28 mL, 3.15 mmol) and triethylamine (1.29 mL, 9.30 mmol) were reacted according to the above general procedure for 35 min to give the *title compound* (510 mg, 67%) as a colourless solid, mp 228–231 °C (lit.⁵⁸ mp 229–231 °C) (Found [ES $^+$]: 275.0101. $\text{C}_{10}\text{H}_8\text{N}_2\text{NaO}_4\text{S}$ [MNa] requires 275.0097); IR (neat) ν_{max} /cm $^{-1}$ 3489 (N–H), 3367 (N–H), 1697 (C=O), 1624 (C–O); ^1H NMR (400 MHz, d_6 -DMSO) δ_{H} /ppm 8.90 (1H, d, J = 2.1 Hz, 7-CH), 8.37 (1H, d, J = 8.7 Hz, 4-CH), 8.19 (1H, dd, J = 8.7, 2.1 Hz, 5-CH), 7.30 (2H, bs, NH₂), 3.83 (3H, s, Me); ^{13}C NMR (101 MHz, d_6 -DMSO) δ_{C} /ppm 164.3 (C), 148.4 (C), 147.0 (C), 138.8 (C), 135.6 (C), 124.2 (CH), 119.8 (CH), 118.5 (CH), 99.8 (C), 51.7 (Me); m/z (ES) 253 (MH $^+$, 20%), 252 (M $^{+}$, 100%). ^1H NMR spectroscopic analyses were in good agreement with literature data.⁶³

Methyl 3-amino-5-phenylbenzo[b]thiophene-2-carboxylate (5f). 4-Fluoro-[1,1'-biphenyl]-3-carbonitrile (14f) (0.30 g, 1.52 mmol), methyl thioglycolate (0.14 mL, 1.59 mmol) and triethylamine (0.65 mL, 4.74 mmol) were reacted according to a modified general procedure for 15 min. After cooling in a stream of compressed air, EtOAc (25 mL) was added and the solution was washed sequentially with water (3 × 25 mL) and brine (3 × 25 mL), dried over MgSO₄ and evaporated *in vacuo*. Purification by flash chromatography on silica gel, eluting with light petroleum–EtOAc (4 : 1), gave the *title compound* (380 mg, 85%) as a colourless solid, mp 96–97 °C (Found [ES $^+$]: 284.0741. $\text{C}_{16}\text{H}_{14}\text{NO}_2\text{S}$ [M] requires 284.0740); IR (neat) ν_{max} /cm $^{-1}$ 3439 (N–H), 3338 (N–H), 2949 (C–H), 1658 (C=O); ^1H NMR (400 MHz, d_6 -DMSO) δ_{H} /ppm 8.54 (1H, s, 4-CH), 7.91 (1H, d, J = 8.5 Hz, 7-CH), 7.84 (1H, d, J = 8.5 Hz, 6-CH), 7.78 (2H, d, J = 7.5 Hz, 2',6'-PhH), 7.51 (2H, app t, J = 7.5 Hz, 3',5'-PhH), 7.39 (1H, t, J = 7.5 Hz, 4'-PhH), 7.26 (2H, bs, NH₂), 3.80 (3H, s, Me); ^{13}C NMR (101 MHz, d_6 -DMSO) δ_{C} /ppm 164.8 (C),

149.9 (C), 139.6 (C), 138.0 (C), 136.2 (C), 132.2 (C), 129.0 (CH), 127.5 (CH), 127.2 (CH), 126.7 (CH), 123.6 (CH), 121.0 (CH), 95.0 (C), 51.2 (Me); m/z (ES) 284 (MH $^+$, 100%).

Methyl 3-aminobenzo[b]thiophene-2-carboxylate (5g). 2-Fluorobenzonitrile (14g) (0.16 mL, 1.5 mmol), methyl thioglycolate (0.135 mL, 1.5 mmol) and triethylamine (0.63 mL, 4.5 mmol) were reacted according to the above general procedure for 15 min to give the *title compound* (203 mg, 65%) as a purple solid, mp 107.6–107.8 °C (MeOH–H₂O) (lit.,⁵⁸ mp 110–111 °C) (Found [ES $^+$]: 208.0434. $\text{C}_{10}\text{H}_{10}\text{NO}_2\text{S}$ [M] requires 208.0432); IR (neat) ν_{max} /cm $^{-1}$ 3434 (N–H), 3337 (N–H), 1659 (C=O), 1520, 1289 (C–O); ^1H NMR (500 MHz, d_6 -DMSO) δ_{H} /ppm 8.13 (1H, d, J = 8.1 Hz, 4-CH), 7.82 (1H, d, J = 8.1 Hz, 7-CH), 7.50 (1H, m, 6-CH), 7.39 (1H, m, 5-CH), 7.15 (2H, bs, NH₂), 3.78 (3H, s, Me); ^{13}C NMR (101 MHz, d_6 -DMSO) δ_{C} /ppm 164.8 (C=O), 149.8 (3-C), 138.8 (7 α -C), 131.4 (3 α -C), 128.5 (6-CH), 123.8 (5-CH), 123.1 (4-CH), 123.1 (7-CH), 94.4 (2-C), 51.2 (Me); m/z (ES) 207 (M $^{+}$, 93%), 176 (30), 175 (100), 147 (34), 146 (37). NMR spectroscopic analyses were in good agreement with literature data.^{59,68}

Methyl 3-amino-5-(trifluoromethyl)benzo[b]thiophene-2-carboxylate (5h). 2-Fluoro-5-(trifluoromethyl)benzonitrile (14h) (0.50 g, 2.6 mmol), methyl thioglycolate (0.24 mL, 2.73 mmol) and triethylamine (1.14 mL, 8.20 mmol) were reacted according to the above general procedure for 20 min to give the *title compound* (640 mg, 88%) as a colourless solid, mp 140–141 °C (Found [ES $^+$]: 298.0121. $\text{C}_{11}\text{H}_8\text{F}_3\text{NNaO}_2\text{S}$ [MNa] requires 298.0120); IR (neat) ν_{max} /cm $^{-1}$ 3468 (N–H), 3335 (N–H), 1664 (C=O); ^1H NMR (400 MHz, d_6 -DMSO) δ_{H} /ppm 8.67 (1H, bs, 4-CH), 8.09 (1H, d, J = 8.5 Hz, 7-CH), 7.79 (1H, dd, J = 8.5, 1.5 Hz, 6-CH), 7.34 (2H, s, NH₂), 3.82 (3H, s, Me); ^{13}C NMR (101 MHz, d_6 -DMSO) δ_{C} /ppm 164.5 (C), 149.3 (C), 142.5 (C), 131.3 (C), 124.9 (q, $^2J_{\text{C}-\text{F}} = 32.7$ Hz, C), 124.5 (q, $^1J_{\text{C}-\text{F}} = 272.1$ Hz, C), 124.4 (C–H), 124.1 (m, C–H), 120.7 (m, C–H), 96.2 (C), 51.4 (Me); m/z (ES) 276 (MH $^+$, 100%).

