NPR

REVIEW



View Article Online

Cros ←click Cite this: *Nat.*

CrossMark ← click for updates

Cite this: Nat. Prod. Rep., 2015, 32, 273

Triterpenoids

Robert A. Hill* and Joseph D. Connolly

Covering: 2012. Previous review: Nat. Prod. Rep., 2013, 29, 1028-1065

Received 30th July 2014 DOI: 10.1039/c4np00101j

www.rsc.org/npr

This review covers the isolation and structure determination of triterpenoids reported during 2012 including squalene derivatives, lanostanes, holostanes, cycloartanes, cucurbitanes, dammaranes, euphanes, tirucallanes, tetranortriterpenoids, quassinoids, lupanes, oleananes, friedelanes, ursanes, hopanes, serratanes, isomalabaricanes and saponins; 348 references are cited.

- 1. Introduction
- 2. The squalene group
- 3. The lanostane group
- 4. The dammarane group
- 4.1 Tetranortriterpenoids
- 4.2 Quassinoids
- 5. The lupane group
- 6. The oleanane group
- 7. The ursane group
- 8. The hopane group
- 9. Miscellaneous compounds
- 10. References

1. Introduction

There is continued interest in the anticancer activities of triterpenoids¹⁻³ and their potential for treatment or prevention of diabetes and Alzheimer's disease.⁴ The oral absorption and metabolism of triterpenoid saponins has been reviewed.⁵ Surveys of triterpenoids from *Ceriops*,⁶ and *Ilex*⁷ species and *Sapindus mukorossi*,⁸ have appeared.

2. The squalene group

Sapelenins G 1–J 4 are further anti-inflammatory squalene derivatives from the bark of Cameroonian *Entandrophragma cylindricum*.⁹ The highly oxidised squalene derivative 5 has been isolated from Peruvian *Protium subserratum*.¹⁰ The structures of saiyacenols A 6 and B 7, from the red alga *Laurencia viridis*, support the accepted pathway for the formation of the aplysiols.¹¹ The mechanism of triterpene biosynthesis in *Botryococcus braunii* has been reviewed.¹²

3. The lanostane group

New protostanes include alisol Q acetate **8** (ref. 13) and alisol X **9** (ref. 14) from *Alisma orientale* and the epoxy-ketone **10** from the leaves of *Aglaia odorata*.¹⁵ The tetraterpenes abiestetranes A **11** and B **12**, from *Abies fabri*,¹⁶ and abibalsamins A **13** and B **14**, from the oleoresin of *Abies balsamea*,¹⁷ appear to have arisen by Diels–alder cycloadditions of rearranged lanostanes with the monoterpene β -myrcene. The structure of **13** was confirmed by X-ray analysis. A series of rearranged lanostanes, neoabiestrines A **15**–F **20**, has been reported from *Abies recurvata*.¹⁸ The structures of neoabiestrine A **15** were confirmed by X-ray analysis. The compounds showed some cytotoxic activity. The mariesane lactone **21** has been obtained from *Abies sibirica*.¹⁹ The tetranor derivative **22** and the 3,4-secolanostane **23** have been found in *Abies holophylla*.²⁰

Pseudoferic acids A 24, B 25 and C 26 are interesting new 16,24-cyclised lanostanes from Pseudolarix kaempferi.²¹ The stems of Schisandra glaucescens contain the ring A-cleaved lanostanes schiglausins A 27-H 34, together with the intact derivatives schiglausins I 35 and J 36.²² The structure of schiglausin A 27 was confirmed by X-ray analysis. Schiglausin H 34 is the methyl ester of micranoic acid A. An impressive array of rearranged, ring A-cleaved and intact lanostanes, kadpolysperins A 37-N 50, has been isolated from Kadsura polysperma.23 Kadcoccitones A 51 and B 52 are unusual rearranged lanostane derivatives from Kadsura coccinea where they occur with kadcoccitone C 53, whose structure was confirmed by X-ray analysis.24 Secococcinic acids G 54-K 58 are further constituents of Kadsura coccinea.²⁵ Three esters 59-61 of 3-epidehydrotumulosic acid have been obtained from Wolfiporia extensa.26 Other new lanostanes include lanosta-5,15-dien-3a-ol 62 from Arctium lappa²⁷ and inonotsuoxodiols B 63 and C 64, epoxyinonotsudiol 65 and methoxyinonotsutriol 66 from Inonotus obliguus.²⁸

School of Chemistry, Glasgow University, Glasgow, G12 8QQ, UK. E-mail: bob.hill@ glasgow.ac.uk



Sarasinosides N–R are 30-norlanostane glycosides from the sponge *Lypastrotethya* sp. with the new genins **67**, **68**, **69** and the 30-*nor*-18(13 \rightarrow 14)-*abeo*-derivative **70**.²⁹ Scillanostaside F, with the new genin **71**, and scillanostaside G, with a known tetranorlanostane genin, have been isolated from the bulbs of *Scilla scilloides*.³⁰



New compounds from *Ganoderma sinense* include the pentanor derivative ganosineniol A 72, ganoderic acids J_c 73 and J_d 74, ganodermatetraol 75, the glucosyl ester ganosinoside A 76 with a known genin, ganolucidic acid γ_a 77, ganolucidate F 78, ganoderiol J 79 and methyl lucidenate H_a 80.³¹ Ganodermacetal 81 is an acetonide from *Ganoderma amboinense*³² and lucialdehyde E 82 is a further compound from *Ganoderma lucidum*.³³ Reviews have appeared on the triterpenoids of *Ganoderma lucidum*^{34,35} and lanostanes from fungi.³⁶

Astraodoric acids A **83**–D **86** are metabolites of the mushroom *Astraeus odoratus*.²⁹ The structure of astraodoric acid B **84** was confirmed by X-ray analysis. The structure of astrakurkurol **87**, from the Indian edible mushroom *Astaeus hygrometricus*, was also confirmed by X-ray analysis.³⁷ The corresponding lactone, astrakurkurone **88**, was also obtained. Two highly acetylated lanostanes, coprinacins A **89** and B **90**, have been

















This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

(cc)) BY

Open Access Article. Published on 24 October 2014. Downloaded on 7/19/2025 1:07:28 AM.











reported from *Coprinus cinereus*.³⁸ Other fungal metabolites include **91–94** from *Antrodia camphorate*³⁹ and formitoside K **95**, a glucoside with a new genin, from *Fomitopsis nigra*.⁴⁰

Cucumarioside A8 is a lanostane saponin with the new genin 96 from the sea cucumber *Eupentacta fraudatrix*.⁴¹ The sea cucumber *Apostichopus japonicus* is the source of several saponins, 26-*nor*-25-oxoholotoxin A₁ with the new genin 97 and holototoxins D–G.⁴² Holototoxins F and G have the new genin **98** while the genins of D and E are known. Cucumariosides B_1 and B_2^{43} and cucumariosides H_2 , H_3 and H_4 ,⁴⁴ from *Eupentacta fraudatrix*, all have known genins. Holostane saponins and their biological activity have been reviewed.⁴⁵

Interesting new structures continue to appear from *Schisandra* species. Schilancitrilactones A **99**, B **100** and C **101**







have been reported from *Schisandra lancifolia*.⁴⁶ The structures of all the schilancitrilactones were confirmed by X-ray analyses. New structures from *Schisandra sphenanthera* include preschisanartanins E **102–J 107** and sphenadilactones D **108–**F **110**.⁴⁷ Isoschicagenin C **111**, preschisanartanins K **112–M 114**,

schisdilactones A 115–G 121 and schinesdilactones A 122 and B 123 constitute an impressive array of new derivatives from *Schisandra chinensis*.⁴⁸ Schiglausins K 124–O 128 are simple ring A-cleaved cycloartanes from *Schisandra glaucescens*.⁴⁹







112 R¹ = R⁴ = R⁴ = H; R² = OH **113** R¹ = R² = H; R³ = OH; R⁴ = Me **114** R¹ = OH; R² = R³ = H; R⁴ = Me; 22*E*



115 $R^1 = H$; $R^2 = Me$ **116** $R^1 = H$; $R^2 = Me$; 20-epimer **117** $R^1 = OH$; $R^2 = Me$ **118** $R^1 = OH$; $R^2 = Me$; 20-epimer **119** $R^1 = R^2 = H$





An interesting approach to the "dereplication, residual complexity and rational naming" of the *Actea* triterpenoids has the potential to be applied to other groups of natural products with the same inherent problems.⁵⁰ Pseudolaridimers A **129** and

B **130**, from *Pseudolarix amabilis*, appear to have arisen by a Diels–Alder reaction involving a cycloartane and a labdane.⁵¹ The structure of **129** was confirmed by an X-ray analysis of the corresponding methyl ester. The cycloartanes lygodipenoids A

NPR





131 and B 132, from Lygodium japonicum, have an additional cyclopropane in their side chains.52 Thirty new cycloartane proteasome inhibitors 133-162 have been reported from Neoboutonia melleri.53 An interesting UV light-induced inversion of the configurations at C-9 and C-10 was observed in this series. Caloncobic acids A 163 and B 164 and caloncobalactones A 165 and B 166 are constituents of the leaves of Caloncoba glauca.54 Compounds 167-172 are new cycloartanes from the leaves of Homonoia riparia.55 Neoabiestrines G 173-I 175 have been

reported from Abies recurvata.18 The structure of neoabiestrine H 174 was confirmed by X-ray analysis. Other new cycloartanes include the trinor derivatives 176 and 177 from Abies holophylla,²⁰ rotundusolide C 178 from the rhizomes of Cyperus rotundus,⁵⁶ 179-182 from the aerial parts of Atemisia lagocephala,⁵⁷ three esters 183-185 of cyclomargenol from Krameria pauciflora,58 euphonerins A 186-G 192 from the leaves of Euphorbia neriifolia,⁵⁹ cycloccidentalic acids A 193 and B 194 and the related saponins cycloccidentalisides I 195-V 199 from





151 R = Ac 152 R = H 153 R = Ac; 9,10-diepimer

154 R = H **155** R = OH

156 $R^1 = R^4 = H$; $R^2 = OAc$; $R^3 = OH$ **157** $R^1 = OH$; $R^2 = OAc$; $R^3 = H$; $R^4 = H$ **158** $R^1 = R^2 = R^4 = H$; $R^3 = H$ **159** $R^1 = R^2 = R^3 = H$; $R^4 = Rha$







163 $R^1 = O$; $R^2 = \alpha OH$; $R^3 = CH_2$ **164** $R^1 = \beta OH$, H; $R^2 = \beta OH$; $R^3 = CH_3$, OH



167 $\mathbb{R}^1 = \mathbb{H}; \mathbb{R}^2 = \mathbb{R}^3 = \beta OH, \mathbb{H}; \mathbb{R}^4 = \mathbb{H}_2$ **168** $\mathbb{R}^1 = \mathbb{H}; \mathbb{R}^2 = \mathbb{R}^3 = \mathbb{R}^4 = \beta OH, \mathbb{H}$ **169** $\mathbb{R}^1 = \mathbb{H}; \mathbb{R}^2 = \mathbb{R}^4 = \beta OH, \mathbb{H}; \mathbb{R}^3 = \mathbb{H}_2$ **170** $\mathbb{R}^1 = Xyl; \mathbb{R}^2 = \mathbb{H}_2; \mathbb{R}^3 = \mathbb{R}^4 = \beta OH, \mathbb{H}$ **171** $\mathbb{R}^1 = Xyl; \mathbb{R}^2 = \mathbb{R}^4 = \mathbb{H}, \beta OH; \mathbb{R}^3 = \mathbb{H}_2$ **172** $\mathbb{R}^1 = Xyl; \mathbb{R}^2 = \mathbb{R}^3 = \beta OH, \mathbb{H}; \mathbb{R}^4 = \mathbb{H}_2$

R¹0











186 $R^1 = \beta OH, H; R^2 = H; R^3 = CH_3, OH$ **187** $R^1 = \beta OH, H; R^2 = OH; R^3 = CH_3, H$ **188** $R^1 = \beta OH, H; R^2 = \alpha OH; R^3 = CH_2; 25,26-dihydro$ **190** $R^1 = O; R^2 = \beta OH; R^3 = \beta OH, H$



Cassia occidentalis,⁶⁰ glaucartanoic acids A **200** and B **201** from the fruit of *Caloncoba glauca*⁶¹ and **202** from the leaves of *Aglaia exima*.⁶² Compounds **203**, **204** and the acetonide **205**, a likely artefact, have been obtained from the resin of *Commiphora opobalsamum*.⁶³ The structures of **203** and **205** were confirmed by X-ray analyses.

New cycloartanes continue to be isolated from *Cimicifuga* species. *Cimicifuga foetida* is the source of compounds **206–209**

(ref. 70) and of 24-*epi*-cimigenol-3-one **210** and the xyloside foetinoside **211**.⁶⁴ The ring A-cleaved derivative **212**, the cimigenol arabinosides **213–215**, the 25-dehydrocimigenol arabinoside **216**, compounds **217–219** and the shengmanol arabinoside **220** have all been isolated from the roots of *Cimicifuga heracleifolia*.⁶⁵ Other new compounds include the galactopyranosides **221–223**, all with new genins, from the roots

NPR



R O O H





209

206 R = O **210** R = H₂; 7,8 β -dihydro; 24-epimer

207 R = O **208** R = β OH, H; 7,8-didehydro; 1,2-dihydro



215 $R^1 = R^2 = H$; $R^3 = Ac$ **216** $R^1 = R^3 = H$; $R^2 = Ac$; 25,26-anhydro

211



of *Cimicifuga simplex*⁶⁶ and isocimipodocarpaside **224** from *Cimicifuga racemosa*.⁶⁷

Six cycloartane saponins with the new genins 225-228 have been obtained from Astragalus angustifolius.68 Cycloquivinoside A 229 is a new saponin from Astragalus chivensis.⁶⁹ The new genin cycloartane-36,6a,166,20S,24R,25-hexol 230 has been identified in the saponins of Astragalus stereocalyx70 and in saponins from Astragalus schottianus.⁷¹ Nervisides A 231-C 233, from Nervilia fordii, all have new genins.72 Other cycloartane saponins with new genins include curculigosaponins N and O from Curculigo orchioides with the genin 234 (ref. 73) and two saponins from Thalictrum fortune with the genins 235 and 236.74 The ring-A cleaved cycloartanes sootependial 237 and sootepenoic acid 238 have been isolated from the exudates of Gardenia sootepensis.75 Novel cycloartane saponins with known genins include cycloascidoside from Astragalus mucidus,76 cyclogaleginoside C from Astragalus galegiformis and cycloascauloside D from Astragalus caucasicus,77 hareftosides A-D from

Astragalus hareftae,⁷⁸ neoastragaloside I from Astragalus membranaceus⁷⁹ and unnamed saponins from Astragalus erinaceus.⁸⁰

A new nitrogen-containing cucurbitane, endecaphyllacin C 239, has been reported from the tubers of Hemsleya endecaphylla.81 Eleaocarpucins A 240-H 247 are 16,23-epoxy derivatives from *Eleaocarpus chinensis*.⁸² The new cucurbitanes jinfushanencins A 248 and B 249 occur in the tubers of Hemsleya penxianensis, together with the glycosides jinfushanosides E-K.83 Jinfushanoside K has the new genin 250. Other new cucurbitanes include 251-253 from the fruit of Momordica charantia,⁸⁴ 10β-hydroxybryodulcosigenin 254 from Saniculiphyllum guangxiense,⁸⁵ six new compounds 255-260 from the leaves of Momordica charantia86 and isoarvenin III 261 from the fruit of Trichosanthes kirilowii.87 The 3,4-seco-cucurbitane 262 is a constituent of Russula lepida and Russula amarissima.88 The unlikely 10a-methyl lanostane structure 263 has been proposed for a compound from Momordica charantia.89 Perhaps the compound is a cucurbitane! Reviews have appeared on







cucurbitacins and bottle gourd toxicity,⁹⁰ medicinally important plants of the Cucurbitaceae⁹¹ and the anticancer activity of the cucurbitacins.^{92,93}

4. The dammarane group

A review of ginsenoside derivatives and their antitumour activity has been published.⁹⁴ New saponins from the stems and leaves of *Panax ginseng* include ginsenosides Rh₁₄-Rh₁₇ with the new

genins **264** (Rh₁₅) and **265** (Rh₁₇)⁹⁵ and ginsenosides Rh₁₈– Rh₂₀.⁹⁶ The new genin **266** of ginsenoside Rh₁₈ was also isolated together with the dammarane **267**.⁹⁶ Two new saponins of 20*S*-protopanaxatriol have also been found in *Panax ginseng* together with the dammarane **268**.⁹⁷ The biological activity of an artefact **269** from the acid hydrolysate of *Panax ginseng* has been investigated.⁹⁸ The hexanordammarane saponin, ginsenoside R₁₀ **270**, has been isolated from the stems and leaves of *Panax quinquefolium*.⁹⁹





Saponins with the new genins 271 and 272 (ref. 100) and 273 (ref. 101) have been isolated from *Gynostemma pentaphyllum*. Cyclocariosides D–G, with the new genins 274 and 275, cyclocarioside H, with a known genin, and the dammarane cyclocarin A 276 are constituents of the leaves of *Cyclocarya paliurus*.¹⁰² Dammarane saponins with known genins have been

isolated from the roots of *Machilus yaoshansis*.¹⁰³ This paper also reports a revision the C-23 configuration of *Gynostemma pentaphylla* saponins as in 277.

