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Two-colour fluorescent imaging in organisms using self-assembled nano-systems of upconverting nanoparticles and molecular switches†

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The water-insolubility of a potentially versatile photoresponsive 'turn-on' fluorescence probe was overcome by incorporating it into a nano-assembly containing an upconverting nanoparticle wrapped in an amphiphilic polymer. The appeal of the nano-system is not only in the ability to turn "on" and "off" the fluorescence from the organic chromophore using UV and visible light, it is in the fact that the nanoparticle acts as a static probe because it emits red and green light when excited by near infrared light, which is not effected by UV and visible light. This dual-functioning emission behaviour was demonstrated in live organisms.

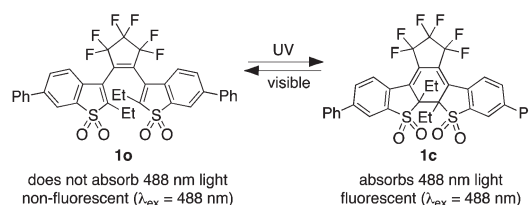
Introduction

Although the use of conventional fluorescent probes¹ for bio-imaging applications has proven experimentally and clinically beneficial, the use of 'turn-on' fluorescent probes² has its own merits. These probes are engineered to emit read-out signals only after they are activated by an external stimulus.² When the external stimulus involves a chemical reaction or interaction, the appearance of emission is a convenient way to validate site-specific binding or detect biological events in cells or living organisms. On the other hand, photoresponsive probes that are turned 'on' and 'off' using two colours of stimulating light when and where the user desires adds the component of time to imaging methods. This benefit offers the ability to separate the spatially indistinguishable fluorophores temporally, and allows the reconstruction of images with subdiffraction

resolution to improve the quality of images.³ These features explain why 'turn-on' fluorescent probes are frequently used as tools for cell imaging where they can detect chemical species⁴ and imaging proteins in cells,⁵ while photoactive 'turn-on' fluorescent probes can be used for super-resolution imaging.³

A recently reported example of a probe that changes from a dark, non-emissive state to a bright, emissive state was based on the reversible photochemical reaction of the disulfone **1o** shown in Scheme 1.⁶ This appealing system has an impressive emission contrast ratio between its two states and is based on the well known ring-closing and ring-opening reactions of diarylethenes using two different colours of light.⁷ In the case of this diarylethene, its non-emissive ring-open isomer **1o** is quantitatively converted to its highly emissive ring-closed isomer **1c** using UV light (**1o** → **1c**). The broad emission of **1c** is between 500 nm and 650 nm when using light of 488 nm as the excitation source. The emission can be turned off using visible light, which triggers the ring-opening reaction (**1c** → **1o**) and regenerates the original non-emissive compound.

We have demonstrated how multi-component, water-soluble nano-systems are self-assembled when an amphiphilic polymer wraps itself around inorganic upconverting nanoparticles (UCNPs).⁸ The amphiphilic nature of the polymer ensures the entire nano-system has good compatibility with aqueous environments while providing a lipophilic layer close to the surface of the nanoparticles. It is within this lipophilic



Scheme 1 Photoinduced ring-closing and ring-opening reactions of the diarylethene (**1o** and **1c**).

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layer that hydrophobic compounds such as **1o** are entrapped if they are present during the self-assembly process.

In this article, we describe the 'turn-on' properties of a self-assembled nano-system containing compound **1o**, which becomes reversibly emissive when exposed to UV and visible light. The appeal of the system is that the nanoparticles display their own emission when they absorb several photons of near-infrared (NIR) light and convert them into red and green ones.⁹ The resulting system is multi-modal (it has two fluorescent probes). The emission from the UCNPs (stimulated by NIR light) is always 'on' and can always be used as a read-out signal even when the signal from the 'turn-on' probe is 'off'. Using UV light to trigger the ring-closing reaction of **1o** and consequently turning the emission from the organic chromophore 'on' provides a greater level of reliability and a reduction in false positive signals when tracking biochemical events.¹⁰ We demonstrate the use of our nano-system imaging agent in live *Caenorhabditis elegans* nematodes.