Methyl (E)-3-amino-5-(2-cyclopropylvinyl)benzo[b]thiophene-2-carboxylate (5i). (E)-5-(2-Cyclopropylvinyl)-2-fluorobenzonitrile (14i) (0.25 g, 1.33 mmol), methyl thioglycolate (0.12 mL, 1.39 mmol) and triethylamine (0.57 mL, 4.12 mmol) were reacted according to a modified general procedure for 35 min. After cooling in a stream of compressed air, EtOAc (25 mL) was added and the solution was washed sequentially with water (3 × 25 mL) and brine (3 × 25 mL), dried over MgSO₄ and evaporated *in vacuo*. Purification by flash chromatography on silica gel, eluting with light petroleum–EtOAc (4 : 1), gave the *title compound* (210 mg, 58%) as a colourless solid, mp 131–132 °C (Found [ES $^+$]: 274.0899. $\text{C}_{15}\text{H}_{16}\text{NO}_2\text{S}$ [M] requires 274.0896); IR (neat) ν_{max} /cm $^{-1}$ 3481 (N–H), 3365 (N–H), 1672 (C=O); ^1H NMR (400 MHz, d_6 -DMSO) δ_{H} /ppm 8.12 (1H, d, J = 1.6 Hz, 4-CH), 7.71 (1H, d, J = 8.5 Hz, 7-CH), 7.50 (1H, dd, J = 8.5, 1.6 Hz, 6-CH), 7.12 (2H, bs, NH₂), 6.53 (1H, d, J = 15.8 Hz, CH), 5.93 (1H, dd, J = 15.8, 9.1 Hz, CH), 3.78 (3H, s, Me), 1.69–1.55 (1H, m, CH), 0.86–0.80 (2H, m, CHH), 0.56–0.49 (2H, m, CHH); ^{13}C NMR (101 MHz, d_6 -DMSO) δ_{C} /ppm 164.8 (C), 149.7 (C), 136.9 (C), 135.2 (C), 133.7 (CH), 131.9 (C), 126.4 (CH), 126.4 (CH), 123.1 (CH), 119.4 (CH),



94.9 (C), 51.2 (Me), 14.4 (CH), 7.1 (CH₂); *m/z* (ES) 274 (MH⁺, 100%).

(3R)-3-Methyl-9-nitro-1,2,3,4-tetrahydro-5H-[1]benzothieno-[3,2-*e*][1,4]diazepin-5-one (6b). A solution of (*R*)-methyl 3-((2-((*tert*-butoxycarbonyl)amino)propyl)amino)-5-nitrobenzo[*b*]-thiophene-2-carboxylate (**18**) (0.30 g, 0.73 mmol) in TFA (10% *v/v* in CH₂Cl₂) (5.7 mL, 7.3 mmol) was stirred at room temperature for 4.5 h. When salt formation was confirmed by TLC analysis [*R*_f 0.3 in MeOH-CH₂Cl₂ (1:9)] the mixture was evaporated *in vacuo*. The residue was dissolved in a mixture of MeOH (5 mL) and NaOMe (25 wt% in MeOH; 1 mL, 4.4 mmol) and warmed to 50 °C for 2 h before heating at reflux for 2 h. The reaction mixture was then allowed to cool to room temperature, cooled in an ice bath, neutralized by the addition of hydrochloric acid (1 M) and stirred at 0 °C for 30 min. The resulting solid was isolated by filtration under reduced pressure, washed with water and dried in air to give the *title compound* (0.18 g, 100%) as a red powder, mp 337.4–342.4 °C (dec.) (Found [ES⁺]: 278.0590. C₁₂H₁₈N₃O₃S [MH] requires 278.0594); [α]_D²⁴ +110.6 (c 0.04, MeOH); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 3307 (N-H), 1598 (C=O), 1501 (NO₂), 1438 (C-C), 1318 (NO₂), 1102 (C-N), 736 (N-H); ¹H NMR (500 MHz, *d*₆-DMSO) $\delta_{\text{H}}/\text{ppm}$ 9.03 (1H, d, *J* = 2 Hz, 10-CH), 8.22 (1H, dd, *J* = 9, 2 Hz, 8-CH), 8.05 (1H, d, *J* = 9 Hz, 7-CH), 8.02 (1H, m, 1-NH), 7.95 (1H, d, *J* = 3 Hz, 4-NH), 3.60 (1H, m, 3-CH), 3.39 (2H, m, 2-CH₂), 1.16 (3H, d, *J* = 7 Hz, Me); ¹³C NMR (125 MHz, *d*₆-DMSO) $\delta_{\text{C}}/\text{ppm}$ 164.2 (5-C), 145.3 (C), 144.2 (C), 141.8 (C), 132.9 (C), 123.8 (7-CH), 120.8 (8-CH), 118.6 (10-CH), 108.6 (C), 50.6 (2-CH₂), 47.6 (3-CH), 18.6 (Me); *m/z* (ES) 277 (M⁺, 87%), 262 (24), 234 (29), 206 (30).

Methyl 5-nitrobenzo[*b*]thiophene-2-carboxylate (10), prepared under ambient conditions. Methyl thioglycolate (0.48 mL, 5.4 mmol) and K₂CO₃ (0.89 g, 6.5 mmol) were added sequentially to a solution of 2-chloro-5-nitrobenzaldehyde (1.01 g, 5.4 mmol) in DMF (6.5 mL) and the mixture was stirred at room temperature for 17 h. The reaction was then quenched in ice-water and the resulting solid collected, washed with water and dried *in vacuo* to give the *title compound* (1.22 g, 95%) as an off-white solid, mp 213.3–217.6 °C (lit.,⁶⁹ mp 213–215 °C) (Found [FTMS + p NSI]: 255.0435. C₁₀H₁₁N₂O₄S [MNH₄] requires 255.0434); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 3093 (C-H), 1701 (C=O), 1528 (NO₂), 1439 (C-C), 1342 (NO₂), 1302 (C-O); ¹H NMR (500 MHz, CDCl₃) $\delta_{\text{H}}/\text{ppm}$ 8.80 (1H, d, *J* = 2 Hz, 4-CH), 8.32 (1H, dd, *J* = 9, 2 Hz, 6-CH), 8.20 (1H, s, 3-CH), 8.01 (1H, d, *J* = 9 Hz, 7-CH), 4.00 (3H, s, Me); ¹³C NMR (125 MHz, CDCl₃) $\delta_{\text{C}}/\text{ppm}$ 162.2 (C), 147.4 (C), 145.9 (C), 138.3 (C), 137.2 (C), 130.7 (3-CH), 123.6 (7-CH), 121.2 (9-CH), 120.9 (7-CH), 52.9 (Me); *m/z* (EI) 237 (M⁺, 100%), 206 (63), 160 (25). ¹H NMR spectroscopic analyses were in good agreement with literature data.^{53,70}

