NPR

REVIEW

Chemistry of the pheromones of mealybug and scale insects

Yunfan Zou[a] and Jocelyn G. Millar*[ab]

Covering: 1979 to 2014

This article comprehensively reviews the syntheses of all known sex pheromones of scales and mealybugs, describes how they were identified, and how the synthetic pheromones are used in insect management.

1   Introduction

Mealybugs and scales (Order Hemiptera, Superfamily Coccoidea) comprise a group of small herbivorous insects with sucking mouthparts that colonize a wide variety of plants. Heavy infestations may result in direct plant damage and yield losses. Equal or greater indirect damage may be caused by the proliferation of sooty molds and other fungi on the insects' honeydew excretions. More recently, their importance as major agricultural pests has substantially increased due to their role as vectors of plant pathogens, particularly leafroll viruses. Furthermore, quarantine restrictions may prohibit importation of potentially infested agricultural commodities, particularly fruits, unless they have undergone expensive disinfestation treatments. The systematics, biology, and economic importance of identified scales and mealybugs are comprehensively summarized on the Scale Net website (http://www.sel.barc.usda.gov/scalenet/scalenet.htm), and methods for their management, including the use of pheromone-based techniques, have been recently reviewed.[1]

Most mealybugs and scales reproduce sexually, with the sedentary, wingless females producing powerful sex pheromones to attract the fragile and ephemeral males for mating. These pheromones have several interesting characteristics. First, they possess extraordinary biological activity, with lures baited with a few micrograms of pheromone remaining attractive for periods of at least several months under field conditions. Second, the structures of many of the pheromones are chemically interesting, usually consisting of irregular terpenoids, some of which possess unique terpenoid skeletons. These unique structures, coupled with the tiny amounts in which they are produced, have presented challenges for both their identification and their synthesis. Third, unlike most other insects, in which related species usually create unique pheromone signals by using different ratios or subsets of shared compounds, scales and mealybugs appear to create unique pheromone channels by producing species-specific structures. Several reviews of scale and mealybug pheromone chemistry and applications have been published, but all are outdated, and/or are focused on a subset of species.[2–5] Thus, this review will summarize the literature on the identification and synthesis of all known scale and mealybug pheromones, and conclude with a short summary of practical applications and commercialization of the pheromones.

aDepartment of Entomology, University of California, Riverside, CA 92521, USA. E-mail: Jocelyn.millar@ucr.edu; Fax: +1 951 827 3086; Tel: +1 951 827 5821

bDepartment of Chemistry, University of California, Riverside, CA, 92521, USA

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1067**

2 Methods for isolation and identification of scale and mealybug pheromones

Adult female scales and mealybugs can live for periods of several months, during which time they will continue to produce pheromone at the rate of a few ng per day until they are mated. Once mated, pheromone production is greatly reduced or ceases altogether, and so a key to identifying their pheromones is to produce large cohorts of virgin females which will release pheromone for weeks at a time. Thus, large colonies of the insects are established on factitious hosts such as potato sprouts or squash fruit, specifically chosen because they will last for a number of weeks without rotting. Cohorts of virgin females are then created in one of two ways. First, the developing males can be manually removed, often helped by the fact that they move off the host fruit to pupate.[6] The second, and more efficient and convenient method, takes advantage of the fact that the males undergo a complete metamorphosis to the adult stage whereas the females do not. Thus, males are much more sensitive to insect growth regulating hormones (IGRs) than females, and treatment of mixed sex cohorts of immatures with a discriminating dose of an IGR by spraying or dipping infested host materials into an IGR solution, will selectively kill the males, leaving large cohorts of virgin females on the host.[7] The female-infested material is then placed in glass chambers swept continuously with clean air, with the headspace volatiles collected on an adsorbent such as Porapak Q or activated charcoal, changed at daily to weekly intervals. The trapped volatiles are recovered by elution with solvents such as pentane or methylene chloride. Comparison of the profile of volatiles with that from collections from uninfested host material usually allows the insect-produced compounds to be singled out for identification. Alternatively, analysis of the extracts by gas chromatography coupled with electroantennogram detection, using antennae of males as living detectors, allows the unequivocal location of pheromone compounds in the extracts.[8]

As with other insect pheromones that are usually present in only microgram amounts in even composite extracts, most identifications of scale and mealybug pheromones have used spectroscopic methods, particularly mass spectrometry, in combination with microscale chemical tests to determine the presence or absence of specific functional groups. Some of these tests have included catalytic hydrogenation to determine the numbers of rings and C=C double bonds, ozonolysis to determine number and positions of double bonds, base hydrolysis (sometimes followed by reesterification) to confirm presence of esters, and carbon skeleton determination by high-temperature reduction of all functional groups (for full descriptions of these and other microchemical tests applied to semiochemical identification, see ref. 9). Further fragments of information about the structures have been garnered from high resolution mass spectrometry to obtain molecular formulae, and from use of retention indices to estimate molecular size and the possible presence of polar functional groups. For some of the more complex or unusual structures, identification by microscale NMR was unavoidable, sometimes requiring several years of insect rearing to collect enough material to obtain spectra.[8] In fact, reviewing some of the clever methods that have been used to isolate and identify microgram amounts of these volatile pheromones makes an interesting read.

3 Overview of scale and mealybug pheromones

To date, sex pheromones of 24 scale and mealybug species have been identified, from the families Diaspididae (7 species), Margarodidae (3 species), and Pseudococcidae (1 sp. in the subfamily Phenacoccinae, 13 spp. in the subfamily Pseudococcinae). For diaspidid and pseudococcid pheromones, all known examples are terpenoid derivatives, with many of them constituting esters of irregular, non-head-to-tail

Dr Yunfan Zou was born in Beijing, China, in 1975. After receiving his BSc in Chemistry and MSc in Organic Chemistry, both from Tsinghua University, he joined the group of Professor Michael Pirrung at Duke University, completing his doctoral studies in 2006. He has been working with Professor Jocelyn Millar at UC Riverside since 2007, first as a postdoctoral scholar, and currently as a research specialist. As a synthetic organic chemist, his research is focused on developing efficient and practical syntheses of newly discovered semiochemicals, both to confirm their identifications and to provide material for field trials.
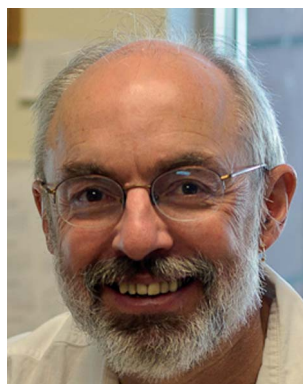
Jocelyn Millar holds the rank of Distinguished Professor in the Department of Entomology and Cooperating Faculty Member in the Department of Chemistry at the University of California, Riverside. He obtained his PhD in organic chemistry with A.C. Oehlschlager at Simon Fraser University in Canada in 1983, then did postdoctoral studies with R. M. Silverstein in Syracuse, New York, and E. W. Underhill at the National Research Council of Canada before joining UC Riverside in 1988. He is the author or coauthor of >275 papers, 25 book chapters and reviews, and 4 edited books on insect chemical ecology.

1068 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

terpene alcohols. They can be grouped into 4 structural categories: acyclic mono- or sesquiterpene alcohols, alcohols with a cyclopropane ring (chrysanthemol), mono- or sesquiterpenols with a cyclobutane ring, and monoterpene alcohols with a cyclopentane ring. The acid portion of these esters is variable, consisting of simple straight-chain acids such as acetic, propanoic, and butanoic acids, through to chiral acids such as 2-methylbutanoic and 2-acetoxy-3-methylbutanoic acids, and unsaturated acids such as senecioic and 3-methyl-3-butenoic acids. Exceptions include the hemiterpenol ester pheromone of the Matsumoto mealybug, and the degraded terpenoid ketone pheromone of *Aulacaspis murrayae*, which nevertheless contains the non-head-to-tail, 1′–2 linkage of two isoprene units that characterizes many of the acyclic pheromones. In contrast, the pheromones of margorodid scales are clearly of polyketide origin. In the descriptions of the identifications and syntheses, the pheromones are grouped by structural classes rather than by taxonomic relatedness, to highlight the structural similarities.

3.1 Acyclic pheromones

3.1.1 Ester of a hemiterpenol: Matsumoto mealybug pheromone.
The sex pheromone of the Matsumoto mealybug, *Crisicoccus matsumotoi* (Siraiwa), was identified by Tabata and coworkers as 3-methyl-3-butenyl 5-methylhexanoate **1**.[10] This is the first and to date only report of a hemiterpene-based pheromone structure from a scale or mealybug. In addition, the 5-methylhexanoic acid moiety is rare in insect pheromones. The pheromone was prepared by esterification of the alcohol with the acid using a dehydrative ester condensation catalyst, dimesitylammonium pentafluorobenzenesulfonate.[10]

1
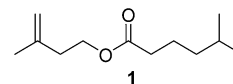
3.1.2 Esters of geraniol and nerol isomers: San Jose scale pheromone.
Female San Jose scale were found to produce three closely related compounds **2**, **3**, and **4**, as pheromone components, in a ratio of 48.5 : 46.7 : 4.8 (Fig. 1).[11–13]

Male scales were attracted to any one of the three compounds, and the compounds did not seem to act synergistically. The first synthesis was designed so that all three compounds could be made from a shared intermediate **9** (Scheme 1A).[12] Thus, orthoester Claisen rearrangement of the enolate **6** from 2-methyl-2-propen-1-ol **5** gave ester **7**, which was reduced and converted to the bromide **9**. Copper-catalyzed regioselective addition of the corresponding Grignard reagent **10** to 3-butyn-1-yl propanoate **11** gave the pheromone **2**. Alternatively, the Grignard reagent was added in the same fashion to TMS-protected 3-butyn-1-ol, followed by deprotection and esterification (not shown). Anderson subsequently developed a one-step procedure to the core alcohol structure **14** by alkylation of the dianion of 3-methyl-3-buten-1-ol **12** with 1-bromo-3-methyl-3-butene **13** (Scheme 1B).[13] Chong's group, apparently unaware of this synthesis, subsequently published a very similar route.[14]

Weiler and coworkers (Scheme 2) alkylated the dianion of methyl acetoacetate **15** with 1-bromo-3-methyl-3-butene **13**, then converted the resulting β-ketoester **16** to the enolate **17** to protect the ketone from reduction, and reduced the ester to an alcohol, with concomitant hydrolysis of the enolate during the workup to give **18**. Esterification followed by conversion of the ketone to a methylene completed the synthesis of **2**.[15]
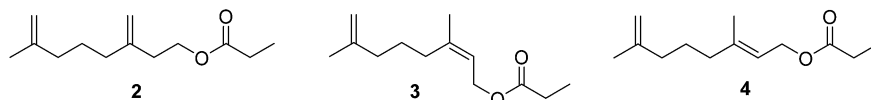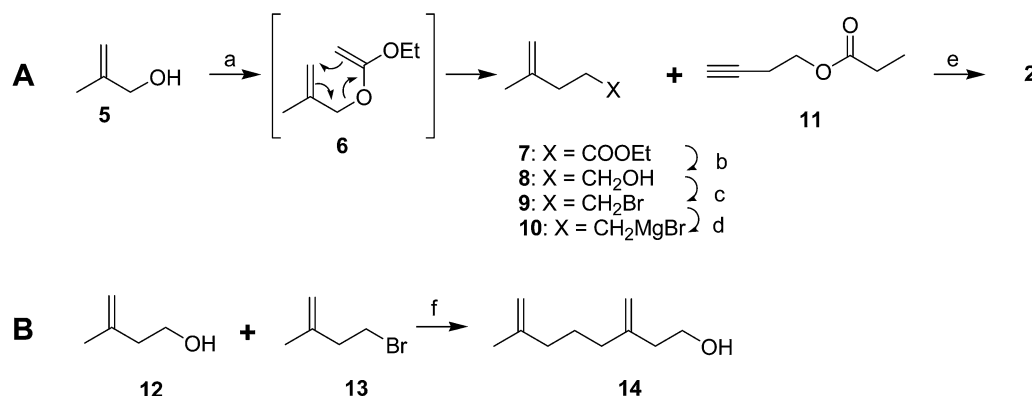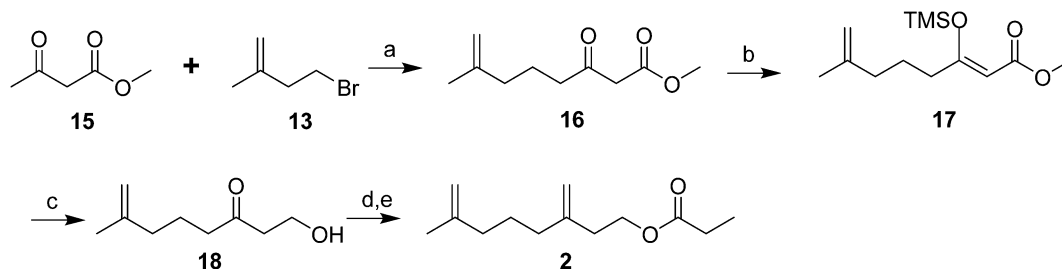

Fig. 1 The three components of San Jose scale pheromone.

Scheme 1 Two syntheses of **2**, the most abundant component of the San Jose scale pheromone. Reagents and conditions: (a) EtCOOH, (EtO)$_3$CH, reflux, 68%; (b) LiAlH$_4$, THF, 90%; (c) Ph$_3$PBr$_2$, pyridine, CH$_3$CN, 70%; (d) Mg, ether; (e) CuBr, Me$_2$S, −45−20 °C, 42%. (f) 2 BuLi, TMEDA, hexane, 0−20 °C, ~40%.

Scheme 2 Weiler and coworkers' synthesis of **2**. Reagents and conditions: (a) NaH, BuLi, THF, 70%; (b) TMSiCl, Et$_3$N, ether, 97%; (c) LiAlH$_4$, ether, 89%; (d) (EtCO)$_2$O, pyridine, 90%; (e) CH$_2$Br$_2$, Zn, TiCl$_4$, THF, 83%.

Lombardo and Weedon attempted a photochemical deconjugation of ethyl 3,7-dimethyl-2,7-octadienoate, but the desired product was obtained only in low yield in a mixture of byproducts; the synthesis was not useful and is mentioned only for completeness.[16] Veselovskii and coworkers also developed a low-yielding synthesis but there was insufficient detail given to determine whether the resulting product was pure or a mixture of isomers.[17]

The 2nd and 3rd pheromone components are (E)- and (Z)-isomers. Anderson and coworkers prepared the (Z)-isomer by regio- and stereoselective addition of Grignard reagent **10** to 1-propyne **19**, and reaction of the resulting vinyl cuprate intermediate **20** with CO$_2$, esterification with diazomethane, reduction of the ester **22**, and acylation (Scheme 3A).[13] The (E)-isomer was prepared by analogous copper-catalyzed reaction of 4-methyl-4-pentenyllithium **24** with methyl 2-butynoate **25**, reduction of the ester **26**, and acylation (Scheme 3B).[13]

In another synthesis of the two isomers using synthons with preset stereochemistry, Moiseenkov and coworkers alkylated 1-iodo-3-methyl-3-butene with an allylic (E)-hydroxysulfone or a related (Z)-hydroxysulfonamide, but because the preparation of these two synthons was not described, the overall utility of the syntheses is not clear.[18] A related scheme formed the carbon skeleton by sequential alkylation of bis(arylsulphonyl)methane with (E–Z)-4-chloro-3-methyl-2-buten-1-ol and ethyl 2-methyl-prop-2-en-1-yl carbonate but suffered from low overall yield and incomplete control of the alkene stereochemistry.[19] A 12-step synthesis of a mixture of the two isomers starting from methyl 4-oxopentanoate is mentioned only for completeness.[20]

In contrast, Weiler's group prepared the (E)- and (Z)-isomers by stereoselective formation of the (E)- and (Z)-enolates of β-ketoester **16** (the intermediate from the synthesis of the first pheromone component), trapping the enolates with diethylchlorophosphate,
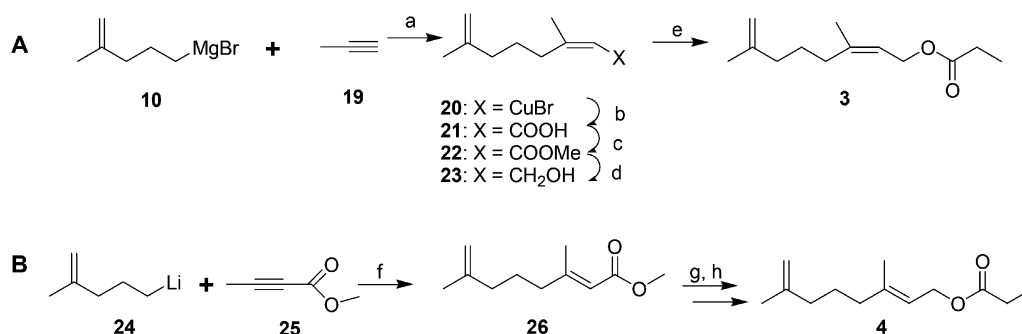
Scheme 3 Anderson *et al.* synthesis of the 2nd and 3rd components of the San Jose scale pheromone. Reagents and conditions: (a) CuBr, Me$_2$S, ether; (b) HMPA, CO$_2$, (EtO)$_3$P, −15 °C; (c) CH$_2$N$_2$, ether, 47% from **10**; (d) DIBAL, benzene, quant.; (e) (EtCO)$_2$O, pyridine, 20 °C, 62%. (f) CuI, TMEDA, ether, −60 °C, 85%; (g) DIBAL, benzene, 20 °C, 90%; (h) (EtCO)$_2$O, pyridine, 85 °C, 79%.
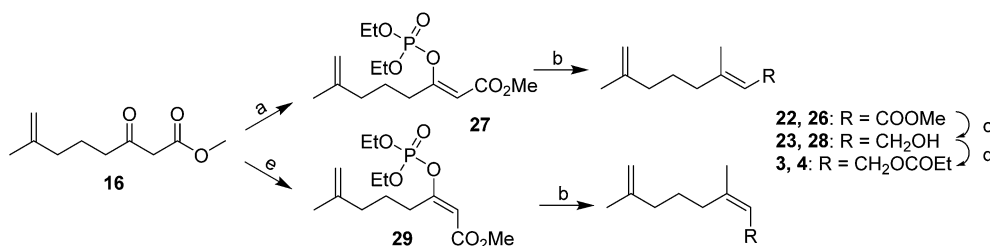
Scheme 4 Weiler's synthesis of the (E)- and (Z)-isomers constituting the minor components of the San Jose scale pheromone by selective formation of the (Z)- and (E)-enolates respectively. Reagents and conditions: (a) NaH, (EtO)$_2$POCl, THF, 90–95%; (b) MeMgCl, MeCu, THF, 68–70%; (c) DIBAL, ether, 98% for (E)-isomer, 82% for (Z)-isomer; (d) (EtCO)$_2$O, pyridine, DMAP, ether, 96% for (E)-isomer, 92% for (Z)-isomer; (e) Et$_3$N, DMAP, HMPA, (EtO)$_2$POCl, 97%.

1070 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

followed by alkylation with dimethyllithium cuprate with no loss of stereochemical integrity (Scheme 4).[15] Thus, treatment of the ketoester **16** with NaH in THF gave, after trapping with diethyl chlorophosphate, the (Z)-enol phosphate **27**, and reaction with MeMgCl with copper catalysis gave the trisubstituted alkene **22** as a 98E/2Z mixture. Choice of the reaction conditions and Grignard reagent counterion were critical to the maintenance of high stereochemical integrity. In contrast, formation of the enolate with Et$_3$N in HMPA, trapping as the (E)-enol phosphate **29**, and alkylation gave the Z-isomer **26** (98Z/2E). The syntheses were completed by reduction of the terminal esters **22** and **26** and acylation of the resulting alcohols **23** and **28**.

Rather than trying to make the trisubstituted alkenes stereoselectively, Novak et al. used readily available geraniol **30** or its acetate **31** or propanoate esters **32** as starting materials (Scheme 5).[21] Thus, regioselective chlorination with t-butyl hypochlorite with migration of the double bond produced the corresponding allylic chlorides **33–35** with a terminal methylene, with no loss of the stereochemical integrity of the other double bond. Alcohol **33** and acetate **34** were reduced with LiAlH$_4$, followed by esterification with propanoyl chloride, whereas the chloride **35** from geranyl propanoate was reduced directly to the final products with NaBH$_4$ and NaI in DMF. The same series of reactions using nerol as starting material produced the (Z)-alkene analogs. Minor variations involving different methods of removing the allylic leaving group were later published.[22]

3.1.3 Esters of lavandulol and related compounds. The vine mealybug pheromone (S)-**36**, the minor component of the pink hibiscus mealybug pheromone (2R,2'S)-**37**, and both components of the banana mealybug pheromone ((R)-**38** and

(R)-**39**) are all esters of lavandulol. The alcohol portion of the Japanese mealybug pheromone **40** and passionvine mealybug pheromone **41** are regioisomers of lavandulol. The Comstock mealybug pheromone (R)-**42** is also discussed with this group because of its obvious structural similarities to the other pheromones, and its likely genesis from lavandulol (Fig. 2). Chiral lavandulol can be prepared by a 3-step synthesis using Evans' chiral auxiliary,[23,24] or by enzymatic kinetic resolution of commercially available, racemic lavandulol.[25,26] The esters then are readily prepared by standard methods (acid chloride or anhydride and a base).

Vine mealybug pheromone. The sex pheromone of the vine mealybug, *Planococcus ficus*, was identified from mass spectrometry and microchemical tests as (S)-(+)-lavandulyl senecioate **36**.[27] Racemic lavandulyl senecioate proved to be as attractive to male mealybugs as the insect produced (S)-enantiomer, indicating no inhibition by the unnatural enantiomer. Lavandulol also was identified as a minor component in extracts from virgin females, but it was not attractive alone, and inhibited attraction at higher doses.[24] In Israeli populations, Zada and coworkers found another compound in headspace volatiles, (S)-(+)-lavandulyl isovalerate.[28] As in California, males were only attracted to the same major component **36**. During esterification of lavandulol with senecioyl chloride, choice of base was important; use of Et$_3$N resulted in extensive deconjugation of the senecioate, whereas pyridine yielded the desired ester cleanly.[27]

Pink hibiscus mealybug pheromone minor component. The sex pheromone of the pink hibiscus mealybug, *Maconellicoccus hirsutus* (Green), was identified by Zhang and coworkers as a 1 : 5 mixture of (R)-lavandulyl (S)-2-methylbutanoate **37** and
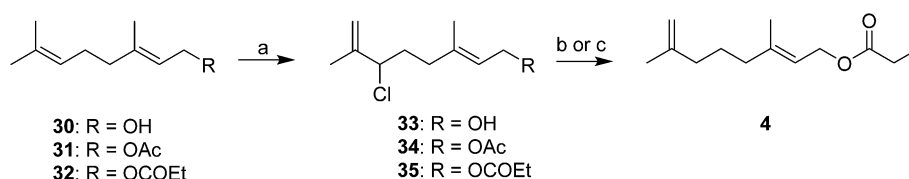
Scheme 5 Synthesis of one of the minor components of San Jose scale pheromone from geraniol or geranyl esters. Reagents and conditions: (a) t-BuOCl, silica gel, hexane, 0–20 °C, 61–78%; (b) for alcohol and acetate, LiAlH$_4$, THF, then EtCOCl, Et$_3$N, CH$_2$Cl$_2$, ~52–56%; (c) for propanoate ester, NaBH$_4$, NaI, DMF, 58%.

30: R = OH
31: R = OAc
32: R = OCOEt

33: R = OH
34: R = OAc
35: R = OCOEt

4

(S)-36
vine mealybug
Planococcus ficus

(2R,2'S)-37
pink hibiscus mealybug II
Maconellicoccus hirsutus

(R)-38
banana mealybug
Dysmicoccus grassii

(R)-39

40
Japanese mealybug
Planococcus kraunhiae

41
passionvine mealybug
Planococcus minor
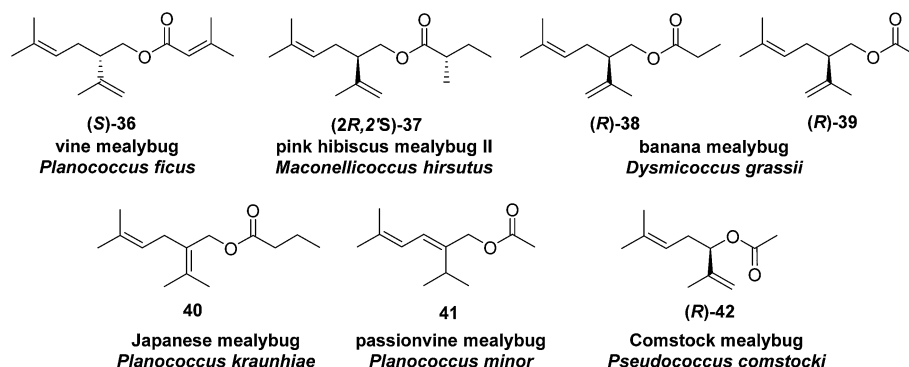
(R)-42
Comstock mealybug
Pseudococcus comstocki

Fig. 2 Esters of lavandulol and related compounds.

(R)-2,2-dimethyl-3-(1-methylethylidene)cyclobutylmethyl (S)-2-methylbutanoate [(R)-maconelliyl (S)-2-methylbutanoate (see Fig. 6, compound 257).[29] The alcohol portion of the minor component 37 is the enantiomer of the alcohol moiety of vine mealybug pheromone 36; it was prepared by a known synthesis.[23] The chirality–bioactivity relationship of 36 is discussed below with the description of the major component 257.

Banana mealybug pheromone. The sex pheromone of the banana mealybug, *Dysmiccocus grassii* Leonardi, was identified by de Alfonso and coworkers as (R)-(−)-lavandulyl propionate 38 and acetate 39 in a 6 : 1 ratio.[30] The major component 38 was more active than the minor component 39, and the two components acted additively rather than synergistically. The unnatural (S)-enantiomers were neither attractive nor inhibitory. The compounds were synthesized by kinetic resolution of racemic lavandulol,[25] followed by esterification with the appropriate acid chloride or anhydride.

Japanese mealybug pheromone. The sex pheromone of the Japanese mealybug, *Planococcus kraunhiae* (Kuwana), was identified by Sugie and coworkers as 2-isopropyliden-5-methyl-4-hexen-1-yl butyrate (fujikonyl butyrate) 40.[31] The alcohol portion, fujikonol, is a regioisomer of lavandulol, and the pheromone was generated in 4 steps from racemic lavandulol 43 by oxidation to aldehyde 44, acid catalyzed isomerization to the conjugated aldehyde 45, reduction, and esterification (Scheme 6).[32]

Passionvine mealybug pheromone. The sex pheromone of the passionvine mealybug, *Planococcus minor* (Maskell), was identified by Ho and coworkers as (E)-2-isopropyl-5-methyl-2,4-hexadienyl acetate 41.[33] To confirm the gross structure and determine the stereochemistry, a nonstereoselective synthesis based on Wittig reaction of a semi-stabilized allylic ylide was used to produce both isomers, which could only be separated on milligram scale by HPLC. The (E)-isomer attracted male mealybugs in laboratory bioassays, whereas the (Z)-isomer was antagonistic. Because this route was not suitable for production of the pheromone for practical use, Millar developed a short, stereoselective synthesis of 41 (Scheme 7)[34] by recognizing that the substitution pattern of the trisubstituted alkene 51 was ideally set up for copper-catalyzed, regio- and stereoselective *anti*-addition of an isopropyl Grignard reagent to propargylic alcohol 49. The stereoselectivity was proposed to result from formation of cyclic intermediate 50.[35] In the event, addition of

isopropylmagnesium bromide to enynol 49 gave dienol 51 with >99% (E)-selectivity. Acetylation completed the 3-step synthesis.
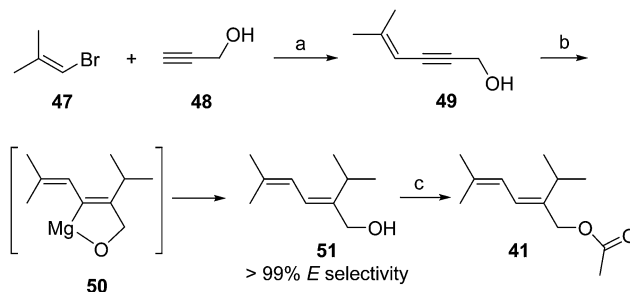
Comstock mealybug pheromone. The Comstock mealybug pheromone was the first such compound identified from a Pseudococcidae species, and so in what follows, we have described the identification in some detail to give the reader a sense of the methods that were available at that time. The pheromone was identified independently and virtually simultaneously by two groups. The first team, led by Barbara Bierl-Leonhardt, used the insect growth regulator method to generate cohorts of virgin females, collecting ~30 mg of pheromone over 2–3 years (~5 million female-day-equivalents).[36] The pheromone was identified primarily by mass spectral interpretation and microchemical tests, including hydrolysis and reacetylation to prove it was an acetate ester, catalytic reduction to determine that there were 2 C=C double bonds, ozonolysis to determine their positions, carbon skeleton determination by high temperature reduction to remove all functional groups, determination that the alcohol was a secondary alcohol by use of retention indices, and silylation of the alcohol portion followed by mass spectrometry to determine the position of the secondary alcohol.

The racemic pheromone was readily synthesized (Scheme 8),[37] with the key step being the rearrangement of epoxide 55 to the secondary allylic alcohol 56. Derivatization with Mosher's acid chloride gave two diastereomeric esters which were separable by GC, and corresponding derivatization of the alcohol from the natural pheromone gave one peak, verifying that the insect produced only one enantiomer, but it was not yet possible to determine which enantiomer it was, although the natural pheromone was shown to have a positive optical rotation. Fortunately, the racemic material was as attractive to males as the purified natural compound or as virgin females, which also suggested that the pheromone was a single component.

The parallel identification effort of Negishi *et al.* generated cohorts of virgin females by physical removal of males.[6] The pheromone structure was narrowed down to 6 possibilities by mass spectral interpretation and microchemical tests, followed by NMR of 6–7 μg of pheromone isolated from 600 000 females. The correct structure was determined by a two-step synthesis, albeit in low yield (Scheme 9A).[38] Thus, slow addition of a mixture of methacrolein 57 and 3-methyl-3-butenyl bromide 58
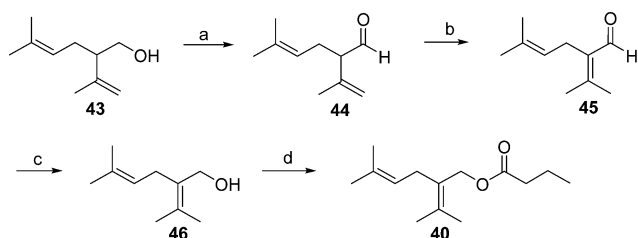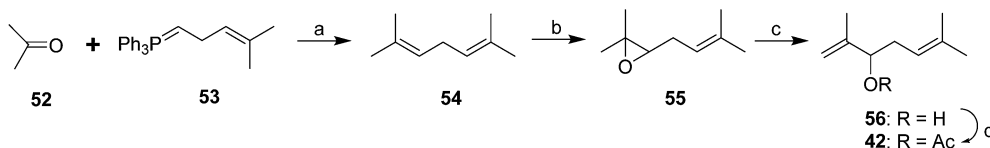

Scheme 6 Synthesis of the Japanese mealybug pheromone 40. Reagents and conditions: (a) Dess–Martin periodinane, CH_2Cl_2, room temp, overnight, 77%; (b) 1 M aq. HCl, THF, 50 °C, 24 h, 78%; (c) $NaBH_4$, EtOH, room temp, 4 h, 87%; (d) butyric anhydride, pyridine, 40 °C, overnight, 83%.

Scheme 7 Synthesis of the passionvine mealybug pheromone 41. Reagents and conditions: (a) CuI, $(Ph_3P)_2PdCl_2$, pyrrolidine, 40%; (b) *i*-PrMgBr, CuI, THF, 64%; (c) AcCl, pyridine, Et_2O, 97%.

1072 | *Nat. Prod. Rep.*, 2015, 32, 1067–1113

This journal is © The Royal Society of Chemistry 2015

Scheme 8 First racemic synthesis of the Comstock mealybug pheromone **42**. Reagents and conditions: (a) Solvent not stated; (b) m-CPBA, solvent not stated; (c) Al(i-PrO)$_3$, toluene, reflux; (d) Ac$_2$O, pyridine.

to Li metal in THF at room temp gave an 11.4% yield of the alcohol precursor **56** of the pheromone. In a subsequent improvement, reaction of 2-methylpropenyllithium **60** with 2-(methylethenyl)oxirane **59** proceeded in much better (60%) yield (Scheme 9B).[39]

The first chiral syntheses of the pheromone enantiomers were accomplished with Sharpless asymmetric epoxidation of an appropriate trisubstituted olefin alcohol **64**, followed by rearrangement and opening of the two enantiomeric epoxides **65** to produce the desired secondary allylic alcohol enantiomers in ~90% ee (Scheme 10).[40] The alcohol precursor **64** was produced in three steps from β-methallyl 2-pyridyl sulfide **61**.