Results and discussion

The self-assembled nano-system **1o-NP** is shown in Fig. 1 and was synthesized using the reported one-pot protocol^{8d} by treat-

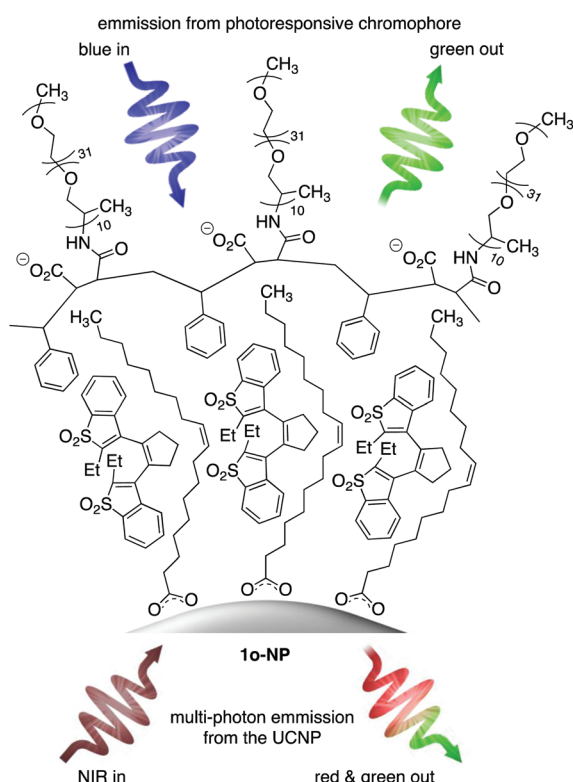


Fig. 1 The self-assembled nano-system (**1o-NP**) containing the amphiphilic polymer wrapped around the lanthanide-doped upconverting nanoparticles with encapsulated diarylethene chromophores. The green emission from the organic chromophore is turned 'on' using UV light to ring-close the diarylethene. The multiphoton emission from the UCNPs is always on when irradiated with NIR light.

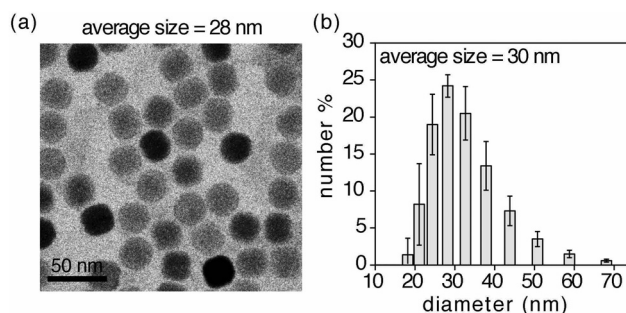


Fig. 2 (a) TEM image of the nano-assemblies (**1o-NP**). (b) DLS measurements of the size distribution of the nano-assemblies. The average sizes calculated using each technique are provided. For the DLS measurements, the original aqueous dispersion (0.4 mg mL^{-1} as determined by TGA; $0.8 \times 10^{-6} \text{ M}$ **1o**) was diluted 100 times with water and this diluted sample was employed for the measurements. The sample was incubated at 25°C for 2 min before the DLS measurements were acquired.

ing a CHCl_3 dispersion of oleate-coated lanthanide-doped (2 mol% Er^{3+} and 20% Yb^{3+}) upconverting nanoparticles ($\text{NaYF}_4:\text{ErYb}$) and poly(styrene-*co*-maleic anhydride) with poly(propylene glycol)bis(2-aminopropyl ether) to ring-open the anhydride groups. Introducing the hydrophobic chromophore (**1o**) and replacing the organic solvent with basic water promotes the self-assembly process and produces the nano-system **1o-NP**, which were isolated by passing the aqueous sample through syringe filters.[‡]

Transmission electron microscopy (TEM) images of **1o-NP** (Fig. 2a) show an average diameter of 28 nm for the nano-systems (Fig. S1†). The average size is measured to be slightly larger (30 nm) by dynamic light scattering (DLS) because this technique measures the average hydrodynamic radius, which includes the polymer shell (Fig. 2b). This organic shell cannot be directly observed by TEM due to its lack of contrast on the TEM grid. The shell thickness of 2 nm is substantially smaller than what it would be if all the polyethylene glycol chains were fully extended (11 nm) suggesting the polymer shell exists in a compressed form as has been previously observed.^{8a,b}

