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Transition metal diamine complexes with antimicrobial activity against *Staphylococcus aureus* and methicillin-resistant *S. aureus* (MRSA)[†]

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Pentaalkylcyclopentadienyl (Cp^{*R}) iridium (Ir) and cobalt (Co) 1,2-diamine complexes were synthesized. Susceptibility of *Staphylococcus aureus* and recent patient methicillin-resistant *S. aureus* (MRSA) isolates to the transition metal-diamine complexes were measured by broth microdilution and reported as the MIC and MBC. Hemolytic activities of the transition metal-complexes as well as toxicity toward Vero cells were also measured. The transition metal complex of Cp^{*R}Ir with *cis*-1,2-diaminocyclohexane, had strong antibiotic activity against *S. aureus* and MRSA (MIC = 4 $\mu\text{g mL}^{-1}$, MBC = 8 $\mu\text{g mL}^{-1}$) strains and killed 99% of *S. aureus* cells in 6 hours. Stronger antibiotic activity was associated with the presence of octyl linked to the cyclopentadienyl group and cyclohexane as the diamine backbone. Activity was greatly diminished by tri- or tetramethylation of the nitrogen of the diamine. A cyclopentadienylcobalt complex of *cis*-1,2-diaminocyclohexane also showed significant anti-microbial activity against both *S. aureus* and MRSA strains. The absence of hemolytic activity, Vero cell cytotoxicity and the significant anti-microbial activity of several members of the family of compounds reported suggest this is an area worth further development.

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Introduction

Nosocomial infections involving *Staphylococcus aureus* and methicillin-resistant *S. aureus* (MRSA) continue to present major challenges in the United States and Europe.^{1–3} There is also evidence of an alarming increase in *S. aureus* skin infections in children.^{4,5} While both *S. aureus* and MRSA infections occur more frequently amongst persons with weakened defences against infection in hospitals and healthcare facilities, MRSA infections are also of increasing concern in the community at large.⁶ Many *S. aureus* and MRSA infections are intractable due to antibiotic resistance and their tendency to be localized in high densities, such as in abscesses or biofilms.⁷ Colonization of in-dwelling catheters leads to biofilm formation and results in catheters serving as sources of continual infection in patients.⁷ One strategy to combat these infections is to develop novel and effective anti-infective agents.

There has been an extensive body of work reporting on transition metal compounds with anti-cancer activity.⁸ Sadler

and co-workers have synthesized and investigated the cytotoxic properties of a number of half-sandwich type organometallic compounds.^{9,10} Others have also reported on the cytotoxic behavior of different variants of half-sandwich compounds.^{11,12}

By comparison, the work on small inorganic/organometallic compounds as antimicrobials has been sparse.¹³ The antimicrobial properties of the Ag⁺ ion have long been known.^{14,15} More recently new compounds incorporating the silver ion with ligands such as *N*-heterocyclic carbenes have been shown to possess anti-microbial activity as well.^{16–18} In many other cases, anti-bacterial activity stems from either making a metal complex of a known organic anti-bacterial^{19,20} or making an organometallic analog of a known antibiotic by replacing an aryl group with an organometallic moiety such as ferrocene.^{21,22} Recently, an intriguing polypeptide with three organometallic groups has been shown to be a potent antimicrobial.²³ A review on small organometallic compounds as antibacterial agents appeared recently.²⁴

Previously, we have reported that compounds based on simple organometallic complexes comprised of either iridium (Ir) or rhodium (Rh) chelated by amino acids were potent *in vitro* inhibitors of members of the genus *Mycobacterium*.²⁵ Herein we extend our documentation of the strong antibiotic properties of a different family of transition metal complexes containing 1,2-diamine ligands to *S. aureus* and MRSA. Based on the encouraging initial results we believe that this work could lead to the identification of new therapeutic targets

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[†] Electronic supplementary information (ESI) available: A. Proton NMR spectra of the most active compounds reported in this paper, complexes 3-Ir, 4-Ir and 5-Ir. B. Complete table of MICs and MBCs for all compounds and all strains of *S. aureus* and MRSA tested. See DOI: 10.1039/c5md00228a

and development of a new class of urgently needed drugs to address the growing problem of difficult-to-treat *S. aureus* and MRSA infections.

Results and discussion

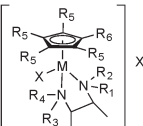
Chemistry

In a previous paper, we reported on cyclopentadienyl rhodium and iridium amino acid complexes that exhibited significant anti-mycobacterial activity.²⁵ None of the amino acid compounds displayed any significant anti-staphylococcal activity. As part of that mycobacterial study, we also made complexes of two well-known drugs used in the treatment of tuberculosis (TB): isoniazid and ethambutol.²⁶ Neither [Cp*Ir(isoniazid)Cl]Cl nor [Cp*Ir(ethambutol)Cl]Cl were active against an extensive panel of mycobacteria with MIC > 125 $\mu\text{g mL}^{-1}$. However, our practice is to screen all compounds first against a broad panel of bacteria, yeast and fungi and we noted that the ethambutol compound had moderate activity against *S. aureus* (MIC = 35 $\mu\text{g mL}^{-1}$). Noting that ethambutol is a substituted 1,2-diaminoethane, we turned our attention to complexes of other diamine ligands. That replacement of the chelating amino acids with chelating 1,2-diamines was successful in producing the anti-staphylococcal compounds described in this work.

Complexes of rhodium and iridium were synthesized according to the general procedure outlined in Scheme 1. The penta-alkyl substituted Cp rings shown in the scheme may be pentamethyl (Cp*) or tetramethylalkyl (Cp^R) the starting cyclopentadienes being synthesized as described previously.²⁷ Using this route, an extensive series of compounds were prepared, fourteen of which are listed in Table 1, 1-Ir through 14-Ir. No significant difference in activity was found between Rh and Ir compounds, so only Ir compounds are reported here.

