



Cite this: *Nanoscale*, 2015, 7, 11746

Quenching of the upconversion luminescence of $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ and $\text{NaYF}_4:\text{Yb}^{3+}, \text{Tm}^{3+}$ nanophosphors by water: the role of the sensitizer Yb^{3+} in non-radiative relaxation†

Riikka Arppe,^{*a} Iko Hyppänen,^b Niina Perälä,^a Riikka Peltomaa,^a Martin Kaiser,^c Christian Würth,^c Simon Christ,^{a,d} Ute Resch-Genger,^c Michael Schäferling^{a,c} and Tero Soukka^a

We have studied the mechanisms of water-based quenching of the upconversion photoluminescence of upconverting nanophosphors (UCNPs) via luminescence decay measurements for a better understanding of the non-radiative deactivation pathways responsible for the relatively low upconversion luminescence efficiency in aqueous solutions. This included both upconversion luminescence measurements and the direct excitation of emissive energy states of Er^{3+} and Yb^{3+} dopants in $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ UCNPs by measuring the decays at 550 and 655 nm upon 380 nm excitation and at 980 nm upon 930 nm excitation, respectively. The luminescence intensities and decays were measured from both bare and silanized $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ and $\text{NaYF}_4:\text{Yb}^{3+}, \text{Tm}^{3+}$ UCNPs in H_2O and D_2O . The measurements revealed up to 99.9% quenching of the upconversion photoluminescence intensity of both Er^{3+} and Tm^{3+} doped bare nanophosphors by water. Instead of the multiphonon relaxation of excited energy levels of the activators, the main mechanism of quenching was found to be the multiphonon deactivation of the Yb^{3+} sensitizer ion caused by OH-vibrations on the surface of the nanophosphor. Due to the nonlinear nature of upconversion, the quenching of Yb^{3+} has a higher order effect on the upconversion emission intensity with the efficient Yb–Yb energy migration in the ~35 nm nanocrystals making the whole nanophosphor volume susceptible to surface quenching effects. The study underlines the need of efficient surface passivation for the use of UCNPs as labels in bioanalytical applications performed in aqueous solutions.

Received 1st April 2015,
Accepted 15th June 2015
DOI: 10.1039/c5nr02100f
www.rsc.org/nanoscale

1. Introduction

Upconverting nanophosphors (UCNPs) have increasingly gathered interest in numerous applications. Decreasing the size of

UCNPs to sub-10 nm levels while maintaining the strong luminescence intensity is a subject of intensive research for *e.g.* imaging purposes.^{1,2} However, the upconversion (UC) luminescence efficiency is known to be size-dependent due to decreasing number of emitting ions and increasing surface-to-volume ratio leading to more strongly emerging surface quenching effects.^{3–6} To overcome these limitations, the non-radiative deactivation channels and surface effects must be understood better. There has been discussion about whether the size-dependent luminescence originates from phonon-mediated energy transfer processes,^{4,7} surface effects related to high energy vibrations of ligands, surfactants³ and surrounding solvents⁵ containing OH- and CH-groups, or from increased surface defect density,⁵ or from a combination of these effects.

For the use of UCNPs as reporters in bioanalytical assays and biological environments they must be rendered water dispersible. It is well known that water quenches the luminescence of lanthanides and also lanthanide doped nanocrystals.^{8,9} This is commonly ascribed to high energy vibrational modes of OH-groups (3200–3600 cm^{-1}) which

^aUniversity of Turku, Department of Biotechnology, 20014 Turun yliopisto, Turku, Finland. E-mail: riikka.arppe@utu.fi

^bUniversity of Turku, Department of Chemistry, Laboratory of Materials Chemistry and Chemical Analysis, Vatselankatu 2, 20014 Turun yliopisto, Turku, Finland

^cBAM Federal Institute of Materials Research and Testing, Division 1.10 Biophotonics, Richard-Willstätter-Str. 11, 12489 Berlin, Germany

^dUniversity of Regensburg, Institute of Analytical Chemistry, Chemo- and Biosensors, 93040 Regensburg, Germany

† Electronic supplementary information (ESI) available: Transmission electron micrographs of the bare and silanized UCNPs, X-ray powder diffraction spectra of the $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ and $\text{NaYF}_4:\text{Yb}^{3+}, \text{Tm}^{3+}$ nanophosphors, FT-IR spectra of oleic acid capped and bare UCNPs, the repetitiveness study of luminescence lifetime measurement, decay curves and table of lifetimes of upconversion emission of bare $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ in different proportions of water in H_2O and time points at 550 nm, and upconversion luminescence spectra of bare $\text{NaYF}_4:\text{Yb}^{3+}, \text{Er}^{3+}$ in both D_2O and H_2O at different excitation power densities. See DOI: 10.1039/c5nr02100f



result in an increased non-radiative relaxation of the excited states.⁸ In Er^{3+} doped UCNPs, the OH-groups quench the luminescence of Er^{3+} ions by facilitating multiphonon relaxation of the $^4\text{I}_{11/2} \rightarrow ^4\text{I}_{13/2}$ and $^2\text{H}_{11/2}/^4\text{S}_{3/2} \rightarrow ^4\text{F}_{9/2}$ transitions (Fig. 1). Both relaxation pathways favor the population of the $^4\text{F}_{9/2}$ state from which the red luminescence occurs at the expense of the green luminescence.^{8,10} The quenching mechanism of water has not yet been studied in detail for Tm^{3+} doped nanophosphors.^{6,11} Also Yb^{3+} has been reported to be quenched by water in molecular complexes and chelates.^{12,13}

Wang *et al.* (2010) showed that by covering the $\text{NaGdF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ core with a thin inactive NaGdF_4 shell, the quenching effect of 20 vol% water decreased from 80% to 35% compared with the core material only.⁶ Also, an inactive NaYF_4 shell has been reported to protect the luminescence of the core from surface-related quenching effects.^{14,15} Therefore, water molecules on the surface of the nanophosphor affect only the luminescent lanthanide dopants and the ions comprising the inactive shell or the host crystal are unaffected. The lanthanide ions inside the “core only” nanophosphors and those on the surface should show different emission intensities because the surface dopants are more prone to surface quenching effects.^{5,6} In theory, however, the excitation energy can travel *via* Yb–Yb energy migration from the core regions of an UCNP to the surface dopants where the energy is coupled with the vibrational modes of the environment.^{2,16,17} Gargas *et al.* suggested that the entire volume of their 8 nm nanophosphor was energetically coupled to the surface.² Also Er–Er and Tm–Tm energy transfer may occur but is less probable,

due to the very low concentration of the activators and thus a larger distance between two individual ions.

In addition, the absorption of 980 nm excitation light by water has been suggested to be involved in the decrease of upconversion luminescence intensity.¹⁸ The absorptivity of water at 980 nm is 0.485 cm^{-1} .^{18,19} Thus, the transmission of light in a path length of 1 cm in water is 61.6%. Therefore, the energy of the excitation light is gradually attenuated which further affects the resulting upconversion emission intensity.

The rational design of bright UCNPs for bioanalytical applications performed in aqueous solutions encouraged us to systematically study the influence of water on the upconversion luminescence intensity and decay of $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ and $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ nanophosphors. Furthermore, the effect of water on the intensity and decay of the directly excited (at 380 nm) luminescence of the Er^{3+} emission and on the luminescence decay at 980 nm upon 930 nm excitation originating from the sensitizer Yb^{3+} were studied. Also, comparative luminescence studies were performed in D_2O to identify the luminescence deactivation pathways. In addition, because surface functionalization is required for biomolecule coupling, we studied the effect of a thick silica shell on the luminescence quenching by water.

2. Experimental

Materials

$\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ ($x_{\text{Yb}} = 0.17$, $x_{\text{Er}} = 0.03$) and $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ ($x_{\text{Yb}} = 0.20$, $x_{\text{Tm}} = 0.005$) nanophosphors were synthesized in

