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Acyclic cucurbit[n]uril-type molecular containers: influence of glycoluril oligomer length on their function as solubilizing agents†

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We present the synthesis of a series of six new glycoluril derived molecular clips and acyclic CB[n]-type molecular containers (**1–3**) that all feature SO_3^- solubilizing groups but differ in the number of glycoluril rings between the two terminal dialkoxyaromatic sidewalls. We report the X-ray crystal structure of **3b** which shows that its dialkoxynaphthalene sidewalls actively define a hydrophobic cavity with high potential to engage in π - π interactions with insoluble aromatic guests. Compounds **1–3** possess very good solubility characteristics (≥ 38 mM) and undergo only very weak self-association ($K_s < 92 \text{ M}^{-1}$) in water. The weak self-association is attributed to unfavorable $\text{SO}_3^- \cdots \text{SO}_3^-$ electrostatic interactions in the putative dimers **1₂–4₂**. Accordingly, we created phase solubility diagrams to study their ability to act as solubilizing agents for four water insoluble drugs (PBS-1086, camptothecin, β -estradiol, and ziprasidone). We find that the containers **3a** and **3b** which feature three glycoluril rings between the terminal dialkoxy-*o*-xylylene and dialkoxynaphthalene sidewalls are less efficient solubilizing agents than **4a** and **4b** because of their smaller hydrophobic cavities. Containers **1** and **2** behave as molecular clip type receptors and therefore possess the ability to bind to and thereby solubilize aromatic drugs like camptothecin, ziprasidone, and PBS-1086.

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Introduction

A ubiquitous problem facing the pharmaceutical industry is that an estimated 40–70% of new drug candidates are so poorly soluble that they cannot be formulated on their own.¹ Accordingly, there is a real need for the development of new tools that improve the solubility of poorly soluble drugs that enable their formulation. To date, a wide variety of methods that enhance the rate and extent of dissolution have been developed including solid dispersions and nanocrystalline solid forms of the drug.² Other methods that have been demonstrated to improve solubility include the preparation of salts, higher solubility prodrugs, dendrimer-drug systems, and designed co-crystalline forms of the drug.³ However, the most attractive approach to improving solubility of insoluble drugs from the point of view of supramolecular chemists relies on the use of cyclodextrin molecular containers (e.g. HP- β -CD,

SBE- β -CD, Fig. 1) as solubilizing excipients.⁴ Cyclodextrin derivatives are currently used to formulate a number of drugs that are administered to humans.

In recent years, we and others, have been actively investigating the synthesis, molecular recognition properties, and applications of an alternative class of molecular container compounds known as cucurbit[n]urils (CB[n], $n = 5, 6, 7, 8, 10, 14$; Fig. 1).⁵ The great interest in CB[n] compounds arises from the availability of a homologous series of hosts that display both high affinity (K_a up to 10^{17} M^{-1}) and high selectivity toward their guests in water.⁶ In addition, the inherent stimuli responsiveness (pH, chemical, electrochemical, photochemical) of CB[n] guest complexes have made CB[n] popular components to create functional systems including molecular machines, chemical sensors, affinity capture phases, and gas purification materials.⁷ Accordingly, workers in the CB[n] field envisioned that CB[n] compounds – particularly CB[7] with its good water solubility – might be good substitutes for cyclodextrins in pharmaceutical applications. In recent years, the macrocyclic CB[n] compounds were demonstrated to have low *in vitro* and *in vivo* toxicity⁸ and have been used to solubilize, protect, activate, and deliver pharmaceutical agents.⁹ CB[8] has been used as a glue to create polymer hydrogels and nanocapsules based on ternary complex formation.¹⁰ Derivatives of macrocyclic CB[n] have even been used for targeted drug delivery applications.¹¹

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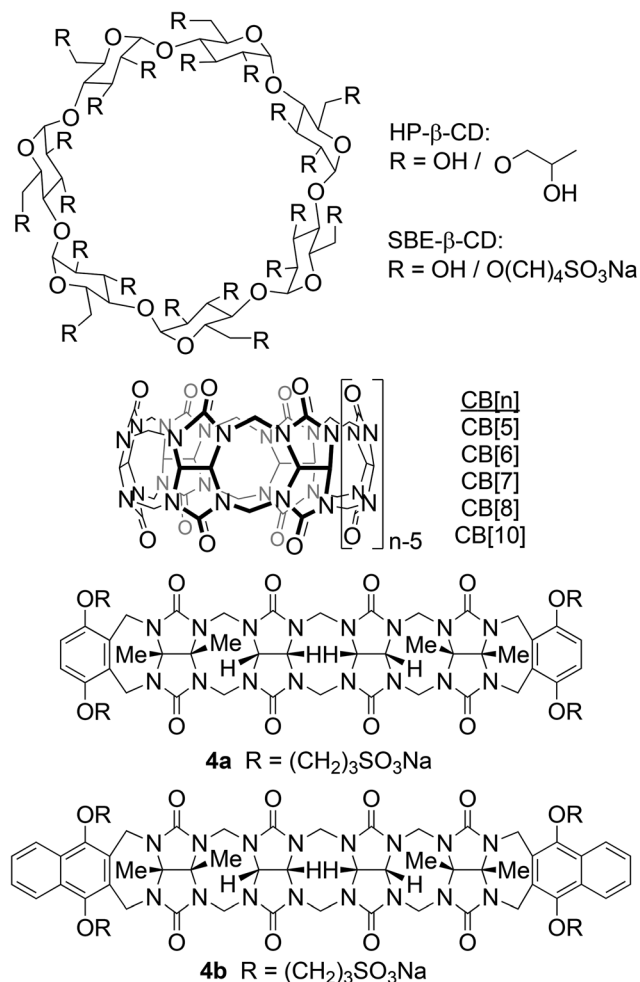


Fig. 1 Structures of solubilizing molecular containers.

Over the years, both the Isaacs and Sindelar groups have worked toward an improved understanding of the mechanism of CB[n] formation – especially with regard to the intermediate methylene bridged glycoluril dimers and higher oligomers¹² – and the preparation of new members of the cucurbit[n]uril family. These studies have resulted in the preparation of glycoluril dimer based molecular clips,¹³ CB[n] analogues,¹⁴ inverted CB[n],¹⁵ nor-seco-CB[n],¹⁶ bambus[n]urils,¹⁷ CB[n] derivatives,^{11a,12e,18} CB[n] dimers,¹⁹ and acyclic CB[n]-type receptors.^{12d,g,j,20} Recently, the Isaacs group used acyclic CB[n] containers as solubilizing excipients²¹ for insoluble drugs and reported the influence of the nature of the solubilizing groups²² and aromatic sidewalls²³ on their function. In this paper we explore the influence of the length of the central glycoluril oligomer on their ability of acyclic CB[n]-type receptors 1–4 to solubilize four insoluble drugs (Fig. 2).

Results and discussion

This results and discussion section is organized as follows. First, we present the synthesis of six new containers (1–3),

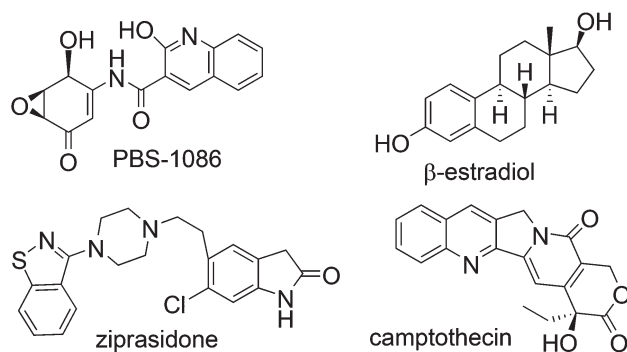


Fig. 2 Structures of insoluble drugs used in this study.

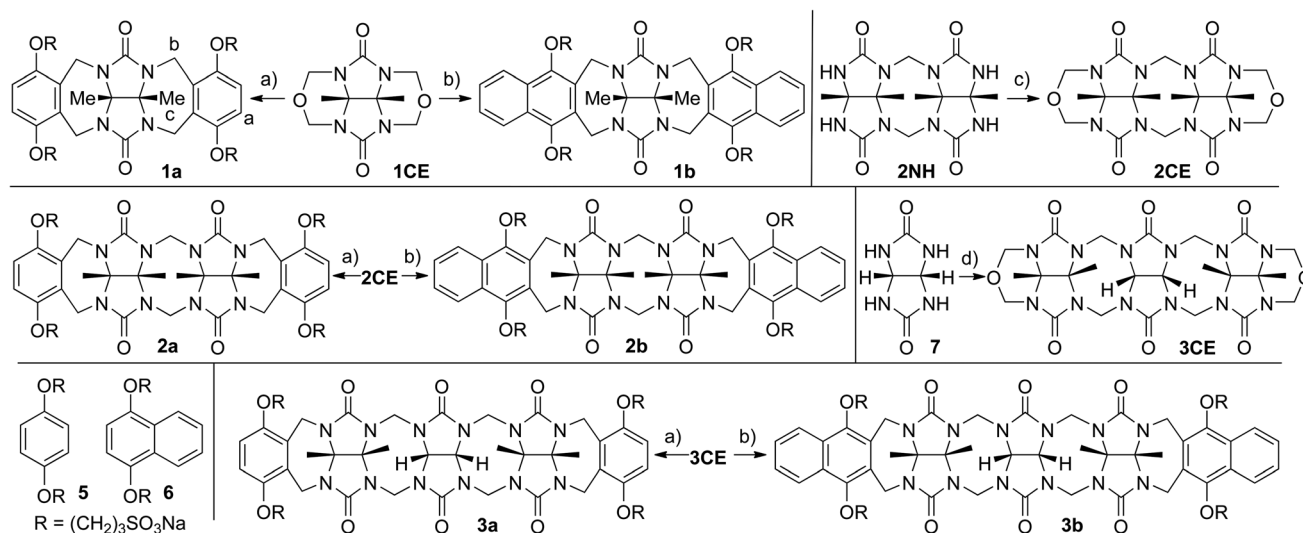
studies of their inherent solubility, and studies of their tendency toward self-association. Next, we present the X-ray crystal structure of **3b** and compare it to the X-ray crystal structures of previously reported glycoluril monomer, dimer, and tetramer derived containers. Next, we report the ability of 1–3 to act as solubilizing agents for four insoluble drugs. Finally, we compare the trends in the solubilization data for 1–3 with that of previously reported **4a** and **4b**.