It was also decided that the cobalt congeners should be investigated since the lighter, first-row transition elements would be less likely to have general cytotoxic behaviors. The route shown in Scheme 1 is not applicable for cobalt, so a

Table 1 General structure and numbering for complexes in this paper. The table lists the various diamine ligand substitutions (R₁, R₂, R₃ and R₄), the C₅Me₄R substitution as well as the metal



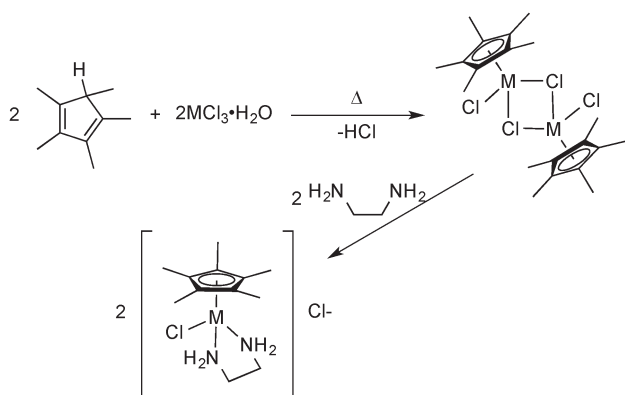
Cpd	R ₁ , R ₂ , R ₃	R ₄	R ₅	R ₆	X	Ligand
1-Ir	H	H	CH ₃	CH ₃	Cl	–CH ₂ –CH ₂ –
2-Ir	H	H	CH ₃	C ₈ H ₁₇	Cl	–CH ₂ –CH ₂ –
3-Ir	H	H	CH ₃	CH ₃	Cl	cis-Cyclohexane
4-Ir	H	H	CH ₃	C ₈ H ₁₇	Cl	cis-Cyclohexane
5-Ir	H	H	CH ₃	C ₆ H ₁₃	Cl	cis-Cyclohexane
6-Ir	H	H	CH ₃	CH ₃	Cl	trans-Cyclohexane
7-Ir	H	H	CH ₃	C ₈ H ₁₇	Cl	trans-Cyclohexane
8-Ir	H	H	CH ₃	C ₁₂ H ₂₅	Cl	cis-Cyclohexane
9-Ir	CH ₃	CH ₃	CH ₃	CH ₃	Cl	–CH ₂ –CH ₂ –
10-Ir	CH ₃	H	CH ₃	CH ₃	Cl	–CH ₂ –CH ₂ –
11-Ir	H	C ₇ H ₇	CH ₃	CH ₃	Cl	–CH ₂ –CH ₂ –
12-Ir	H	C ₁₀ H ₇	CH ₃	C ₈ H ₁₇	Cl	–CH ₂ –CH ₂ –
13-Ir	H	H	CH ₃	CH ₃	Cl	(1 <i>S</i> ,2 <i>S</i>)-1,2-Bis(4-fluorophenyl)
14-Ir	H	H	CH ₃	CH ₃	Cl	(1 <i>S</i> ,2 <i>S</i>)-1,2-Bis(4-methoxyphenyl)
15-Co	H	H	H	H	I	cis-Cyclohexane
16-Co	H	H	H	H	I	trans-Cyclohexane

new synthesis based on cyclopentadienylcobaltdicarbonyl, CpCo(CO)₂, was developed²⁸ (Scheme 2). Only the 1,2-diaminocyclohexane compounds of cobalt, 15-Co and 16-Co, are discussed herein.

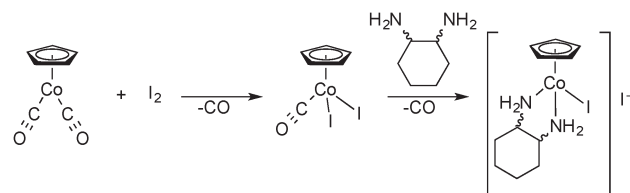
Characterization of compound structure and purity

All compounds reported in this paper were characterized by standard analytical techniques: ¹H NMR spectroscopy, ¹³C NMR spectroscopy and High Resolution Mass Spectrometry. All spectroscopies were completely consistent with the proposed structures. In addition, to assure the purity of the complexes by examining the NMR spectroscopies for the presence of any contaminants, such as solvents, compounds were submitted for C, H analyses which showed purity within 0.4% of calculated values.

All complexes described in this paper are quite water soluble, even complex 12-Ir containing a decyl chain on the Cp ligand. This is not surprising since the compounds are cationic with a chloride counterion. Complexes that had been



Scheme 1 Outline of the 2-step synthesis of iridium complexes with 1,2-diamines as the organic ligand. Ring and diamine substitution is delineated in Table 1.



Scheme 2 Outline of the synthesis of cobalt-containing complexes with 1,2-diaminocyclohexane.



dissolved in D₂O were monitored by ¹H NMR spectroscopy and show no changes in physical appearance of the solution nor any change in the NMR spectra over months of monitoring. An indicator that, in general, these diamine complexes are stable in these *in vitro* conditions is that there was no anti-mycobacterial activity of the ethambutol complex. At the highest concentrations used, even a few percent decomposition of the ethambutol complex would release a sufficient amount of free ethambutol to show anti-mycobacterial activity and none was observed.

More to the point, a standard solution of complex **5-Ir** in the growth medium was prepared and tested for activity against *S. aureus* at the time of preparation, at 6 months following preparation and at one year following preparation. The standard solution was maintained at ambient temperature during that period of time. The MIC of **5-Ir** against *S. aureus* was reproducibly 4 µg mL⁻¹ over the one year testing period showing no decomposition of the complex in the culture medium over a 12 month period of time.

Anti-staphylococcal activities and complex structures

Sixteen different complexes that were tested against *S. aureus* and various laboratory and patient strains of MRSA are listed in Table 1. A partial listing of the antimicrobial activities are listed in Table 2 while a more complete table can be found in the ESI.† While a number of iridium complexes showed activity against *S. aureus*, two of those compounds, **4-Ir** and **5-Ir** were notable for their low MICs against both *S. aureus* and MRSA isolates (Table 2). Further, the MBCs for **5-Ir** were almost the same as the MICs, demonstrating its bactericidal activity.

The data in Table 2 not only demonstrate that these complexes exhibit potent activity against both *S. aureus* and MRSA but also provide preliminary insight into the

structure–activity relationships (SARs) for these complexes. Our results thus far have suggested the following conclusions:

Substitutions to the amino groups with either four (**9-Ir**) or three (**10-Ir**) methyl groups on the nitrogen atoms leads to a loss of activity (Table 2). We speculate that the hydrogen bond contributions of the NH-functionalities on opposite sides of the diamine core are important to activity.

Several observations support the conclusion that the active species in these experiments is the organometallic complex and not the metal-free ligand nor the ligand-free metal. First, all starting materials and complex intermediates were tested against the *S. aureus* panel and showed no activity. The fact that only some of these complexes display any activity against staph strains is good evidence that the complexes are not decomposing to form the metal ion or some other indiscriminate metal fragment – if that were the case *all* of the compounds would show activity. These observations lead us to conclude that these complexes are robust and do not dissociate under conditions of the assay to yield a common iridium or rhodium species that would account for activity.

