Enzymatic hydrolysis of lignocellulosic polysaccharides in the presence of ionic liquids

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Biofuels and -chemicals can be produced from carbohydrates in lignocellulosic biomass. For an efficient total enzymatic hydrolysis of the plant cell wall polysaccharides, a pretreatment step is required. Ionic liquids (ILs) have recently gained considerable interest as solvents for cellulose and lignocellulosic biomass and pretreatment of lignocellulose with ILs is currently an extensively studied concept. However, the applicability of ILs in an integrated process, in which enzymatic hydrolysis is done in the same vessel as the IL pretreatment without IL removal and substrate washing between the process steps, suffers from the fact that cellulose-dissolving ILs severely inactivate the cellulases used to catalyse the polysaccharide hydrolysis. This article reviews research on cellulase activity, stability and action in hydrolysis in cellulose-dissolving ILs, and different routes to increase the cellulase performance in these reaction systems. Impressive advances have recently been made in discovering and developing cellulases and other glycosyl hydrolases with increased IL-tolerance. Different cellulase stabilisation techniques and the design of enzyme-friendly cellulose-dissolving ILs are also discussed. In light of the recent developments, the integrated enzymatic hydrolysis of polysaccharides in the presence of ILs may well prove to be a potential route for utilizing lignocellulosic biomass as feedstock in biofuel and -chemical production.

Introduction

The rapidly growing global need for energy, chemicals and materials, price and feedstock security issues, as well as the concern for global warming, directs us from the use of fossil to renewable raw material sources. Lignocellulosic biomass is

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a renewable, carbon neutral and widely available non-food raw material for the production of fuels, platform chemicals and polymeric materials. Lignocellulosic biomass can be obtained from diverse sources including forestry side-streams, such as logging and wood processing mill residues, removed biomass from forest management and land clearing operations, and agricultural sources such as crop residues (corn stovers, straw), perennial grasses and energy and woody crops. It has been estimated that 5–8% of the annual lignocellulosic production would be enough to cover the annually used fossil oil. Cellulose is a linear homopolymer consisting of anhydroglucose units linked together by $\beta$-1,4-glycosidic bonds. Cellulose and hemicelluloses form covalent bonds with each other, known as lignin-carbohydrate complexes (LCCs).

The main components of lignocellulosics are cellulose, hemicelluloses and lignin, in various ratios depending on the source of biomass. Together these polymers form a complex matrix, which is highly recalcitrant towards depolymerization. Cellulose and hemicelluloses are polysaccharides. Cellulose is a linear homopolymer consisting of anhydroglucose units linked together by $\beta$-1,4-glycosidic bonds. The anhydroglucose units are distorted 180° to each other so that the smallest repeating unit in the cellulose chain is the anhydrocellobiose unit (Fig. 1).

Fig. 1 Structure of cellulose and its intra- and interchain hydrogen bonding. The anhydrocellobiose unit is the basic structural element of cellulose. Reproduced with permission from ref. 162.

The degree of polymerization (DP) of cellulose can range from 20 (laboratory synthesized) to 10 000 (bacterial celluloses). Crystalline cellulose has a number of polymorphs, which may carry saccharide branches and other substituents such as acetyl groups and sugar acids. Typical saccharides in hemicellulose are glucose, xylose, mannose, galactose, arabino, rhamnose and glucuronic, galacturonic and 4-O-Me-glucuronic acid. Hemicellulose composition and structure greatly vary depending on its source. The DP of different hemicelluloses varies but is in general an order of magnitude smaller than in cellulose. Unlike cellulose, many hemicelluloses are soluble in alkaline aqueous solutions, or even in neutral water. Lignin is an irregular, branched polymer built up from three different phenylpropanoid monomers, 4-hydroxy-cinnamyl alcohol, coniferyl alcohol, and sinapyl alcohol, which differ from each other in the degree of methoxylation of the aromatic ring. Due to its aromatic constituents, lignin has considerable hydrophobic character, in contrast to the hydrophilic polysaccharides. Lignin and hemicelluloses form covalent bonds with each other, known as lignin-carbohydrate complexes (LCCs).

Lignocelluloses can be converted to fuel molecules or chemicals through various routes or be burned directly for energy, although burning represents the lowest added value to the raw material. Biomass gasification and pyrolysis are techniques for producing charcoal, fuels, heat, energy and chemicals from biomass. A much studied route for converting lignocellulose to liquid fuel is to hydrolyse its polysaccharides, cellulose and hemicelluloses to monosaccharides and ferment them further to ethanol or other fuel molecules. The conversion of lignocellulose to monosaccharides and then further to liquid fuel is usually achieved in a three-step process consisting of: (1) pretreatment of the lignocellulosic feedstock, (2) hydrolysis of the feedstock polysaccharides to monosaccharides, and (3) microbial fermentation of the liberated monosaccharides to the target product, e.g. ethanol or sugar acids. The pretreatment step is necessary to open up the structure of the usually highly hydrolysis-resistant lignocellulosic matrix. Without pretreatment, enzymatic hydrolysis yields are typically low (~20% of glucan in feedstock).

The hydrolysis may be carried out either by enzymatic or mineral acid hydrolysis. Enzymatic hydrolysis offers several advantages over acid hydrolysis: less formation of undesirable by-products, no need for corrosion resistant processing equipment, less acid waste and the potential for almost complete conversion. In the presence of an acid, glucose is dehydrated to 5-hydroxymethylfurfural (HMF), which effectively inhibits the subsequent microbial fermentation. Enzymatic cellulose hydrolysis is catalysed by cellulases, i.e. glycosyl hydrolases specialized in the hydrolysis of the 1,4-$\beta$-glycosidic bonds. In the total hydrolysis, cellulases are generally applied as cocktails of different cellulase activities. Traditionally, three cellulase activities have been considered: endoglucanases, which catalyse the random cleavage of the cellulose chains especially in the amorphous regions, causing rapid reduction in the cellulose DP while liberating cello-oligomers in the process; cellobiophydrolyases (exoglucanases), which catalyse the cleavage of cellobiose from the cellulose chain ends; and $\beta$-glucosidases, which catalyse the hydrolysis of the liberated cello-oligomers to glucose. In addition to glycosyl hydrolyses, it has recently...
been suggested that also oxidative enzymes, known as lytic polysaccharide mono-oxigenases (LPMOs, recently reviewed by Horn et al.\textsuperscript{16}), contribute to the total hydrolysis of cellulose. The synergistic action of cellulases in hydrolysis has been studied extensively and at least seven different modes of cellu-

lase synergy have been described.\textsuperscript{7}

Hemicellulose hydrolysis is important both for removing the hemicellulose, which shields the cellulose from hydrolysis, and for producing monosaccharides from hemicellulose for further fermentation. In addition to glucose, also other sac-
charides, such as xylose\textsuperscript{17} and galactose,\textsuperscript{18} may be used as feedstock for various fermentations. A large battery of hemi-

cellulases (e.g. xyulasanes, mannanases, esterases and α-glucuro-

nonidases), which work in synergy, is needed for efficient hydrolysis of diverse hemicelluloses.\textsuperscript{19} Cellulases and hemi-
cellulases are often closely related, both structurally and regard-

ing their catalytic mechanism. Many glycosyl hydrolases have a modular structure, meaning that the proteins contain a carbohydrate-binding module (CBM), which has been shown to significantly promote the enzymes’ ability to catalyse the hydrolysis of solid substrates.\textsuperscript{20,21}

Ionic liquids (ILs) are by definition salts with melting points <100 °C. Usually ILs consist of an organic and rather bulky cation and either an organic or inorganic anion, which in the simplest case can be a one-atom ion such as a halogen. The low melting point of these salts has mainly been attributed to the mismatched sizes of the cation and the anion and the bulkiness and often asymmetry of the cation, as well as the charges being delocalized on several atoms, leading to weak Coulombic interactions and low crystal lattice energies and preventing crystallization.\textsuperscript{22,23} ILs have often been called green solvents mainly due to their negligible vapour pressure, which mitigates any emissions of volatile organic compounds (VOCs), and they also have excellent solvent properties due to their capability to dissolve solutes of varying polarity.\textsuperscript{24,25} In 2002, Swatloski et al. reported the dissolution of cellulose in ILs (in 1-butyl-3-methylimidazolium chloride, [BMIM]Cl, and other related ILs).\textsuperscript{26} This discovery has been followed by the dissolution of both hardwood and softwood\textsuperscript{27–30} as well as annual and perennial plants, such as switchgrass\textsuperscript{31} in ILs.