New dammaranes include the rearranged aglinone 278 and aglinin E 279 from the bark of *Aglaia smithii*,¹⁰⁴ the *nor*-derivatives 280–283 from *Dysoxylum hainanense*,¹⁰⁵ the



0

н

OH

270



R²

HO

ОН

٠H

`OEt

GlcO

ΟН

269

Å



268 R¹ = R² = OH; R³ = H **271** R¹ = R² = H; R³ = OH



`OMe









27-*nor*-derivative **284** from *Dipterocarpus obtusifolius*¹⁰⁶ and the two probable artefacts **285** and **286** from the leaves of *Aglaia odorata*.¹⁵

Novel dammarane saponins with known genins include acerbosides A and B from *Hovenia acerba*,¹⁰⁷ chikusetsusaponins FK_1 - FK_7 , FH_1 , FH_2 , FM and FT_1 - FT_4 from *Panax japonicus*,^{108,109} gypenbiosides A and B from *Gynostemma*









pentaphyllum,¹¹⁰ 20*R*-pseudoginsenoside F_{11} (ref. 111) and quinquenosides Ja and Jb¹¹² from *Panax quinquefolium*.

New compounds from the stem bark of *Aphanamixis* grandifolia include the tautomeric tirucallane

cyclopentenones 287 and 288 (ref. 113) and the tirucallanes 289 and 290.114 Compounds 287 and 288 were originally named aphragranins A and B, names used by the authors for previous compounds. Their names have been changed to aphagraones A and B.¹¹⁵ Two further derivatives 291 and 292 were obtained from the leaves and twigs of Aphanamixis grandifolia.¹¹⁶ Some of these compounds clearly incorporate the extraction solvent. Aphanamgrandins A 293-J 302 constitute a series of 2,3-seco- and 3,4-seco-tirucallanes from the stems of Aphanamixis grandifolia.117 They were accompanied by the ring-A intact derivatives aphanamgrandin J 303 and the dienone 304. The structures of aphanamgramins A 293 and B 294 were confirmed by X-ray analyses. Other new tirucallanes include the 21-nor-derivative dysoxylentin A 305 from Dysoxylum lenticellatum,¹¹⁸ ixoroid 306 from the flowers of Ixora coccinea,119 307 and 308 from the stem bark of Araliopsis synopsis,¹²⁰ capulin 309 from Capuronianthus mahafalensis,¹²¹ 310-312 from the stem bark of Melia toosendan,¹²² dysohainanin F 313 from Dysoxylum hainanense123 and











Review



24,25,26-trihydroxytirucall-7-en-3-one **314** from *Salacia* hainanensis.¹²⁴ The 19(10 \rightarrow 9)-abeotirucallane **315** has been reported from *Euphorbia mellifera*.¹²⁵

Chisiamols G **316** and H **317** (ref. 126) and chisopanins L **318**–O **321** (ref. 108 and 127) are new apotirucallanes from *Chisocheton paniculatus*. A series of compounds from the leaves and twigs of *Melia toosendan* includes the apoeuphane mesendanin K **322**, the euphanes mesendanins L **323**–P **327**, the tirucallanes mesendanins Q **328**–T **331** and the apotirucallane mesendanin U **332**.¹²⁸ The apotirucallane **332** was also isolated from *Dysoxylum hainanense* and named dysohainanin E.¹²³ Other new apotirucallanes include piscidinones A **333** and B **334** from *Walsura trifoliata*,¹²⁹ agladorals A **335–E 339** from *Aglaia odorata* var. *microphyllina*¹³⁰ and trichostemonate **340** from the roots of *Walsura trichostemon*.¹³¹ The structure of **340** was confirmed by X-ray analysis. Toonaciliatavarins A **341–H 348** are constituents of *Toona ciliata*.¹³²

4.1 Tetranortriterpenoids

A review of the structures and biological activities of limonoids from *Cipadessa* species has been published.¹³³ Walsucochinoids

A 349 and B 350 are rearranged limonoids, with an aromatic ring D, from Walsura cochinchinensis.134 Andhraxylocarpins A 351, B 352, C 353 and E 354 are new skeletal types from *Xylocarpus granatum*.¹³⁵ The structures of **349**, **351** and **353** were confirmed by X-ray analyses. Andhraxylocarpin D 355 is the same as chisomicin A, published in 2011. Hortia oreadica is the source of the interesting rearranged limonoids 356-363, related to the known hortiolide A.¹³⁶ The structure of hortiolide C 358 was confirmed by X-ray analysis. The configurations at C-5 and C-10 of hortiolide E 362 and 12-hydroxyhortiolide E 363 are wrongly drawn in the paper. Carapanolides A 364 and B 365 are 9,10-seco mexicanolide derivatives from the seeds of Carapa guianensis.137 Other new structural types include citriolide A 366 from the seeds of Citrus reticulate, 138 aphanamixoid A 367 from Aphanamixis polystachya139 and chukrasones A 368 and B 369 from Chukrasia tabularis.140 The structure of aphanamixoid A 367 was confirmed by X-ray analysis. It was accompanied in the extract by aphanamixoid B 370.

Toonayunnanins A **371–**L **382** constitute a group of assorted limonoids from the leaves of *Toona ciliata* var. *yunnanensis*.¹⁴¹ Toonayunnanin I **378** is the same as toonaciliatin P, published in 2011. Another group of compounds, toonaciliatones B **383–**F





387, has been obtained from the seeds of *Toona ciliata*.¹⁴² Meliatoosenins E **388**, F **389**, I **390**, J **391**, L **392**–N **394** and P **395**–S **398** are new compounds from the fruit of *Melia toosendan*.¹⁴³ Meliatoosenins G, H, K and O, claimed as new compounds, have all been published previously. Other new compounds include andirolides H **399**–P **407** from *Carapa guianensis* flowers,¹⁴⁴ ceramicines J **408**–L **410** from *Chisocheton ceramicus*¹⁴⁵ and walsuranins A **411**–C **413** from *Walsura yunnanensis*.¹⁴⁶ The ring A-cleaved compounds, aphanalides A **414–**H **421**, have been reported from the fruit of *Aphanamixis polystachya*.¹⁴⁷ The structures of aphanalides A **414** and C **416** were confirmed by X-ray analyses. Aphanagranins A **422–**D **425** are constituents of *Aphanamixis grandifolia*.¹⁴⁸ Other species producing multiple new compounds include *Munronia unifoliata* with munronoids A **426–**J **435** (ref. 149) and K **436–**O **440** (ref. 150) and *Dysoxylum hainanense* with dysohainanins A **441–**D **444**.¹²³ 6-O-Deacetylseverinolide **445** is a constituent of *Atalantia buxifolia*.¹⁵¹













 $\begin{array}{l} \textbf{377} \ \textbf{R} = \textbf{CH}_3 \\ \textbf{378} \ \textbf{R} = \textbf{CH}_2 \textbf{OAc} \\ \textbf{379} \ \textbf{R} = \textbf{CH}_3; \ \textbf{1,2-didehydro} \\ \textbf{380} \ \textbf{R} = \textbf{CHO}; \ \textbf{1,2-didehydro} \end{array}$





383 $R^1 = O$; $R^2 = R^3 = H$ **385** $R^1 = \alpha OAc$, H; $R^2 = OH$; $R^3 = H$ **386** $R^1 = \alpha OAc$, H; $R^2 = H$; $R^3 = OH$



O OAc OAc 387





392 R^1 = Tig; R^2 = H; R^3 = α OMe **393** R^1 = H; R^2 = Ac; R^3 = α OMe **394** R^1 = R^2 = Ac; R^3 = β OMe



411

NPR

296 | Nat. Prod. Rep., 2015, 32, 273-327

408

409 R¹ = OAc; R² = H

410 R¹ = CH₃; R² = OH

413

412



414 R¹ = HCO; R² = Me₂CHCH₂CO **415** R¹ = H; R² = Me₂CHCH₂CO **416** R¹ = H; R² = Et(Me)CHCH(OH)CO **422** R¹ = R² = H



417



418 $R^1 = H$; $R^2 = Et(Me)CHCH(OH)CO$; $R^3 = H$ **419** $R^1 = H$; $R^2 = Et(Me)CHCH(OH)CO$; $R^3 = Ac$ **420** $R^1 = HCO$; $R^2 = Et(Me)CHCH(OH)CO$; $R^3 = H$ **421** $R^1 = HCO$; $R^2 = Me_2CHCH_2CO$; $R^3 = H$

O





438 $R^1 = R^3 = Ac; R^2 = \alpha OTig, H$





The seemingly unending flow of phragmalin/bussein derivatives and variants continues and is matched only by the proliferation of confusing trivial names. These points are well illustrated by *Chukrasia tabularis* and *Chukrasia tabularis* var. *velutina*, the sources of chubularisins A **446**–R **463**,¹⁵² chuktabrins C **464–J 471** and chuktabularins U **472–X 475**,¹⁵³ tabulalins G **476–I 478**,¹⁵⁴ chukvelutins D **479–F 481** (ref. 155) and R310B8 **482** and velutinalides A **483** and B **484**.¹⁵⁶

Chabularisin O **460** is the same as chuktabularin V **474**. The structure of chuktabrin C **464** was confirmed by X-ray analysis. Heytrijumalins A **485–I 493** have been isolated from the twigs and leaves of *Heynea trijuga*.¹⁵⁷ Other new compounds in this group include hisomicines D **494** and E **495** from *Chisocheton ceramicus*,¹⁵⁸ malayanines A **496** and B **497** from *Chisocheton ceramicus*,¹⁵⁹ senegalensions A **498–**C **500** from *Khaya senegalensis*,¹⁶⁰ kotschyins D **501–**H **505** from *Pseudocedrela kotschyi*,¹⁶¹ soymidins A **506** and B **507** from *Soymida febrifuga*¹⁶² and swietenin J **508** from *Swietenia macrophylla*.¹⁶³ Five new mexicanolide derivatives, heytrijunolides A **509–**E **513**, have been obtained from *Heynea trijuga*.¹⁶⁴ Mollucensins R **514–**Y **521** are new mexicanolide and phragmalin derivatives from the seeds of *Xylocarpus moluccensis*.¹⁶⁵

4.2 Quassinoids

The compounds isolated from *Brucea javanica* and their pharmacology have been reviewed.¹⁶⁶ Bruceanic acids E **522** and F **523**, buruceanic acid E methyl ester **524**, javanic acids A **525** and B **526** and javanicolide H **527** are new compounds from the





448 R¹ = Et; R² = R³ = R⁴ = H; R⁵ = Ac; R⁶ = ⁱPrCO **449** R¹ = Me; R² = EtCO; R³ = Ac; R⁴ = R⁵ = H; R⁶ = ⁱPrCO **450** R¹ = Me; R² = ⁱPrCO; R³ = Ac; R⁴ = R⁵ = H; R⁶ = ⁱPrCO **451** R¹ = ⁱPr; R² = R⁵ = R⁶ = H; R³ = Ac; R⁴ = OAc **452** R¹ = ⁱPr; R² = H; R³ = Ac; R⁴ = OAc; ; R⁵ = R⁶ = Ac



453 R' = OAc; R² = 'PrCC **454** R¹ = H; R² = EtCO





458 $R^1 = Ac; R^2 = CH_3$ **459** $R^1 = Ac; R^2 = H$



460 R¹ = R² = Ac; R³ = ⁱPrCO; R⁴ = R⁵ = H **461** R¹ = H; R² = Ac; R³ = EtCO; R⁴ = Me; R⁵ = H **462** R¹ = Ac; R² = H; R³ = ⁱPrCO; R⁴ = Me; R⁵ = H **463** R¹ = H; R² = Ac; R³ = Ac; R⁴ = Me; R⁵ = Me





seeds of *Brucea javanica*.¹⁶⁷ Other new quassinoids include picrasin K **528** from *Quassin amara*¹⁶⁸ and odyendanol **529** from the fruit of *Odyendyea gabonensis*.¹⁶⁹

5. The lupane group

Reviews covering the sources and biological properties of betulinic acid¹⁷⁰ and the antineoplastic effects of lupeol¹⁷¹ have







appeared. The 3,4-secolupane derivatives acanthosessiligenins I **530** and II **531** and acanthosessiliosides A **532**–F **537** have been isolated from the fruit of *Acanthopanax sessiliflorus*.¹⁷² The 17,18-secolupane **538** has been found in the roots of *Taraxacum platycarpum* together with 3β-acetoxylup-18-en-21-one **539**, the neolupane derivatives **540–542** and the migrated lupane **543**.¹⁷³ Lup-20(29)-ene-2β,3β-diol **544** is a constituent of *Salacia hainanensis*¹²⁴ and the related ketones **545** and **546** have be found in *Fagus hayatae*.¹⁷⁴ Other simple lupane derivatives include glochitriol **547** from *Glochidion lanceolarium*,¹⁷⁵ bengalensinone