Comparison between **1-Ir** and **2-Ir** where the only difference is the substitution on the Cp* ring of an octyl group for one of the methyl groups shows an increase in activity of **2-Ir**, the octyl substituted ring complex. Similarly, comparison between **3-Ir**, **4-Ir** and **5-Ir**, all complexes with the *cis*-1,2-diaminocyclohexane ligand, shows that there is an increase in activity for **4-Ir** and **5-Ir** where one methyl group of the Cp* ligand is substituted with either a hexyl or an octyl group. It is tempting to ascribe this effect to a simple increase in hydrophobicity of the complexes, but a more specific interaction may be at play. Evidence for this is that **8-Ir** with the significantly longer dodecyl chain on the Cp* ring has little activity.

Aryl groups on the ethylenediamine backbone also resulted in activity but the range of compounds is too small to draw any significant conclusions. However, greater activity is observed for 4-fluorophenyl groups, **13-Ir**, than for 4-methoxyphenyl groups, **14-Ir**.

The most dramatic difference in activity is observed for both iridium (**3-Ir** vs. **6-Ir**) and cobalt (**15-Co** vs. **16-Co**) where the sole difference is the use of the *cis*- vs. the *trans*-isomer of 1,2-diaminocyclohexane. Both isomers complex strongly with the iridium and cobalt, but only the complexes with the *cis*-isomer show significant activity against *S. aureus*.

Time-kill measurements

Survival of *S. aureus* strain ATCC 6358 and MRSA strain 34380 when exposed to 8 µg mL⁻¹ **4-Ir** (*i.e.*, MIC/MBC) after 1, 2, 3, and 6 h exposure at 37 °C were measured in two independent experiments (Fig. 1). **4-Ir** killed both the antibiotic-susceptible *S. aureus* and the methicillin-resistant *S. aureus* strains (Fig. 2).

Table 2 MICs (MBCs) of 1,2-diamine complexes of Cp complexes of iridium and cobalt against *S. aureus* and MRSA strains in µg mL⁻¹

Compound	<i>S. aureus</i> 6358	MRSA patient	MRSA 43300	MRSA 34380
1-Ir	296 (600)	296 (250)	296 (600)	600 (600)
2-Ir	124 (124)	63 (63)	124 (124)	124 (124)
3-Ir	34 (34)	34 (34)	34 (68)	34 (34)
4-Ir	7 (16)	7 (7)	7 (8)	7 (7)
5-Ir	4 (4)	4 (4)	4 (4)	4 (4)
6-Ir	262 (524)	262 (524)	262 (524)	262 (524)
7-Ir	108 (108)	108 (108)	216 (432)	432 (432)
8-Ir	198 (400)	198 (400)	198 (250)	400 (400)
9-Ir	>500	>500	>500	>500
10-Ir	>500	>500	>500	>500
11-Ir	>500	>500	>500	>500
12-Ir	97 (97)	97 (97)	97 (97)	97 (97)
13-Ir	26 (26)	26 (26)	26 (52)	26 (26)
14-Ir	62.5 (62.5)	52 (52)	104 (104)	104 (104)
15-Co	11 (22)	22 (22)	44 (44)	44 (44)
16-Co	>500	>500	>500	>500
Vanco	0.7 (0.7)	0.7 (0.7)	0.7 (0.7)	0.7 (0.7)



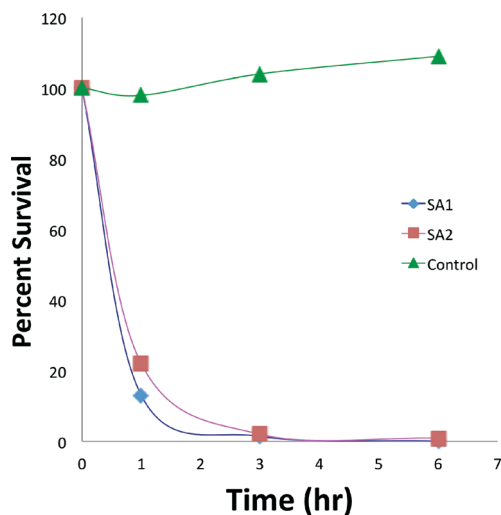


Fig. 1 Killing *S. aureus* strain ATCC 6358 with 4-Ir.

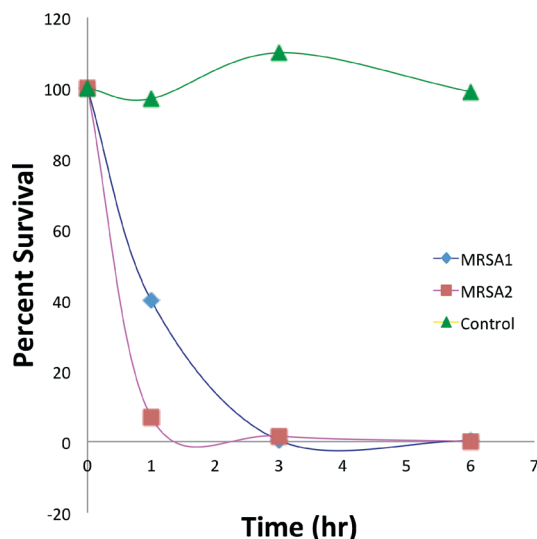


Fig. 2 Killing of MRSA strain 34380 with 4-Ir.

Hemolytic activities

The complexes with strong antibiotic activity (*i.e.*, 3-Ir, 4-Ir, and 5-Ir) were tested for hemolytic activity (<10% hemolysis of sheep red blood cells). These complexes showed no hemolytic activity at their MICs or MBCs and hemolytic activity at the 10% level was not observed until concentrations of greater than 250 $\mu\text{g mL}^{-1}$ were reached.

Mammalian toxicity

While the cobalt complexes discussed above were not tested, the iridium complexes that showed significant activity against *S. aureus* and MRSA strains showed no evidence of toxicity against Vero cell line ATCC CCL-81 (ref. 29) at the highest concentrations tested of 500 $\mu\text{g mL}^{-1}$. In addition, compound 4-Ir was tested for toxicity in mice and it was

determined that no detrimental toxicological effects were present at doses of 5 mg kg^{-1} .²⁹

Experimental

Materials

Unless otherwise stated, synthetic work was carried out in air with untreated solvents. Commercially available reagents including all diamines were obtained from Sigma-Aldrich, St. Louis, MO 63103. Deuterated solvents for NMR spectroscopy were obtained from Cambridge Isotope Laboratories.

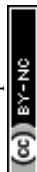
General procedure

A round bottom flask was charged with appropriate amounts of the respective $[\text{Cp}^*\text{IrCl}_2]_2$ (ref. 9, 27, 30) in dichloromethane with magnetic stirring. After 10 minutes the 1,2-diamine was added in a 2.1 molar excess drop wise to the stirring solution. After the addition of the 1,2-diamine the solution was allowed to react for an additional 30 minutes. The complexes were recrystallized from dichloromethane and ether or hexanes and collected on a fine fritted filter as yellow crystalline powders. ^1H NMR and ^{13}C NMR spectra were collected on a Varian MR-400 NMR spectrometer. High-resolution mass spectrometry data was collected on Agilent 6220 Accurate Mass TOF LC-MS. Elemental analyses were performed by Atlantic Microlabs, Norcross, GA.