The (R)-enantiomer of **42** had a positive optical rotation, and was much more active in bioassays, indicating that the natural pheromone was (R). Another synthesis of the (R)-enantiomer was accomplished by kinetic resolution of the alcohol precursor **56** to the pheromone (generated in 5 steps) by selective asymmetric epoxidation of the (S)-alcohol, leaving the (R)-enantiomer unchanged (not shown).[41]

Larchevêque and Petit devised a shorter and more efficient synthesis of either enantiomer, using chiral ethyl glycidates **66** (readily prepared from the enantiomers of serine in three steps) as chiral synthons (Scheme 11).[42] Thus, regioselective opening of the epoxide in (R)-**66** with diisobutenyllithium cuprate **67**
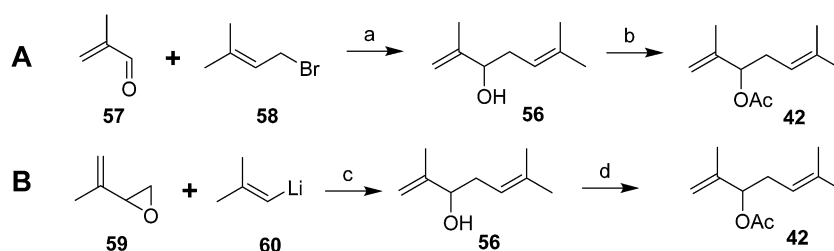
Scheme 9 Racemic syntheses of the Comstock mealybug pheromone **42**. Reagents and conditions: (a) Li, THF, room temp; 11.4%; (b) Ac$_2$O, pyridine, 77%. (c) THF, 60%; (d) AcCl, pyridine, ether, 77%.
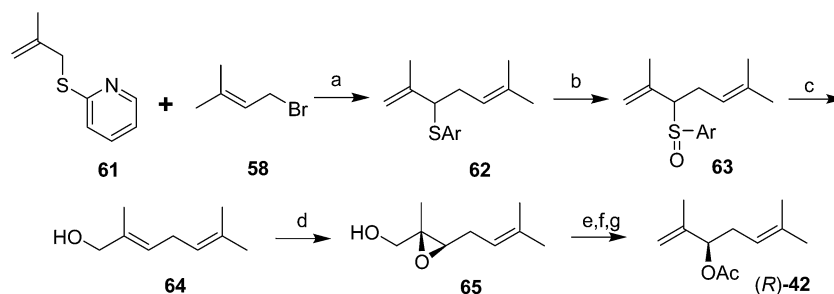

Scheme 10 Mori and Ueda's chiral synthesis of the Comstock mealybug pheromone. Reagents and conditions: (a) BuLi, THF, −70 °C-room temp, 85%; (b) m-CPBA, CH$_2$Cl$_2$, −30–10 °C, quant.; (c) Et$_2$NH, MeOH, 46%; (d) t-BuOOH, Ti(i-PrO)$_4$, diethyl (D)-(−)-tartrate, CH$_2$Cl$_2$, −23 °C; aq. tartaric acid, 59%; (e) TosCl, pyridine; NaI, acetone, reflux, 82%; (f) Zn, AcOH, 0 °C-room temp, 79%; (g) Ac$_2$O, pyridine, 80%.
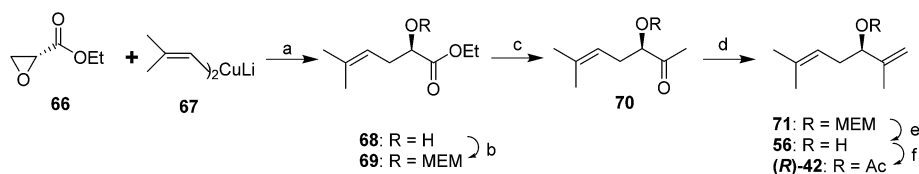
Scheme 11 Larchevêque and Petit's synthesis of one enantiomer of the Comstock mealybug pheromone from ethyl (R)-glycidate. Reagents and conditions: (a) Ether, −60 °C, 90%; (b) MEMCl, diisopropylethylamine, CH$_3$CN, 85%; (c) MeLi, TMEDA, ether/pentane, −100 °C, 80%; (d) Ph$_3$-PCH$_3$Br, BuLi, ether, 50%; (e) pyridinium p-toluenesulfonate, EtOH, reflux, 88%; (f) Ac$_2$O, pyridine, 90%.
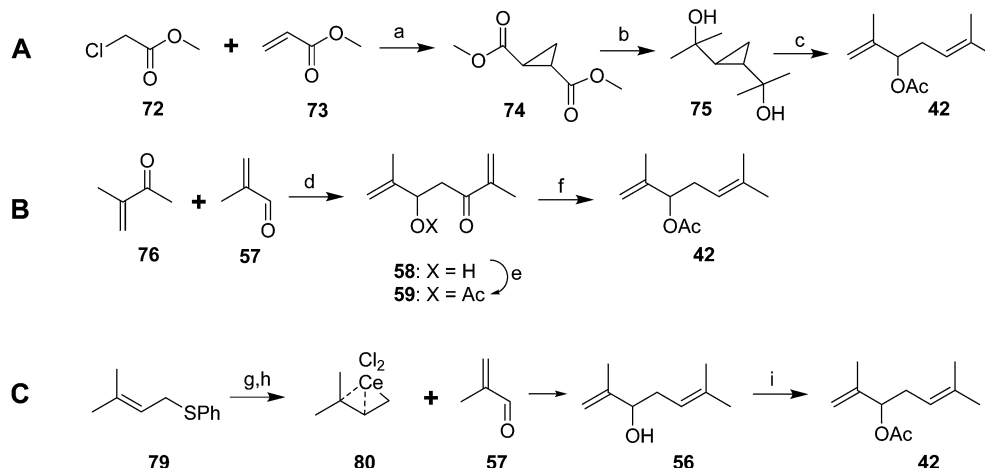
Scheme 12 Several short syntheses of racemic Comstock mealybug pheromone. Reagents and conditions: (a) NaOMe, then NaOMe, MeOH, 66%; (b) MeLi, ether, quant; (c) AcOH, Ac$_2$O, 80 °C, 74%. (d) LDA, ether, −60 °C; (e) Ac$_2$O, pyridine; (f) p-TsNHNH$_2$, NaBH$_3$CN, AcOH, 70 °C. (g) Li 1-(dimethylamino)naphthalenide, THF, −78 °C; (h) CeCl$_3$; (i) Ac$_2$O, pyridine.

gave the chiral hydroxyester **68**. MEM protection of the alcohol followed by low temperature reaction of **69** with MeLi gave ketone **70**. Straightforward Wittig olefination, removal of the protecting group, and acetylation completed the synthesis. Use of the TBDMS protecting group instead of MEM resulted in lower yields.[42]

Several relatively short and efficient syntheses of the racemate were subsequently developed. For example, base-catalyzed tandem Michael addition/cyclization of methyl chloroacetate **72** and methyl acrylate **73** followed by equilibration of the crude product in MeOH/MeONa gave a 66% yield of the *trans*-diester cyclopropane **74** (Scheme 12A).[43] Treatment of the diester with MeLi gave the crystalline diol **75** almost quantitatively, reaction of which with AcOH/Ac$_2$O gave the racemic pheromone **42** (74%, ~50% overall). Alternatively, condensation of 3-methyl-3-buten-2-one **76** and methacrolein **57**, acetylation, and removal of the ketone with concomitant migration of a double bond provided another short route to racemic **42** in ~50% yield (Scheme 12B).[44]

A third synthesis produced alcohol **56** from reaction of 1-methylethenylmagnesium bromide with 4-methyl-3-pentenal (generated from acrolein in 3 steps), for an overall 30% yield (not shown).[45] A fourth carried out a 4-step sequence in one pot, with 45% isolated yield (Scheme 12C).[46] The critical feature was the transmetallation of the initial lithium anion with CeCl$_3$ to produce the cerium complex **80** in order to direct attack of the least substituted carbon towards the carbonyl of methacrolein **57**, giving alcohol **56** in 54% yield.

Other, less efficient approaches that have been explored include the photooxidation of 2,6-dimethyl-2,5-heptadiene[47] and a rather long 9-step route centered on a sila-Cope rearrangement.[48]

3.1.4 Esters of sesquiterpenols, and solanone, a degraded sesquiterpenoid. The core structures of the sex pheromones of three diaspidid species, California red scale, yellow scale, and white peach scale are based on sesquiterpenols with the unusual 1′–2 linkage between the first two isoprene units that characterizes the lavandulol-based structures described above,
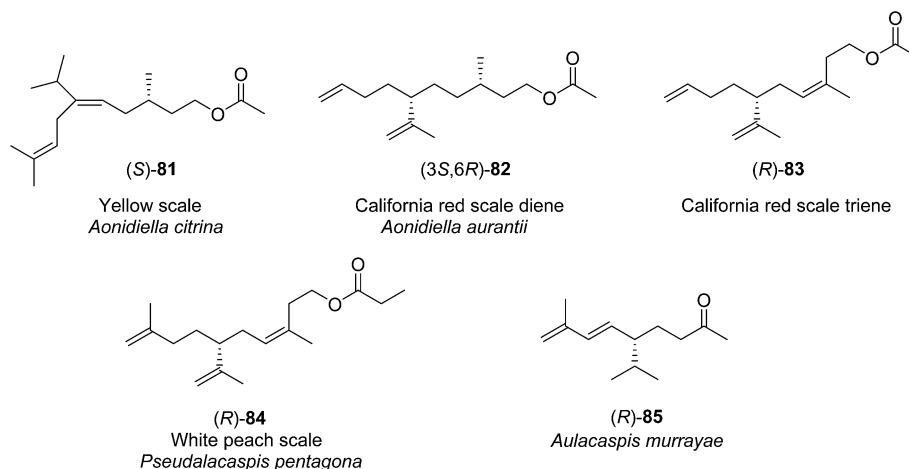
Fig. 3 Structures of the pheromones of California red scale, yellow scale, white peach scale, and the scale *Aulacaspis murrayae*.

and then with a third isoprene unit attached in normal fashion. The structures of the yellow scale and white peach scale pheromones contain the entire sesquiterpenoid skeleton, whereas the two components of the red scale pheromone have been truncated by one carbon. The pheromone of *Aulacaspis murrayae* is also discussed with this group because it was likely derived from degradation of a sesquiterpenoid (Fig. 3).

Yellow scale pheromone. The basic structure of the yellow scale pheromone 81 was identified *via* a combination of mass and ^{1}H NMR spectral data, and the results of microchemical tests (catalytic hydrogenation, LiAlH$_4$ reduction and reacetylation,

ozonolysis), but there was insufficient data to determine the absolute configuration or the stereochemistry of the double bond.[49] Thus, the first nonstereoselective synthesis by Anderson and Henrick was designed to provide both the (*Z*)- and (*E*)-isomers (Scheme 13).[50] Monoprotection of 3-methyl-1,5-pentanediol 86, pyridinium chlorochromate oxidation, and reaction of the resulting aldehyde 88 with the anion from diethyl 1-ethoxycarbonyl-2-methylpropylphosphonate 89 gave trisubstituted alkene 90 as a 4 : 1 mixture of (*Z*)- and (*E*)-isomers. Reduction of the ester, acetylation of the resulting allylic alcohol 91, and alkylation of 92 with diisobutenyllithium cuprate
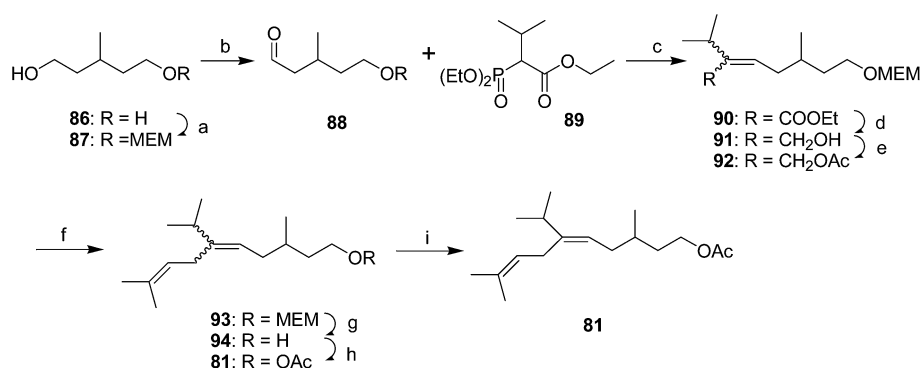
Scheme 13 First nonstereoselective synthesis of yellow scale pheromone. Reagents and conditions: (a) BuLi, THF, MEMCl, 47%; (b) PCC, NaOAc, CH$_2$Cl$_2$, 82%; (c) NaH, THF, 58%, 4 : 1 (*Z*–*E*); (d) DIBAL, benzene, 90%; (e) Ac$_2$O, pyridine, quantitative; (f) diisobutenyllithium cuprate, ether, −25 °C, 60%; (g) Cl$_3$CCOOH, EtOH/H$_2$O, 70 °C; (h) Ac$_2$O, pyridine, 65% over 2 steps; (i) preparative GC.
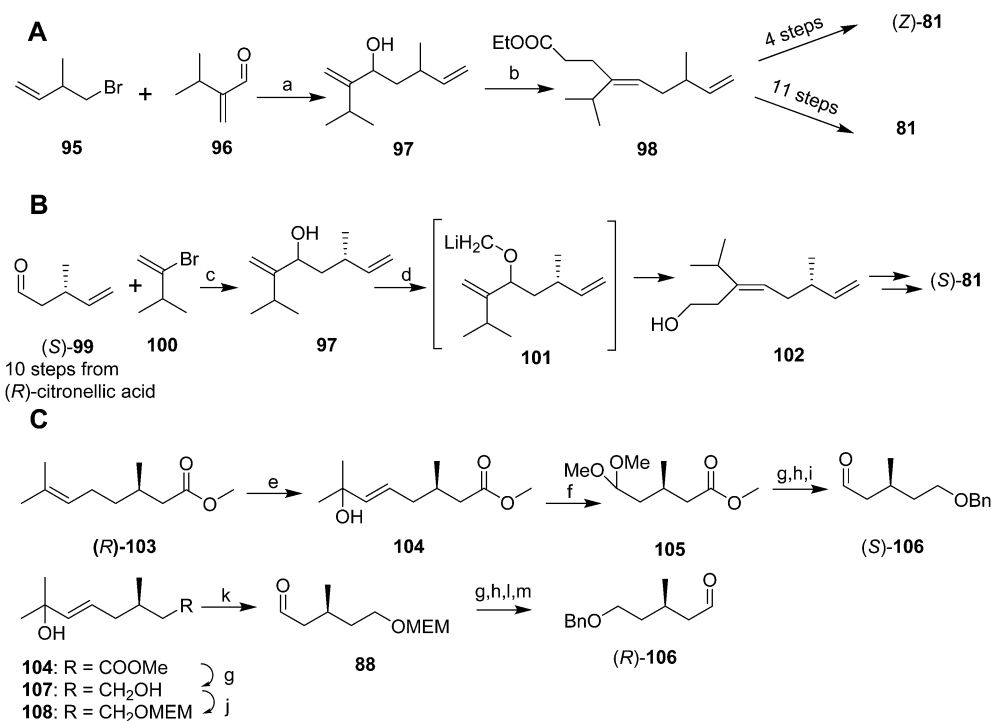
Scheme 14 Three syntheses of yellow scale pheromone by Mori's group. Reagents and conditions: (a) Mg, ether, 34%; (b) ethyl orthoacetate, H$^+$, 135 °C, 63%. (c) Mg, THF, 30%; (d) KH, 18-crown-6, THF, then ICH$_2$SnBu$_3$, then −78 °C, BuLi, 44%. (e) (PhSe)$_2$, H$_2$O$_2$, CH$_2$Cl$_2$, then *t*-BuOOH, 83%; (f) O$_3$, MeOH, −78 °C, then Me$_2$S, then MeOH, PTSA, 83%; (g) LiAlH$_4$, ether, 87%; (h) NaH, DME, PhCH$_2$Br; (i) HClO$_4$, THF/H$_2$O, ~quantitative. (j) NaH, MEMCl; (k) O$_3$, MeOH, −78 °C, then Me$_2$S; (l) MeOH, PTSA, 40 °C, 36% over 6 steps; (m) CrO$_3$·2Py.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1075**

completed the carbon skeleton. After deprotection and acetylation, the resulting isomers were readily separable by preparative GC, with the (E)-isomer **81** matching the insect-produced compound.

Mori and coworkers developed three syntheses of the pheromone in rapid succession, with the first aimed at determining the double bond geometry,[51] the second at developing a chiral synthesis producing one enantiomer,[52] and the third at providing both enantiomers.[53] These syntheses were long and low-yielding, have been previously reviewed in detail, and so they are only summarized here. In the first synthesis (Scheme 14A),[51] reaction of the Grignard reagent prepared from 1-bromo-2-methyl-3-butene **95** with 2-(1-methylethyl)acrolein **96** followed by orthoester Claisen rearrangement gave ester **98** which was elaborated to the (Z)-isomer of **81** in 4 steps. The correct (E)-isomer **81** was generated from ester **98** in 11 steps, using a reaction sequence that included inversion of the double bond by the sequence of epoxidation, opening the epoxide with lithium diphenylphosphide, methylation of the alkyldiphenylphosphine intermediate to form the phosphonium salt, and elimination of methyldiphenylphosphine oxide.

Mori's second synthesis used enantiomerically pure (R)-citronellic acid, in turn generated from (R)-pulegone, as the source of chirality (Scheme 14B).[52] Thus, aldehyde **99**, generated in 10 steps from (R)-citronellic acid, was reacted with the Grignard reagent from 3-methyl-2-bromo-1-butene **100** (generated in 2 steps and 9% yield from 3-methyl-1-butene, and requiring spinning band distillation purification) to give chiral alcohol **97**. O-Alkylation with iodomethyltributyltin and transmetallation with lithium to give **101** then induced a 2,3-Wittig rearrangement to yield (E)-alcohol **102** stereoselectively, which was elaborated to the (S)-enantiomer (S)-**81** in the same sequence of steps used in their previous synthesis.

The third synthesis produced both pheromone enantiomers from methyl (R)-citronellate **103** (Scheme 14C).[53] Thus, regiospecific Markovnikov addition of phenylselenenic acid (generated in situ from diphenyldiselenide and H_2O_2) to the alkene, followed by oxidation of the alkylphenylselenium intermediate and elimination gave allylic alcohol **104**. Ozonolysis and reductive quenching with dimethylsulfide, and protection of the aldehyde as the dimethylacetal, followed by reduction of the

ester and protection of the resulting alcohol as the benzyl ether yielded a chiral synthon **105** with differentiated terminals. Deprotection of the dimethyl acetal and reaction of the aldehyde (S)-**106** with the Grignard reagent from vinyl bromide **100** provided the key intermediate required for the 2,3-Wittig rearrangement used in the previous synthesis to establish the (E)-trisubstituted double bond, and further elaboration to the final product. Alternatively, the primary alcohol of diol **107** was MEM protected, followed by ozonolysis, reduction of the resulting aldehyde (S)-**88** with $LiAlH_4$ to the alcohol, and protection as the benzyl ether, giving a second chiral synthon with differentiated ends. Removal of the MEM protecting group and oxidation gave the (R)-enantiomer of aldehyde **106**, which was then carried through to (R)-**81** as described above.
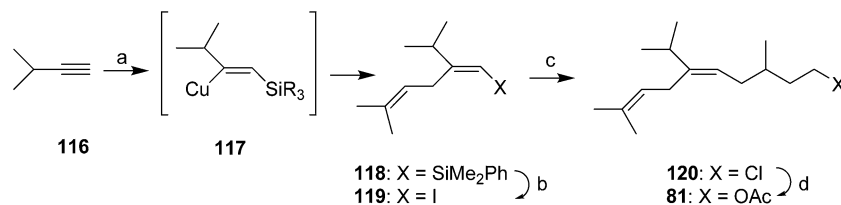
A synthesis from Julia's group controlled the stereochemistry of the trisubstituted double bond by stereoselective elimination of AcOH from a vicinal acetoxysulfone **114** (Scheme 15).[54] Thus, (R)-3-methylvalerolactone **109**, generated by desymmetrization of 3-methylpentanedioc acid dimethyl ester with pig liver esterase to give the (R)-half-ester, followed by selective reduction of the free acid with borane and acid-catalyzed closure of the lactone, was alkylated with the anion of sulfone **110** with attack at the carbonyl of the lactone, giving hydroxyketone **111**. Protection of the alcohol, reduction of the ketone and acetylation gave vicinal acetoxysulfone **114**, which stereoselectively eliminated AcOH when treated with powdered NaOH in ether to give the (E)-trisubstituted alkene **115** in 98% stereochemical purity. Reaction of the sulfone acetate **114** with isopropylmagnesium bromide with $FeCl_3$ catalysis unfortunately gave an ~1 : 1 mixture of the (E)-alkene **94** (with no loss of stereochemical purity) and the product from hydrogenolysis instead of alkylation. Deprotection and acetylation completed the synthesis of (S)-**81**, with separation of the desired trisubstituted alkene from the disubstituted byproduct by flash chromatography on silica gel impregnated with silver nitrate.

Millar developed a short, convergent, and highly stereoselective synthesis via silylcupration of an acetylene (Scheme 16).[55] Thus, regiospecific addition of bis(phenyldimethylsilyl) lithium cuprate to 3-methyl-1-butyne **116** and trapping of the intermediate vinyl cuprate **117** with prenyl bromide gave **118** in 77% yield. Replacement of the R_3Si group with iodine, and
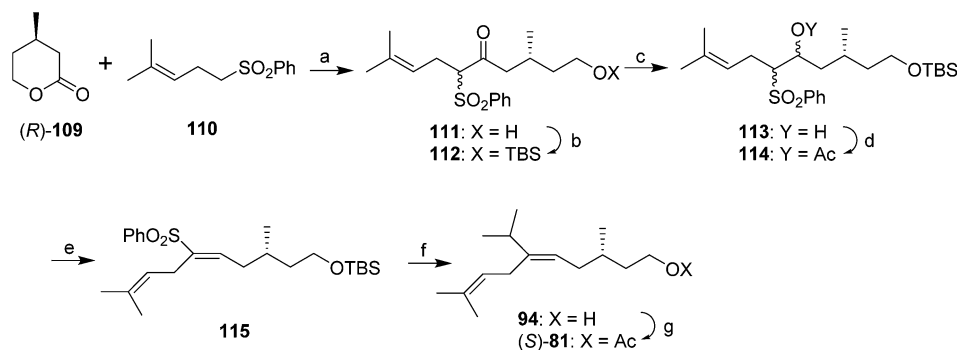
Scheme 15 Julia's synthesis of yellow scale pheromone. Reagents and conditions: (a) BuLi, THF/TMEDA, 62%; (b) TBDMSiCl, Et₃N, DMAP, CH₂Cl₂, 97%; (c) NaBH₄, MeOH/H₂O, 92%; (d) Ac₂O, Et₃N, DMAP, CH₂Cl₂, 95%; (e) powdered NaOH, ether, 94%; (f) i-PrMgCl, FeCl₃, ether, −78–20 °C, then TBAF·3H₂O, THF, 36%; (g) Ac₂O, Et₃N, DMAP, CH₂Cl₂, 96%.

**1076** | Nat. Prod. Rep., 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

Scheme 16 Millar's synthesis of yellow scale pheromone. Reagents and conditions: (a) (PhMe$_2$Si)$_2$CuLi, THF, −78 °C, then prenyl bromide, 77%; (b) I$_2$, MeCN, 51%; (c) 4-chloro-2-methylbutylzinc iodide, (Ph$_3$P)$_4$Pd, THF; (d) NaOAc, HMPA, 75 °C, 42% from the vinyl iodide **119**.

reaction of the resulting unstable vinyl iodide **119** with 4-chloro-2-methylbutylzinc iodide with (Ph$_3$P)$_4$Pd catalysis, and conversion of the crude coupling product **120** to the acetate **81** completed the synthesis, with complete control of the alkene stereochemistry. Although the racemate was prepared, the synthesis could be readily modified to produce either
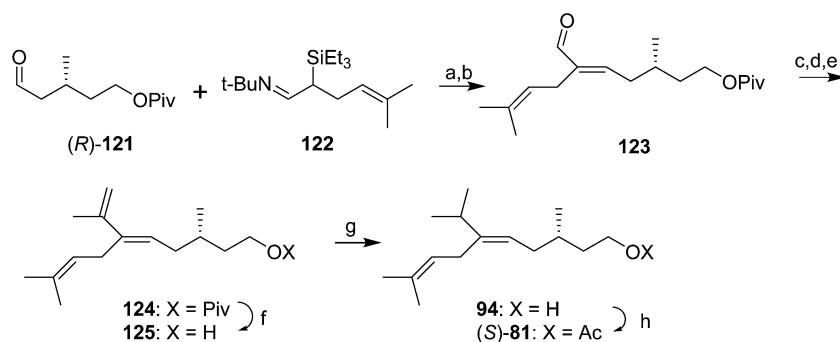
Scheme 17 Baudouy and Sancho's synthesis of yellow scale pheromone. Reagents and conditions: (a) s-BuLi, THF, −78−0 °C, then CF$_3$COOH and water, 66%; (b) pyridinium hydrochloride, CH$_2$Cl$_2$, 100%; (c) MeMgI, ether, −70 to −35 °C; (d) (COCl)$_2$, DMSO, Et$_3$N, CH$_2$Cl$_2$; (e) MePh$_3$PBr, BuLi, THF, 20 °C-reflux; 90% from the aldehyde; (f) DIBAL, solvent not stated, quantitative; (g) (Ph$_3$P)$_3$RhCl, H$_2$, benzene, quantitative; (h) Ac$_2$O, Et$_3$N, solvent not stated, 94%.
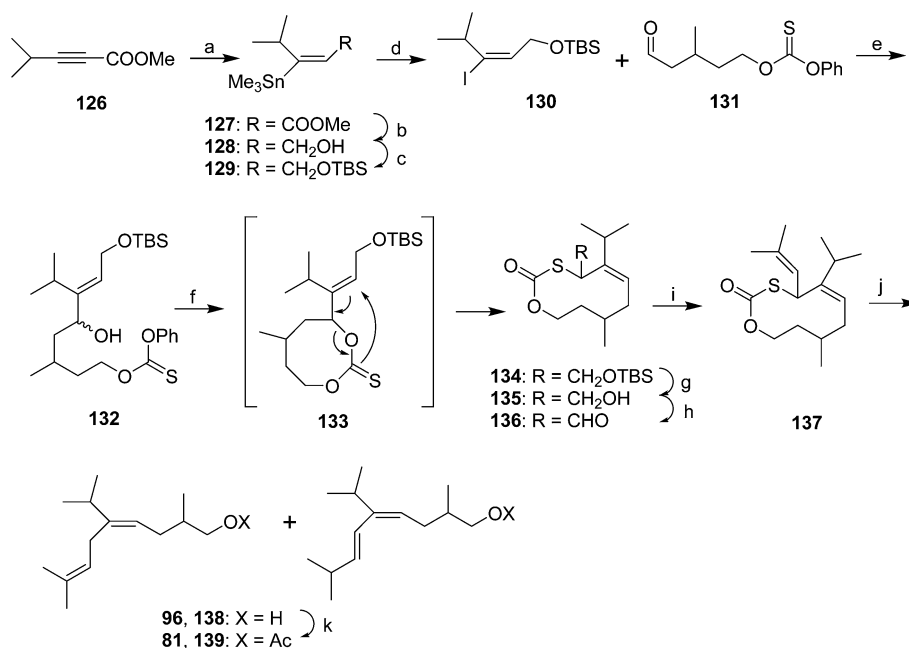
Scheme 18 Synthesis of racemic yellow scale pheromone *via* [3,3]-sigmatropic rearrangement of a cyclic thiocarbonate. Reagents and conditions: (a) Me$_3$SnCu·SMe$_2$, THF, −78 °C, 87%; (b) DIBAL, pentane/toluene, ∼-45 °C, 82%; (c) TBDMSiOTf, pyridine, quantitative; (d) I$_2$, ether, quantitative; (e) t-BuLi, ether, −78 °C; (f) LiHMDS, THF, 72% from stannane **128**; (g) TBAF, THF, 97%; (h) (COCl)$_2$, DMSO, CH$_2$Cl$_2$, −78 °C, then Et$_3$N; (i) i-PrPh$_3$PBr, BuLi, ether, 46−60% from alcohol **135**; (j) Li, NH$_3$ or LDBB, THF/HMPA; (k) Ac$_2$O, DMAP, CH$_2$Cl$_2$, 60% over 2 steps.
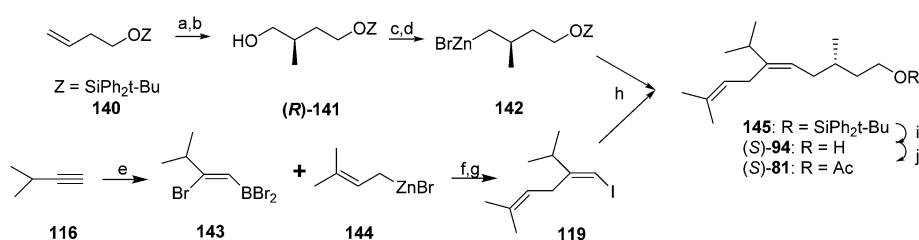
72% ee, was readily upgraded to 98% ee by a lipase-catalyzed kinetic resolution. The resulting alcohol (R)-**141** was then converted to an iodide, ready for coupling. To construct the key trisubstituted alkene, addition of BBr_3 across the triple bond of 3-methyl-1-butyne **116**, Pd catalyzed replacement of the bromide on carbon 2 with prenylzinc bromide, and conversion of the resulting alkenyl boron dibromide to the vinyl iodide **119** proceeded with complete regio- and stereochemical control, in 77% yield. The first synthon was then lithiated and transmetallated with $ZnBr_2$ to give **142**. Pd-catalyzed reaction between organozinc **142** and vinyl iodide **119** then provided the completed carbon skeleton in very high stereochemical purity. Removal of the silyl group and acetylation completed the synthesis, providing (S)-**81** in 34% yield over 6 steps. As an interesting side note, during the course of the development and optimization of the coupling between allylzinc bromides and bromoborane intermediates from addition of BBr_3 to terminal alkynes (e.g., **143** + **144** → **119**), the authors found that allylzinc bromide itself did not react, but any other allylzinc bromide with substituents on either the 2 or 3 positions, reacted readily.

Pheromone components of California red scale. The pheromone of California red scale was the first pheromone identified from the insect order Hemiptera, using material from an astonishing 400 million female-days of collection.[59] The identification made extensive use of microchemical tests, along with ^{1}H NMR. The pheromone consists of diene **82** and triene **83**. The insect produces the (3S,6R)-enantiomer of **82**, but the (3S,6S)-

diastereomer is also active, and in fact, males respond well to the full blend of 4 stereoisomers, indicating that the three unnatural isomers are not inhibitory. The more unsaturated compound **83** also has the (6R)-configuration.

The challenges in synthesizing the two compounds are different because the diene component has two chiral centers but no stereochemistry associated with the double bonds, whereas the triene component has a single chiral center, and a trisubstituted double bond in which the stereochemistry must be controlled. Consequently, despite their superficial similarity, quite different synthetic strategies and intermediates were used in their syntheses.

Diene component of the California red scale pheromone. The first reported synthesis of the diene component **82** as a diastereomeric mixture used an $AlCl_3$-catalyzed ene reaction of citronellyl acetate **146** and methyl propiolate **147** to give diester **148** (Scheme 20).[60a] Selective reduction of the conjugated double bond, followed by a 4-step sequence to add the additional carbon and convert it to a terminal double bond (reaction of the methyl ester with phenylsulfonylmethyllithium to add one carbon, reduction of the resulting ketone to the secondary alcohol, acetylation, and reduction of the geminal sulfone acetate with sodium amalgam in EtOH) and reacetylation to replace the acetate which had come off during this sequence, completed the synthesis of **82** in about 17% overall yield. Another route using an ene reaction between citronellyl acetate and formaldehyde gave the desired alcohol product in good
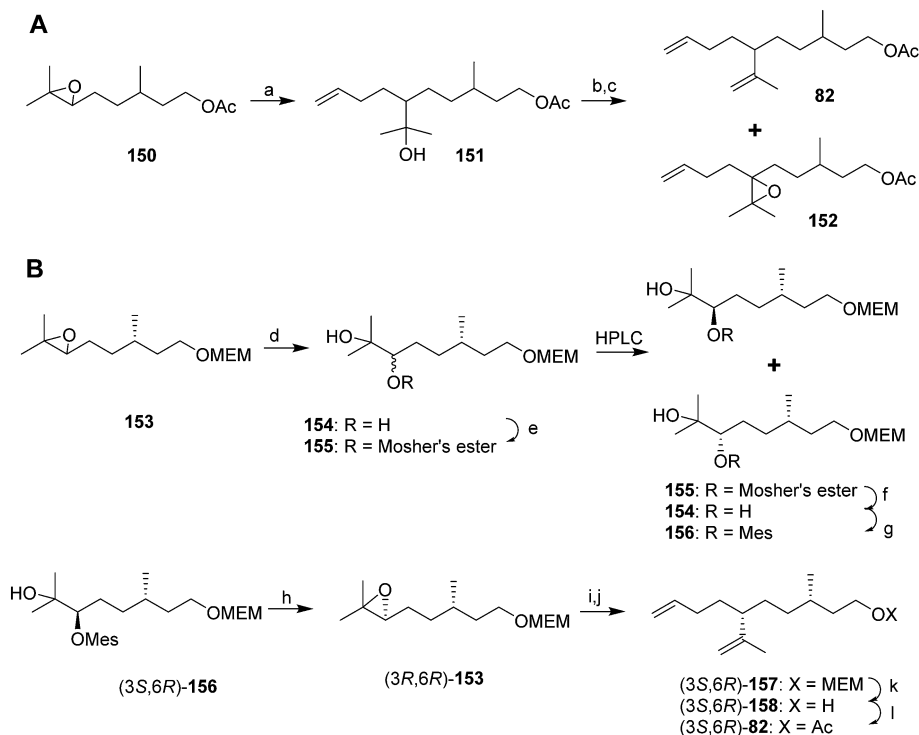
Scheme 21 Anderson's syntheses of racemic and chiral diene **82**. Reagents and conditions: (a) di(3-butenyl)lithium cuprate, ether, −20−4 °C, then Ac_2O, pyridine, 46%; (b) MesCl, Et_3N, 0 °C-room temp; (c) m-CPBA, $CHCl_3$, room temp, prep. TLC, 73%. (d) $HClO_4$, THF/H_2O, 67%; (e) Mosher's acid chloride, DMAP, MeCN, then HPLC; (f) $LiAlH_4$, THF, 73%; (g) MesCl, Et_3N, CH_2Cl_2; (h) KOH, MeOH, 62% over 2 steps; (i) di(3-butenyl) lithium cuprate, ether, −20−4 °C, 70%; (j) MesCl, Et_3N, 97%; (k) BuLi, hexane, then H_3O^+, quant.; (l) Ac_2O, Et_3N, ether, then m-CPBA, CH_2Cl_2, room temp, prep. TLC, 37%.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1079**

yield, but attempts to convert this alcohol to the pheromone produced only complex mixtures of products (not shown).[60b]

A cleaner and more practical synthesis of 82 by the authors who identified the pheromone made clever use of available chiral synthons by first developing a short nonstereoselective synthesis from racemic citronellol to prove the basic structure (Scheme 21A).[61] Thus, the epoxide of (±)-citronellyl acetate 150 (or citronellyl pivalate) was reacted with di(3-butenyl)lithium cuprate to regioselectively open the epoxide, and the resulting tertiary alcohol 151 was then mesylated followed by elimination in the same pot, giving a 4 : 1 mixture of the desired isomer 82 with the tetrasubstituted double bond isomer. The latter was removed by selective epoxidation of the more electron rich tetrasubstituted double bond and chromatographic separation of the epoxide 152 from the desired product 82. When the acetate ester was used in the alkylation step, the acetate was partially removed, requiring reacetylation of the crude product, whereas this was not the case with the pivalate ester, but with the pivalate route, the pivalate had to be removed later and replaced with acetate.