548 from *Ficus bengalensis*,¹⁷⁶ 3β-hydroxylupane-28,29-dioic acid **549** from *Aglaia duperreana*¹⁷⁷ and the norlupane derivatives **550**, **551** and **552** from *Dipterocarpus obtusifolius*.¹⁰⁶

Two lupane saponins with the new genin **553** have been isolated from *Schefflera venulosa*.¹⁷⁸ New lupane saponins with known genins include ilekudinchoside E from *Ilex kudincha*,¹⁷⁹ stellatosides C, D and E, the methyl esters of stellatosides B and C and thurberoside A from *Stenocereus eruca*¹⁸⁰ and unnamed saponins from *Liquidambar formosana*¹⁸¹ and *Cichorium intybus*.¹⁸²





503 R^1 = OAc, H; R^2 = Me; R^3 = O





Oleanderocioic acid **554**, a lupane ester with the unusual *cis*-4-acetylcinnamic acid, is claimed to be a constituent of *Nerium oleander*.¹⁸³ Other new lupane esters include the nonanoyl ester of lupeol **555** from *Dorstenia harmsiana*,¹⁸⁴ kurramanoic acid **556** from *Nepeta clarkei*,¹⁸⁵ the myristoyl esters **557** from *Glochidion wrightii*¹⁸⁶ and **558** from *Sinocalamus affinis*,¹⁸⁷ the *cis*-feruloyl ester **559** from *Panax ginseng*,¹⁸⁸ the palmitoyl ester **560** from *Cichorium intybus*¹⁸² and the stearoyl ester **561** from *Ocimum sanctum*.¹⁸⁹

6. The oleanane group

Reviews have appeared on the origins, biosynthesis and biological activity of oleanolic acid¹⁹⁰ and the beneficial effects of arjunolic acid in type 1 diabetes.¹⁹¹ *Fagus hayatae* is the source of the 1,10-secooleanane derivative **562** where it is found with 3α ,23-dihydroxy-3-oxoolean-12-en-28-oic acid **563** and 3β ,12 α ,23-trihydroxyoleanan-28,13 β -olide **564**.¹⁷⁴ Sentulic acid from *Sandoricum koetjape* has been identified as 3,4-secooleana-4(23),12-diene-3,27-dioic acid **565**.¹⁹² The 17,22-secooleanenol



566, from *Pyrenacantha kaurabassana*, has the unusual 18α*H*configuration.¹⁹³ Zizimauritic acids A **567**, B **568** and C **569** are ring-A contracted 20,21-secooleanane derivatives from *Ziziphus mauritiana*.¹⁹⁴ The zizimauritic acids also have the 18α*H*configuration. A ring-E contracted oleanane derivative **570** has been isolated from the bark of *Diospyros decandra*.¹⁹⁵ Platycodonoids A **571** and B **572** are 28-noroleanane derivatives from the roots of *Platycodon grandiflorum*.¹⁹⁶ The 30-*nor* derivatives euscaphic acids G **573** and H **574**, together with their likely precursor euscaphic acid I **575**, are present in *Euscaphis japonica*.¹⁹⁷ Further noroleanane derivatives include 30-noroleanolic acid **576** from *Olea europea*,¹⁹⁸ the 24,30-dinor derivatives **577** and **578** and the 24-*nor* compounds **579** and **580** from the roots of *Paeonia emodi*¹⁹⁹ and **581** from *Pilea cavaleriei*.²⁰⁰ A 4900 year old oak wood sample found in a freshwater sediment has produced six noroleanane derivatives **582–587** all lacking oxygenation at C-3.²⁰¹

Several olean-18-ene derivatives have been identified including olean-18-ene- 1β , 2α , 3β -triol **588** from *Salvia atropatana*,²⁰² **589–596** from *Cassine xylocarpa*, **597** from *Maytenus jelskii*,²⁰³ olean-18-ene- 1α , 3β -diol **598** from *Juglans sinensis*²⁰⁴ and 2α , 3α ,24-trihydroxyolean-18-en-28-oic acid **599** from

Review



530 R¹ = Me; R² = OH; R³ = H **531** R¹ = R² = H; R³ = Glc



535 R¹ = Glc; R² = R³ = R⁴ = H **536** R¹ = R² = H; R³ = OH; R⁴ = Glc

Eucalyptus exserta.²⁰⁵ The stem bark of *Terminalia arjuna* is the source of oleaterminaloic acids A **600**, B **601** and C **602** together with the 3-glucoside oleaterminalide **603**.²⁰⁶ Other new simple oleanane derivatives include pseuderanic acid **604** from *Pseuderanthemum carruthersii*,²⁰⁷ punicaone **605** from *Punica granatum*,²⁰⁸ turformosinic acid **606** from *Turpinia formosana*,²⁰⁹ 3β,23,28-trihydroxyolean-12-en-11-one **607** from *Aster yomena*,²¹⁰ 3α,29-dihydroxyolean-12-ene-23,28-dioic acid **608** from *Schefflera farinosa*,²¹¹ oleana-9(11),12-diene-1β,3β-diol **609** from *Salvia xanthocheila*,²¹² 1-*epi*-castanopsol **610** from *Simira glaziovii*,²¹³ the 29,22-olide **611** from *Celastrus orbiculatus*²¹⁴ and the epoxy aldehyde **612** from *Tetraena mongolica*.²¹⁵

New oleanane esters include the caffeoyl derivatives **613–615**, from *Tetraena mongolica*,²¹⁵ the coumaroyl ester **616** from *Rubia schumanniana*,²¹⁶ the ferruloyl ester **617** from *Saniculiphyllum guangxiense*,⁸⁵ the palmitates **618** from *Anemone rivularis*,²¹⁷ **619** from *Barringtonia asiatica*²¹⁸ and **620** from *Lobelia sessilifolia*.²¹⁹ Four esters **621–624** of the 19(18 \rightarrow 17)-*abeo*-28-

noroleanane phlomstetraol B have been isolated from *Leonurus heterophyllus*.²²⁰

Two oleanane saponins, with the new genin olean-12ene2 β ,15 α ,23-triol **625**, have been isolated from *Ammannia auriculata*.²²¹ Antoniosides E–J are oleanane saponins from *Antonia ovata*.²²² Antioniosides E–H have new genins that are esters of olean-12-ene-3 β ,15 α ,16 α ,21 β ,22 α ,23,28-heptol **626**. Sorbifoliasides G–J, from *Xanthoceras sorbifolia*, have known genins and sorbifoliaside K has the new genin oleana-12,15diene-3 β ,21 β ,22 α ,28-tetrol **627**.²²³ A variety of new 30-noroleanane genins, **628–631**, are included in akemisaponins A–K from *Akebia trifoliate*.²²⁴ Two saponins from *Camellia japonica* have been assigned the names camelliosides E and F that have been used previously.²²⁵ Camellioside E has the new genin 28norolean-12-ene-3 β ,16 α ,17 β -triol **632**. Four saponins have been isolated from *Astragallus angustifolius* including the new genin olean-12-ene-3 β ,21 β ,22 α ,24,29-pentol **633**.⁶⁸

New oleanane saponins with known genins that have been assigned trivial names are listed in Table 1.







544 $R^1 = R^2 = \beta OH, H$ **545** $R^1 = O; R^2 = \alpha OH, H$ **546** $R^1 = \beta OH, H; R^2 = O$













0

H₃C(CH₂)₁₄





















R¹

R³

589 R¹ = βOH, H; R² = H; R³ = OH **590** R¹ = O; R² = H; R³ = OH **591** R¹ = O; R² = R³ = OH **592** R¹ = O; R² = OH; R³ = H **593** R¹ = βOH, H; R² = OH; R³ = H

594 $R^1 = \beta OH$, H; $R^2 = \alpha OH$, H **595** $R^1 = O$; $R^2 = \alpha OH$, H **596** $R^1 = R^2 = O$

 R^2











Open Access Article. Published on 24 October 2014. Downloaded on 7/19/2025 1:07:28 AM.

ЪОН

′СО₂Н





HO 621 R = H 622 R = OMe 623 R = H; Z-isomer 624 R = OMe; Z-isomer



619 R¹ = OH; R² = CH₃

620 R¹ = H; R² = CHO



629 R = CHO **630** R = CHO; Δ^{19} -isomer **631** R = CHO; $\Delta^{20(29)}$ -isomer

Table 1

Trivial name	Plant species	Ref	the 13,27-cyclized o	
	T lane species	1.01.	C 636, from Donell	
Aesculiosides G1–G16	Aesculus glabra	226	analyses. ²⁹¹ The d	
Afrocyclamins A and B	Cyclamen africanum	227		
Anemonerivulariside A	Anemone rivularis	228		
Asperosaponins A–C	Dipsacus Asper	229		
Besylvosides I–VI	Aegilops geniculata	230	Table 2	
Bigeloviis A and B	Salicornia bigelovii	231		
Bodiniosides C and D	Elsholtzia bodinieri	232	Plant species	
Bonarienosides A–E	Hydrocotyle bonariensis	233		
Bunkankasaponin F	Xanthoceras sorbifolia	234	Abrus precatorius	
Centelloside D	Centella asiatica	235	Abuta grandifolia	
Chakasaponin IV	Camellia sinensis	236	Acanthopanax senticos	
Dumortierinoside A methyl ester	Isolatocereus dumortieri	237	Acanthophyllum evpso	
Elmalienosides A–C	Cephalaria elmaliensis	238	Alhagi maurorum	
Entadosides A–D	Entada phaseoloides	239	Anemone amurensis	
Gardenisides A–C	Gardenia jasminoides	240	Aralia elata	
Guaianin P	Guaiacum officinale	241	Aralia taibaiensis	
Gummososide A, gummososide A	Stenocereus alamosensis	237	Ardisia gigantifolia	
methyl ester			Callicarpa integerrima	
Hareftoside E	Astragalus hareftae	78	Chenopodium foliosun	
Hederifoliosides A-E	Cyclamen hederifolium	242	Clematis argentilucida	
Ilexsaponins D–F	Ilex pubescens	243	Cvperus rotundus	
Ilexsaponin D (duplicate name)	Ilex pubescens	244	Dizygotheca elegantiss	
Kalopanaxsaponins L and M	Kalopanax pictus	245	Eragrostis tef	
Lobatosides L and M	Actinostemma lobatum	246	Grangea maderaspata	
Lonimacranthoides IV and V	Lonicera macranthoides	247	Gvmnema svlvestre	
Mimengosides H and I	Buddleja lindleyana	248	Gvmnocladus chinensi.	
Oleiferasaponin A ₁	Camellia oleifera	249	Gvpsophila trichotoma	
Paviosides A–H	Aesculus pavia	250	Kalopanax pictus	
Pleurosaponins A–K	Pleurospermum	251	Kalopanax septemlobu	
	kamtschaticum		Maesa lanceolata	
Polygalasaponins LI–LIII	Polygala japonica	252	Patrinia scabiosifolia	
Raddeanosides R ₂₀ -R ₂₂	Anemone raddeana	253	Pometia pinnata	
Sanchakasaponins A–D	Camellia japonica	254	Salicornia europaea	
Sanchakasaponins E–H	Camellia japonica	255	Salicornia herbacea	
Sandrosaponin XI	Ferula hermonis	256	Samanea saman	
Senaciapittosides A and B	Pittosporum senacia	257	Sideroxylon obtusifoliı	
Silenoviscoside F	Silene viscidula	258	Tarenna grevei	
Sorbifoliasides A–F	Xanthoceras sorbifolia	234	Wedelia chinensis	
Teaseedsaponins A–L	Camellia sinensis	259	Xanthoceras sorbifolia	
Treleaseside A	Stenocereus eruca	180		

The sources of new oleanane saponins with known genins that have not been assigned trivial names are listed in Table 2.

Pachanosides I1 and D1 are pachanane saponins with known genins from *Isolatocereus dumortieri*.²³⁷ The structures of the 13,27-cyclized oleananes donellanic acids A **634**, B **635** and C **636**, from *Donella ubanguiensis*, were all established by X-ray analyses.²⁹¹ The donellanic acids are accompanied by two

e 2 t species Ref. s precatorius 260 a grandifolia 261 thopanax senticosus 262 thophyllum gypsophiloides 263 gi maurorum 264none amurensis 265 ia elata 266 a taibaiensis 267 sia gigantifolia 268 icarpa integerrima 269 10podium foliosum 270 atis argentilucida 271 and 272 erus rotundus 273 gotheca elegantissima 274 rostis tef 275 igea maderaspatana 276 nema sylvestre 277 nocladus chinensis 278 ophila trichotoma 279 panax pictus 245 panax septemlobus 280 sa lanceolata 281 inia scabiosifolia 2.82 etia pinnata 283 cornia europaea 284 cornia herbacea 285 anea saman 286 roxylon obtusifolium 287

288 289

290



further 13,17-cyclized compounds that have been assigned the tentative structures **637** and **638**. Ilelic acid B **639** is a rearranged oleanane with a seven-membered C-ring from *Ilex latifolia*.²⁹² Four taraxerane derivatives **640–643** have been identified in *Saussurea graminea*.²⁹³

Cassinolide **644**, from *Cassine xylocarpa*, is a 2,3secofriedelan-3,24-olide derivative.²⁹⁴ *Maytenus robusta* is the source of the 3,4-secofriedelan-3,11β-olide **645** (ref. 295) and friedelane-3β,11β-diol **646**.²⁹⁶ Reissantiadiol, from *Reisantia grahamii*, has been identified as friedelane-2 α ,3 α -diol **647**.²⁹⁷





Other simple friedelane triterpenoids include 30-hydroxyfriedel-1-en-3-one **648** from *Salacia hainanensis*,¹²⁴ **649–653** from *Celastrus vulcanicola* and **654–658** from *Maytenus jelskii*²⁹⁸ and glaucalactone **659** from *Caloncoba glauca*.⁶¹

A review of the pharmacology of celastrol, a norfriedlane derivative, has been published.²⁹⁹ Hypoglaside A **660** is a dinorfriedelane glucoside from *Tryperygium hypoglaucum*.³⁰⁰ *Celastrus orbiculatus* is the source of a range of norfriedelane and methyl-migrated derivatives.³⁰¹ The $25(9 \rightarrow 8)$ -*abeo*

norfriedelane **661** and the 25(9 \rightarrow 7)-*abeo* derivative **662** are accompanied by the 8,14-*seco* compounds **663** and **664**. A biosynthetic pathway to these rearranged friedelane derivatives has been proposed. Also present in *Celastrus orbiculatus* are the "dimeric" norfriedelanes celastrolines A α **665** and A β **666** and isocelastroline A α **667**, together with celastrolines B α **668** and B β **669** that have a linkage to a podocarpane derivative.





7. The ursane group

Ursane triterpenoids have shown potential as anticancer drugs.³⁰² Reviews on the anticancer activities of ursolic acid³⁰³ and acetyl-11-keto- β -boswellic acid (AKBA)³⁰⁴ have been published.