In 2006, Dadi et al. reported that the dissolution and regen-

eration of microcrystalline cellulose (MCC) in [BMIM]Cl dra-

matically accelerated the subsequent enzymatic hydrolysis kinetics,\textsuperscript{12} which stimulated extensive research on ionic liquid pretreatments of pure cellulosic substrates and later also of natural and industrially more relevant lignocellulose samples. The regeneration procedure of Dadi et al. included a wash of the regenerated substrate between the regeneration and enzymatic hydrolysis steps (Fig. 2). Kamiya et al. proposed another process alternative, the so-called one-pot or \textit{in situ} hydrolysis, in which IL pretreatment and enzymatic hydrolysis steps were integrated with no separation of IL between the steps.\textsuperscript{32} It was, however, already in 2003 demonstrated by Turner et al. that cellulose-dissolving ILs ([BMIM]Cl studied) severely inactivate cellulases.\textsuperscript{33} The one-pot hydrolysis offers some benefits over the two-step regeneration procedure, as will be discussed later, but it is a necessary task to design systems in which the cellulases can remain hydrolytically active in high concentrations of IL to make the one-pot hydrolysis technically attractive.

### Factors affecting enzymatic hydrolysis of ionic liquid-pretreated biomass

#### Dissolution of cellulose and lignocellulosic biomass in ionic liquids

Cellulose is insoluble in water and conventional organic sol-

vents but can, however, be dissolved in certain “exotic” solvent systems by either derivatizing or non-derivatizing dissolution mechanisms. The Lyocell process, in which regenerated cellulose fibers are produced by dissolving cellulose in N-methyl morpholine-N-oxide (NMMO) monohydrate and regenerated by spinning the cellulose solution in a water bath, is in industrial use.\textsuperscript{34} Cellulose dissolution in the LiCl/N,N-dimethylacetamide (DMAc) and tetrabutylammonium fluoride/ dimethylsulfoxide (DMSO) systems is of interest for research applications.\textsuperscript{35}

A patent by Graenacher from 1934 is considered the first report on dissolving cellulose in an IL type solution.\textsuperscript{36} More recently, cellulose dissolution in a pure IL was reported by Swatloski et al. in 2002 who found that cellulose dissolves well in [BMIM]Cl and to some extent in [BMIM]Cl with Br\textsuperscript{−} and SCN\textsuperscript{−} anions, but not with BF\textsubscript{4}\textsuperscript{−} or PF\textsubscript{6}\textsuperscript{−} anions.\textsuperscript{26} Another important observation in this study was that the cellulose solubility in [BMIM]Cl was extremely water sensitive, with even 1% of added water leading to a complete loss of cellulose solubility. 1-Allyl-3-methylimidazolium chloride ([AMIM]Cl) was the first cellulose-dissolving IL carrying functionalized substitu-

ents on the cation (Fig. 3).\textsuperscript{37,38} In later studies, also non-

halogen ILs were observed to dissolve cellulose efficiently, including 1-ethyl-3-methylimidazolium acetate ([EMIM]AcO)\textsuperscript{27}.

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**Fig. 2** Processing schemes for IL pretreatment with subsequent enzymatic hydrolysis of plant cell wall polysaccharides through the one-pot and the regeneration with washing procedures.
and dialklyphosphonates, such as 1,3-dimethylimidazolium dimethylphosphate (DMIMDMP).39

The dissolution and dissolution mechanisms of both cellulose and the complete lignocellulosic matrix have been extensively reviewed in recent years4,40–44 and only the most central points affecting enzymatic hydrolysis will be highlighted in this review. Cellulose-dissolving ILs are typically based on cations such as imidazolium, pyridinium, pyrrolidinium, chol linium, tetrabutylammonium and alkylalkyloxyammonium. Typical anions are halogens, carboxylates, amides, imides, thiocyanates, phosphates, sulphates, sulphonates and dichloroaluminates (Fig. 4). For many years practically all research on cellulose dissolution in ILs was done with imidazolium-based ILs, but recently several new classes of ILs have been described to dissolve cellulose, including ILs based on the organic superbases tetramethylguanidine (TMG)45 and 1,5-diazabicyclo-[4.3.0]non-5-ene (DBN),46 1-hexylpyridinium chloride (HPy)Cl,47 alkylalkyloxamonium amino acids,48 and phase-separable ILs based on tetraalkylphosphonium cations, which together with DMSO dissolve cellulose, but upon addition of water form their own, recyclable phase.49

Cellulose-dissolving ILs are, in general, hydrophilic, whereas hydrophobic ILs do not dissolve cellulose or other carbohydrates.50 The basicity of the IL’s anion has been considered a key property for cellulose dissolution, as the anion is believed to break up the hydrogen bond network which keeps the cellulose together.51,52 Cellulose solubility in ionic liquid solvent systems has been correlated to e.g. Hildebrand–Hansen solubility parameters and Kamlet-Taft parameters,53,54 which have been found to be especially practical.

The role of the IL cation in cellulose dissolution appears to be somewhat unclear. The imidazolium cation has been proposed to have hydrophobic interactions with the hydrophobic face of the cellulose (Fig. 5),55,56 although other results suggest only weak interactions between the imidazolium and carbohydrates.57 Also the C-2 proton in the imidazolium has been simulated to interact as a weak hydrogen bond donor with the cellulose hydroxyl groups during dissolution58 and cation acidity has recently been suggested to be an important parameter for predicting cellulose solubility in certain cases.46

Cellulose dissolution rates are greatly affected by the IL’s viscosity. Heating naturally reduces the solution viscosity and thus increases cellulose dissolution rates.39 The high viscosity of ILs and especially polymer solutions in ILs not only renders dissolution of cellulose in IL laborious, but also hampers the general processability of IL solutions. Furthermore, the cellulose DP and origin will have an impact on the dissolution behaviour. The dissolution process is very moisture sensitive, which is challenging as moisture can be introduced from air moisture (IL hygroscopicity) and the cellulose itself generally contains some water.

Due to their complex structures, ILs are able to interact with solutes through a variety of different interactions, including dispersive, π–π, n–π, hydrogen bonding, dipolar, and ionic/charge–charge interactions.58 The unique properties of certain ILs to dissolve all the components of lignocellulosic biomass are probably due to this set of possible dissolution interactions that ILs have. [AMIM]Cl is especially rich in π electrons due to the allyl substituent, which in part is likely to explain its high dissolution ability for wood; [AMIM]Cl is able to dissolve not only the polysaccharides but also the lignin due to the extra set of π electrons interacting with the aromatic lignin.57,58 In a comparison between different ILs, [AMIM]Cl was found to be

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**Fig. 3** The most used ILs for lignocellulose pretreatment: 1-ethyl-3methylimidazolium acetate ([EMIM]AcO), 1-allyl-3-methy limidazolium chloride ([AMIM]Cl), 1-butyl-3-methylimidazolium chloride ([BMIM]Cl) and dialkylimidazolium dialkylphosphates, among which the most known is 1,3-dimethylimidazolium dimethylphosphate (DMIMDMP).39