Methyl 5-nitrobenzo[*b*]thiophene-2-carboxylate (10), using microwave-assisted conditions. A mixture of 2-chloro-5-nitrobenzaldehyde (0.75 g, 4.0 mmol), methyl thioglycolate (0.45 mL, 5 mmol) and K₂CO₃ (0.67 g, 4.8 mmol) in DMF (4.5 mL) was irradiated at 90 °C for 15 min (hold time) in a pressure-rated glass tube (35 mL) using a CEM Discover microwave synthesizer by moderating the initial power (100 W). After cooling in a flow

of compressed air, the reaction mixture was poured into water and the resulting solid filtered under reduced pressure, washed with water and dried in air to give the *title compound* (0.84 g, 88%) as an off-white solid, mp 216.8–218.5 °C (lit.,⁶⁹ mp 213–215 °C), with identical spectroscopic properties.

5-Nitrobenzo[*b*]thiophene-2-carboxylic acid (11), using conductive heating. A solution of aqueous NaOH (1 M; 5 mL, 5 mmol) was added to a solution of methyl 5-nitrobenzo[*b*]-thiophene-2-carboxylate (**10**) (0.40 g, 1.7 mmol) in MeOH (5.5 mL) and the mixture was heated at reflux for 3 h. After cooling to room temperature, the solution was acidified with 1 M HCl (aq.) and the resulting solid filtered under reduced pressure and dried in air to give the *title compound* (0.36 g, 95%) as an off-white powder, mp 241.4–242.7 °C (lit.,⁷¹ mp 239–241 °C) (Found [TOFMS]: 224.0020. C₉H₆NO₄S [MH] requires 224.0018); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 2844 (br, O-H), 1670 (C=O), 1600 (C-C), 1511 (NO₂), 1344 (NO₂), 1313 (C-O); ¹H NMR (500 MHz, CD₃OD) $\delta_{\text{H}}/\text{ppm}$ 8.87 (1H, d, *J* = 2 Hz, 4-CH), 8.30 (1H, dd, *J* = 9, 2 Hz, 6-CH), 8.24 (1H, s, 3-CH), 8.15 (1H, d, *J* = 9 Hz, 7-CH); ¹³C NMR (125 MHz, CD₃OD) $\delta_{\text{C}}/\text{ppm}$ 164.8 (C), 148.9 (5-C), 147.5 (C), 140.3 (C), 140.0 (C), 132.0 (3-CH), 125.1 (7-CH), 122.3 (4-CH), 121.9 (6-CH); *m/z* (EI) 223 (M⁺, 100%), 195 (38), 149 (37). ¹H NMR spectroscopic analyses were in good agreement with literature data.⁵³

5-Nitrobenzo[*b*]thiophene-2-carboxylic acid (11), using microwave-assisted heating. A mixture of methyl 5-nitrobenzo[*b*]-thiophene-2-carboxylate (**10**) (0.20 g, 0.84 mmol), aqueous NaOH solution (1 M; 2.5 mL) and MeOH (3.5 mL) was irradiated at 100 °C for 3 min (hold time) in a pressure-rated glass tube (10 mL) using a CEM Discover microwave synthesizer by moderating the initial power (100 W). After cooling in a flow of compressed air, the reaction mixture was diluted with water, acidified with 1 M HCl and the resulting solid was filtered under reduced pressure, washed with water and dried in air to give the *title compound* (0.16 g, 94%) as an off-white powder, mp 241.3–242.1 °C (lit.,⁷¹ mp 239–241 °C), with identical spectroscopic properties.

3-Bromo-5-nitrobenzo[*b*]thiophene-2-carboxylic acid (12). According to a modified literature procedure,⁵⁷ bromine (1.4 mL, 27 mmol) was added portion-wise to a solution of 5-nitrobenzo[*b*]thiophene-2-carboxylic acid (**11**) (1.0 g, 4.5 mmol) and anhydrous sodium acetate (1.13 g, 13 mmol) in glacial acetic acid (28 mL) under N₂. A reflux condenser was fitted and the solution heated at 55 °C for 27 h. After cooling to room temperature, the solution was poured into ice-water and the resulting solid isolated by vacuum filtration and dried in air to give the *title compound* (1.02 g, 75%) as a yellow powder, mp 309.5–316.8 °C (lit.,⁴⁰ mp 307–309 °C) (Found [TOF MS]: 301.9126. C₉H₅⁷⁹BrNO₄S [MH] requires 301.9123); IR (neat) $\nu_{\text{max}}/\text{cm}^{-1}$ 2961 (br, O-H), 1701 (C=O), 1600 (C-C), 1511 (NO₂), 1347 (NO₂), 1270 (C-O), 620 (C-Br); ¹H NMR (500 MHz, CD₃OD) $\delta_{\text{H}}/\text{ppm}$ 8.83 (1H, d, *J* = 2 Hz, 4-CH), 8.39 (1H, dd, *J* = 9, 2 Hz, 6-CH), 8.21 (1H, d, *J* = 9 Hz, 7-CH); *m/z* (EI) 303 (M^[⁸¹Br], 98%), 301 (M^[⁷⁹Br], 96), 293 (17), 291 (13).