Synthesis of $[(\eta^5\text{-C}_5\text{Me}_5)\text{Ir}(\text{ethylenediamine})\text{Cl}]\text{Cl}$ (1-Ir). Following the general procedure, 100 mg (0.125 mmol) of $[\text{Cp}^*\text{IrCl}_2]_2$ was combined with 15.9 mg (0.264 mmol) of ethylenediamine in dichloromethane. Yield: 91.7 mg (0.217 mmol) (86%) HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{12}\text{H}_{23}\text{N}_2[^{193}\text{Ir}]\text{Cl}$ 423.1257; found 423.1265. Anal. calcd. $\text{C}_{12}\text{H}_{23}\text{Cl}_2\text{N}_2\text{Ir}$ C: 31.44, H: 5.06; found C: 31.65, H: 5.10. ^1H NMR (400 MHz, CDCl_3) δ 6.81 (br s, 2H), 5.74 (br s, 2H), 3.27 (m, 2H), 3.07 (d, 2H), 1.89 (s, 15H, 5CpMe) ppm. ^{13}C NMR (101 MHz, CDCl_3) δ 93.2 (CpC), 92.8 (CpC), 92.2 (CpC), 54.1 (CH_2) ppm.

Synthesis of $[(\eta^5\text{-C}_4\text{Me}_4\text{C}_8\text{H}_{17})\text{Ir}(\text{ethylenediamine})\text{Cl}]\text{Cl}$ (2-Ir). Following the general procedure, 100 mg (0.112 mmol) of $[\text{Cp}^*(\text{octyl})\text{IrCl}_2]_2$ was combined with 14.2 mg (0.235 mmol) of ethylenediamine in dichloromethane. Yield: 85.1 mg (0.152 mmol) (68%) HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{19}\text{H}_{37}\text{N}_2[^{193}\text{Ir}]\text{Cl}_2$ 556.1963; found 556.1959. ^1H NMR (400 MHz, CDCl_3) δ 5.41 (br s, 2H, NH), 4.65 (br s, 2H, NH), 3.35 (d, 2H, CH_2), 3.07 (d, 2H, CH_2), 2.14 (m, 2H), 1.96 (m, 2H), 1.84 (s, 15H, 5CpMe), 1.43 (m, 2H), 1.25 (br m, 4H), 1.05–0.96 (m, 4H), 0.84 (m, 3H, CH_3) ppm.

Synthesis of $[(\eta^5\text{-C}_5\text{Me}_5)\text{Ir}(\text{cis-1,2-diaminocyclohexane})\text{Cl}]\text{Cl}$ (3-Ir). Following the general procedure, 87.1 mg (0.109 mmol) of $[\text{Cp}^*\text{IrCl}_2]_2$ was combined with 26.2 mg (0.229 mmol) of *cis*-1,2-diaminocyclohexane. Yield: 100.7 mg (0.211 mmol) (96%). HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{16}\text{H}_{29}\text{N}_2[^{193}\text{Ir}]\text{Cl}$ 477.1747; found 477.1752. Anal. calcd. $\text{C}_{16}\text{H}_{28}\text{Cl}_2\text{N}_2\text{Ir}(\text{H}_2\text{O})$ C: 36.19, H: 5.71; found C 35.83, H: 5.73. ^1H NMR (400 MHz, CDCl_3) δ 5.72 (s, 2H, NH_2), 3.27 (m, 1H, CH), 3.09 (m, 1H, CH), 2.34–2.14 (m, 2H), 2.08 (d, $J = 12.6$ Hz, 2H), 1.91 (s, 15H, CpMe), 1.65 (q, $J = 12.9$ Hz, 2H, CH_2), 1.43 (q, $J = 12.9$ Hz,



2H), 1.26 (m, 2H), 1.22 (m, 2H, CH₂) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 93.53 (CpC), 92.25 (CpC), 84.75 (CpC), 84.04 (CpC), 34.9, 32.05, 31.72, 28.14, 26.93, 21.41, 10.44, 9.74 ppm.

Synthesis of [(η⁵-C₄Me₄C₈H₁₇)Ir(*cis*-1,2-diaminocyclohexane)Cl]Cl (4-Ir). Following the general procedure, 50.0 mg (0.056 mmol) of [Cp*(^{octyl})IrCl₂]₂ was combined with 24.3 mg (0.212 mmol) of *cis*-1,2-diaminocyclohexane. Yield: 53.1 mg (0.092 mmol) (83%). HRMS/ESI⁺ (*m/z*): calcd. for C₂₃H₄₃N₂[¹⁹³Ir]Cl 575.2751; found 575.2764. Anal calc. C₂₃H₄₃Cl₂N₂Ir C: 45.23, H: 7.10; found C: 45.19, H: 7.15. ¹H NMR (400 MHz, CDCl₃) δ 3.38 (2, H), 3.10 (m, 2H), 2.24 (t, *J* = 8.0 Hz, 4H), 2.08 (m, 2H), 1.84 (s, CpMe), 1.81 (s, CpMe), 1.78 (s, CpMe), 1.76 (s, CpMe) 1.64 (t, *J* = 8.0 Hz, 2H, CH₂), 1.43 (p, *J* = 8.1 Hz, 4H), 1.32 (m, 4H, 2CH₂), 1.16 (m, 4H), 0.86 (t, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 88.2 (CpC), 86.2 (CpC), 86.1 (CpC), 51.39 (CH), 49.34 (CH), 31.88 (CH₂), 31.87 (CH₂), 30.59 (CH₂), 29.69 (CH₂), 29.50 (CH₂), 29.28 (CH₂), 29.07 (CH₂), 27.61 (CH₂), 26.44 (CH₂), 26.22 (CH₂), 25.17 (CH₂), 24.12 (CH₂), 15.25 (CH₃) ppm.