Synthesis of Containers 1–3

Scheme 1 shows the synthesis of new compounds 1–3. For the preparation of molecular clips **1a** and **1b** we followed the path blazed by Nolte and co-workers and allowed glycoluril bis(cyclic ether) **1CE** to react with the corresponding dialkoxyaromatic walls (**5** and **6**) at 70 °C in TFA–Ac₂O to deliver molecular clips **1a** and **1b** in high yield.^{21,24} To prepare molecular clips **2a** and **2b** we first transformed the known glycoluril dimer **2NH**²⁵ into bis(cyclic ether) **2CE** by treatment with paraformaldehyde in TFA at reflux (40%). Subsequently, we reacted **2CE** with **5** or **6** in TFA/Ac₂O at 70 °C to yield molecular clips **2a** and **2b** in good yield. For the preparation of acyclic CB[n] compounds **3a** and **3b** we first needed to prepare the glycoluril trimer **3CE**. The preparation of **3CE** was accomplished by reaction of glycoluril (**7**) with **1CE** in methanesulfonic acid with careful temperature control (10 °C for 2 h and then 23 °C for 2 h). Subsequently, we allowed **3CE** to react with **5** or **6** in TFA/Ac₂O at 70 °C to give **3a** (48%) and **3b** (59%) in good yields.

We were fortunate to obtain the X-ray crystal structure of acyclic CB[n]-type receptor **3b** (Fig. 3). Fig. 3 also shows the X-ray crystal structure of previously prepared molecular clips **1ester** and **2ester** (Fig. 4) and acyclic CB[n]-type receptor **4b** to illustrate the geometrical change that occurs across the homologous series of receptors 1–4 upon elongation of the glycoluril oligomer backbone. As can be readily seen, as one increases the number of glycoluril rings the receptors change from molecular clips with divergent to nearly parallel aromatic sidewalls (**1** and **2**) to containers with a well defined hydrophobic cavity despite their acyclic nature (**3** and **4**). Overall, as a result of the glycoluril trimer backbone, **3b** is C-shaped and the two naphthalene rings help to define a cavity that is occupied by a molecule of acetone in the crystal. The mean planes of the aromatic rings of **3b** are oriented at an average angle of





Scheme 1 Synthesis of new containers 1–3. Conditions: (a) TFA–Ac₂O (1 : 1), 70 °C, 5; (b) TFA–Ac₂O (1 : 1), 70 °C, 6; (c) TFA, paraformaldehyde, reflux; (d) 1CE, MeSO₃H, 10 °C to 23 °C.

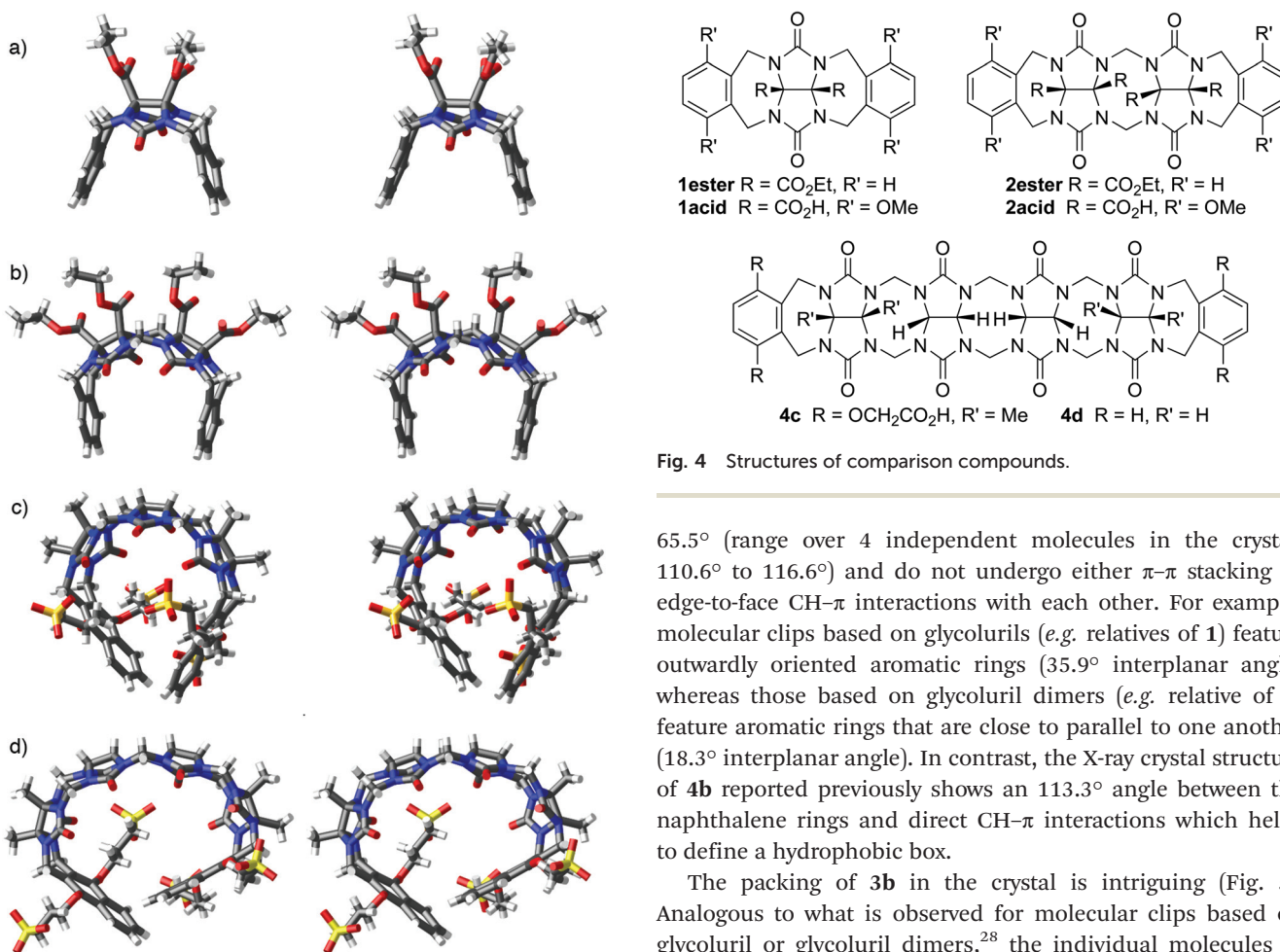


Fig. 4 Structures of comparison compounds.

65.5° (range over 4 independent molecules in the crystal: 110.6° to 116.6°) and do not undergo either π – π stacking or edge-to-face CH– π interactions with each other. For example, molecular clips based on glycolurils (*e.g.* relatives of 1) feature outwardly oriented aromatic rings (35.9° interplanar angle) whereas those based on glycoluril dimers (*e.g.* relative of 2) feature aromatic rings that are close to parallel to one another (18.3° interplanar angle). In contrast, the X-ray crystal structure of 4b reported previously shows an 113.3° angle between the naphthalene rings and direct CH– π interactions which helps to define a hydrophobic box.