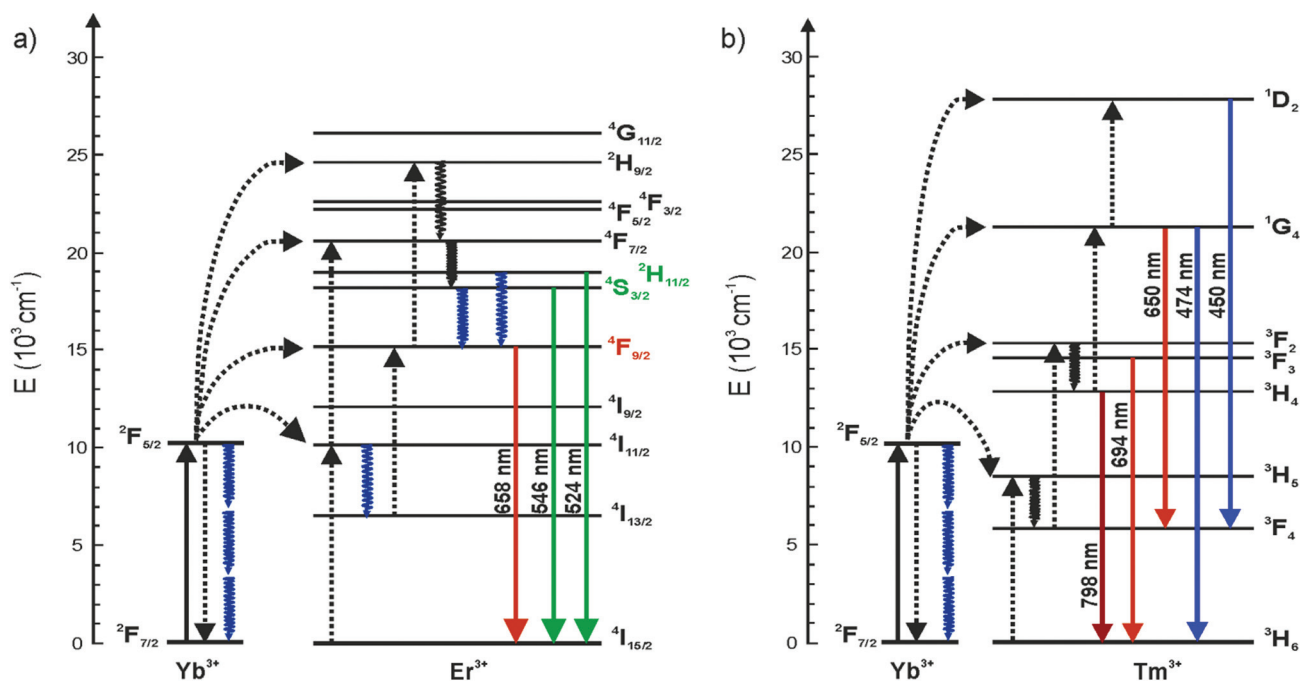


Fig. 1 Energy level diagrams of (a) $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ and (b) $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$. Solid, dotted and wavy arrows represent photon absorption or emission, energy transfer and relaxation processes, respectively. The blue wavy arrows denote the increased multiphonon relaxations caused by OH-vibrations.



octadecene/oleic acid.²⁰ Their size distribution was measured with transmission electron microscopy (TEM) and the crystal structure was studied with X-ray powder diffraction (XPD). The oleic acid (OA) capping was removed using a modified method of Bogdan *et al.*¹⁰: 20 mg of OA-capped UCNPs in toluene were recovered by centrifuging for 30 min, 16 837g (Eppendorf Centrifuge 5418, Hamburg, Germany) and by removing the supernatant carefully by pipetting. The UCNP-pellet was redispersed into 1 ml of 0.1 M HCl (pH 1) by ultrasonication (Finnsonic m03, Finnsonic Oy, Lahti, Finland) and the acid wash was left to perform in rotation overnight. During the acid wash the OA-molecules coordinated to the lanthanides were protonated and thus disengaged from the surface of the UCNP. The UCNPs were precipitated by adding 500 μ l of methanol and 150 μ l of isopropanol and centrifuged as before. The UCNPs were washed once with 1 ml of toluene to remove the residue OA and then the bare UCNPs were dispersed to 100 μ l of ethanol.

Both Er³⁺- and Tm³⁺-doped UCNPs were also surface-modified by a silanization with 60 μ l of tetraethylorthosilicate (TEOS) using a reverse-microemulsion technique by Wilhelm *et al.*²¹ after which the UCNPs with silica shell were dispersed into 200 μ l of ethanol. No functional groups were added to the silica.

Both bare and silanized UCNP-dispersions were divided into two and the ethanol was evaporated with MiVac centrifugal concentrator (Genevac Ltd Suffolk, UK). Then, the pellets were heated to 80 °C for overnight to remove all coordinated water and thereafter, dispersed into 500 μ l of either ultrapure mQ-H₂O or D₂O (99.9% purity, Sigma-Aldrich, St. Louis, MO) and stored in rotation at room temperature. The absence of OA in the bare UCNPs was evaluated with FT-IR (Vertex 70, Bruker Optics, Billerica, MA). As a reference, the IR-spectrum of OA-capped UCNP was measured.

Luminescence lifetime measurements in the time domain

Luminescence lifetimes in the time domain were measured using three instruments: a modular luminometer with pulsed excitation (method 1), a modified multimode microtiter plate reader (method 2), and a commercial fluorometer equipped with a pulsed μ -flash xenon lamp and a phosphorescence accessory (method 3).

In method 1, an analog-to-digital converter NI USB-6251 (National Instruments, Austin, TX) was used to generate square-wave excitation pulse profile for the laser driver WDL3343 (Wavelength Electronics, Inc., Bozeman, MT). The excitation source was a NIR laser diode L9418-04 (Hamamatsu Photonics, Hamamatsu City, Japan) providing max. 1.2 W at 976 nm. The optical part of the luminometer consisted of tubular excitation and emission chambers (Thorlabs, Inc., Newton, NJ) in a right-angle configuration. The sample chamber was an aluminum cube equipped with a Peltier element for thermoelectric cooling/heating and a 5 mm quartz NMR tube for liquid samples. In the excitation light path, a RG850 long-pass filter with a cutoff at 850 nm (Edmund Optics, Barrington, NJ) was used to ensure a pure NIR excitation. In the emission light path, an extended hot mirror filter (Edmund Optics) with a good transmission at visible

wavelengths was used to exclude the scattered excitation radiation. A long-pass filter with a cutoff at 750 nm (Newport Spectra-Physics GmbH, Germany) was used to select the 800 nm emission of thulium. Other emission wavelengths were selected with interference filters with a suitable band-pass, 544/10 nm, 650/10 nm (Thorlabs), 470/20 nm (Chroma Technology Corp., Bellows Falls, VT). The detector at the end of emission chamber was a head-on R1464 photomultiplier (Hamamatsu). The photomultiplier signal was amplified in a high-speed current amplifier DHPA-100 (Femto Messtechnik GmbH, Germany). The amplified signal was recorded with the NI USB-6251 A/D converter, which was connected to a computer *via* USB and controlled with a computer program written in LabVIEW 8.5 (National Instruments). The pulse profile consisted of a 20 ms excitation pulse (1.2 W) followed by a 30 ms delay period. During a single measurement the pulse profile was cycled 10 000 times resulting a measurement time of 8.5 minutes. Same settings and measurement times were used for all samples, and thus, there was no need for normalization.

In method 2, a modified Plate Chameleon fluorometer (Hidex Oy, Turku, Finland) equipped with a 980 nm laser diode as excitation source was used.²² The sample in 100 μ l volume in clear Greiner polypropylene microtiter plate (Sigma-Aldrich) was repeatedly exposed to 2 ms excitation light pulses. The upconversion emission wavelengths were selected by optical band-pass filters 535/40 nm, 665/70 nm, 470/20 nm (Chroma Technology Corp.) or 795/5 nm (Edmund Optics). Neutral density filters were used to damp the obtained signals, when necessary. Same settings and measurement times were used for all samples, and thus, there was no need for normalization.

In method 3, the luminescence decay measurements were carried out with a commercial Edinburgh Instruments spectrofluorometer FSP-920 equipped with a μ -flash lamp 920H (pulse width 3 μ s) and a red extended PMT (R2658P) from Hamamatsu. The Yb³⁺ luminescence band ²F_{5/2} → ²F_{7/2} was excited with 930 nm excitation and detected at 980 nm (slit width was set to 20 nm optical band width for both excitation and detection). The green ⁴S_{3/2} → ⁴I_{15/2} and red ⁴F_{9/2} → ⁴I_{15/2} Er³⁺ luminescence bands were excited at 378 nm and detected at 550/655 nm (slit width was set to 4 nm and 8 nm optical band width for excitation and detection, respectively), and a 400 nm long-pass was used to avoid detection of scattered excitation light. The decays of the directly excited luminescence of Er³⁺ dopants were normalized in respect to the highest obtained signal. The decays of directly excited luminescence of Yb³⁺ dopants were normalized to correspond a measurement time of 60 min.

For the method 1, an UCNP-concentration of 1 mg mL⁻¹ was used. For the method 2, the concentration was 40–50 μ g mL⁻¹ and for the method 3, 40 mg mL⁻¹ UCNP-concentration was used due to the low signal levels of the directly excited luminescence. The luminescence profiles were analyzed with the exponential decay fitting with Origin 8.0 (Originlab Corporation, Northampton, MA). Fittings of the decay profiles of the upconversion luminescence were started 50–200 μ s after the excitation pulse was turned off.



Luminescence intensity and spectral measurements

The modified Plate Chameleon fluorometer was also used for measurement of upconversion photoluminescence intensity. The measurements were done at UCNP-concentration of 40 $\mu\text{g mL}^{-1}$ or 2.2 $\mu\text{g mL}^{-1}$ in 100 μl volume from clear polypropylene 96-well microtiter plates. A 2 s excitation pulse was used and the emission was measured simultaneously. A standard Plate Chameleon fluorometer with Xenon flash lamp as UV-light source (Hidex Oy) was used to measure the directly excited (at 380 nm) luminescence emission intensities of the $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ nanophosphors at 660 nm using 665/45 nm (Chroma Technology Corp.) and 365/50 nm (bk Interferenzoptik Elektronik GmbH, Nabburg, Germany) optical filters, respectively, with a 80 μs delay and a 420 μs measurement window.