The 18,19-secoursane derivative **670** has been isolated from the bark of *Diospyros decandra* together with **671**.¹⁹⁵ The 17,22*seco* derivative **672** has been found in both *Salvia palaestina* and *Salvia syriaca*.³⁰⁵ Euscaphic acids J **673**, K **674** and L **675** have been isolated from *Euscaphis japonica*.¹⁹⁷ Euscaphic acid L **675** has a contracted ring-A. Negundonorins A **676** and B **677**, from *Vitex negundo*, are 24-nor and 28-noruranes, respectively.³⁰⁶ Further 24-norursanes include ulmoidol A **678** from *Eucommia ulmoides*³⁰⁷ and the related compounds **679** and **680** from *Dipsacus chinensis*.³⁰⁸ 30-Norurs-11-en-3 α -ol **681** has been identified in the roots of *Alhagi camelorum*.³⁰⁹

6β,20β-Dihydroxyurs-12-en-28-oic acid **682**, from leaves of *Psidium guajava*, is unusual as it lacks oxygenation at C-3.³¹⁰ The structure of 3β,19α,23,24-tetrahydroxyurs-12-en-28-oic acid **683**, from *Nauclea officinalis*, was confirmed by X-ray analysis.³¹¹ It is accompanied by the corresponding 2β,3β,19α,24-tetrahydroxy compound **684**. Further simple ursane derivatives include the acetal **685** from *Juglans sinensis*,²⁰⁴ rhododendric acid A **686**

NPR





HC

нό

686

HO

`CO₂Me

Kirmanoic acid **702**, from *Nepeta clarkei*, is an unusual alkylphenyl ether.¹⁸⁵ New ursane esters include the isoferuloyl ester **703** from *Eucalyptus exserta*,²⁰⁵ ehretiolide **704** from *Ehretia longifolia*,³²¹ the 30-*cis*-coumaryl ester **705** from *Rubia schumanniana*,²¹⁶ 3β-tetradecanoyloxyurs-12-en-28,19β-olide **706** from *Lysimachia clethroides*³²² and 3-*epi*-cecropic acid **707** from *Dipterocarpus obtusifolius*.¹⁰⁶

CO₂H

687

Sanguisoside A is an ursane saponin from *Sanguisorba officinalis* with the new 18,19-*seco*-ursane genin **708**.³²³ Centelloside E from *Centella asiatica* has the new genin **709** (ref. 235) and the genins **710** and **711** are present in saponins from *Actinidia valvata*.³²⁴











713

AcC

HO

HO

Ursane saponins with known genins include asphorins A and B from *Asphodelus tenuifolius*,³²⁵ asprellanosides A and B from *Ilex asprella*,³²⁶ elasticoside from *Ficus elastica*,³²⁷ ilekudinchosides F and G from *Ilex kudincha*,³²⁸ ilemaminosides A and B from *Ilex mamillata*,³²⁹ ilexasosides A-H from *Ilex asprella*,³³⁰ phillyriside A from *Stenocereus eruca*,¹⁸⁰ and unnamed saponins from *Diospyros decandra*,¹⁹⁵ *Juglans sinensis*,²⁰⁴ *Lantana camara*³³¹ and *Psidium guajava*.³³²

712

The taraxastane hydroperoxide derivatives **712** and **713** have been isolated from the roots of *Taraxacum platycarpum*.¹⁷³ Psiguanin A, which is 2α ,3β-dihydroxytaraxast-20-en-28-oic acid **714**, has been found in the leaves of *Psidium guajava*.³¹⁵ Other new taraxastane derivatives include chlorotolpidiol **715** and tolpidiol A **716** from *Tolpis proustii* and *Tolpis lagopoda*,³²⁰ pergularines A **717** and B **718** from *Pergularia tomentosa*³³³ and calotroprocerol A **719**, calotroprocerone A **720** and calotroproceryl acetates A **721** and B **722** from *Calotropis procera*.³³⁴ Two taraxastane saponins with known genins have been found in the roots of *Ilex pubescens*.³³⁵

714

The 27(14 \rightarrow 13)-*abeo*-urs-14-enone derivative 723, from *Rubia schumanniana*, has the unusual β -configuration for the migrated methyl.²¹⁶ The rearranged ursanes ilelic acids A 724, C



725 and D **726** have been isolated from *Ilex latifolia*.²⁹² The structure of ilelic acid D **726** was confirmed by X-ray analysis.

8. The hopane group

3,4-Secohop-22(29)-en-3-oic acid 727 has been isolated from *Maytenus robusta*.²⁹⁵ The scale insect pathogenic fungus *Aschersonia calendulina* is the source of two hopane metabolites 728 and 729.³³⁶ The leaves of *Hybanthus austro-caledonicus* produce 3-epiwoodwardinic acid 730.³³⁷ Plakohopanoid 731, a C₃₂ hopanoid ester of a manosyl myoinostol, has been isolated from the sponge *Plakortis cf. lita*.³³⁸ The structure of plakohopanoid 731 implies that it is of bacterial origin. This is the first example of a biologically produced C₃₂ hopanoic acid. Such

acids were considered to only be geohopanoids formed by abiotic degradation of bacteriohopanoids. Oppositifolone 732, from *Glinus oppositifolius*, has a $29(22 \rightarrow 21)$ -*abeo*-hopane skeleton and is the 3-ketone of spergulagenin A.³³⁹

Pteroxygonumnol **733**, from the roots of *Pteroxygonum giraldii*, has been identified as 2β ,25: 19β ,28-diepoxyarborin-9(11)-ene- 3β , 7β -diol.³⁴⁰ Five 25-norarborinane derivatives **734**–**738** have been isolated from bamboo stems, *Sinocalamus affinis*.¹⁸⁷ The structure of **734** was confirmed by X-ray analysis. *Atalantia retusa* is the source of the 17,21-*seco*-arborane derivative retusinol **739**.³⁴¹ Peniciside **740** is a fernane metabolite of *Penicillium* sp.169³⁴² and 3β -acetoxyfern-7-en-6-one **741** is a constituent of *Scorzonera latifolia*.³⁴³

Review





 $R^1 = \alpha OH$, H; $R^2 = R^3 = H$ $R^1 = \alpha OH$, H; $R^2 = OH$; $R^3 = H$ $R^1 = O$; $R^2 = R^3 = H$ $R^1 = \alpha OH$, H; $R^2 = H$; $R^3 = OH$



ŅН



R³

R²

9. Miscellaneous compounds

The unlikely structure **742** has been proposed for erucaoic acid from *Sonchus eruca*.³⁴⁴ The club moss *Lycopodium phlegmaria* is the source of the serratane esters lycophlegmariols A **743**–D **746** (ref. 345) and four esters **747–750** have been found in *Palhinhaea cernua* (syn. *Lycopodium cernuum*).³⁴⁶ The D:B-friedobaccharane derivatives leonatriol **751** and the corresponding ketone leonatriolone **752** have been isolated from *Cassine xylocarpa* and *Celastrus vulcanicola*, respectively.²⁹⁴ Globostelletins J **753–**S **761**, from the marine sponge *Rhabdastrella globostellata*, are isomalabaricanes with cyclopentane side chains.³⁴⁷

10. References

- 1 J. M. R. Patlolla and C. V. Rao, *Curr. Pharm. Biotechnol.*, 2012, 13, 147–155.
- 2 S. H. Safe, P. L. Prather, L. K. Brents, G. Chadalapaka and I. Jutooru, *Anti-Cancer Agents Med. Chem.*, 2012, **12**, 1211– 1220.
- 3 M. K. Shanmugam, A. H. Nguyen, A. P. Kumar, B. K. H. Tan and G. Sethi, *Cancer Lett.*, 2012, **320**, 158–170.
- 4 M.-C. Yin, Biomedicine, 2012, 2, 2-9.
- 5 J. Wang, J. Shan, L. Di, S. Wang and B. Cai, *Zhongcaoyao*, 2012, **43**, 196–200.
- 6 H. Wang, M.-Y. Li and J. Wu, *Chem. Biodiversity*, 2012, **9**, 1–11.
- 7 S. K. Kothiyal, S. C. Sati, M. S. M. Rawat, M. D. Sati, D. K. Semwal, R. B. Semwal, A. Sharma, B. Rawat and A. Kumar, *Nat. Prod. J.*, 2012, **2**, 212–224.
- 8 A. Upadhyay and D. K. Singh, *Rev. Inst. Med. Trop. Sao Paulo*, 2012, **54**, 273–280.
- 9 S. F. Kouam, S. Kusari, M. Lamshoeft, O. K. Tatuedom and M. Spiteller, *Phytochemistry*, 2012, 83, 79–86.
- 10 J. Lokvam and P. V. A. Fine, *Molecules*, 2012, **17**, 7451–7457.
- F. Cen-Pacheco, F. Mollinedo, J. A. Villa-Pulgarin, M. Norte, J. J. Fernandez and A. Hernandez Daranas, *Tetrahedron*, 2012, 68, 7275–7279.
- 12 S. Okada, Kagaku to Seibutsu, 2012, 50, 93-102.
- H.-G. Jin, Q. Jin, A. Ryun Kim, H. Choi, J. H. Lee, Y. S. Kim,
 D. G. Lee and E.-R. Woo, *Arch. Pharmacal Res.*, 2012, 35, 1919–1926.
- 14 N. Xu, H. Zhang and X. Xie, Zhongcaoyao, 2012, 43, 841-843.
- O. Yodsaoue, J. Sonprasit, C. Karalai, C. Ponglimanont, S. Tewtrakul and S. Chantrapromma, *Phytochemistry*, 2012, **76**, 83–91.
- 16 Y.-L. Li, S.-D. Zhang, H.-Z. Jin, J.-M. Tian, Y.-H. Shen, X.-W. Yang, H.-L. Li and W.-D. Zhang, *Tetrahedron*, 2012, 68, 7763–7767.
- 17 S. Lavoie, J. Legault, C. Gauthier, V. Mshvildadze, S. Mercier and A. Pichette, *Org. Lett.*, 2012, **14**, 1504–1507.
- 18 Y.-L. Li, Y.-X. Gao, X.-W. Yang, H.-Z. Jin, J. Ye, L. Simmons, N. Wang, A. Steinmetz and W.-D. Zhang, *Phytochemistry*, 2012, **81**, 159–164.
- 19 G. R. Wang, Y. L. Li, W. D. Zhang, X. W. Yang, W. C. Liu, J. Ye, Z. J. Zhu and H. Chen, *Chin. Chem. Lett.*, 2012, 23, 1251–1253.

- 20 J.-H. Xia, S.-D. Zhang, Y.-L. Li, L. Wu, Z.-J. Zhu, X.-W. Yang, H.-W. Zeng, H.-L. Li, N. Wang, A. Steinmetz and W.-D. Zhang, *Phytochemistry*, 2012, 74, 178–184.
- 21 X.-D. Wu, J. He, Y. Shen, L.-B. Dong, Z.-H. Pan, G. Xu, X. Gong, L.-D. Song, Y. Leng, Y. Li, L.-Y. Peng and Q.-S. Zhao, *Tetrahedron Lett.*, 2012, 53, 800–803.
- 22 J. Zou, L.-B. Yang, J. Jiang, Y.-Y. Diao, X.-N. Li, J. Huang, J.-H. Yang, H.-L. Li, W.-L. Xiao, X. Du, S.-Z. Shang, J.-X. Pu and H.-D. Sun, *Planta Med.*, 2012, **78**, 472–479.
- 23 K. Dong, J.-X. Pu, X. Du, J. Su, X.-N. Li, J.-H. Yang, W. Zhao, Y. Li and H.-D. Sun, *Tetrahedron*, 2012, **68**, 4820–4829.
- 24 C.-Q. Liang, Y.-M. Shi, R.-H. Luo, X.-Y. Li, Z.-H. Gao, X.-N. Li, L.-M. Yang, S.-Z. Shang, Y. Li, Y.-T. Zheng, H.-B. Zhang, W.-L. Xiao and H.-D. Sun, *Org. Lett.*, 2012, 14, 6362–6365.
- 25 N. Wang, Z.-l. Li, D.-d. Song, W. Li, Y.-h. Pei, Y.-k. Jing and H.-M. Hua, *Planta Med.*, 2012, **78**, 1661–1666.
- 26 G. She, N. Zhu, S. Wang, Y. Liu, Y. Ba, C. Sun and R. Shi, *Chem. Cent. J.*, 2012, **6**, 39.
- 27 S. Jeelani and M. A. Khuroo, *Nat. Prod. Res.*, 2012, **26**, 654–658.
- 28 N. Handa, T. Yamada and R. Tanaka, *Phytochem. Lett.*, 2012, 5, 480–485.
- 29 J.-H. Lee, J.-E. Jeon, Y.-J. Lee, H.-S. Lee, C. J. Sim, K.-B. Oh and J. Shin, *J. Nat. Prod.*, 2012, **75**, 1365–1372.
- 30 M. Ono, Y. Takatsu, T. Ochiai, S. Yasuda, Y. Nishida, T. Tanaka, M. Okawa, J. Kinjo, H. Yoshimitsu and T. Nohara, *Chem. Pharm. Bull.*, 2012, **60**, 1314–1319.
- 31 J.-Q. Liu, C.-F. Wang, Y. Li, H.-R. Luo and M.-H. Qiu, *Planta Med.*, 2012, **78**, 368–376.
- 32 S.-X. Yang, Z.-C. Yu, Q.-Q. Lu, W.-Q. Shi, H. Laatsch and J.-M. Gao, *Phytochem. Lett.*, 2012, 5, 576–580.
- 33 B.-J. Ma, Y. Zhou, Y. Ruan, J.-C. Ma, W. Ren and C.-N. Wen, J. Antibiot., 2012, 65, 165–167.
- 34 Y. Li, Z. Zhu, W. Yao and R. Chen, *Zhongguo Zhongyao Zazhi*, 2012, 37, 165–171.
- 35 Y. Li, Z.-M. Zhu, W.-X. Yao and R.-Y. Chen, *Med. Plant*, 2012, 3, 75–81.
- 36 J.-L. Rios, I. Andujar, M.-C. Recio and R.-M. Giner, *J. Nat. Prod.*, 2012, **75**, 2016–2044.
- 37 T. K. Lai, G. Biswas, S. Chatterjee, A. Dutta, C. Pal, J. Banerji,
 N. Bhuvanesh, J. H. Reibenspies and K. Acharya, *Chem. Biodiversity*, 2012, 9, 1517–1524.
- 38 Y.-Z. Liu, Y.-Y. Li, Y.-F. Sun, Z.-H. Zheng, S.-Y. Song, W.-J. Su and Y.-M. Shen, *Helv. Chim. Acta*, 2012, **95**, 282–285.
- 39 H.-C. Huang, C.-C. Liaw, H.-L. Yang, Y.-C. Hseu, H.-T. Kuo, Y.-C. Tsai, S.-C. Chien, S. Amagaya, Y.-C. Chen and Y.-H. Kuo, *Phytochemistry*, 2012, 84, 177–183.
- 40 I.-K. Lee, J.-Y. Jung, J.-H. Yeom, D.-W. Ki, M.-S. Lee, W.-H. Yeo and B.-S. Yun, *Mycobiology*, 2012, **40**, 76–78.
- 41 A. S. Silchenko, A. I. Kalinovsky, S. A. Avilov,
 P. V. Andryjashchenko, P. S. Dmitrenok, V. I. Kalinin and
 V. A. Stonik, *Biochem. Syst. Ecol.*, 2012, 44, 53–60.
- 42 Z. Wang, H. Zhang, W. Yuan, W. Gong, H. Tang, B. Liu,
 K. Krohn, L. Li, Y. Yi and W. Zhang, *Food Chem.*, 2012, 132, 295–300.