The synthesis was then repeated using (R)- and (S)-citronellols, giving diastereomeric mixtures of (3R,6R/S)- and (3S,6R/S)-isomers respectively. Only the latter was attractive to male

scales, and so the epoxide from MEM-protected (S)-citronellol 153 was opened to give diastereomeric diols 154, the Mosher's ester derivatives of which (155) were separable by HPLC (Scheme 21B). Reduction of the purified (3S,6R)-hydroxy ester to the diol 154, mesylation, and ring closure to reform the epoxide (3R,6R)-153, followed by the alkylation and elimination sequence as before, gave intermediate 157. Although removal of the MEM protecting group by standard methods failed, the authors developed a two-step sequence of treatment with strong base followed by aqueous acid that delivered the alcohol (3S,6R)-158 in quantitative yield. Routine acetylation completed the synthesis, and the resulting (3S,6R)-82 exactly matched the naturally produced compound. The (3S,6S)-diastereomer was made in analogous fashion from the other hydroxyester diastereomer.

Baudouy and Maliverney exploited (−)-dihydrocarvone 159 as a source of both stereocenters in the pheromone (Scheme 22).[62] Thus, straightforward manipulations transformed (−)-dihydrocarvone 159 into bromoester 161. DIBAL reduction of the ester yielded the corresponding alcohol with no epimerization at the adjacent chiral center. The authors initially attempted to alkylate the bromoalcohol with allylmagnesium chloride prepared from the typical 98% pure Mg sold for
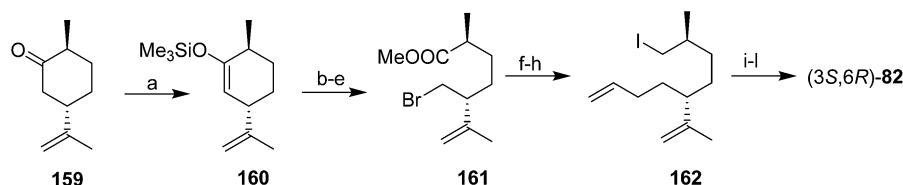
Scheme 22 Synthesis of (3S,6R)-82 from (−)-dihydrocarvone. Reagents and conditions: (a) LDA, Me₃SiCl, DME; (b) O₃, CH₂Cl₂/MeOH, then NaBH₄; (c) CH₂N₂, ether, 90% over 3 steps; (d) MesCl, Et₃N, ether; (e) Bu₄NBr, benzene; (f) DIBAL, heptane, 0 °C; (g) allyl MgCl, DME, reflux, 72%; (h) I₂, imidazole, Ph₃P, MeCN, 98%; (i) NaCN, DMF, 95%; (j) NaOH, EtOH/H₂O; (k) LiAlH₄, ether, 90% over 2 steps; (l) Ac₂O, pyridine, 94%.
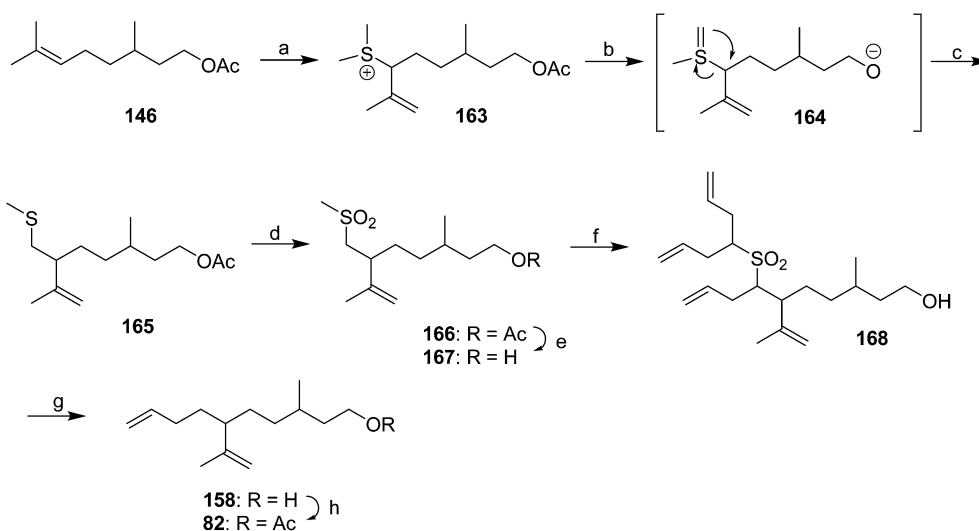
Scheme 23 Synthesis of racemic red scale diene component from citronellyl acetate. Reagents and conditions: (a) DMSO, (CF₃CO)₂O, CH₂Cl₂, −10 °C, 90%; (b) MeONa, MeOH, reflux, 59%; (c) reacetylate, conditions and yield not given; (d) m-CPBA, ether, −40 °C–room temp, 77%; (e) LiAlH₄, THF, −20 °C, 95%; (f) Excess BuLi, excess allyl bromide, THF/HMPA, −60 °C, 61%; (g) Na, NH₃/THF, 52%; (h) conditions and yield not stated.

1080 | Nat. Prod. Rep., 2015, 32, 1067–1113

This journal is © The Royal Society of Chemistry 2015

making Grignard reagents, with CuI catalysis, resulting in 65% of the desired coupling product plus 20% of a byproduct from transmetallation of the bromide. This byproduct was completely eliminated by using high purity Mg (99.8%) to prepare the Grignard reagent, and by carrying out the coupling in refluxing DME without the CuI catalyst. The synthesis was then completed by a straightforward sequence (conversion of the alcohol to iodide **162**, one-carbon chain extension with NaCN, hydrolysis to the acid, reduction, and acetylation). Alternatively, iodide **162** was chain extended by reaction with the anion of phenylmethylsulfide, followed by reaction with MeI, NaI, and DMF to give the chain-extended iodide, which was converted to acetate (3S,6R)-**82** by treatment with tetramethyl-ammonium acetate in DMF.

Another stereoselective synthesis used (R)-limonene as the source of the chiral isopropenyl group at C6, with the second chiral center at C3 being generated by an asymmetric Michael reaction, giving a product with a 9 : 1 d.r. (not shown).[63]

Dragan and coworkers used some interesting and unusual chemistry to devise a synthesis of the racemic pheromone from citronellyl acetate **146** (Scheme 23).[64] Thus, treatment of acetate **146** with DMSO and triflic anhydride gave allylic sulfide **163**, presumably via nucleophilic attack of the alkene on the activated acylsulfoxonium cation from DMSO. Treatment with strong base then resulted in a 1,2-Stevens rearrangement, possibly as shown in Scheme 23. Oxidation of the resulting sulfide **165** to the sulfone **166**, reduction of the acetate to the alcohol **167**, deprotonation with 5 equivalents of BuLi and reaction with 5 equivalents of allyl bromide gave the trialkylated product **168**. The large excesses were necessary in order to

produce a reasonable monoalkylation yield at the desired methylene position. Reductive removal of the sulfone and acetylation completed the synthesis.

A few additional modifications have appeared since 1989. For example, Zhang et al. stereoselectively dihydroxylated (S)-citronellyl N-phenylcarbamate with the fungus Aspergillus niger.[65] The resulting diol could be converted in several steps to the (3S,6S)-epoxide of citronellyl acetate, an intermediate used in previous syntheses (**150**, Scheme 21). Auer et al. developed a 14-step synthesis from (R)-(+)-limonene in which a key step was electrolysis of an iodide intermediate derived from limonene with a very large excess (160 equiv) of methyl crotonate.[66] The resulting 2 : 1 mixture of the desired product with a cyclic byproduct were only separable by HPLC, limiting the utility of the synthesis. Kefalas and Ragoussis reported a 3-step synthesis from (S)-citronellyl acetate **146** (Scheme 24),[67] using regiose-lective allylic oxidation, conversion of the resulting alcohol **169** to chloride **170**, and S_N2' alkylation of the chloride using 3-butenylmagnesium bromide with copper catalysis, giving the S_N2' (**82**) and S_N2 products in a 20 : 1 ratio. The stereochemistry at C6 was not controlled, but reports had indicated that male scale were attracted to both the (3S,6R)- and (3S,6S)-diastereo-mers, so this did not appear to be an issue for practical use.

Triene component of the California red scale pheromone. The first synthesis of the triene component of the red scale phero-mone **83** was designed to be flexible enough to produce all 4 stereoisomers (Scheme 25).[59] (R)- and (S)-carvone were chosen as suitable sources of the chiral center. Thus, epoxidation of (S)-carvone **171** followed by hydrolysis of the epoxide **172** and Pb(OAc)₄ oxidation gave ester acetal **174**. Treatment with
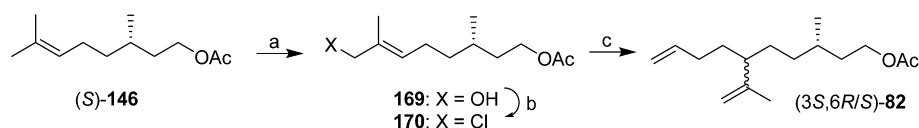
Scheme 24 3-step synthesis of technical grade diene **82**. Reagents and conditions: (a) SeO₂, t-BuOOH, then NaBH₄, 52%; (b) Ph₃P, CCl₄, 75%; (c) 3-butenylmagnesium bromide, CuCN·2LiCl, THF, −78 °C, 78%.
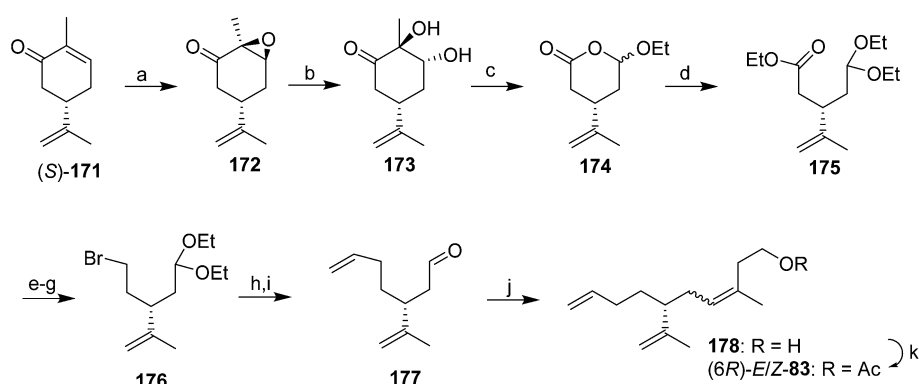
Scheme 25 First nonstereoselective synthesis of the triene component of red scale pheromone. Reagents and conditions: (a) H₂O₂, NaOH, MeOH, 77%; (b) HClO₄, THF/H₂O, 65 °C, 27%; (c) Pb(OAc)₄, EtOH/benzene; 98%; (d) HC(OEt)₃, PTSA, EtOH, 94%; (e) LiAlH₄, THF, 98%; (f) TosCl, pyridine, 90%; (g) NaBr, HMPA; 93% (h) CH₂=CHLi, THF, 78%; (i) PTSA, acetone/water, 96%; (j) Ph₃P=C(CH₃)CH₂CH₂OLi, THF, 49%, Z/E ~ 1 : 1; (k) Ac₂O, pyridine, 92%.

This journal is © The Royal Society of Chemistry 2015

*Nat. Prod. Rep.*, 2015, **32**, 1067–1113 | 1081

triethyl orthoformate and acid in EtOH opened the ring and gave a synthon **175** with differentiated ends. Reduction of the ester, conversion of the resulting alcohol to bromide **176**, and two-carbon chain extension with ethenyllithium and deprotection gave dienal **177**, which was subjected to Wittig reaction to complete the carbon skeleton, followed by acetylation. The (Z)- and (E)-isomers of **83** were separated by preparative GC, and the opposite enantiomers were generated from (R)-carvone. The (R,Z)-isomer matched the insect-produced compound.

Since that first synthesis, a number of other strategies have been explored, using synthons derived from the chiral pool (*e.g.*, enantiomers of carvone or limonene), or induction of asymmetry in achiral precursors. Control of the stereochemistry of the trisubstituted alkene also has been addressed by a number of different approaches.

In the first general strategy exploiting the chiral pool, dienal **177** or the corresponding alcohol have been used in several routes. For example, two routes used chiral aldehyde **177** generated from (R)-limonene as a key intermediate. In the first,

Wittig reaction between aldehyde **177** and the ylide from 3-hydroxy-1-methylpropyltriphenylphosphonium iodide, analogous to the route used by Roelofs *et al.*[59] gave a 67 : 33 mixture of the (Z)- and (E)-isomers of the penultimate alcohol in 75% yield (not shown).[63] The second route achieved excellent control over the alkene geometry, but at the cost of a considerably longer route (Scheme 26).[68] Thus, reaction between aldehyde **177** and silylimine **179** (generated in 4 steps from 1,4-butanediol in 63% yield) followed by hydrolysis gave α,β-unsaturated aldehyde **180** in an 87 : 13 ratio favoring the desired (E)-isomer, and the undesired (Z)-isomer was cleanly isomerized to the (E)-isomer by treatment of the mixture with pyridinium hydrochloride. After reduction of the aldehyde to the corresponding alcohol **181**, the synthesis was completed by one of two sequences in similar yields, *i.e.*, reductive removal of the benzyl protecting group, selective conversion of the allylic alcohol of **182** to the chloride, reduction of the chloride to the methyl group, and acetylation, or conversion of the allylic alcohol to a sulfate leaving group, reduction to the methyl, and simultaneous
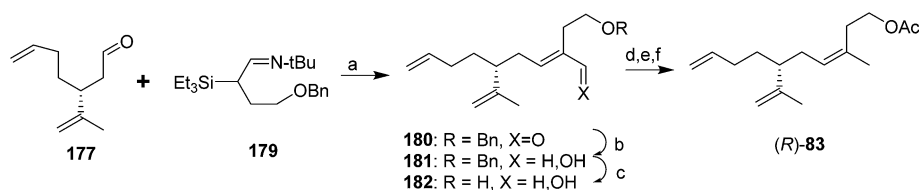
Scheme 26 Baudouy and Prince's route to (R)-**83**. Reagents and conditions: (a) s-BuLi, THF, −78−0 °C, then aq. oxalic acid; pyridinium hydrochloride, CH₂Cl₂, 66%; (b) LiAlH₄, ether, 100%; (c) Li/NH₃/ether, 88%; (d) NCS, Me₂S/CH₂Cl₂; (e) LiAlH₄, ether; 85% over 2 steps; (f) Ac₂O, pyridine, 94%.
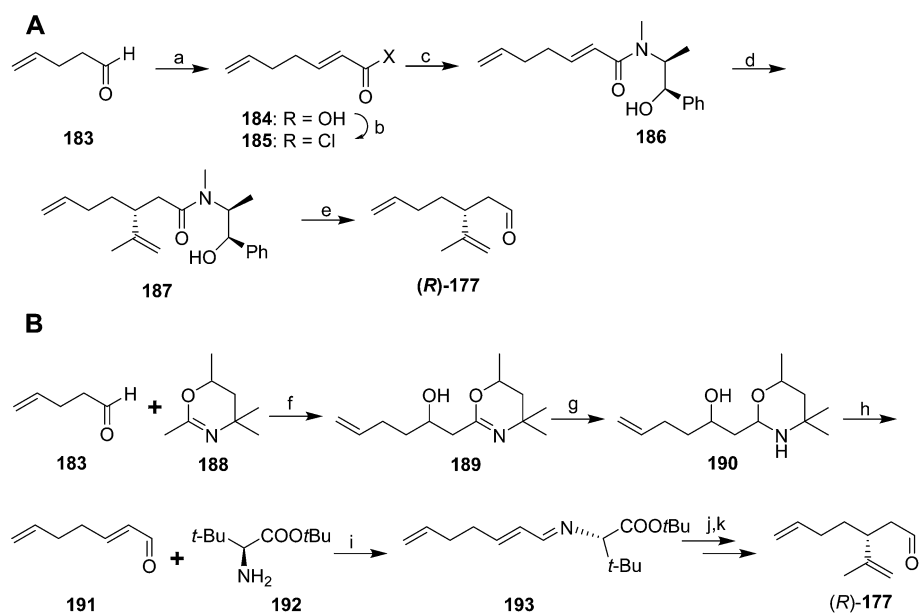
Scheme 27 Two routes to chiral aldehyde (R)-**177** based on asymmetric induction. Reagents and conditions: (a) malonic acid, pyridine, pyrrolidine, reflux, 79%; (b) SOCl₂, 0 °C–reflux, 94%; (c) (L)-ephedrine, proton sponge®, THF, 88%; (d) isopropenylmagnesium bromide, THF/ether, −60 to −20 °C, 51−69%; (e) DIBALH, hexane, then MeOH, H₃O⁺, 21%. (f) BuLi, THF, −78 °C to room temp, 83%; (g) NaBH₄, pH 6−8, 98%; (h) H₃O⁺, reflux, 45%; (i) t-butyl (S)-2-amino-3,3-dimethylbutyrate, benzene, 0 °C; (j) isopropenylmagnesium bromide, THF/ether, −70 to −15 °C; (k) H₃O⁺, 43% over 3 steps.

removal of the benzyl protecting group and acetylation with acetic anhydride with perchloric acid catalysis (not shown). Chiral dienal **177** also has been produced from camphor, but in a very long route (13 steps).[69]

In an interesting comparative study, Whittaker and coworkers used three different methods of asymmetric induction to generate key dienal **177** enantioselectively.[70] The first (Scheme 27A) began with two-carbon chain extension of 4-pentenal **183** with malonic acid and pyridine/pyrrolidine. Although the acid **184** was obtained as an E/Z mixture, the conditions used for conversion to the acid chloride **185** resulted in isomerization of the (Z)-isomer, producing pure (E)-2,6-heptadienoyl chloride **185**. Reaction with (L)-ephedrine in the presence of 1,8-bis(dimethylamino)naphthalene (proton sponge®) gave α,β-unsaturated amide **186**. 1,4-Addition of 2-propenylmagnesium bromide and cleavage of the amide with DIBAL gave the aldehyde (R)-**177**, with an unfortunate 21% yield in the cleavage step. The second route (Scheme 27B) proceeded via addition of lithiated 2,4,4,6-tetramethyl-5,6-dihydro-1,3-oxazine **188** to 4-pentenal **183**. Reduction of the oxazine **189** to the cyclic aminal **190** and acid hydrolysis gave 2,6-heptadienal **191** as the pure (E)-isomer. This route was selected from among

several tried because the others had problems with low yield, awkward byproducts, or incomplete control of the double bond stereochemistry. Reaction of enal **191** with t-butyl (S)-2-amino-3,3-dimethylbutyrate **192** gave conjugated imine **193**. Enantioselective addition of isopropenylmagnesium bromide created the chiral center, followed by hydrolysis of the imine to aldehyde (R)-**177**. A third route (not shown) used an oxazoline chiral auxiliary to successfully introduce the chiral center, but it proved impossible to remove the auxiliary group without degradation of the product.

In yet another variation, Normant's group formed a chiral acetal with a twofold axis of symmetry by transacetalization of (E)-2,6-heptadienal diethyl acetal with (2R,3R)-2,3-butanediol. 1,4-Addition of isopropenyl copper in the presence of LiBr, BF₃, and Bu₃P followed by acidic hydrolysis gave key aldehyde **177**, but in only 85% ee (not shown).[71]

Racemic dienal **177** has been generated by reaction of 4-pentenal **183** with the stabilized ylid from triethylphosphono-2-propionate, reduction to the alcohol **195**, Claisen orthoester rearrangement with triethylorthoacetate via **196**, reduction to the alcohol **198**, and PCC oxidation to dienal **177** (Scheme 28).[72]
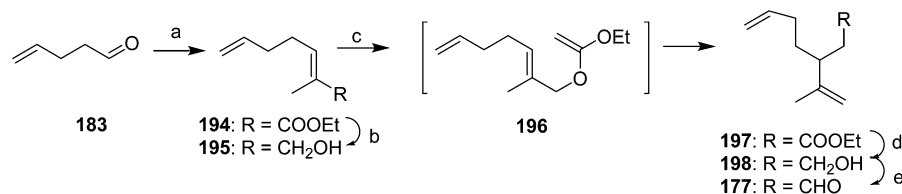
Scheme 28 Synthesis of racemic aldehyde **177** by Claisen rearrangement. Reagents and conditions: (a) triethyl phosphono-2-propionate, NaH, ether, 0 °C, 74%; (b) LiAlH₄, ether, 0 °C, 81%; (c) (EtO)₃CH, H⁺, 140 °C, 82%; (d) LiAlH₄, ether, 0 °C, 81%; (e) PCC, CH₂Cl₂, 55%.
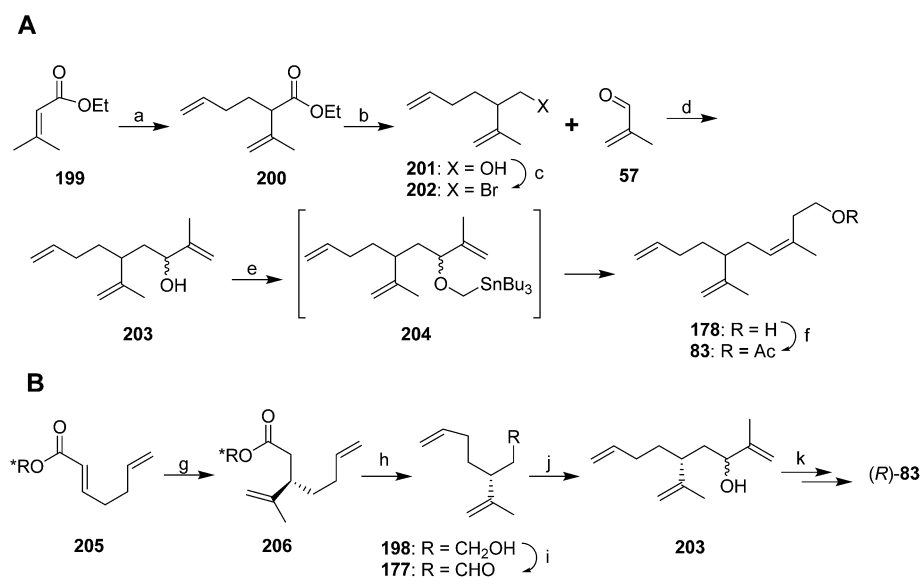
Scheme 29 Syntheses of racemic and chiral triene **83** based on 2,3-Wittig rearrangements. Reagents and conditions: (a) LDA, THF/HMPA, 1-bromo-3-butene, −78 °C, 80%; (b) LiAlH₄, solvent not given; (c) Ph₃P, NBS, solvent not given, 93% over 2 steps; (d) Mg, ether, −20 °C, 52%; (e) KH, Bu₃SnCH₂I, THF, then BuLi, −78 °C; (f) Ac₂O, pyridine, 83% over 2 steps. (g) CH₂=CH(Me)Cu·PBu₃, BF₃, −78–40 °C, >90%; (h) LiAlH₄, 96%; (i) (COCl)₂, DMSO, −78 °C, then Et₃N, 85%; (j) CH₂=CCH₃Li, ether, -78 °C, 85%; (k) steps, ∼71%.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1083**

A second group of strategies focused on controlling the stereochemistry of the trisubstituted double bond. In the first synthesis of this type, Still and Mitra developed a short and efficient synthesis in which the stereochemistry of the trisubstituted double bond was controlled by the geometry of the transition state of a 2,3-Wittig rearrangement (Scheme 29A).[73] Thus, ethyl 3,3-dimethacrylate **199** was deconjugatively alkylated at the α-position with 1-bromo-3-butene, and the ester **200** was reduced to the alcohol **201** which was converted to the corresponding bromide **202**. Reaction of the Grignard reagent from bromide **202** with methacrolein **57** gave allylic alcohol **203**, the anion of which was alkylated with iodomethyltributyltin to give **204**. Transmetallation with BuLi then induced a 2,3-sigmatropic rearrangement, giving the desired (Z)-isomer of the alcohol precursor **178** to the pheromone with 95–96% isomeric purity. The high stereoselectivity was suggested to be a consequence of the large R-group in the 5-membered transition state adopting a pseudoaxial orientation for steric reasons. The synthesis was completed by acetylation.

Oppolzer and Stevenson generated intermediate **203** by enantioselective alkylation of (E)-acid **205** esterified with a

Re-face directing chiral auxiliary (Scheme 29B).[74] Reaction with CH₂=CH(Me)Cu·PBu₃ and BF₃, followed by reduction to the alcohol **198**, oxidation to the aldehyde **177**, and reaction with 2-propenyllithium gave the key intermediate **203** from the Still and Mitra synthesis but with the chiral center fixed. Alcohol **203** was then carried through to the final product (R)-**83** (98.2% ee) via the (Z)-selective 2,3-sigmatropic rearrangement developed by Still and Mitra (see above). The key allylic alcohol **203** has also been generated in 14 steps from camphor.[69]

Cooke and Burman used a trisubstituted alkene synthon with the desired stereochemistry in place rather than trying to control the stereochemistry during reactions (Scheme 30).[75] Thus, spinning band distillation of dimethyl glutaconate gave the (Z)-isomer **207** in high purity, which was reduced to diol **208**. Regioselective halogenation and protection of the homoallylic alcohol gave key intermediate **209**. The second required intermediate, ylide **212**, was formed by acylation of stabilized triphenylphosphorane **211**, followed by elimination of HCl to give the terminal enone **212** as a methyl vinyl ketone equivalent. Sequential conjugate addition of allyllithium and alkylation of the resulting enolate **213** with bromide **209** inserted two of the
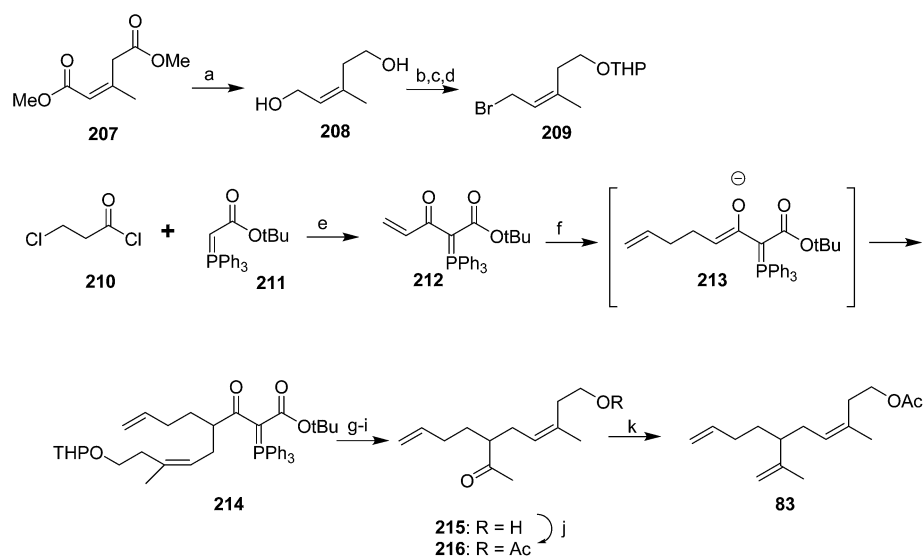
Scheme 30 Synthesis of triene **83** from a synthon with the double bond geometry already in place. Reagents and conditions: (a) AlH₃, solvent not given; (b) NCS, Me₂S; (c) dihydropyran, H⁺, 74%; (d) NaBr, N-methylpyrrolidinone, 72%; (e) benzene, 5–25 °C, 91%, then NaOMe, MeOH, 74%; (f) (allyl)₄Sn, BuLi, THF, −78–0 °C, then **212**, 0–27 °C, 87%; (g) AcOH, 90 °C, 86%; (h) K₂CO₃, MeOH, 97%; (i) aq. EtOH, pH 9–10, reflux, 84%; (j) Ac₂O, pyridine, 92%; (k) Ph₃PCH₃Br, BuLi, THF, 57%.
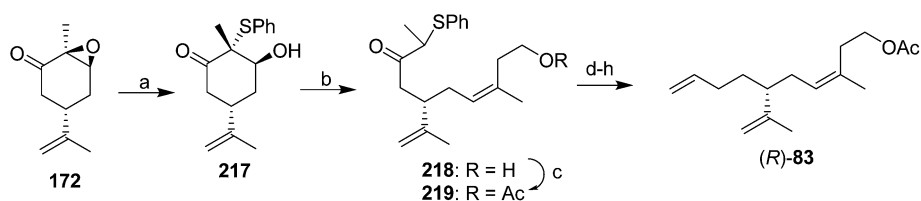
Scheme 31 Synthesis of triene (R)-**83** from a synthon with the double bond geometry already in place. Reagents and conditions: (a) PhSH, Et₃N, MeCN; (b) 3-hydroxy-1-methylpropyltriphenyl-phosphonium bromide, BuLi, THF/HMPA, 0 °C, 83%; (c) Ac₂O, pyridine, room temp; (d) m-CPBA, CH₂Cl₂, −5 °C, 85% over 2 steps; (e) 135 °C, 0.15 mm Hg; (f) NaBH₄, CeCl₃·7H₂O; (g) Ac₂O, pyridine, 52% over 3 steps; (h) HCO₂NH₄, PdCl₂·(PPh₃)₂, dioxane, reflux.

three alkenes. Decarbalkoxylation and simultaneous replacement of the THP with acetate by treatment with hot AcOH, followed by cleavage of the acetate with K_2CO_3 in MeOH and the ylide with aqueous ethanolic NaOH gave ketone **215**. Acetylation and Wittig reaction to install the terminal methylene completed the synthesis.

In their stereoselective synthesis of the trisubstituted alkene (*R*)-**83** (Scheme 31),[76] Caine and Crews regioselectively opened the epoxide ring of epoxidized (*S*)-carvone **172** with thiophenol, and under the conditions of a Wittig olefination, the resulting keto alcohol underwent an *in situ* retroaldol condensation, with the resulting aldehyde being captured by the ylide from 3-hydroxy-1-methylpropyltriphenylphosphonium bromide. The high (*Z*)-selectivity was attributed to unfavorable electrostatic interactions between three negatively charged groups in the betaine intermediate that would lead to the (*E*)-isomer. Although the stereochemistry of the trisubstituted alkene was controlled, 5 further steps were required to remove the keto and sulfide groups and place the terminal alkene, the final product was contaminated with 11% of the isomer with the terminal double bond moved in one position, and yields were not given for all steps, so the efficiency of the synthesis is unclear.

Celebuski and Rosenblum exploited the ability of (η^1-allyl) ferrocenyliron complexes to undergo electrophilic addition to form a dihapto iron-olefin cation, followed by regio- and stereoselective addition of a nucleophile, to simultaneously install both the trisubstituted double bond and one of the two terminal double bonds (Scheme 32).[77] Thus, reaction of allylic chloride

221, prepared in one step from commercial diene **220**, with $C_5H_5Fe(CO)_2Na$ (FpNa, prepared from Fp_2 by reduction with Na/Hg amalgam) gave allylic organometallic **222** *via* S_N2' and/or spontaneous rearrangement of the product from S_N1 reaction. The second synthon was prepared in two steps from 3,6-dihydro-4-methyl-2*H*-pyran **223**, by ring-opening with acetyl chloride catalyzed by Zeise's salt, and conversion of the resulting acetoxychloride **224** to the iodide **225** with NaI in acetone. Fortuitously, stirring the 1 : 1 mixture of (*E*/*Z*)-isomers of **225** with the Fp-diene complex **222** in nitromethane for several days resulted in exclusive production of the desired (*Z*)-isomer of the pheromone because only the (*Z*)-isomer of iodide **225** reacted with the complex, and the iodide apparently isomerized under the reaction conditions so that as the (*Z*)-isomer was consumed, it was replaced by continuing isomerization of the (*E*)-isomer. The specificity of the alkylation reaction was proposed to be due to anchimeric assistance to ionization by the acetoxy group, which would be possible with the (*Z*)- but not the (*E*)-isomer of iodide **225**.