Upconversion emission spectra were measured at a UCNP-concentration of 1 mg mL^{-1} with a PC2000-CCD optical fiber spectrometer (Ocean Optics, Inc., Dunedin, FL). A laser diode driver 5060 (Newport, Irvine, CA) was used to control a fiber-coupled NIR laser diode IFC-975-008 (Optical Fiber Systems, Inc., Chelmsford, MA) providing 6 W at 973 nm. The optical part of the system was the same tubular chamber setup used in the lifetime measurement method 1, but without interference filters in the emission site. At the end of the emission chamber was an optical fiber connected to the PC2000 spectrometer. The OOIrrad software was used to record the upconversion emission spectra. The spectral response of the system was calibrated with a tungsten halogen light source LS-1-CAL (Ocean Optics Inc.). The laser beam diameter was estimated to be ~ 3 mm when focused on samples.

3. Results and discussion

The UCNPs were characterized with TEM, XPD and FT-IR. The $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ were rod-shaped with dimensions of 31–38 nm, and the $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ nanophosphors were sphere-shaped and 29–32 nm in diameter (Fig. S1 in the ESI†). Their crystal structure was pure hexagonal according to the XPD-analysis (Fig. S2a in the ESI†). The FT-IR spectra were measured from oleic acid-capped and bare UCNPs to investigate the surface modification (Fig. S2b in the ESI†). The OA-capped UCNPs exhibited similar stretches reported by Cao *et al.*²³ for OA-capped UCNPs: a multipeak band at ~ 2925 and 2852 cm^{-1} , which were assigned to the asymmetric and symmetric stretching vibrations of methylene (CH_2) in the long alkyne chain of OA, and stretches at 1558, 1446 and 1421 cm^{-1} , which were assigned to the asymmetric and symmetric stretching vibrations of the carboxylic group of the bound OA ligand. These vibrations were absent in the bare UCNPs, indicating the total removal of OA from the UCNP surface. The 7–10 nm silica shell was visualized with TEM (Fig. S1 in the ESI†). The bare and silanized UCNPs were used for the study of quenching mechanism of water.

It has been reported that OH-groups quench the luminescence of $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ UCNPs by facilitating multiphonon relaxation of ${}^2\text{H}_{11/2}/{}^4\text{S}_{3/2} \rightarrow {}^4\text{F}_{9/2}$ and ${}^4\text{I}_{11/2} \rightarrow {}^4\text{I}_{13/2}$ transitions

of Er^{3+} -ions. Both of these relaxation pathways favor the population of the ${}^4\text{F}_{9/2}$ state (either directly or *via* ${}^4\text{I}_{13/2} \rightarrow {}^4\text{F}_{9/2}$ transition, respectively), from which the red luminescence occurs at the expense of green luminescence.^{8,10} Thus, the red-to-green luminescence ratio increases in the presence of water. This effect of the OH-vibrations was also confirmed here by measurement of upconversion luminescence spectra of both bare, *i.e.* without any ligand capping which was removed by acid washing,¹⁰ and silanized $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ nanophosphors in both D_2O and H_2O (Fig. 2). The red-to-green ratio increased from 1.6 in D_2O to 2.4 in H_2O with bare particles, and from 1.8 to 2.2 with silanized particles, respectively. The quenching efficiency of water, calculated using eqn (1), for the green upconversion emission of bare and silanized Er^{3+} -doped particles by water was 86% and 85%, respectively.

$$\eta(\text{quenching}) = 1 - \frac{I_Q}{I} \quad (1)$$

here I_Q is the upconversion luminescence intensity in the presence of the quencher, *i.e.* water, and I is the upconversion luminescence intensity in the absence of the quencher, *i.e.* in D_2O . However, also the red emission was quenched by water up to 79% and 82%, respectively.

The upconversion photoluminescence decays of both bare and silanized Er^{3+} -doped nanophosphors in D_2O and H_2O were studied using method 1 (see Experimental). The resulting decay curves are illustrated in Fig. 3 and the lifetimes with amplitude fractions are listed in Table 1. The repetitiveness of the lifetime measurements was confirmed by measuring the upconversion luminescence decay of the same sample twice using method 1 (Fig. S3 in the ESI†). Similar intensity quenching to the spectral measurements is observed also in the intensity levels of the decay curves in water when compared with the intensity in D_2O . The reduction in the luminescence intensity is reflected by the decay behavior, *i.e.*, major changes in the lifetimes of the different emission peaks as well as in the respective decay amplitudes were observed when the two solvents were compared. In the presence of water the amplitude fraction of the shorter decay was reduced from that in D_2O and the lifetime τ_1 of green emission was shortened by 54%. The τ_1 of the red emission was shortened by only 11%. Thus, the luminescence with the shorter decay probably originates from the surface of the nanophosphors which is most affected by the water molecules. However, also the longer lifetime τ_2 at 544 nm was shortened by 25% by water, and a reduction of up to 41% was observed at 650 nm. Thus, it seems that most of the particle volume can be affected by surface quenchers.

The upconversion decay time of the green emitting state ${}^4\text{S}_{3/2} \rightarrow {}^4\text{I}_{15/2}$ was shortened more by water than that of the red emitting state ${}^4\text{F}_{9/2} \rightarrow {}^4\text{I}_{15/2}$, which can be explained by the increased multiphonon relaxation ${}^4\text{S}_{3/2} \rightarrow {}^4\text{F}_{9/2}$. When an intermediate energy level is deactivated *via* OH-vibration, the emissive states become less populated, which leads to intensity quenching of upconversion emission. However, this should not affect the decay of the populated emissive states. Thus, the 54% decrease in the green luminescence lifetime originates



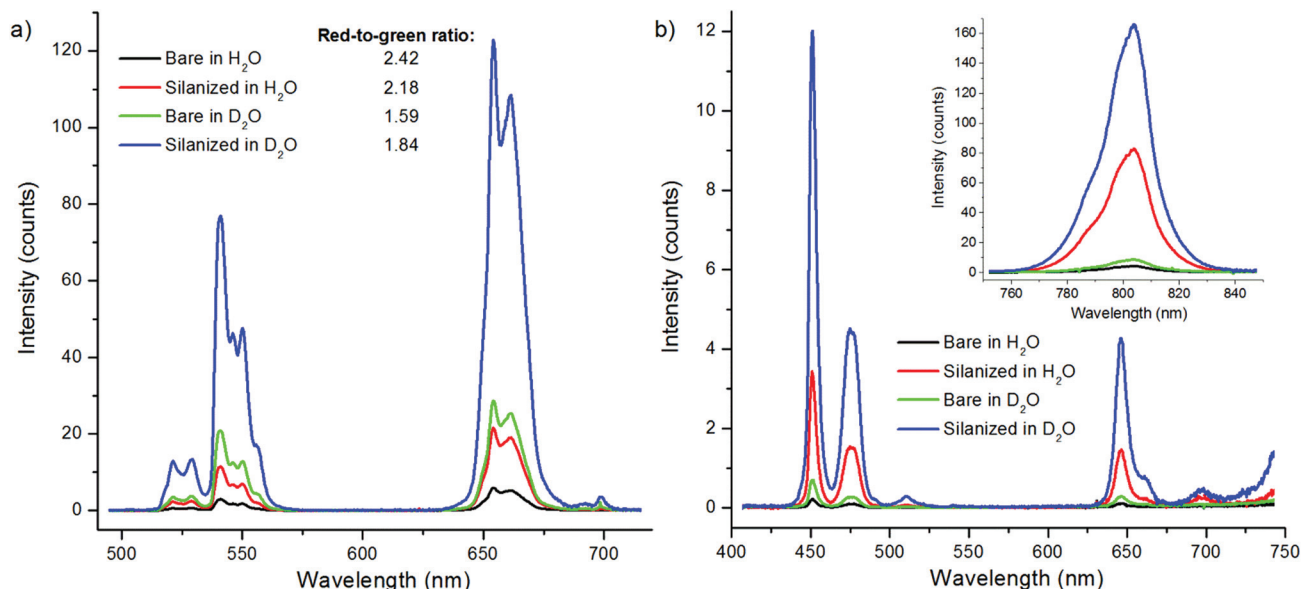


Fig. 2 Upconversion luminescence spectra of both bare and silanized (a) $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ and (b) $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ nanophosphors in H_2O and D_2O .