- 43 A. S. Silchenko, A. I. Kalinovsky, S. A. Avilov,
 P. V. Andryjaschenko, P. S. Dmitrenok, E. A. Martyyas and
 V. I. Kalinin, *Nat. Prod. Commun.*, 2012, 7, 1157–1162.
- 44 A. S. Silchenko, A. I. Kalinovsky, S. A. Avilov, P. V. Andryjaschenko, P. S. Dmitrenok, E. A. Yurchenko and V. I. Kalinin, *Nat. Prod. Res.*, 2012, **26**, 1765–1774.
- 45 S.-K. Kim and S. W. A. Himaya, *Adv. Food Nutr. Res.*, 2012, 65, 297–319.
- 46 X. Luo, Y.-M. Shi, R.-H. Luo, S.-H. Luo, X.-N. Li, R.-R. Wang, S.-H. Li, Y.-T. Zheng, X. Du, W.-L. Xiao, J.-X. Pu and H.-D. Sun, Org. Lett., 2012, 14, 1286–1289.
- 47 F. He, X.-Y. Li, G.-Y. Yang, X.-N. Li, X. Luo, J. Zou, Y. Li, W.-L. Xiao and H.-D. Sun, *Tetrahedron*, 2012, **68**, 440–446.
- 48 J.-R. Wang, T. Kurtan, A. Mandi and Y.-W. Guo, *Eur. J. Org. Chem.*, 2012, **2012**, 5471–5482.
- 49 J. Zou, J. Jiang, Y.-Y. Diao, L.-B. Yang, J. Huang, H.-L. Li, X. Du, W.-L. Xiao, J.-X. Pu and H.-D. Sun, *Fitoterapia*, 2012, 83, 926–931.
- 50 F. Qiu, A. Imai, J. B. McAlpine, D. C. Lankin, I. Burton, T. Karakach, N. R. Farnsworth, S.-N. Chen and G. F. Pauli, *J. Nat. Prod.*, 2012, 75, 432–443.
- 51 B. Li, D.-Y. Kong, Y.-H. Shen, H. Yuan, R.-C. Yue, Y.-R. He, L. Lu, L. Shan, H.-L. Li, J. Ye, X.-W. Yang, J. Su, R.-H. Liu and W.-D. Zhang, *Org. Lett.*, 2012, 14, 5432–5435.
- 52 Q.-H. Han, X. Liu, W.-Q. Yao, Z.-B. Cheng, T.-T. Lin, C. Song and S. Yin, *Planta Med.*, 2012, 78, 1971–1975.
- 53 C. Long, J. Beck, F. Cantagrel, L. Marcourt, L. Vendier, B. David, F. Plisson, F. Derguini, I. Vandenberghe, Y. Aussagues, F. Ausseil, C. Lavaud, F. Sautel and G. Massiot, *J. Nat. Prod.*, 2012, 75, 34–47.
- 54 J. D. Simo Mpetga, Y. Shen, P. Tane, S.-F. Li, H.-P. He, H. K. Wabo, M. Tene, Y. Leng and X.-J. Hao, *J. Nat. Prod.*, 2012, 75, 599–604.
- 55 I. Lee, J. Kim, Y. S. Kim, N. H. Yoo, C.-S. Kim, K. Jo, J.-H. Kim, T. T. Bach and J. S. Kim, *J. Nat. Prod.*, 2012, 75, 1312–1318.
- 56 J.-L. Yang and Y.-P. Shi, Planta Med., 2012, 78, 59-64.
- 57 L. P. Ponomarenko, A. I. Kalinovsky, E. A. Martyyas, R. V. Doudkin, P. G. Gorovoy and V. A. Stonik, *Phytochem. Lett.*, 2012, 5, 118–122.
- 58 M. A. Ramirez-Cisneros, M. Y. Rios, R. Rios-Gomez and A. B. Aguilar-Guadarrama, *Planta Med.*, 2012, 78, 1942– 1948.
- 59 K. Toume, T. Nakazawa, T. Hoque, T. Ohtsuki, M. A. Arai, T. Koyano, T. Kowithayakorn and M. Ishibashi, *Planta Med.*, 2012, **78**, 1370–1377.
- 60 S.-F. Li, Y.-T. Di, R.-H. Luo, Y.-T. Zheng, Y.-H. Wang, X. Fang, Y. Zhang, L. Li, H.-P. He, S.-L. Li and X.-J. Hao, *Planta Med.*, 2012, **78**, 821–827.
- 61 J. D. S. Mpetga, M. Tene, H. K. Wabo, S.-F. Li, L.-M. Kong, H.-P. He, X.-J. Hao and P. Tane, *Phytochem. Lett.*, 2012, 5, 183–187.
- K. Awang, X.-M. Loong, K. H. Leong, U. Supratman, M. Litaudon, M. R. Mukhtar and K. Mohamad, *Fitoterapia*, 2012, 83, 1391–1395.
- 63 J.-L. Yang and Y.-P. Shi, *Phytochemistry*, 2012, **76**, 124–132.

- 64 L. Lu, J.-C. Chen, Y. Li, C. Qing, Y.-Y. Wang, Y. Nian and M.-H. Qiu, *Chem. Pharm. Bull.*, 2012, **60**, 571–577.
- 65 Y. Nian, H.-Y. Wang, J. Su, L. Zhou, G. Feng, Y. Li and M.-H. Qiu, *Tetrahedron*, 2012, **68**, 6521–6527.
- 66 H. X. Kuang, Y. Su, Q. H. Wang, L. Wu, B. Y. Yang,
 Z. B. Wang and Y. G. Xia, *Planta Med.*, 2012, 78, 622–625.
- 67 M. K. Jamróz, M. H. Jamróz, J. C. Dobrowolski, J. A. Gliński and M. Gleńsk, *Spectrochim. Acta, Part A*, 2012, 93, 10–18.
- 68 D. Gülcemal, M. Masullo, E. Bedir, M. Festa, T. Karayildirim, O. Alankus-Caliskan and S. Piacente, *Planta Med.*, 2012, **78**, 720–729.
- 69 T. K. Naubeev, K. K. Uteniyazov, M. I. Isaev, V. V. Kachala and A. S. Shashkov, *Chem. Nat. Compd.*, 2012, **48**, 810–812.
- 70 F. N. Yalçin, S. Piacente, A. Perrone, A. Capasso, H. Duman and I. Calis, *Phytochemistry*, 2012, 73, 119–126.
- 71 F. Karabey, I. A. Khan and E. Bedir, *Phytochem. Lett.*, 2012, 5, 320–324.
- 72 L.-B. Wei, J.-M. Chen, W.-C. Ye, X.-S. Yao and G.-X. Zhou, J. Asian Nat. Prod. Res., 2012, 14, 521–527.
- 73 A.-X. Zuo, Y. Shen, Z.-Y. Jiang, X.-M. Zhang, J. Zhou, J. Lue and J.-J. Chen, *J. Asian Nat. Prod. Res.*, 2012, **14**, 407–412.
- 74 X.-T. Zhang, S.-W. Ma, H.-Y. Jiao and Q.-W. Zhang, J. Asian Nat. Prod. Res., 2012, 14, 327–332.
- 75 K. Pudhom, T. Nuanyai and K. Matsubara, *Chem. Pharm. Bull.*, 2012, **60**, 1538–1543.
- 76 T. K. Naubeev, A. A. Zhanibekov and M. I. Isaev, *Chem. Nat. Compd.*, 2012, 48, 813–815.
- 77 M. D. Alaniya and T. I. Gigoshvili, *Chem. Nat. Compd.*, 2012, 48, 914–916.
- 78 I. Horo, E. Bedir, M. Masullo, S. Piacente, F. Özgökçe and Ö. Alankuş-Çalişkan, *Phytochemistry*, 2012, 84, 147–153.
- 79 M.-m. Zhang, Y.-l. Liu, Z. Chen, X.-r. Li, Q.-m. Xu and S.-l. Yang, *Zhongcaoyao*, 2012, 43, 1462–1470.
- 80 T. Savran, D. Gülcemal, M. Masullo, T. Karayildirim, E. Polat, S. Piacente and Ö. Alankuş-Çalişkan, *Rec. Nat. Prod.*, 2012, 6, 230–236.
- 81 J.-C. Chen, Z.-Z. Xu, L.-X. Yang, X.-Y. He, C. Chen, G.-H. Zhang, Y.-T. Zheng and M.-H. Qiu, *Chem. Nat. Compd.*, 2012, 48, 591–593.
- 82 L. Pan, Y. Yong, Y. Deng, D. D. Lantvit, T. N. Ninh, H. Chai,
 E. J. Carcache de Blanco, D. D. Soejarto, S. M. Swanson and
 A. D. Kinghorn, *J. Nat. Prod.*, 2012, 75, 444–452.
- 83 J.-C. Chen, L. Zhou, Y.-H. Wang, R.-R. Tian, Y.-X. Yan, Y. Nian, Y. Sun, Y.-T. Zheng and M.-H. Qiu, *Nat. Prod. Bioprospect.*, 2012, 2, 138–144.
- 84 Y.-W. Liao, C.-R. Chen, Y.-H. Kuo, J.-L. Hsu, W.-L. Shih, H.-L. Cheng, T.-C. Huang and C.-I. Chang, *Nat. Prod. Commun.*, 2012, 7, 1575–1578.
- 85 C.-A. Geng, X.-Y. Huang, L.-G. Lei, X.-M. Zhang and J.-J. Chen, *Chem. Biodiversity*, 2012, **9**, 1508–1516.
- 86 J. Zhang, Y. Huang, T. Kikuchi, H. Tokuda, N. Suzuki, K.-I. Inafuku, M. Miura, S. Motohashi, T. Suzuki and T. Akihisa, *Chem. Biodiversity*, 2012, 9, 428–440.
- 87 X.-M. Fan, G. Chen, Y. Sha, X. Lu, M.-X. Shen, H.-M. Ma and Y.-H. Pei, *J. Asian Nat. Prod. Res.*, 2012, **14**, 528–532.
- 88 M. Clericuzio, C. Cassino, F. Corana and G. Vidari, *Phytochemistry*, 2012, **84**, 154–159.

- 89 B. G. Panlilio, A. P. G. Macabeo, M. Knorn, P. Kohls, P. Richomme, S. F. Kouam, D. Gehle, K. Krohn, S. G. Franzblau, Q. Zhang and M. A. M. Aguinaldo, *Phytochem. Lett.*, 2012, 5, 682–684.
- 90 S. L. Deore, A. Parab, B. A. Baviskar and S. S. Khadabadi, *Pharm. Rev.*, 2012, **10**, 125–127.
- 91 K. Dhiman, A. Gupta, D. K. Sharma, N. S. Gill and A. Goyal, *Asian J. Clin. Nutr.*, 2012, 4, 16–26.
- 92 X. Chen, J. Bao, J. Guo, Q. Ding, J. Lu, M. Huang and Y. Wang, *Anti-Cancer Drugs*, 2012, 23, 777–787.
- 93 J. L. Rios, I. Andujar, J. M. Escandell, R. M. Giner and M. C. Recio, *Curr. Pharm. Des.*, 2012, **18**, 1663–1676.
- 94 M. Cao, H.-S. Yu, X.-B. Song and B.-P. Ma, *Yaoxue Xuebao*, 2012, 47, 836–843.
- 95 K.-K. Li, C.-M. Yao and X.-W. Yang, *Planta Med.*, 2012, 78, 189–192.
- 96 K.-K. Li, X.-B. Yang, X.-W. Yang, J.-X. Liu and X.-J. Gong, *Fitoterapia*, 2012, **83**, 1030–1035.
- 97 H.-Y. Ma, H.-Y. Gao, J. Huang, B.-H. Sun and B. Yang, J. Nat. Med., 2012, 66, 576–582.
- 98 W. Li, J. Y. Yin, Z. Y. Cong, Y. Liu, Y. Zhang, Y. Y. Li and Q. Meng, *Chem. Nat. Compd.*, 2012, 48, 1017–1020.
- 99 J.-P. Liu, D. Lu and P.-Y. Li, *Nat. Prod. Res.*, 2012, **26**, 744-748.
- 100 L. Ma, W.-J. Xiang, P. Van Khang, Y. Liang, Y. Xiao and L.-H. Hu, *Planta Med.*, 2012, **78**, 597–605.
- 101 L. Shi, X.-J. Meng, J.-Q. Cao and Y.-Q. Zhao, *Nat. Prod. Res.*, 2012, **26**, 1419–1422.
- 102 S. Li, B. Cui, Q. Liu, L. Tang, Y. Yang, X. Jin and Z. Shen, *Planta Med.*, 2012, **78**, 290–296.
- M. Gan, M. Liu, L. Gan, S. Lin, B. Liu, Y. Zhang, J. Zi,
 W. Song and J. Shi, *J. Nat. Prod.*, 2012, 75, 1373–1382.
- 104 D. Harneti, R. Tjokronegoro, A. Safari, U. Supratman, X.-M. Loong, M. R. Mukhtar, K. Mohamad, K. Awang and H. Hayashi, *Phytochem. Lett.*, 2012, 5, 496–499.
- 105 F. Wang and Y. Guan, Fitoterapia, 2012, 83, 13-17.
- 106 P. Khiev, O.-K. Kwon, H.-H. Song, S.-R. Oh, K.-S. Ahn, H.-K. Lee and Y.-W. Chin, *Chem. Pharm. Bull.*, 2012, 60, 955–961.
- 107 F.-F. Xu, X.-Q. Zhang, J. Zhang, B. Liu, J. Jiang, W.-J. Wang, M.-H. Gao, R.-W. Jiang and W.-C. Ye, *J. Asian Nat. Prod. Res.*, 2012, **14**, 135–140.
- 108 K. Yoshizaki and S. Yahara, *Chem. Pharm. Bull.*, 2012, **60**, 354–362.
- 109 K. Yoshizaki, M. Murakami, H. Fujino, N. Yoshida and S. Yahara, *Chem. Pharm. Bull.*, 2012, **60**, 728–735.
- 110 L. Shi, F. Lu, H. Zhao and Y.-Q. Zhao, *J. Asian Nat. Prod. Res.*, 2012, **14**, 856–861.
- 111 J.-P. Liu, F. Wang, P.-Y. Li and D. Lu, Nat. Prod. Res., 2012, 26, 731–735.
- 112 H. T. Nguyen and Y. Shoyama, *Chem. Pharm. Bull.*, 2012, **60**, 1329–1333.
- 113 J.-S. Wang, Y. Zhang, X.-B. Wang, D.-D. Wei, J. Luo, J.-G. Luo, M.-H. Yang, H.-Q. Yao, H.-B. Sun and L.-Y. Kong, *Tetrahedron Lett.*, 2012, 53, 1705–1709.
- 114 Y. Zhang, J. Wang, P. Wang and L. Kong, *Chin. J. Chem.*, 2012, **30**, 1356–1360.

