Fast electrically assisted regeneration of on-chip SERS substrates†

T.-A. Meier,a E. Poehler,a F. Kemper,b O. Pabst,b H.-G. Jahnke,c E. Beckert,b A. Robitzki c and D. Belder*a

A microfluidic chip approach utilising integrated electrically connected stationary SERS targets based on inkjet-printed silver nanoparticles is presented. It enables multiple interference-free consecutive surface-enhanced Raman measurements inside chip channels by electrically assisted regeneration of the stationary SERS substrate. Thereby it circumvents common adsorption and memory effect problems associated with stationary SERS targets allowing multiple consecutive measurements in a continuous-flow system.

Introduction

Within the past ten years surface-enhanced Raman spectroscopy (SERS) has become increasingly important within the field of microfluidics due to its potential for label-free, non-destructive and sensitive on-chip detection.1 In addition, it provides structural information making it an attractive tool for usage in a wide application area. Fields of interest are chemical2–4 and biological5–7 analyses as well as detection in separation techniques8–10 and reaction monitoring.11

Traditionally, suspended nanoparticles as SERS substrates have played a key role in microfluidic SERS measurements. They can be used straightforwardly in both continuous12 and segmented-flow13–15 systems. The use of particle suspensions to promote SERS-detection in microfluidics, however, can be problematic as it can contaminate or even clog microfluidic channels13,17 and it can also interfere negatively with downstream processes. Stationary substrates such as roughed silver surfaces18 or immobilized nanoparticles19,20 provide interesting alternatives with the additional promise of being more suitable for quantitative analytics.21,22

The utilisation of stationary substrates in flow-through systems is, however, hampered by the strong adsorption of analyte molecules to the SERS substrate, causing a so-called memory effect.23 This often irreversible adsorption of analytes to the SERS-active surface causes sample carry-over and impedes the broad application of SERS as detection technique in advanced lab-on-a-chip applications such as flow synthesis24–27 or separation techniques.28 Especially in chip-based separation techniques there is a great demand for label-free detection methods.29–32 The corresponding application of SERS could thus be a break-through in chip electrophoresis, chip chromatography or flow chemistry. A prerequisite, however, is a fast desorption of molecules from such immobile SERS-active substrates present in integrated detection flow cells. This is necessary to avoid both, signal carry-over and a potential blockade of the SERS-active spots.4 To the best of our knowledge a necessary fast and straightforward regeneration of SERS surfaces immobilised in microfluidic devices has not yet been described. Literature on conventional macroscopic flow cells, however, indicates that this could be achieved by electrically assisted desorption.33–36

We present a microfluidic system using electrically assisted substrate regeneration for inline SERS detection in chip-based systems. A chip-integrated electrically regenerable SERS substrate is used giving access to sensitive interference-free Raman spectroscopic monitoring of different successively transported compounds in flow (see Fig. 1).

Experimental section

Chemicals

MicroChem SU-8 2050 polymer and developer were purchased from Micro Resist Technology (Germany) and Sylgard 184 Polydimethylsiloxane (PDMS) from Dow Corning (Germany). 2-Propanol was purchased from Roth (Germany). Crystal violet (CV), brilliant green (BG), malachite green (MG) and phosphoric acid were obtained from Sigma Aldrich (Germany). Suntronic EMD5603 silver ink was purchased from Sun Chemical (USA). Purified water was used as solvent.

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ITO/silver electrode

The bottom layer of the microfluidic chip, containing an indium-tin-oxide (ITO)/silver electrode was fabricated using a commercial microscope glass slide (Carl Roth, Germany). On top, a 5 mm broad and 500 nm thick strip of ITO was sputtered (CREAMET® 500, Creavac, Germany) approximately 2 cm parallel shifted from the short edge.

Orthogonally, a row of 100 μm broad silver spots was inkjet-printed in the middle of the ITO strip (see Fig. 1). Printing was performed using a commercial inkjet deposition system (Omnijet 100, Unijet, South Korea) equipped with printing cartridges that exhibit a nominal droplet volume of 10 pL (DMC-11610, Dimatix Fujifilm, USA). Silver structures were printed using commercial silver nanoparticle dispersion in an ethanol/ethanediol mixture with a solid content of 20 wt% and particle diameters of 30 to 50 nm. After printing, the structures were partially sintered in a convection oven and used for microfluidic chip fabrication (see ESI S1†).

Microfluidic setup

Microfluidic streams were applied using neMESYS high precision syringe pumps (Cetoni, Germany) equipped with gas-tight syringes (Hamilton, Switzerland) linked to the chip inlets with Teflon tubes (Supelco, USA). The microfluidic structure itself consists of four feeding channels with pressure-stabilising serpentine. Downstream the fluids are lead through a zigzag-shaped mixing structure before flowing over the printed silver ink dots. All channels were approximately 150 μm wide and 100 μm high. As an exception, the area containing the dots was oval-formed with a maximum width of 650 μm. A schematic sketch of the microfluidic system can be seen in Fig. 1. The first of the four channels is used for flushing with phosphate buffer solution (pH = 3, 22.89 mM), while the other three provide the model analyte solution streams from crystal violet (1 × 10⁻⁵ mol L⁻¹) brilliant green (1 × 10⁻⁴ mol L⁻¹) and malachite green (5 × 10⁻⁵ mol L⁻¹). All solutions were pumped with 5 μL min⁻¹.