**Fig. 4** Typical ion constituents of cellulose-dissolving ILs.

**Fig. 5** Proposed dissolution mechanisms for cellulose in imidazolium-based ionic liquids: (A) interactions between the basic anion and the cellulose hydroxyl groups break up the hydrogen bond network which keeps the cellulose together.51,52 (B) Hydrophobic interactions take place between the hydrophobic face of cellulose and the imidazolium ring. (C) The acidic proton at the C-2 position on the imidazolium ring interacts with the cellulose hydroxyl groups.
the most efficient for wood dissolution, whereas [EMIM]AcO was the most efficient for dissolving cellulose.27

Lignocellulose pretreatments with ionic liquids

Lignocellulosic feedstocks need to be pretreated prior to enzymatic hydrolysis due to their inherent recalcitrance towards degradation. The pretreatment of biomass has been calculated to be the second largest cost factor in sugar-based biofuel production, after the feedstock itself.59 The main factors contributing to the lignocellulosic recalcitrance are cellulose crystallinity, DP and surface accessibility and the presence of hemicellulose and lignin,60 which form a physical barrier around the cellulose.51,62 Furthermore, enzymes can bind non-specifically to lignin.63 The cellulose DP has a direct impact on cellbiohydrolysis action as these cellulases need free cellulose chain ends to work on, whereas the DP is not known to influence the action of endoglucanases.7,64 A pretreatment should reduce the substrate recalcitrance by modifying at least one and preferably several of the above mentioned substrate properties. A pretreatment should in the ideal case have low capital and operating costs and be effective on many different substrate types, the side streams (hemicelluloses, lignin) should be easy to recover for further use and the formation of by-products (e.g. furfurals) should be avoided, because these inhibit downstream microbial fermentations.15,60 The pretreated substrates should be efficiently hydrolysed by enzymes at high substrate loadings with short residence times and low enzyme dosage.65 In current processes, the pretreatment is a bottle-neck and no completely satisfying procedure has to date been designed.

Conventional pretreatment methods are typically divided into physical (milling), chemical (alkali and acid treatment) and biological pretreatments (treatment with wood-degrading fungi).60 Pretreatments are often combinations of the above mentioned methods, such as e.g. physicochemical pretreatments which include organosolv, ammonia freeze (or fibre) explosion (AFEX), steam explosion and ammonia recycle percolation (ARP). Conventional pretreatment methods have been extensively reviewed in the recent literature and will not be covered here.60,66-68 In the operational demonstration and industrial scale bioethanol plants, the applied pretreatment methods have been steam/hydrothermal pretreatment, in some cases combined with H2SO4 or SO2 catalysis.3

ILs hold great promise for biomass pretreatment and are widely applicable to different types of lignocellulosic feedstocks, due to their unique potential to dissolve the complete lignocellulosic matrix. The first reports of using ILs for cellulose pretreatment were published by Dadi et al., who dissolved and regenerated MCC in [BMIM]Cl and later in [AMIM]Cl.12,69 After optimization, the regenerated cellulose had even 90 times faster initial enzymatic hydrolysis rates than untreated MCC, which was attributed to the transformation of crystalline cellulose I to amorphous cellulose. IL pretreatment of pure cellulose substrates (especially MCC) has thereafter been widely studied.70-74 MCC serves well as a model substance for studying the effects of IL pretreatment, but cannot be con-sidered an industrially relevant substrate for glucose production. IL pretreatments for the hydrolysis of industrially relevant substrates, such as wheat,75,76 rice straw,77-80 wood,81-85 corn stover,86-88 switchgrass,89-92 bagasse,47,90-92 cotton waste textiles,93 fibre sludge94 and other lignocellulosic feedstocks, have been extensively studied. In mid-2014, the number of articles dealing with IL pretreatments of lignocellulosics or components thereof is close to 600 and is rapidly growing. Several reviews of this topic have recently been published.44-46,95-97 The most frequently applied ILs in pretreatments are those also known as good cellulose solvents: [EMIM]AcO, [AMIM]Cl, [BMIM]Cl and dialkylimidazolium dialkylphosphat ILs (Fig. 3).

Impact of ionic liquid pretreatment on lignocellulose substrate properties. The effect of IL pretreatment on substrate properties depends on both the type of substrate (native lignocellulosic biomass vs. pure cellulose), the type of IL and the pretreatment conditions. The main effects of IL pretreatments can be categorized into: (1) decrystallization or crystallinity transformation from cellulose I to cellulose II, (2) extraction of hemicelluloses and lignin and (3) partial reduction in cellulose DP. In several studies, disruption of LCCs, which are likely to impede enzymatic hydrolysis, has also been suggested to take place during IL pretreatments.31,81,98

Crystallinity analysis of cellulose regenerated from IL solution has shown that the cellulose precipitates either as amorphous cellulose,12,99 cellulose II,71,81 a mixture of amorphous cellulose and cellulose II,91,100 or a mixture of amorphous cellulose and cellulose I,69,74 depending on dissolution and regeneration conditions and the type of cellulose. Cellulose II has, especially when hydrated, been shown to have a higher enzymatic digestibility than native cellulose I, probably due to changes in the hydrogen bonding network in the cellulose crystallites.101 The crystallinity changes during IL pretreatment have been found to be dependent on the type of substrate and in particular to vary when comparing pure cellulose and lignocellulose samples.

Many ILs, e.g. cholinium amino acids77,78 and cholinium mono- and dicarboxylates,98 are suitable for hemicellulose and lignin extraction but leave the cellulose mostly unaltered. Pretreatment with these ILs has been reported to be as efficient as with [EMIM]AcO in some cases.98 It has, however, been suggested that lignin cannot to a high degree be extracted from wood by lignin-dissolving ILs, which do not dissolve cellulose, because the lignin is partly trapped inside the lignocellulosic matrix in wood.53

Depolymerization of the biomass polysaccharide fraction has been reported in conjunction with IL treatments and is beneficial for enzymatic hydrolysis, as cellbiohydrolyases are directly dependent on finding cellulose chain ends as starting points for hydrolysis.7 Bagasse and MCC pretreatment with [HPy]Cl led to significant depolymerization of the substrate polysaccharides,47 whereas somewhat conflicting results have been reported on cellulose depolymerization during pretreatments in e.g. [EMIM]AcO.47,91 Subjecting the pretreatment mixture to microwave irradiation during pretreatment has

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been shown to cause significant additional depolymerization in comparison with simple heating; microwaving and sonication have also been noticed to promote the breakdown of the crystalline cellulose structure during pretreatment.\textsuperscript{76,100,102} Several groups have studied the addition of acid catalysts to the cellulose solutions during IL pretreatment to cause further DP reductions.\textsuperscript{85,89,103} This procedure can be seen as a combination of pretreatment and acid hydrolysis in IL. Acid hydrolysis in IL solution has recently been reviewed by e.g. Tadesse and Luque.\textsuperscript{96}

**Novel approaches to Ionic liquid pretreatments and economic considerations.** Although the first studies on IL pretreatment of lignocellulosics used complete dissolution of the feedstock prior to regeneration, an increasing number of studies have reported efficient IL pretreatment procedures in which the substrate is only partially dissolved or only certain components are dissolved. Wu et al. demonstrated that corn stover could be pretreated in [EMIM]AcO in up to 50% (w/w) solid loading.\textsuperscript{88} For high solid loadings the pretreatment efficiency was not at its optimum, but the digestibility increase was constant for solid loadings in the range of 5–33% (w/w). Under these processing conditions, both disruption of cellulose crystallinity and lignin extraction took place. Techno-economic evaluations have shown that >30% biomass loadings are necessary to make the IL pretreatment economical in economic evaluations have shown that >30% biomass loadings during the pretreatment step, the regeneration procedure was more feasible, whereas both the routes were equally economical at high, 50% biomass loadings. The one-pot procedure was significantly more sustainable regarding water usage. Recovering a pure lignin fraction as a by-product is expected to greatly enhance the economic viability of bioethanol production, although issues such as lignin price and the risk for rapid market saturation are difficult to predict.\textsuperscript{105}

Developing efficient routes for IL recycling is vital if ILs are to be used in biorefineries. To recycle the IL, the simplest proposed procedure is to evaporate the anti-solvent from the IL and then reuse the IL.\textsuperscript{72,75,84,88,90} A major drawback is the high energy consumption for dehydrating the recycled IL by e.g. evaporation or reverse osmosis.\textsuperscript{106} Alternatively, IL recovery can be achieved by creating an aqueous-IL biphasic system by mixing the IL phase with concentrated salt solutions.\textsuperscript{107–109} Serious problems can in both cases be expected with lignin and other components accumulating into the IL.\textsuperscript{65,110} Recently, phase-separable and cellulose-dissolving tetra-alkylphosphonium ILs, which form their own recyclable IL phase upon addition of water, have been reported.\textsuperscript{49} The development of cellulose-dissolving acid–base conjugate ILs, such as those based on the TMG or DBN superbases, offers a new recycling alternative through distillation.\textsuperscript{45,46} This approach would avoid the problems with accumulation of non-precipitated biomass components into the recycled IL.