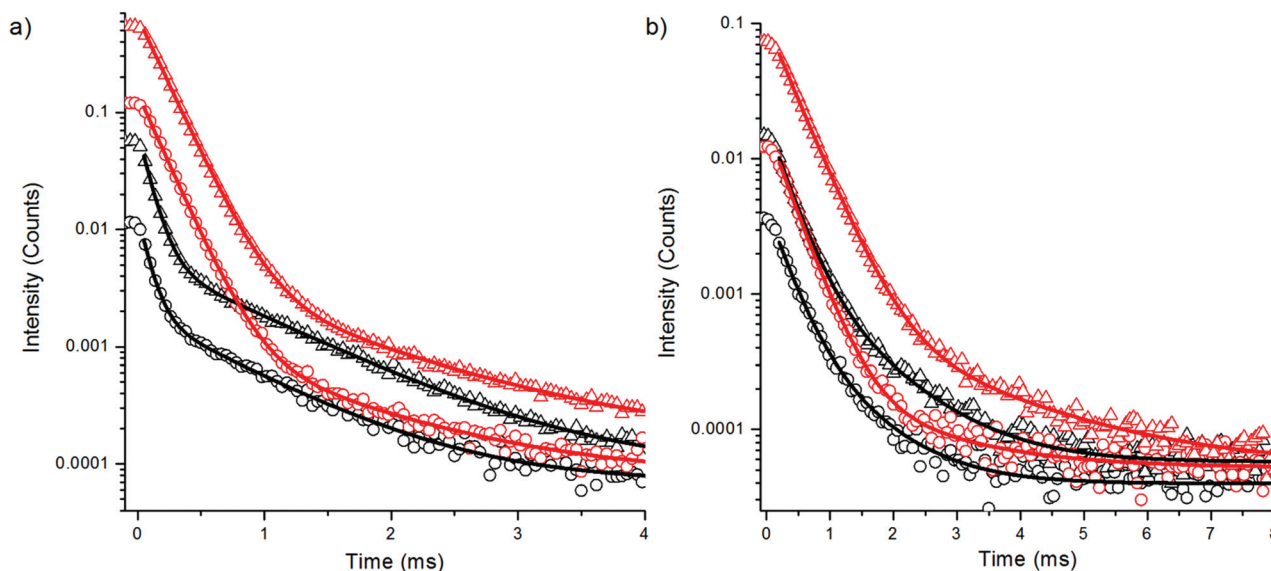


Fig. 3 Decay curves of upconversion emission of bare (circles) and silanized (triangles) $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ in H_2O (black) and D_2O (red) (a) at 544 nm and (b) at 650 nm upon 980 nm excitation with 20 ms pulse width.

Table 1 Decay times and amplitude fractions of green and red upconversion emission transitions of $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$

	Bare in H_2O	Bare in D_2O	Silanized in H_2O	Silanized in D_2O
$^4\text{S}_{3/2} \rightarrow ^4\text{I}_{15/2}$ (green emission)				
τ_1 (μs)	81 ± 1.5 (86.2%)	177 ± 0.5 (99.1%)	92 ± 0.5 (91.8%)	185 ± 0.4 (99.2%)
τ_2 (μs)	755 ± 10.8 (13.8%)	1003 ± 55.0 (0.9%)	843 ± 5.6 (8.2%)	1070 ± 55 (0.8%)
$^4\text{F}_{9/2} \rightarrow ^4\text{I}_{15/2}$ (red emission)				
τ_1 (μs)	296 ± 10.8 (85.4%)	334 ± 2.5 (98.4%)	300 ± 3.8 (90.7%)	383 ± 0.9 (99.0%)
τ_2 (μs)	849 ± 47.3 (14.6%)	1440 ± 129.3 (1.6%)	966 ± 24.4 (9.3%)	1862 ± 72.3 (1.0%)



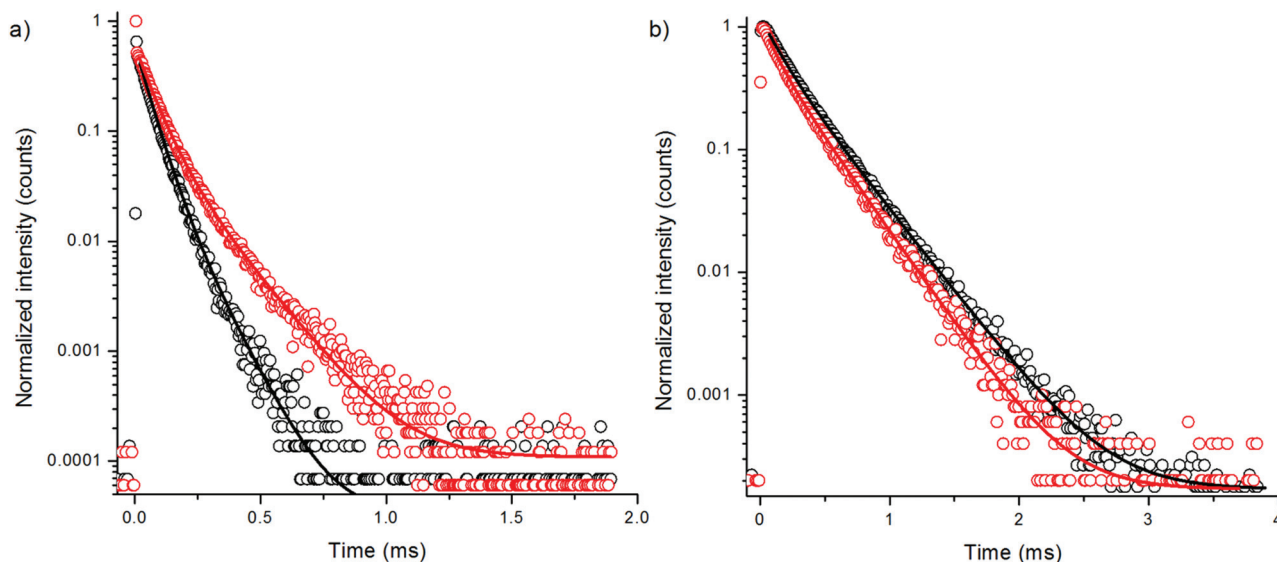


Fig. 4 Decay curves of 380 nm excited emission of bare NaYF₄:Yb³⁺,Er³⁺ (a) at 550 nm and (b) at 655 nm in H₂O (black) and in D₂O (red) using 3 μs excitation pulse width.

only from the increased multiphonon relaxation $^4S_{3/2} \rightarrow ^4F_{9/2}$ and the deactivation of $^4I_{11/2} \rightarrow ^4I_{13/2}$ has no effect on the lifetime of the green emission. The decay time of red emitting state $^4F_{9/2} \rightarrow ^4I_{15/2}$ was less affected by water and thus cannot explain the major intensity quenching of the red upconversion luminescence. The pH of the solvent had no effect on the upconversion photoluminescence intensities or lifetimes of bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors in the range of 3–7 (data not shown).

Surface modification of the nanophosphors *via* silanization provided little protection against the water. The lifetime reduction of the luminescence of silanized nanophosphors in H₂O was similar to the bare particles when compared to the luminescence decays of silanized nanophosphors in D₂O. The amplitude fraction of the main decay component increased in silanized nanophosphors in H₂O, when compared to the corresponding fraction of bare particles. Thus, silica offered partial shielding against OH-vibrations but not enough to prevent the intensity quenching completely. The silica shell might be somewhat porous allowing the water molecules to reach the UCNP surface.

To further study the effect of OH-vibrations on the non-radiative relaxation of the excited states of bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors, we measured the luminescence decays of both $^4S_{3/2} \rightarrow ^4I_{15/2}$ and $^4F_{9/2} \rightarrow ^4I_{15/2}$ transitions by direct excitation with 380 nm radiation using method 3. Excitation populates the higher energy state $^4G_{11/2}$ of Er³⁺ and the subsequent relaxations the emissive states. Thus, the measurement excluded all effects of lower-lying energy level relaxations. The decay curves revealed that the $^4S_{3/2}$ state is indeed quenched by water *via* increased multiphonon relaxation $^4S_{3/2} \rightarrow ^4F_{9/2}$ resulting in attenuated green emission, but the red emission state is not significantly affected by water (Fig. 4, Table 2).

Table 2 Luminescence decay times and amplitude fractions of green and red emissive transitions of NaYF₄:Yb³⁺,Er³⁺ upon 380 nm excitation

	Bare in H ₂ O	Bare in D ₂ O
$^4S_{3/2} \rightarrow ^4I_{15/2}$ (green emission)		
τ_1 (μs)	51 ± 1.1 (86.3%)	63 ± 0.7 (80.4%)
τ_2 (μs)	102 ± 9.1 (13.7%)	155 ± 3.9 (19.6%)
$^4F_{9/2} \rightarrow ^4I_{15/2}$ (red emission)		
τ_1 (μs)	145 ± 2.6 (50.8%)	114 ± 4.2 (51.8%)
τ_2 (μs)	324 ± 2.2 (49.2%)	288 ± 3.1 (48.2%)

Thus, there is no direct deactivation of the $^4F_{9/2}$ state due to OH-vibration. The lifetime of the red luminescence in H₂O was longer than in D₂O, which is probably caused by the increased multiphonon relaxation of $^4S_{3/2} \rightarrow ^4F_{9/2}$ transition in the presence of high energy OH-vibrations, which favors the population of the $^4F_{9/2}$ state in water. The amplitude fraction of the main decay component of green luminescence did not change in the two solvents. Interestingly, the red luminescence contained two lifetimes with about equal amplitude fractions.