Electronic circuit

The outlet tube of the microfluidic chip was equipped with an in-house developed flow-through platinum electrode.³⁷ This electrode was connected to the cathode of a power supply (HCV 35–6500, F.u.G. Elektronik, Germany). The anode was connected to the ITO/silver electrode imprinted on the bottom glass slide (Fig. 1). A potential of 100 V, resulting in a current of 44 μA, was applied short-timed for substrate cleaning purpose.

Raman measurements

For SERS measurements the microfluidic chip was placed onto an inverted IX71 epifluorescence microscope (Olympus, Japan) which was part of a modular confocal Raman system. The system was equipped with a 473 nm laser (Cobolt, Sweden) as excitation light source, with the laser beam passing a
Results and discussion

The setup described above was tested by acquiring SERS spectra from model compounds. We investigated if SERS substrates which are contaminated with adsorbed analytes can be regenerated by applying electrical potentials via an integrated ITO electrode. For that purpose crystal violet was first flushed through the chip including the silver dots. When the analyte solution approaches the detection area with the solvent and the corresponding generation of reactive and convection promoting gas at the surface supports the desorption of different analytes in flow applications such as lab-on-a-chip.

double neutral density filter wheel for intensity tuning. Excitation intensity was kept at 0.5 mW for all experiments. The laser beam was focussed by a 40-fold objective (Olympus, Japan) onto the individual silver spots to obtain SERS signals. The scattered light passes an edge filter (473 nm), which is eliminating Rayleigh and anti-Stokes scattering, and a 150 μm wide entrance slit. Raman measurements were taken with an Acton SP2750 monochromator (Princeton instruments, USA) using a 600 lines mm⁻¹ grating and a peltier-cooled EM-CCD camera (ProEM 1600, Princeton instruments, USA) as detector. Acquisition time in all experiments was 1000 ms.

deviations of the absolute signal intensities are quite high (RSE = 33%) as shown in Fig. 3. As similar results are achieved if several fresh substrates are compared (RSE = 40%, n = 5, ESI S2†) the strong deviations in absolute intensities can be attributed to the inhomogeneity of the sintered silver particle layer. This could in principle be overcome using better defined targets such as engineered nanostructures allowing for a more quantitative analysis.

In order to investigate the electrical regeneration process a bit closer we studied the influence of the applied potentials and electrical currents. For this purpose the voltages were varied between 0 and 120 V in a corresponding series of experiments. It was found that an increase in voltage correlates with an accelerated regeneration speed (see ESI S2†). We also observed a degradation of the ITO surface above 100 V could, which could be circumvented by using alternative conductive sub-layers such as sputtered platinum. As the effective applied voltages of around 100 V are far beyond any relevant electrochemical potential including that for the decomposition of the solvent water, it appears that analyte specific electrochemical processes are less relevant. Although the actual desorbing mechanism is still unclear initial indications suggest that, electrolytic decomposition of the solvent and the corresponding generation of reactive and convection promoting gas at the surface supports the desorption. This assumption is supported by the finding that an increase in current at constant voltages, using buffer solutions of different conductivity, accelerates substrate regeneration (see ESI S4†). The regeneration of the SERS substrate, or more specifically the removal of the analytes from the surface, could also be monitored via fluorescence microscopy. This was shown for crystal violet which showed strong decrease of emission if applying an electric potential as above (see ESI S5†).

The presented system reveals great potential for the detection of different analytes in flow applications such as...
separations or segmented flow. In order to study if this can, in principle, be achieved with the presented approach, three different analytes, crystal violet (1 × 10^{-5} mol L^{-1}) brilliant green (1 × 10^{-4} mol L^{-1}) and malachite green (5 × 10^{-5} mol L^{-1}), were sequentially flushed over the target. In the aftermath of every injection, 5 minutes flushing with phosphate buffer solution was performed to monitor the memory effect in the absence of an applied potential. For monitoring an indicator wavenumber at 1585 cm^{-1} of three compounds in a microflow process with the same regeneration target.

Conclusions

We introduce an approach to perform multiple SERS measurements inside a single microfluidic device. A PDMS/glass hybrid-chip was built using moulding techniques embedding contactable silver spots as stationary SERS substrates. With the aid of integrated electrodes contacting the silver electrode potentials was applied to promote analyte desorption. This led to fast and effective electrical substrate regeneration as a proof-of-concept this approach allowed to investigate three different analytes flowed in a sequential manner over the same SERS substrate avoiding common memory effects.

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References

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