**Enzymatic hydrolysis of lignocellulosic polysaccharides in the presence of ionic liquids**

Regeneration of cellulose from IL has been shown to be a highly effective pretreatment for enzymatic hydrolysis.\textsuperscript{12} A thorough washing of the regenerated substrate is, however, needed, as even relatively small amounts of IL in the substrate may induce severe inactivation of the cellulases and also inhibit downstream bioprocessing.\textsuperscript{73,91} Results from IL pretreatment studies have shown that the cellulose recovery after precipitation may exceed 100%, suggesting that considerable amounts of IL are trapped in the regenerated material.\textsuperscript{111} Furthermore, the high dilution ratio of the IL caused by the regeneration and subsequent washing presents challenges to economical IL recycling.\textsuperscript{106} The one-pot procedure proposed by Kamiya et al., in which pretreatment and enzymatic hydrolysis are carried out without removing the IL between the steps, has the potential to avoid excessive IL dilution during regeneration and washing, and to avoid losing IL trapped in the regenerated substrate.\textsuperscript{32} Reported low enzyme activities in aqueous solutions of cellulose-dissolving ILs are, however, a serious drawback in the one-pot hydrolysis.\textsuperscript{23}
General factors affecting enzyme activity in ionic liquid solutions

Biocatalysis in organic solvents was pioneered by Klíbanov in the 1980s, showing that enzymes can be used in synthetic applications of organic solvents. The first study of the use of an enzyme (alkaline phosphatase) in an IL, aqueous ethylammonium nitrate ([EtNH₂][NO₃]), was published by Magnuson et al. in 1984. During the last decade, a number of good reviews have been published on both the fundamentals and applications of biocatalysis in ILs. ILs differ in many properties from conventional molecular solvents, and it is not yet completely clear to what extent the laws governing biocatalysis in organic solvents can be applied in IL solutions. Water is the natural medium for most enzymes, interacting with them sufficiently to dissolve them, but not to unfold their active structure. In buffer, the enzymes show a higher stability due to the “salting out” effect, i.e., repulsions between the charged ions in buffer, and the protein’s hydrophobic interior keeps the protein folded. The stabilization energy of folded, active proteins is in most cases very low, calculated to be 30–60 kJ mol⁻¹ in the dissolved state. Most enzymes reported to be active in ILs are lipases, which work at water–oil interfaces. In many cases, increased enzymatic activity, stability and selectivity have been encountered in IL matrices.

Three types of solution should be taken into account when dealing with enzyme stability and action in ILs: anhydrous hydrophobic ILs, anhydrous hydrophilic ILs, and aqueous hydrophilic ILs. Generally, hydrophilic ILs have been considered to be destabilizing and hydrophobic ILs stabilizing for enzymes. Whereas hydrophobic ILs stabilize suspended enzymes, carbohydrates have very low solubilities in these ILs, thus rendering them of marginal interest as solvents for biocatalytic reactions on dissolved polysaccharides. Hydrophobic ILs do not dissolve enzymes, but rather suspend them. Enzyme-dissolving ILs often inactivate the enzymes, with the exception of a few ILs, e.g., [Chol]H₂PO₄. Some hydrophilic ILs may promote refolding of a denatured protein, as has been demonstrated with hen egg white lysozyme in [EtNH₂][NO₃].

In the early studies with enzymes in ILs, diverse problems were encountered regarding IL purity, unexpected pH shifts and precipitation of buffer salts. Especially, impurities in ILs have led to conflicting results. The main impurities in ILs are often residual halides and water, which cause problems especially in reverse hydrolysis reactions. Very small chloride impurities have been shown to lead to even complete lipase inactivation and different purification methods such as washing an IL with an aqueous Na₂CO₃ solution greatly increased the lipase activity. Enzyme stability in hydrophilic solvents has been predicted by the solutions' Hildebrand solubility parameter (δ), dielectric constant (ε), dipole moment (μ) or octanol-water partition coefficient (log P), but applying these parameters in IL systems does not always appear to be straightforward. In aqueous salt solutions, the protein stability has with success been related to the Hofmeister series or the kosmotropicity and chaotropicity of the IL ions. IL ions have been found to mainly follow the Hofmeister series in their effect on protein stability, although exceptions exist and especially the effect of larger ions with greater complexity is harder to predict. It is also important to notice that the Hofmeister series is only applicable when the aqueous IL is sufficiently diluted for the ions to be dissociated from each other. The imidazolium cations common in cellulose-dissolving ILs have been ordered into the following series based on the destabilising effect on proteins: [BMMIm]⁺ > [EMIm]⁺ > [DMIm]¹. Cholinium cations have been found to be stabilizing, whereas increasing the cation hydrophobicity has been reported to decrease enzyme melting points. Results published by Kaar et al. suggest the enzyme inactivation to be mostly dependent on the anion. Nucleophilic anions are possibly able to coordinate to positively charged surface residues on the enzyme and cause conformational changes. It has been found to depend strongly on the anion whether enzyme inactivation is reversible or irreversible. As discussed by Lee et al., enzymes appear to be active in anhydrous ILs containing BF₄⁻, PF₆⁻, bis(trifluoromethylsulphonyl) imide (Tf₂N⁻) or SbF₆⁻ anions (i.e. hydrophilic ILs), but not in anhydrous ILs with NO₃⁻, acetate, trifluoroacetate or halide anions (i.e. hydrophobic ILs). Large anions spread their charge on several atoms and are thus believed to form weaker hydrogen bonds with the enzymes and less disruption of the structure. Carbohydrate-dissolving ILs have anions which form strong hydrogen bonds, thus also exerting a denaturing effect on enzymes.

The inactivating effect of both hydrophilic organic solvents as well as ILs has been suggested to be caused by dehydration of essential water molecules from proteins, but this inactivation mechanism was deemed unlikely by Kaar et al. when studying a lipase in IL, as dehydration did not return its lost activity. It has been proposed that enzyme incubation in hydrophobic ILs such as [BMMIm]PF₆ could protect the essential water molecules from stripping. Having a high number of charged amino acid residues on the protein surface appears to stabilize enzymes in strong salt solutions and ILs.