As is typically done for photochromic compounds, the ring-closing (**1o-NP** \rightarrow **1c-NP**) and ring-opening (**1c-NP** \rightarrow **1o-NP**) reactions of the chromophores entrapped within the nano-assemblies are best monitored using UV-vis absorption spectroscopy. When an aqueous dispersion of **1o-NP** is exposed to 365 nm light,[§] the characteristic bands in the visible region of the spectrum (centered at 460 nm) corresponding to the ring-closed isomer **1c** appear (Fig. 3a). These bands overlap with those for the ring-closed isomer of the free chromophore in CH_3CN . Due to the scattering of the nano-systems in the UV-vis region of the spectrum, the decrease in the higher-energy bands (250 nm to 300 nm) that are characteristic of the ring-closing reaction of diarylethenes (see the spectra of **1o** and **1c** in Fig. 3a) cannot be observed. In either case, the appearance of the visible bands explains the change in colour of both solutions from colourless to yellow. The spectral changes stop within a minute (30 s for **1o-NP** in water and 60 s for **1o** in



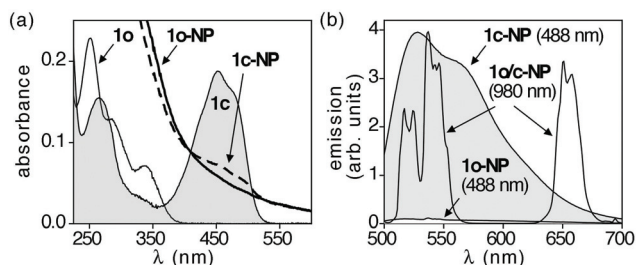


Fig. 3 (a) UV-vis absorption spectra of a CH₃CN solution (1×10^{-5} M) of chromophore **1o** before (white shaded area) and after exposure to 365 nm light (grey shaded area), and an aqueous dispersion (0.8×10^{-6} M)** of the nano-assemblies (**1o**-NP) before (solid line) and after exposure to 365 nm light (broken line). (b) Emission spectra for an aqueous dispersion of the nano-assemblies (0.8×10^{-6} M)** when excited at 488 nm and 980 nm before (**1o**-NP) and after exposure to 365 nm light (**1o**-NP). Because the emission from the UCNP are not affected by the states of the organic chromophores, the two spectra generated with NIR light (**1o**-NP) are shown as superimpositions. The concentrations reported here are concentrations of **1o** in free solution or in encapsulated nano-assemblies.

CH₃CN) at which time the chromophores have reached their photostationary states. While the amount of ring-closed isomer in the photostationary state of the nano-assemblies (**1c**-NP) could not be directly measured, it is safe to assume the behaviour is similar to that of the free chromophores,^{8b,d} which ring-close to generate **1c** quantitatively as measured by ¹H NMR spectroscopy. Based on this assumption, the average loading in **1o**-NP was calculated to be 65 chromophores per nanoparticle, which is in accordance with the number of organic molecules encapsulated in the previous reported nano-assemblies synthesized by similar methods.^{8a,b,d} When either coloured solution (**1c**-NP in water and **1o** in CH₃CN) is exposed to visible light of wavelengths greater than 434 nm,[¶] it reverts back to its colourless state as the ring-open isomers are regenerated. This ring-closing/ring-opening cycle can be repeated without major changes in the spectra (Fig. S6†).

The emission behavior of the nano-assemblies is illustrated in Fig. 3b. When the aqueous dispersion of **1o**-NP is excited with 980 nm light, it generates green emission (also shown in photographs in Fig. S4 and S5†). This emission is attributed solely to the UCNP since neither chromophore **1o** nor **1c** absorbs NIR light, and exposing a CH₃CN solution of **1c** produces no observable fluorescence. As is typical for NaYF₄:ErYb/NaYF₄ nanoparticles, two sets of relatively sharp bands are observed in the region between 500 and 700 nm (Fig. 3b). The higher energy green emission (504–568 nm) correspond to the [²H_{11/2}, ⁴S_{3/2}] → ⁴I_{15/2} transitions, while the lower energy red emission (627–684 nm) are a result of the ⁴F_{9/2} → ⁴I_{15/2} transitions.