- 115 J.-S. Wang, Y. Zhang, X.-B. Wang, D.-D. Wei, J. Luo, J.-G. Luo, M.-H. Yang, H.-Q. Yao, H.-B. Sun and L.-Y. Kong, *Tetrahedron Lett.*, 2012, 53, 4030.
- 116 X.-Y. Wang, G.-H. Tang, C.-M. Yuan, Y. Zhang, L. Hou, Q. Zhao, X.-J. Hao and H.-P. He, *Nat. Prod. Bioprospect.*, 2012, 2, 222–226.
- 117 Q. Zeng, B. Guan, J.-J. Qin, C.-H. Wang, X.-R. Cheng, J. Ren, S.-K. Yan, H.-Z. Jin and W.-D. Zhang, *Phytochemistry*, 2012, 80, 148–155.
- 118 T. Tang, S.-G. Liao, Z. Na, Y. Li and Y.-K. Xu, *Tetrahedron Lett.*, 2012, **53**, 1183–1185.
- 119 M. A. Versiani, A. Ikram, S. Khalid, S. Faizi and I. A. Tahiri, *Nat. Prod. Commun.*, 2012, 7, 831–834.
- 120 E. N. Happi, A. T. Tcho, J. C. Sirri, J. D. Wansi, B. Neumann, H.-G. Stammler, J. Wandji and N. Sewald, *Phytochem. Lett.*, 2012, **5**, 423–426.
- 121 T. Fossen, P. Rasoanaivo, C. S. Manjovelo, F. H. Raharinjato, S. Yahorava, A. Yahorau and J. E. S. Wikberg, *Fitoterapia*, 2012, 83, 901–906.
- 122 Q. Zhao, Y. Song, C. Feng and H. Chen, Arch. Pharmacal Res., 2012, 35, 1903–1907.
- 123 W.-X. Liu, G.-H. Tang, H.-P. He, Y. Zhang, S.-L. Li and X.-J. Hao, *Nat. Prod. Bioprospect.*, 2012, **2**, 29–34.
- 124 J. Huang, Z.-h. Guo, P. Cheng, B.-h. Sun and H.-Y. Gao, *Phytochem. Lett.*, 2012, 5, 432–437.
- 125 I. Valente, M. Reis, N. Duarte, J. Serly, J. Molnár and M.-J. U. Ferreira, *J. Nat. Prod.*, 2012, 75, 1915–1921.
- 126 F. Zhang, X.-F. He, W.-B. Wu, W.-S. Chen and J.-M. Yue, *Nat. Prod. Bioprospect.*, 2013, **2**, 235–239.
- 127 M.-H. Yang, J.-S. Wang, J.-G. Luo, X.-B. Wang and L.-Y. Kong, *Can. J. Chem.*, 2012, **90**, 199–204.
- 128 S.-H. Dong, X.-F. He, L. Dong, Y. Wu and J.-M. Yue, *Helv. Chim. Acta*, 2012, **95**, 286–300.
- M. S. A. Rao, G. Suresh, P. Ashok Yadav, K. Rajendra Prasad,
 V. Lakshma Nayak, S. Ramakrishna, C. V. Rao and
 K. S. Babu, *Tetrahedron Lett.*, 2012, 53, 6241–6244.
- 130 J. Liu, S.-P. Yang, G. Ni, Y.-C. Gu and J.-M. Yue, *J. Asian Nat. Prod. Res.*, 2012, **14**, 929–939.
- 131 J. Sichaem, T. Aree, S. Khumkratok, J. Jong-aramruang and S. Tip-pyang, *Phytochem. Lett.*, 2012, **5**, 665–667.
- 132 F. Zhang, J.-S. Wang, Y.-C. Gu and L.-Y. Kong, *J. Nat. Prod.*, 2012, **75**, 538–546.
- 133 A. K. R. Bandi and D.-U. Lee, *Chem. Biodiversity*, 2012, 9, 1403–1421.
- 134 M.-L. Han, H. Zhang, S.-P. Yang and J.-M. Yue, *Org. Lett.*, 2012, **14**, 486–489.
- 135 J. Li, M.-Y. Li, T. Bruhn, D. C. G. Goetz, Q. Xiao, T. Satyanandamurty, J. Wu and G. Bringmann, *Chem.– Eur. J.*, 2012, 18, 14342–14351.
- 136 V. G. P. Severino, P. A. C. Braga, M. F. d. G. F. da Silva, J. B. Fernandes, P. C. Vieira, J. E. Theodoro and J. A. Ellena, *Phytochemistry*, 2012, **76**, 52–59.
- 137 T. Inoue, Y. Nagai, A. Mitooka, R. Ujike, O. Muraoka, T. Yamada and R. Tanaka, *Tetrahedron Lett.*, 2012, **53**, 6685–6688.
- 138 J. Liao, T. Xu, Y.-H. Liu and S.-Z. Wang, *Nat. Prod. Res.*, 2012, **26**, 756–761.

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

ppen Access Article. Published on 24 October 2014. Downloaded on 7/19/2025 1:07:28 AM.

View Article Online

- 139 J.-Y. Cai, Y. Zhang, S.-H. Luo, D.-Z. Chen, G.-H. Tang, C.-M. Yuan, Y.-T. Di, S.-H. Li, X.-J. Hao and H.-P. He, Org. Lett., 2012, 14, 2524-2527.
- 140 H.-B. Liu, H. Zhang, P. Li, Z.-B. Gao and J.-M. Yue, Org. Lett., 2012, 14, 4438-4441.
- 141 J.-Q. Liu, C.-F. Wang, Y. Li, J.-C. Chen, L. Zhou and M.-H. Qiu, Phytochemistry, 2012, 76, 141-149.
- 142 S.-Y. Jiang, J.-Q. Liu, J.-J. Xia, Y.-X. Yan and M.-H. Qiu, Helv. Chim. Acta, 2012, 95, 301-307.
- 143 Y. Zhang, C.-P. Tang, C.-Q. Ke, X.-Q. Li, H. Xie and Y. Ye, Phytochemistry, 2012, 73, 106-113.
- 144 Y. Tanaka, A. Sakamoto, T. Inoue, T. Yamada, T. Kikuchi, T. Kajimoto, O. Muraoka, A. Sato, Y. Wataya, H.-S. Kim and R. Tanaka, Tetrahedron, 2012, 68, 3669-3677.
- 145 C. P. Wong, M. Shimada, A. E. Nugroho, Y. Hirasawa, T. Kaneda, A. H. A. Hadi, S. Osamu and H. Morita, J. Nat. Med., 2012, 66, 566-570.
- 146 L. Jiang, Chem. Nat. Compd., 2012, 48, 1013-1016.
- 147 J.-S. Wang, Y. Zhang, X.-B. Wang and L.-Y. Kong, Tetrahedron, 2012, 68, 3963-3971.
- 148 L. Tong, Y. Zhang, H. He and X. Hao, Chin. J. Chem., 2012, 30, 1261-1264.
- 149 Y.-H. Ge, J.-X. Zhang, S.-Z. Mu, Y. Chen, F.-M. Yang, Y. Lu and X.-J. Hao, Tetrahedron, 2012, 68, 566-572.
- 150 Y.-h. Ge, K.-x. Liu, J.-x. Zhang, S.-z. Mu and X.-j. Hao, J. Agric. Food Chem., 2012, 60, 4289-4295.
- 151 T. Yang, Y.-B. Zeng, Z.-K. Guo, W.-J. Zuo, S.-S. Ma, S.-S. Li, W.-L. Mei and H.-F. Dai, J. Asian Nat. Prod. Res., 2012, 14, 581-585.
- 152 H.-B. Liu, H. Zhang, P. Li, Y. Wu, Z.-B. Gao and J.-M. Yue, Org. Biomol. Chem., 2012, 10, 1448-1458.
- 153 J. Luo, Y. Li, J.-S. Wang, J. Lu, X.-B. Wang, J.-G. Luo and L.-Y. Kong, Chem. Pharm. Bull., 2012, 60, 195-204.
- 154 Y. Li, J. Luo, Q. Wang and L.-Y. Kong, Heterocycles, 2012, 85, 3035-3041.
- 155 J. Luo, Y. Li, J.-S. Wang, J. Lu and L.-Y. Kong, Phytochem. Lett., 2012, 5, 249-252.
- 156 X.-L. Chen, H.-L. Liu and Y.-W. Guo, Planta Med., 2012, 78, 286-290.
- 157 W. Yang, L. Kong, Y. Zhang, G. Tang, F. Zhu, S. Li, L. Guo, Y. Cheng, X. Hao and H. He, Planta Med., 2012, 78, 1676-1682.
- 158 I. A. Najmuldeen, A. H. A. Hadi, K. Mohamad, K. Awang, K. A. Ketuly, M. R. Mukhtar, H. Taha, N. Nordin, M. Litaudon, F. Gueritte, A. E. Nugroho and H. Morita, Heterocycles, 2012, 84, 1265-1270.
- 159 S.-L. Chong, K. Awang, M. T. Martin, M. R. Mokhtar, G. Chan, M. Litaudon, F. Gueritte and K. Mohamad, Tetrahedron Lett., 2012, 53, 5355-5359.
- 160 C.-M. Yuan, Y. Zhang, G.-H. Tang, S.-L. Li, Y.-T. Di, L. Hou, J.-Y. Cai, H.-M. Hua, H.-P. He and X.-J. Hao, Chem.-Eur. J., 2012, 7, 2024-2027.
- 161 F. Dal Piaz, N. Malafronte, A. Romano, D. Gallotta, M. A. Belisario, G. Bifulco, M. J. Gualtieri, R. Sanogo, N. De Tommasi and C. Pisano, Phytochemistry, 2012, 75, 78-89.

- 162 P. A. Yadav, G. Suresh, K. R. Prasad, M. S. A. Rao and K. S. Babu, Tetrahedron Lett., 2012, 53, 773-777.
- 163 J.-Q. Liu, C.-F. Wang, J.-C. Chen and M.-H. Qiu, Nat. Prod. Res., 2012, 26, 1887-1891.
- 164 W. Yang, L.-M. Kong, S.-F. Li, Y. Li, Y. Zhang, H.-P. He and X.-J. Hao, Nat. Prod. Bioprospect., 2012, 2, 145-149.
- 165 J. Li, M.-Y. Li, G. Feng, J. Zhang, M. Karonen, J. Sinkkonen, T. Satyanandamurty and J. Wu, J. Nat. Prod., 2012, 75, 1277-1283.
- 166 M. Chen, R. Chen, S. Wang, W. Tan, Y. Hu, X. Peng and Y. Wang, Int. J. Nanomed., 2012, 8, 85-92.
- 167 J.-H. Liu, N. Zhao, G.-J. Zhang, S.-S. Yu, L.-J. Wu, J. Qu, S.-G. Ma, X.-G. Chen, T.-Q. Zhang, J. Bai, H. Chen, Z.-F. Fang, F. Zhao and W.-B. Tang, J. Nat. Prod., 2012, 75, 683-688.
- 168 N. Cachet, F. Ho-A-Kwie, M. Rivaud, E. Houel, E. Deharo, G. Bourdy and V. Jullian, *Phytochem. Lett.*, 2012, 5, 162–164.
- 169 S. M. M. Donkwe, E. N. Happi, J. D. Wansi, B. N. Lenta, K. P. Devkota, B. Neumann, H.-G. Stammler and N. Sewald, Planta Med., 2012, 78, 1949-1956.
- 170 M. G. Moghaddam and F. B. H. Ahmad, Asian J. Chem., 2012, 24, 4843-4846.
- 171 L. Zhang and Y.-c. Zhang, Guoji Zhongliuxue Zazhi, 2012, 39, 113-116.
- 172 D.-Y. Lee, K.-H. Seo, D.-S. Lee, Y.-C. Kim, I.-S. Chung, G.-W. Kim, D.-S. Cheoi and N.-I. Baek, J. Nat. Prod., 2012, 75, 1138-1144.
- 173 T. Warashina, K. Umehara and T. Miyase, Chem. Pharm. Bull., 2012, 60, 205-212.
- 174 Y.-C. Lai, C.-K. Chen, S.-F. Tsai and S.-S. Lee, Phytochemistry, 2012, 74, 206-211.
- 175 C.-x. Yang, Z.-k. Zhang, N. Liu, B. Wei and X.-l. Su, Zhongcaoyao, 2012, 43, 1471-1474.
- 176 N. Riaz, M. A. Naveed, M. Saleem, B. Jabeen, M. Ashraf, S. A. Ejaz, A. Jabbar and I. Ahmed, J. Asian Nat. Prod. Res., 2012, 14, 1149-1155.
- 177 H. Zhang, H.-H. Xu, Z.-J. Song, L.-Y. Chen and H.-J. Wen, Fitoterapia, 2012, 83, 1081-1086.
- 178 L.-F. Peng, W.-J. Xia, L. He and T. Cui, Chin. J. Nat. Med., 2012, 10, 81-83.
- 179 W. Zuo, Q. Wang, W. Li, Y. Sha, X. Li and J. Wang, Magn. Reson. Chem., 2012, 50, 325-328.
- 180 K. Kakuta, T. Koike, K. Kinoshita, S. Ito, K. Koyama and K. Takahashi, Heterocycles, 2012, 85, 1377-1392.
- 181 J. Yu, S. Liu and L. Xuan, Nat. Prod. Res., 2012, 26, 630-636.
- 182 R. Kumari, M. Ali and V. Aeri, J. Asian Nat. Prod. Res., 2012, 14, 7-13.
- 183 B. S. Siddiqui, N. Khatoon, S. Begum, A. D. Farooq, K. Qamar, H. A. Bhatti and S. K. Ali, Phytochemistry, 2012, 77, 238-244.
- 184 H. M. P. Poumale, K. P. Awoussong, R. Randrianasolo, C. C. F. Simo, B. T. Ngadjui and Y. Shiono, Nat. Prod. Res., 2012, 26, 749-755.
- 185 J. Hussain, N. Ur Rehman, H. Hussain, A. Al-Harrasi, L. Ali, T. S. Rizvi, M. Ahmad and Mehjabeen, Fitoterapia, 2012, 83, 593-598.

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

ppen Access Article. Published on 24 October 2014. Downloaded on 7/19/2025 1:07:28 AM.