Synthesis of [(η⁵-C₄Me₄C₆H₁₃)Ir(*cis*-1,2-diaminocyclohexane)Cl]Cl (5-Ir). Following the general procedure, 50 mg (0.057 mmol) of [IrCp*(^{hexyl})Cl₂]₂ was combined with 15.9 mg (0.139 mmol) of *cis*-1,2-diaminocyclohexane. Yield: 52.0 mg (0.095 mmol) (85%). HRMS/ESI⁺ (*m/z*): calcd. for C₂₁H₃₉N₂[¹⁹³Ir]Cl 547.2509; found 547.2541. Anal calc. C: 43.29, H: 6.75; found C: 43.078, H: 6.74. ¹H NMR (400 MHz, CDCl₃) δ 6.51 (s, 1H, NH), 5.71 (s, 1H, NH), 4.03 (m, 1H, CH), 3.31 (m, 1H, CH), 2.24 (t, *J* = 8.5, 2H), 1.89–1.81 (2s, 15H, CpMe), 1.78 (m, 4H), 1.47–1.34 (m, 4H), 1.34–1.12 (m, 2H), 0.92–0.76 (m, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 87.41 (CpC), 86.14 (CpC), 47.6 (CH), 46.31 (CH), 35.55 (CH₂), 32.64 (CH₂), 29.69 (CH₂), 29.50 (CH₂), 27.61 (CH₂), 26.44 (CH₂), 26.22, 24.12 (CH₂), 12.45 (CH₃), 10.4 (CpMe) ppm.

Synthesis of [(η⁵-C₅Me₅)Ir(*trans*-1,2-diaminocyclohexane)Cl]Cl (6-Ir). Following the general procedure, 47.1 mg (0.059 mmol) of [Cp*IrCl₂]₂ was combined with 15.2 mg (0.133 mmol) of *trans*-1,2-diaminocyclohexane. Yield 48.1 mg (0.101 mmol) (81%). Calcd. for C₁₆H₂₉N₂[¹⁹³Ir]Cl 477.1765; found 477.1761. ¹H NMR (400 MHz, CDCl₃) δ 5.68 (s, 2H, NH₂) 3.27 (m, 1H, CH), 3.15 (m, 1H, CH), 2.25 (m, 2H), 2.08 (d, *J* = 11.6 Hz, 2H), 1.84 (2s, 15H, CpMe), 1.65 (q, *J* = 11.6 Hz, 2H, CH₂), 1.43 (q, *J* = 12.9 Hz, 2H) 1.26 (m, 2H), 1.22 (m, 2H, CH₂) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 94.85 (CpC), 93.25 (CpC), 84.75 (CpC), 84.04 (CpC), 34.9, 32.05, 30.82, 28.14, 27.33, 21.41, (cyclohexane) 10.18 (CpMe) ppm.

Synthesis of [(η⁵-C₄Me₄C₈H₁₇)Ir(*trans*-1,2-diaminocyclohexane)Cl]Cl (7-Ir). Following the general procedure, 75.0 mg (0.083 mmol) of [Cp*(^{octyl})IrCl₂]₂ was combined with 36.4 mg (0.318 mmol) of *trans*-1,2-diaminocyclohexane. Yield: 71.0 mg (0.119 mmol) (65%). HRMS/ESI⁺ (*m/z*): calcd for C₂₃H₄₃N₂[¹⁹³Ir]Cl 575.2755; found 575.2771. ¹H NMR (400 MHz, CDCl₃) δ 3.56 (2, NH₂), 3.36 (m, 2H), 2.75 (t, *J* = 9.5 Hz, 4H), 2.17 (m, 2H), 1.86 (s, CpMe), 1.84 (s, CpMe), 1.81 (s, CpMe), 1.76 (s, CpMe), 1.64 (t, *J* = 9.5 Hz, 2H, CH₂), 1.43 (p, *J* = 8.1 Hz, 4H), 1.37 (m,

4H, 2CH₂), 1.16 (m, 4H), 0.91 (t, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 88.7 (CpC), 88.5 (CpC), 84.2 (CpC), 51.56 (CH), 49.65 (CH), 32.88 (CH₂), 30.77 (CH₂), 30.59 (CH₂), 29.69 (CH₂), 29.50 (CH₂), 29.28 (CH₂), 29.07 (CH₂), 26.81 (CH₂), 26.44 (CH₂), 25.44 (CH₂), 25.17 (CH₂), 24.12 (CH₂), 15.18 (CH₃) ppm.

Synthesis of [(η⁵-C₄Me₄C₁₂H₂₅)Ir(*cis*-1,2-diaminocyclohexane)Cl]Cl (8-Ir). Following the general procedure, 50.0 mg (0.051 mmol) of [Cp*(^{dodecyl})IrCl₂]₂ was combined with 12.0 (0.105 mmol) mg of *cis*-1,2-diaminocyclohexane. Yield: 39.2 mg (0.062 mmol) (60%). HRMS/ESI⁺ (*m/z*): calcd. for C₂₇H₅₁N₂[¹⁹³Ir]Cl 631.3448; found 631.3475. ¹H NMR (400 MHz, CDCl₃) δ 5.69 (s, 1H, NH), 4.24 (s, 1H, NH), 3.24 (m, 1H, CH), 3.05 (m, 1H, CH), 2.39–2.19 (bm, 6H), 1.85 (s, 15H, CpMe), 1.71–1.62 (m, 4H), 1.61–1.52 (m, 4H), 1.51–1.45 (m, 4H), 1.40 (m, 6H), 1.35–1.15 (m, 10H), 0.84 (m, 3H, CH₃) ppm.

Synthesis of [(η⁵-C₅Me₅)Ir(*N,N,N',N'*-tetramethylethylenediamine)Cl]Cl (9-Ir). Following the general procedure, 50.0 mg (0.062 mmol) of [Cp*IrCl₂]₂ was combined in dichloromethane with 18.1 mg (0.155 mmol) of *N,N,N',N'*-tetramethylethylenediamine. Yield: 42.2 mg (0.088 mmol). HRMS/ESI⁺ (*m/z*): calcd. for C₁₆H₃₁N₂[¹⁹³Ir]Cl 479.1805; found 479.1804. ¹H NMR (400 MHz, CDCl₃) δ 3.62 (m, 3H, CH₃), 3.23 (s, 3H, CH₃), 2.88 (m, 3H, CH₃), 2.42 (m, 2H, CH₂) 1.89 (s, 15H, CpMe) ppm.

Synthesis of [(η⁵-C₅Me₅)Ir(*N,N,N'*-trimethylethylenediamine)Cl]Cl (10-Ir). Following the general procedure, 50.0 mg (0.062 mmol) of [Cp*IrCl₂]₂ was combined in dichloromethane with 17.8 mg (0.174 mmol) of *N,N,N'*-trimethylethylenediamine dissolved in 10 mL of methanol. Yield: 36.2 mg (0.078 mmol). HRMS/ESI⁺ (*m/z*): calcd. for C₁₅H₂₉N₂[¹⁹³Ir]Cl 465.1648; found 465.1650. ¹H NMR (400 MHz, CDCl₃) δ 4.65 (br s, 1H, NH), 3.42 (m, 3H, CH₃), 3.12 (m, 2H, CH₂), 2.65 (m, 3H, CH₃), 1.87 (s, 15H, CpMe) ppm.