When light at 488 nm is used as excitation source, no fluorescence is observed from the aqueous dispersion of **1o**-NP as expected since neither component (**1o** or the UCNP) absorbs at 488 nm (Fig. 3b). After exposure to 365 nm light to trigger the ring-closing reaction (**1o**-NP → **1c**-NP) an intense, broad

emission band between 500 and 700 nm is observed when the sample is excited at 488 nm. This behavior parallels that of the free chromophore in CH₃CN (Fig. S3†). The emission of **1c**-NP when it is excited with 980 nm light was almost identical to that of **1o**-NP (Fig. S3†), indicating the emission from UCNP in the nano-assemblies are not effected by the states of surrounding diarylethene molecules. The emission from the organic chromophores could be turned ‘off’ again by exposing the aqueous dispersion to light at wavelengths greater than 480 nm to regenerate the ring-open isomer (**1o**-NP).

The use of the nano-systems as both conventional and ‘turn-on’ fluorescent probes was demonstrated using *Caenorhabditis elegans* nematodes L4 hermaphrodites, which were incubated for 3 h with **1o**-NP in NaCl buffer.‡ Importantly, the nematodes were still viable after the treatment and incubation. After immobilizing the nematodes with NaN₃ in NaCl buffer, they were washed and isolated by centrifugation, then mounted on agarose gel pads and imaged by using 2-photon fluorescence microscopy. The results are shown in the images in Fig. 4.

As anticipated, no fluorescence was observed when the animals are exposed to 488 nm light. However, the emission resulting from the upconverting process of NIR light by the UCNP in **1o**-NP could be clearly observed in the digestive tract of the nematodes, indicating that they had ingested the nano-particles. Exposing the animals to 365 light for 30 s results in clearly observable emission when excited at 488 nm due to the ring-closing reaction already demonstrated. Once again, the emission from the UCNP was not effected. Importantly, the

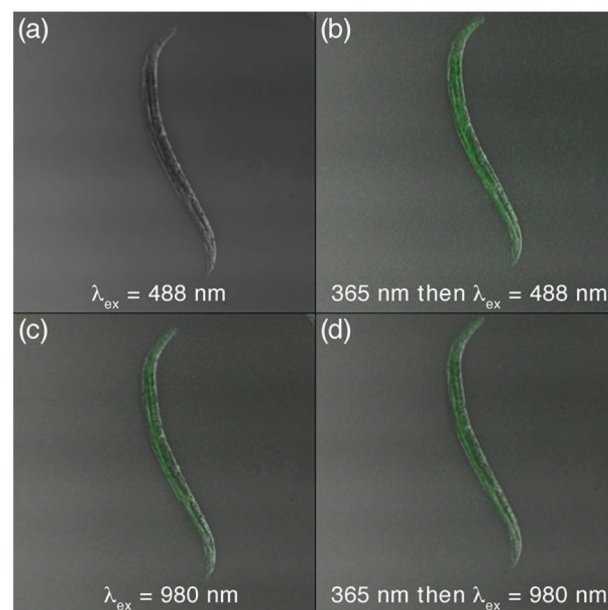


Fig. 4 Fluorescence microscopy images of a representative *C. elegans* when excited with 488 nm light (a) before and (b) after exposure to 365 nm light, and when excited with 980 nm light (c) before and (d) after exposure to 365 nm light. The images are the superimpositions of white light images and fluorescent images.



entire process caused no observable damage to the nematodes, and the nematodes were still mobile before, during, and after the imaging experiments.||

Conclusions

In this article, we show how we have successfully used water-dispersible nano-assemblies containing photoresponsive diarylethene chromophores and upconverting nanoparticles as dual-functioning fluorescent probes for live organism imaging. The NIR-generated emission from the UCNPs can be used as one (static) probe as it does not change. The second emission can be modulated by toggling the photoresponsive compound between its dark (**1o**) and bright (**1c**) state using UV and visible light. Systems such as this not only have the capacity to enhance the quality of images through 'turn-on' mechanism,² they can also reduce false positive signals due to the presence of a second probe.

Acknowledgements

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Notes and references

‡ See ESI† for details.