The packing of 3b in the crystal is intriguing (Fig. 5). Analogous to what is observed for molecular clips based on glycoluril or glycoluril dimers,²⁸ the individual molecules of 3b interact with one another *via* CH– π and π – π interactions between their naphthalene sidewalls in a head-to-tail fashion to yield linear tape like assemblies. The distance between the



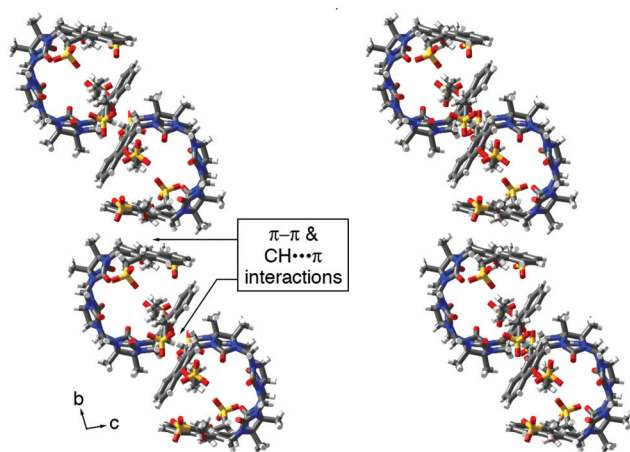


Fig. 5 Illustration of the packing of molecules of **3b** into linear assemblies along the *b*-axis. Color code: C, gray; H, white; N, blue; O, red; S, yellow.

mean planes of the naphthalene ring averages 3.66 Å (range 3.60 to 3.78 Å). These tapes extend along both the *b*-axis and the *c*-axis; they are alternately arranged in a criss-cross fashion reminiscent of building method of a log cabin home as one extends along the *a*-axis. The channels defined by the packing of the linear assemblies are filled with the solubilizing arms and solvating water molecules.

Inherent solubility of 1–4

Given our interest in assessing the performance of **1–3** as solubilizing excipients for insoluble drugs we first sought to measure their inherent solubility. For this purpose, we stir an excess of container with 20 mM phosphate buffered D₂O and readjust the pD to 7.4 until equilibrium is reached and then remove excess insoluble container by centrifugation and filtration. An aliquot of the filtrate is diluted by a known factor and combined with a solution of benzene-1,3,5-tricarboxylic acid as internal standard (1.00 mM); the relative integrals of diagnostic resonances in the ¹H NMR spectrum for the container (**1a**: 6.87 ppm, **1b**: 7.56 ppm, **2a**: 6.99 ppm, **2b**: 7.84 ppm, **3a**: 7.02 ppm, **3b**: 7.95 ppm) versus those of the internal standard (8.38 ppm) were used to determine the inherent solubility of each container. The results are summarized in Table 1. As can be readily seen, the solubility of all the containers bearing dialkoxy-*o*-xylylene sidewalls is higher than 100 mM which is advantageous for their use as solubilizing excipients. In contrast, the solubility of containers with naphthalene sidewalls are variable. Containers **2b** and **3b**

display high solubility (>300 mM) whereas **1b** and **4b** are modest (38 and 14 mM, respectively).

Containers 1–3 do not self-associate strongly

In order for molecular containers to have high potential for use as solubilizing excipients they must not be strongly self-associated which would reduce their ability to form container drug complexes. Accordingly, we studied the self-association of **1–3** by preparing solutions of different concentrations of **1–3** and monitoring the changes in the ¹H NMR chemical shifts. The chemical shifts of the protons on the aromatic sidewall were particularly sensitive to self-association. Fig. 6 shows the changes in chemical shift of *H_a*, *H_b*, and *H_c* as a function of [2a]. The changes in chemical shift were fitted to the standard two-fold self-association model^{21,29} implemented within Scientist 3.0™ (ESI†) which yielded a self-association constant (*K_s*) of 12 M^{−1}. Table 1 gives the self-association constants measured for **1–3** (ESI†) and also presents the known values for **4** from the literature²¹ for comparison. As expected, the *K_s* values measured for acyclic CB[n]-type receptors **3a** and **3b** are quite low (<50 M^{−1}) which is advantageous for their use as solubilizing agents. Given the well-known propensity for glycoluril and glycoluril dimer derived molecular clips to undergo self-association,^{13a,26,28a,d,30} we were surprised that the *K_s* values for **1** and **2** were low (<100 M^{−1}). For purposes of comparison, the structures and self-association constants measured previously for compounds **1acid** (*K_s* = 1840 M^{−1})²⁶ and **2acid** (*K_s* = 41 700 M^{−1})^{13a} which differ from **1a** and **2a** in the spatial orientation of their solubilizing groups are shown in Fig. 4. Because the CO₂[−] solubilizing groups of **1acid** and **2acid** are on their convex face, unfavorable carboxylate–carboxylate electrostatic interactions are avoided upon formation of dimers **1acid**₂ and **2acid**₂. In contrast, the SO₃[−] groups of **1–4** are directed toward each other within the putative dimeric species **1**₂–**4**₂ which results in unfavorable electrostatic sulfonate–sulfonate interactions which decreases the propensity of **1–4** to dimerize. The self-association constant of

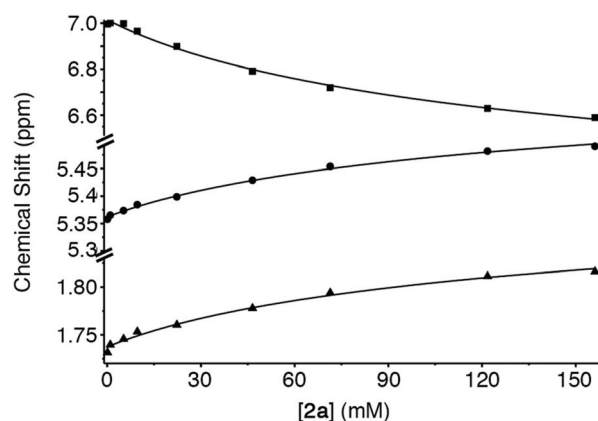


Fig. 6 Plot of chemical shift versus [2a]. The solid line represents the best global fit of the data to a two-fold self-association model with *K_s* = 12 M^{−1}. Conditions: 20 mM sodium phosphate buffered D₂O, pD 7.4, room temperature. Key: *H_a*, ■; *H_b*, ●; *H_c*, ▲.

Table 1 Inherent solubility values of **1–4** and their self-association constant *K_s* determined in 20 mM sodium phosphate buffer, pD 7.4

	1a	1b	2a	2b	3a	3b	4a^a	4b^a
Solub. (mM)	466	38	396	343	102	336	105	14
<i>K_s</i> (M ^{−1})	30	92	12	6	3	49	47	624

^a Values from the literature.²¹



1a (2a) is 61-fold (3500-fold) lower than that measured for **1acid (2acid)** which amounts to a destabilization of the dimer by 2.4 (4.8) kcal mol⁻¹ which we attribute to electrostatics. The recognition of the importance of electrostatic interactions that discourage self-association of **1–4** provides a rationale for the superior binding constants observed previously for **4a** and **4c** relative to **4d**^{12g,j} which lacks the anionic solubilizing groups which translates into the superior solubilizing abilities of **4a** and **4b**.²³

Construction of phase solubility diagrams for 1–3 with insoluble drugs

In order to assess the ability of **1–3** as solubilizing agents for insoluble drugs we created phase solubility diagrams (PSD)^{29,31} for the six new containers **1–3**. Phase solubility diagrams are plots of concentration of container on the *x*-axis versus concentration of solubilized poorly soluble drug on the *y*-axis. Several types of PSDs are possible (*A_L*, *A_P*, *A_N*; Fig. 7) although linear (*A_L*-type) PSDs are most common for cyclodextrin molecular containers. Containers that display *A_L*-type PSDs behave according to eqn (1) where *S₀* is the inherent solubility of the drug, *K_a* is the binding constant for the container drug complex, and slope is the slope of the PSD.²⁹ In this paper, we generally observed *A_L*-type PSDs except for **2a** and **2b** with ziprasidone and **2a** with PBS-1086 which displayed *A_P*-type behaviour. We consider PSDs with slopes ≥ 0.5 (e.g. a 50 mM solution of container solubilizes 25 mM drug) as indication that a given container is a very good solubilizing agent for a given drug. If we substitute slope = 0.5 into eqn (1) then it is easy to show that *K_a* × *S₀* = 1.²³ Alternatively, if we want to solubilize a drug with inherent solubility of 1 × 10⁻⁵ M (1 × 10⁻⁶ M) then the container must display a binding constant of 1 × 10⁵ M⁻¹ (1 × 10⁶ M⁻¹) to achieve a slope of 0.5.

$$K_a = \frac{\text{slope}}{S_0(1 - \text{slope})} \quad (1)$$

In this section we report the phase solubility behavior of **1–3** with four insoluble drugs (camptothecin, PBS-1086, β-estradiol, and ziprasidone) which we have previously studied with **4a** and **4b**.^{21,23} To create the PSDs we prepare a solution of container of known concentration and stir it with an excess of solid insoluble drug. After equilibrium is reached, the excess of insoluble drug is removed by centrifugation and filtration. The concentration of drug in the filtrate is determined by ¹H NMR spectroscopy after addition of benzene-1,3,5- tricar-

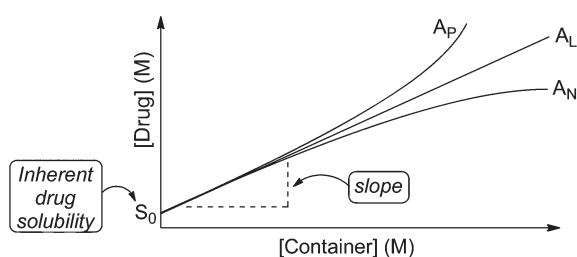


Fig. 7 Idealized phase solubility diagrams.