Synthesis of [(η⁵-C₅Me₅)Ir(*N*-(1-benzyl)ethylenediamine)Cl][Cl] (11-Ir). Following the general procedure: 50.0 mg (0.062 mmol) of [Cp*IrCl₂]₂ was combined in dichloromethane with 19.8 mg (0.171 mmol) of *N*-(1-benzyl)ethylenediamine dissolved in 10 mL of methanol. Yield: 42.3 mg (0.082 mmol). HRMS/ESI⁺ (*m/z*): calcd. for C₁₉H₂₉N₂[¹⁹³Ir]Cl 513.1649; found 513.1648. ¹H NMR (400 MHz, CDCl₃) δ 7.39 (m, 4H, CH), 7.26 (d, 1H, CH), 5.68 (d, 2H, NH), 5.52 (s, 1H, NH), 4.57 (s, 2H), 2.1 (br s, 2H), 1.73 (s, 15H, CpMe), 1.35 (s, 2H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 125.6 (aryl), 124.9 (aryl), 88.2 (CpC), 87.1 (CpC), 55.2 (CH₂), 51.4 (CH₂), 12.5 (CH₃) ppm.

Synthesis of [(η⁵-C₅Me₄C₈H₁₇)Ir(*N*-(1-naphthyl)ethylenediamine)Cl]Cl (12-Ir). Following the general procedure, 50.0 mg (0.056 mmol) of [Cp*(^{octyl})IrCl₂]₂ was combined in dichloromethane with 32.1 mg (0.171 mmol) of *N*-(1-naphthyl)ethylenediamine dissolved in 10 mL of methanol. Yield: 40.6 mg (0.062 mmol) (55%). HRMS/ESI⁺ (*m/z*): calcd. for C₂₉H₄₃N₂[¹⁹³Ir] 647.2321; found 647.2394. Anal calc. C₂₉H₄₃Cl₂N₂Ir C: 49.71, H: 6.46; found C: 48.86, H: 5.96. ¹H NMR (400 MHz, CDCl₃) δ 7.55 (d, *J* = 6.9 Hz, 1H, ArH), 7.41 (d, *J* = 6.9 Hz, 1H, ArH), 7.14 (m, 4H, ArH), 4.88 (m, 2H), 3.72 (d, *J* = 7.5 Hz, 1H), 3.60 (d, *J* = 7.5 Hz, 1H), 3.11 (s, 2H), 2.36 (t, *J* = 11.5 Hz, 2H), 2.02 (s, 3H,



CpMe), 1.99 (s, 3H, CpMe) 1.93 (s, 3H, CpMe), 1.89 (s, 3H, CpMe), 1.79 (t, $J = 12.6$, 2H, CH_2), 1.72 (d, 1H, CH), 1.52–1.28 (m, 4H), 1.16 (m, $J = 11.5$ Hz, 3H, CH_3) ppm. ^{13}C NMR (101 MHz, CDCl_3) δ 129.9 (aryl), 127.9 (aryl), 127.8 (aryl), 86.2 (CpC), 86.1 (CpC), 60.39 (CH_2), 58.2 (CH_2), 31.88 (CH_2), 31.87 (CH_2), 30.59 (CH_2), 35.69 (CH_2), 32.50 (CH_2), 27.28 (CH_2), 26.07 (CH_2), 27.55 (CH_2), 22.30 (CH_2), 14.15 (CH_3) ppm.

Synthesis of $[(\eta^5\text{-C}_5\text{Me}_5)\text{Ir}(1\text{S},2\text{S})\text{-}1,2\text{-bis}(4\text{-fluorophenyl})\text{-ethylenediamine}]\text{Cl}$ (13-Ir). Following the general procedure, 50.0 mg (0.062 mmol) of $[\text{Cp}^*\text{IrCl}_2]_2$ was combined with 42.3 mg (0.131 mmol) (1S,2S)-1,2-bis(4-fluorophenyl)-ethylenediamine dihydrochloride in a 50/50 mix of dichloromethane and methanol. 15 mg of sodium bicarbonate was added to the solution mixture. Yield: 67.5 mg (0.110 mmol) (88%) HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{24}\text{H}_{29}\text{N}_2\text{F}_2[^{193}\text{Ir}]\text{Cl}$ 611.1622; found 611.1636. Anal. calc. C: 44.51, H: 4.58; found C: 44.58, H: 4.67. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) 7.29 (dd, $J = 8.5$, 5.5 Hz, 2H), 7.22 (dd, $J = 8.5$, 5.5 Hz, 2H), 7.10–7.02 (m, 4H), 6.81 (m, 1H), 6.24–6.12 (m, 1H, NH), 5.58 (m, 1H, NH), 4.61 (t, $J = 11.5$ Hz, 1H), 4.40 (t, $J = 11.5$, 1H), 1.72 (s, 15H, CpMe) ppm. ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) 144.8 (aryl), 135.1 (aryl), 132.5 (aryl), 129.7 (aryl), 85.8 (CpC), 26.7 (CH), 8.95 (CpMe) ppm.

Synthesis of $[(\eta^5\text{-C}_5\text{Me}_4\text{C}_8\text{H}_{17})\text{Ir}(1\text{S},2\text{S})\text{-}1,2\text{-bis}(4\text{-methoxyphenyl})\text{-ethylenediamine}]\text{Cl}$ (14-Ir). Following the general procedure, 50.0 mg (0.056 mmol) of $[\text{IrCp}^*\text{Cl}_2]_2$ was combined with 46.2 mg (0.145 mmol) of (1S,2S)-1,2-bis(4-methoxyphenyl)-ethylenediamine dihydrochloride in a 50/50 mix of dichloromethane and methanol. 17 mg of sodium bicarbonate was added to the solution mixture. Yield 48.3 mg (0.076 mmol) (67%). HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{26}\text{H}_{35}\text{N}_2\text{O}_2[^{193}\text{Ir}]\text{Cl}$ 635.2008; found: 635.2005. ^1H NMR (400 MHz, CDCl_3) δ 7.49 (d, $J = 8.5$ Hz, 1H), 7.38 (d, $J = 8.5$ Hz, 1H), 6.71 (d, $J = 8.6$ Hz, 1H), 6.62 (d, $J = 8.6$ Hz, 1H), 4.79 (m, 1H, NH), 3.67 (m, 2H), 3.55 (m, 2H), 2.01 (s, 2H), 1.88 (s, 15H, CpMe), 1.59 (s, 6H, 2CH_3) ppm. ^{13}C NMR (101 MHz, CDCl_3) δ 159.56 (aryl), 130.24 (aryl), 129.78 (aryl), 129.60 (aryl), 114.32 (aryl), 114.16 (aryl), 86.68 (CpC), 77.19 (CpC), 63.27 (CH), 55.11 (CH), 54.99 (CH), 9.52 (CpMe), 9.31 (CpMe) ppm.