McCullough *et al.* cleverly manipulated the regiochemistry of alkylation of allyl anion intermediates in their synthesis of racemic **83** (Scheme 33).[46] Thus, deprotonation of methallyl phenyl sulfide and regioselective α-alkylation of the anion **226** with 1-bromo-3-butene gave **227**. Reductive lithiation with lithium 4,4'-di(*tert*-butyl)biphenyl (LDBB), transmetallation with titanium, and reaction with formaldehyde resulted in regioselective alkylation at the most substituted terminus of the metallated intermediate. Conversion of the resulting alcohol
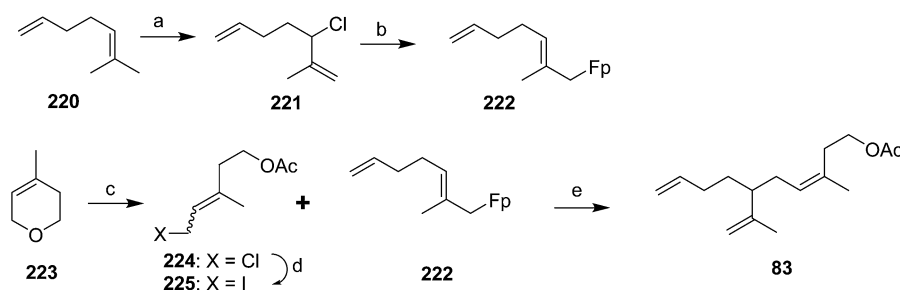
Scheme 32 Synthesis of racemic triene **83** from a synthon with the double bond geometry already in place. Reagents and conditions: (a) $Ca(OCl)_2$, CH_2Cl_2/H_2O, 73%; (b) Fp_2, Na/Hg, DME, 57%; (c) AcCl, toluene, $K[PtCl_3(C_2H_4)]$, 75%; (d) NaI, acetone, no yield reported; (e) **225** + **222**, nitromethane, room temp, 42%.
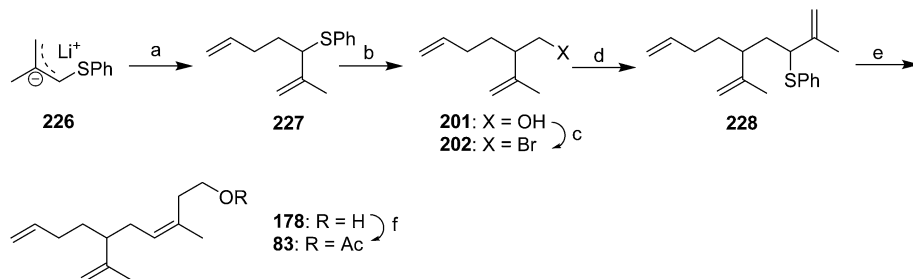
Scheme 33 Synthesis of racemic triene **83** by regiocontrol of alkylation. Reagents and conditions: (a) 1-bromo-3-butene, *s*-BuLi, THF, −78 °C, 85%; (b) lithium 4,4'-(di-*t*-butyl)biphenyl (LDBB), THF, −78 °C, then Ti(*i*-PrO)$_4$, then CH_2O, 79%; (c) CBr_4, PPh_3, MeCN, 81%; (d) methallyl phenyl sulfide, *s*-BuLi, THF, −78 °C, 78%; (e) LDBB, THF −78 °C, then $CeCl_3$, then CH_2O; (f) Ac_2O, pyridine, 54%.

201 to bromide **202**, followed by another regioselective alkylation with lithiated methallyl phenyl sulfide **226** at the α-position gave allylic sulfide **228**. Reductive lithiation with LDBB, transmetallation with CeCl₃, and reaction with formaldehyde resulted in both regioselective alkylation at the least substituted terminus this time, and high selectivity for the (Z)-alkene, due to the configuration of the anion remaining fixed by complexation with cerium. Acetylation completed the synthesis.

Vasil'ev *et al.* addressed the alkene stereochemistry problem with a (Z)-selective, 1,4-dihydrogenation of a conjugated diene precursor (Scheme 34).[78] Thus, aldehyde **229**, generated in ~34% yield in 4 steps from ethyl acetoacetate, subjected to Wadsworth–Horner–Emmons olefination gave triene **230** in ~79% yield, as a 65 : 35 mixture of (2E,4E)- and (2Z,4E) isomers. 1,4-Dihydrogenation of this mixture with a methyl benzoate-chromium tricarbonyl catalyst gave the desired (3Z)-alkene **231** as the sole product, which was carried through to the pheromone **83** in several straightforward steps.

Most recently, Hesse *et al.* constructed the trisubstituted alkene with the desired (Z)-stereochemistry using stereoselective protodeboronation and conversion of an (E)-allylic boronic ester into a (Z)-trisubstituted alkene (Scheme 35).[79] Thus, vinylboronate ester **234** was prepared in two steps from 2-butyn-1-ol **232** by TBDMS-protection of the alcohol followed by copper-catalyzed hydroboration with pinacolatoborane. Deprotonation of the 2,4,6-triisopropylbenzoate ester **235** (prepared in 4 steps and 63% yield from (R)-limonene) and reaction with the boronic ester **234** gave the expected ate complex, which upon heating rearranged to the allylic intermediate **236**. Treatment with TBAF·3H₂O and heating resulted in sequential removal of the protecting group and protodeboronation to give alcohol **178**, with >20 : 1 (Z/E) selectivity. The stereoselectivity was

attributed to steric interactions disfavoring the (E)-isomer in the proposed six-membered ring transition state during protodeboronation. Acetylation completed the synthesis.

Pheromone of white peach scale. The pheromone of the white peach scale was originally identified as propionate ester (R)-**84**, using a combination of liquid and preparative gas chromatography to isolate the active compound, and spectroscopic data with microchemical tests to identify it.[80] However, there may be different pheromone races or possibly even cryptic species of white peach scale. Specifically, populations from France were found to produce the corresponding alcohol **241** in larger quantities than the propanoate ester, and males from this population required both compounds for optimal response.[81] In contrast, Florida males responded best to the propanoate alone, and responses were decreased by higher doses of the alcohol in combination with the ester.[82] Unfortunately, the original paper describing the identification of the propanoate from the Florida population did not mention whether the alcohol was present in the aeration extracts.

The core structure of the white peach scale pheromone alcohol **241** and ester **84** differs from that of the triene ester component of the red scale pheromone **83** by a single methyl group on carbon 9, and so the syntheses of white peach scale pheromone usually involved minor modifications to those routes. In fact, the initial syntheses were somewhat easier than those for red scale pheromone because the product from ozonolysis of limonene was perfectly set up for straightforward elaboration to the desired structure. Thus, limonene **237** was ozonized in MeOH with a trace of acid and quenching with Me₂S, producing protected ketoaldehyde **238** in one step (Scheme 36A).[80] Wittig reaction to convert the ketone to a terminal methylene, deprotection of the aldehyde, and a second
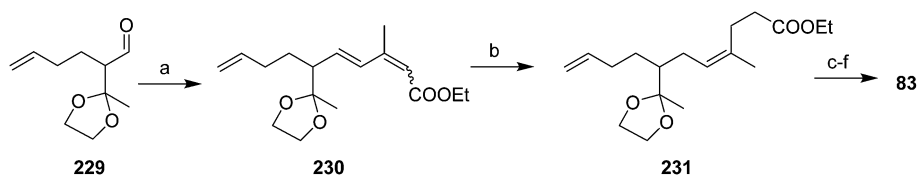
Scheme 34 Synthesis of racemic triene **83** by a (Z)-selective, 1,4-dihydrogenation. Reagents and conditions: (a) triethyl 3-methyl-4-phosphonocrotonate, KOH, 18-crown-6, benzene, 69%; (b) H₂, 80 atm, (PhCOOMe)Cr(CO)₃, acetone, 120 °C, 52%; (c) LiAlH₄, ether; (d) oxalic acid, H₂O/acetone, reflux, 52% over 2 steps; (e) Ph₃P=CH₂, THF/DMSO; (f) Ac₂O, Na₂CO₃, 100 °C, 45% over 2 steps.
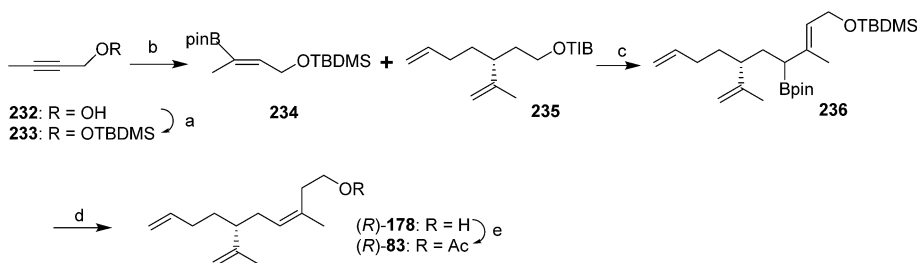
Scheme 35 Synthesis of triene (R)-**83** from a synthon with the double bond geometry already in place. Reagents and conditions: (a) TBDMSiCl, imidazole, DMAP, CH₂Cl₂, 99%; (b) CuCl (5%), PPh₃ (6%), t-BuOK (20%), B₂pin₂, MeOH/THF, room temp; (c) s-BuLi, TMEDA, Et₂O, −78 °C-reflux; (d) TBAF·3H₂O, room temp-45 °C, 73% over 2 steps; (e) Ac₂O, pyridine, 99%.

1086 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

Wittig reaction with the lithium salt of 3-triphenylphosphor-anylidene-1-butanol completed the carbon skeleton, and the (E)- and (Z)-isomers were separated by HPLC before final esterification with propanoic anhydride. Thus, by using both enantiomers of limonene as starting materials, all possible isomers were generated, and the insect-produced compound was determined to be the (Z)-isomer by GC, and the (R)-enantiomer via bioassays.

In a significant improvement to this route, generation of Ph₃P=C(CH₃)CH₂CH₂OLi by reaction of ethylidenetriphenylphosphorane with ethylene oxide followed by deprotonation, as opposed to generation of the phosphoranylidene by reaction of the lithium salt of 3-triphenylphosphoranylidene-1-propanol with MeI gave a much cleaner and higher yielding Wittig reaction.[83] In an interesting final modification, the authors discovered that deprotonation of the betaine **242** initially formed by reaction between ethylidenetriphenylphosphorane and aldehyde **240**, followed by reaction with ethylene oxide and elimination of triphenylphosphine oxide, gave 98% (Z)-selectivity, albeit in low (15%) yield (Scheme 36B).[83] Because male scale appeared to be indifferent to the (E)-isomer, a recently filed patent used essentially the same route to prepare the pheromone as blends of the (E)- and (Z)-isomers for field use.[84]

The synthesis of Vasil'ev et al. was identical to their synthesis of the red scale pheromone (See Scheme 34) with the exception that 1-bromo-3-methyl-3-butene was used to make the protected ketoaldehyde starting material in place of 1-bromo-3-butene.[78]

Pheromone of the scale Aulacapis murrayae. Ho and coworkers very recently identified the pheromone of *A. murrayae* as the (R)-enantiomer of the degraded irregular terpenoid solanone.[85] This compound was originally identified from tobacco, and

several syntheses of the racemate have been published.[86] However, only chiral syntheses will be described because male scales are inhibited by the (S)-enantiomer.[85]

In a communication on a method for regio- and stereo-selective preparation of conjugated dienes from allylic alcohols, Mori and coworkers prepared racemic solanone, and multigram quantities of a precursor **245** that could be transformed into (S)-solanone in two easy steps (Scheme 37).[87] However, the paper did not describe how the precursor **244** was prepared, nor, surprisingly, did it report carrying the precursor through to (S)-solanone (S)-**85**. Thus, the utility of this synthesis is hard to assess, nor is it clear whether it could be readily adapted to the synthesis of the (R)-enantiomer.

The only published synthesis of (R)-solanone **85** began with asymmetric induction using Evans' chiral oxazolidinone chemistry to place the chiral center (Scheme 38).[85] Dihydroxy-lation of the intermediate **246** resulted in spontaneous lac-tonization to the γ-lactone, followed by acetylation to give **247**. Reduction of the ester and lactone simultaneously, and selective formation of the acetonide from the vicinal diol gave **248**. Swern oxidation of the primary alcohol followed by Horner–Wadsworth–Emmons olefination with diethyl-(2-methylallyl)phosphonate gave the 1,3-diene **249** as one stereoisomer. One-pot cleavage of the acetonide and oxidative cleavage of the resulting diol gave an aldehyde which was reduced to alcohol **250** and converted to the tosylate. Displacement of the tosylate with cyanide, followed by reaction of the resulting nitrile **251** with MeMgBr and aqueous workup gave a few milligrams of (R)-solanone (R)-**85**, suffi-cient for bioassays to verify its bioactivity.
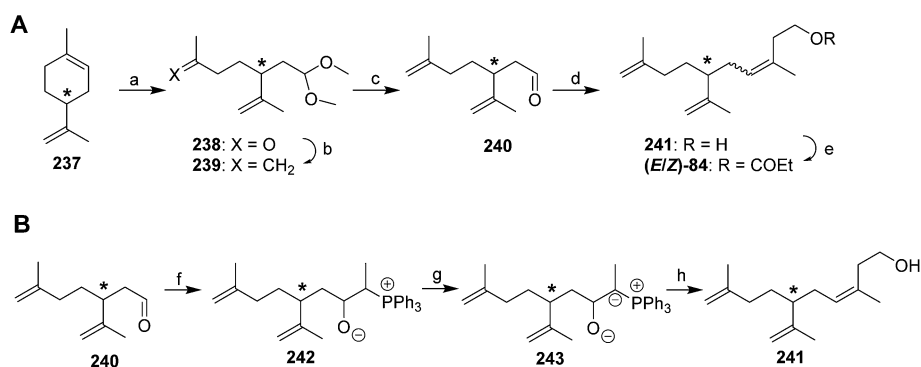
Scheme 36 Syntheses of white peach scale pheromone enantiomers from either enantiomer of limonene. Reagents and conditions: (a) O₃, MeOH, Me₂S, H⁺, 80%; (b) Ph₃P=CH₂, THF, 75%; (c) PTSA, acetone/H₂O, 96%; (d) Ph₃P=C(CH₃)CH₂CH₂OLi, THF, 48%; (e) (EtCO)₂O, pyridine, 90%. (f) BuLi, THF, Ph₃PEtBr, −78 °C (g) BuLi, −78 °C; (h) ethylene oxide, −5 °C, 15%.
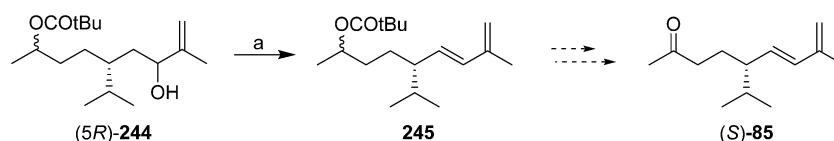
Scheme 37 Mori and coworkers' partially described synthesis of (S)-solanone. Reagents and conditions: (a) MesCl, Et₃N, CH₂Cl₂, then diiso-propylethylamine, HMPA, 140–180 °C, 64%.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1087**
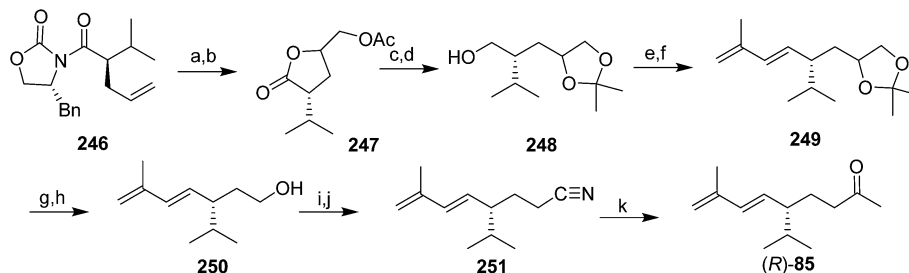
Scheme 38 Synthesis of (R)-solanone **85**. Reagents and conditions: (a) OsO₄, NMO, MeCN, room temp; (b) Ac₂O, pyridine, DMAP, 0 °C, 98% over 2 steps; (c) LiBH₄, THF, room temp; (d) 2,2-dimethoxypropane, H⁺, DMF, room temp, 84% over 2 steps; (e) (COCl)₂, DMSO, CH₂Cl₂, −78 °C, then Et₃N; (f) diethyl (2-methylallyl)phosphonate, BuLi, THF, HMPA, −78 °C-room temp, 84% over 2 steps; (g) NaIO₄, AcOH, H₂O, room temp; (h) NaBH₄, MeOH, 0 °C, 86% over 2 steps; (i) TosCl, Et₃N, DMAP, CH₂Cl₂, room temp; (j) NaCN, DMSO, room temp, 73% over 2 steps; (k) CH₃MgBr, toluene, reflux, then H₂O, 53%.

3.2 Cyclic pheromones

3.2.1 Cyclopropane-containing pheromones: esters of chrysanthemol.
To date, two mealybug pheromones with cyclopropane rings have been identified, both of which are esters of (1R,3R)-chrysanthemol (Fig. 4).

Madeira mealybug pheromone. The sex pheromone of the Madeira mealybug, *Phenacoccus madeirensis* Green, was identified by Ho and coworkers as (1R,3R)-[2,2-dimethyl-3-(2-methyl-prop-1-enyl)cyclopropyl]methyl (R)-2-methylbutanoate [(1R,3R)-chrysanthemyl (R)-2-methylbutanoate] **252**.[88] The absolute configuration was determined by comparison of retention times of the acid and alcohol portions with those of enantiopure standards on a chiral column. (1R,3R)- and (1S,3S)-Chrysanthemol were obtained by resolution of racemic *trans*-chrysanthemol by lipase-catalyzed acylation with succinic anhydride.[26] 2-Methylbutanoic acid was resolved by formation of diastereomeric amides with (S)-α-methylbenzylamine, separation by semipreparative HPLC, and hydrolysis of the separated diastereomers in refluxing 70% H₂SO₄, without epimerization. (R)-Lavandulyl (R)-2-methylbutanoate was identified as a minor component in extracts from virgin females, but it was not attractive to males, nor did it act synergistically or additively with the main component **252**.

Ho and coworkers also studied the relationship between chirality and bioactivity of the stereoisomers of **252** (Fig. 5).[89] The insect-produced (1R,3R,2′R)-isomer was most attractive, whereas the (1R,3R,2′S)-diastereomer (opposite stereochemistry in the acid portion) was inhibitory. In contrast, the (1S,3S,2′R)-isomer (opposite stereochemistry in the alcohol portion) was neither attractive nor inhibitory. The diastereomers prepared from the *cis*-alcohol (1R,3S,2′R and 1S,3R,2′R) were also inhibitory.

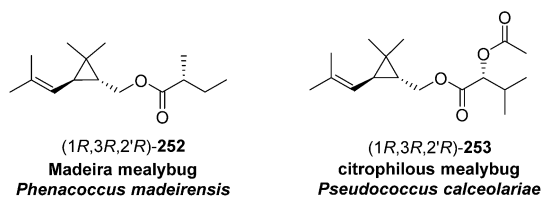*Citrophilous mealybug pheromone.* The sex pheromone of the citrophilous mealybug, *Pseudococcus calceolariae*, was identified jointly by New Zealand and Chilean teams as [2,2-dimethyl-3-(2-methylprop-1-enyl)cyclopropyl]methyl 2-acetoxy-3-methyl-butanoate (chrysanthemyl 2-acetoxy-3-methylbutanoate **253**, and readily synthesized by esterification of *trans*-chrysanthemol with 2-acetoxy-3-methylbutyryl chloride.[90] Unelius and coworkers subsequently determined the absolute configuration to be (1R,3R)-chrysanthemyl (R)-2-acetoxy-3-methylbutanoate by comparison of the retention times of the acid and alcohol portions with synthetic standards on a chiral column.[91] The correct stereoisomer of the pheromone was readily prepared from (1R,3R)-chrysanthemol (from LiAlH₄ reduction of commercial (1R,3R)-chrysanthemic acid) and (R)-2-acetoxy-3-methylbutyryl chloride (from acetylation of the hydroxyacid with Ac₂O and pyridine, then conversion to the acid chloride with oxalyl chloride). The (R)-configuration in the acid moiety was necessary for activity. The non-natural stereoisomers were at most slightly inhibitory, so that relatively inexpensive racemic chrysanthemol could be used for commercial synthesis of the pheromone.

3.2.2 Cyclobutane-containing pheromones: esters of the monoterpenols planococcyl alcohol and maconelliol.
Four pheromones with a cyclobutane core structure have been identified from three mealybug and one scale species (Fig. 6).

Citrus mealybug pheromone. The sex pheromone of the citrus mealybug, *Planococcus citri* (Risso), was identified by Bierl-Leonhardt and coworkers as (1R,3R)-3-isopropenyl-2,2-dime-thylcyclobutylmethyl acetate [(1R,3R)-planococcyl acetate] **254**.[92] The natural enantiomer and the racemate were equally active,
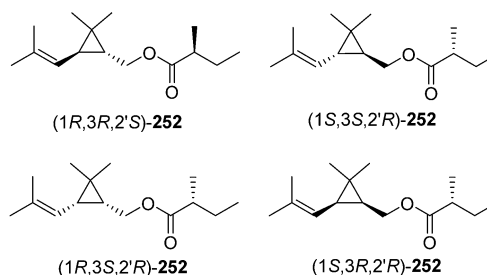

Fig. 4 Cyclopropane-containing pheromones (esters of chrysanthemol).

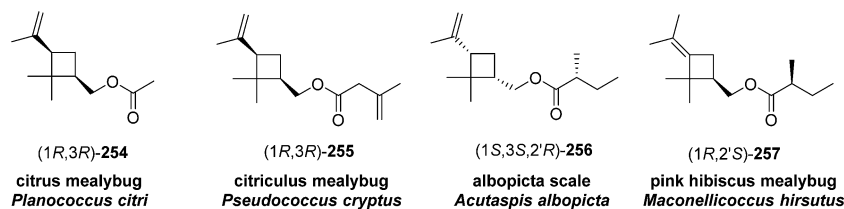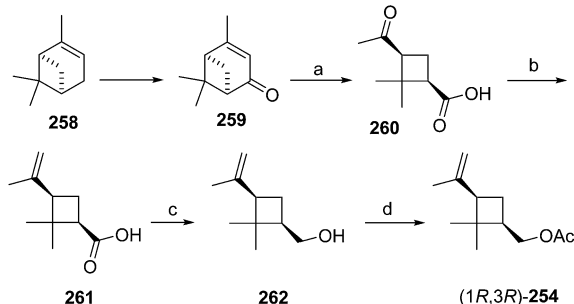**Fig. 5** Stereoisomers of Madeira mealybug pheromone.

Fig. 6 Scale and mealybug pheromones with a cyclobutane-containing monoterpenol core.

indicating no inhibition by the unnatural (1*S*,3*S*)-enantiomer.[93] The *trans* isomers were much less active than **254**.[92] Several syntheses have been reported, most of which used (+)-α-pinene **258** as an obvious starting material because it contains the required cyclobutane skeleton, and precursors to all the functional groups with the correct substitution pattern and stereochemistry. Thus, (+)-α-pinene **258**, or (*R*)-(+)-verbenone **259** prepared by allylic oxidation of (+)-α-pinene, were subjected to oxidative ring opening. Syntheses reported before 1989 have been thoroughly reviewed[2] and will not be discussed further here.

Scheme 39 summarizes Passaro's synthesis of (*R*)-**254**,[94] which incorporated two major improvements to a previous route.[95] Specifically, the crucial step of ruthenium-catalyzed oxidative ring opening with concomitant decarboxylation of (*R*)-(+)-verbenone **259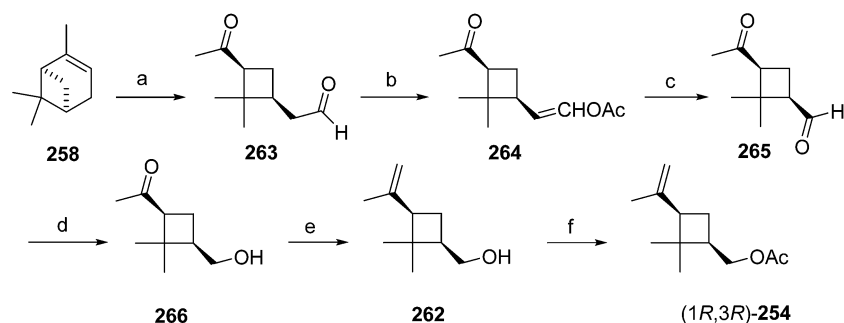** was carried out in aqueous *tert*-butanol instead of $CCl_4 : CH_3CN : H_2O$ (2 : 1 : 4). This modified procedure afforded (+)-*cis*-pinononic acid **260** in excellent yield (94%), was easier to perform, and simplified product isolation. Secondly, methylenation of **260** using $Zn/CH_2Br_2/TiCl_4$ (ref. 96) was superior to Wittig reactions in larger scale preparations. $LiAlH_4$ reduction of acid **261** and acetylation of the resulting alcohol **262** gave the desired pheromone **254** in >40% overall yield from verbenone.

Kukovinets *et al.* used essentially the same route and intermediates, but slightly different reagents, for example, employing ozonolysis for the oxidative cleavage of the ring.[97]

Another route began with direct oxidative cleavage of the ring in (+)-α-pinene **258** by ozonolysis to give ketoaldehyde **263** (Scheme 40).[93] Selective conversion of the aldehyde to enol acetate **264** followed by a second ozonolysis removed one carbon, and chemoselective reduction[98] of the aldehyde of **265** with $Zn(BH_4)_2$ in mixed ether solvents gave pinononic alcohol **266**, which was subjected to Wittig reaction and acetylation to afford the pheromone **254**. All intermediates were used as crude materials, and only the final product was purified by chromatography to provide technical grade (85% pure) pheromone, which proved to be as active as the pure pheromone in field trials.

Citriculus mealybug pheromone. The sex pheromone of the citriculus mealybug, *Pseudococcus cryptus* Hempel, was identified by Arai and coworkers as (1*R*,3*R*)-3-isopropenyl-2,2-dimethylcyclobutylmethyl 3-methyl-3-butenoate **255** [[(1*R*,3*R*)-planococcyl 3-methyl-3-butenoate].[99] The alcohol portion was identical to that of the citrus mealybug pheromone.

Scheme 41 summarizes Nakahata's 5-step synthesis that utilized all ten carbon atoms of the starting material (+)-α-pinene **258**.[100] Thus, treatment of (+)-α-pinene with $Pb(OAc)_4$

Scheme 39 Passaro's synthesis of the citrus mealybug pheromone (1*R*,3*R*)-**254** from (+)-α-pinene **258**. Reagents and conditions: (a) RuCl₃, NaIO₄, *t*-BuOH, H₂O, 94%; (b) Zn, CH₂Br₂, TiCl₄; (c) LiAlH₄, Et₂O; (d) Ac₂O, pyridine, 44% for 3 steps.

Scheme 40 Zada's synthesis of the citrus mealybug pheromone (1*R*,3*R*)-**254** from (+)-α-pinene **258**. Reagents and conditions: (a) O₃, NaHCO₃ (cat.), then Me₂S, 78%; (b) Ac₂O, Et₃N, DMAP, 100% (*E*−*Z* = 1 : 1); (c) O₃, NaHCO₃ (cat.), then Me₂S, 86%; (d) Zn(BH₄)₂, DME, Et₂O, THF; (e) Ph₃P═CH₂, THF; (f) Ac₂O, pyridine, 35% for 3 steps.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1089**
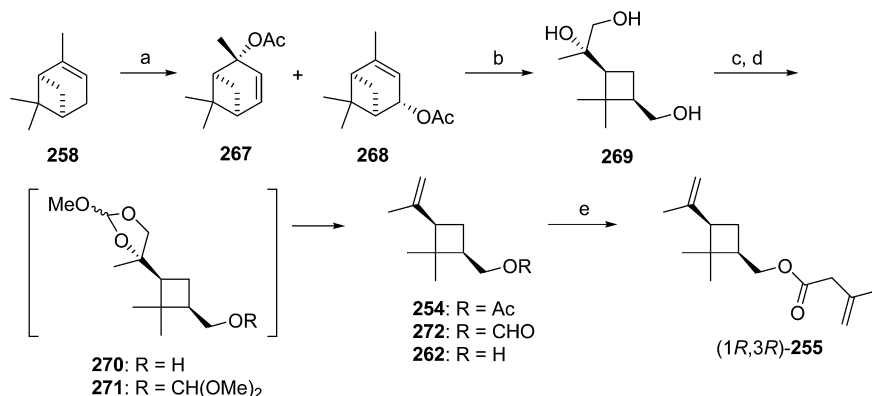
Scheme 41 Synthesis of the citriculus mealybug pheromone (1R,3R)-255 from (+)-α-pinene 258. Reagents and conditions: (a) Pb(OAc)$_4$, C$_6$H$_6$, 75%, 267 : 268 = 95 : 5; (b) O$_3$, MeOH, then NaBH$_4$, and finally K$_2$CO$_3$, 72%; (c) HC(OMe)$_3$, PPTS, THF; (d) Ac$_2$O, then aq. NaOH, 88% for 2 steps; (e) 3-methyl-3-butenoic acid, DCC, DMAP, 91%.

gave 267 with a small amount of allylic rearrangement product 268 (95 : 5 ratio). The order of addition and careful drying of the Pb(OAc)$_4$ were crucial for minimizing the formation of 268. Ozonolysis with reductive workup, followed by methanolysis of the acetate gave triol 269. 269 was converted into a mixture of acetate 254 and formate 272 (3 : 4 ratio) by conversion of the vicinal diols into olefins via 2-methoxy-1,3-dioxolane derivatives 270 and 271 and subsequent treatment with Ac$_2$O.[101] The mixture of 254 and 272 was hydrolyzed to alcohol 262 in a one-pot operation. The alcohol 262 was esterified with 3-methyl-3-butenoic acid in the presence of DCC and DMAP. The use of only a catalytic amount of DMAP (about 0.1 equivalent) was essential for preventing migration of the double bond of the acid portion into conjugation.

Albopicta scale pheromone. The pheromone of the invasive scale *Acutaspis albopicta* was recently identified as (1S,3S)-3-isopropenyl-2,2-dimethylcyclobutylmethyl (R)-2-methyl-butanoate 256, in which the alcohol portion is the enantiomer of that found in the pheromones of the citrus and citriculus mealybugs.[102] The alcohol portion was synthesized from (S)-(−)-verbenone by the method of Passaro et al. (see Scheme 39),[94] the acid portion was made by kinetic resolution of 2-methyl-butanol with Amano lipase PF, followed by Jones oxidation of the resolved alcohols, and the ester linkage was made by

treatment of a CH$_2$Cl$_2$ solution of the acid and alcohol portions with the coupling reagent 1-ethyl-3-[(3-dimethylamino)propyl] carbodiimide hydrochloride (EDC) and DMAP.[102] The other stereoisomers were made by synthesis of the enantiomeric alcohol portion from (R)-(+)-verbenone, and esterifications with commercially available (S)-2-methylbutanoic acid or (R)-2-methylbutanoic prepared as described above. Bioassays of all 4 stereoisomers determined that the absolute configuration of the monoterpenol portion was critical, with the (1R,3R,2′R) and (1R,3R,2′S) diastereomers being inactive, whereas the (1S,3S)-alcohol esterified with either enantiomer of 2-methylbutanoic acid were equally active, indicating that the male scale did not discriminate the absolute configuration of the acid portion of the pheromone.
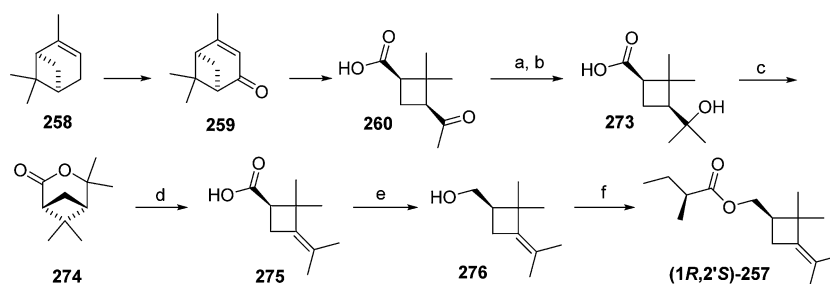
Pink hibiscus mealybug pheromone (major component). As mentioned earlier, the sex pheromone of the pink hibiscus mealybug, *Maconellicoccus hirsutus* (Green), was identified by Zhang and coworkers as a 1 : 5 mixture of (R)-lavandulyl (S)-2-methylbutanoate 37 and (R)-maconelliyl (S)-2-methylbutanoate 257.[29] Maconelliol 276, the alcohol portion of 257, is structurally similar to the planococcyl alcohol portion of the citrus and citriculus mealybug pheromones, with a methylethylidene group on the ring instead of an isopropenyl group. Zhang et al. developed an enantioselective synthesis of 257 (Scheme 42)[103]

Scheme 42 Synthesis of (1R,2′S)-257, the major component of the pink hibiscus mealybug pheromone from (+)-α-pinene 258. Reagents and conditions: (a) MeLi/Et$_2$O, 1.0 equiv, THF, −20 °C, 15 min; (b) MeMgCl/THF, 1.6 equiv, −10 °C, 30 min, room temp, 1 h, 89%; (c) POCl$_3$, pyridine, room temp, 24 h, 75%; (d) PTSA, C$_6$H$_6$, reflux, 24 h, 78%; (e) LiAlH$_4$, Et$_2$O, room temp, overnight, 88%; (f) (S)-2-methylbutanoic acid, (COCl)$_2$, DMF, C$_6$H$_6$, room temp, 1.5 h, 79%.

starting from (+)-*cis*-pinononic acid **260**, a key intermediate in the citrus mealybug pheromone synthesis (see Scheme 39). Reaction of **260** (94% ee) with 3 equivalents of MeMgCl resulted in partial loss of enantiomeric purity (80% ee) due to double epimerization during nucleophilic addition. However, reaction with 1 equivalent of MeLi followed by 1.6 equivalents of MeMgCl gave tertiary alcohol **273** with no loss of chirality. Thinking to prevent decarboxylation during dehydration, **273** was converted to its methyl ester, but when the ester was subjected to conventional dehydration methods, the desired methylethylidene product was contaminated with the isopropenyl isomer, which was difficult to separate. However, treatment of the free acid **273** under the same conditions resulted in lactone **274** as the only product, which was cleanly converted to the desired methylethylidene product **275** by treatment with acid. LiAlH$_4$ reduction and condensation of the resulting alcohol **276** with (*S*)-2-methylbutanoic acid gave the desired ester (1*R*,2′*S*)-**257** in good yield.

Zhang and coworkers carried out a systematic study of the relationship between chirality and biological activity of the stereoisomers of **257**.[104] Their results suggested that the

stereochemistry of the acid portion is more critical than that of the alcohol portion. Thus, the (*R*)-alcohol esterified with the (*S*)-acid was attractive, whereas the (*R*,*R*)-diastereomer was inhibitory. The chiral center in the alcohol portion was less important to attraction or inhibition. Importantly, this was the first report of enantiomeric antagonism in any mealybug or scale insects. Furthermore, optimal attraction was critically dependent on dose, with doses above 10 μg resulting in decreased attraction.