Methyl 3-bromo-5-nitrobenzo[*b*]thiophene-2-carboxylate (13), from acid 12. According to a modified literature pro-



cedure,⁵⁷ iodomethane (0.46 mL, 7.4 mmol) was added to a solution of 3-bromo-5-nitrobenzo[*b*]thiophene-2-carboxylic acid (**12**) (1.12 g, 3.7 mmol) and K_2CO_3 (1.28 g, 9.2 mmol) in DMF (15 mL). After stirring at room temperature for 3 h, the reaction mixture was quenched with saturated aqueous NH_4Cl solution, poured into excess water and filtered under reduced pressure. The collected solid was washed with water and dried in air to give the *title compound* (1.09 g, 93%) as an off-white powder, mp 211.5–212.5 °C (lit.,⁴⁰ mp 211–212 °C) (Found [TOF MS]: 315.9279. $C_{10}H_7^{79}BrNO_4S$ [MH] requires 315.9279); IR (neat) ν_{max}/cm^{-1} 3101 (C–H), 1691 (C=O), 1600 (C–C), 1514 (NO₂), 1347 (NO₂), 1089 (C–O), 1052 (C–O), 617 (C–Br); ¹H NMR (500 MHz, CDCl₃) δ_H/ ppm 8.90 (1H, d, J = 2 Hz, 4–CH), 8.38 (1H, dd, J = 9, 2 Hz, 6–CH), 8.00 (1H, d, J = 9 Hz, 7–CH), 4.02 (3H, s, Me); ¹³C NMR (125 MHz, CDCl₃) δ_C/ ppm 161.0 (C), 146.5 (5–C), 144.6 (C), 138.9 (C), 131.0 (3–C), 123.8 (7–CH), 122.2 (6–CH), 121.3 (4–CH), 115.6 (3–CH), 53.0 (Me); m/z (EI) 317 ($M^{[81]Br}^+$, 100%), 315 ($M^{[79]Br}^+$, 94), 286 (58), 284 (55).

Methyl 3-bromo-5-nitrobenzo[*b*]thiophene-2-carboxylate (13), from 3-aminobenzothiophene 5b. Following the procedure of Iaroshenko *et al.*,⁶⁰ CuBr₂ (0.94 g, 4.2 mmol) was added to a solution of *tert*-butyl nitrite (0.45 mL, 3.8 mmol) in dry MeCN (11 mL) at 0 °C under argon. Methyl 3-amino-4-nitrobenzo[*b*]thiophene-2-carboxylate (**5b**) (0.69 g, 2.7 mmol) was then added portion-wise and the solution kept at 0 °C until nitrogen evolution ceased. The reaction mixture was allowed to warm to room temperature, stirred for 2 h and then poured into dilute hydrochloric acid (10%; 25 mL). The aqueous mixture was extracted with EtOAc (3 × 30 mL) and the organic extracts were combined, dried (Na₂SO₄) and evaporated *in vacuo* to give the *title compound* (0.86 g, 99%) as an orange solid, mp 212.3–213.0 °C (acetone) (lit.,⁴⁰ mp 211–212 °C), with identical spectroscopic and spectrometric properties.

(E)-5-(2-Cyclopropylvinyl)-2-fluorobenzonitrile (14i). To a solution containing 5-bromo-2-fluorobenzonitrile (**14a**) (3.0 g, 15.0 mmol), Pd(PPh₃)₂Cl₂ (1.0 g, 1.5 mmol) and *trans*-2-cyclopropylvinylboronic acid pinacol ester (3.7 mL, 18 mmol) in DME/EtOH/H₂O (7 : 2 : 3) (50 mL) was added aqueous Na₂CO₃ solution (1 M; 27 mL). The resulting mixture was placed under an atmosphere of nitrogen and heated at 80 °C for 16 h after which time the reaction was cooled to room temperature and filtered through Celite. Purification by flash column chromatography on silica gel, eluting with light petroleum-EtOAc (9 : 1), gave the *title compound* (2.4 g, 86%) as a yellow oil (Found [ES⁺]: 188.0868. $C_{12}H_{11}NF$ [MH] requires 188.0870); IR (neat) ν_{max}/cm^{-1} 3007 (C–H), 2496 (CN), 1672 (C=C); ¹H NMR (400 MHz, CDCl₃) δ_H/ ppm 7.53–7.46 (2H, m), 7.16–7.08 (1H, m), 6.39 (1H, d, J = 15.7 Hz), 5.71 (1H, dd, J = 15.7, 9.0 Hz), 1.65–1.53 (1H, m), 0.92–0.84 (2H, m), 0.59–0.52 (2H, m); ¹³C NMR (101 MHz, CDCl₃) δ_C/ ppm 161.7 (d, J = 258.2 Hz), 137.9, 135.3 (d, J = 3.5 Hz), 131.8 (d, J = 7.9 Hz), 130.0, 124.2, 116.5 (d, J = 20.3 Hz), 101.5 (d, J = 15.7 Hz), 77.2, 14.7, 7.6; m/z (ES) 188 (MH⁺, 100%).

Methyl 3-aminothieno[2,3-*b*]pyridine-2-carboxylate (16). 2-Chloro-3-pyridinecarbonitrile (**15b**) (35 mg, 0.25 mmol), methyl thioglycolate (0.22 mL, 0.25 mmol) and triethylamine (0.10 mL, 0.72 mmol) were reacted according to the above

general procedure for 15 min to give the *title compound* (36 mg, 69%) as a yellow solid, mp 191 °C (dec.) (lit.,¹³ mp 194–196 °C) (Found [ES⁺]: 209.0380. $C_9H_9N_2O_2S$ [MH] requires 209.0385); IR (neat) ν_{max}/cm^{-1} 3417 (N–H), 3314, 3202, 2943, 1679 (C=O), 1291 (C–O), 1127 (C–O); ¹H NMR (500 MHz, *d*₆-DMSO) δ_H/ ppm 8.68 (1H, dd, J = 4.6, 1.6 Hz, 6–CH), 8.54 (1H, dd, J = 8.1, 1.6 Hz, 4–CH), 7.46 (1H, dd, J = 8.1, 4.6 Hz, 5–CH), 7.30 (2H, bs, NH₂), 3.80 (3H, s, Me); ¹³C NMR (101 MHz, *d*₆-DMSO) δ_C/ ppm 164.7 (C=O), 159.7 (7–C), 150.7 (6–CH), 147.7 (3–C), 131.4 (4–CH), 125.5 (3–C), 119.4 (5–CH), 93.2 (2–C), 51.4 (Me); m/z (EI) 208 (M⁺, 100%), 177 (28), 176 (93), 148 (60). ¹H NMR spectroscopic analyses were in good agreement with literature data.¹³