Synthesis of $[(\eta^5\text{-C}_5\text{Me}_5)\text{Ir}(\text{ethambutol})\text{Cl}][\text{Cl}]$. Following the general procedure, 100 mg (0.125 mmol) of $[\text{Cp}^*\text{IrCl}_2]_2$ was combined with 72.8 mg (0.262 mmol) ethambutol dihydrochloride in a 50/50 mix of dichloromethane and methanol. 20 mg of sodium bicarbonate was added to the solution mixture. Yield: 67.7 mg (0.113 mmol) (45%). HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{20}\text{H}_{39}\text{N}_2\text{O}_2[^{193}\text{Ir}]\text{Cl}_2$ 602.2018; found 602.2022. ^1H NMR (400 MHz, D_2O) δ 4.13, (m, 4H) 3.23 (m, 2H), 2.21 (m, 2H, 2CH), 1.86 (s, 15H, CpMe), 1.21 (m, 3H) ppm.

Synthesis of $[(\eta^5\text{-C}_5\text{H}_5)\text{Co}(\text{cis-}1,2\text{-diaminocyclohexane})]\text{I}$ (15-Co). 100 mg (0.240 mmol) of $\text{CpCo}(\text{CO})\text{I}_2$ was dissolved in dichloromethane and allowed to stir for 10 minutes under nitrogen. 2.1 molar equivalents of *cis*-1,2-diaminocyclohexane was added dropwise *via* syringe to the solution and allowed

to stir for 20 minutes under nitrogen. The solution immediately evolved CO gas from the reaction. The solution turned a clear red-brown color. The solvent and excess *cis*-1,2-diaminocyclohexane were removed using a rotary evaporator leaving a dark red-brown powder. The powder was then dissolved in deionized water and filtered on a fine frit. The filtrate was collected and the water was removed to yield 76 mg (0.208 mmol) 86%. HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{11}\text{H}_{19}\text{N}_2[\text{Co}]\text{I}$ 365.0033; found 364.9924. ^1H NMR (400 MHz, methanol- d) δ 4.8 (s, H, Cp), 3.29 (m, 2H, 2CH), 1.79 (m, 4H, 2CH_2), 1.35 (m, 4H, 2CH_2), 1.6 (m, 4H), 1.35 (m, 4H, 2CH_2) ppm.

Following the same synthetic route as the previous cobalt complex, \pm -*trans*-1,2-diaminocyclohexane was used to synthesize $[(\eta^5\text{-C}_5\text{H}_5)\text{Co}(\pm\text{-trans-}1,2\text{-diaminocyclohexane})]\text{I}$ (16-Co). Yield 67 mg (0.184 mmol) 76%. HRMS/ESI⁺ (m/z): calcd. for $\text{C}_{11}\text{H}_{19}\text{N}_2[\text{Co}]\text{I}$ 365.0041; found 365.0052. ^1H NMR (400 MHz, methanol- d) δ 5.18 (s, H, Cp), 3.41 (m, 2H, 2CH), 1.82 (m, 4H, 2CH_2), 1.42 (m, 4H, 2CH_2), 1.53 (m, 4H), 1.44 (m, 4H, 2CH_2) ppm.

Staphylococcal strains and measurement of MIC and MBC

Staphylococcus aureus strains ATCC 6358 and ATCC 29213 were obtained from the American Type Culture Collection (ATCC), the unrelated methicillin-resistant *S. aureus* strain 43330 was obtained from Danville Community Hospital, and the recent patient isolates of MRSA (523000, 522870, 34864, 36361, 53016, and 34380) were obtained from Georgetown University Medical Center.³¹ MICs were measured by broth microdilution of fresh overnight cultures according to the Clinical and Laboratory Standards Institute (CLSI) guidelines with cation-adjusted Mueller–Hinton broth and an inoculum of 10^5 CFU mL^{-1} .³¹ Stocks of the compounds were dissolved in Mueller–Hinton broth. The MIC ($\mu\text{g mL}^{-1}$) was defined as the lowest concentration of compound completely inhibiting the appearance of turbidity by eye and confirmed by absorbance 540 nm. The MBC ($\mu\text{g mL}^{-1}$) was defined as the lowest concentration of compound reducing the colony count by 99.9% of the colony count in the initial, compound-free, inoculated well after 24 hr incubation. All results represent the average of three independent measurements. Both MIC and MBC experiments were also performed using brain heart infusion broth (BHIB), resulting in similar data for both MIC and MBC.³²

Time-kill measurements

A single colony of the strain was inoculated into 2 mL of Mueller–Hinton broth in a 16×125 mm screw cap tube and incubated overnight at 37 °C without aeration. One mL of that culture was inoculated into 9 mL of cation-adjusted Mueller–Hinton broth in a 125 mL flask and the culture incubated at 37 °C with aeration (60 rpm) until reaching mid-log phase (2×10^8 CFU mL^{-1}). A dilution of that culture was then added to 20 mL of Mueller–Hinton broth to reach a final density of 10^5 CFU mL^{-1} . A volume of the complex stock in Mueller–Hinton broth was added to the *S. aureus* or MRSA suspension to reach its MIC and immediately and at 1, 3,



and 6 hr, a 1 mL sample was removed and surviving CFU mL⁻¹ were measured by spreading 0.1 mL (in triplicate) of undiluted, 10⁻¹, 10⁻², and 10⁻³ fold diluted suspensions on Mueller–Hinton agar and incubating plates at 37 °C for 1 day. The results are reported as percent (%) surviving cells at each time point from two independent measurements.³²

Toxicity studies

Adapting the protocol of Babiceanu *et al.*,³³ the iridium complexes were tested for toxicity against Vero cell line ATC CCL-81. While the cobalt complexes were not tested, the iridium compounds studied showed no toxicity at the maximum level tested of 500 µg mL⁻¹.

Hemolysis measurements

Hemolysis measurements were performed as described in Maisuria *et al.*³¹ and results reported at the lowest concentration of compound yielding 10% hemolysis (IC_{10%}).

Conclusions

The transition metal complexes described here show specific activity against *S. aureus* strains and lack activity against other microorganisms (*i.e.*, *Escherichia coli*, *Mycobacterium smegmatis*, *Candida albicans*, and *Aspergillus niger*). In addition, the specificity of the complexes that show activity indicates that there are specific structure/activity relationships that must be met. This is most dramatically noted in the high activity observed for complexes of the *cis* isomer of 1,2-diaminocyclohexane (3-Ir and 15-Co) and the total lack of activity of the complexes made with the *trans* isomer of 1,2-diaminocyclohexane (6-Ir and 16-Co). We have begun work on elucidating the mechanism of action of this novel class of anti-microbial agents and hope to report on those findings in the near future. The combination of high and specific activity for certain complexes, the low cytotoxicity as tested with Vero cells and the low hemolytic activity all suggest that this transition metal platform may prove to be useful in overcoming antibiotic resistance in *S. aureus*.