The quenching of Tm³⁺-doped nanophosphors by water has been studied less than the quenching of Er³⁺-doped nanophosphors. In the energy level diagram of NaYF₄:Yb³⁺,Tm³⁺ (Fig. 1b), the vibrational relaxations occur only in intermediate energy levels and the energy difference of these levels are too small to be caused by OH-vibrations. However, the intensity of the upconversion luminescence is quenched as much as that of Er³⁺-doped particles (Fig. 2b). The quenching efficiency, calculated from the spectra, of 470 nm upconversion emission of bare and silanized Tm³⁺-doped particles by water was 61% and 66%, respectively. The red emission at 650 nm was quenched 57% and 66%, respectively, and the near infrared (NIR) emis-



sion was quenched 50% in both bare and silanized nanophosphors. Interestingly, silanization seemed to protect the luminescence from quenching more efficiently than in Er^{3+} -doped nanophosphors.

The upconversion luminescence decays of $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ nanophosphors were measured at 470 nm, 650 nm and 800 nm with method 1 (Fig. 5, Table 3). The luminescence decay times of the bare nanophosphors did not differ significantly from the decay times of silanized UCNPs even though the intensity reduction was as large in the decay measurements as in spectral measurement (Fig. 2b). Only 5–25% decrease in lifetimes was found in water compared to D_2O . This suggests that no direct relaxations occur from the emissive states $^1\text{G}_4$ and $^3\text{H}_4$ by the influence of OH-vibration. The 470 nm and 650 nm emissions occur from the same energy

level $^1\text{G}_4$ which can also be seen in the similar decays obtained.

The upconversion photoluminescence intensities of both bare and silanized UCNPs were measured at increasing proportion of H_2O in D_2O and compared to the intensity measured at 100 vol% D_2O . Fig. 6 illustrates the quenching efficiency of water on upconversion luminescence intensity of $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ at both 535 nm and 665 nm. The luminescence of bare particles at both green and red emission wavelengths was quenched up to 99.9%. At 80 vol% H_2O the luminescence intensities were already below 1% when compared to the intensity in D_2O . Similar results have been obtained by Zhang *et al.*²⁴ With silanized particles, the water quenching was reduced to 65% for the green upconversion emission and 55% for the red emission. Therefore, the silica

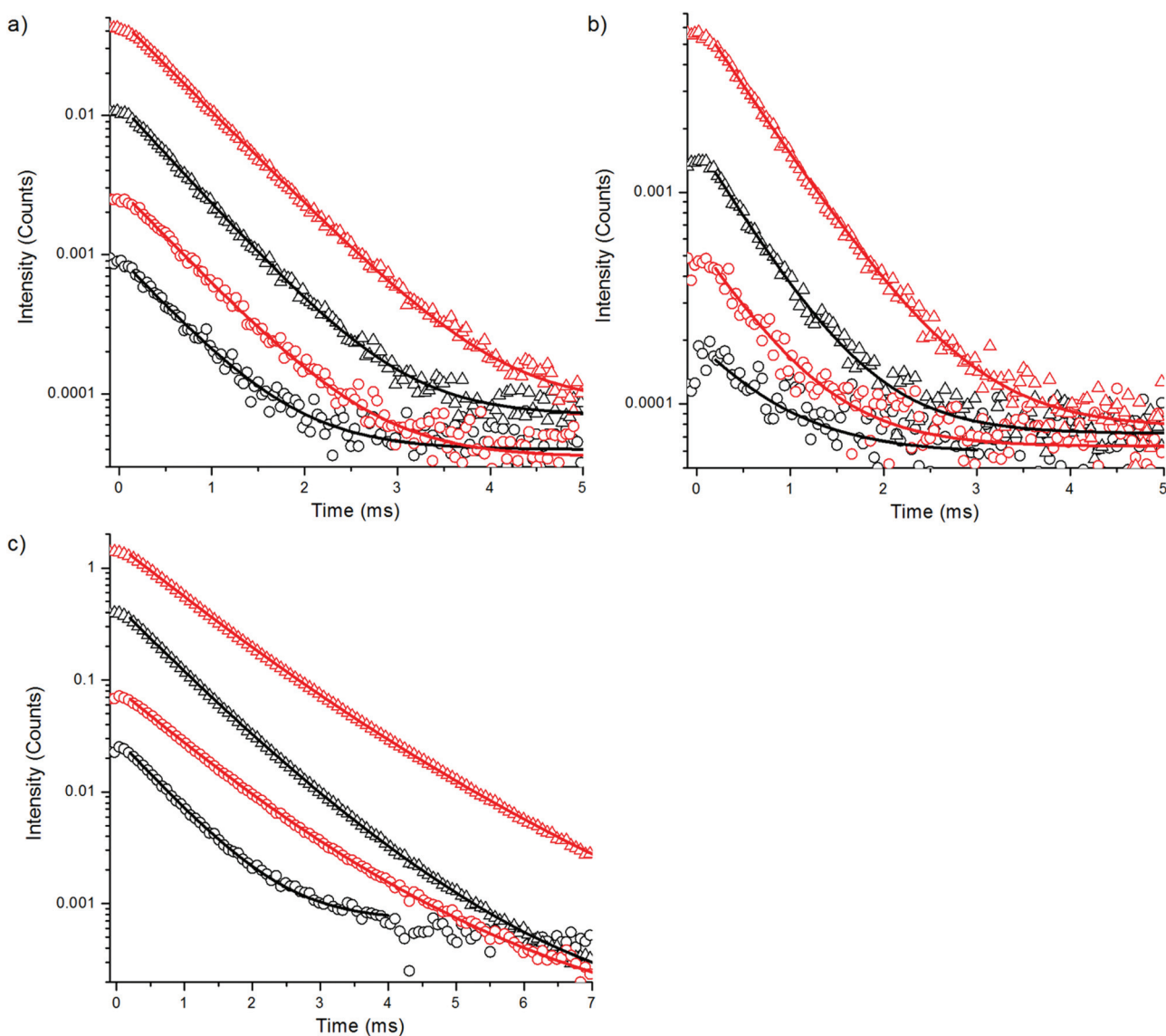
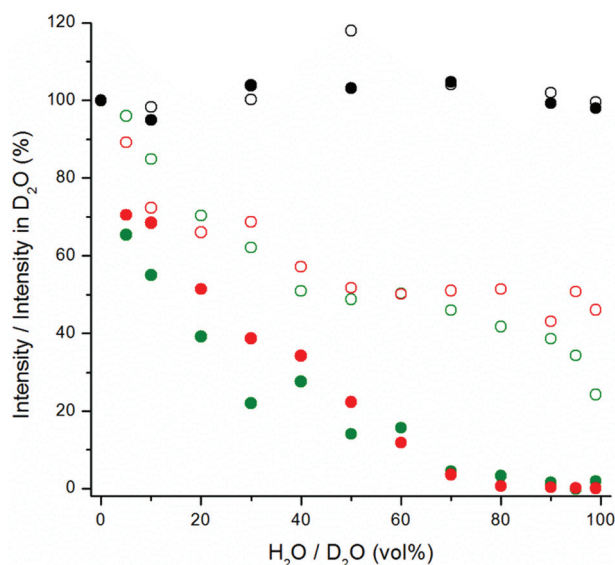
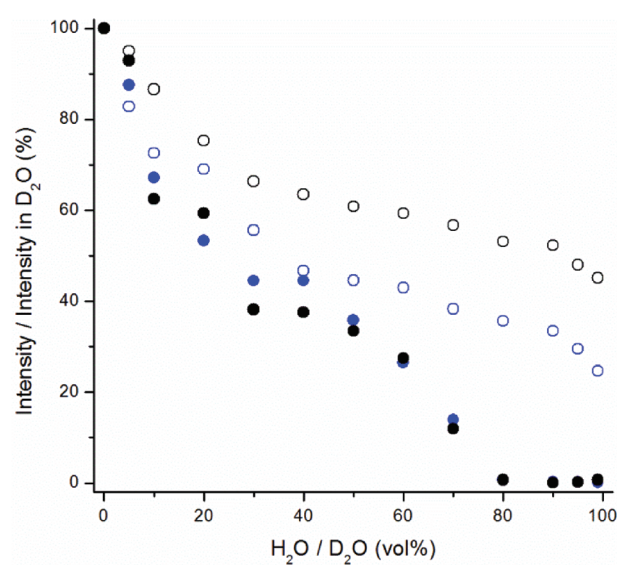


Fig. 5 Decay curves of upconversion emission of bare (circles) and silanized (triangles) $\text{NaYF}_4:\text{Yb}^{3+},\text{Tm}^{3+}$ in H_2O (black) and D_2O (red) at (a) 470 nm, (b) at 650 nm and (c) at 800 nm using 20 ms excitation pulse width.