***tert*-Butyl [(2*R*)-1-aminopropan-2-yl]carbamate (17).** According to a modified literature procedure,⁷² a solution of Boc-D-Ala-OH (0.50 g, 2.64 mmol) and HOBT·H₂O (0.41 g, 3.0 mmol) in CH₂Cl₂ (20 mL) was cooled to 0 °C and EDCI·HCl (0.58 g, 3.0 mmol) was added. The solution was allowed to warm to room temperature and stirred for 30 min. The solution was then cooled to 0 °C, aqueous NH₃ (18.1 M; 0.6 mL) was added drop-wise and the mixture was stirred at room temperature for 30 min. Any solid residue was removed by filtration and the filtrate was washed sequentially with water (20 mL) and brine (2 × 20 mL), dried (MgSO₄) and evaporated *in vacuo*. Purification by flash column chromatography on SiO₂, gradient eluting with MeOH–CH₂Cl₂ (from 3–7% MeOH), gave Boc-D-Ala-NH₂ (0.37 g, 75%) as colourless crystals, mp 127.5–128.2 °C (lit.,⁷³ mp 120–121 °C). According to a modified literature procedure,⁷⁴ BH₃·SMe₂ (2 M in THF; 13.5 mL, 27 mmol) was added portion-wise to a solution Boc-D-Ala-NH₂ (0.5 g, 2.7 mmol) in dry THF (9 mL) under N₂ at 0 °C. The reaction mixture was allowed to warm to room temperature, stirred for 18 h, evaporated *in vacuo* and treated with MeOH (3 × 15 mL), stirring and evaporating *in vacuo* each time. Purification by ion exchange chromatography using an Isolute SPE SCX-2 flash column, eluting first with MeOH and then with a solution of NH₃ in MeOH (2 M), gave the *title compound* (0.39 g, 84%) as a colourless oil [R_f 0.3 in MeOH–CH₂Cl₂ (1 : 9)] (Found [ES⁺]: 175.1443. $C_8H_{19}N_2O_2$ [MH] requires 175.1441); $[\alpha]_D^{24} -5.7$ (c 1.1, MeOH); IR (neat) ν_{max}/cm^{-1} 2973 (C–H), 1685 (C=O), 1521 (N–H), 1364 (CH₃ deformation), 1247 (C–O), 1165 (C–N), 1045 (C–O), 872 (N–H); ¹H NMR (500 MHz, CDCl₃) δ_H/ ppm 4.78 (1H, m, NH_{Boc}), 3.60 (1H, m, 2–CH), 2.70 (1H, m, 1–CHH), 2.59 (1H, m, 1–CHH), 1.40 (9H, s, CMe₃), 1.08 (3H, d, J = 7 Hz, 3–Me); ¹³C NMR (125 MHz, CDCl₃) δ_C/ ppm 155.7 (C), 79.1 (C), 48.6 (2–CH), 47.4 (4–CH₂), 28.4 (CMe₃), 18.5 (Me). ¹H NMR spectroscopic analyses were in good agreement with literature data.⁷⁵

(*R*)-Methyl 3-((*tert*-butoxycarbonyl)amino)propylamino-5-nitrobenzo[*b*]thiophene-2-carboxylate (18). According to a modified literature procedure,⁷⁶ (*R*)-*tert*-butyl (1-aminopropan-2-yl)carbamate (17) (35 mg, 0.20 mmol) was added to a stirred mixture of methyl 3-bromo-5-nitrobenzo[*b*]thiophene-2-carboxylate (**13**) (50 mg, 0.16 mmol), Cs₂CO₃ (73 mg, 0.22 mmol), (\pm)-BINAP (13 mg, 0.02 mmol) and Pd(OAc)₂ (2.0 mg, 8.0 μ mol) in dry toluene (1.0 mL) under N₂ and the mixture was irra-



diated at 150 °C for 75 min (hold time) in a pressure-rated glass tube (10 mL) using a CEM Discover microwave synthesizer by moderating the initial power (200 W). After cooling in a flow of compressed air, the reaction mixture was partitioned between water (20 mL) and EtOAc (30 mL). The aqueous layer was further extracted with EtOAc (2 × 30 mL) and the organic extracts were combined, washed with brine (30 mL), dried (Na₂SO₄) and evaporated. Purification by flash column chromatography on SiO₂ (dry load), gradient eluting with light petroleum to Et₂O-light petroleum (1 : 1), gave the *title compound* (50 mg, 76%) as a red solid, mp 182.9–184.0 °C (Found [ES⁺]: 410.1378. C₁₈H₂₄N₃O₆S [MH] requires 410.1380); [α]_D²⁴ +55.3 (c 0.1, MeOH); IR (neat) ν_{max} /cm⁻¹ 3355 (N-H), 2954 (C-H), 1672 (C=O), 1588 (NO₂) 1508 (C-C), 1325 (NO₂), 1230 (C-O), 1060 (C-N); ¹H NMR (500 MHz, *d*₆-DMSO) δ_{H} /ppm 8.94 (1H, m, 4-CH), 8.27 (1H, d, *J* = 9 Hz, 6-CH), 8.15 (1H, d, *J* = 9 Hz, 7-CH), 7.54 (1H, m, 3-NH), 6.87 (1H, d, *J* = 6 Hz, 2'-NH), 3.81 (4H, m, 1'-CHH and OMe), 3.76 (1H, m, 2'-CH), 3.57 (1H, m, 1'-CHH), 1.32 (9H, s, CMe₃), 1.12 (1H, d, *J* = 6 Hz, 3'-Me); ¹³C NMR (125 MHz, *d*₆-DMSO) δ_{C} /ppm 164.0 (C), 155.1 (C), 150.3 (3-C), 145.2 (C), 144.3 (C), 131.1 (3 α -C), 124.7 (7-CH), 121.6 (6-CH), 120.5 (4-CH), 99.5 (2-C), 77.6 (CMe₃), 51.7 (OMe), 50.5 (1'-CH₂), 46.2 (2'-CH), 28.1 (CMe₃), 18.3 (3'-Me); *m/z* (EI) 410 (MH⁺, 100%), 409 (M⁺, 70).