§ Standard hand-held lamps used for visualizing TLC plates were used to carry out the ring-closing and photolysis reactions at 365 nm (1.4 mW cm⁻²). Irradiation was carried out in low-light conditions to minimize interference from ambient light.

¶ Visible light irradiation was carried out using the light of a 150 W tungsten source that was passed through a 434 nm cutoff filter to eliminate higher-energy light. Irradiation was carried out in low-light conditions to minimize interference from ambient light.

|| The quantity of NaN₃ buffer in the experiment was carefully controlled to assure paralyzing the nematodes to the level where they were immobile enough to take pictures with good resolution and focus while not killing them.

** This value is the total concentration of organic chromophores entrapped within the nano-assemblies obtained using the concentration of the ring-closed isomer (**1c**) at its photostationary state (as measured by UV-vis spectroscopy) divided by percentage of ring-closed isomers in this state (as measured by ¹H NMR spectroscopy). See ESI† for details.

- 1 R. Weissleder and M. J. Pittet, *Nature*, 2008, **452**, 580.
- 2 H. Kobayashi and P. L. Choyke, *Acc. Chem. Res.*, 2011, **44**, 83.
- 3 (a) H. Bo, M. Bates and X. Zhuang, *Annu. Rev. Biochem.*, 2009, **78**, 993; (b) M. Fernandez-Suarez and A. Y. Ting, *Nat. Rev.*, 2008, **9**, 929; (c) F. M. Raymo, *Phys. Chem. Chem. Phys.*, 2013, **15**, 14840.
- 4 (a) X. Chen, T. Pradhan, F. Wang, J. S. Kim and J. Yoon, *Chem. Rev.*, 2012, **112**, 1910; (b) J. Chan, S. C. Dodani and C. J. Chang, *Nat. Chem.*, 2012, **4**, 973; (c) M. E. Jun, B. Roy and K. H. Ahn, *Chem. Commun.*, 2011, **47**, 7583.
- 5 Y. Hori and K. Kikuchi, *Curr. Opin. Chem. Biol.*, 2013, **17**, 644.
- 6 K. Uno, H. Niikura, M. Morimoto, Y. Ishibashi, H. Miyasaka and M. Irie, *J. Am. Chem. Soc.*, 2011, **133**, 13558.
- 7 (a) M. Irie, T. Fukaminato, K. Matsuda and S. Kobatake, *Chem. Rev.*, 2014, **114**, 12174; (b) B. L. Feringa, *Molecular Switches*, Wiley-VCH, Weinheim, 2010; (c) L. Ubaghs, D. Sud and N. R. Branda, *Handbook in Thiophene-Based Materials: Applications in Organic Electronics and Photonics*, ed. I. D. Perepichka and D. Perepichka, John Wiley & Sons, Chichester, 2009, vol. 2; (d) H. Tian and S. Yang, *Chem. Soc. Rev.*, 2004, **33**, 85.
- 8 (a) T. Wu, S. Kaur and N. R. Branda, *Org. Biomol. Chem.*, 2015, **13**, 2017; (b) T. Wu, D. Wilson and N. R. Branda, *Chem. Mater.*, 2014, **26**, 4313; (c) T. Wu, M. Barker, C.-J. Carling, K. M. Arafah, J.-C. Boyer and N. R. Branda, *Angew. Chem., Int. Ed.*, 2013, **52**, 11106; (d) T. Wu, J.-C. Boyer, M. Barker, D. Wilson and N. R. Branda, *Chem. Mater.*, 2013, **25**, 2495.
- 9 (a) M. Nyk, R. Kumar, T. Y. Ohulchanskyy, E. J. Bergey and P. N. Prasad, *Nano Lett.*, 2008, **8**, 3834; (b) M. Haase and H. Schäfer, *Angew. Chem., Int. Ed.*, 2011, **50**, 5808; (c) S. Gai, C. Li, P. Yang and J. Lin, *Chem. Rev.*, 2014, **114**, 2343; (d) Y. Park, K. T. Lee, Y. D. Suh and T. Hyeon, *Chem. Soc. Rev.*, 2015, **44**, 1302.
- 10 J.-H. Lee and J. Cheon, *Anal. Sci. Technol.*, 2011, **2**, A71.