Table 3 Decay times and amplitude fractions of blue, red and NIR upconversion emission transitions of NaYF₄:Yb³⁺,Tm³⁺

	Bare in H ₂ O	Bare in D ₂ O	Silanized in H ₂ O	Silanized in D ₂ O
¹ G ₄ → ³ H ₆ (blue emission)				
τ ₁ (μs)	593 ± 10.9 (100%)	625 ± 3.9 (100%)	596 ± 1.9 (100%)	650 ± 1.0 (100%)
¹ G ₄ → ³ F ₄ (red emission)				
τ ₁ (μs)	708 ± 79.4 (100%)	618 ± 18.0 (100%)	590 ± 7.0 (100%)	656 ± 2.7 (100%)
³ H ₄ → ³ H ₆ (NIR emission)				
τ ₁ (μs)	666 ± 2.5 (100%)	808 ± 18.6 (86.2%)	668 ± 4.7 (84.8%)	859 ± 4.5 (86.9%)
τ ₂ (μs)		1592 ± 137.0 (13.8%)	1139 ± 22.4 (15.2%)	1526 ± 26.3 (13.1%)

**Fig. 6** The quenching effect of increasing proportion of water in D₂O on upconversion emission at 535 nm (green) and at 665 nm (red), and on 380 nm excited luminescence at 665 nm (black), of bare (filled symbols) and silanized (open symbols) NaYF₄:Yb³⁺,Er³⁺ nanophosphors.**Fig. 7** The quenching effect of increasing proportion of water in D₂O on upconversion emission at 470 nm (blue) and at 800 nm (black) of bare (filled symbols) and silanized (open symbols) NaYF₄:Yb³⁺,Tm³⁺ nanophosphors.

shell provides some protection against the water quenching but it does not offer complete passivation of the UCNPs surface. Similar results were obtained also with NaYF₄:Yb³⁺,Tm³⁺ particles and at both 470 and 800 nm emission wavelengths (Fig. 7). The luminescence of bare Tm-doped nanophosphors was also quenched by up to 99.9% in water. When 380 nm light was used to excite the NaYF₄:Yb³⁺,Er³⁺ nanophosphors directly, the emission intensity at 665 nm remained constant independent of the proportion of H₂O in D₂O (Fig. 6).

The quenching efficiency of water in this experimental setup was much higher than in the spectral measurements above. The differences between the measurement setup using the plate Chameleon and the spectrometer are UCNPs-concentration (40 μg mL⁻¹ vs. 1 mg mL⁻¹, respectively), measurement time (2 seconds per well in microtiter plate assay vs. 10 s–2 min in spectral measurement) and excitation power density (>40 mW cm⁻² and 85 W cm⁻², respectively). The path lengths of the excitation light in the two setups were approximately the same (4.2–4.6 mm). The effects of these parameters on the quenching efficiency were further studied.

The water quenching long-time kinetics was studied using different UCNPs concentrations and proportions of water in D₂O (Fig. 8). The bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors in D₂O were diluted to different vol% of H₂O in D₂O and their upconversion luminescence at 535 nm was measured in 1 min intervals using the plate Chameleon. The luminescence intensities were compared with the intensities obtained from samples in D₂O at each time point. In 99 vol% H₂O, the initial quenching efficiency of water was 75%, which, however, slowly increased to 98.4%. In 75 vol% H₂O the quenching efficiency increased from 71% to 88% during the 40 minutes. The decrease in luminescence intensities can partly be caused by the adsorption of the nanophosphors onto the walls of the microtiter plate, but this should be the same in water and in D₂O and thus cannot explain the faster signal drop in water. Therefore, it seems that the D₂O molecules coordinated on the surface of the nanophosphors are slowly exchanged by H₂O molecules. In addition, a UCNPs-concentration dependence on the quenching rate of water was found (Fig. 8b). When UCNPs-concentration was decreased from 40 μg mL⁻¹ to 2.2 μg mL⁻¹,



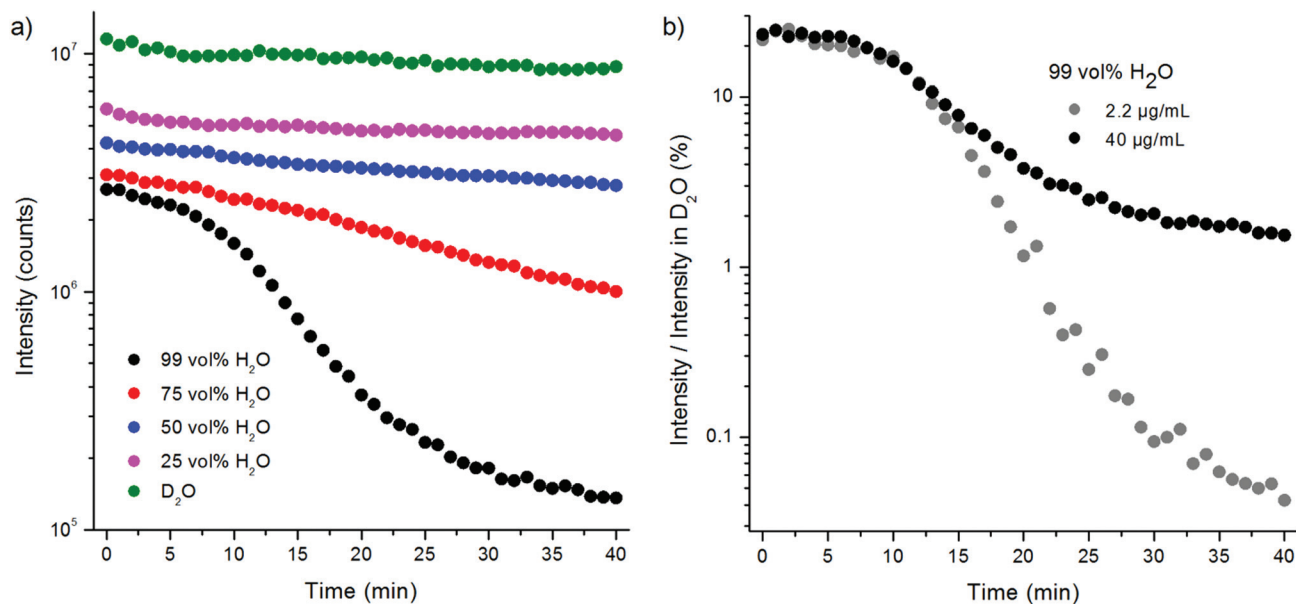


Fig. 8 (a) Kinetics of water quenching of upconversion photoluminescence of bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors at 535 nm after diluting the UCNPs in D₂O to increasing proportions of water (UCNP-concentration 40 μg mL⁻¹). (b) Concentration dependence of the kinetics of water quenching of upconversion photoluminescence of bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors at 535 nm after diluting to 99 vol% H₂O medium at two UCNPs-concentrations.

the quenching efficiency in 99 vol% water increased from the initial efficiency of 78% to 99.96% after 40 minutes. *I.e.*, as the UCNPs-concentration decreased 18-fold, the luminescence intensity at 40 min time point decreased 321-fold. Therefore, the UCNPs-concentration dependence can account for the differences in quenching efficiencies of water found in the two experimental setups. The concentration dependence in the quenching efficiency of water is most likely due to nanophosphor aggregation. Ultrasonication of the bare UCNPs in 99 vol% water led to an increase in the quenching efficiency in comparison to the unsonicated sample due to the increase in surface-to-volume ratio by disruption of the aggregates. Nanophosphor aggregation, which reduces the surface-to-volume ratio, is enhanced in higher UCNPs-concentrations.

The slow decrease in upconversion luminescence was confirmed to have been caused by quenching instead of *e.g.* slow adsorption of the upconverting nanophosphors onto the walls of the microtiter plate wells by measuring the luminescence decays at different time points after preparing fresh samples. The decays were measured from samples in 99 and 75 vol% H₂O in D₂O and in 100 vol% D₂O at time points 1–6 min, 10–16 min and 25–31 min (one measurement lasted for 5 minutes) using method 2 (Fig. S4, Table S1 in the ESI†). In D₂O, the lifetimes remained constant at the three time points. In 99 vol% H₂O, however, there was a 29% decrease in the main decay component measured at 25–31 min when compared to the decay measured at 1–6 min. In 75 vol% H₂O, the corresponding decrease was 20%. Also, the intensity decrease could be observed from the decay curves in H₂O samples whereas in D₂O, the intensity levels remained the same.

The effect of excitation power density on the quenching efficiency of water was studied by measuring the upconversion luminescence spectrum of bare NaYF₄:Yb³⁺,Er³⁺ in both water and D₂O using different excitation powers from 57 to 85 W cm⁻² (Fig. 9, Fig. S5 in the ESI†). Surprisingly, the quenching efficiency of water increased in decreasing power density, *i.e.*, a larger proportion of the luminescence was quenched in water compared to the luminescence in D₂O when lower power densities were used. An enhanced dependence on excitation power density was found for the quenching of the red emission compared to the green. However, the relative quenching of the green emission peaks was stronger. The results suggest that at higher excitation power densities the surface dopants are probably quenched as much as at lower densities, but the core of the UCNPs is able to produce more emission rather than deactivate through relaxations. At higher power densities, a larger population of Yb³⁺ ions is excited and thus there are probably fewer unexcited Yb³⁺ ions for the energy migration to occur, leading to the decreased overall quenching. This study shows that the quenching effect of water on upconversion photoluminescence is inversely dependent on the excitation power density, and the dependency is linear at the power range studied here. In the future, the power density range should be increased to study the luminescence behavior also at higher powers.