Ester of a cyclobutane-containing sesquiterpenol: oleander scale pheromone. The oleander scale pheromone is the only example so far of a scale or mealybug pheromone incorporating a sesquiterpenoid cyclobutane structure. Its identification required the integration of fragments of information from a variety of analysis methods.[105] High resolution tandem MS provided the molecular formula (C$_{17}$H$_{28}$O$_2$) and the formulae of a number of fragments obtained from the parent molecule and some derivatives. Microchemical reactions showed the presence of two C=C double bonds, one ring, and an acetate ester, but this still did not allow determination of the complete structure. Subsequently, ^{1}H and ^{13}C spectra acquired on 900 μg of pheromone accumulated over several years yielded the basic structure but not the relative and absolute stereochemistries, nor were the authors successful in preparing crystalline derivatives. Thus, the final structure was proven by synthesis of isomers of known configuration, and bioassays of the two possible enantiomers showed that the insect-produced compound likely had the (1*R*,2*S*)-configuration (Fig. 7).

The goal of the first synthetic effort was to develop stereoselective syntheses of all four possible stereoisomers with known absolute configurations, in large enough amounts to
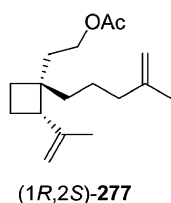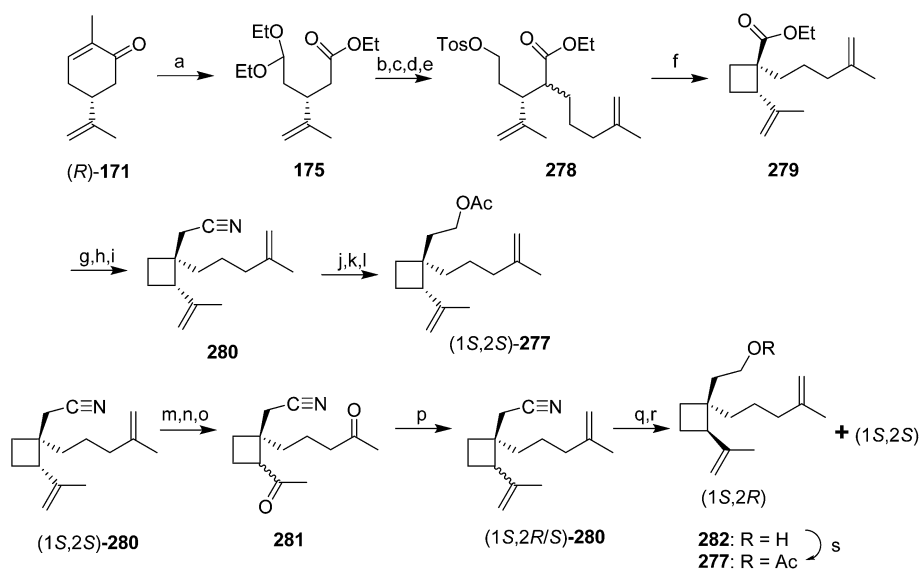

(1*R*,2*S*)-**277**
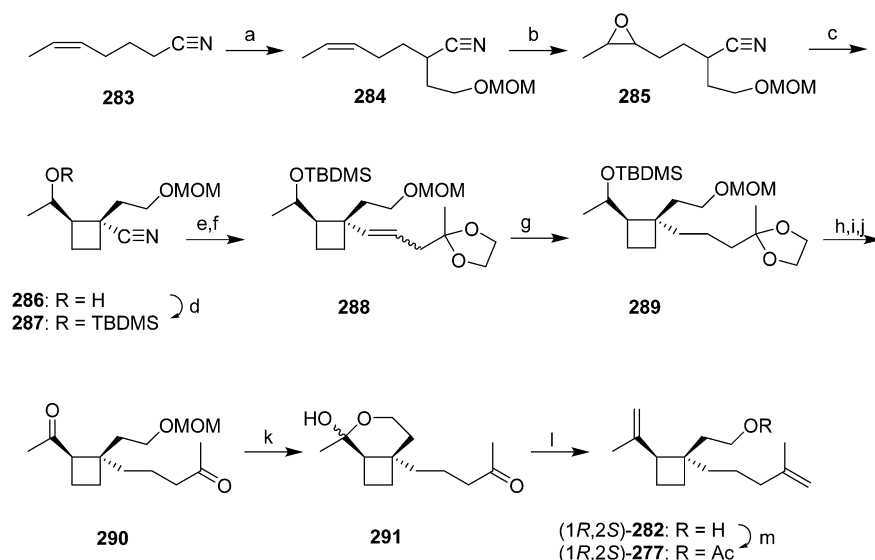
Fig. 7 Oleander scale pheromone.

Scheme 43 First syntheses of stereoisomers of the oleander scale pheromone. Reagents and conditions: (a) 3 steps from (*R*)-carvone, 34%; (b) KHMDS, THF/HMPA, 5-iodo-2-methyl-1-pentene, −20–0 °C, 60% based on recovered starting material; (c) PTSA, acetone/water 1 : 1; (d) NaBH$_4$, EtOH, 0 °C; (e) TosCl, pyridine, 43% over 3 steps; (f) LiHMDS, THF/HMPA, −10 °C, 46%; (g) LiAlH$_4$, THF, 86%; (h) TosCl, DMAP, CH$_2$Cl$_2$, 0 °C, 86%; (i) NaCN, H$_2$O/HMPA, 87%; (j) DIBAL, CH$_2$Cl$_2$, −20 °C, aq. workup; (k) NaBH$_4$, EtOH, 0 °C, 44% over 2 steps; (l) Ac$_2$O, pyridine; 70%. (m) OsO$_4$, NMO, acetone/H$_2$O; (n) NaIO$_4$, MeOH/H$_2$O, 64%; (o) DBU, benzene, reflux, 2 : 3 mixture of epimers; (p) Ph$_3$PCH$_3$Br, BuLi, THF, −70 °C-room temp, 75% over 2 steps; (q) DIBAL, CH$_2$Cl$_2$, −20 °C, then H$_3$O$^+$; (r) NaBH$_4$, EtOH, 32% from nitrile; (s) Ac$_2$O, pyridine, 70%.

This journal is © The Royal Society of Chemistry 2015

*Nat. Prod. Rep.*, 2015, **32**, 1067–1113 | **1091**

provide material for bioassays, and without having to resolve scalemic intermediates (Scheme 43).[106] (*R*)- and (*S*)-carvone were chosen as readily accessible chiral synthons, and the initial steps of the synthesis were modeled after those used in the synthesis of grandisol, a monoterpenoid analog containing the same 1,1,2-trisubstituted cyclobutane motif. Thus, (*R*)-(−)-carvone **171** was converted to acetal ester **175** in 3 steps, followed by alkylation of the corresponding enolate to install the longest side chain. Hydrolysis of the acetal, reduction of the aldehyde, and tosylation of the resulting alcohol then yielded the key intermediate **278** for formation of the cyclobutane ring. After careful optimization of the conditions, the cyclization proceeded in 46% yield, giving an unexpectedly high 19 : 1 ratio of the two possible diastereomers of **279**. The order of the steps was critical because it proved impossible to install the long side chain if the cyclization was performed first. With the 1,1,2-

trisubstituted cyclobutane core assembled, straightforward functional group manipulations then gave (1*S*,2*S*)-**277**, in 14 steps from (*R*)-carvone.

The authors then tried to take advantage of the high stereocontrol obtained in the cyclization step to develop a synthesis of the other diastereomer, by installing the acetoxyethyl sidechain or a suitable precursor before cyclizing, and then converting the carboethoxy group to the 4-methyl-4-pentenyl sidechain. However, several variations on this theme were not successful. Instead, (1*S*,2*S*)-**280** from the first synthesis was oxidized to the diketone **281**, and the ketone directly attached to the ring was epimerized with base, giving an inseparable 2 : 3 mixture of diastereomers of **281**. Wittig reactions then installed the two terminal methylenes, followed by conversion of the nitrile to the alcohol diastereomers **282**, which were separable by flash chromatography. Acetylation of each alcohol then

Scheme 44 Petschen's synthesis of racemic and chiral forms of oleander scale pheromone. Reagents and conditions: (a) LDA, Br(CH₂)₂OMOM, THF/HMPA, 61%; (b) *m*-CPBA, CH₂Cl₂, 92%; (c) LiHMDS, THF, 0 °C-reflux, 60%; (d) TBDMSiCl, imidazole, DMF, 98%; (e) DIBAL, hexane, then H₃O⁺, 90%; (f) Ph₃P=CHCH₂C(OCH₂CH₂O)CH₃, THF, 82%; (g) H₂, Pd/C, EtOH, 90%; (h) TBAF, THF, 98%; (i) PDC, DMF, 97%; (j) Amberlyst A15 resin, acetone/H₂O, 95%; (k) Dowex 50W-X4, MeOH, then SiO₂, hexane, acetone/H₂O; (l) excess Ph₃P=CH₂, THF, 45% over 2 steps; (m) Ac₂O, Et₃N, DMAP, CH₂Cl₂, 98%.
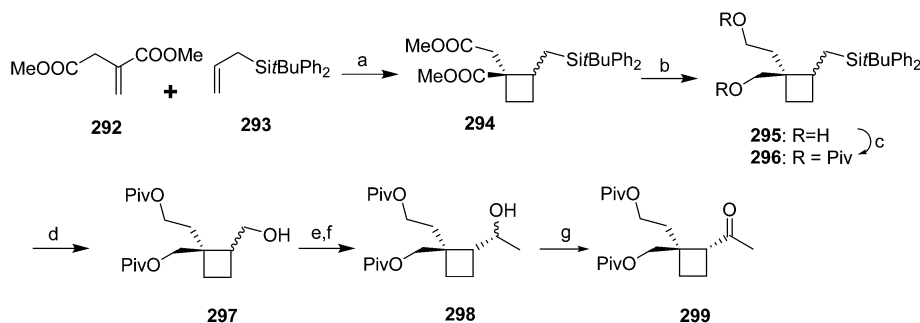
Scheme 45 Synthesis of an advanced intermediate for the oleander scale pheromone. Reagents and conditions: (a) TiCl₄, CH₂Cl₂, room temp – reflux, 96%; (b) LiAlH₄, THF, 93%; (c) PivCl, Et₃N, DMAP, CH₂Cl₂, 90%; (d) BF₃·2AcOH, ClCH₂CH₂Cl, reflux, then KF, NaHCO₃, H₂O₂, THF/MeOH, 1 : 1, 36−48% over 2 steps; (e) PDC, molecular sieve; (f) MeMgBr, THF, −10−25 °C, ~72%, *anti* and *syn* isomers separated by chromatography; (g) TPAP, NMO, 4A molecular sieve, CH₂Cl₂, −10 °C-room temp, 95%.

1092 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

completed the syntheses. The other two stereoisomers were obtained with the same sequences of reactions from (S)-carvone. With all stereoisomers available, the insect-produced compound was shown to have the (1R*,2S*) relative configuration, the two enantiomers of which did not resolve by chiral stationary phase chromatography. However, bioassays showed that the natural compound was almost certainly the (1R,2S)-enantiomer.

The key step in a second synthesis of the racemic pheromone was a regio- and stereocontrolled intramolecular 4-*exo-tet*-cyclization of a *cis*-epoxynitrile precursor **285** (Scheme 44).[107a] The *cis*-stereochemistry of the epoxide was important to ensure high regioselectivity, allowing preferential formation of the cyclobutane ring. Thus, nitrile **283** was alkylated with MOM-protected 2-bromoethanol, then epoxidized with *m*-CPBA. Use of MOM rather than other more bulky protecting groups proved crucial to later steps in the synthesis. Optimization of the epoxide opening and cyclization conditions resulted in a 60% yield of **286**, with a 79 : 21 ratio favoring the desired *trans* product. Protection of the resulting alcohol, conversion of the nitrile to an aldehyde, and Wittig reaction then installed the precursor to the third side chain, giving **289**. Removal of the TBDMS protecting group, PDC oxidation, and removal of the ketal protecting group gave the diketone **290**. Double Wittig reaction with $Ph_3P=CH_2$ then gave a precursor with the two terminal methylene groups installed. However, the final MOM protecting group could not be removed cleanly under a variety of conditions. Instead, the MOM group was removed from diketone **290** by treatment with an acid catalyst in MeOH, followed by hydrolysis of the resulting acetal to the hemiacetal **291**. Double Wittig reaction of this precursor with a large excess of

$Ph_3P=CH_2$ then gave the penultimate precursor (1R,2S)-**282**, and acetylation completed the synthesis. In an alternative sequence, all three protecting groups were removed from **289** to give the ketonediol, followed by selective acetylation of the primary alcohol, oxidation of the secondary alcohol, and double Wittig olefination. The latter step resulted in partial removal of the acetate ester, which was easily reinstalled with acetic anhydride, Et_3N, and DMAP. This alternative sequence (not shown) produced a similar yield from **289** as the first series of steps.

With a synthesis of the racemic pheromone in hand, the authors then carried out kinetic resolutions of intermediate **286** with lipase PS or PFL and vinyl acetate to provide the enantiomers of **286**, each in >98% ee, which were then carried through to the final products by the route described above. Both enzymes selectively esterified the (1R,2S,5R)-enantiomer of alcohol **286**.[107b]

In an entirely different approach, Schmidt et al. used a $TiCl_4$-promoted [2 + 2] cycloaddition to construct trisubstituted cyclobutane intermediate **294** in 96% yield and an *anti/syn* ratio of 2 : 1 (Scheme 45).[108a] Reduction of the esters and protection of the resulting alcohols, followed by a modified Tamao–Fleming oxidation gave diprotected triol **297**. Several straightforward steps then gave advanced intermediate **299**. At any stage the diastereomers were apparently separable, so the diastereomers could be carried forward as a mixture or as separate isomers. However, the synthesis does not yet seem to have been carried forward to completion.[108b]

3.2.3 Pheromones from cyclopentanyl and cyclopentenyl monoterpenols.
To date, three cyclopentane-based pheromone structures have been found (Fig. 8), all from members of the
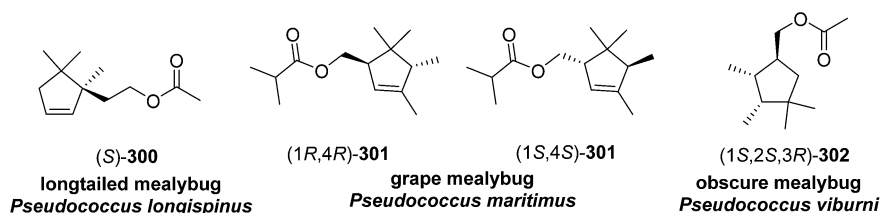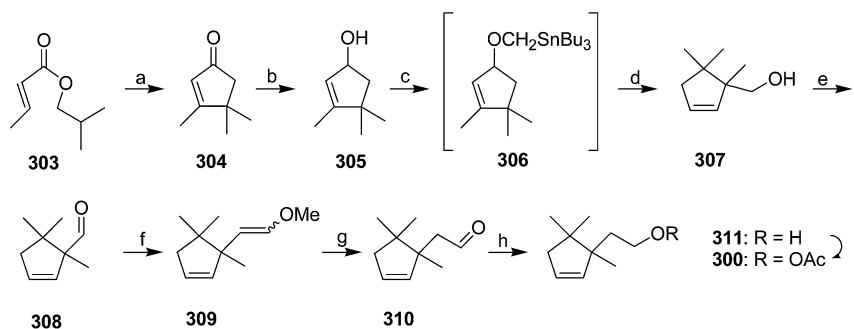

Fig. 8 Mealybug pheromones with cyclopentanyl and cyclopentenyl substructures.

Scheme 46 Millar group's first synthesis of longtailed mealybug pheromone *via* 2,3-Wittig rearrangement. Reagents and conditions: (a) polyphosphoric acid, 100 °C; (b) LiAlH$_4$, ether, 89%; (c) KH, THF, Bu$_3$SnCH$_2$I, 91%; (d) BuLi, THF, −100 °C, 25%; (e) PCC, CH$_2$Cl$_2$; (f) MeOCH$_2$PPh$_3$Cl, BuLi, THF, 53% over 2 steps; (g) H$_3$O$^+$, 86%; (h) NaBH$_4$, EtOH, 95%; (i) AcCl, pyridine, ether, 77%.

mealybug genus *Pseudococcus*. The pheromones of the long-tailed and obscure mealybugs were the first members of their respective terpene skeletal classes to be found anywhere in nature, whereas the alcohol portion of the grape mealybug pheromone had been previously found in the secretions of a carrion beetle.[109] The pattern of methylation of the longtailed mealybug pheromone suggests that it may be derived from geraniol, whereas the grape and obscure mealybug pheromones may be derived from cyclization of lavandulol to a cyclobutane intermediate followed by ring expansion and methyl migration.

Longtailed mealybug pheromone. The pheromone of the longtailed mealybug, *Pseudococcus longispinus*, was identified by Millar and coworkers as 2-(1,5,5-trimethylcyclopent-2-en-1-yl) ethyl acetate **300**.[110] The first synthesis of the racemate used a 2,3-Wittig rearrangement to set the two contiguous quarternary centers while simultaneously placing the double bond correctly (Scheme 46).[110] Thus, cyclization of isobutyl 2-butenoate **303** in hot polyphosphoric acid yielded multigram quantities of cyclopentenone **304**. This interesting reaction represents an example of a general rearrangement of α,β-unsaturated esters to cyclopentenones, and presumably proceeds *via* a fragmentation-recombination-Nazarov cyclization pathway.[111] Reduction and *O*-alkylation with Bu$_3$SnCH$_2$I, followed by transmetallation

with BuLi at low temperature, gave the rearranged cyclopentenol **307**, albeit in low yield. A straightforward series of reactions extended the chain by one carbon *via* aldehyde **310**, followed by reduction of the aldehyde and acetylation.

The second synthesis used an entirely different strategy to construct the cyclopentane ring from an acylic precursor, *via* insertion of a carbene into a methine C–H bond to close the ring (Scheme 47).[112] Thus, β-ketoester **313**, from γ-alkylation of methylacetoacetate **15** with isobutyl iodide **312**, was converted to diazo ester **314**. Rh-catalyzed cyclization proceeded in excellent yield, followed by alkylation of the resulting ketoester **315** with methyl iodide to complete the assembly of the vicinal quarternary carbons. Conversion of the ketone of **316** to the enol triflate **317** and Pd-catalyzed reduction placed the double bond, giving **318**. The remainder of the synthesis used the same series of steps described above, *i.e.*, one-carbon extension of the side chain, reduction of the resulting aldehyde, and acetylation, producing >5 g of the racemic pheromone.

To encourage commercialization of the pheromone, a shorter 6-step sequence was then developed (Scheme 48).[113] The first two steps were the same as those used in the first synthesis, but then instead of using a 2,3-Wittig rearrangement, an Ireland-Claisen rearrangement was used. This produced the
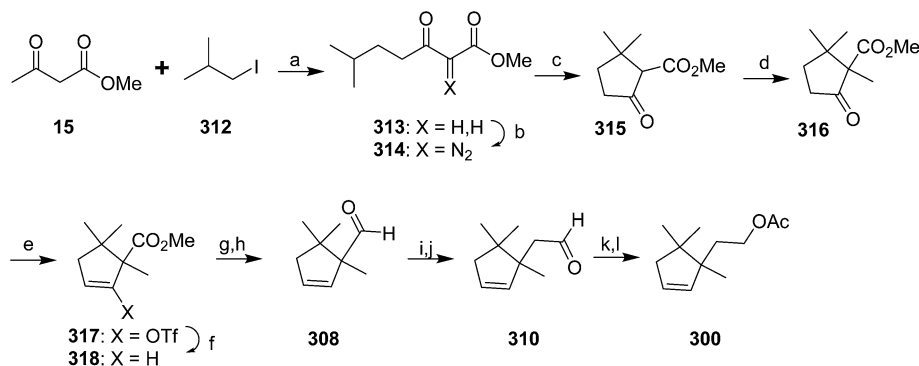

Scheme 47 Millar group's 2nd synthesis of longtailed mealybug pheromone, based on an intramolecular cyclization. Reagents and conditions: (a) NaH, BuLi, THF, 43%; (b) *p*-acetamidobenzenesulfonyl azide, Et$_3$N, MeCN, 96%; (c) Rh$_2$(OAc)$_4$, CH$_2$Cl$_2$, 91%; (d) K$_2$CO$_3$, MeI, acetone, 84%; (e) LDA, *N*-phenyl-bis(trifluoromethanesulfonimide), THF, 90%; (f) Pd(OAc)$_2$, Ph$_3$P, Bu$_3$N, HCOOH, DMF, 82%; (g) LiAlH$_4$, Et$_2$O; (h) (COCl)$_2$, DMSO, CH$_2$Cl$_2$, Et$_3$N; (i) MeOCH$_2$PPh$_3$Cl, NaHMDS, THF, 72% over 3 steps; (j) H$_3$O$^+$, THF; (k) LiAlH$_4$, Et$_2$O, 85% over 2 steps; (l) AcCl, pyridine, Et$_2$O, 95%.
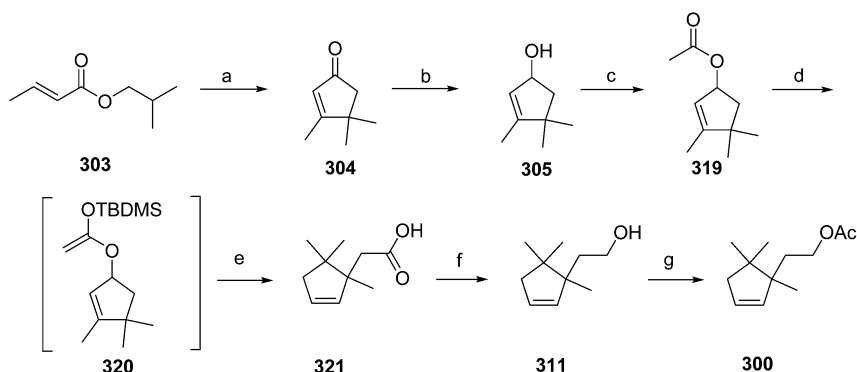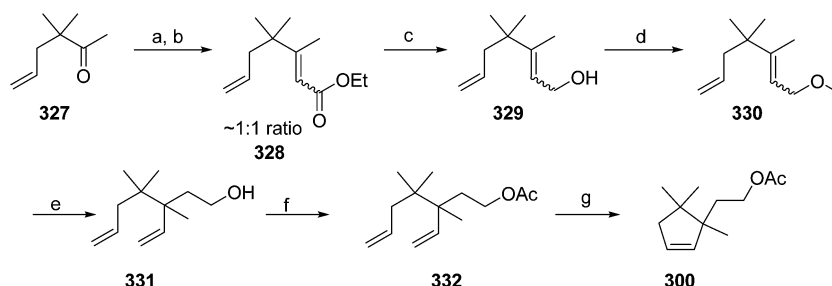

Scheme 48 Synthesis of the longtailed mealybug pheromone **300** based on Ireland-Claisen rearrangement. Reagents and conditions: (a) polyphosphoric acid; (b) LiAlH$_4$, Et$_2$O; (c) Ac$_2$O, DMAP, pyridine, CH$_2$Cl$_2$, 78% for 2 steps; (d) LDA, TBDMSiCl, THF; (e) reflux, then NaOH, acidic workup, 50% from **319**; (f) LiAlH$_4$, Et$_2$O, 95%; (g) AcCl, pyridine, Et$_2$O, 95%.
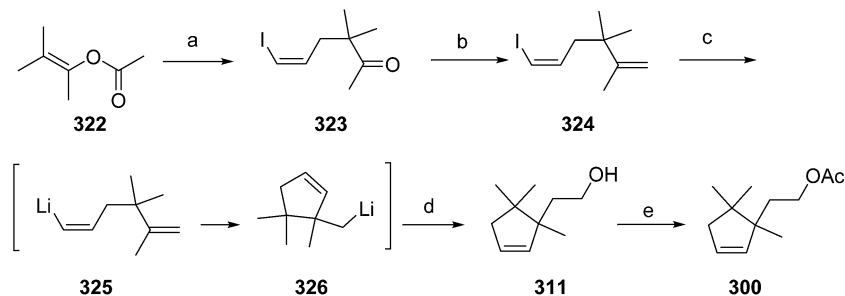
Scheme 49 Synthesis of the longtailed mealybug pheromone **300** based on intramolecular carbolithiation. Reagents and conditions: (a) MeLi, DME, (Z)-3-bromo-1-iodopropene, 64%; (b) Zn, cat. PbCl$_2$, CH$_2$Br$_2$, THF, then TiCl$_4$, 53%; (c) 2.2 eq. t-BuLi, −78 °C, 10 min, then TMEDA, room temp, 1 h; (d) (CH$_2$O)$_n$, −78 °C, then room temp, 1 h, finally H$_3$O$^+$, 65% over 2 steps; (e) AcCl, pyridine, Et$_2$O, 86%.

Scheme 50 Synthesis of the longtailed mealybug pheromone **300** based on ring-closing metathesis. Reagents and conditions: (a) ethoxyacetylene, n-BuLi, THF, −78 °C; (b) Sc(OTf)$_3$, CH$_2$Cl$_2$–EtOH, room temp, 71% for 2 steps; (c) DIBAL, CH$_2$Cl$_2$, −20 °C, 92%; (d) ethyl vinyl ether, Hg(OAc)$_2$, 47% (77% based on recovered starting material); (e) DIBAL, CH$_2$Cl$_2$, −40 °C, 69%; (f) Ac$_2$O, Et$_3$N, CH$_2$Cl$_2$; (g) Grubbs II catalyst (10 mol%), CH$_2$Cl$_2$, room temp, 63% for 2 steps.

complete carbon skeleton of the pheromone, in contrast to the 2,3-Wittig rearrangement route which required several extra steps to add an additional carbon. The Ireland-Claisen rearrangement of **319** proceeded smoothly through the silyl ketene acetal **320** to give γ,δ-unsaturated carboxylic acid **321** in respectable yield. LiAlH$_4$ reduction of acid **321** and acetylation completed the synthesis.

Bailey and Bakonyi recently developed a concise synthesis of **300** based on intramolecular carbolithiation (Scheme 49).[114] Alkylation of the lithium enolate derived from **322** gave ketone **323**. When standard Wittig reaction conditions were employed to convert **323** to **324**, a large quantity of alkyne was generated as a side product by dehydrohalogenation of the vinyl iodide. In contrast, Utimoto's olefination procedure,[115] using a CH$_2$Br$_2$–TiCl$_4$–Zn system with catalytic PbCl$_2$ cleanly afforded (Z)-1-iodo-4,4,5-trimethyl-1,5-hexadiene **324**. The key 5-exo-trig ring closure via lithiation, cyclization, and trapping the resulting anion with paraformaldehyde proceeded efficiently (65%) in one pot to give **311**, and acetylation completed the synthesis.

The Reddy group developed a short and efficient synthesis of **300** based on ring-closing metathesis (Scheme 50).[116] The overall strategy is similar to the total synthesis of racemic herbertenediol.[117] Thus, olefination of sterically hindered ketone **327** led to α,β-unsaturated ester **328**. Because the Wittig reaction and its Horner–Wadsworth–Emmons variant were not successful, a two-step Meyer–Schuster rearrangement[118] was utilized for the desired transformation. DIBAL reduction, followed by Hg-promoted vinylation gave allyl vinyl ether **330**.

Treatment of **330** with DIBAL at −40 °C led to clean Claisen rearrangement and in situ reduction of the resulting aldehyde to give the key intermediate **331**. Acetylation and ring-closing metathesis then afforded the target **300**. Alternatively, alcohol **331** was subjected to ring-closing metathesis followed by acetylation to give **300** in slightly lower yield over the two steps (53 vs. 63%).

The Reddy group then modified the synthesis to prepare both enantiomers of the pheromone (Scheme 51).[119] Thus, racemic alcohol **331** was esterified with Harada's camphorsultam phthalic (CSP) acid to produce a mixture of diastereomers **333** and **334**,
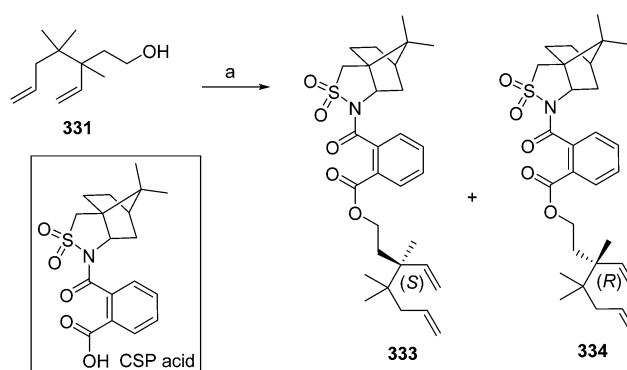

Scheme 51 Formation of separable diastereomeric derivatives of **331** en route to synthesis of the (R)- and (S)-enantiomers of the longtailed mealybug pheromone **300**. Reagents and conditions: (a) dicyclohexylcarbodiimide, DMAP, CSP acid, CH$_2$Cl$_2$.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1095**

which were separated by preparative HPLC. The relative and absolute configurations of **334** were determined through X-ray crystal structure analysis, revealing that the chiral center in the intermediate had the (R)-configuration. Each enantiomer was then hydrolyzed and converted to the respective pheromone enantiomers. In field trials, the (S)-(+)-enantiomer was highly attractive to male mealybugs, indicating that this was almost certainly the insect-produced enantiomer. The (R)-enantiomer was neither attractive nor inhibitory, so the racemic pheromone is entirely sufficient for practical purposes.

Grape mealybug pheromone. The sex pheromone of the grape mealybug, *Pseudococcus maritimus*, was identified by the Millar

group as *trans-α-necrodyl isobutyrate* **301**.[8] Because the insect is difficult to maintain in culture, it took several years to obtain enough purified pheromone to determine the structure by microprobe NMR. Furthermore, the pheromone was produced in very small quantities so gas chromatography coupled to electroantennogram detection had to be used to locate the pheromone as a small peak in a complex background.[8] The core monoterpene alcohol structure *trans-α-necrodol* **342** had been previously identified from a carrion beetle, and synthesized several times. Scheme 52 summarizes the synthesis reported by the Millar group,[120] which combined and optimized elements from previous syntheses of necrodol isomers, including a
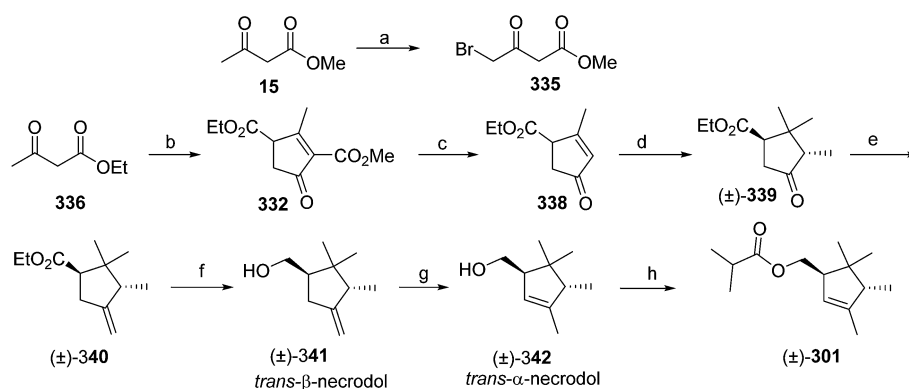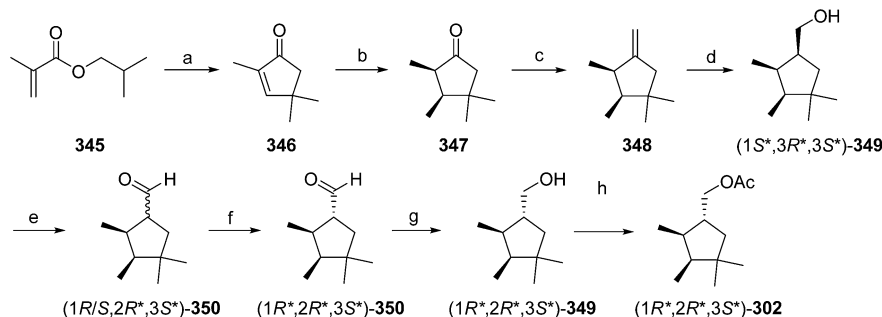

Scheme 52 Synthesis of the racemic grape mealybug pheromone **301**. Reagents and conditions: (a) Br₂, CHCl₃; (b) NaH, THF, **335**; (c) NaI, AcOH, diglyme, 28% over 3 steps; (d) Me₂Zn, Ni(acac)₂, THF, then HMPA, MeI, 64%; (e) TiCl₄, CH₂Br₂, Zn, THF, CH₂Cl₂, 67%; (f) LiAlH₄, Et₂O, 97%; (g) Li, ethylenediamine, 65%; (h) isobutyryl chloride, Et₃N, DMAP (cat.), CH₂Cl₂, 93%.
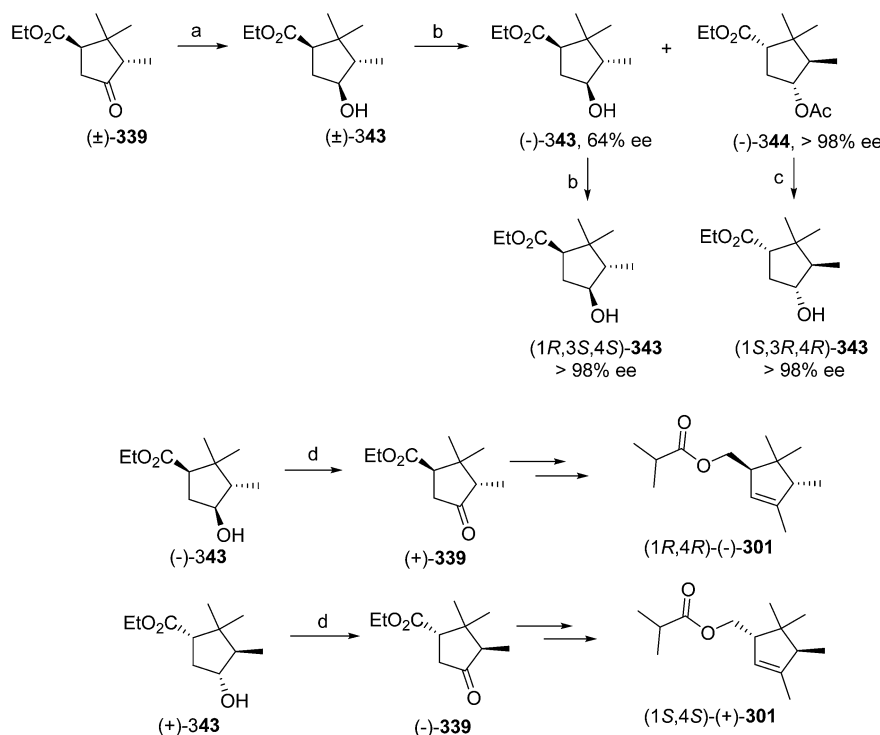
Scheme 53 Synthesis of the (1R,4R)- and (1S,4S)-enantiomers of the grape mealybug pheromone **301**. Reagents and conditions: (a) NaBH₄, EtOH; (b) vinyl acetate, Amano AK lipase; (c) K₂CO₃, MeOH; (d) DMSO, (COCl)₂, Et₃N, CH₂Cl₂.