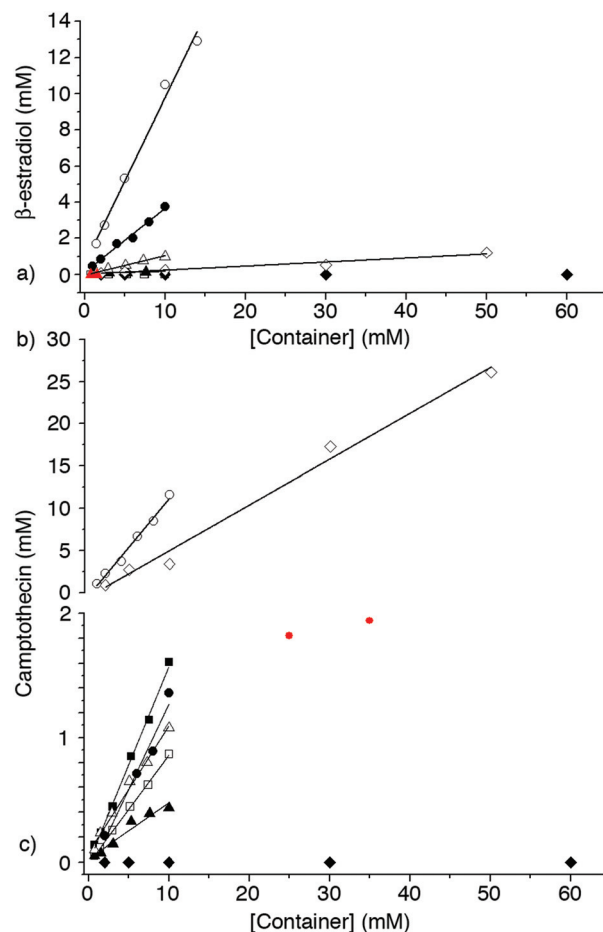


Fig. 8 Phase solubility diagrams created for: (a) **1–4** with β-estradiol, (b) **1b** and **4b** with camptothecin, and (c) **1a**, **2**, **3**, and **4a** with camptothecin. Conditions: 20 mM sodium phosphate buffer, room temperature, pD 7.4. Symbols: **1a**, ◆; **1b**, ◇; **2a**, ■; **2b**, □; **3a**, ▲; **3b**, △; **4a**, ●; **4b**, ○. Data points colored red were not used for linear fitting.

oxylic acid (1.00 mM) as a non-binding internal standard using the relative integrals of diagnostic resonances of the drug versus those of the internal standard. Fig. 8a shows the PSDs for solutions of containers **1–4** with the insoluble drug β-estradiol. As can be readily seen most of the PSDs are of the *A_L*-type and it is therefore appropriate to perform a linear fitting of the data to determine the value of the slope. The slope values range from 0 to 0.92 for **4b** which indicates that the affinity of the various containers toward β-estradiol differ significantly. Next, we substituted the measured slope values and the known inherent solubilities (*S₀*)²³ into eqn (1) to determine the affinity constant (*K_a*, M⁻¹) for the interaction of each container β-estradiol complex (Table 2). In Table 2, the values of slope and *K_a* and their uncertainty are given to two significant figures which reflects the limitations of phase solubility measurements. Similar experiments were performed for camptothecin with containers **1–3** and compared with the data previously reported for **4** (Fig. 8b,c and Table 2).^{21,23} Fig. 8c shows that the PSD for **4a** and camptothecin displays a plateau region above [4a] = 10 mM which reflects the limited solubility

Table 2 Inherent solubility (S_0 , μM) and slope values calculated from the linear region of the PSDs for containers **1–4** with the four poorly soluble drugs. The corresponding K_a (M^{-1}) values were calculated using eqn (1)^a

		Camptothecin	Ziprasidone	PBS-1086	β -estradiol
1a	S_0	54 ± 3.9	63 ± 3.5	4.5 ± 0.90	8.8 ± 0.42
	slope	—	—	—	—
1b	K_a	—	—	—	—
	slope	0.54 ± 0.032	—	—	0.022 ± 0.0026
2a	K_a	$2.2(\pm 0.16) \times 10^4$	—	—	$2.6(\pm 0.32) \times 10^3$
	slope	0.16 ± 0.0043	NL	NL	—
2b	K_a	$3.5(\pm 0.27) \times 10^3$	—	—	—
	slope	0.087 ± 0.0012	NL	0.046 ± 0.0040	—
3a	K_a	$1.8(\pm 0.13) \times 10^3$	—	$1.1(\pm 0.24) \times 10^4$	—
	slope	0.045 ± 0.0051	0.070 ± 0.012	0.032 ± 0.0018	0.013 ± 0.0011
3b	K_a	$8.7(\pm 1.2) \times 10^2$	$1.2(\pm 0.22) \times 10^3$	$7.4(\pm 1.5) \times 10^3$	$1.5(\pm 0.14) \times 10^3$
	slope	0.10 ± 0.0051	0.38 ± 0.026	0.14 ± 0.0056	0.11 ± 0.0058
4a^b	K_a	$2.0(\pm 0.18) \times 10^3$	$9.8(\pm 0.95) \times 10^3$	$3.6(\pm 0.74) \times 10^4$	$1.4(\pm 0.10) \times 10^4$
	slope	0.14 ± 0.0070	1.1 ± 0.19	0.71 ± 0.027	0.35 ± 0.019
4b^b	K_a	$2.9(\pm 0.26) \times 10^3$	TL	$5.5(\pm 1.2) \times 10^5$	$6.2(\pm 0.48) \times 10^4$
	slope	1.1 ± 0.059	0.43 ± 0.052	0.89 ± 0.0043	0.92 ± 0.053
	K_a	TL	$1.2(\pm 0.20) \times 10^4$	$1.9(\pm 0.38) \times 10^6$	$1.3(\pm 0.86) \times 10^6$

^a NL = non-linear PSD; — = could not be determined because the slope is too small to measure. TL = too large to be determined from PSD.^b Lit. data.^{21,23}

of the **4a** camptothecin complex. The data points shown in red (Fig. 8c) are excluded from the linear fitting used to calculate slope and K_a . Using the same procedures, we generated PSDs for systems comprising containers **1–3** with ziprasidone and the developmental anti-cancer agent PBS-1086³² and calculated K_a values (Table 1 and ESI†) for the container drug complexes.

Interpretation of the trends in the K_a values and solubilization efficiency

This section discusses the trends in the K_a values as a function of the structural variables of the containers and the guests.

Influence of the number of glycoluril rings. As shown in Fig. 3, the crystal structures of **1–4** demonstrate that compounds **1** and **2** are molecular clips with divergent and slightly convergent aromatic sidewalls, respectively. In contrast, containers **3** and **4** are more properly defined as acyclic CB[n]-type receptors because they feature more fully formed ureidyl C=O portals and closed cavities that are fully defined by the glycoluril rings and the aromatic sidewalls. Accordingly, it is appropriate to compare the solubilization properties of **1** and **2** and separately, **3** and **4**. We find that container **1a** is uniformly unsuccessful as a solubilizing agent, whereas container **2a** is able to solubilize camptothecin, ziprasidone, and PBS-1086. We attribute the better solubilizing ability of **2a** to the more nearly parallel alignment of its aromatic walls (centroid to centroid distance = 7.40 \AA) which preorganizes it to engage in π - π interactions with drugs that contain aromatic rings. In contrast, the behavior of containers **1b** and **2b** toward the four drugs is drug dependent. For example, container **1b** performs better for camptothecin and β -estradiol but **2b** performs better for ziprasidone and PBS-1086. In this regard it is noteworthy that the PSDs for **2a** and **2b** with ziprasidone are non-linear which is indicative of higher order container_n-drug complexes.

The behavior of acyclic CB[n]-type containers **3** and **4** toward the four drugs exhibit clear trends. For example, container **3a** binds less strongly ($K_a \approx 10^3 - 10^4 \text{ M}^{-1}$) to all four drugs than **4a** does ($2.9 \times 10^3 < K_a < 5.5 \times 10^5 \text{ M}^{-1}$). We attribute this result to the larger hydrophobic cavity of **4a** relative to **3a** and the more fully formed ureidyl C=O portals of **4a** which results in stronger ion-dipole interactions with cationic drugs (e.g. ziprasidone). Similarly, container **3b** ($2.0 \times 10^3 < K_a < 3.6 \times 10^4 \text{ M}^{-1}$) does not perform as well as **4b** ($1.2 \times 10^4 < K_a < 1.9 \times 10^6 \text{ M}^{-1}$) as a solubilizing agent for these four drugs. Accordingly, we conclude that container **4** is a more efficient solubilizing agent than container **3** although the relative *in vivo* efficacy of drugs formulated by the different containers will depend on the ability of the drugs to be released by dilution and competition from endogenous cationic small molecules (e.g. spermine).³³

Influence of aromatic sidewall. The influence of the nature of the aromatic sidewall (e.g. benzene *versus* naphthalene) is clear cut for containers **1**, **3**, **4**. In most cases the containers with the longer aromatic sidewall (e.g. naphthalene) display higher K_a values toward the drugs. For example, container **1b** binds nicely ($K_a = 2.2 \times 10^4$ and $2.6 \times 10^3 \text{ M}^{-1}$) toward camptothecin and β -estradiol whereas the binding of **1a** to these drugs could not be detected. Similarly, the ratio of the K_a values of **3b** *versus* **3a** toward camptothecin (2.3), ziprasidone (8.2), PBS-1086 (4.9), and β -estradiol (9.3) indicate that **3b** is a significantly better host than **3a**. Containers **4b** and **4a** display a similar trend with the ratio of K_a values for **4b** *versus* **4a** toward camptothecin (>6.4), PBS-1086 (3.5), and β -estradiol (21). Interestingly, ziprasidone which is both narrow and cationic at pD 7.4 prefers to bind to the smaller cavity of container **4a** relative to **4b**. We conclude that the containers with substituted naphthalene sidewalls generally outperform those with *o*-xylylene sidewalls because they possess larger hydrophobic cavities which should increase the number of solvating



H₂O molecules expelled upon binding^{6d,34} and result in the formation of π - π interactions between larger π -surfaces particularly for insoluble drugs that contain aromatic rings.