Cellulose-dissolving ILs are usually basic which means that the medium pH will not be at the enzyme optimum in IL solutions. The question whether the pH effect is a major reason for enzyme inactivation e.g. in the case of cellulases is under debate. IL media usually have high viscosities, leading to lower mass transfer rates and slower reactions. In many cases the reaction system can be diluted with a low-viscosity organic solvent, which has also been shown to be applicable for cellulose-solutions with good success for increasing the cellulose dissolution kinetics. A third general effect of the presence of ILs in a reaction medium is an increased ionic strength, which affects the action of most enzymes. In a study by Engel et al. the cumulative effect of increased viscosity and ionic strength was compared to the inactivation measured at the corresponding concentration of [DMIm]DMP at fixed pH, and it could be concluded that the
presence of [DMIM]DMP also had other inactivation mechanisms than those caused by the high viscosity and ionic strength.132

Cellulase activity and stability in ionic liquid solutions

Many reports have been published on the activity, stability and action in hydrolysis experiments of mesophilic cellulases from e.g. the *Trichoderma reesei* and *Aspergillus niger* cellulase systems, either with the cellulases in a monocomponent form or as cellulase cocktails (Table 1). Of the monocomponent cellulases studied in ILs, endoglucanases are the most studied. Studies of monocomponent cellobiohydrolases, β-glucosidases and various hemicellulases have also been published. In addition to the mesophilic cellulases, an increasing number of studies have been concentrating on cellulases derived from different extremophilic sources, as thermo-, salt- and alkalistability appears to be linked to tolerance to cellulose-dissolving ILs. Carboxymethylcellulose (CMC) has been the most employed substrate in the activity and stability measurements for endoglucanases, but studies of the hydrolysis of different solid substrates have also been increasingly reported. Three different types of measures for cellulolytic performance in IL prevail: activity measurements (initial reaction velocity), stability (retained cellulolytic activity after incubation in IL) and long term hydrolysis experiments. Occasionally, cellulase stability in IL solution has also been studied by other, non-reaction based techniques, such as fluorescence spectroscopy or differential scanning calorimetry (DSC).33,136,140 The use of computational methods such as molecular dynamics has also contributed with interesting knowledge on glycosyl hydrolase–IL interactions.141

Turner *et al.* were the first to report cellulase activity in cellulose-dissolving ILs.33 *Trichoderma reesei* cellulase was found to be much inactivated in [BMIM]Cl, which was proposed to be due to the high concentration of Cl⁻ ions. A high degree of dilution of the IL solution with water led to refolding of the cellulase, suggesting the inactivation to be reversible in [BMIM]Cl for the studied enzyme. In another early study, a *Humicola insolens* cellulase was found to be stable in [BMIM]-PF₆ and [BMIM]BF₄, which do not dissolve cellulose, but the cellulase was inactivated in cellulose-dissolving [BMIM]Cl.142

Salvador *et al.* found the activity of *A. niger* cellulase to correlate linearly with the water activity in [BMIM]Cl solutions.143 The cellulase was noticed to regain its activity upon dilution with buffer after incubation in 10% [BMIM]Cl, supporting the earlier observations by Turner *et al.* about reversible inactivation in this IL.133 Cellulase inactivation in ILs has been shown to be temperature dependent, as a cellulase mixture had only minor activity losses during incubation in 30% [EMIM]AcO at 4 °C, but more rapid inactivation at 50 °C.144

Engel *et al.* made activity measurements on α-cellulose, which, being a solid substrate, represents a more practical substrate for activity measurements than the usual soluble substrates.132 Interestingly, cellulases have been found to show increased thermal stability in the presence of regenerated cellulose, indicating that inactivation kinetics measured in IL solutions without substrate may not give the complete picture of cellulase inactivation in practice.73 In a study by Ebner *et al.* residual endoglucanase activity measurements were done on both soluble CMC and regenerated pulp after incubation in 90–100% (w/w) [EMIM]AcO, whereby it was noticed that the endoglucanase retained residual activity for significantly longer times in the presence of (dissolved) pulp, although the different inactivation kinetics measured may have also been due to different sensitivities and substrates of the two activity assays used.145 Engel *et al.* found that cellulases retained their activity towards soluble substrates to a higher degree than towards α-cellulose in aqueous [DMIM]DMP, demonstrating that the outcome of activity measurements in IL solutions may also be dependent on the substrate used.132

Wolski *et al.* have reported a screening method using green fluorescent protein as a probe for determining protein stability in ILs with fluorescence measurements.146 Preserved protein fluorescence was found to correlate with a well-retained activity of cellulase in IL solutions. [DMIM]DMP and [EMIM]lactate were identified by this method as potentially enzyme-compatible ILs for one-pot cellulose hydrolysis and *T. reesei* cellulases were found to retain their activity in up to 40% [DMIM]DMP or [EMIM]lactate and *A. niger* β-glucosidase in up to 60% [DMIM]DMP in validation experiments. Bose *et al.* studied the stability of a *T. reesei* cellulase cocktail in eight different ILs and found tris(2-hydroxyethyl)methylammonium methylsulphate ([HEMA]MeSO₄) to stabilize this enzyme mixture at temperatures up to 115 °C.136 Although [HEMA]MeSO₄ showed great promise in view of its cellulase compatibility, this IL is only capable of dissolving ~1% of cellulose,149 which may limit its applicability to biomass pretreatment. Zhi *et al.* studied the cellulase stability in a series of dialkylphosphate ILs with increasing alkyl substituent size.147 [DMIM]DMP, with the smallest alkyl substituents, had the highest cellulase compatibility. In a comparison of the cellulase compatibility of the most common cellulose-dissolving ILs ([DMIM]DMP, [BMIM]-Cl, [EMIM]AcO and [AMIM]Cl]), cellulase activity was observed to generally decrease with 70–85% in the presence of 10% (v/v) IL as compared to buffer systems.132 In some cases larger differences in relative cellulase activity were noticed for the same IL from different manufacturers than between different ILs. [DMIM]DMP could be concluded to be the least cellulase-inactivating of the studied ILs. Likewise, Yang *et al.* identified [DMIM]DMP as the most cellulase-compatible IL out of a set of six phosphate ILs and optimized different hydrolysis parameters in this IL.70 Substituted imidazoles (i.e. uncharged imidazole derivatives) have been shown to cause considerable inhibition of glycosyl hydrolases and the inhibition efficiency was very dependent on the substituent types and positions on the imidazole ring.148 The results suggest that some imidazole derivatives interact in a very precise manner with glycosyl hydrolases, thus inhibiting them.

In a comparison of four commercial cellulase preparations derived from *Aspergillus sp.*, *A. niger*, *Trichoderma viride* and *T. reesei*, the cellulase extract from *Aspergillus* sp. was found to retain a remarkable activity in 30% (v/v) of five different ILs,
### Table 1  Published studies on the action of cellulases and other glycosyl hydrolases in ionic liquid solutions. The table entries highlight the used substrates, enzymes, ionic liquids, the type of measurement (activity, hydrolysis, stability) and the main results of each study. Abbreviations: CMC = carboxymethylcellulose; MCC = microcrystalline cellulose; 4-NP = 4-nitrophenol; 4-MUC = 4-methylumbelliferyl-β-D-cellobioside; CBM = carbohydrate-binding module; PEG = polyethylene glycol; DSC = differential scanning calorimetry. IL ions: [BMIM] = 1-butyl-3-methylimidazolium; [EMIM] = 1-ethyl-3-methylimidazolium; [DMIM] = 1,3-dimethylimidazolium; [TMIM] = 1,2,3-trimethylimidazolium; [AMIM] = 1-ethyl-3-methylimidazolium; [BMP] = 1-butyl-3-methylpyridinium; [DMAP] = dimethyl phosphate; [HMIM] = 1-hydroxyethyl-3-methylimidazolium; DEP = diethylphosphate; [TMG] = tetramethylguanidinium; DBP = dibutylphosphate; HEMA = tris-(2-hydroxyethyl)methylammonium; [E(OH)MIM] = 1-hydroxyethyl-3-methylimidazolium; DBN = 1,5-diazabicyclo-[4.3.0]non-1-allyl-3-methylimidazolium; DEP = diethylphosphate; [TMG] = tetramethylguanidinium; DBP = dibutylphosphate; HEMA = tris-(2-hydroxyethyl)methylammonium; [E(OH)MIM] = 1-hydroxyethyl-3-methylimidazolium; DBN = 1,5-diazabicyclo-[4.3.0]non-1-allyl-3-methylimidazolium.