1096 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

Scheme 56 Enantioselective synthesis of the obscure mealybug pheromone (1S,2S,3R)-**302**. Reagents and conditions: (a) orthoester Claisen rearrangement; (b) alkaline hydrolysis, 71% over 2 steps; (c) PivCl, Et₃N, THF, then X_N–Li, 83%; (d) NaHMDS, MeI, THF, 74%; (e) NaBH₄, THF/H₂O, 71%; (f) Swern oxidation, CH₂Cl₂, then Ph₃P=CHCO₂Me (one pot), 83%; (g) Me₂CuLi, TMSiCl, CH₂Cl₂, 81%; (h) O₃, MeOH/CH₂Cl₂, then NaBH₄, 80%; (i) Tf₂O, 2,6-lutidine, CH₂Cl₂; (j) NaHMDS, 72% for 2 steps; (k) DIBAL, CH₂Cl₂, 94%; (l) Ac₂O, pyridine, 85%.

of the racemate, followed by vibrational circular dichroism analysis to assign the absolute stereochemistry of each enantiomer.[129] The assignment was subsequently confirmed by enantioselective synthesis of the natural (1S,2S,3R)-enantiomer by Hashimoto and coworkers starting from known carboxylic acid **355**, prepared from prenyl alcohol **354** in two steps (Scheme 56).[130] The stereocenter at C-3 was established by asymmetric methylation of **356**, directed by Evans' chiral auxiliary. Reductive cleavage of the chiral auxiliary, and Swern oxidation followed by Wittig olefination in one pot produced γ-alkyl-α,β-unsaturated ester **359**. The stereocenter at C-2 was established by diastereoselective conjugate addition of an organocopper reagent to **359**. After several failed attempts, Yamamoto's method[131] using Me₂CuLi/TMSiCl in CH₂Cl₂ gave the desired product **360** with complete diastereoselectivity, rationalized by a Felkin-Anh type transition state model. Compound **360** was ozonized with a reductive workup to afford alcohol **361**, followed by triflation. The stereocenter at C-1 was established in the final cyclization step. A single

stereoisomer was obtained, probably due to the greater thermodynamic stability of **362** as compared to the corresponding epimer. DIBAL reduction of ester **362** and acetylation of the resulting alcohol gave (1S,2S,3R)-**302**.

The Reddy group subsequently reported an efficient synthesis of the (1R,2R,3S)-enantiomer from commercial D-(−)-pantolactone **363** (Scheme 57).[132] Thus, benzyl-protected lactol **364** was converted into α,β-unsaturated ester **365** by Wittig olefination and tosylation of the resulting alcohol. The two chiral centers at C-1 and C-2 were installed in one pot by a tandem conjugate addition/cyclization reaction, giving **366**. The alcohol from LiAlH₄ reduction of ester **366** was protected with the TBDPS group, followed by removal of the benzyl group, oxidation of the resulting secondary alcohol to ketone **368**, and Tebbe methylenation, furnishing alkene **369**. Hydrogenation of the exocyclic double bond using Wilkinson's catalyst established the chiral center at C-3 diastereoselectively giving **370**. Deprotection and acetylation of the resulting alcohol gave (1R,2R,3S)-**302**.
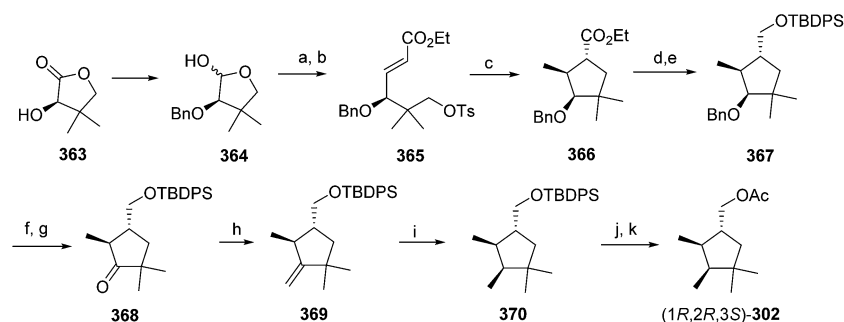
Scheme 57 Enantioselective synthesis of the unnatural, (1R,2R,3S)-enantiomer of the obscure mealybug pheromone. Reagents and conditions: (a) Ph₃P=CHCO₂Et, PhMe, reflux, 8 h, 75%; (b) TosCl, pyridine, DMAP, rt, 24 h, 88%; (c) Me₂CuLi, TMSiCl, THF, −78 °C to room temp, 24 h, 94%; (d) LiAlH₄, THF, 0 °C, 0.5 h, 95%; (e) TBDPSiCl, imidazole, pyridine, room temp, 93%; (f) 10% Pd/C, H₂, MeOH, 91%; (g) Dess-Martin periodinane, CH₂Cl₂, 0 °C, 1 h, 89%; (h) Tebbe's reagent, THF, 0 °C, 45 min, 89%; (i) Rh(PPh₃)₃Cl, H₂, THF, t-BuOH (1 : 1), room temp, 85%; (j) TBAF, THF, 60 °C, 1 h, 86%; (k) Ac₂O, pyridine, DMAP, CH₂Cl₂, room temp, 1 h, 87%.

3.3 Polyketide pheromones of pine bast scales

As their names suggest, pine bast scales in the genus *Matsucoccus* develop exclusively on pine, and their pheromones are markedly different than those of other scale and mealybug species. Specifically, the pattern of methylation for all of the five pheromone compounds identified from pine bast scales clearly indicates that they are products of the polyketide rather than the terpenoid biosynthetic pathway. As might be expected for pheromones of congeneric species, the structures share a
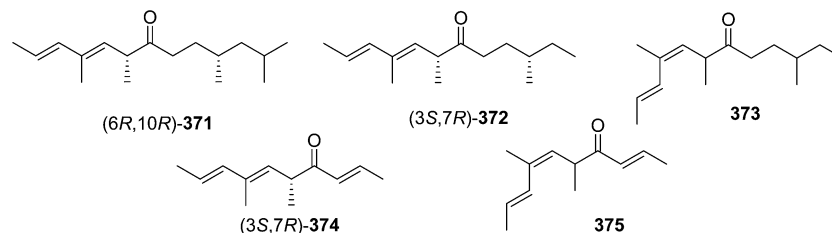
Fig. 9 Structures of pheromone components of pine bast scales.
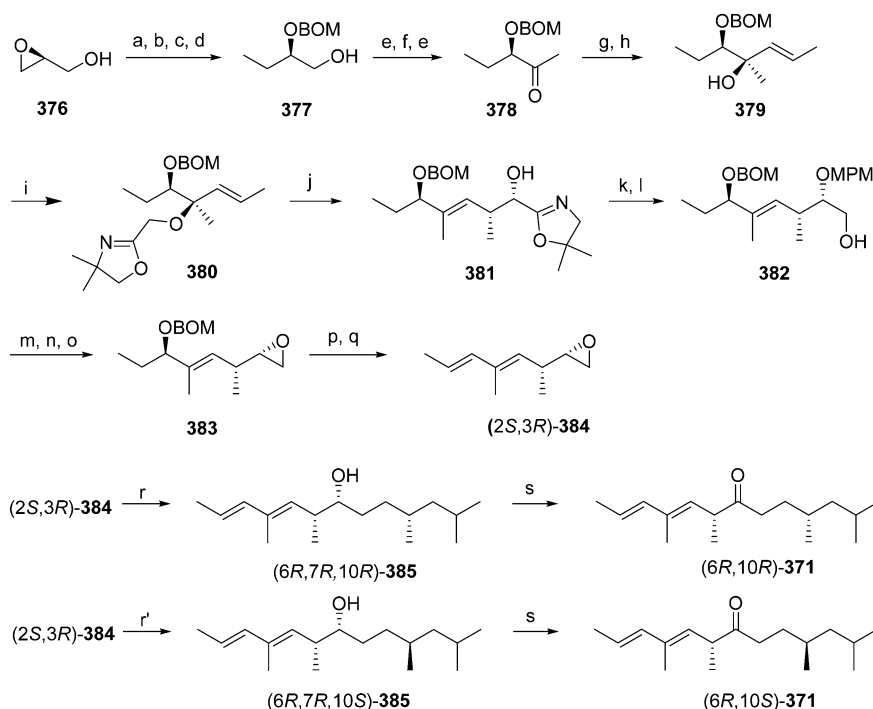
Scheme 58 Cywin et al. enantioselective synthesis of (6R,10R)-371 and (6R,10S)-371. Reagents and conditions: (a) TBDMSiCl, imidazole, CH_2Cl_2, 0 °C; (b) MeMgBr, CuI, THF; (c) $PhCH_2OCH_2Cl$, *i*-PrNEt₂, LiAlH₄; (d) TBAF, THF, 58% over 4 steps; (e) Swern oxidation; (f) MeMgBr, THF, then e, 85% over 3 steps; (g) MeC≡CMgBr, THF; (h) LiAlH₄, THF; 81% over 2 steps; (i) 2-(chloromethyl)-4,5-dihydro-4,4-dimethyloxazole, KH, DME; (j) *n*-BuLi, THF, −78 °C; (k) *p*-MeOPhCH₂Cl, KH, DME; (l) TFA, H₂O, 24 h, then LiAlH₄, THF, 44% over 4 steps; (m) TosCl, pyridine; (n) 1M aq. Ce(NH₄)₂(NO₃)₆, MeCN; (o) NaOMe, MeOH; 74% over 3 steps; (p) Na, NH₃, −78 °C; (q) MeOC(O)NSO₂NEt₃, benzene, 50 °C; 92% over 2 steps; (r) (S)-2,4-dimethylpentylMgBr, CuI, THF, 47%; (r') (R)-2,4-dimethylpentylMgBr, CuI, THF, 47%; (s) (n-Pr)₄NRuO₄, NMO, 4A sieves, CH_2Cl_2, 98%.
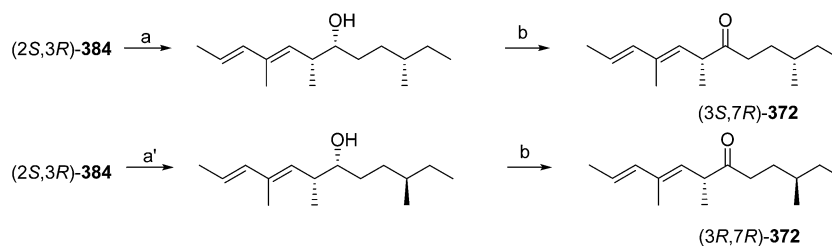
Scheme 59 Cywin's enantioselective synthesis of (3S,7R)-372 and (3R,7R)-372. Reagents and conditions: (a) (S)-2-methylbutylMgBr, CuI, THF, 60%; (a') (R)-2-methylbutylMgBr, CuI, THF, 58%; (b) (n-Pr)₄NRuO₄, NMO, 4A sieves, CH_2Cl_2.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1099**

common structural motif on the left hand side of the molecules as drawn in Fig. 9, with variable chains appended on the right hand side, and two of the structural pairs vary only in the stereochemistry of one double bond. Because most of the synthetic strategies used to make these compounds have taken advantage of the shared structural motif, the syntheses will be described collectively rather than being broken out by species as was done in earlier sections.
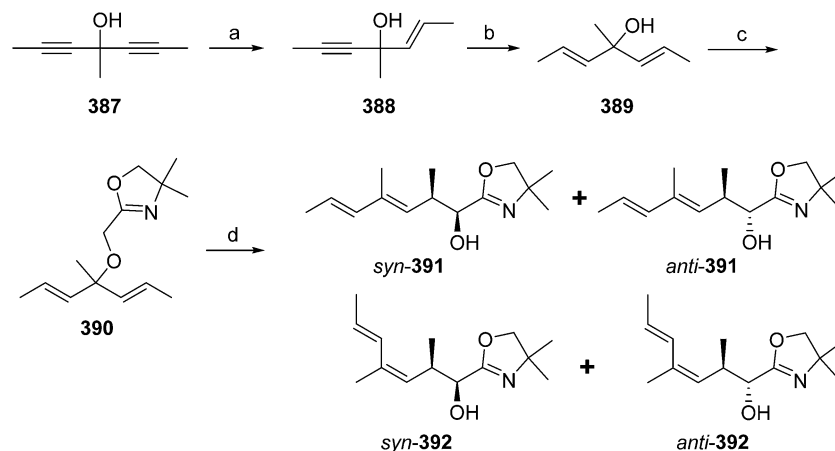
Scheme 60 Preparation and [2,3]-Wittig rearrangement of tertiary bis-allyllic ether **390**. Reagents and conditions: (a) LiAlH$_4$, 98%; (b) LiAlH$_4$, 89%; (c) 2-(chloromethyl)-4,5-dihydro-4,4-dimethyloxazole, KH, 90%; (d) KH, 100%.
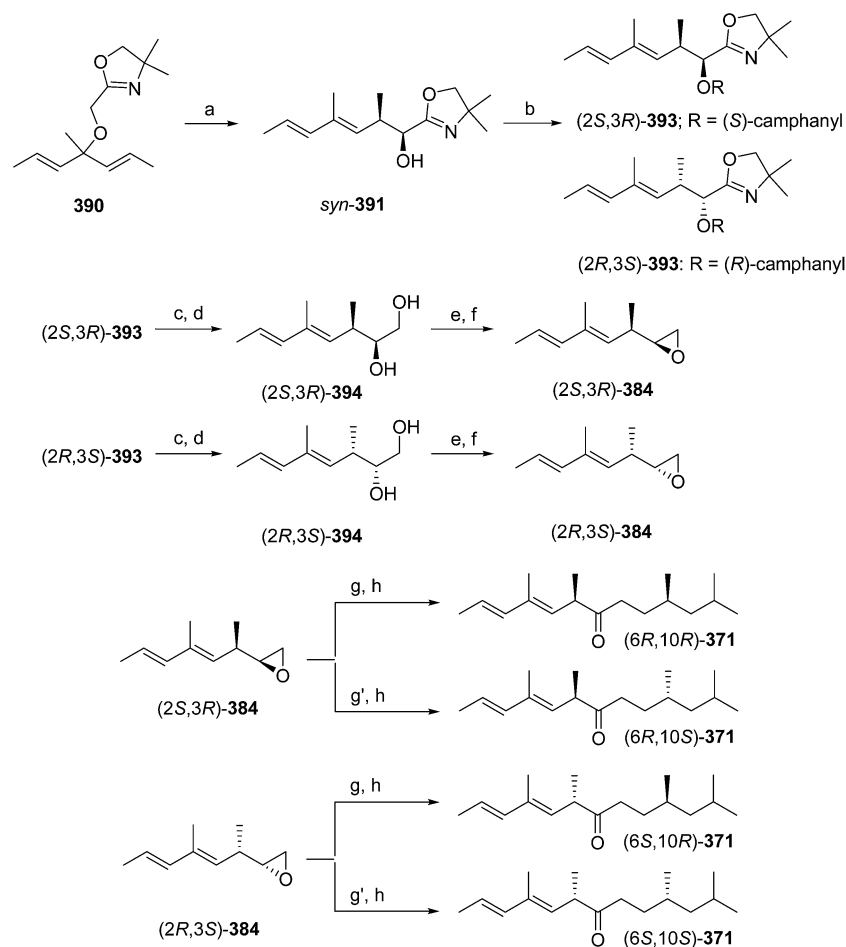
Scheme 61 Shi's enantioselective synthesis of all four stereoisomers of **371**. Reagents and conditions: (a) *n*-BuLi, THF/HMPA = 7 : 3, −100 °C, 0.5 h, 95%; (b) (*S*)-camphanyl chloride and (*R*)-camphanyl chloride, recrystallization; (c) TFA, H$_2$O, THF, 25 °C, 24 h; (d) LiAlH$_4$, THF, 0 °C, 4 h, 87% for 2 steps; (e) TosCl, pyridine, CH$_2$Cl$_2$, −30 °C, 48 h; (f) NaOMe, MeOH, 0 °C, 2 h, 63% for 2 steps; (g) (*S*)-2,4-dimethylpentylMgBr, CuI, THF; (g') (*R*)-2,4-dimethylpentylMgBr, CuI, THF; (h) (*n*-Pr)$_4$NRuO$_4$, NMO, 4A molecular sieves, 25 °C, 0.5 h, 82% for 2 steps for (6*R*,10*R*)-**371**.

1100 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

The first pine bast scale pheromone was identified by a joint effort among US, Chinese, and Korean scientists as (2*E*,4*E*)-4,6,10,12-tetramethyltrideca-2,4-dien-7-one (matsuone) **371**, for

the red pine bast scale *Matsucoccus resinosae* in the US, and two related and possibly synonymous species indigenous to Asia known as black pine bast scale, *M. matsumurae* in China and *M.*
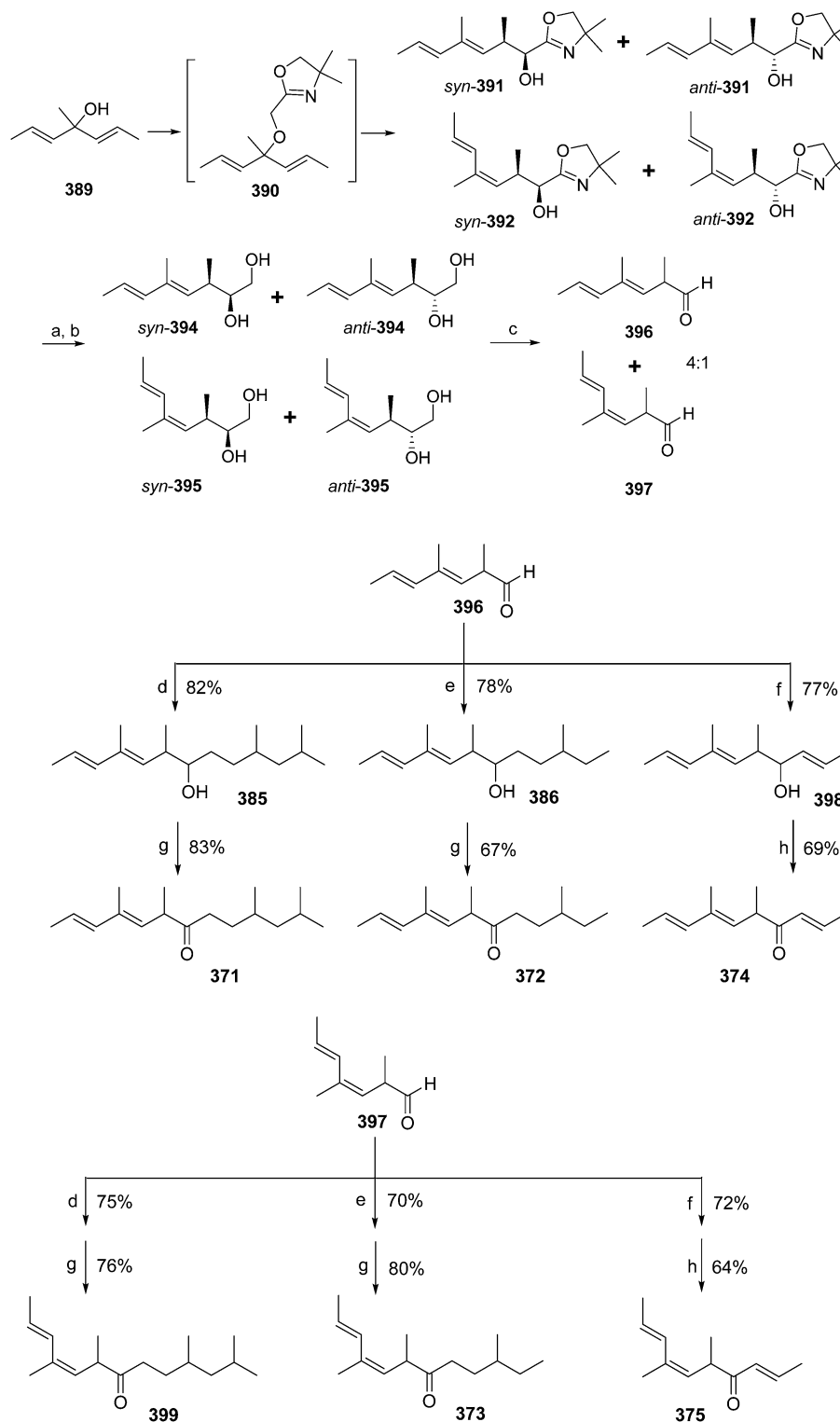
Scheme 62 Shi's geometrically selective synthesis of all the *Matsucoccus* pheromones **371-375** and **399** as racemic and diastereomeric mixtures. Reagents and conditions: (a) TFA, H$_2$O, THF, 25 °C, 24 h; (b) LiAlH$_4$, THF, 0 °C, 4 h, 87% for 2 steps; (c) NaIO$_4$, dioxane:H$_2$O (1 : 1), room temp, 2 h, 55%; (d) 3,5-dimethylhexylMgBr, THF, room temp, 1 h; (e) 3-methylpentylMgBr, THF, room temp, 1 h; (f) (*E*)-CH$_3$CH═CHBr, *t*-BuLi, THF, −115 °C, 1 h; (g) Jones' reagent, 0 °C, 5 min; (h) (*n*-Pr)$_4$NRuO$_4$, NMO, CH$_2$Cl$_2$, room temp, 0.5 h.

thunbergianae in Korea.[133] Subsequently, the sex pheromone of the maritime pine bast scale *M. feytaudi*, native to Africa and Europe, was identified as a mixture of (8E,10E)-3,7,9-trimethyldodeca-8,10-dien-6-one **372** and its (8Z,10E)-isomer **373** (**373** is ~3% of **372**).[134] The pheromone of the Israeli pine bast scale *M. josephi* was identified as a 75 : 25 mixture of (2E,6E,8E)-5,7-dimethyldeca-2,6,8-trien-4-one **374** and its (2E,6Z,8E)-isomer **375**.[135] Enantioselective syntheses of stereoisomers of **371**, **372**, and **374**, followed by bioassays or GC comparison with natural pheromones determined the absolute stereochemistry of **371** as (6R,10R), **372** as (3S,7R), and **374** as 5R (Fig. 9). Thus, all these *Matsucoccus* pheromones share the same (R)-configured ketone motifs, with the remaining parts of the molecules differing and conferring species specificity. The diastereomeric (6R,10S)-**371** was also active, whereas the other two diastereomers were not.[136,137b] Similarly, the unnatural diastereomer (3R,7R)-**372** was attractive to male *M. feytaudi*, whereas the other two diastereomers of **372** were inactive.[138] The unnatural (5S)-**374** also was not attractive to *M. josephi*.[139]

Cywin and coworkers reported the first enantioselective synthesis of two diastereomers of **371**, (6R,10R)-**371** and (6R,10S)-**371** respectively (Scheme 58).[140] The convergent synthesis started from commercially available (S)-(−)-glycidol **376**. Routine transformations gave methyl ketone **378**. Chelation-controlled addition of propynyl magnesium bromide, followed by LiAlH$_4$ reduction afforded tertiary allylic alcohol **379** with the desired stereochemistry. O-Alkylation with 2-(chloromethyl)-4,5-dihydro-4,4-dimethyloxazole and treatment of the resulting ether **380** with n-BuLi resulted in [2,3]-Wittig rearrangement to give a mixture of **381** (major product) and its (Z)-isomer (minor product). The mixture was converted to alcohol **382** and its (Z)-isomer, which were readily separable by flash chromatography. After conversion of **382** to epoxide **383**, reductive removal of the (benzyloxy)methyl (BOM) protecting group and dehydration using the Burgess protocol installed the second (E)-double bond, giving **384**. The copper-catalyzed coupling of (E,E)-diene epoxide (2S,3R)-**384** with (S)- and (R)-2,4-dimethylpentylmagnesium bromide, followed by oxidation, afforded (6R,10R)-**371** and (6R,10S)-**371**. Comparison of the 500 MHz ^{1}H NMR spectra of the two synthetic diastereomers with that of the natural pheromone established the relative stereochemistry of **371** as 6R*,10R* (6,10-*syn*).

Following the same route, Cywin and coworkers reported the first enantioselective synthesis of two diastereomers of the
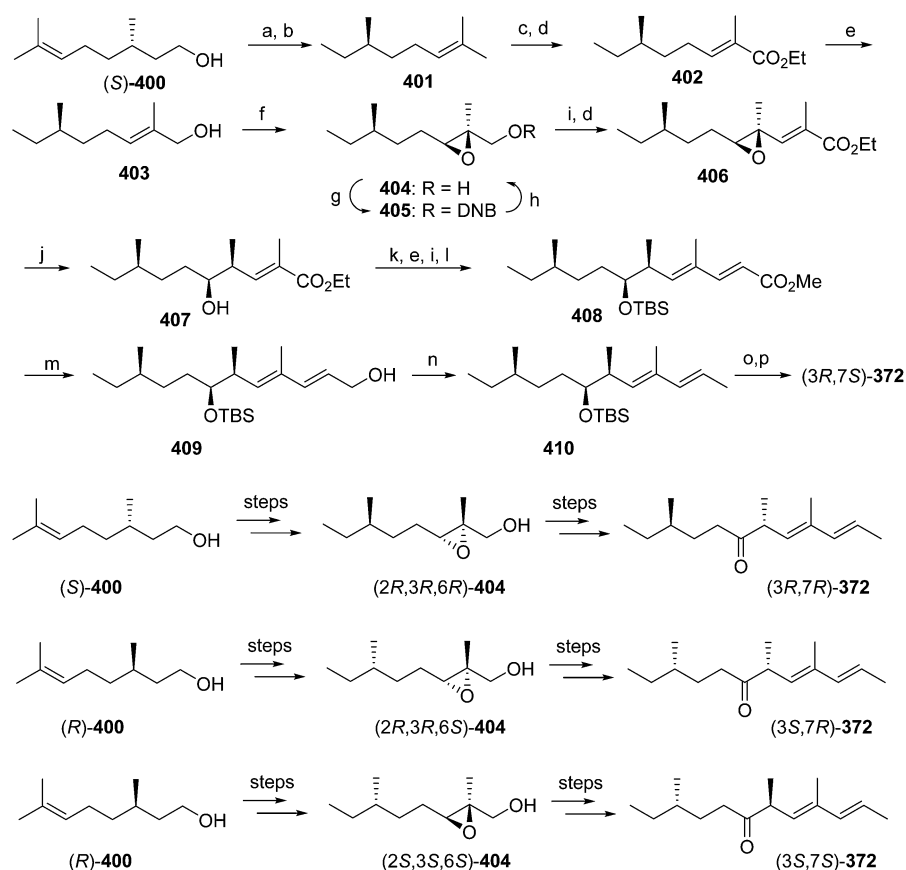
Scheme 63 Mori's enantioselective synthesis of all four stereoisomers of **372**. Reagents and conditions: (a) TosCl, pyridine; (b) LiAlH$_4$, THF, 79% over 2 steps; (c) O$_3$, NaHCO$_3$, MeOH/CH$_2$Cl$_2$; (d) Ph$_3$P=C(Me)CO$_2$Et, benzene, 81% over 2 steps; (e) DIBALH, Et$_2$O, 95%; (f) (iPrO)$_4$Ti, diethyl L-(+)-tartrate, t-BuOOH, 63%; (g) 3,5-DNBCl, pyridine, Et$_2$O, recrystallization, 45%; (h) NaOH, MeOH/THF/H$_2$O, 98%; (i) Swern oxidation, then d, 87%; (j) Pd$_2$(dba)$_3$·CHCl$_3$, Ph$_3$P, HCO$_2$H, Et$_3$N, dioxane, 78%; (k) TBDMSiCl, imidazole, DMF, then e, then i, 81%; (l) (MeO)$_2$P(O)CH$_2$CO$_2$Me, n-BuLi, THF, 96%; (m) DIBALH, Et$_2$O, 91%; (n) SO$_3$·pyridine, THF, then LiAlH$_4$, THF, 59%; (o) TBAF, THF, prep. HPLC, 71%; (p) Swern oxidation, 94%.

1102 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

major component of the sex pheromone of *M. feytaudi*, (3*S*,7*R*)-**372** and (3*R*,7*R*)-**372** (Scheme 59).[141] The convergent synthesis employed the advanced intermediate described in Scheme 58, epoxide (2*S*,3*R*)-**384** and (*S*)- and (*R*)-2-methylbutylmagnesium bromide. Comparison of the 500 MHz ¹H NMR spectra of the two synthetic diastereomers with that of the natural pheromone established the relative stereochemistry of **372** as 3*S**,7*R** (3,7-*syn*), the same as that of matsuone **371**.

Shi and coworkers used an analogous diastereoselective [2,3]-Wittig rearrangement of tertiary bisallylic ether **390** to provide a direct entry to conjugated diene systems containing an (*E*)-disubstituted double bond, a stereochemically defined trisubstituted double bond, and a homoallylic hydroxyl group, key structural elements of **371**, **372**, and **374** (Scheme 60).[137a] Rearrangement of easily prepared **390** favored the formation of

syn-**391** under all reaction conditions, but the ratio of products was clearly influenced by reaction conditions. Use of a lithium base at low temperature with THF/HMPA solvent produced optimum yields of *syn*-**391** (*syn*-**391**: 93.6%, *anti*-**391**: 6.4%).