Methyl 3-((1-amino-2-chloroethylidene)amino)-5-((E)-2-cyclopropylvinyl)benzo[b]thiophene-2-carboxylate hydrochloride (19). Methyl (E)-3-amino-5-(2-cyclopropylvinyl)benzo[b]thiophene-2-carboxylate (**5i**) (200 mg, 0.73 mmol) was suspended in a solution of HCl in 1,4-dioxane (4 M; 3 mL) and chloroacetonitrile (91 μ L, 1.43 mmol) was added drop-wise. The reaction mixture was stirred at room temperature for 16 h and the suspension was filtered under reduced pressure, washed with light petroleum (60 mL) and dried *in vacuo* for 6 h to give the *title compound* (110 mg, 39%) as a colourless solid, mp 195–197 °C (dec.) (Found [ES⁺]: 317.0512. C₁₆H₁₄³⁵ClN₂OS [MH – MeOH] requires 317.0515); IR (neat) ν_{max} /cm⁻¹ 3332 (br, N-H), 3001 (C-H), 2675 (C-H), 1676 (C=O), 1620 (C=C); ¹H NMR (400 MHz, CD₃OD) δ_{H} /ppm 7.91 (1H, d, *J* = 8.6 Hz, 7-CH), 7.72 (1H, d, *J* = 1.5 Hz, 4-CH), 7.67 (1H, dd, *J* = 8.6, 1.5 Hz, 6-CH), 6.60 (1H, d, *J* = 15.8 Hz, CH), 5.96 (1H, dd, *J* = 15.8, 9.1 Hz, CH), 4.73 (2H, s, CH₂), 3.94 (3H, s, Me), 1.65–1.55 (1H, m, CH), 0.87–0.79 (2H, m, CHH), 0.59–0.48 (2H, m, CHH); ¹³C NMR (101 MHz, CD₃OD) δ_{C} /ppm 166.3 (C), 162.4 (C), 138.9 (C), 138.3 (CH), 138.0 (C), 135.8 (C), 130.5 (C), 130.4 (C), 127.7 (CH), 127.3 (CH), 124.6 (CH), 119.6 (CH), 53.4 (Me), 40.1 (CH), 15.5 (CH), 7.8 (CH₂, CH₂); *m/z* (ES) 349 (M⁺³⁵ClH⁺, 100%).

(E)-8-(2-Cyclopropylvinyl)-2-((dimethylamino)methyl)benzo-[4,5]thieno[3,2-*d*]pyrimidin-4(3*H*)-one (20). Compound **19** (70 mg, 0.18 mmol) was added to a solution of dimethylamine in ethanol (33%; 3 mL) and the reaction mixture was stirred at room temperature for 16 h. The solvent was evaporated *in vacuo* and the resulting colourless solid was dissolved in MeOH–CH₂Cl₂ (1 : 1; 5 mL) and loaded onto an SCX cartridge (1 g). The cartridge was flushed with MeOH (3 × 5 mL) and then eluted with a solution of NH₃ in MeOH (2 M; 8 mL) to

give the *title compound* as a colourless solid (55 mg, 94%), mp 202–203 °C (dec.) (Found [ES⁺]: 326.1324 C₁₈H₂₀N₃OS [MH] requires 326.1322); IR (neat) ν_{max} /cm⁻¹ 2873 (br, C-H), 1672 (C=O), 1587 (C=C); ¹H NMR (400 MHz, *d*₆-DMSO) δ_{H} /ppm 12.41 (1H, bs, NH), 8.09 (1H, d, *J* = 1.5 Hz, 4-CH), 8.03 (1H, d, *J* = 8.5 Hz, 7-CH), 7.69 (1H, dd, *J* = 8.5, 1.5 Hz, 6-CH), 6.68 (1H, d, *J* = 15.8 Hz, CH), 6.01 (1H, dd, *J* = 15.8, 9.2 Hz, CH), 3.52 (2H, s, CH₂), 2.29 (6H, s, Me), 1.66–1.56 (1H, m, CH), 0.85–0.78 (2H, m, CHH), 0.61–0.54 (2H, m, CHH); ¹³C NMR (101 MHz, *d*₆-DMSO) δ_{C} /ppm 158.5 (C), 157.2 (C), 152.7 (C), 138.3 (C), 136.2 (CH), 135.2 (CH), 134.6 (C), 126.5 (C), 126.2 (CH), 123.9 (CH), 122.0 (C), 119.7 (CH), 61.3 (CH₂), 45.1 (Me, Me), 14.7 (CH), 7.2 (CH₂, CH₂); *m/z* (ES) 326 (MH⁺, 100%). ¹H NMR spectroscopic analyses were in good agreement with literature data for the corresponding HCl salt.¹⁵

Conclusions

This method for the microwave-assisted synthesis of 3-amino-benzo[b]thiophenes is rapid, simple to carry out and generally high yielding. It has been applied in the synthesis of a range of functional benzothiophenes and their 7-aza analogues and constitutes a very efficient route to the corresponding 3-halo-benzothiophenes using diazonium chemistry. Applications of this process have been shown in the synthesis of the thieno-[2,3-*b*]pyridine core motif of LIMK1 inhibitors using 2-halopyridine-3-carbonitriles, the benzo[4,5]thieno[3,2-*e*][1,4]diazepin-5(2*H*)-one scaffold of MK2 inhibitors with functionality suitable for subsequent modification using Buchwald–Hartwig chemistry, and a benzo[4,5]thieno[3,2-*d*]pyrimidin-4-one target as a chemical tool for inhibition of PIM kinases. In the case of inhibitors of MK2, the use of this approach has been shown to be superior to traditional bromination chemistry using the parent benzothiophene heterocycle. Given the speed, efficiency and reliability of these methods, and their ability to incorporate a wide range of functionality, these approaches are likely to find application in providing chemical tools, rapidly, reliably and efficiently, for advancing studies in chemical biology, as well as to access targets in medicinal chemistry.

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Notes and references

- 1 K. Koike, Z. Jia, T. Nikaido, Y. Liu, Y. Zhao and D. Guo, *Org. Lett.*, 1999, **1**, 197.



2 J. Huang, H. Luo, L. Wang, Y. Guo, W. Zhang, H. Chen, M. Zhu, Y. Liu and G. Yu, *Org. Lett.*, 2012, **14**, 3300.

3 Y. Ni, K. Nakajima, K. Kanno and T. Takahashi, *Org. Lett.*, 2009, **11**, 3702.

4 K. G. Pinney, a. D. Bounds, K. M. Dingeman, V. P. Mocharla, G. R. Pettit, R. Bai and E. Hamel, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 1081.

5 R. Romagnoli, P. G. Baraldi, M. D. Carrion, C. L. Cara, D. Preti, F. Fruttarolo, M. G. Pavani, M. A. Tabrizi, M. Tolomeo, S. Grimaudo, A. Di Cristina, J. Balzarini, J. A. Hadfield, A. Brancale and E. Hamel, *J. Med. Chem.*, 2007, **50**, 2273.

6 T. Chonan, D. Wakasugi, D. Yamamoto, M. Yashiro, T. Oi, H. Tanaka, A. Ohoka-Sugita, F. Io, H. Koretsune and A. Hiratake, *Bioorg. Med. Chem.*, 2011, **19**, 1580.