The integrated intensities of spectra in Fig. S5 (in the ESI†) were plotted against the excitation power densities in a log–log curve (Fig. 9b). The slopes of the power density dependency curves in D₂O for the green emission transitions ²H_{11/2}/⁴S_{3/2} → ⁴I_{15/2} and the red emitting state ⁴F_{9/2} → ⁴I_{15/2} were 2.1 and 2.6,



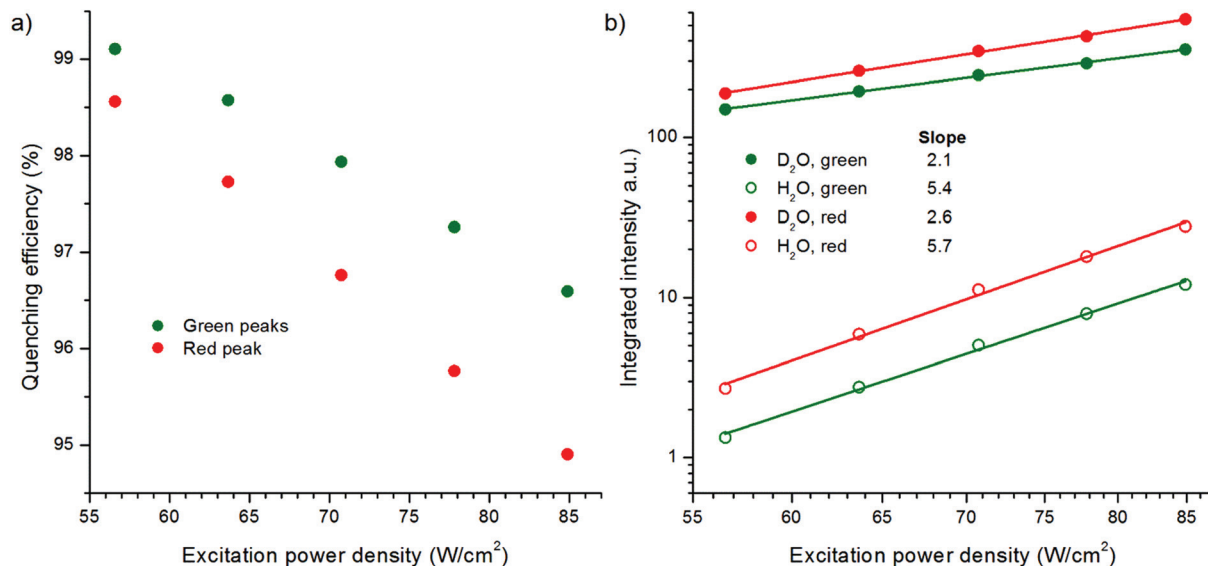


Fig. 9 (a) The quenching efficiency of upconversion photoluminescence of bare NaYF₄:Yb³⁺,Er³⁺ nanophosphors by water at different excitation power densities, and (b) intensities of the red (636–684 nm) and green (510–565 nm) upconversion emission peaks in D₂O or in H₂O plotted together with the excitation power density in a log–log curve.

respectively, which are similar with those reported in the literature.¹⁴ However, in the presence of the high energy OH-vibrations in water, the slopes increased to 5.4 and 5.7, respectively. To our knowledge, this is the first time that the power dependency of upconversion photoluminescence has been measured in water. Usually, the power density dependencies are studied from a dry UCNP powder or the sample environment is not even mentioned.^{14,25,26}

The absorptivity of water at 980 nm, which is 0.485 cm^{-1} ,^{18,19} has been suggested to be one of the quenching mechanisms of upconversion photoluminescence.¹⁸ In the experimental setups with 4.2–4.6 mm path length, ~20% of the excitation light was absorbed. The absorption of the excitation light by water should have a similar effect on the upconversion luminescence intensity as the decrease of excitation power density. Because the excitation power depen-

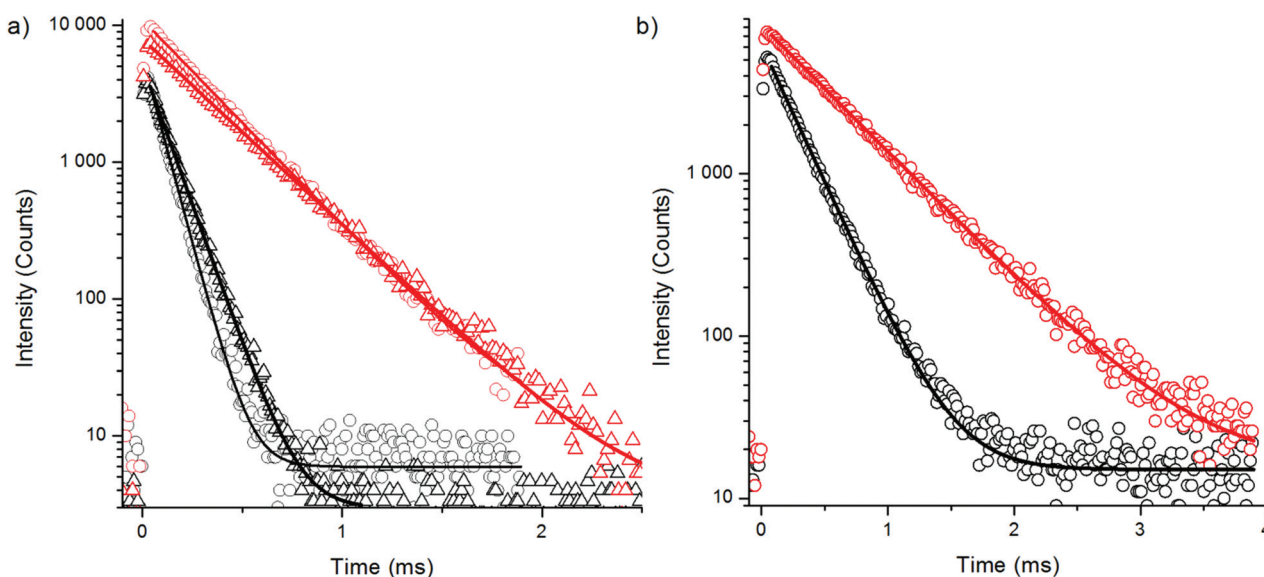


Fig. 10 Decay curves of 930 nm excited ytterbium emission at 980 nm in H₂O (black) and D₂O (red) of (a) bare (circles) and silanized (triangles) NaYF₄:Yb³⁺,Er³⁺ and (b) bare NaYF₄:Yb³⁺,Tm³⁺ using 3 μs excitation pulse width.



Table 4 Luminescence lifetimes and amplitude fractions of NaYF₄:Yb³⁺,Er³⁺ and NaYF₄:Yb³⁺,Tm³⁺ emission at 980 nm from ²F_{5/2} to ²F_{7/2} transition of Yb³⁺ upon 930 nm excitation

	Bare in H ₂ O	Bare in D ₂ O	Silanized in H ₂ O	Silanized in D ₂ O
NaYF ₄ :Yb ³⁺ ,Er ³⁺				
τ ₁ (μs)	79 ± 0.3 (100%)	288 ± 0.5 (100%)	106 ± 0.3 (100%)	319 ± 0.5 (100%)
NaYF ₄ :Yb ³⁺ ,Tm ³⁺				
τ ₁ (μs)	256 ± 0.7 (100%)	556 ± 1.0 (100%)		

dence of red upconversion luminescence intensity is slightly higher than that of the green luminescence,¹⁴ the absorption of the excitation light by water should affect the red emission peak more. The opposite was observed in water, *i.e.*, the green peak was quenched more in relation to the red peak (Fig. 2a and 9, Fig. S5 in the ESI†). Therefore, the absorption of NIR light by water cannot be the main mechanism in quenching of the upconversion photoluminescence.

To find an additional explanation for the strong quenching efficiency of water we studied the luminescence decay kinetics of the Yb³⁺ sensitizer emission at 980 nm upon 930 nm excitation for both bare NaYF₄:Yb³⁺,Er³⁺ and NaYF₄:Yb³⁺,Tm³⁺ nanophosphors and corresponding UCNPs with a silica shell in H₂O and D₂O using method 3 (Fig. 10, Table 4). The luminescence lifetime of bare NaYF₄:Yb³⁺,Er³⁺ was shortened by 73% and the lifetime of bare NaYF₄:Yb³⁺,Tm³⁺ nanophosphors was shortened by 54% in water when compared to D₂O. This major quenching in Yb³⁺ emission lifetime implicates that neither the increased non-radiative relaxation of the intermediate or emissive states of Er³⁺, nor the water absorption of excitation light at 980 nm, are the source of large quenching observed in upconversion luminescence intensities, but it is the ²F_{5/2} state of Yb³⁺ which is deactivated in water. The third vibrational overtone of OH-stretching can bridge the gap of ²F_{5/2} → ²F_{7/2} transition (~10 200 cm⁻¹) causing relaxation to ground state. The silica shell did not help much to prevent the quenching as revealed by the similar reduction in luminescence lifetime found for silica encapsulated UCNPs in water and that found for bare particles. In D₂O, also the intensity of the Yb³⁺ emission is enhanced.