This transformation could be applied in two ways. First, carrying out the [2,3]-Wittig rearrangement under the conditions described above gave (±)-*syn*-**391** in excellent yield, which could be isolated in over 99% purity by recrystallization from ether. Then, derivatization of the racemate with (*S*)-camphanyl chloride gave a mixture of two diastereomers, from which (2*S*,3*R*)-**393** could be selectively crystallized. Derivatization of racemic (±)-*syn*-**391** with (*R*)-camphanyl chloride similarly gave (2*R*,3*S*)-**393** in > 99% de. The two enantiomers were then transformed to epoxides (2*S*,3*R*)-**384** and (2*R*,3*S*)-**384**, respectively. The copper-catalyzed coupling of the epoxides with (*S*)-
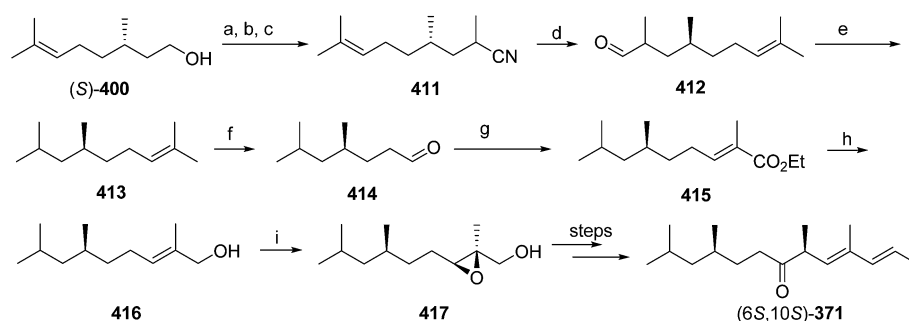
Scheme 64 Mori's enantioselective synthesis of (6*S*,10*S*)-**371**. Reagents and conditions: (a) TosCl, pyridine; (b) NaCN, DMF; 95% over 2 steps; (c) LDA, MeI, THF; 80%; (d) DIBALH, Et₂O; (e) H₂NNH₂·H₂O, KOH, H₂O/ethylene glycol, 89% over 2 steps; (f) O₃, NaHCO₃, MeOH/CH₂Cl₂, then Me₂S; (g) Ph₃P=C(Me)CO₂Et, benzene, 85% over 2 steps; (h) DIBALH, Et₂O, 99%; (i) (iPrO)₄Ti, diethyl ʟ-(+)-tartrate, *t*-BuOOH, CH₂Cl₂, 81%.
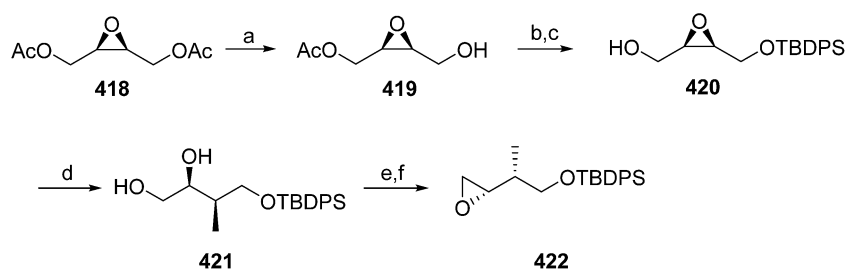
Scheme 65 Preparation of intermediate epoxide **422**. Reagents and conditions: (a) Pig pancreatic lipase, phosphate buffer/isopropyl ether, 0 °C, 71%; (b) TBDPSiCl, Et₃N, DMAP, CH₂Cl₂, quant; (c) K₂CO₃, MeOH, 0 °C, 98%; (d) Me₃Al, pentane, CH₂Cl₂, 62%; (e) TosCl, pyridine; (f) K₂CO₃, MeOH, 85% for 2 steps.
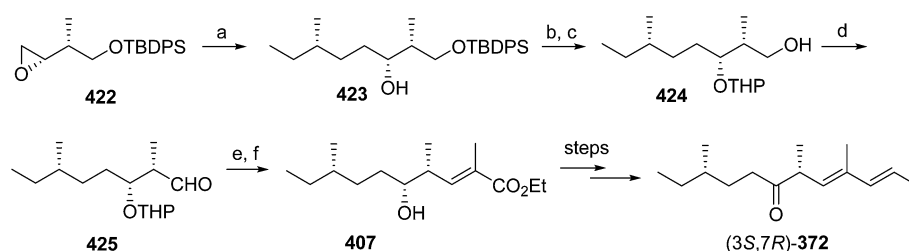
Scheme 66 Mori's improved synthesis of (3*S*,7*R*)-**372**. Reagents and conditions: (a) (*S*)-2-methylbutylmagnesium bromide, CuBr, THF, 93%; (b) dihydropyran, PPTS, CH₂Cl₂, 95%; (c) TBAF, THF, 95%; (d) Swern oxidation; (e) Ph₃P=C(Me)CO₂Et, benzene; (f) PTSA, EtOH, 79% over 3 steps.

and (R)-2,4-dimethylpentylmagnesium bromide, respectively, followed by oxidation, afforded the four stereoisomers of **371** (Scheme 61).[137b]

This transformation was then extended to generate the (E,E)- and (Z,E)-conjugated dienes in one pot, providing an easy access to the geometric isomers of all three of the *Matsucoccus*

pheromones (Scheme 62).[137c] In the presence of two equivalents of KH, the tertiary allylic alcohol **389** was alkylated to afford ether **390**, which without isolation underwent the desired [2,3]-Wittig rearrangement to give a mixture of four racemic, conjugated dienes (*syn*-**391** : *anti*-**391** : *syn*-**392** : *anti*-**392** = 7 : 1 : 1 : 1). The mixture was converted directly into the
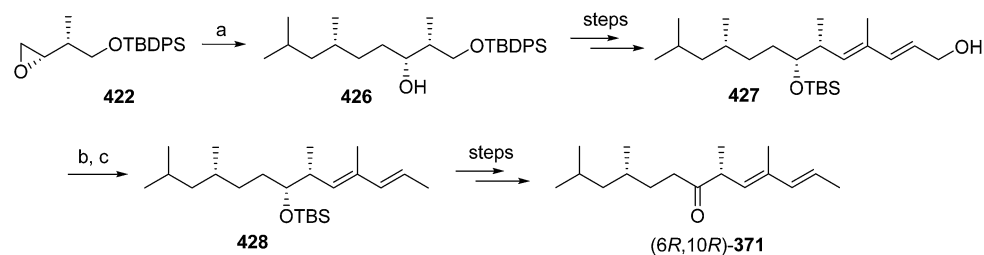
Scheme 67 Mori's improved synthesis of (6R,10R)-**371**. Reagents and conditions: (a) (S)-2,4-dimethylpentylMgBr, CuBr, THF, 96%; (b) n-BuLi, Mes$_2$O, THF; (c) LiBEt$_3$H, THF, 73% for 2 steps.
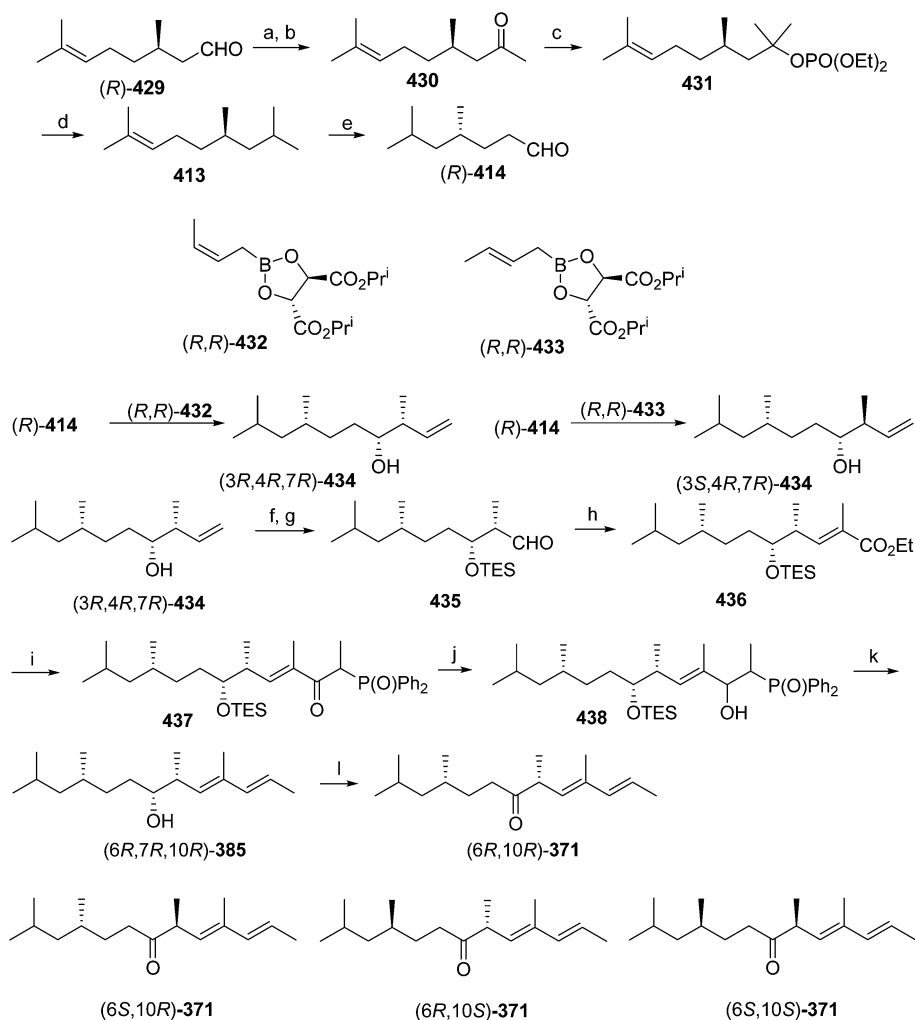
Scheme 68 Lin and Xu's enantioselective synthesis of all four stereoisomers of **371**. Reagents and conditions: (a) MeMgI, Et$_2$O, 98%; (b) PCC, CH$_2$Cl$_2$, room temp, 96%; (c) i. MeMgI, Et$_2$O, ii. n-BuLi, THF, −20 to 0 °C, iii. ClPO(OEt)$_2$, TMEDA, −20 °C to room temp, 95%; (d) Li, EtNH$_2$, −30 °C, quant.; (e) O$_3$, CH$_2$Cl$_2$, −78 °C, then Ph$_3$P; (f) TESiCl, imidazole, DMF, quant.; (g) O$_3$, CH$_2$Cl$_2$, −78 °C, then Ph$_3$P, room temp; (h) Ph$_3$P=C(Me) CO$_2$Et, PhMe, 80 °C, 36 h, 76% over 2 steps; (i) Ph$_2$P(O)Et, n-BuLi, THF, −78 °C, 95%; (j) DIBAL, Et$_2$O, −78 °C, 74%; (k) NaH, DMF, 40 °C, 65%; (l) Swern oxidation, 90%.

1104 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

corresponding mixture of four dienediols **394** and **395**. Periodate cleavage then afforded a mixture of aldehydes **396** and **397** (4 : 1 ratio), which were separated by flash chromatography. Addition of the appropriate Grignard or lithium reagents, followed by oxidation, yielded geometrically pure final products. This geometrically selective synthesis provided the naturally occurring sex pheromone components **371–375**, along with **399**, the geometric isomer of **371**, each as a racemic and diastereomeric mixture. Whereas **373** and **375** were found as trace components in the insects, **399** has not yet been reported as a natural product.

Mori's synthesis of all four stereoisomers of **372**, the major component of the sex pheromone of *M. feytaudi*, built up the chain from the opposite end (Scheme 63).[142] Thus, synthesis of (3*R*,7*S*)-**372** started from (*S*)-(−)-citronellol **400**, which provided the required chiral center at C-3 in **372**. Routine transformations led to allylic alcohol **403**. Sharpless asymmetric epoxidation of **403** with diethyl L-(+)-tartrate, formation and recrystallization of the dinitrobenzoate derivatives **405** to improve the enantiomeric purity, oxidation of the resulting epoxyalcohol **404** and Horner–Wittig olefination gave epoxy-ester **406**. Palladium-catalyzed reductive cleavage of the epoxide gave **407**, establishing the chiral center at C-7 in **372**. Standard chain elongation steps and removal of the terminal, allylic functional group completed the carbon skeleton, and deprotection and oxidation of the resulting alcohol completed the synthesis, giving (3*R*,7*S*)-**372**. Following the same route, (3*R*,7*R*)-**372** was synthesized from (*S*)-(−)-citronellol **400** employing diethyl D-(−)-tartrate in the Sharpless asymmetric epoxidation step. Similarly, (3*S*,7*R*)-**372** and (3*S*,7*S*)-**372** were synthesized from (*R*)-(+)-citronellol (*R*)-**400**, employing diethyl D-(−)-tartrate and diethyl L-(+)-tartrate in the asymmetric epoxidation steps, respectively.

Following a similar route, Mori's group synthesized (6*S*,10*S*)-matsuone **371** from (*S*)-citronellol (*S*)-**400** and (6*R*,10*R*)-**371** from (*R*)-citronellol (*R*)-**400** (Scheme 64).[143]

Mori's group developed an improved synthesis of (3*S*,7*R*)-**372** from **420**, a versatile building block in pheromone synthesis (Scheme 65).[144] The epoxy alcohol **420** was prepared from optically active epoxide **419**, derived from pig pancreatic lipase (PPL)-catalyzed asymmetric hydrolysis of *meso*-diacetate **418**.[145] Stereoselective cleavage of the epoxy ring with trimethylaluminum gave **421** as the major regioisomer, which was converted to epoxide **422** by selective tosylation of the primary alcohol and base induced closure of the epoxide ring.

The copper-catalyzed coupling of epoxide **422** with (*S*)-2-methylbutylmagnesium bromide gave **423**, which after several standard operations afforded **424**, which was carried through to (3*S*,7*R*)-**372** as described in Mori's previous synthesis (Scheme 66).

Mori's group used a similar route to synthesize (6*R*,10*R*)-**371** (Scheme 67).[146] Thus, reaction of epoxide **422** with (*S*)-2,4-dimethylpentylmagnesium bromide gave alcohol **426**. An improvement over the previous synthesis was a more efficient removal of the primary hydroxyl group of **427**, which was converted to the corresponding mesylate, and then reduced with lithium triethylborohydride (Super-hydride) to **428**.

Lin and Xu developed enantioselective syntheses of all four stereoisomers of matsuone **371** from (*R*)- or (*S*)-citronellal, which provided the chiral center at C-10 in **371** (Scheme 68).[147] Several straightforward steps converted (*R*)-citronellal **429** to aldehyde (*R*)-**414**, which was then subjected to asymmetric Aldol reactions to insert the second stereocenter. The (*R*)- or (*S*)-configuration at C-6 of **371** was determined in the transition state by employing the (*Z*)- or (*E*)-boronates **432** and **433**, respectively. After protection of the hydroxyl group, alcohol **434** was subjected to ozonolysis and Wittig reaction to afford α,β-unsaturated ester **436**. Reaction of **436** with the anion of ethyldiphenylphosphine oxide gave the keto-phophonate **437** which was reduced with DIBAL to give alcohol **438** with good selectivity (*threo–erythro* = 6 : 1). The *threo* isomer underwent
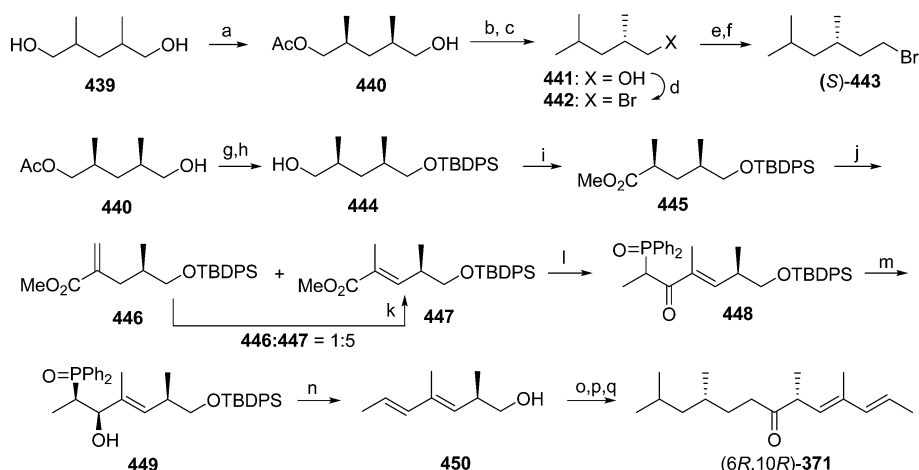
Scheme 69 Lin and Xu's enzyme-based synthesis of (6*R*,10*R*)-**371**. Reagents and conditions: (a) vinyl acetate, pig pancreatic lyase, wet THF, 28 °C; (b) TosCl, pyridine, 92%; (c) LiAlH₄, Et₂O, reflux, 90%; (d) Ph₃P, CBr₄, CH₂Cl₂, 79%; (e) Mg, Et₂O, reflux, then (HCHO)ₙ, reflux, 74%; (f) Ph₃P, CBr₄, CH₂Cl₂, 82%; (g) TBDPSiCl, imidazole, DMF, 98%; (h) K₂CO₃, MeOH, 40 °C, 95%; (i) i. Jones oxidation, 0 °C, ii. CH₂N₂, Et₂O, 0 °C, 83%; (j) i. LDA, THF, 0 °C; ii. PhSeBr, −78 °C, iii. H₂O₂, pyridine, CH₂Cl₂, 85%; (k) RhCl₃·3H₂O, EtOH, reflux, quant.; (l) Ph₂P(O)Et, *n*-BuLi, −78 °C, 98%; (m) NaBH₄, CeCl₃, EtOH, −78 °C, 92%; (n) NaH, DMF, 40 °C, 90%; (o) Swern oxidation; (p) (*S*)-1-bromo-3,5-dimethylhexane, Mg, THF, CuI, reflux, 55% over 2 steps; (q) Swern oxidation, 92%.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1105**

base-induced elimination and concomitant deprotection to give, after oxidation, the desired pheromone (6*R*,10*R*)-**371**. The other three stereoisomers were prepared in the same manner from alcohol (3*S*,4*R*,7*R*)-**434** and the corresponding alcohols from (*S*)-citronellal (*S*)-**429**.

Lin and Xu reported a second approach to the synthesis of (6*R*,10*R*)-**371**, in which both chiral centers of **371** were controlled *via* manipulation of a bifunctional building block **440**, obtained by the lipase-catalyzed transesterification of *meso*-2,4-dimethyl-1,5-propanediol **439** (Scheme 69).[148] Thus, bromide **443** was prepared from **440** in five routine steps. Synthesis of the second required synthon **450** also started with **440**. Thus, methyl ester **445**, obtained from **440** in three steps, was subjected to phenylselenenylation and oxidative

elimination to afford alkene **447**, along with a small amount of isomeric **446**, which was converted quantitatively to **447** by refluxing in anhydrous ethanol with catalytic RhCl₃. Treatment of α,β-unsaturated ester **447** with the lithium salt of ethyl diphenylphosphine oxide, followed by regioselective reduction with NaBH₄/CeCl₃ furnished **449** with good selectivity (*threo–erythro* = 4 : 1). After removal of the *erythro* isomer by flash chromatography, the *threo* isomer underwent base-induced elimination and concomitant deprotection to give the diene homoallylic alcohol **450**. Oxidation to the aldehyde, copper-catalyzed reaction with the Grignard reagent derived from **443**, and Swern oxidation gave (6*R*,10*R*)-**371**.

Zegelman and coworkers reported a relatively short synthesis of (±)-**374** and (±)-**375**, the racemates of both
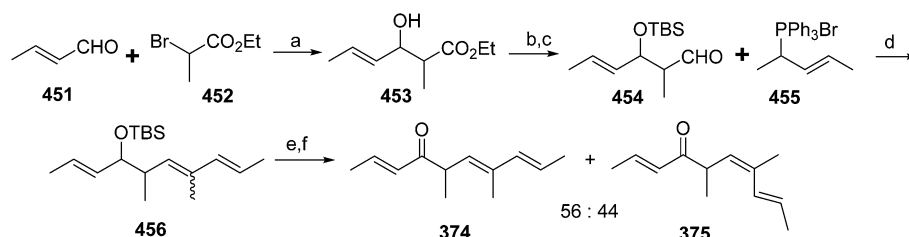
Scheme 70 Zegelman's synthesis of (±)-**374** and (±)-**375**. Reagents and conditions: (a) Zn–Cu, Et₂O, room temp, 84%; (b) TBDMSiCl, imidazole, DMF, room temp; (c) DIBAL, PhMe, −78 °C, 63% over 2 steps; (d) *n*-BuLi, THF, 0–10 °C, 72%; (e) TBAF, THF, room temp; (f) PCC, NaOAc, CH₂Cl₂, room temp, 54% over 2 steps.
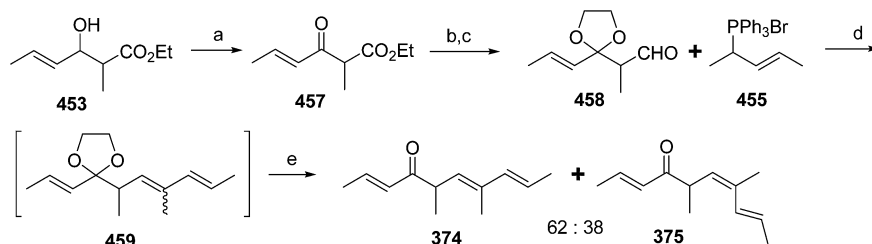
Scheme 71 Zegelman's modified synthesis of (±)-**374** and (±)-**375**. Reagents and conditions: (a) PCC, NaOAc, CH₂Cl₂, room temp, 62%; (b) HO(CH₂)₂OH, PTSA, CHCl₃, reflux; (c) DIBAL, PhMe, −78 °C, 48% for 2 steps; (d) *n*-BuLi, THF, 0–10 °C; (e) aq. HCl, THF, room temp, 51% over 2 steps.
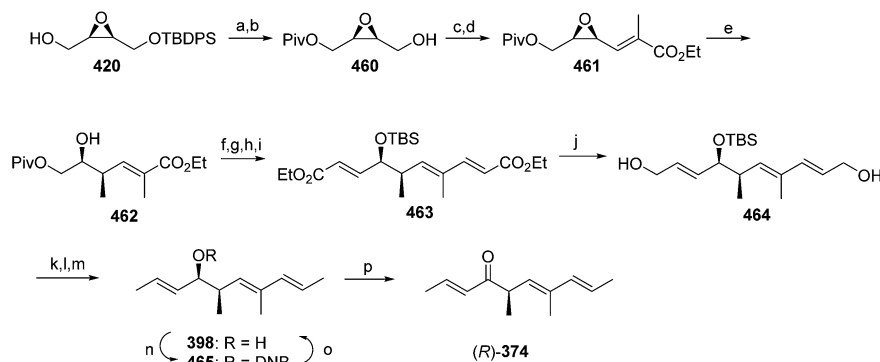
Scheme 72 Mori's enantioselective synthesis of (*R*)-**374**. Reagents and conditions: (a) PivCl, pyridine, CH₂Cl₂, 93%; (b) TBAF, HF, H₂O/THF, 95%; (c) Swern oxidation; (d) Ph₃P=C(Me)CO₂Et, THF, 63% over 2 steps; (e) Me₃Al (10 equiv.), H₂O (6 equiv.), CH₂Cl₂, 89%; (f) TBDMSiCl, imidazole, DMF, 91%; (g) DIBAL, Et₂O, 88%; (h) Swern oxidation, (i) (EtO)₂P(O)CH₂CO₂Et, *n*-BuLi, THF, 61% over 2 steps; (j) DIBAL, Et₂O, 98%; (k) *n*-BuLi, Mes₂O, THF; (l) LiBEt₃H, THF; (m) TBAF, THF, SiO₂–AgNO₃ chromatography; (n) 3,5-dinitrobenzoyl chloride, pyridine, recrystallization; (o) K₂CO₃, MeOH/THF; (p) Swern oxidation, 92%.

components of the sex pheromone of *M. josephi* (Scheme 70).[149] Reformatsky reaction between crotonaldehyde **451** and ethyl 2-bromopropanoate **452** gave hydroxyester **453**, which was protected and reduced to aldehyde **454**. Wittig reaction of **454** with the anion from allylic phosphonium salt **455** gave a mixture of (*E*)- and (*Z*)-isomers. Removal of the TBDMS protecting group followed by oxidation with pyridinium chlorochromate produced a mixture of (±)-**374** and (±)-**375** in a ratio of 56 : 44.

A modified route led straight from the Wittig product to the pheromone (Scheme 71).[149]

Bioassays showed that pheromonal activity was due mainly to the *E*-isomer (±)-**374**, whereas the minor *Z*-isomer (±)-**375** was of low activity, but not inhibitory.

Mori and Amaike reported an enantioselective synthesis of both enantiomers of **374** (Scheme 72),[150] which used many of the same steps and synthons as in his syntheses of matsuone **371**. Thus, the reactive ends of **420** were interchanged by sequential protection of the free hydroxyl and deprotection of the other hydroxyl, followed by Swern oxidation and Horner–Wittig reaction. In a key step, cleavage of the epoxy ring of **461** with trimethylaluminum in the presence of a small amount of
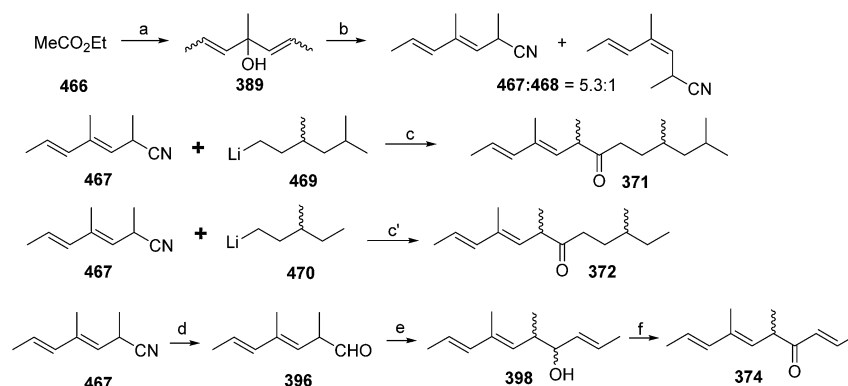
Scheme 73 Watanabe's concise syntheses of **371**, **372**, and **374** as racemic and diastereomeric mixtures. Reagents and conditions: (a) 1-propenylmagnesium bromide, 95%; (b) TMSCN, TMSOTf, 56% (**467** : **468** = 5.3 : 1); (c) Et₂O, then H₃O⁺, 70%; (c′) Et₂O, then H₃O⁺, 73%; (d) DIBAL, then H₃O⁺; (e) (*E*)-1-propenyllithium, THF, −78 °C, 67% over 2 steps; (f) PDC, DMF, 49%.
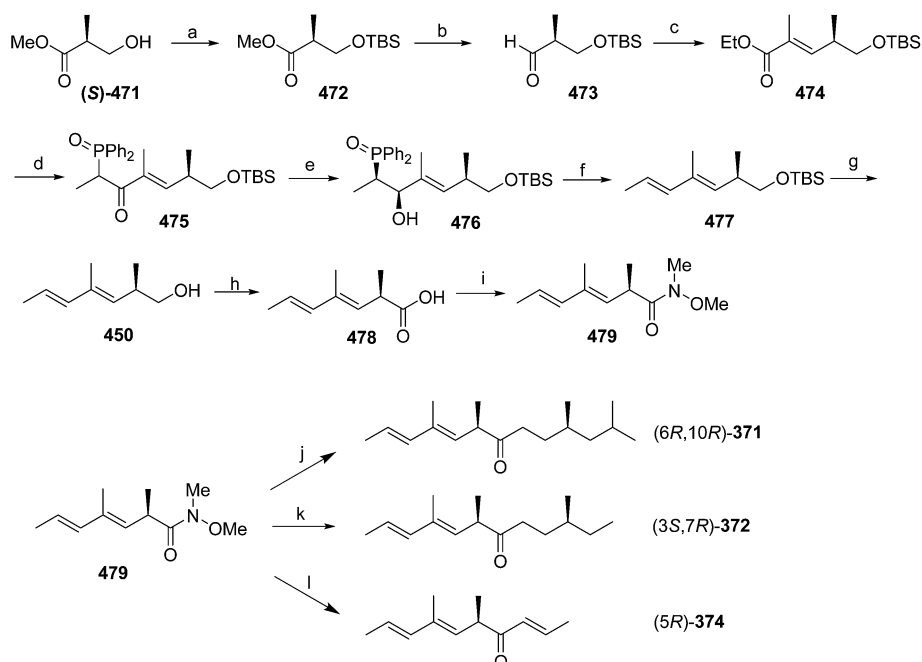
Scheme 74 The Mori group's syntheses of (6*R*,10*R*)-**371**, (3*S*,7*R*)-**372**, and (5*R*)-**374** from one key intermediate. Reagents and conditions: (a) TBDMSiCl, imidazole, 98%; (b) DIBAL, hexanes/CH₂Cl₂; (c) Ph₃P=C(Me)CO₂Et, benzene, 83% over 2 steps (*E−Z* = 95 : 5); (d) Ph₂P(O)Et, *n*-BuLi, THF, 98%; (e) NaBH₄, CeCl₃·7H₂O, EtOH, then silica gel chromatography, 66%; (f) NaH, DMF; (g) TBAF, THF, 90% over 2 steps; (h) PDC, DMF; (i) HNMe(OMe)·HCl, EDC·HCl, DMAP, diisopropylethylamine, CH₂Cl₂, 38% over 2 steps; (j) (*S*)-3,5-dimethylhexylMgBr, THF, 89%; (k) (*S*)-3-methylpentylMgBr, THF, 90%; (l) (*E*)-propenylMgBr, THF, 80%.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | **1107**

water regio- and stereoselectively placed the chiral methyl group, giving **462** in excellent yield. Chain elongation, removal of the two terminal hydroxyls, deprotection and oxidation of the resulting secondary alcohol afforded (*R*)-**374**. (*S*)-**374** was prepared in analogous fashion from the antipode of **460**, in turn prepared from **420** in 4 simple steps.

Because the unnatural isomers were not inhibitory, Watanabe developed short syntheses of *Matsucoccus* sex pheromones **371**, **372**, and **374** as mixtures of isomers for a study of practical pest control (Scheme 73).[151] The key reaction was Lewis acid-mediated S_N2' cyanation of symmetric tertiary alcohol **389** to afford common intermediate **467** as the major product, which could be separated from the undesired minor isomer **468** by flash chromatography. Nucleophilic addition of (±)-3,5-dimethylhexyllithium **469** and (±)-3-methylpentyllithium **470** followed by aqueous hydrolysis of the resulting imines afforded **371** and **372** respectively, as racemic and diastereomeric mixtures. However, reaction of **467** with (*E*)-1-propenyllithium resulted in the recovery of **467**, presumably due to the lower nucleophilicity of the vinyl anion. Therefore, **467** was reduced to aldehyde **396**, which then underwent nucleophilic addition of (*E*)-1-propenyllithium, followed by PDC oxidation to give racemic **374**.

Mori's group developed yet another synthetic route to (6*R*,10*R*)-**371**, (3*S*,7*R*)-**372**, and (5*R*)-**374** with a synthon from the chiral pool, methyl (*S*)-2-methyl-3-hydroxypropanoate **471** (Scheme 74).[152] The route incorporated a number of steps from previous routes. Thus, several routine steps gave α,β-unsaturated ester **474**, which was converted to diene homoallylic alcohol **450** as described by Lin and Xu (Scheme 69). Two steps converted **450** to the key intermediate, Weinreb amide **479**, followed by coupling with appropriate organometallic reagents to give (6*R*,10*R*)-**371**, (3*S*,7*R*)-**372**, and (5*R*)-**374**.

Somewhat surprisingly, given the interesting challenges in the syntheses of the pine bast scale pheromones, there have been no further syntheses reported since 2003.

4 Practical applications of scale and mealybug pheromones

Pheromones can be used for detection and sampling of scales and mealybugs, and potentially for direct control. In practice, the pheromones are ideal for detection and sampling because they are powerful and species-specific attractants, very small amounts of pheromone (usually a few micrograms) are required per lure, and the pheromone structures are quite stable so that pheromone lures can have field lifetimes of several months or more. Because such small doses of pheromone are required per lure, relatively expensive, multistep syntheses of the pheromones are still feasible. For example, at a dose rate of 20 micrograms per lure, 1 g of pheromone is sufficient for 50 000 lures. Furthermore, in most cases, male scales and mealybugs are insensitive to the presence of the unnatural enantiomers or other stereoisomers, and so less expensive racemic pheromone can be used for most applications. Consequently, pheromone lures for many species are available from commercial suppliers.

Furthermore, because the pheromones are species-specific, no special expertise is required to identify the trapped insects; if male scales or mealybugs are detected in a pheromone-baited trap, it is almost certain that they are males of the species that produces that particular pheromone. The one documented exception has been the cross-attraction of the scale *Quadraspidiotus zonatus* to the pheromone of the San Jose scale, *Q. perniciosus*.[153] Conversely, field trials with composite lures containing the pheromones of citrus, obscure, and longtailed mealybugs showed that there was minimal interference among these pheromones, suggesting that it may be possible to use "generic" mealybug lures that attract several species simultaneously.[154] This may be useful in crops that can be infested with several species simultaneously because the control measures are usually the same no matter what species is present.

In terms of detection, pheromone-baited sticky traps can detect very low level and dispersed populations that would be virtually impossible to find by any other method. For example, the pheromone of the passionvine mealybug was used to show that this species had invaded Florida from the Caribbean islands, but populations were extremely low, possibly as a result of control by natural enemies.[155] Pheromone-baited traps were also used as a sensitive method of tracking the expanding ranges of the invasive vine mealybug as it invaded California vineyards,[24] and the pink hibiscus mealybug in several US states.[156] As a proactive measure against another invasive species, the pheromone of the albopicta scale was identified so that it could be used as a tool for early detection of this insect if it invaded California from Mexico.[102] As a further example of the powerful attraction of these pheromones, when developing protocols for using yellow scale pheromone to detect infestations, lure doses had to be lowered to 1–5 micrograms per lure, both to prevent traps from becoming completely covered with male scales and making them very difficult to count, and because higher doses apparently attracted male scales from substantial distances, providing the false impression that the orchard in which the trap was hung was infested, when the actual infestation might be some distance away.[157]

Pheromone-baited traps have also been used extensively in monitoring established populations of scales and mealybugs, particularly for monitoring the flight phenology of adults as an aid to making control decisions. It has proven more difficult to correlate trap catches with infestation levels to develop economic thresholds that, when exceeded, trigger control measures, for two interlinked reasons. First, as mentioned above, the pheromones are so powerful that they can attract males from some distance away, and second, scales and mealybugs tend to form highly clumped distributions, rather than being randomly distributed throughout a crop. Thus, high trap catches may not necessarily indicate high populations throughout the crop. Nevertheless, by judicious use of lower doses of pheromone to decrease the range of attraction of pheromone lures, in some cases it has been possible to develop good correlations between pheromone trap catches and infestation levels as determined by visual counts or other means.[24,154,158] The interested reader is referred to a more extensive discussion of the development of pheromone-baited traps for monitoring purposes.[3]

1108 | *Nat. Prod. Rep.*, 2015, **32**, 1067–1113

This journal is © The Royal Society of Chemistry 2015

Pheromones can be used to control insect populations in at least three ways: (1) disruption of mating by permeating the atmosphere of a crop with pheromone so that males cannot find females, (2) mass trapping, in which sufficient numbers of pheromone-baited traps are deployed to remove a large percentage of males, and (3) attract-and-kill, in which pheromone is mixed with a toxicant in a matrix applied in multiple droplets throughout a crop, so that males that are attracted and contact the droplets are killed. The feasibility of using these three methods for controlling scales and mealybugs hinges on a number of economic and biological factors. First and foremost, if the pheromones cannot be made in multikilo scale at a price that is competitive with other control measures, mating disruption at least will not be economically feasible. For mass trapping and attract-and-kill, which use much smaller quantities of pheromone, the costs of deploying and servicing large arrays of traps or attract-and-kill droplets may still present significant barriers to implementation. For attract-and-kill, there is also the additional requirement that formulations containing a toxicant may be subject to additional registration requirements. A number of other points also must be considered, including:

1. The number of generations per year, and the overall adult activity periods. Clearly the costs and technical issues relating to control of a species with a single generation per year and a relatively short adult activity period will be different than those involved in controlling a multivoltine species that is present for many months.

2. Female scales and mealybugs can live for several months, during which time they will continue to produce pheromone until they are mated. Thus, even if a substantial percentage of the available males is removed, the remaining males may still mate many of the available females because males can mate multiple times.[159] Thus, almost all of the males must be removed or otherwise prevented from mating in order to achieve good control.

3. Conversely, adult males live for at most a few days under field conditions.

4. The highly clumped, non-random distribution of scales and mealybugs will also hinder pheromone-based control measures, because males and females will be close together in clumps, particularly in heavy infestations. Thus, males may serendipitously find females.

5. Pheromone formulations must have effective field lifetimes of many weeks. However, these issues are readily dealt with because scale and mealybug pheromones are relatively stable, and longevity of lures or other formulations can be readily adjusted by appropriate choice of release device.

6. The size of the market for any one pheromone is relatively small. That is, the species specificity of pheromones is a major advantage from the viewpoint of nontarget effects, and a major disadvantage because, unlike a broad-spectrum insecticide which is effective against multiple insects on numerous crops, a scale or mealybug pheromone will only affect a single target species.