- 186 X. Zhang, J. Chen and K. Gao, *Biochem. Syst. Ecol.*, 2012, 45, 7–11.
- 187 L. Xiong, M. Zhu, C. Zhu, S. Lin, Y. Yang and J. Shi, *J. Nat. Prod.*, 2012, **75**, 1160–1166.
- 188 J. A. Kim, J. H. Son, S. Y. Yang, S. B. Song, G. Y. Song and Y. H. Kim, Arch. Pharmacal Res., 2012, 35, 647–651.
- 189 M. Zaffer Ahmad, M. Ali and S. R. Mir, *J. Pharm. Res.*, 2012, 5, 548–550.
- 190 J. Pollier and A. Goossens, Phytochemistry, 2012, 77, 10-15.
- 191 P. Manna and P. C. Sil, Free Radical Res., 2012, 46, 815-830.
- 192 M. Efdi, M. Ninomiya, E. Suryani, K. Tanaka, S. Ibrahim, K. Watanabe and M. Koketsu, *Bioorg. Med. Chem. Lett.*, 2012, 22, 4242–4245.
- 193 J. J. Omolo, V. Maharaj, D. Naidoo, T. Klimkait, H. M. Malebo, S. Mtullu, H. V. M. Lyaruu and C. B. de Koning, *J. Nat. Prod.*, 2012, 75, 1712–1716.
- 194 C.-J. Ji, G.-Z. Zeng, J. Han, W.-J. He, Y.-M. Zhang and N.-H. Tan, *Bioorg. Med. Chem. Lett.*, 2012, 22, 6377–6380.
- 195 S. Sutthivaiyakit, C. Seeka, T. Kritwinyu,
 S. Pisutchareonpong and N. Chimnoi, *Tetrahedron Lett.*, 2012, 53, 1713–1716.
- 196 Q. Zhan, F. Zhang, L. Sun, Z. Wu and W. Chen, *Molecules*, 2012, **17**, 14899–14907.
- 197 L.-J. Zhang, J.-J. Cheng, C.-C. Liao, H.-L. Cheng, H.-T. Huang, L.-M. Y. Kuo and Y.-H. Kuo, *Planta Med.*, 2012, **78**, 1584–1590.
- 198 I. Khlif, K. Hamden, M. Damak and N. Allouche, *Chem. Nat. Compd.*, 2012, **48**, 799–802.
- 199 M. A. Tantry, J. A. Dar, M. A. Khuroo and A. S. Shawl, *Phytochem. Lett.*, 2012, **5**, 253–257.
- 200 H.-C. Ren, R.-D. Qin, Q. Wang, W. Cheng, Q.-Y. Zhang and H. Liang, J. Asian Nat. Prod. Res., 2012, 14, 1032–1038.
- 201 G. Schnell, P. Schaeffer, E. Motsch and P. Adam, *Org. Biomol. Chem.*, 2012, **10**, 8276–8282.
- 202 Z. Habibi, Z. Cheraghi, S. Ghasemi and M. Yousefi, *Nat. Prod. Res.*, 2012, **26**, 1910–1913.
- 203 A. A. Osorio, A. Munoz, D. Torres-Romero, L. M. Bedoya, N. R. Perestelo, I. A. Jimenez, J. Alcami and I. L. Bazzocchi, *Eur. J. Med. Chem.*, 2012, **52**, 295–303.
- 204 H. Yang, H.-J. Cho, S. H. Sim, Y. K. Chung, D.-D. Kim, S. H. Sung, J. Kim and Y. C. Kim, *Bioorg. Med. Chem. Lett.*, 2012, 22, 2079–2083.
- 205 J. Li, H. Xu, W. Tang and Z. Song, *Fitoterapia*, 2012, **83**, 383–387.
- 206 R. A. Kaskoos, M. Ali and K. J. Naquvi, *J. Pharm. Res.*, 2012, 5, 2368–2372, 2365 pp.
- 207 T. N. Vo, P. L. Nguyen, L. T. Tuong, L. M. Pratt, P. N. Vo,
 K. P. P. Nguyen and N. S. Nguyen, *Chem. Pharm. Bull.*,
 2012, 60, 1125–1133.
- 208 H.-Z. Jiang, Q.-Y. Ma, H.-J. Fan, W.-J. Liang, S.-Z. Huang, H.-F. Dai, P.-C. Wang, X.-F. Ma and Y.-X. Zhao, *J. Braz. Chem. Soc.*, 2012, 23, 889–893.
- 209 H.-C. Huang, C.-T. Chiou, P.-C. Hsiao, C.-C. Liaw, L.-J. Zhang, C.-L. Chang, I.-S. Chen, W.-C. Chen, K.-H. Lee and Y.-H. Kuo, *Molecules*, 2012, **17**, 1837–1851.
- 210 Q. Jin, H.-G. Jin, A. R. Kim and E.-R. Woo, *Helv. Chim. Acta*, 2012, **95**, 1455–1460.

- 211 T. K. L. Giang, T. T. Nguyen, T. H. A. Nguyen, V. S. Tran and H. C. Dao, *Tap Chi Hoa Hoc*, 2011, **49**, 738–742.
- 212 S. Gandomkar, M. Yousefi, Z. Habibi and M. A. As'habi, *Nat. Prod. Res.*, 2012, **26**, 648–653.
- 213 M. F. de Araujo, R. Braz-Filho, M. G. de Carvalho and I. J. C. Vieira, *Quim. Nova*, 2012, **35**, 2202–2204.
- 214 J.-J. Li, J. Yang, F. Lu, Y.-T. Qi, Y.-Q. Liu, Y. Sun and Q. Wang, *Chin. J. Nat. Med.*, 2012, **10**, 279–283.
- 215 S.-A. Tang, L.-L. Ding, H.-Y. Zhai, N. Qin and H.-Q. Duan, J. Asian Nat. Prod. Res., 2012, 14, 838–843.
- 216 B. Kuang, J. Han, G.-Z. Zeng, X.-Q. Chen, W.-J. He and N.-H. Tan, *Nat. Prod. Bioprospect.*, 2012, **2**, 166–169.
- 217 C.-C. Zhao, J.-H. Shao and J.-D. Fan, *Chem. Nat. Compd.*, 2012, **48**, 803–805.
- 218 C. Y. Ragasa, D. L. Espineli and C.-C. Shen, *Nat. Prod. Res.*, 2012, **26**, 1869–1875.
- 219 J. Sun, X. Wang, H. Zhang and J. Yang, *Chem. Nat. Compd.*, 2012, **48**, 416–418.
- 220 Y. Liu, M. Kubo and Y. Fukuyama, J. Nat. Prod., 2012, 75, 1353–1358.
- 221 A. A. Gohar, G. T. Maatooq, E. M. Mrawan, A. A. Zaki and Y. Takaya, *Nat. Prod. Res.*, 2012, 26, 1328–1333.
- A. Alabdul Magid, N. Lalun, C. Long, N. Borie, H. Bobichon,
 C. Moretti and C. Lavaud, *Phytochemistry*, 2012, 77, 268–274.
- 223 L. Yu, X. Tang, L. Chen, M. Wang, J. Jian, S. Cao, X. Wang, N. Kang and F. Qiu, *Fitoterapia*, 2012, 83, 1636–1642.
- 224 S. Iwanaga, T. Warashina and T. Miyase, *Chem. Pharm. Bull.*, 2012, **60**, 1264–1274.
- 225 S. Nakamura, T. Moriura, S. Park, K. Fujimoto, T. Matsumoto, T. Ohta, H. Matsuda and M. Yoshikawa, *J. Nat. Prod.*, 2012, 75, 1425–1430.
- 226 W. Yuan, P. Wang, G. Deng and S. Li, *Phytochemistry*, 2012, 75, 67–77.
- 227 S. Bencharif-Betina, T. Miyamoto, C. Tanaka, Z. Kabouche, A.-C. Mitaine-Offer and M. A. Lacaille-Dubois, *Helv. Chim. Acta*, 2012, **95**, 1573–1580.
- 228 C. T. A. Minh, M. K. Nguyen, P. T. Thuong, I. H. Hwang, D. W. Kim and M. K. Na, *Biochem. Syst. Ecol.*, 2012, 44, 270–274.
- 229 D. Ji, Y. Wu, B. Zhang, C.-F. Zhang and Z.-L. Yang, *Fitoterapia*, 2012, **83**, 843–848.
- 230 M. Scognamiglio, B. D'Abrosca, V. Fiumano, A. Chambery,
 V. Severino, N. Tsafantakis, S. Pacifico, A. Esposito and
 A. Fiorentino, *Phytochemistry*, 2012, 84, 125–134.
- 231 Q.-z. Wang, X.-f. Liu, Y. Shan, F.-q. Guan, Y. Chen, X.-y. Wang, M. Wang and X. Feng, *Fitoterapia*, 2012, 83, 742–749.
- 232 H.-Z. Li, L.-Z. Fu, H.-M. Li, R.-T. Li and X.-L. Deng, *Phytochem. Lett.*, 2012, **5**, 572–575.
- 233 T. K. Tabopda, A.-C. Mitaine-Offer, T. Miyamoto, C. Tanaka, J.-F. Mirjolet, O. Duchamp, B. T. Ngadjui and M.-A. Lacaille-Dubois, *Phytochemistry*, 2012, 73, 142–147.
- 234 L. Yu, X. Wang, X. Wei, M. Wang, L. Chen, S. Cao, N. Kang and F. Qiu, *Bioorg. Med. Chem. Lett.*, 2012, 22, 5232–5238.
- 235 X.-X. Weng, J. Zhang, W. Gao, L. Cheng, Y. Shao and D.-Y. Kong, *Helv. Chim. Acta*, 2012, **95**, 255–260.

- 236 H. Matsuda, M. Hamao, S. Nakamura, H. Kon'i, M. Murata and M. Yoshikawa, *Chem. Pharm. Bull.*, 2012, **60**, 674–680.
- 237 K. Kakuta, M. Baba, S. Ito, K. Kinoshita, K. Koyama and K. Takahashi, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 4793–4800.
- 238 N. B. Sankahya and S. Kirmizigul, *Planta Med.*, 2012, 78, 828–833.
- 239 Y. Iwamoto, S. Sugimoto, L. Harinantenaina, K. Matsunami and H. Otsuka, *J. Nat. Med.*, 2012, **66**, 321–328.
- 240 J. Wang, J. Lu, C. Lv, T. Xu and L. Jia, *Fitoterapia*, 2012, **83**, 1396–1401.
- 241 N. Saba, R. Khatoon, Z. Ali and V. U. Ahmad, *J. Chem. Soc. Pak.*, 2012, **34**, 448–450.
- 242 H. Altunkeyik, D. Gulcemal, M. Masullo, O. Alankus-Caliskan, S. Piacente and T. Karayildirim, *Phytochemistry*, 2012, **73**, 127–133.
- 243 L. Li, Y.-X. He, M.-L. Gou and C. Dai, *J. Asian Nat. Prod. Res.*, 2012, 14, 1169–1174.
- 244 Z.-x. Zhao, R.-x. Zeng, J. Jin, C.-z. Lin, T.-q. Xiong, J.-y. Cai and C.-c. Zhu, *Zhongcaoyao*, 2012, 43, 1267–1269.
- 245 T. H. Quang, T. T. N. Nguyen, C. V. Minh, P. V. Kiem, H.-J. Boo, J.-W. Hyun, H.-K. Kang and Y. H. Kim, *Phytochem. Lett.*, 2012, 5, 177–182.
- 246 W. Li, J. Cao, Y. Tang, L. Zhang, Q. Xie, H. Shen and Y. Zhao, *Fitoterapia*, 2012, **83**, 147–152.
- 247 Y. Chen, Y. Shan, Y. Y. Zhao, Q. Z. Wang, M. Wang, X. Feng and J. Y. Liang, *Chin. Chem. Lett.*, 2012, 23, 325–328.
- 248 D.-L. Wu, Y.-K. Wang, J.-S. Liu, X.-C. Wang and W. Zhang, *J. Asian Nat. Prod. Res.*, 2012, **14**, 342–347.
- 249 X.-F. Zhang, Y.-Y. Han, G.-H. Bao, T.-J. Ling, L. Zhang, L.-P. Gao and T. Xia, *Molecules*, 2012, **17**, 11721–11728.
- 250 V. Lanzotti, P. Termolino, M. Dolci and P. Curir, *Bioorg. Med. Chem.*, 2012, **20**, 3280–3286.
- 251 I. K. Lee, S. U. Choi and K. R. Lee, *Chem. Pharm. Bull.*, 2012, **60**, 1011–1018.
- 252 C. Li, J. Fu, J. Yang, D. Zhang, Y. Yuan and N. Chen, *Fitoterapia*, 2012, **83**, 1184–1190.
- 253 F. Li, C.-R. Sun, B. Chen, L.-S. Ding and M.-K. Wang, *Phytochem. Lett.*, 2012, 5, 258–261.
- 254 K. Fujimoto, S. Nakamura, S. Nakashima, T. Matsumoto, K. Uno, T. Ohta, T. Miura, H. Matsuda and M. Yoshikawa, *Chem. Pharm. Bull.*, 2012, **60**, 1188–1194.
- 255 S. Nakamura, K. Fujimoto, S. Nakashima, T. Matsumoto, T. Miura, K. Uno, H. Matsuda and M. Yoshikawa, *Chem. Pharm. Bull.*, 2012, 60, 752–758.
- 256 Z. Z. Ibraheim, W. M. Abdel-Mageed and M. Jaspars, *Biochem. Syst. Ecol.*, 2012, **40**, 86–90.
- 257 J. Linnek, A.-C. Mitaine-Offer, T. Paululat and M.-A. Lacaille-Dubois, *Magn. Reson. Chem.*, 2012, 50, 798– 802.
- 258 W. Xu, J. Fang, Z. Zhu, J. Wu and Y. Li, *Nat. Prod. Res.*, 2012, **26**, 2002–2007.
- 259 M. Myose, T. Warashina and T. Miyase, *Chem. Pharm. Bull.*, 2012, **60**, 612–623.
- 260 Z.-H. Xiao, F.-Z. Wang, A.-J. Sun, C.-R. Li, C.-G. Huang and S. Zhang, *Molecules*, 2012, 17, 295–302.
- 261 C. Sayagh, C. Long, C. Moretti and C. Lavaud, *Phytochem. Lett.*, 2012, 5, 188–193.