7 L. Berrade, B. Aisa, M. J. Ramirez, S. Galiano, S. Guccione, L. R. Moltzau, F. O. Levy, F. Nicoletti, G. Battaglia, G. Molinaro, I. Aldana, A. Monge and S. Perez-Silanes, *J. Med. Chem.*, 2011, **54**, 3086.

8 K. C. Lee, B. S. Moon, J. H. Lee, K.-H. Chung, J. A. Katzenellenbogen and D. Y. Chi, *Bioorg. Med. Chem.*, 2003, **11**, 3649.

9 C. D. Jones, M. G. Jevnikar, A. J. Pike, M. K. Peters, L. J. Black, A. R. Thompson, J. F. Falcone and J. A. Clemens, *J. Med. Chem.*, 1984, **27**, 1057.

10 Editorial, *Lancet Oncol.*, 2006, **7**, 443; V. G. Vogel, J. P. Costantino, D. L. Wickerham, W. M. Cronin, R. S. Cecchini, J. N. Atkins, T. B. Bevers, L. Fehrenbacher, E. R. Pajon, J. L. Wade, A. Robidoux, R. G. Margolese, J. James, S. M. Lippman, C. D. Runowicz, P. A. Ganz, S. E. Reis, W. McCaskill-Stevens, L. G. Ford, V. C. Jordan and N. Wolmark, *JAMA*, 2006, **295**, 2727.

11 P. Lu, M. L. Schrag, D. E. Slaughter, C. E. Raab, M. Shou and A. D. Rodrigues, *Drug Metab. Dispos.*, 2003, **31**, 1352.

12 J. D. Croxtall and G. L. Plosker, *Drugs*, 2009, **69**, 339.

13 R. Romagnoli, P. G. Baraldi, M. K. Salvador, D. Preti, M. A. Tabrizi, M. Bassetto, A. Brancale, E. Hamel, I. Castagliuolo, R. Bortolozzi, G. Basso and G. Viola, *J. Med. Chem.*, 2013, **56**, 2606.

14 B. E. Sleebs, A. Levit, I. P. Street, H. Falk, T. Hammonds, A. C. Wong, M. D. Charles, M. F. Olson and J. B. Baell, *Med. Chem. Commun.*, 2011, **2**, 977.

15 Z.-F. Tao, L. A. Hasvold, J. D. Leverson, E. K. Han, R. Guan, E. F. Johnson, V. S. Stoll, K. D. Stewart, G. Stamper, N. Soni, J. J. Bouska, Y. Luo, T. J. Sowin, N.-H. Lin, V. S. Giranda, S. H. Rosenberg and T. D. Penning, *J. Med. Chem.*, 2009, **52**, 6621.

16 D. R. Anderson, M. J. Meyers, R. G. Kurumbail, N. Caspers, G. I. Poda, S. A. Long, B. S. Pierce, M. W. Mahoney and R. J. Mourey, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 4878.

17 D. R. Anderson, M. J. Meyers, R. G. Kurumbail, N. Caspers, G. I. Poda, S. A. Long, B. S. Pierce, M. W. Mahoney, R. J. Mourey and M. D. Parikh, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 4882.

18 H. Mikkers, M. Nawijn, J. Allen, C. Brouwers, E. Verhoeven, J. Jonkers and A. Berns, *Mol. Cell. Biol.*, 2004, **24**, 6104.

19 S. Kumar, J. Boehm and J. C. Lee, *Nat. Rev. Drug Discovery*, 2003, **2**, 717.

20 M. C. Bagley, T. Davis, P. G. S. Murziani, C. S. Widdowson and D. Kipling, *Pharmaceuticals*, 2010, **3**, 1842.

21 D. Kipling, T. Davis, E. L. Ostler and R. G. Faragher, *Science*, 2004, **305**, 1426.

22 T. Davis, M. F. Haughton, C. J. Jones and D. Kipling, *Ann. N. Y. Acad. Sci.*, 2006, **1067**, 243.

23 T. Davis, D. M. Baird, M. F. Haughton, C. J. Jones and D. Kipling, *J. Gerontol., Ser. A*, 2005, **60**, 1386.

24 A. Schlapbach and C. Huppertz, *Future Med. Chem.*, 2009, **1**, 1243.

25 N. Ronkina, A. Kotlyarov and M. Gaestel, *Front. Biosci.*, 2008, **13**, 5511.

26 R. J. Mourey, B. L. Burnette, S. J. Brustkern, J. S. Daniels, J. L. Hirsch, W. F. Hood, M. J. Meyers, S. J. Mnich, B. S. Pierce, M. J. Saabye, J. F. Schindler, S. A. South, E. G. Webb, J. Zhang and D. R. Anderson, *J. Pharmacol. Exp. Ther.*, 2010, **333**, 797.

27 J. S. Daniels, Y. Lai, S. South, P.-C. Chiang, D. Walker, B. Feng, R. Mireles, L. O. Whitley, J. W. McKenzie, J. Stevens, R. Mourey, D. Anderson and J. W. Davis II, *Drug Metab. Lett.*, 2013, **7**, 15.

28 T. Davis, M. J. Rokicki, M. C. Bagley and D. Kipling, *Chem. Cent. J.*, 2013, **7**, 18.

29 M. C. Bagley, M. Baashen, J. Dwyer, P. Milbeo, D. Kipling and T. Davis, in *Microwaves in Drug Discovery and Development: Recent Advances*, ed. J. Spencer and M. C. Bagley, Future Science Ltd., 2014, ch. 5, pp. 86–104.

30 M. C. Bagley, M. Baashen, V. L. Paddock, D. Kipling and T. Davis, *Tetrahedron*, 2013, **69**, 8429.

31 T. Davis, M. C. Dix, M. J. Rokicki, C. S. Widdowson, A. J. C. Brook, D. Kipling and M. C. Bagley, *Chem. Cent. J.*, 2011, **5**, 83.

32 M. C. Bagley, T. Davis, M. C. Dix, V. Fusillo, M. Pigeaux, M. J. Rokicki and D. Kipling, *J. Org. Chem.*, 2009, **74**, 8336.