In practice, mass trapping has only been attempted with the citrus mealybug. Whereas numbers of males were reduced in treated blocks relative to controls, damage levels were unaffected. Furthermore, it appeared that the traps were attracting more males into the treated areas, partially counteracting male removal by the traps.[160]

Mating disruption was first explored for California red scale in the 1980s,[161] and has been followed up in a series of studies in the last decade[162] which demonstrated decreased infestation levels and significantly less crop damage in treated areas *versus* untreated controls. Thus, this system has considerable promise biologically, if the economic hurdles to producing large quantities of the pheromone can be surmounted. Similarly, a mating disruption trial with San Jose scale was moderately successful but was not pursued further because of the prohibitive cost of the pheromone.[163]

The vine mealybug represents the most successful and to date only commercially viable use of mating disruption for control of scales or mealybugs, primarily because the pheromone can be made economically in large scale. Initial results from trials using a microencapsulated formulation were reported in 2006,[164,165] and since then, other dispenser types have been developed, including reservoir dispensers and "puffers" that release puffs of pheromone at timed intervals.[166–168] Vine mealybug mating disruption is most effective when used in combination with early-season insecticide treatments to reduce initial populations to low levels, which are then maintained with the disruptant treatment.[167] In California, mating disruption of vine mealybug is now used on thousands of acres of wine grapes annually, and the active formulation is being registered for use in 8 additional countries by the manufacturer (Suterra LLC, Bend OR, USA).

5   Conclusions

We hope that this review has given the reader an appreciation of the fascinating chemistry of scale and mealybug pheromones. Although the biosyntheses of these compounds have not yet been studied, it is clear that they fall into two major groups, based on irregular terpenoid biosynthetic pathways for the diaspidid and pseudococcid species, and polyketides for the margorodid species. Synthesis of these pheromones in pure form can be challenging, requiring control of one to several chiral centers and the geometries of di- and trisubstituted alkenes. However, because most species are not inhibited by the presence of unnatural isomers, much shorter, non-stereoselective syntheses can be used to prepare "technical" grade pheromones which are usually adequate for practical purposes. Although numerous stereoselective and non-stereoselective syntheses of the various compounds have been developed, faster, more efficient, and more economical syntheses are still highly desirable in order to accelerate the development and adoption of these useful natural products for pest management.

Following the progression of syntheses from the late 1970s to the present is also an interesting study in the development of synthetic chemistry methodology. For example, syntheses of chiral pheromones in the 1980s made extensive use of chiral synthons available from natural sources, or derivatives thereof.
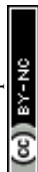
Alternatively, diastereomeric derivatives were formed from reaction of a chiral derivatizing agent with the racemate of the desired synthon, followed by separation of the diastereomers by fractional crystallization or chromatography (Schemes 21, 51 and 61). The desired chiral synthon was then released from the purified diastereomer by hydrolysis or some other appropriate reaction. Both of these methods were limited by the number and types of chiral synthons or reagents that were available from natural sources.

This pool was then expanded somewhat by using enzymes to produce chiral synthons, either by kinetic resolution of racemates (*i.e.*, enzyme-catalyzed reactions selective for one enantiomer) (Schemes 19, 53, and Madeira mealybug pheromone synthesis), or the related enzyme-catalyzed desymmetrization of achiral compounds containing two or more chiral centers such as meso compounds (Schemes 15, 65, 69 and 72).

However, the great leap forward that largely freed synthetic chemists from the limitations of the chiral synthons available from natural sources came with the introduction and widespread adoption of methods for asymmetric induction in achiral precursors, by a variety of methods. For example, temporarily attached chiral auxiliaries could be used to direct the stereoselective formation of new bonds (Schemes 27A, 27B, 29B, 38 and 56). Alternatively, chiral catalysts were developed that favored the production of one particular stereoisomer due to the three-dimensional assembly of the various reactants in the transition state (Scheme 19). In this day and age when the use of chiral catalysts is commonplace, it may be hard for younger chemists to appreciate the impact of a reaction such as the Sharpless asymmetric epoxidation on the field of organic synthesis. It was truly a turning point, in terms of demonstrating that generation of chirality by asymmetric induction was not only possible but practically feasible. In the intervening years, literally thousands of stereoselective reactions and applications have been developed, that have revolutionized the science of organic synthesis, both for small-scale applications in research laboratories, and in large-scale industrial applications.

Another area in which the advances in synthetic methodology are apparent from this review is in the formation of carbon–carbon bonds. Thus, in the syntheses from the 1980s, we see some of the first applications of copper-catalyzed reactions, or reactions using stoichiometric amounts of organocuprates. Subsequent years illustrate not only the expansion of the breadth and scope of these reactions, but introduction of additional coupling methods using other organometallics such as organostannanes (Schemes 14, 18, 29A and 46) and organozinc reagents (Schemes 7, 16 and 19), and reactions catalyzed by palladium complexes (Scheme 19). These include not only nucleophilic displacements, but also the regio- and stereoselective addition of organometallics to alkenes and alkynes (Schemes 1, 3, 7, 19 and 49). Olefin metathesis has also been used in pheromone synthesis (Scheme 50).

However, it is also useful to keep in mind that old reactions are not necessarily outdated or inefficient, particularly given that chiral catalysts containing rare elements such as palladium, gold, and ruthenium can be breathtakingly expensive.

Rather, the different routes to a target compound, and the reagents required, should be carefully considered in light of the particular requirements of a project, including the required amount and stereochemical purity of a target compound.

To date, many of the pheromones described above have proven enormously useful for detection of infestations, particularly for invasive species. They have also found widespread use in monitoring scale and mealybug populations and their seasonal dynamics in numerous crops, providing valuable data on population cycles that inform pest management decisions. However, the use of pheromone-based methods for direct crop protection, particularly on a large scale, will not be competitive with other crop protection methods for a number of species because of the prohibitive cost of producing the required pheromones in sufficient quantities. This barrier to implementation may be insurmountable for some species because of the structural complexity of their pheromones.

6 References

1 J. C. Franco, A. Zada and Z. Mendel, in *Biorational control of arthropod pests*, ed. I. Ishaaya and A. R. Horowitz, Springer Science + Business Media B.V., 2009, pp. 233–278.

2 K. Mori, in *The Total Synthesis of Natural Products*, ed. J. ApSimon, John Wiley & Sons, New York, 1992, vol. 9, pp. 280–332.

3 E. Dunkelblum, in *Pheromones of Non-Lepidopteran Insects Associated with Agricultural Plants*, ed. J. Hardie and A. K. Minks, CABI publishing, 1999, pp. 251–276.

4 J. G. Millar, K. M. Daane, J. S. McElfresh, J. A. Moreira and W. J. Bentley, in *Semiochemicals in Pest and Weed Control*, ed. R. J. Petroski, M. R. Tellez and R. W. Behle, ACS Symposium Series 906, American Chemical Society, Washington, DC, 2005, pp 11−27.

5 Y. Zou, S. P. Chinta and J. G. Millar, in *Pest Management with Natural Products*, ed. J. J. Beck, J. R. Coats, S. O. Duke and M. E. Koivunen, ACS Symposium Series 1141, American Chemical Society, Washington, DC, 2013, pp. 125–143.

6 T. Negishi, M. Uchida, Y. Tamaki, K. Mori, T. Ishiwatari, S. Asano and K. Nakagawa, *Appl. Entomol. Zool.*, 1980, **15**, 328.

7 D. S. Moreno, J. Fargerlund and J. G. Shaw, *J. Econ. Entomol.*, 1976, **69**, 292.

8 B. A. Figadère, J. S. McElfresh, D. Borchardt, K. M. Daane, W. Bentley and J. G. Millar, *Tetrahedron Lett.*, 2007, **48**, 8434.

9 A. B. Attygalle, in *Methods in Chemical Ecology. Volume 1. Chemical Methods*, ed. J. G. Millar and K. F. Haynes, Chapman and Hall, Norwell, MA, 1998, pp. 207–294.

10 J. Tabata, Y. Narai, N. Sawamura, S. Hiradate and H. Sugie, *Naturwissenschaften*, 2012, **99**, 567.

11 M. J. Gieselmann, R. E. Rice, R. A. Jones and W. L. Roelofs, *J. Chem. Ecol.*, 1979, **5**, 891.

12 R. J. Anderson, H. R. Chinn, K. Gill and C. A. Henrick, *J. Chem. Ecol.*, 1979, **5**, 919.

13 R. J. Anderson, M. J. Gieselmann, H. R. Chinn, K. G. Adams, C. A. Henrick, R. E. Rice and W. L. Roelofs, *J. Chem. Ecol.*, 1981, **7**, 695.

14 K. H. Yong, J. A. Lotoski and J. M. Chong, *J. Org. Chem.*, 2001, **66**, 8248.

15 (*a*) M. Alderdice, C. Spino and L. Weiler, *Tetrahedron Lett.*, 1984, **25**, 1643; (*b*) M. Alderdice, C. Spino and L. Weiler, *Can. J. Chem.*, 1993, **71**, 1955.

16 D. A. Lombardo and A. C. Weedon, *Tetrahedron Lett.*, 1986, **27**, 5555.

17 V. V. Veselovskii, S. P. Skorobogatov, M. A. Novikova and A. M. Moiseenkov, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1990, 591, English translation: *Russ. Chem. Bull.*, 1990, **39**, 513.

18 A. M. Moiseenkov, R. I. Ishchenko, V. V. Veselovskii, V. N. Odinokov, E. V. Polunin, B. G. Kovalev, B. A. Cheskis and G. A. Tolstikov, *Khim. Prir. Soedin.*, 1989, 422, English translation: *Chem. Nat. Compd.*, 1989, **25**, 366.

19 D. Ferroud, J. M. Gaudin and J. P. Genet, *Tetrahedron Lett.*, 1986, **27**, 845.

20 U. P. Dhokte and A. S. Rao, *Synth. Commun.*, 1988, **18**, 811.

21 L. Novak, L. Poppe, C. Szantay and E. Szabo, *Synthesis*, 1985, 939.

22 V. N. Odinokov, O. S. Kukovinets, R. A. Zainullin, E. Yu. Tsyglintseva, V. R. Sultanmuratova, V. V. Veselovskii, B. A. Dragan, T. Ya. Rubinskaya, B. A. Cheskis, A. M. Moiseenkov and G. A. Tolstikov, *Khim. Prir. Soedin.*, 1989, 419, English translation: *Chem. Nat. Compd.*, 1989, **25**, 364.

23 G. Cardillo, A. D'Amico, M. Orena and S. Sandri, *J. Org. Chem.*, 1988, **53**, 2354.

24 J. G. Millar, K. M. Daane, J. S. McElfresh, J. A. Moreira, R. Malakar-Kuenen, M. Guillen and W. J. Bentley, *J. Econ. Entomol.*, 2002, **95**, 706.

25 A. Zada and M. Harel, *Tetrahedron: Asymmetry*, 2004, **15**, 2339.

26 A. Zada and E. A. Dunkelblum, *Tetrahedron: Asymmetry*, 2006, **17**, 230.

27 D. M. Hinkens, J. S. McElfresh and J. G. Millar, *Tetrahedron Lett.*, 2001, **42**, 1619.

28 A. Zada, E. Dunkelblum, F. Assael, M. Harel, M. Cojocaru and Z. Mendel, *J. Chem. Ecol.*, 2003, **29**, 977.

29 A. Zhang, D. Amalin, S. Shirali, M. S. Serrano, R. A. Franqui, J. E. Oliver, J. A. Klun, J. R. Aldrich, D. E. Meyerdirk and S. L. Lapointe, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, **101**, 9601.

30 I. de Alfonso, E. Hernandez, Y. Velazquez, I. Navarro and J. Primo, *J. Agric. Food Chem.*, 2012, **60**, 11959.

31 H. Sugie, M. Teshiba, Y. Narai, T. Tsutsumi, N. Sawamura, J. Tabata and S. Hiradate, *Appl. Entomol. Zool.*, 2008, **43**, 369.

32 J. Tabata, *Appl. Entomol. Zool.*, 2013, **48**, 229.

33 H.-Y. Ho, C.-C. Hung, T.-H. Chuang and W.-L. Wang, *J. Chem. Ecol.*, 2007, **33**, 1986.

34 J. G. Millar, *Tetrahedron Lett.*, 2008, **49**, 315.

35 J. G. Duboudin and B. Jousseaume, *J. Organomet. Chem.*, 1979, **168**, 1.

36 B. A. Bierl-Leonhardt, D. S. Moreno, M. Schwarz, H. S. Forster, J. R. Plimmer and E. D. DeVilbiss, *Life Sci.*, 1980, **27**, 399.

37 B. A. Bierl-Leonhardt, D. S. Moreno, M. Schwarz, H. S. Forster, J. R. Plimmer and E. D. DeVilbiss, *J. Chem. Ecol.*, 1982, **8**, 689.

38 M. Uchida, K. Nakagawa, T. Negishi, S. Asano and K. Mori, *Agric. Biol. Chem.*, 1981, **45**, 369.

39 R. I. Ishchenko, V. V. Veselovskii, A. M. Moiseenkov, B. A. Cheskis and B. G. Kovalev, *Khim. Prir. Soedin.*, 1989, 132, English translation: *Chem. Nat. Compd.*, 1989, **25**, 118.

40 K. Mori and H. Ueda, *Tetrahedron*, 1981, **37**, 2581.

41 N. Nakagawa and K. Mori, *Agric. Biol. Chem.*, 1984, **48**, 2799.

42 M. Larcheveque and Y. Petit, *Bull. Soc. Chim. Fr.*, 1989, 130.

43 L. Skattebol and Y. Stenstrom, *Acta Chem. Scand.*, 1989, **43**, 93.

44 P. Baeckstroem and L. Li, *Synth. Commun.*, 1990, **20**, 1481.

45 S. K. Kang and C. S. Park, *Org. Prep. Proced. Int.*, 1990, **22**, 627.

46 (*a*) T. Cohen and M. Bhupathy, *Acc. Chem. Res.*, 1989, **22**, 152; (*b*) D. W. McCullough, M. Bhupathy, E. Piccolino and T. Cohen, *Tetrahedron*, 1991, **47**, 9727.

47 P. Baeckstrom, F. Bjokling, H.-E. Hogberg and T. Norin, *Acta Chem. Scand.*, 1984, **B38**, 779.

48 Y. Fall, N. V. Bac and Y. Langlois, *Tetrahedron Lett.*, 1986, **27**, 3611.

49 M. J. Gieselmann, D. S. Moreno, J. Fargerlund, H. Tashiro and W. L. Roelofs, *J. Chem. Ecol.*, 1979, **5**, 27.

50 R. J. Anderson and C. A. Henrick, *J. Chem. Ecol.*, 1979, **5**, 773.

51 T. Suguro, W. L. Roelofs and K. Mori, *Agric. Biol. Chem.*, 1981, **45**, 2509.

52 S. Masuda, S. Kuwahara, T. Suguro and K. Mori, *Agric. Biol. Chem.*, 1981, **45**, 2515.

53 K. Mori and S. Kuwahara, *Tetrahedron*, 1982, **38**, 521.

54 E. Alvarez, T. Cuvigny, C. Harve du Penhoat and M. Julia, *Tetrahedron*, 1988, **44**, 119.

55 J. G. Millar, *Tetrahedron Lett.*, 1989, **30**, 4913.

56 R. Baudouy and M.-R. Sancho, *Tetrahedron*, 1991, **47**, 10015.

57 (*a*) S. Harusawa, H. Osaki, S. Takemura, R. Yoneda and T. Kurihara, *Tetrahedron Lett.*, 1992, **33**, 2543; (*b*) S. Harusawa, S. Takemura, H. Osaki, R. Yoneda and T. Kurihara, *Tetrahedron*, 1993, **49**, 7657; (*c*) S. Harusawa, S. Takemura, R. Yoneda and T. Kurihara, *Tetrahedron*, 1993, **49**, 10577.

58 Z. Xu and E. Negishi, *Org. Lett.*, 2008, **10**, 4311.

59 W. Roelofs, M. Gieselmann, A. Carde, H. Tashiro, D. S. Moreno, C. A. Henrick and R. J. Anderson, *J. Chem. Ecol.*, 1978, **4**, 211.

60 (*a*) B. B. Snider and D. Rodini, *Tetrahedron Lett.*, 1978, **19**, 1399; (*b*) B. B. Snider and G. B. Phillips, *J. Org. Chem.*, 1983, **48**, 464.

61 R. J. Anderson, K. G. Adams, H. R. Chinn and C. A. Henrick, *J. Org. Chem.*, 1980, **45**, 2229.

62 R. Baudouy and C. Maliverney, *Tetrahedron*, 1988, **44**, 471.

63 D. Becker and Y. Sahali, *Tetrahedron*, 1988, **44**, 4541.

64 V. A. Dragan, V. V. Veselovskii and A. M. Moiseenkov, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1989, 1143, English translation: *Russ. Chem. Bull.*, 1989, **38**, 1038.

65 X.-M. Zhang, A. Archelas and R. Furstoss, *Tetrahedron: Asymmetry*, 1992, **3**, 1373.

66 L. Auer, C. Weymuth and R. Scheffold, *Helv. Chim. Acta*, 1993, **76**, 810.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | 1111

67 P. Kefalas and N. Ragoussis, *Synthesis*, 1995, 644.

68 R. Baudouy and P. Prince, *Tetrahedron*, 1989, **45**, 2067.

69 J. H. Hutchinson and T. Money, *Can. J. Chem.*, 1985, **63**, 3182.

70 M. Whittaker, C. R. McArthur and C. C. Leznoff, *Can. J. Chem.*, 1985, **63**, 2844.

71 P. Mangeney, A. Alexakis and J. F. Normant, *Tetrahedron Lett.*, 1987, **28**, 2363.

72 S. M. Kher and G. H. Kulkarni, *Synth. Commun.*, 1990, **20**, 495.

73 W. C. Still and A. Mitra, *J. Am. Chem. Soc.*, 1978, **100**, 1927.

74 W. Oppolzer and T. Stevenson, *Tetrahedron Lett.*, 1986, **27**, 1139.

75 M. P. Cooke Jr and D. L. Burman, *J. Org. Chem.*, 1982, **47**, 4955.

76 D. Caine and E. Crews, *Tetrahedron Lett.*, 1984, **25**, 5359.

77 J. Celebuski and M. Rosenblum, *Tetrahedron*, 1985, **41**, 5741.

78 A. A. Vasil'ev, A. L. Vlasyuk, G. V. Kryshtal and E. P. Serebryakov, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1995, 2026, English translation: *Russ. Chem. Bull.*, 1995, **44**, 1946.

79 M. J. Hesse, C. P. Butts, C. L. Willis and V. K. Aggarwal, *Angew. Chem., Int. Ed.*, 2012, **51**, 12444.

80 R. R. Heath, J. R. McLaughlin, J. H. Tumlinson, T. R. Ashley and R. E. Doolittle, *J. Chem. Ecol.*, 1979, **5**, 941.

81 J. Einhorn, H. Bianchi and C. Benassy, *C. R. Acad. Sci., Ser. III*, 1983, **296**, 861.

82 J. R. McLaughlin, *J. Chem. Ecol.*, 1990, **16**, 749.

83 R. R. Heath, R. E. Doolittle, P. E. Sonnet and J. H. Tumlinson, *J. Org. Chem.*, 1980, **45**, 2910.

84 N. Ishibashi, M. Nakamura, F. Mochizuki and T. Fukumoto, *U.S. Pat.* Appl. Publ.,20110172304 A1 20110714, 2011.

85 H.-Y. Ho, B. S. Kuarm, C.-H. Ke, Y.-K. Ma, H.-J. Lee, C.-C. Cheng, K. K. Liu and J. G. Millar, *J. Chem. Ecol.*, 2014, **40**, 379.

86 (*a*) R. R. Johnson and J. A. Nicholson, *J. Org. Chem.*, 1965, **30**, 2918; (*b*) A. Kohda, K. Nagayoshi, K. Maemoto and T. Sato, *J. Org. Chem.*, 1983, **48**, 425; (*c*) O. S. Park, H. J. Kim, W. K. Chae and W. Y. Lee, *Bull. Korean Chem. Soc.*, 1993, **14**, 639.

87 T. Kitahara, T. Matsuoka, H. Kiyota, Y. Warita, H. Kurata, A. Horiguchi and K. Mori, *Synthesis*, 1994, 692.

88 H.-Y. Ho, Y.-T. Su, C.-H. Ko and M.-Y. Tsai, *J. Chem. Ecol.*, 2009, **35**, 724.

89 H.-Y. Ho, C.-H. Ko, C.-C. Cheng and Y.-T. Su, *J. Econ. Entomol.*, 2011, **104**, 823.

90 A. M. El-Sayed, C. R. Unelius, A. Twidle, V. Mitchell, L.-A. Manning, L. Cole, D. M. Suckling, M. F. Flores, T. Zaviezo and J. Bergmann, *Tetrahedron Lett.*, 2010, **51**, 1075.

91 C. R. Unelius, A. M. El-Sayed, A. Twidle, B. Bunn, T. Zaviezo, M. F. Flores, V. Bell and J. Bergmann, *J. Chem. Ecol.*, 2011, **37**, 166.

92 B. A. Bierl-Leonhardt, D. S. Moreno, M. Schwarz, J. Fargerlund and J. R. Plimmer, *Tetrahedron Lett.*, 1981, **22**, 389.

93 A. Zada, E. Dunkelblum, M. Harel, F. Assael, S. Gross and Z. Mendel, *J. Econ. Entomol.*, 2004, **97**, 361.

94 L. C. Passaro and F. X. Webster, *J. Agric. Food Chem.*, 2004, **52**, 2896.

95 P. H. J. Carlsen and W. Odden, *Acta Chem. Scand., Ser. B*, 1984, **38**, 501.

96 L. Lombardo, *Org. Synth.*, 1987, **65**, 81.

97 O. S. Kukovinets, T. I. Zvereva, V. G. Kasradze, F. Z. Galin, L. L. Frolova, A. V. Kuchin, L. V. Spirikhin and M. I. Abdullin, *Chem. Nat. Compd.*, 2006, **42**, 216.

98 B. C. Ranu and R. Chakraborty, *Tetrahedron Lett.*, 1990, **31**, 7663.

99 T. Arai, H. Sugie, S. Hiradate, S. Kuwahara, N. Itagaki and T. Nakahata, *J. Chem. Ecol.*, 2003, **29**, 2213.

100 T. Nakahata, N. Itagaki, T. Arai, H. Sugie and S. Kuwahara, *Biosci., Biotechnol., Biochem.*, 2003, **67**, 2627.

101 M. Ando, H. Ohhara and K. Takase, *Chem. Lett.*, 1986, **15**, 879.

102 J. G. Millar, S. P. Chinta, J. S. McElfresh, L. J. Robinson and J. G. Morse, *J. Econ. Entomol.*, 2012, **105**, 497.

103 (*a*) A. Zhang, J. Nie and A. Khrimian, *Tetrahedron Lett.*, 2004, **45**, 9401; (*b*) A. Zhang and J. Nie, *J. Agric. Food Chem.*, 2005, **53**, 2451.

104 (*a*) A. Zhang and D. Amalin, *Environ. Entomol.*, 2005, **34**, 264; (*b*) A. Zhang, S. Wang, J. Vitullo, A. Roda, C. Mannion and J. C. Bergh, *Chem. Senses*, 2006, **31**, 621.

105 J. Einhorn, A. Guerrero, P.-H. Ducrot, F.-D. Boyer, M. Gieselmann and W. Roelofs, *Proc. Natl. Acad. Sci. U. S. A.*, 1998, **95**, 9867.

106 F.-D. Boyer and P.-H. Ducrot, *Eur. J. Org. Chem.*, 1999, 1201–1211.

107 (*a*) I. Petschen, A. Parrilla, M. P. Bosch, C. Amela, A. A. Botar, F. Camps and A. Guerrero, *Chem.–Eur. J.*, 1999, **5**, 3299; (*b*) I. Petschen, M. P. Bosch and A. Guerrero, *Tetrahedron: Asymmetry*, 2000, **11**, 1691.

108 (*a*) A. W. Schmidt, J. R. Suresh, G. Theumer and H.-J. Knolker, *Chem. Lett.*, 2007, **36**, 1478; (*b*) A. W. Schmidt and H.-J. Knolker, *Synlett*, 2010, 2207.

109 T. Eisner, M. Deyrup, R. Jacobs and J. Meinwald, *J. Chem. Ecol.*, 1986, **12**, 1407.

110 J. G. Millar, J. A. Moreira, J. S. McElfresh, K. M. Daane and A. S. Freund, *Org. Lett.*, 2009, **11**, 2683.

111 J. M. Conia and M. L. Leriverend, *Bull. Soc. Chim. Fr.*, 1970, 2981.

112 Y. Zou and J. G. Millar, *J. Org. Chem.*, 2009, **74**, 7207.

113 Y. Zou and J. G. Millar, *Synlett*, 2010, 2319.

114 W. F. Bailey and J. M. Bakonyi, *J. Org. Chem.*, 2013, **78**, 3493.

115 K. Takai, T. Kakiuchi, Y. Kataoka and K. Utimoto, *J. Org. Chem.*, 1994, **59**, 2668.

116 S. E. Kurhade, V. Siddaiah, D. Bhuniya and D. S. Reddy, *Synthesis*, 2013, **45**, 1689.

117 A. Srikrishna and G. Satyanarayana, *Tetrahedron*, 2006, **62**, 2892.

118 D. A. Engel and G. B. Dudley, *Org. Biomol. Chem.*, 2009, **7**, 4149.

119 R. Ramesh, P. S. Swaroop, R. G. Gonnade, C. Thirupathi, R. A. Waterworth, J. G. Millar and D. S. Reddy, *J. Org. Chem.*, 2013, **78**, 6281.

120 Y. Zou, K. M. Daane, W. J. Bentley and J. G. Millar, *J. Agric. Food Chem.*, 2010, **58**, 4977.

121 J.-M. Galano, G. Audran, L. Mikolajezyk and H. Monti, *J. Org. Chem.*, 2001, **66**, 323.

122 H. Pamingle, R. L. Snowden and K. H. Schulte-Elte, *Helv. Chim. Acta*, 1991, **74**, 543.

123 J. G. Millar, S. L. Midland, J. S. McElfresh and K. M. Daane, *J. Chem. Ecol.*, 2005, **31**, 2999.

124 J. G. Millar and S. L. Midland, *Tetrahedron Lett.*, 2007, **48**, 6377.

125 (a) N. Krause, *Angew. Chem., Int. Ed.*, 1994, **33**, 1764; (b) N. Krause, S. Ebert and A. Haubrich, *Liebigs Ann./Recl.*, 1997, 2409.

126 Y. Zou and J. G. Millar, *Tetrahedron Lett.*, 2011, **52**, 4224.

127 C. Morel-Fourrier, J.-P. Dulcère and M. Santelli, *J. Am. Chem. Soc.*, 1991, **113**, 8062.

128 (a) W. Barth and L. A. Paquette, *J. Org. Chem.*, 1985, **50**, 2438; (b) F. Kazmierczak and P. Helquist, *J. Org. Chem.*, 1989, **54**, 3988.

129 B. Figadere, F. J. Devlin, J. G. Millar and P. J. Stephens, *Chem. Commun.*, 2008, 1106.

130 K. Hashimoto, A. Morita and S. Kuwahara, *J. Org. Chem.*, 2008, **73**, 6913.

131 N. Asao, S. Lee and Y. Yamamoto, *Tetrahedron Lett.*, 2003, **44**, 4265.

132 A. K. Hajare, L. S. Datrange, S. Vyas, D. Bhuniya and D. S. Reddy, *Tetrahedron Lett.*, 2010, **51**, 5291.

133 G. N. Lanier, Y.-T. Qi, J. R. West, S. C. Park, F. X. Webster and R. M. Silverstein, *J. Chem. Ecol.*, 1989, **15**, 1645.

134 J. Einhorn, P. Menassieu, C. Malosse and P. H. Ducrot, *Tetrahedron Lett.*, 1990, **31**, 6633.

135 E. Dunkelblum, Z. Mendel, F. Assael, M. Harel, L. Kerhoas and J. Einhorn, *Tetrahedron Lett.*, 1993, **34**, 2805.

136 S. C. Park, A. J. Wi and K. Mori, *Kor. J. Appl. Entomol.*, 1994, **33**, 250.

137 (a) X. Shi, F. X. Webster, J. Kallmerten and J. Meinwald, *Tetrahedron Lett.*, 1995, **36**, 7197; (b) X. Shi, F. X. Webster and J. Meinwald, *Tetrahedron Lett.*, 1995, **36**, 7201; (c) X. Shi, F. X. Webster and J. Meinwald, *Tetrahedron*, 1995, **51**, 10433.

138 H. Jactel, P. Menassieu, M. Lettere, K. Mori and J. Einhorn, *J. Chem. Ecol.*, 1994, **20**, 2159.

139 E. Dunkelblum, R. Gries, G. Gries, K. Mori and Z. Mendel, *J. Chem. Ecol.*, 1995, **21**, 849.

140 C. L. Cywin, F. X. Webster and J. Kallmerten, *J. Org. Chem.*, 1991, **56**, 2953.

141 C. L. Cywin and J. Kallmerten, *J. Nat. Prod.*, 1991, **54**, 1664.

142 K. Mori and S. Harashima, *Liebigs Ann. Chem.*, 1993, 391.

143 K. Mori and S. Harashima, *Liebigs Ann. Chem.*, 1993, 993.

144 K. Mori, T. Furuuchi and H. Kiyota, *Liebigs Ann. Chem.*, 1994, 971.

145 J.-L. Brevet and K. Mori, *Synthesis*, 1992, 1007.

146 K. Mori, T. Furuuchi and H. Kiyota, *Liebigs Ann. Chem.*, 1995, 2093.

147 G.-Q. Lin and W.-C. Xu, *Tetrahedron Lett.*, 1993, **34**, 5931.

148 G.-Q. Lin and W. C. Xu, *Bioorg. Med. Chem.*, 1996, **4**, 375.

149 L. Zegelman, A. Hassner, Z. Mendel and E. Dunkelblum, *Tetrahedron Lett.*, 1993, **34**, 5641.

150 K. Mori and M. Amaike, *J. Chem. Soc., Perkin Trans. 1*, 1994, 2727.

151 H. Watanabe, T. Watanabe, T. Kitahara and K. Mori, *Biosci., Biotechnol., Biochem.*, 1997, **61**, 127.

152 S. Kurosawa, M. Takenaka, E. Dunkelblum, Z. Mendel and K. Mori, *ChemBioChem*, 2000, **1**, 56.

153 J. E. Frey and B. Frey, *Mol. Ecol.*, 1995, **4**, 777.

154 R. A. Waterworth, R. A. Redak and J. G. Millar, *J. Econ. Entomol.*, 2011, **104**, 555.

155 A. Roda, J. G. Millar, J. Rascoe, S. Weihman and I. Stocks, *J. Econ. Entomol.*, 2012, **105**, 2052.

156 A. Francis, K. A. Bloem, A. L. Roda, S. L. Lapointe, A. Zhang and O. Onokpise, *Fla. Entomol.*, 2007, **90**, 440.

157 E. E. Grafton-Cardwell, J. G. Millar, N. V. O'Connell and L. M. Hanks, *J. Agric. Urban Entomol.*, 2000, **17**, 75–88.

158 V. M. Walton, K. M. Daane and K. L. Pringle, *Crop Prot.*, 2004, **23**, 1089.

159 R. A. Waterworth, I. M. Wright and J. G. Millar, *Ann. Entomol. Soc. Am.*, 2011, **104**, 249.

160 J. C. Franco, S. Gross, E. B. da Silva, P. Suma, A. Russo and Z. Mendel, *An. Inst. Super. Agron., Univ. Tec. Lisboa*, 2003, **49**, 353.

161 A. Hefetz, S. Kronenberg, B. A. Peleg and I. Bar-Zakay, in *Citriculture: Proc. 6th Int. Citrus Cong.*, ed. R. Goren, N. Goren and K. Mendel, Middle-East, Tel Aviv, Israel, March 6-11, 1988.

162 (a) S. Vacas, C. Alfaro, V. Navarro-Llopis and J. Primo, *Bull. Entomol. Res.*, 2009, **99**, 415–423; (b) S. Vacas, C. Alfaro, V. Navarro-Llopis and J. Primo, *Pest Manage. Sci.*, 2010, **66**, 745; (c) S. Vacas, C. Alfaro, J. Primo and V. Navarro-Llopis, *J. Pest Sci.*, 2014, DOI: 10.1007/s10340-014-0623-1; (d) S. Vacas, P. Vanaclocha, C. Alfaro, J. Primo, M. J. Verdú, A. Urbaneja and V. Navarro-Llopis, *Pest Manage. Sci.*, 2012, **68**, 142.

163 R. E. Rice, C. A. Atterholt, M. J. Delwiche and R. A. Jones, *IOBC-WPRS Bulletin*, 1997, **20**, 151.

164 K. Daane, W. Bentley, V. Walton, R. Malakar-Kuenen, J. Millar, C. Ingels, E. Weber and C. Gispert, *Calif. Agric.*, 2006, **60**, 31.

165 V. M. Walton, K. M. Daane, W. J. Bentley, J. G. Millar, T. E. Larsen and R. Malakar-Kuenen, *J. Econ. Entomol.*, 2006, **99**, 1280.

166 A. Cocco, M. Coinu, A. Lentini, G. Serra and G. Delrio, *IOBC-WPRS Bulletin*, 2011, **67**, 215.

167 K. M. Daane, W. J. Bentley, R. J. Smith, D. R. Haviland, E. A. Wever, M. Battany, C. Gispert and J. G. Millar, Planococcus mealybugs, in *Grape Pest Management*, ed. L. J. Bettiga, Publication 3343, University of California Agriculture and Natural Resources Publication 3343, Oakland CA, 3rd edn, 2013, pp. 246–260.

168 D. J. Langone, Efficacy of pheromone mating disruption for vine mealybug control, MSc. Thesis, Fresno State University, Fresno CA, 2014, p. 75, http://hdl.handle.net/10211.3/105368.

This journal is © The Royal Society of Chemistry 2015

Nat. Prod. Rep., 2015, **32**, 1067–1113 | 1113