The quenching effect of water by relaxation of the excited state of Yb³⁺ ion in UCNPs explains why the upconversion photoluminescence intensity of both Er³⁺ and Tm³⁺-doped nanophosphors is quenched so efficiently (Fig. 6 and 7). Because upconversion is a multiphoton process, the quenching of the sensitizer ion Yb³⁺ has a higher order effect on the upconversion luminescence. This was observed also as an increase in the slope of the power density dependency curve in water in comparison with the slope in D₂O (Fig. 9b). Further, due to the larger amount and short distances between individual Yb³⁺ ions, there seems to be an efficient energy migration between Yb³⁺ ions in the nanocrystal, which leads to the whole volume of the nanoparticle being susceptible to non-radiative

deactivation *via* surface OH-vibrations. Even the silica shell does not prevent the quenching of the excited state of Yb³⁺. A steep dependence on the excitation power density was found for the quenching efficiency of upconversion photoluminescence by water. The exchange of D₂O molecules to H₂O molecules was fairly slow and also the UCNPs-concentration affected the quenching efficiency.

4. Conclusions

Our results underline the strong influence of the surrounding water molecules on the intensity and decay behavior of the upconversion photoluminescence of nanometer-sized upconverting nanophosphors. The increased non-radiative relaxation of energy states of Er³⁺-dopants by high energy vibrational modes of OH-groups is, however, only partly responsible for the strong quenching observed in the upconversion luminescence intensity. As a major source of non-radiative deactivation, we identified the increased relaxation of the Yb³⁺ sensitizer ion, reflected by the considerably shortened luminescence lifetimes of its directly (at 930 nm) excited emission at 980 nm in water. Because upconversion photoluminescence is a multiphoton process, the quenching of the sensitizer can be expected to have a higher order effect on the luminescence intensity. Furthermore, it seems that Yb–Yb energy migration is very efficient in these nanocrystals, which makes the whole ~35 nm nanophosphors very susceptible to surface quenching effects. This clearly explains why up to 99.9% of the upconversion photoluminescence intensity can be quenched by water. A steep dependency on excitation power density was also found. In addition, the UCNPs-concentration had an effect on the quenching efficiency and the exchange of D₂O molecules to H₂O molecules was fairly slow.

Silica shell provided only little protection against the quenching effect of water. To improve the UCNPs-labels for bioanalytical applications, which are performed in aqueous solutions, better surface passivation is required to prevent this quenching and enhance the upconversion luminescence. Our findings about the role of Yb³⁺ quenching are of high importance for the rational design of new core–shell nanophosphors as a prerequisite for upconversion nanoparticles with improved brightness as most likely all photon upconverting materials containing Yb³⁺ dopants at or near the nanophosphor surface can be quenched by water. The surface passivation must prevent Yb–Yb energy migrations from reaching the nanophosphor surface, which is in contact with water and thus most vulnerable to quenching. Possible options reach from inactive NaYF₄ shells to layers of amphiphilic or hydrophobic polymers without high energy vibrational modes. An interesting option for a surface passivation through surface coating with amphiphilic polymers is the ligand attraction method, in which a bilayer with both hydrophobic and hydrophilic sections is formed.²⁷



Acknowledgements

This study was supported by Tekes, the Finnish Funding Agency for Technology and Innovation, the National Doctoral Programme of Advanced Diagnostic Technologies and Applications (DIA-NET). We gratefully acknowledge financial support from the Federal Ministry for Economic Affairs and Energy (MNPQ program BMWI 11/12), DFG (Deutsche Forschungsgemeinschaft) for a Heisenberg research fellowship (SCHA 1009/10-1), and the Erasmus Student Network. The authors would like to acknowledge the COST Action CM1403 funded by the European Union. The authors would also like to thank Emilia Palo and Vishal Kale for synthesizing the UCNPs, Joni Juvonen for TEM-image analysis, and Milla Suominen and Antti Viinikanoja for helping with the FT-IR analysis.

Notes and references

- 1 A. D. Ostrowski, E. M. Chan, D. J. Gargas, E. M. Katz, G. Han, P. J. Schuck, D. J. Milliron and B. E. Cohen, *ACS Nano*, 2012, **6**, 2686.
- 2 D. J. Gargas, E. M. Chan, A. D. Ostrowski, S. Aloni, M. V. P. Altoe, E. S. Barnard, B. Sanii, J. J. Urban, D. J. Milliron, B. E. Cohen and P. J. Schuck, *Nat. Nanotechnol.*, 2014, **9**, 300.
- 3 J. Shan, M. Uddi, N. Yao and Y. Ju, *Adv. Funct. Mater.*, 2010, **20**, 3530.
- 4 S. Schietinger, L. d. S. Menezes, B. Lauritzen and O. Benson, *Nano Lett.*, 2009, **9**, 2477.
- 5 J. Zhao, Z. Lu, Y. Yin, C. McRae, J. A. Piper, J. M. Dawes, D. Jin and E. M. Goldys, *Nanoscale*, 2013, **5**, 944.
- 6 F. Wang, J. Wang and X. Liu, *Angew. Chem., Int. Ed.*, 2010, **49**, 7456.
- 7 G. Liu, *Chem. Soc. Rev.*, 2015, **44**, 1635.
- 8 J.-C. Boyer, M.-P. Manseau, J. I. Murray and F. C. J. M. van Veggel, *Langmuir*, 2009, **26**, 1157.
- 9 S. V. Eliseeva and J. C. Bunzli, *Chem. Soc. Rev.*, 2010, **39**, 189.
- 10 N. Bogdan, F. Vetrone, G. A. Ozin and J. A. Capobianco, *Nano Lett.*, 2011, **11**, 835.
- 11 M. Pedroni, F. Piccinelli, T. Passuello, S. Polizzi, J. Ueda, P. Haro-González, L. Martinez Maestro, D. Jaque, J. García-Solé, M. Bettinelli and A. Speghini, *Cryst. Growth Des.*, 2013, **13**, 4906.
- 12 E. G. Moore, M. Seitz and K. N. Raymond, *Inorg. Chem.*, 2008, **47**, 8571.
- 13 A. Beeby, I. M. Clarkson, R. S. Dickins, S. Faulkner, D. Parker, L. Royle, A. S. de Sousa, J. A. G. Williams and M. Woods, *J. Chem. Soc., Perkin Trans. 2*, 1999, 493.
- 14 Y. Wang, L. Tu, J. Zhao, Y. Sun, X. Kong and H. Zhang, *J. Phys. Chem. C*, 2009, **113**, 7164.
- 15 G. S. Yi and G. M. Chow, *Adv. Funct. Mater.*, 2006, **16**, 2324.
- 16 L. Tu, X. Liu, F. Wu and H. Zhang, *Chem. Soc. Rev.*, 2015, **44**, 1331.
- 17 Q. Su, S. Han, X. Xie, H. Zhu, H. Chen, C.-K. Chen, R.-S. Liu, X. Chen, F. Wang and X. Liu, *J. Am. Chem. Soc.*, 2012, **134**, 20849.
- 18 Q. Zhan, J. Qian, H. Liang, G. Somesfalean, D. Wang, S. He, Z. Zhang and S. Andersson-Engels, *ACS Nano*, 2011, **5**, 3744.
- 19 K. Ahrar, A. Gowda, S. Javadi, A. Borne, M. Fox, R. McNichols, J. U. Ahrar, C. Stephens and R. J. Stafford, *J. Vasc. Interv. Radiol.*, 2010, **21**, 555.
- 20 M. Ylihärsilä, E. Harju, R. Arppe, L. Hattara, J. Hölsä, P. Saviranta, T. Soukka and M. Waris, *Clin. Microbiol. Infect.*, 2012, **19**, 551.
- 21 S. Wilhelm, T. Hirsch, W. M. Patterson, E. Scheucher, T. Mayr and O. S. Wolfbeis, *Theranostics*, 2013, **3**, 239.
- 22 T. Soukka, K. Kuningas, T. Rantanen, V. Haaslahti and T. Lövgren, *J. Fluoresc.*, 2005, **15**, 513.
- 23 T. Cao, Y. Yang, Y. Gao, J. Zhou, Z. Li and F. Li, *Biomaterial*, 2011, **32**, 2959.
- 24 F. Zhang, R. Che, X. Li, C. Yao, J. Yang, D. Shen, P. Hu, W. Li and D. Zhao, *Nano Lett.*, 2012, **12**, 2852.
- 25 J. Shan, M. Uddi, R. Wei, N. Yao and Y. Ju, *J. Phys. Chem. C*, 2010, **114**, 2452.
- 26 J. F. Suyver, A. Aebischer, S. García-Revilla, P. Gerner and H. U. Güdel, *Phys. Rev. B: Condens. Matter*, 2005, **71**, 125123.
- 27 S. Jiang, K. Y. Win, S. Liu, C. P. Teng, Y. Zheng and M.-Y. Han, *Nanoscale*, 2013, **5**, 3127.