Conclusions

In summary, we have reported the synthesis of a series of molecular container compounds (**1–3**) that differ in the number of glycoluril rings that connect the two terminal substituted *o*-xylylene or naphthalene sidewalls. Compounds **1–3** display very good aqueous solubility (>38 mM) and somewhat surprisingly do not undergo significant self-association in aqueous solution ($K_s \leq 92 \text{ M}^{-1}$). We trace the low values of K_s observed for **1** and **2** to unfavorable sulfonate-sulfonate electrostatic interactions (2.4 to 4.8 kcal mol⁻¹) that would occur in the putative dimeric complexes **1**₂ and **2**₂. The X-ray crystal structure of acyclic CB[n] type receptor **3b** shows a cavity shaped by its two naphthalene walls that do not engage in π - π or CH $\cdots\pi$ interactions. PSDs were created for **1–3** with four insoluble drugs (camptothecin, ziprasidone, PBS-1086, and β -estradiol) and compared with the PSDs measured previously for **4a** and **4b**. We find that the acyclic CB[n]-type containers containing larger numbers of glycoluril rings (e.g. **4**) and the larger naphthalene sidewalls (e.g. **4a** versus **4b**) are generally superior solubilizing agents for insoluble drugs. For the molecular clip receptors **1** and **2** the results are less clear cut although the naphthalene walled compounds often display higher K_a values than the *o*-xylylene walled analogues and more often exhibit the desirable A_L-type PSDs. When combined with our previous studies on the influence of the nature of the solubilizing group (e.g. anionic versus neutral versus cationic) and the aromatic sidewalls (benzene, two isomeric naphthalenes, alkylated variants), this study allows us to conclude that compounds **4a** and **4b** are most efficient as solubilizing agents for insoluble drugs and are best positioned for further development as solubilizing excipients for real world pharmaceutical applications. In this regard the higher values of K_a exhibited by acyclic CB[n]-type receptors compared to HP- β -cyclodextrin promises to broaden the range of drugs that can be efficiently solubilized using molecular container technology and to do so at lower concentrations of container.

Experimental

General experimental

Starting materials were purchased from commercial suppliers and were used without further purification or were prepared by literature procedures. Compounds **1CE** and **2NH** were prepared according to literature procedures.^{21,25} Melting points were measured on a Meltemp apparatus in open capillary tubes and are uncorrected. IR spectra were measured on a JASCO FT/IR 4100 spectrometer and are reported in cm⁻¹. NMR spectra were measured on commercial spectrometers operating at 600 MHz and 400 MHz for ¹H and 125 MHz for

¹³C. Mass spectrometry was performed using a JEOL AccuTOF electrospray instrument (ESI⁺) or on a Bruker 12T Apex IV FT-ICR mass spectrometer.

Compound 1a. A solution of dimethylglycoluril bis(cyclic ether) **1CE** (0.650 g, 2.56 mmol) in TFA (7 mL) was mixed with compound **5** (3.96 g, 10.2 mmol). The mixture was stirred and heated at 70 °C for 3 h and then was poured into MeOH (70 mL). The solid was collected by filtration and was dried under high vacuum. The crude solid was recrystallized two times from a mixture of water and acetone (1 : 2, v/v, 20 mL). The solid was dissolved in water (10 mL) and adjusted to pH = 7 by adding 1 M aqueous NaOH. The solvent was removed by rotary evaporation and then the solid was further dried under high vacuum to yield compound **1a** (2.16 g, 83%) as a white solid. M.p. >280 °C. IR (ATR, cm⁻¹): 2925w, 1684s, 1477s, 1437m, 1355w, 1311m, 1261s, 1195s, 1095m, 1051s, 800m, 768m, 616m, 532m. ¹H NMR (600 MHz, D₂O): δ 6.85 (s, 4H), 5.18 (d, J = 16.2, 4H), 4.14 (d, J = 16.2, 4H), 4.05–3.90 (m, 8H), 3.15–2.95 (m, 8H), 2.20–2.05 (m, 8H), 1.77 (s, 6H). ¹³C NMR (125 MHz, D₂O, 1,4-dioxane as internal reference): δ 157.2, 149.4, 127.6, 114.4, 78.2, 68.2, 47.4, 34.4, 23.2, 15.0 (10 out of the 12 expected resonances are observed). High-Res MS (ESI): m/z 925.1607 ([M – 4Na + 3H]⁺, C₃₄H₄₅N₄O₁₈S₄, calculated for 925.1612).

Compound 1b. A solution of dimethylglycoluril bis(cyclic ether) **1CE** (0.650 g, 2.56 mmol) in TFA (7 mL) was mixed with compound **6** (4.57 g, 10.2 mmol). The mixture was stirred and heated at 70 °C for 3 h and then was poured into MeOH (70 mL). The solid was collected by filtration and was dried under high vacuum. The crude solid was recrystallized twice from a mixture of water and acetone (1 : 2, v/v, 20 mL). The solid was dissolved in water (10 mL) and adjusted to pH = 7 by adding 1 M aqueous NaOH. The solvent was removed by rotary evaporation and then the solid was further dried under high vacuum to yield compound **1b** (2.60 g, 91%) as a white solid. M.p. >280 °C. IR (ATR, cm⁻¹): 2939w, 2858w, 1690m, 1468s, 1426w, 1344m, 1307w, 1181s, 1047s, 1029s, 952w, 766m, 738m. ¹H NMR (600 MHz, D₂O): δ 8.00–7.95 (m, 4H), 7.60–7.55 (m, 4H), 5.20 (d, J = 16.2, 4H), 4.39 (d, J = 16.2, 4H), 3.65–3.55 (m, 8H), 2.95–2.75 (m, 8H), 2.15–2.05 (m, 8H), 1.81 (s, 6H). ¹³C NMR (125 MHz, D₂O, 1,4-dioxane as internal reference): δ 157.1, 147.8, 126.8, 125.9, 121.3, 78.2, 73.7, 47.6, 36.0, 24.6, 15.5 (11 out of the 14 expected resonances were observed). High-Res MS (ESI): m/z 1091.1392 ([M – Na]⁺, C₄₂H₄₆N₄Na₃O₁₈S₄, calculated for 1091.1377).

Compound 2CE. Compound **2NH** (2.12 g, 5.81 mmol) was dissolved in TFA (58 mL). Paraformaldehyde (872 mg, 29.04 mmol) was added and the reaction mixture stirred and heated at 75–80 °C for 20 h in a pre-heated oil bath. The reaction mixture was poured into MeOH (400 mL) and the resulting precipitate was filtered and washed with MeOH. Compound **2CE** was obtained as white solid (1.04 g, 2.33 mmol, 40%). M.p. >300 °C. IR (ATR, cm⁻¹): 2999w, 2960w, 2875w, 1722s, 1468m, 1440s, 1376m, 1302s, 1232m, 1093m, 1067m, 1001m, 949m, 917m, 881m, 768m, 737m, 668w. ¹H NMR (400 MHz, DMSO-d₆): 5.47 (d, J = 15.8, 2H), 5.15 (d, J =



11.0, 4H), 4.87 (d, $J = 11.0$, 4H), 4.33 (d, $J = 15.8$, 2H), 1.83 (s, 6H), 1.65 (s, 6H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 154.5, 76.4, 72.2, 70.6, 44.3, 17.9, 16.7. HR-MS (ESI): m/z 449.1891 $[(\text{M} + \text{H})^+]$, $\text{C}_{18}\text{H}_{24}\text{N}_8\text{O}_6$, calcd 449.1897.