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<td><em>Trichoderma reesei</em> cellulase</td>
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<td>Activity, stability</td>
<td>Cellulose-dissolving ILSs were found highly inactivating for the first time, PEG stabilization was beneficial for cellulase stability, fluorescence spectroscopy was used for stability measurements.</td>
<td>Turner et al. (2003)⁵¹</td>
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<td><em>Trichoderma reesei</em> cellulase</td>
<td>[EMIM]DEP</td>
<td>Hydrolysis</td>
<td>In situ one pot cellulose hydrolysis in IL was first introduced, hydrolysis took place in up to 40% (v/v) IL.</td>
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<td>CMC</td>
<td>Bacterial cellulases</td>
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<td>Supernatants from thermophilic bacterial consortia</td>
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<td>Cellulase from Continent Biotech (Shanghai) Co.</td>
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<td>[DMIM]DMP, [EMIM]AcO, [EMIM]Lactate</td>
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<td>CMC</td>
<td>Halorhabdus utahensis cellbiohydrolase</td>
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<td>Cotton cellulose</td>
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<td>Regenerated cellulose, cellubiose</td>
<td>Trichoderma reesei Cel7A and Cel7B, A. niger β-glucosidase (Novozym 188)</td>
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<tr>
<td>CMC</td>
<td>Bacterial cellulases</td>
<td>6 different ILs</td>
<td>Activity, stability</td>
<td>IL tolerance linked to thermo- and halophilicity</td>
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<td>Sulfolobus solfataricus endoglucanase</td>
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<td>CMC, MCC, straw, cotton, filter paper</td>
<td>Cellulase powder</td>
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<td>Hydrolysis, stability</td>
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<td>Ilmberger et al. (2012)162</td>
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<td>IL-pretreated switchgrass</td>
<td>Cellulases from a thermophilic bacterial consortia</td>
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<td>Hydrolysis</td>
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<td>Hydrolysis</td>
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<td>Su et al. (2012)190</td>
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<tr>
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<td>Commercial cellulase cocktail</td>
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<td>MCC</td>
<td>Trichoderma reesei Cel5A and Cel7B</td>
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<td>Sodium alginate-immobilized cellulase</td>
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<td>5 imidazolium-based ILs</td>
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<td>Trichoderma longibrachiatum GH11 xylanase</td>
<td>[EMIM]AcO, [EMIM]EtSO₄</td>
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<td>MCC, CMC</td>
<td>Cellulase powder</td>
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<td>Activity, hydrolysis, stability</td>
<td>Cellulase was stabilized by PEGylation of the N-terminal and used for hydrolysis in IL solution</td>
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<td>Xylan</td>
<td>Dictyoglomus thermophilum GH11 xylanase</td>
<td>[EMIM]AcO</td>
<td>Activity, stability</td>
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<td>Paenibacillus taminensis endoglucanases</td>
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<td>MCC, pulp</td>
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<td>[EMIM]AcO, TMG- and DBN-based distillable ILs</td>
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<td>D’Arrigo et al. (2014)¹³¹</td>
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<td>Cotton linters, pulp, CMC</td>
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</table>
among which was the cellulose-dissolving [BMIM]Cl earlier reported to be severely inactivating.\textsuperscript{149} The other cellulases became almost completely inactivated under the corresponding conditions. Also the long term stability of this cellulase was found to be extraordinarily high in 60% (v/v) of IL. This finding shows that IL-tolerant cellulases can be found in already commercialised enzyme products.

\textit{T. reesei} Cel7A (cellobiohydrolase) and Cel7B (endoglucanase) were in one study found to be inactivated in a similar manner in the presence of [DMIM]DMP, with some residual activity in 30% (v/v) of this IL.\textsuperscript{150} The β-glucosidase of \textit{A. niger} was more IL sensitive and lost its activity already in 15% (v/v) of [DMIM]DMP.\textsuperscript{150} Cellulase inactivation in aqueous [DMIM]-DMP has been found to be reversible, and also the storage stability has been found to be sufficiently steady for long hydrolyses in 10% (v/v) [DMIM]DMP, after a first initial rapid decrease in the activity.\textsuperscript{132,150} Wahlström et al. measured the residual endoglucanase activity of \textit{T. reesei} Cel5A and Cel7B after incubation in 90% (v/v) [DMIM]DMP at 45 °C; a short incubation time of 15 min led to an activity decrease of ca. 50%, but the activity then stayed steady at this level for days as verified for Cel5A.\textsuperscript{131} \textit{T. reesei} Cel12A (EGIII) has been found to show both good activity and stability in [BMIM][Cl], in which the enzyme was even stabilised at 75 °C as compared to incubation in buffer.\textsuperscript{151}

There appear to be great differences between the inactivation rates (stability) of cellulases in different cellulose-dissolving imidazolium-based ILs. When the inactivation of endoglucanase activity on CMC was studied in pure [EMIM]-AcO and [DMIM]DMP, the endoglucanase was noticed to be completely inactivated in 4 h in [EMIM]AcO at 40 °C, whereas the activity only declined slowly in [DMIM]DMP and over 60% of the initial activity was retained even after 3 days.\textsuperscript{145,152} Furthermore, the inactivation appeared to be irreversible in [EMIM]AcO, as no regain of activity could be measured when the incubated cellulase mixture was diluted to below 4% of IL,\textsuperscript{145} contrasting with the earlier reported reversibility of cellulase inactivation reported in aqueous [BMIM][Cl] and [DMIM]DMP.\textsuperscript{131,132,143}

\textbf{Substrate binding of cellulases and other glycosyl hydrolases and the role of the carbohydrate-binding module (CBM) in ionic liquids}

Pottkämper et al. proposed the carbohydrate-binding module (CBM) of cellulases to be particularly sensitive to ILs, based on comparing the activity of different cellulase mutants on CMC in several ILs.\textsuperscript{155} In a comparison of \textit{T. reesei} Cel5A and its core domain (CD, \textit{i.e.} the cellulase lacking its CBM), the CBM was found important for MCC hydrolysis in buffer, but not in the studied IL solution.\textsuperscript{131} In the presence of 20% (v/v) [DMIM]DMP the hydrolysis yield of the intact cellulase decreased by over 80%, whereas the action of the CD cellulase was not significantly affected by adding IL to the hydrolysis mixture, which was interpreted as a strong IL-sensitivity of the CBM. This was confirmed with hydrolysis experiments in a follow-up study in which the action of the cellobiohydrolase \textit{T. reesei} Cel7A, the endoglucanase \textit{T. reesei} Cel5A and their CDs was compared in IL solutions.\textsuperscript{154} Binding isotherms measured using tritium-labeled cellulases confirmed that the presence of [DMIM]DMP and [EMIM]AcO seriously decreased the cellulase binding to MCC even at 4 °C. The cellulase binding is temperature-dependent and so it can be anticipated that at hydrolysis temperatures of 40–50 °C cellulase binding is even lower in the presence of ILs. The reasons for the reduced cellulase binding were not elucidated in this study, but the authors suggested several possible explanations: solvent effects interfering with the hydrophobic interactions governing the binding to cellulose, conformational changes in the CBM or even complete unfolding of the CBM. Another interesting result from this study was that the \textit{T. reesei} cellobiohydrolase, with the catalytically active site in a tunnel, was able to bind to cellulose to a much higher degree in IL matrices in comparison with the endoglucanase with the active site in a surface cleft.