- 262 Z.-B. Wang, H. Jiang, Y.-G. Xia, B.-Y. Yang and H.-X. Kuang, *Molecules*, 2012, **17**, 6269–6276.
- 263 E. A. Khatuntseva, V. M. Men'shov, A. S. Shashkov,
 Y. E. Tsvetkov, R. N. Stepanenko, R. Y. Vlasenko,
 E. E. Shults, G. A. Tolstikov, T. G. Tolstikova, D. S. Baev,
 V. A. Kaledin, N. A. Popova, V. P. Nikolin, P. P. Laktionov,
 A. V. Cherepanova, T. V. Kulakovskaya,
 E. V. Kulakovskaya and N. E. Nifantiev, *Beilstein J. Org. Chem.*, 2012, 8, 763–775, no. 787.
- 264 A. Hamed, A. Perrone, U. Mahalel, W. Oleszek, A. Stochmal and S. Piacente, *Phytochem. Lett.*, 2012, 5, 782–787.
- 265 Y. Zhang, X. Huang, L. Wang, Y. Wang, Y. Wang and W. Ye, *Chin. J. Chem.*, 2012, **30**, 1249–1254.
- 266 Y. Zhang, Z. Ma, C. Hu, L. Wang, L. Li and S. Song, *Fitoterapia*, 2012, **83**, 806–811.
- 267 L. Bi, X. Tian, F. Dou, L. Hong, H. Tang and S. Wang, *Fitoterapia*, 2012, **83**, 234–240.
- 268 L.-H. Mu, N.-Y. Wei and P. Liu, *Planta Med.*, 2012, **78**, 617–621.
- 269 C. Zhu, L. Gao, Z. Zhao and C. Lin, *Yaoxue Xuebao*, 2012, 47, 77–83.
- 270 P. T. Nedialkov, Z. Kokanova-Nedialkova, D. Bucherl,
 G. Momekov, J. Heilmann and S. Nikolov, *Nat. Prod. Commun.*, 2012, 7, 1419–1422.
- 271 W. Hai, H. Cheng, M. Zhao, Y. Wang, L. Hong, H. Tang and X. Tian, *Fitoterapia*, 2012, **83**, 759–764.
- 272 M. Zhao, H.-F. Tang, F. Qiu, X.-R. Tian, Y. Ding, X.-Y. Wang and X.-M. Zhou, *Biochem. Syst. Ecol.*, 2012, **40**, 49–52.
- 273 P. Alam, M. Ali and V. Ari, *J. Nat. Prod. Plant Resour.*, 2012, 2, 272–280.
- 274 A. Vassallo, M. Pesca, L. Ambrosio, N. Malafronte,
 N. D. Melle, F. Dal Piaz and L. Severino, *Nat. Prod. Commun.*, 2012, 7, 1427–1430.
- 275 T. S. El-Alfy, S. M. Ezzat and A. A. Sleem, *Nat. Prod. Res.*, 2012, **26**, 619–629.
- 276 S. Badal, Int. J. Chem. Sci., 2012, 10, 1271-1276.
- 277 M.-Q. Zhang, Y. Liu, S.-X. Xie, T.-H. Xu, T.-H. Liu, Y.-J. Xu and D.-M. Xu, *J. Asian Nat. Prod. Res.*, 2012, **14**, 1186–1190.
- 278 W. Qi, D. Yuan, L.-M. Yang, K.-H. Xie, T.-Z. Cai, R. Yang and H.-Z. Fu, *Nat. Prod. Res.*, 2012, **26**, 1436–1441.
- 279 M. Yotova, I. Krasteva, K. Jenett-Siems, P. Zdraveva and S. Nikolov, *Phytochem. Lett.*, 2012, 5, 752–755.
- 280 H. Yao, J. Duan, J. Wang and Y. Li, *Biochem. Syst. Ecol.*, 2012, **42**, 14–17.
- 281 L. O. A. Manguro, P. Lemmen, P. Hao and K.-C. Wong, J. Asian Nat. Prod. Res., 2012, 14, 987–1001.
- 282 L. Gao, L. Zhang, L.-M. Wang, J.-Y. Liu, P.-L. Cai and S.-L. Yang, *J. Asian Nat. Prod. Res.*, 2012, 14, 333–341.
- 283 F. V. Mohammad, M. Noorwala, V. U. Ahmad, A. Zahoor and N. H. J. Lajis, *Nat. Prod. Commun.*, 2012, 7, 1423–1426.
- 284 M. Yin, X. Wang, M. Wang, Y. Chen, Y. Dong, Y. Zhao and X. Feng, *Chem. Nat. Compd.*, 2012, 48, 258–261.
- 285 Y. A. Kim, C.-S. Kong, J. I. Lee, H. Kim, H. Y. Park, H.-S. Lee, C. Lee and Y. Seo, *Bioorg. Med. Chem. Lett.*, 2012, 22, 4318– 4322.
- 286 A. d. P. Barbosa, B. Pereira da Silva and J. P. Parente, *Phytochem. Lett.*, 2012, **5**, 626–631.

- 287 A. P. Oliveira, M. Raith, R. M. Kuster, L. M. Rocha, M. Hamburger and O. Potterat, *Planta Med.*, 2012, 78, 703-710.
- 288 L. Harinantenaina, P. J. Brodie, M. W. Callmander, L. J. Razafitsalama, V. E. Rasamison, E. Rakotobe and D. G. I. Kingston, *Nat. Prod. Commun.*, 2012, 7, 705–708.
- 289 X. Li, Y.-F. Wang, Q.-W. Shi and F. Sauriol, *Helv. Chim. Acta*, 2012, **95**, 1395–1400.
- 290 H. Cui, H. Xiao, X.-K. Ran, Y.-Y. Li, D.-Q. Dou and T.-G. Kang, J. Asian Nat. Prod. Res., 2012, 14, 216–223.
- 291 A. V. B. Djoumessi, L. P. Sandjo, J. C. Liermann, D. Schollmeyer, V. Kuete, V. Rincheval, A. M. Berhanu, S. O. Yeboah, P. Wafo, B. T. Ngadjui and T. Opatz, *Tetrahedron*, 2012, 68, 4621–4627.
- 292 C.-Q. Wang, L. Wang, C.-L. Fan, D.-M. Zhang, X.-J. Huang, R.-W. Jiang, L.-L. Bai, J.-M. Shi, Y. Wang and W.-C. Ye, *Org. Lett.*, 2012, 14, 4102–4105.
- 293 J. Hu, X. Shi, J. Chen, H. Huang and C. Zhao, *Fitoterapia*, 2012, **83**, 55–59.
- 294 M. J. Núñez, A. E. Ardiles, M. L. Martínez, D. Torres-Romero, I. A. Jiménez and I. L. Bazzocchi, *Phytochem. Lett.*, 2012, 5, 244–248.
- 295 G. F. Sousa, L. P. Duarte, A. F. C. Alcantara, G. D. F. Silva, S. A. Vieira-Filho, R. R. Silva, D. M. Oliveira and J. A. Takahashi, *Molecules*, 2012, 17, 13439–13456.
- 296 G. F. Sousa, F. L. Ferreira, L. P. Duarte, G. D. F. Silva, M. C. T. B. Messias and S. A. Vieira Filho, *J. Chem. Res.*, 2012, **36**, 203–205.
- 297 A. Patra, S. Ghosh, S. K. Bandyopadhyay, P. K. Bag, P. Bhowmik and E. Sukumar, J. Indian Chem. Soc., 2012, 89, 805–810.
- 298 A. E. Ardiles, A. Gonzalez-Rodriguez, M. J. Nunez, N. R. Perestelo, V. Pardo, I. A. Jimenez, A. M. Valverde and I. L. Bazzocchi, *Phytochemistry*, 2012, 84, 116–124.
- 299 H.-P. Ding, X.-P. Li, W. Zhang, N. Ding, P. Wang, G.-Q. Li and Y.-X. Li, *Zhongguo Yaolixue Yu Dulixue Zazhi*, 2012, 26, 570–576.
- 300 C.-J. Li, F.-G. Xie, J.-Z. Yang, Y.-M. Luo, X.-G. Chen and D.-M. Zhang, J. Asian Nat. Prod. Res., 2012, 14, 973–980.
- 301 J. Wu, Y. Zhou, L. Wang, J. Zuo and W. Zhao, *Phytochemistry*, 2012, 75, 159–168.
- 302 J. A. R. Salvador, V. M. Moreira, B. M. F. Goncalves, A. S. Leal and Y. Jing, *Nat. Prod. Rep.*, 2012, **29**, 1463–1479.
- 303 L. M. Bershtein, Vopr. Onkol., 2012, 58, 744-747.
- 304 Y.-B. Lu, M.-F. He and J.-H. Wang, *Zhongliu Yanjiu Yu Linchuang*, 2012, **24**, 565–566.
- 305 H. I. Al-Jaber, K. K. Abrouni, M. A. Al-Qudah and M. H. Abu Zarga, *J. Asian Nat. Prod. Res.*, 2012, **14**, 618–625.
- 306 C.-J. Zheng, J. Pu, H. Zhang, T. Han, K. Rahman and L.-P. Qin, *Fitoterapia*, 2012, **83**, 49–54.
- 307 C. Li, L. Li, C. Wang, J. Yang, F. Ye, J. Tian, Y. Si and D. Zhang, *Molecules*, 2012, 17, 13960–13968.
- 308 Y. Saito, Y. Takashima, Y. Okamoto, T. Komiyama, A. Ohsaki, X. Gong and M. Tori, *Chem. Lett.*, 2012, 41, 372–373.
- 309 A. H. Laghari, S. Memon, A. Nelofar and K. M. Khan, *Helv. Chim. Acta*, 2012, **95**, 1556–1560.

- 310 G. Venkateswara Rao, M. R. Sahoo, G. D. Rajesh and T. Mukhopadhyay, *J. Pharm. Res.*, 2012, 5, 1946–1948, 1943 pp.
- 311 J.-Y. Tao, S.-J. Dai, F. Zhao, J.-F. Liu, W.-S. Fang and K. Liu, *J. Asian Nat. Prod. Res.*, 2012, **14**, 97–104.
- 312 Y. H. Choi, W. Zhou, J. Oh, S. Choe, D. W. Kim, S. H. Lee and M. Na, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 6116–6119.
- 313 G. Sun, X. Zhang, X. Xu, J. Yang, M. Zhong and J. Yuan, *Molecules*, 2012, **17**, 504–510.
- 314 H. Wang, X. Wu, T. Zhou, X. Deng and D. Wang, *Zhongyaocai*, 2012, **35**, 396–399.
- 315 M. Shao, Y. Wang, X.-J. Huang, C.-L. Fan, Q.-W. Zhang, X.-Q. Zhang and W.-C. Ye, *J. Asian Nat. Prod. Res.*, 2012, 14, 348–354.
- 316 K. H. Kim, S. U. Choi and K. R. Lee, *Planta Med.*, 2012, 78, 86–89.
- 317 B.-b. Zhang, X.-l. Han, Q. Jiang, Z.-x. Liao, C. Liu and Y.-b. Qu, *Fitoterapia*, 2012, **83**, 1242–1247.
- 318 M. M. Hussain, M. A. Rashid, C. M. Hasan and A. Jabbar, *Int. J. Pharma Sci. Res.*, 2012, 3, 1826–1828.
- 319 J. Sidana, S. Singh, S. K. Arora, W. J. Foley and I. P. Singh, *Pharm. Biol.*, 2012, **50**, 823–827.
- 320 J. Triana, M. Lopez, F. J. Perez, M. Rico, A. Lopez, F. Estevez, M. T. Marrero, I. Brouard and F. Leon, *Molecules*, 2012, 17, 12895–12909.
- 321 Y.-C. Chien, C.-H. Lin, M. Y. Chiang, H.-S. Chang, C.-H. Liao, I.-S. Chen, C.-F. Peng and I.-L. Tsai, *Phytochemistry*, 2012, **80**, 50–57.
- 322 Q.-M. Xu, Y.-L. Liu, Y.-L. Feng, L.-H. Tang and S.-L. Yang, *Chem. Nat. Compd.*, 2012, **48**, 597–600.
- 323 P.-Y. Zhang, S.-H. Qin, H.-X. Zhao, F.-L. Wang, H.-J. Guo and H. Bai, *J. Asian Nat. Prod. Res.*, 2012, **14**, 607–611.
- 324 L.-P. Qu, G.-Y. Zheng, Y.-H. Su, H.-Q. Zhang, Y.-L. Yang, H.-L. Xin and C.-Q. Ling, *Int. J. Mol. Sci.*, 2012, 13, 14865– 14870.
- 325 M. Safder, R. Mehmood, B. Ali, U. R. Mughal, A. Malik and A. Jabbar, *Helv. Chim. Acta*, 2012, **95**, 144–151.
- 326 M. Zhou, M. Xu, X.-X. Ma, K. Zheng, K. Yang, C.-R. Yang, Y.-F. Wang and Y.-J. Zhang, *Planta Med.*, 2012, 78, 1702– 1705.
- 327 E. J. T. Mbosso, J. C. A. Nguedia, F. Meyer, B. N. Lenta, S. Ngouela, B. Lallemand, V. Mathieu, P. Van Antwerpen, A. L. Njunda, D. Adiogo, E. Tsamo, Y. Looze, R. Kiss and R. Wintjens, *Phytochemistry*, 2012, 83, 95–103.
- 328 W.-J. Zuo, H.-F. Dai, Y.-B. Zeng, H. Wang, H.-Q. Chen and J.-H. Wang, *J. Asian Nat. Prod. Res.*, 2012, **14**, 308–313.
- 329 Y.-Y. Che, Z. Liang, N. Li, K.-W. Zeng and P.-F. Tu, *Nat. Prod. Res.*, 2012, 26, 1991–1995.
- 330 L. Wang, Y. Cai, X.-Q. Zhang, C.-L. Fan, Q.-W. Zhang, X.-P. Lai and W.-C. Ye, *Carbohydr. Res.*, 2012, **349**, 39–43.
- 331 I. Kazmi, M. Rahman, M. Afzal, G. Gupta, S. Saleem, O. Afzal, M. A. Shaharyar, U. Nautiyal, S. Ahmed and F. Anwar, *Fitoterapia*, 2012, 83, 142–146.
- 332 J. C. Shu, J. Q. Liu, G. X. Chou and Z. T. Wang, *Chin. Chem. Lett.*, 2012, **23**, 827–830.
- 333 Z. Y. Babaamer, L. Sakhri, H. I. Al-Jaber, M. A. Al-Qudah and M. H. Abu Zarga, *J. Asian Nat. Prod. Res.*, 2012, 14, 1137–1143.

- 334 S. R. M. Ibrahim, G. A. Mohamed, L. A. Shaala, L. M. Y. Banuls, G. Van Goietsenoven, R. Kiss and D. T. A. Youssef, *Phytochem. Lett.*, 2012, 5, 490–495.
- 335 T. Wu, Q.-W. Zhang, X.-Q. Zhang, G. Liu, L. Wang, M.-M. Jiang, Y.-F. Feng and W.-C. Ye, *Nat. Prod. Res.*, 2012, 26, 1408–1412.
- 336 M. Isaka, P. Chinthanom, S. Supothina and S. Mongkolsamrit, *Phytochem. Lett.*, 2012, 5, 734–737.
- 337 M. Monnier, C. Lavaud, M. Litaudon and V. Dumontet, *Biochem. Syst. Ecol.*, 2012, **42**, 10–13.
- 338 V. Costantino, G. Della Sala, A. Mangoni, C. Perinu and R. Teta, *Eur. J. Org. Chem.*, 2012, 2012, 5171–5176.
- 339 C. Y. Ragasa, D. L. Espineli, E. H. Mandia, M.-J. Don and C.-C. Shen, *Chin. J. Nat. Med.*, 2012, **10**, 284–286.
- 340 X. Chai, Y.-F. Su, J. Zhang, S.-L. Yan, Y.-H. Gao and X.-M. Gao, *Helv. Chim. Acta*, 2012, **95**, 127–133.

- 341 C. Y. Ragasa, D. L. Espineli, E. H. Mandia, D. D. Raga, M.-J. Don and C.-C. Shen, Z. Naturforsch., B: J. Chem. Sci., 2012, 67, 426–432.
- 342 X.-H. Yuan, G.-B. Xu, W.-L. Wu, T. Yang and G.-Y. Li, *Arch. Pharmacal Res.*, 2012, **35**, 311–314.
- 343 Ö. B. Acıkara, G. S. Çitoğlu, S. Dall'Acqua, K. Šmejkal, J. Cvačka and M. Žemlička, *Nat. Prod. Res.*, 2012, 26, 1892–1897.
- 344 Z. Muhammad, S. Ahmad, R. Ullah, F. Ullah and S. Jan, *Biomed. Pharmacol. J.*, 2012, 5, 65–70.
- 345 S. Wittayalai, S. Sathalalai, S. Thorroad, P. Worawittayanon,
 S. Ruchirawat and N. Thasana, *Phytochemistry*, 2012, 76, 117–123.
- 346 J. Yan, Z.-Y. Zhou, M. Zhang, J. Wang, H.-F. Dai and J.-W. Tan, *Planta Med.*, 2012, 78, 1387–1391.
- 347 J. Li, H. Zhu, J. Ren, Z. Deng, N. J. de Voogd, P. Proksch and W. Lin, *Tetrahedron*, 2012, 68, 559–565.