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Notes and references

- R. M. Klevens, M. A. Morrison, J. Nadle, S. Petit, K. Gershman, S. Ray, L. H. Harrison, R. Lynfield, G. Dumyati and J. M. Townes, *JAMA*, 2007, **298**, 1763–1771.
- M. E. De Kraker, P. G. Davey, H. Grundmann and B. S. Group, *PLoS Med.*, 2011, **8**, e1001104.
- J. L. Hadler, S. Petit, M. Mandour and M. L. Cartter, *Emerging Infect. Dis.*, 2012, **18**, 917.
- I. Naseri, R. C. Jerris and S. E. Sobol, *Arch. Otolaryngol., Head Neck Surg.*, 2009, **135**, 14–16.
- S. Saxena, P. Thompson, R. Birger, A. Bottle, N. Spyridis, I. Wong, A. P. Johnson, R. Gilbert and M. Sharland, *Emerging Infect. Dis.*, 2010, **16**, 530–533.
- R. Kouyos, E. Klein and B. Grenfell, *PLoS Pathog.*, 2013, **9**, e1003134.
- N. Balaban, O. Cirioni, A. Giacometti, R. Ghiselli, J. B. Braunstein, C. Silvestri, F. Mocchegiani, V. Saba and G. Scalise, *Antimicrob. Agents Chemother.*, 2007, **51**, 2226–2229.
- J. C. Dabrowiak, *Metals in medicine*, John Wiley & Sons, 2009.
- Z. Liu, A. Habtemariam, A. M. Pizarro, S. A. Fletcher, A. Kisova, O. Vrana, L. Salassa, P. C. Bruijninx, G. J. Clarkson, V. Brabec and P. J. Sadler, *J. Med. Chem.*, 2011, **54**, 3011–3026.
- Z. Liu and P. J. Sadler, *Acc. Chem. Res.*, 2014, **47**, 1174–1185.
- R. Payne, P. Govender, B. Therrien, C. M. Clavel, P. J. Dyson and G. S. Smith, *J. Organomet. Chem.*, 2013, **729**, 20–27.
- A. A. Nazarov, C. G. Hartinger and P. J. Dyson, *J. Organomet. Chem.*, 2014, **751**, 251–260.
- T. B. Hadda, M. Akkurt, M. F. Baba, M. Daoudi, B. Bennani, A. Kerbal and Z. H. Chohan, *J. Enzyme Inhib. Med. Chem.*, 2009, **24**, 457–463.
- P. Bragg and D. Rainnie, *Can. J. Microbiol.*, 1974, **20**, 883–889.
- R. Richards, *Microbios*, 1980, **31**, 83–91.
- A. Kascatan-Nebioglu, M. J. Panzner, C. A. Tessier, C. L. Cannon and W. J. Youngs, *Coord. Chem. Rev.*, 2007, **251**, 884–895.
- B. D. Wright, P. N. Shah, L. J. McDonald, M. L. Shaeffer, P. O. Wagers, M. J. Panzner, J. Smolen, J. Tagaev, C. A. Tessier, C. L. Cannon and W. J. Youngs, *Dalton Trans.*, 2012, **41**, 6500–6506.
- L. Oehninger, R. Rubbiani and I. Ott, *Dalton Trans.*, 2013, **42**, 3269–3284.
- J. R. Anaconda, *J. Coord. Chem.*, 2001, **54**, 355–365.
- N. E. A. El-Gamel, *J. Coord. Chem.*, 2009, **63**, 534–543.
- D. Scutaru, L. Tataru, I. Mazilu, M. Vata, T. Lixandru and C. Simionescu, *Appl. Organomet. Chem.*, 1993, **7**, 225–231.
- M. Patra, G. Gasser, M. Wenzel, K. Merz, J. E. Bandow and N. Metzler-Nolte, *Organometallics*, 2010, **29**, 4312–4319.
- M. Wenzel, M. Patra, C. H. Senges, I. Ott, J. J. Stepanek, A. Pinto, P. Prochnow, C. Vuong, S. Langklotz, N. Metzler-Nolte and J. E. Bandow, *ACS Chem. Biol.*, 2013, **8**, 1442–1450.
- M. Patra, G. Gasser and N. Metzler-Nolte, *Dalton Trans.*, 2012, **41**, 6350–6358.
- G. Karpin, J. S. Merola and J. O. Falkinham, *Antimicrob. Agents Chemother.*, 2013, **57**, 3434–3436.
- L. B. Heifets, M. Iseman and P. Lindholm-Levy, *Antimicrob. Agents Chemother.*, 1986, **30**, 927–932.
- D. M. Morris, M. McGeagh, D. De Peña and J. S. Merola, *Polyhedron*, 2014, **84**, 120–135.



- 28 U. Koelle, B. Fuss, M. Belting and E. Raabe, *Organometallics*, 1986, 5, 980–987.
- 29 M. Ehrich, personal communication.
- 30 J. S. Merola, D. Morris and N. De Weerd, *Acta Crystallogr., Sect. E: Struct. Rep. Online*, 2013, 69, m176–m176.
- 31 B. B. Maisuria, M. L. Actis, S. N. Hardrict, J. O. Falkinham Iii, M. F. Cole, R. L. Cihlar, S. M. Peters, R. V. Macri, E. W. Sugandhi, A. A. Williams, M. A. Poppe, A. R. Esker and R. D. Gandour, *Bioorg. Med. Chem.*, 2011, 19, 2918–2926.
- 32 K. Li, L. A. Schurig-Briccio, X. Feng, A. Upadhyay, V. Pujari, B. Lechartier, F. L. Fontes, H. Yang, G. Rao, W. Zhu, A. Gulati, J. H. No, G. Cintra, S. Bogue, Y. L. Liu, K. Molohon, P. Orlean, D. A. Mitchell, L. Freitas-Junior, F. Ren, H. Sun, T. Jiang, Y. Li, R. T. Guo, S. T. Cole, R. B. Gennis, D. C. Crick and E. Oldfield, *J. Med. Chem.*, 2014, 57, 3126–3139.
- 33 M. C. Babiceanu, B. A. Howard, A. C. Rumore, H. Kita and C. B. Lawrence, *Front. Aquat. Microbiol.*, 2013, 4, 196.