Competitive inhibition by ILs with subsequent reduced substrate binding as the reason for the observed glycosyl hydrolyase inactivation has been discussed in some articles. Molecular dynamics simulations by Jaeger and Pfandtner suggested that imidazolium cations are trapped in the active site cleft of a xylanase (xylanase II from Trichoderma longibrachiatum, a GH11 enzyme) and that the imidazoliums may cause competitive inhibition of the substrate binding.\textsuperscript{141} Kinetic measurements also suggested competitive inhibition of the active site in xylanases as the main reason for the observed low hydrolysis efficiency in IL solution.\textsuperscript{155}

\textbf{Hemicellulases and other glycosyl hydrolases in ionic liquids}

Hemicellulases are often close in structure and catalytic mechanism to the cellulases, which is why it is of interest to compare the action of hemicellulases and other related enzymes, \textit{e.g.} glycosyl transferases, to cellulases in IL matrices. Hemicellulases also play an important role in the total enzymatic hydrolysis of lignocellulosic biomass, which typically contains significant amounts of hemicelluloses. Different glycosyl hydrolases have been studied in the synthesis of small oligosaccharides in aqueous systems with ILs added as cosolvents.\textsuperscript{156–158} Thomas et al. studied the activity of two β-glucosidases, a xylanase and two arabinofuranosidases in the presence of three dialkylphosphates and [EMIM]AcO and found the dialkylphosphates to be generally more compatible with these enzymes.\textsuperscript{159} When the activity of two α-galactosidases and a β-glucosidase was studied in imidazolium-based ILs, a strict correlation between the thermostability and IL-tolerance of these glycosyl hydrolases could be established.\textsuperscript{160}

Introducing a new disulphide bridge into the N-terminus of an extremophilic \textit{Dictyoglomus thermophilum} GH11 xylanase increased its activity at elevated temperatures and to some extent in the presence of [EMIM]AcO.\textsuperscript{155} This xylanase was inactivated linearly to the [EMIM]AcO concentration and the enzyme was completely inactivated at 25% of this IL. The main IL effect was not in changing the protein structure, but the IL was proposed to cause severe effects on the substrate binding capability of the xylanase. Similar results were obtained in a
molecular dynamics simulation of the effects of [EMIM]AcO on *Trichoderma longibrachiatum* xylanase II (a GH11 enzyme). 141 This simulation study also demonstrated the strong influence of IL on the protein structure, e.g. there are fluctuations in the structure close to the substrate binding site of this xylanase.

**Discovery and development of ionic liquid-tolerant cellulases**

Metagenomic approaches and mutation experiments have led to encouraging results in finding more IL-tolerant cellulases. Adsul et al. reported several glycosyl hydrolases from *Penicillium janthinellum* mutants which exhibited an improved IL-tolerance. 161 In a screening study by Pottkämper et al., the IL-tolerance of 24 bacterial cellulases from metagenomic libraries was elucidated in a number of different ILs. 153 Most of the cellulases had very low IL-tolerance, but it was found that most cellulases displaying increased IL-tolerance also showed high halotolerance, suggesting the correlation between halotolerance and IL-tolerance. Also, Ilmberger et al. reported that several cellulases with good IL-tolerances were found when screening metagenomic libraries. 162

Cellulases have been screened from different extremophilic and halophilic sources for better enzyme performance in cellulose-dissolving ILs. 162–166 Generally, there appears to be a clear correlation between thermostability and IL-tolerance. Increased IL-tolerance has been reported for enzymes active at high pH originating from solvent-tolerant bacteria. 167, 168 Extremotolerant (high alkali-, thermo- and salt tolerance) cellulases derived from *Paenibacillus tarimensis*, found in some very saline environments, have been reported to have a high tolerance towards the ILs [BMIM]Cl and [EMIM]AcO. 169 The activity and stability of two α-glucosidases and a β-glucosidase in imidazolium-based ILs were found to correlate well with the enzymes’ thermostability in another study. 160 The cellulase inactivation was found to take place through a slow irreversible denaturation which was dependent on the enzyme properties and the specific IL, but the reduced activity was also suggested to be due to non-competitive inhibition by the imidazolium ions and low water activity at high IL concentrations.

When screening cellulases from micro-organisms living in highly saline environments for IL-tolerance, several cellulases with high salt (NaCl) tolerance were found. The high IL-tolerance apparently also correlated with thermostability. 170 These cellulases showed good activity in [BMIM]AcO and [EMIM]AcO in the studied IL concentration range of 0–20% (v/v). The IL-tolerance was explained by a high number of acidic amino acid residues on the cellulases’ surfaces, which are suggested to prevent protein aggregation in highly ionic environments. Similarly, a haloalkaliphilic *Halorhabdus utahensis* cellulase had good IL-tolerance, which was linked to the presence of a large number of negatively charged amino acid residues on the protein surface, a low content of hydrophobic amino acids and a compact packing of the protein structure. 171 A negative charge on the protein surface is anticipated to interact well with water and high ion concentrations, 130 and this hypothesis is supported by many of the reviewed studies.

Gladden et al. screened for IL-tolerant cellulases from a library of thermophilic cellulases derived from a switchgrass-adapted microbial community, and found many cellulases active in 10% (v/v) [EMIM]AcO, and some displaying significant activity even in 40% (v/v) [EMIM]AcO. 172 Both endoglucanase, cellobiohydrolase and β-glucosidase activities were studied in the presence of [EMIM]AcO. The correlation between thermostolerance and IL-tolerance could also be strengthened as those enzymes with the highest optimum temperatures also showed the best IL-tolerance. Some of the IL-tolerant enzymes showed good activity under slightly alkaline conditions, but it was generally concluded that alkali tolerance and IL-tolerance were not strongly correlated for the studied enzymes. Similar results were obtained when the hydrolytic performances of an alkali-tolerant and a thermostable endoglucanase were compared in MCC hydrolysis in [EMIM]AcO and tetrarmethylguanidinium acetate ([TMGH]-AcO). 153 A surprising result of the study by Gladden et al. was that several of the studied cellulases showed increased activity in low (5–10%) concentrations of [EMIM]AcO, and in some cases up to a several fold increase in activity could be measured, compared to the activity in buffer. 173 The isolation of an IL-tolerant cellulase-producing fungus (*Aspergillus fumigatus*) from chemically polluted habitats was recently reported. 173 The thus derived cellulase was used in the hydrolysis of rice straw in the presence of 25% (v/v) [EMIM]DMP with approximately two-fold yields as compared to the buffer reference system.

A β-glucosidase from the hyperthermostable *Pyrococcus furiosus* showed high IL-tolerance and retained full activity even in the presence of 50% [DMIM]MeSO4 at 80 °C, and was inactivated but not irreversibly denatured in 70% of this IL. 174 When Kudou et al. studied the activity of a β-glucosidase from *Thermotoga maritima* in aqueous solutions of [EMIM]AcO, [EMIM]Cl, [BMIM]Cl and [BMIM]AcO during 15 min activity measurements, [BMIM]AcO was found to increase the activity of this cellulase, whereas the other ILs slightly decreased the activity. 175 The authors proposed the increased activity to be due to increased flexibility of the cellulase caused by the presence of [BMIM]AcO. The introduction of an ionic strength activation switch into an endoglucanase by directed evolution was recently reported. 176 The endoglucanase mutant had low activity in buffer but was highly activated when 7.5% (v/v) of [BMIM]Cl was added to the reaction medium. The most IL-tolerant cellulase to date is a hyperthermophilic and halophilic GH12 endoglucanase from *Sulfolobus solfataricus*, which was reported by Klose et al. to hydrolyse dissolved or regenerated MCC well in 80% (v/v) [DMIM]DMP and [EMIM]AcO at 90 °C. 164 In all, very impressive progress has been made in discovering cellulases with high IL-tolerances as compared to the earlier studied mesophilic cellulases.