Compound 3CE. Compound 7 (1.196 g, 8.4 mmol) was dissolved in methane sulfonic acid (40 mL) under N_2 atmosphere. The solution was cooled to 8–12 °C in an ice bath and then 1CE (3.996 g, 15.7 mmol) was added at once. The reaction mixture was stirred at 8–12 °C for 2 h and then the ice bath was removed at the reaction was stirred at room temperature for another 2 h. The reaction was poured into precooled (5 °C) acetone (700 mL) and the resulting precipitate was obtained by filtration. The solid was washed with about 100 mL acetone and then transferred while still wet into a mixture of acetonitrile and water (1:1 (v:v), 100 mL) and sonicated. The resulting precipitate was filtered and suspended in formic acid (2 mL). Finally the solid was thoroughly dried on the frit, washed with water and methanol and dried under vacuum to give 3CE as a white solid (204 mg, 0.332 mmol, 4%). M.p. >300 °C. IR (ATR, cm^{-1}): 3551w, 3455w, 3384w, 2990w, 2949w, 1718s, 1702s, 1656w, 1465m, 1425m, 1315s, 1271m, 1239s, 1181m, 1074m, 1013s, 918m, 851s, 788m, 735m, 663w. ^1H NMR (400 MHz, $\text{DMSO}-d_6$): 5.58–5.52 (m, 6H), 5.16 (d, $J = 10.9$ Hz, 2H), 4.85 (d, $J = 11.0$ Hz, 4H), 4.24 (d, $J = 15.1$ Hz, 4H), 1.81 (s, 6H), 1.64 (s, 6H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$, 30 °C): δ 155.2, 154.8, 154.7, 77.1, 72.4, 70.5, 69.9, 48.5, 17.7, 15.7. HR-MS (ESI): m/z 615.2363 $[(\text{M} + \text{H})^+]$, $\text{C}_{24}\text{H}_{30}\text{N}_{12}\text{O}_8$, calcd 615.2388.

Compound 2a. Compound 2CE (448 mg, 1.0 mmol) was dissolved in TFA- Ac_2O (1:1 (v:v), 8.0 mL). Compound 5 (916 mg, 2.30 mmol) was added. The reaction was stirred and heated in a preheated oil bath for 3 h at 70 °C. Then the mixture was poured into acetone (75 mL). The precipitate was collected by centrifugation. The solid was dissolved in water (10 mL) and precipitated by the addition of EtOH (75 mL). The precipitate was collected by centrifugation, then redissolved in water (40 mL) and reprecipitated by the addition of acetone (120 mL). The precipitate was collected by filtration, dissolved in water and the pH of the solution adjusted to 7 by addition of 1 M aqueous NaOH. The solvent was removed under reduced pressure and the residue dried under vacuum. Compound 2a was obtained as brownish solid (533 mg, 0.475 mmol, 48%). M.p. >300 °C. IR (ATR, cm^{-1}): 3450w, 2946w, 2876w, 1706s, 1464s, 1391w, 1298m, 1179s, 1039s, 883w, 800m, 754m, 657w, 594m. ^1H NMR (400 MHz, D_2O): 6.98 (s, 4H), 5.35 (d, $J = 16.2$, 2H), 5.22 (d, $J = 16.2$, 4H), 4.32 (d, $J = 16.2$, 2H), 4.11 (d, $J = 16.2$, 4H), 4.10–4.00 (m, 4H), 3.90–3.80 (m, 4H), 3.20–3.00 (m, 8H), 2.25–2.10 (m, 8H), 1.74 (s, 6H), 1.67 (s, 6H). ^{13}C NMR (125 MHz, D_2O , 1,4-dioxane as internal reference): δ 156.5, 150.9, 128.9, 116.7, 79.4, 77.5, 70.3, 48.8, 44.3, 35.5, 25.3, 16.2, 16.1. HR-MS (ESI): m/z $[(\text{M} - \text{Na})^-]$, 1185.1880, $\text{C}_{42}\text{H}_{52}\text{N}_8\text{O}_{20}\text{S}_4\text{Na}_3$, calcd 1185.1868.

Compound 2b. Compound 2CE (448.4 mg, 1.00 mmol) was dissolved in TFA- Ac_2O (1:1 (v:v), 6.5 mL). Compound 6 (1.79 g, 4.00 mmol) was added and the reaction mixture stirred and heated for 3 h at 75–80 °C in a pre-heated oil bath. The

solution was poured into MeOH (75 mL) and filtered. The residue was dissolved in water (10 mL) and then acetone (50 mL) was added. The precipitate was obtained by filtration and then dissolved in water. The pH of the solution was adjusted to 7 by the addition of 1 M aqueous NaOH. The solvent was removed under reduced pressure and the residue was dried in high vacuum yielding compound 2b as an off-white solid (752 mg, 0.57 mmol, 57%). M.p. >300 °C. IR (ATR, cm^{-1}): 3446m, 2945w, 2884w, 1715s, 1463s, 1344m, 1308m, 1267w, 1176s, 1098m, 1033s, 901w, 820w, 756m, 595m. ^1H NMR (400 MHz, D_2O): 7.90–7.75 (m, 8H), 5.35 (d, $J = 16.3$, 2H), 5.18 (d, $J = 16.2$, 4H), 4.34 (d, $J = 16.3$, 2H), 4.23 (d, $J = 16.2$, 4H), 3.75–3.65 (m, 4H), 3.30–3.20 (m, 4H), 3.15–3.05 (m, 4H), 3.00–2.85 (m, 4H), 2.10–1.95 (m, 4H), 1.90–1.75 (m, 4H), 1.75 (s, 6H), 1.70 (s, 6H). ^{13}C NMR (150 MHz, D_2O , 1,4-dioxane as internal reference): δ 156.2, 148.2, 128.2, 128.1, 127.4, 123.4, 78.8, 77.4, 74.0, 48.7, 44.0, 36.5, 25.9, 16.5, 16.4. HR-MS (ESI): m/z 677.0957 $[(\text{M} + \text{Na})^{2+}]$, $\text{C}_{50}\text{H}_{56}\text{N}_8\text{O}_{20}\text{S}_4\text{Na}_5$, calcd 677.0940.

Compound 3a. To a solution of compound 3CE (0.91 g, 1.48 mmol) in TFA- Ac_2O (1:1 (v:v), 20.8 mL) compound 5 (1.37 g, 3.44 mmol) was added. The mixture was stirred and heated at 75–80 °C for 3 h and then poured into MeOH (75 mL). The solid was collected by centrifugation. The residue was washed twice with MeOH (45 mL) and dried under vacuum. The off-white solid was dissolved in water and the pH was adjusted to 7 by addition of 1 M aqueous NaOH. The solvent was removed under reduced pressure. The residue was mixed with acetone (100 mL) and the solid collected by centrifugation. The solid was washed once with acetone (45 mL) and MeOH (45 mL) and then dissolved in water. The pH was checked to be at 7. The solvent was removed under reduced pressure and the residue dried under vacuum to yield compound 3a as a yellow solid (983 mg, 0.715 mmol, 48%). M.p. >300 °C. IR (ATR, cm^{-1}): 3444w, 2941w, 1713s, 1466s, 1380w, 1316m, 1237m, 1183s, 1091m, 1037s, 976w, 922w, 841m, 789m, 729w, 593m. ^1H NMR (D_2O , 400 MHz): 7.02 (s, 4H), 5.51 (d, $J = 15.8$, 4H), 5.40 (s, 2H), 5.30 (d, $J = 16.3$, 4H), 4.25 (d, $J = 15.8$, 4H), 4.19 (d, $J = 16.3$, 4H), 4.15–4.00 (m, 8H), 3.20–3.05 (m, 8H), 2.25–2.15 (m, 8H), 1.77 (s, 6H), 1.74 (s, 6H). ^{13}C NMR (125 MHz, D_2O , dioxane as internal reference): δ 157.6, 157.00, 150.9, 128.8, 116.0, 79.6, 78.3, 71.8, 69.7, 49.0, 48.9, 35.6, 25.3, 16.6, 15.8. HR-MS (ESI): m/z 1353.2513 $[(\text{M} - \text{Na} + 2\text{H})^+]$, $\text{C}_{48}\text{H}_{60}\text{N}_{12}\text{O}_{22}\text{S}_4\text{Na}_3$, calcd 1353.2521.

Compound 3b. Compound 3CE (614.6 mg, 1.00 mmol) was dissolved in TFA- Ac_2O (1:1 (v:v), 6.5 mL). Compound 6 (1.79 g, 4.00 mmol) was added and the reaction mixture stirred and heated for 3 h at 72–80 °C in a pre-heated oil bath. The suspension was filtered using a glass frit and the residue washed with MeOH. The solid was dissolved in water (20 mL) and precipitated by the addition of acetone (80 mL). The precipitate was obtained by filtration, and the solid was washed with acetone and then dissolved in water. The pH of the solution was adjusted to 7 by addition of 1 M aqueous NaOH. The solvent was removed under reduced pressure. Finally, the yellowish solid was dried under vacuum to yield compound 3b as a yellowish solid (867 mg, 58.8 mmol, 59%). M.p. >300 °C,



IR (ATR, cm^{-1}): 3441w, 2943w, 2881w, 1716s, 1465s, 1382w, 1345m, 1313m, 1177s, 1079m, 1035s, 950m, 881w, 827m, 788m, 724m, 668m. ^1H NMR (400 MHz, D_2O): 8.00–7.90 (m, 8H), 5.50 (d, $J = 15.9$, 4H), 5.41 (s, 2H), 5.24 (d, $J = 16.4$, 4H), 4.30–4.20 (m, 8H), 4.00–3.90 (br m, 4H), 3/55–3/45 (br m, 4H), 3.08 (t, $J = 8.0$, 8H), 2.10–1.95 (br m, 8H), 1.78 (s, 6H), 1.76 (s, 6H). ^{13}C NMR (150 MHz, D_2O , dioxane as internal reference) δ 157.5, 156.9, 148.8, 128.6, 128.5, 127.6, 123.8, 79.4, 78.1, 74.7, 71.6, 48.8, 36.9, 26.0, 16.5, 16.2 (16 of the 17 expected resonances were observed). HR-MS (ESI): m/z 1453.2811 ($[\text{M} - \text{Na} + 2\text{H}]^+$), $\text{C}_{56}\text{H}_{64}\text{N}_{12}\text{O}_{22}\text{Na}_2$, calcd 1453.2834.