33 M. C. Bagley, T. Davis, M. C. Dix, C. S. Widdowson and D. Kipling, *Org. Biomol. Chem.*, 2006, **4**, 4158.

34 T. Force, K. Kuida, M. Namchuk, K. Parang and J. M. Kyriakis, *Circulation*, 2004, **109**, 1196.

35 D. M. Goldstein, A. Kuglstatter, Y. Lou and M. J. Soth, *J. Med. Chem.*, 2010, **53**, 2345.

36 M. C. Genovese, *Arthritis Rheum.*, 2009, **60**, 317.

37 T. Davis, M. C. Bagley, M. C. Dix, P. G. S. Murziani, M. J. Rokicki, C. S. Widdowson, J. M. Zayed, M. A. Bachler and D. Kipling, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 6832.

38 C. Hou, Q. He and C. Yang, *Org. Lett.*, 2014, **16**, 5040.

39 E. David, J. Perrin, S. Pellet-Rostaing, J. Fournier dit Chabert and M. Lemaire, *J. Org. Chem.*, 2005, **70**, 3569.

40 M. Martin-Smith and S. Reid, *J. Chem. Soc.*, 1960, 938.

41 M. Martin-Smith and M. Gates, *J. Am. Chem. Soc.*, 1956, **78**, 5351.

42 M. Martin-Smith and M. Gates, *J. Am. Chem. Soc.*, 1956, **78**, 6177.

43 J. Fournier dit Chabert, L. Joucla, E. David and M. Lemaire, *Tetrahedron*, 2004, **60**, 3221.



44 K. O. Hessian and B. L. Flynn, *Org. Lett.*, 2003, **5**, 4377.

45 B. L. Flynn, P. Verdier-Pinard and E. Hamel, *Org. Lett.*, 2001, **3**, 651.

46 D. Yue and R. C. Larock, *J. Org. Chem.*, 2002, **67**, 1905.

47 S. Kim, N. Dahal and T. Kesharwani, *Tetrahedron Lett.*, 2013, **54**, 4373.

48 V. Guilarde, M. A. Fernández-Rodríguez, P. García-García, E. Hernando and R. Sanz, *Org. Lett.*, 2011, **13**, 5100.

49 S. Knapp, P. Arruda, J. Blagg, S. Burley, D. H. Drewry, A. Edwards, D. Fabbro, P. Gillespie, N. S. Gray, B. Kuster, K. E. Lackey, P. Mazzafera, N. C. O. Tomkinson, T. M. Willson, P. Workman and W. J. Zuercher, *Nat. Chem. Biol.*, 2013, **9**, 3.

50 T. Higa and A. J. Krubsack, *J. Org. Chem.*, 1976, **41**, 3399.

51 T. Higa and A. J. Krubsack, *J. Org. Chem.*, 1975, **40**, 3037.

52 M. C. Bagley, T. Davis, M. C. Dix, P. G. S. Murziani, M. J. Rokicki and D. Kipling, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 3745.

53 R. A. Zambias and M. L. Hammond, *Synth. Commun.*, 1991, **21**, 959.

54 J. Cai, S. Zhang, M. Zheng, X. Wu, J. Chen and M. Ji, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 806.

55 J. Debray, M. Lemaire and F. Popowycz, *Synlett*, 2013, 37.

56 M. C. Bagley, J. E. Dwyer and P. Milbeo, unpublished work.

57 J. M. Mbere, J. B. Bremner, B. W. Skelton and A. H. White, *Tetrahedron*, 2011, **67**, 6895.

58 J. R. Beck, *J. Org. Chem.*, 1972, **37**, 3224.

59 M. H. Norman, F. Navas III, J. B. Thompson and G. C. Rigdon, *J. Med. Chem.*, 1996, **39**, 4692.

60 V. O. Iaroshenko, S. Ali, S. Mkrtchyan, A. Gevorgyan, T. M. Babar, V. Semeniuchenko, Z. Hassan, A. Villinger and P. Langer, *Tetrahedron Lett.*, 2012, **53**, 7135.

61 I. Abdillahi and G. Kirsch, *Synthesis*, 2011, 1314.

62 I. Abdillahi and G. Kirsch, *Synthesis*, 2010, 1428.

63 A. J. Bridges and H. Zhou, *J. Heterocycl. Chem.*, 1997, **34**, 1163.

64 M. D. Meyer, R. J. Altenbach, F. Z. Basha, W. A. Carroll, S. Condon, S. W. Elmore, J. F. Kerwin Jr., K. B. Sippy, K. Tietje, M. D. Wendt, A. A. Hancock, M. E. Brune, S. A. Buckner and I. Drizin, *J. Med. Chem.*, 2000, **43**, 1586.

65 S. S. Khatana, D. H. Boschelli, J. B. Kramer, D. T. Connor, H. Barth and P. Stoss, *J. Org. Chem.*, 1996, **61**, 6060.

66 R. Ekambaram, E. Enkvist, A. Vaasa, M. Kasari, G. Raidaru, S. Knapp and A. Uri, *ChemMedChem*, 2013, **8**, 909.

67 V. Bertolasi, K. Dudová, P. Šimůnek, J. Černý and V. Macháček, *J. Mol. Struct.*, 2003, **658**, 33.

68 L. N. Tumey, Y. Bennani and D. C. Bom, *WO Pat.*, 014 647 A2, 2006.

69 V. G. Matassa, F. J. Brown, P. R. Bernstein, H. S. Shapiro, T. P. Maduskuie Jr., L. A. Cronk, E. P. Vacek, Y. K. Yee, D. W. Snyder, R. D. Krell, C. L. Lerman and J. J. Maloney, *J. Med. Chem.*, 1990, **61**, 2621.

70 D. L. Boger, B. E. Fink and M. P. Hedrick, *J. Am. Chem. Soc.*, 2000, **122**, 6382.

71 L. Fieser and R. Kennelly, *J. Am. Chem. Soc.*, 1935, **57**, 1611.

72 W. Wu, Z. Li, G. Zhou and S. Jiang, *Tetrahedron Lett.*, 2011, **52**, 2488.

73 Z. Xia and C. D. Smith, *J. Org. Chem.*, 2001, **66**, 3459.

74 A. Marquart and B. Podlogar, *J. Org. Chem.*, 1994, **59**, 2092.

75 Z. Huang, J. Jin, T. D. Machajewski, W. R. Antonios-McCrea, M. McKenna, D. Poon, P. A. Renhowe, M. Sendzik, C. M. Shafer, A. Smith, Y. Xu and Q. Zhang, *WO Pat.*, 115 572 A2, 2009.

76 M.-J. R. P. Queiroz, A. Begouin, I. C. F. R. Ferreira, G. Kirsch, R. C. Calhelha, S. Barbosa and L. M. Estevinho, *Eur. J. Org. Chem.*, 2004, 3679.