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Notes and references

- (a) C. A. Lipinski, *J. Pharmacol. Toxicol. Methods*, 2000, **44**, 235–249; (b) D. J. Hauss, *Adv. Drug Delivery Rev.*, 2007, **59**, 667–676.
- (a) C. Leuner and J. Dressman, *Eur. J. Pharm. Biopharm.*, 2000, **50**, 47–60; (b) R. H. Muller and C. M. Keck, *J. Biotechnol.*, 2004, **113**, 151–170.
- (a) A. K. Patri, J. F. Kukowska-Latallo and J. R. Baker, *Adv. Drug Delivery Rev.*, 2005, **57**, 2203–2214; (b) N. Blagden, M. de Matas, P. T. Gavan and P. York, *Adv. Drug Delivery Rev.*, 2007, **59**, 617–630; (c) A. T. M. Serajuddin, *Adv. Drug Delivery Rev.*, 2007, **59**, 603–616; (d) V. J. Stella and K. W. Nti-Addae, *Adv. Drug Delivery Rev.*, 2007, **59**, 677–694.
- (a) K. Okimoto, R. A. Rajewski, K. Uekama, J. A. Jona and V. J. Stella, *Pharm. Res.*, 1996, **13**, 256–264; (b) R. A. Rajewski and V. J. Stella, *J. Pharm. Sci.*, 1996, **85**, 1142–1169; (c) V. J. Stella and R. A. Rajewski, *Pharm. Res.*, 1997, **14**, 556–567; (d) M. V. Rekharsky and Y. Inoue, *Chem. Rev.*, 1998, **98**, 1875–1917.
- (a) W. A. Freeman, W. L. Mock and N.-Y. Shih, *J. Am. Chem. Soc.*, 1981, **103**, 7367–7368; (b) J. Kim, I.-S. Jung, S.-Y. Kim, E. Lee, J.-K. Kang, S. Sakamoto, K. Yamaguchi and K. Kim, *J. Am. Chem. Soc.*, 2000, **122**, 540–541; (c) A. Day, A. P. Arnold, R. J. Blanch and B. Snushall, *J. Org. Chem.*, 2001, **66**, 8094–8100; (d) S. Liu, P. Y. Zavalij and L. Isaacs, *J. Am. Chem. Soc.*, 2005, **127**, 16798–16799; (e) A. I. Day, R. J. Blanch, A. P. Arnold, S. Lorenzo, G. R. Lewis and I. Dance, *Angew. Chem., Int. Ed.*, 2002, **41**, 275–277; (f) X.-J. Cheng, L.-L. Liang, K. Chen, N.-N. Ji, X. Xiao, J.-X. Zhang, Y.-Q. Zhang, S.-F. Xue, Q.-J. Zhu, X.-L. Ni and Z. Tao, *Angew. Chem., Int. Ed.*, 2013, **52**, 7252–7255.
- (a) W. L. Mock and N.-Y. Shih, *J. Org. Chem.*, 1986, **51**, 4440–4446; (b) S. Liu, C. Ruspice, P. Mukhopadhyay, S. Chakrabarti, P. Y. Zavalij and L. Isaacs, *J. Am. Chem. Soc.*, 2005, **127**, 15959–15967; (c) M. V. Rekharsky, T. Mori, C. Yang, Y. H. Ko, N. Selvapalam, H. Kim, D. Sobransingh, A. E. Kaifer, S. Liu, L. Isaacs, W. Chen, S. Moghaddam, M. K. Gilson, K. Kim and Y. Inoue, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 20737–20742; (d) F. Biedermann, V. D. Uzunova, O. A. Scherman, W. M. Nau and A. De Simone, *J. Am. Chem. Soc.*, 2012, **134**, 15318–15323; (e) L. Cao, M. Sekutor, P. Y. Zavalij, K. Mlinaric-Majerski, R. Glaser and L. Isaacs, *Angew. Chem., Int. Ed.*, 2014, **53**, 988–993.
- (a) L. Isaacs, *Acc. Chem. Res.*, 2014, **47**, 2052–2062; (b) Y. H. Ko, E. Kim, I. Hwang and K. Kim, *Chem. Commun.*, 2007, 1305–1315; (c) A. Hennig, H. Bakirci and W. M. Nau, *Nat. Methods*, 2007, **4**, 629–632; (d) R. Dsouza, A. Hennig and W. Nau, *Chem. – Eur. J.*, 2012, **18**, 3444–3459; (e) D.-W. Lee, K. Park, M. Banerjee, S. Ha, T. Lee, K. Suh, S. Paul, H. Jung, J. Kim, N. Selvapalam, S. Ryu and K. Kim, *Nat. Chem.*, 2011, **3**, 154–159; (f) Y. Ahn, Y. Jang, N. Selvapalam, G. Yun and K. Kim, *Angew. Chem., Int. Ed.*, 2013, **52**, 3140–3144; (g) Y. Miyahara, K. Abe and T. Inazu, *Angew. Chem., Int. Ed.*, 2002, **41**, 3020–3023; (h) J. Del Barrio, P. Horton, D. Lairez, G. Lloyd, C. Toprakcioglu and O. Scherman, *J. Am. Chem. Soc.*, 2013, **135**, 11760–11763.
- (a) G. Hettiarachchi, D. Nguyen, J. Wu, D. Lucas, D. Ma, L. Isaacs and V. Briken, *PLoS One*, 2010, **5**, e10514; (b) V. D. Uzunova, C. Cullinane, K. Brix, W. M. Nau and A. I. Day, *Org. Biomol. Chem.*, 2010, **8**, 2037–2042; (c) R. Oun, R. S. Floriano, L. Isaacs, E. G. Rowan and N. J. Wheate, *Toxicol. Res.*, 2014, **3**, 447–455.
- (a) N. Dong, S.-F. Xue, Q.-J. Zhu, Z. Tao, Y. Zhao and L.-X. Yang, *Supramol. Chem.*, 2008, **20**, 659–665; (b) N. Dong, X. Wang, J. Pan and Z. Tao, *Acta Chim. Sin.*, 2011, **69**, 1431–1437; (c) N. Saleh, A. L. Koner and W. M. Nau, *Angew. Chem., Int. Ed.*, 2008, **47**, 5398–5401; (d) S. Walker, R. Oun, F. J. McInnes and N. J. Wheate, *Isr. J. Chem.*, 2011, **51**, 616–624; (e) Y. J. Jeon, S.-Y. Kim, Y. H. Ko, S. Sakamoto, K. Yamaguchi and K. Kim, *Org. Biomol. Chem.*, 2005, **3**, 2122–2125.
- (a) E. Appel, J. del Barrio, X. Loh and O. Scherman, *Chem. Soc. Rev.*, 2012, **41**, 6195–6214; (b) J. Zhang, R. J. Coulston, S. T. Jones, J. Geng, O. A. Scherman and C. Abell, *Science*, 2012, **335**, 690–694.
- (a) L. Cao, G. Hettiarachchi, V. Briken and L. Isaacs, *Angew. Chem., Int. Ed.*, 2013, **52**, 12033–12037; (b) E. Kim, D. Kim, H. Jung, J. Lee, S. Paul, N. Selvapalam, Y. Yang, N. Lim, C. G. Park and K. Kim, *Angew. Chem., Int. Ed.*, 2010, **49**, 4405–4408; (c) H. Jung, K. M. Park, J.-A. Yang, E. J. Oh, D.-W. Lee, K. Park, S. H. Ryu, S. K. Hahn and K. Kim, *Biomaterials*, 2011, **32**, 7687–7694.



- 12 (a) M. Stancl, J. Svec and V. Sindelar, *Isr. J. Chem.*, 2011, **51**, 592–599; (b) L. Isaacs, *Isr. J. Chem.*, 2011, **51**, 578–591; (c) A. Chakraborty, A. Wu, D. Witt, J. Lagona, J. C. Fettingner and L. Isaacs, *J. Am. Chem. Soc.*, 2002, **124**, 8297–8306; (d) W.-H. Huang, P. Y. Zavalij and L. Isaacs, *J. Am. Chem. Soc.*, 2008, **130**, 8446–8454; (e) D. Lucas, T. Minami, G. Iannuzzi, L. Cao, J. B. Wittenberg, P. Anzenbacher and L. Isaacs, *J. Am. Chem. Soc.*, 2011, **133**, 17966–17976; (f) M. Stancl, M. Necas, J. Taraba and V. Sindelar, *J. Org. Chem.*, 2008, **73**, 4671–4675; (g) M. Stancl, M. Hodan and V. Sindelar, *Org. Lett.*, 2009, **11**, 4184–4187; (h) D. Ma, Z. Gargulakova, P. Y. Zavalij, V. Sindelar and L. Isaacs, *J. Org. Chem.*, 2010, **75**, 2934–2941; (i) M. Stancl, Z. Gargulakova and V. Sindelar, *J. Org. Chem.*, 2012, **77**, 10945–10948; (j) M. Stancl, L. Gilberg, L. Ustrnul, M. Necas and V. Sindelar, *Supramol. Chem.*, 2014, **26**, 168–172.
- 13 (a) L. Isaacs, D. Witt and J. Lagona, *Org. Lett.*, 2001, **3**, 3221–3224; (b) L. Isaacs and D. Witt, *Angew. Chem., Int. Ed.*, 2002, **41**, 1905–1907; (c) A. Wu, A. Chakraborty, J. C. Fettingner, R. A. Flowers II and L. Isaacs, *Angew. Chem., Int. Ed.*, 2002, **41**, 4028–4031.
- 14 (a) J. Lagona, J. C. Fettingner and L. Isaacs, *Org. Lett.*, 2003, **5**, 3745–3747; (b) J. Lagona, J. C. Fettingner and L. Isaacs, *J. Org. Chem.*, 2005, **70**, 10381–10392.
- 15 L. Isaacs, S.-K. Park, S. Liu, Y. H. Ko, N. Selvapalam, Y. Kim, H. Kim, P. Y. Zavalij, G.-H. Kim, H.-S. Lee and K. Kim, *J. Am. Chem. Soc.*, 2005, **127**, 18000–18001.
- 16 (a) W.-H. Huang, S. Liu, P. Y. Zavalij and L. Isaacs, *J. Am. Chem. Soc.*, 2006, **128**, 14744–14745; (b) W.-H. Huang, P. Y. Zavalij and L. Isaacs, *Angew. Chem., Int. Ed.*, 2007, **46**, 7425–7427; (c) W.-H. Huang, P. Y. Zavalij and L. Isaacs, *Org. Lett.*, 2008, **10**, 2577–2580.
- 17 (a) J. Svec, M. Necas and V. Sindelar, *Angew. Chem., Int. Ed.*, 2010, **49**, 2378–2381; (b) V. Havel, J. Svec, M. Wimmerova, M. Dusek, M. Pojarova and V. Sindelar, *Org. Lett.*, 2011, **13**, 4000–4003; (c) J. Svec, M. Dusek, K. Fejfarova, P. Stacko, P. Klan, A. E. Kaifer, W. Li, E. Hudeckova and V. Sindelar, *Chem. – Eur. J.*, 2011, **17**, 5605–5612; (d) V. Havel, V. Sindelar, M. Necas and A. E. Kaifer, *Chem. Commun.*, 2014, **50**, 1372–1374; (e) A. Revesz, D. Schroder, J. Svec, M. Wimmerova and V. Sindelar, *J. Phys. Chem. A*, 2011, **115**, 11378–11386.
- 18 (a) L. Gilberg, M. S. A. Khan, M. Enderesova and V. Sindelar, *Org. Lett.*, 2014, **16**, 2446–2449; (b) L. Cao and L. Isaacs, *Org. Lett.*, 2012, **14**, 3072–3075; (c) B. Vinciguerra, L. Cao, J. R. Cannon, P. Y. Zavalij, C. Fenselau and L. Isaacs, *J. Am. Chem. Soc.*, 2012, **134**, 13133–13140.
- 19 (a) J. B. Wittenberg, P. Y. Zavalij and L. Isaacs, *Angew. Chem., Int. Ed.*, 2013, **52**, 3690–3694; (b) M. Zhang, L. Cao and L. Isaacs, *Chem. Commun.*, 2014, **50**, 14756–14759.
- 20 (a) C. A. Burnett, D. Witt, J. C. Fettingner and L. Isaacs, *J. Org. Chem.*, 2003, **68**, 6184–6191; (b) W.-H. Huang, P. Y. Zavalij and L. Isaacs, *Org. Lett.*, 2009, **11**, 3918–3921; (c) D. Ma, P. Y. Zavalij and L. Isaacs, *J. Org. Chem.*, 2010, **75**, 4786–4795; (d) D. Ma, B. Zhang, U. Hoffmann, M. G. Sundrup, M. Eikermann and L. Isaacs, *Angew. Chem., Int. Ed.*, 2012, **51**, 11358–11362; (e) T. Minami, N. A. Esipenko, B. Zhang, L. Isaacs, R. Nishiyabu, Y. Kubo and P. Anzenbacher, *J. Am. Chem. Soc.*, 2012, **134**, 20021–20024; (f) C. Shen, D. Ma, B. Meany, L. Isaacs and Y. Wang, *J. Am. Chem. Soc.*, 2012, **134**, 7254–7257; (g) T. Minami, N. Esipenko, A. Akdeniz, B. Zhang, L. Isaacs and P. Anzenbacher, *J. Am. Chem. Soc.*, 2013, **135**, 15238–15243.
- 21 D. Ma, G. Hettiarachchi, D. Nguyen, B. Zhang, J. B. Wittenberg, P. Y. Zavalij, V. Briken and L. Isaacs, *Nat. Chem.*, 2012, **4**, 503–510.
- 22 B. Zhang, P. Y. Zavalij and L. Isaacs, *Org. Biomol. Chem.*, 2014, **12**, 2413–2422.
- 23 B. Zhang and L. Isaacs, *J. Med. Chem.*, 2014, **57**, 9554–9563.
- 24 R. P. Sijbesma, A. P. M. Kentgens, E. T. G. Lutz, J. H. van der Maas and R. J. M. Nolte, *J. Am. Chem. Soc.*, 1993, **115**, 8999–9005.
- 25 J.-J. Zhou, X. Yu, Y.-C. Zhao, X. Xiao, Y.-Q. Zhang, Q.-J. Zhu, S.-F. Xue, Q.-J. Zhang, J.-X. Liu and Z. Tao, *Tetrahedron*, 2014, **70**, 800–804.
- 26 L. Isaacs, D. Witt and J. C. Fettingner, *Chem. Commun.*, 1999, 2549–2550.
- 27 D. Witt, J. Lagona, F. Damkaci, J. C. Fettingner and L. Isaacs, *Org. Lett.*, 2000, **2**, 755–758.
- 28 (a) Z.-G. Wang, B.-H. Zhou, Y.-F. Chen, G.-D. Yin, Y.-T. Li, A.-X. Wu and L. Isaacs, *J. Org. Chem.*, 2006, **71**, 4502–4508; (b) N.-F. She, X.-G. Meng, M. Gao, A.-X. Wu and L. Isaacs, *Chem. Commun.*, 2008, 3133–3135; (c) N.-F. She, M. Gao, X.-G. Meng, G.-F. Yang, J. A. A. W. Elemans, A.-X. Wu and L. Isaacs, *J. Am. Chem. Soc.*, 2009, **131**, 11695–11697; (d) A. Wu, P. Mukhopadhyay, A. Chakraborty, J. C. Fettingner and L. Isaacs, *J. Am. Chem. Soc.*, 2004, **126**, 10035–10043; (e) S. Ghosh, A. Wu, J. C. Fettingner, P. Y. Zavalij and L. Isaacs, *J. Org. Chem.*, 2008, **73**, 5915–5925.
- 29 K. A. Connors, *Binding constants*, John Wiley & Sons, New York, 1987.
- 30 (a) A. E. Rowan, J. A. A. W. Elemans and R. J. M. Nolte, *Acc. Chem. Res.*, 1999, **32**, 995–1006; (b) J. N. H. Reek, J. A. A. W. Elemans, R. de Gelder, P. T. Beurskens, A. E. Rowan and R. J. M. Nolte, *Tetrahedron*, 2002, **59**, 175–185.
- 31 T. Higuchi and K. A. Connors, *Adv. Anal. Chem. Inst.*, 1965, **4**, 117–212.
- 32 (a) C. Fabre, N. Mimura, K. Bobb, S.-Y. Kong, G. Gorgun, D. Cirstea, J. Hu, J. Minami, H. Ohguchi, J. Zhang, J. Meshulam, R. D. Carrasco, Y.-T. Tai, P. G. Richardson, T. Hideshima and K. C. Anderson, *Clin. Cancer Res.*, 2012, **18**, 4669–4681; (b) J. Zhang, D. R. Sliskovic and C. E. Ducker (Profectus BioSciences, Inc., USA), *7-oxabicyclo [4.1.0]heptenyl heteroaryl amide derivatives as inhibitors of nf-kb useful for treatment of cancer, inflammation, autoimmune diseases, diabetes, infections, cardiovascular diseases and ischemia-reperfusion injuries*, WO 2010111460, 2010.
- 33 V. J. Stella, V. M. Rao, E. A. Zannou and V. Zia, *Adv. Drug Delivery Rev.*, 1999, **36**, 3–16.
- 34 W. M. Nau, M. Florea and K. I. Assaf, *Isr. J. Chem.*, 2011, **51**, 559–577.