**Enzyme-compatible cellulose-dissolving ionic liquids**

The attempts to design IL- and enzyme-compatible systems have concentrated mainly on discovering or developing more suited enzymes for this purpose, whereas the reverse approach,
Stabilization techniques for cellulases in ionic liquids

Cellulases may be stabilised by employing different enzyme stabilization techniques. Several reviews dealing with various stabilisation procedures for enzymes in IL systems are available.\(^{120,128}\) Jones and Vasudevan reported the cross-linking of cellulase with glutaraldehyde and its use in media containing low concentrations (2% v/v) of 1-ethyl-3-methylimidazolium diethylphosphate ([EMIM]DEP).\(^{177}\) Cellulase immobilization onto sodium alginate has also been reported for saccharification in [DMIM]DMP solutions.\(^{178}\) In a study by Lozano et al. a commercial cellulase was immobilized on a polymeric support, Amberlite XAD4, and subsequently coated with a hydrophobic IL, which stabilized the cellulase against thermal inactivation.\(^{179}\) The immobilized and IL-coated enzyme showed a better stability in [BMIM]Cl, known to be a strongly inactivating IL. Cellulase immobilization on chitosan has also been reported to increase cellulase action on CMC in the presence of [DMIM]DMP.\(^{180}\)

Several successful stabilization techniques for cellulases employ the use of PEG. Turner et al. reported a better IL-tolerance of cellulase after lyophilizing it with PEG.\(^{33}\) Another successful approach was to attach PEG chains to the N-terminal end of a cellulase.\(^{181}\) The PEG chain was suggested to form a hydrophilic region around the cellulase, which protects the cellulase from the IL and also increases the interactions between the modified enzyme and cellulose. \textit{Trichoderma viride} cellulase was stabilised as liposomes, which successfully increased the IL-tolerance of the enzyme.\(^{182}\) Nordwald et al. have reported surface charge modification of cellulases by chemical succinylation as an efficient means of increasing the enzyme stability in IL.\(^{183}\) MCC hydrolysis in 15% (v/v) [BMIM]-Cl showed significantly higher yields with succinylated \textit{T. reesei} cellulase as compared to the native enzyme. Furthermore, cellulase succinylation led to lower cellulase binding to lignin.

Hydrolysis experiments in ionic liquid solutions

Activity and stability measurements in IL solutions are commonly made with soluble substrates and short hydrolysis times (from 5 min to some hours). The use of soluble substrates may not correspond well to total hydrolysis experiments with solid lignocellulosic substrates. For process development and research, it is necessary to study how the ILs affect the hydrolysis of real substrates during longer times. In the original work by Kamiya et al., cellulose regeneration from IL was observed to greatly increase hydrolysis rates, but the presence of the IL used, [EMIM]DEP, halted the subsequent enzymatic cellulose hydrolysis when the IL concentration was >40% (v/v).\(^{32}\) After this initial work, enzymatic hydrolysis of IL-pre-treated substrates in the presence of various amounts of different ILs (one-pot procedure) has been studied with both cellulase cocktails and monocomponent cellulases.

In the total enzymatic hydrolysis of regenerated cellulose and yellow poplar in 15% (v/v) [EMIM]AcO, an almost complete conversion of the regenerated cellulose was achieved, whereas IL-pre-treated yellow poplar had lower hydrolysis yields (33%).\(^{144}\) The hydrolysis of regenerated filter paper in aqueous 10% [DMIM]DMP had doubled yields compared to the corresponding hydrolysis of untreated filter paper.\(^{147}\) When comparing untreated and from [DMIM]DMP regenerated \(\alpha\)-cellulose hydrolysis in buffer and in 10, 20 and 30% (v/v) [DMIM]DMP, it was found that the initial hydrolysis rates were greater for regenerated cellulose even in 30% (v/v) [DMIM]DMP compared to untreated \(\alpha\)-cellulose in buffer.\(^{132}\) However, in prolonged hydrolysates the increasing presence of IL lowered the hydrolysis yields. In the hydrolysis of IL-pre-treated \textit{Miscanthus} with \textit{T. reesei} cellulase in aqueous [DMIM]DMP and [EMIM]lactate,
[DMIM]DMP was found to be significantly more enzyme-compatible than [EMIM]lactate.146 Hydrolysis still took place in 50% (w/w) [DMIM]DMP. High hydrolysis yields have been reported in the hydrolysis of delignified and IL-treated bagasse in the presence of [EMIM]DEP.95 Li et al. reported the successful hydrolysis of straw, cotton and filter paper with a cellulase mixture in 15% (w/v) of [E(OH)MMIM]AcO, an IL specially designed for combined cellulose-solubility and enzyme-compatibility.144 Auxenfans et al. showed that cellulose could be successfully hydrolysed by a T. reesei cellulase cocktail in a one-pot procedure with various amounts of 1-ethyl-3-methylimidazolium methylphosphonate ([EMIM]MeO4[H]PO4)].184 The highest hydrolysis yields were obtained in 10% (v/v) IL and comparably good yields were obtained in up to 40% (v/v) IL, but at higher IL concentrations hydrolysis yields were very low, in agreement with the results obtained previously by Kamiya et al.52

Studying the action of monocomponent cellulases on hydrolysis in IL solutions is necessary to gain in-depth knowledge on how the individual cellulases respond to the presence of IL. The effect of [EMIM]AcO and [DMIM]DMP on the hydrolytic action of T. reesei endoglucanases Cel5A and Cel7B has been studied in long (72 h) hydrolyses on untreated MCC and both ILs were found to be severely harmful to the hydrolysis.131 In 40% (v/v) of ILs there was almost no formation of soluble hydrolysis products. On the other hand, in 90% (v/v) [DMIM]-DMP both endoglucanases were able to reduce the molecular weight of MCC, which was not observed in other hydrolysis matrices, including buffer. [DMIM]DMP did not appear to inactivate the endoglucanases irreversibly, and the treatment of MCC in 90% IL partly dissolved the substrate, rendering it accessible for intrachain scission. In a follow-up study it was found that [DMIM]DMP did not completely inactivate the endoglucanase Cel5A but rather slowed it down, whereas the enzymatic hydrolysis stopped completely in a matter of hours in aqueous [EMIM]AcO.185 In another study, T. reesei Cel12A hydrolysed MCC in low and high concentrations of [BMIM]Cl, but in media containing 1:1 [BMIM]Cl-water, this enzyme was virtually non-active in hydrolysis.151 In addition to imidazolium-based ILs enzymatic cellulose hydrolysis has also been studied in ILs consisting of [TMGH]− and [DBNH]− cation-exchange ILs. These ILs are interesting in combining the cellulose dissolution capability with being distillable. They were, however, found to be at least as harmful to the action of T. reesei Cel5A as the [EMIM]AcO used as the reference IL in this study.

The action of A. niger β-glucosidase (Novozyme 188) on cello-oligomer hydrolysis in aqueous [DMIM]DMP and [EMIM]-AcO has been studied in long hydrolysis experiments.185 [EMIM]AcO was significantly more harmful to enzyme performance than [DMIM]DMP. The β-glucosidase was sensitive to the increase of pH but the different basicity of the studied ILs could not alone explain the difference in their impact on the enzyme. Long-time hydrolysis (72 h) of MCC with T. reesei cellobiohydrolase Cel7A and endoglucanase Cel5A has also been studied in aqueous [DMIM]DMP and [EMIM]AcO.154 In comparison, the cellobiohydrolase was somewhat more tolerant towards the presence of IL than the endoglucanase Cel5A.

New innovative concepts for enzymatic lignocellulose hydrolysis in IL matrices have recently been proposed. Klose et al. added a hyperthermophilic endoglucanase gene to induce cellulase production into a plant.164 The enzyme is not active under normal plant growth conditions, whereas under pretreatment conditions with high temperatures (90 °C) it becomes active and starts degrading the plant cell wall from the inside. This cellulase also had an extraordinary IL-tolerance, meaning that ILs can be used for pretreatment. The combination ofIL pretreatment with [EMIM]AcO, [EMIM]DEP and [EMIM][Cl], enzymatic hydrolysis and fermentation to ethanol in a one-pot procedure has been demonstrated by Nakashima et al.186 The employed yeast displayed cellulases on the cell surface, but additional free cellulases needed to be added for an efficient cellulose hydrolysis. The yeast was capable of fermenting the liberated saccharides to ethanol in a maximum IL concentration of 200 mM.

The one-pot or in situ hydrolysis is still a new concept and suffers from problems with enzyme activity in the presence of ILs. Cellulases with high IL-tolerance have, as has been described in this review, recently been reported and potentially offer a solution to this problem. Another challenge needing further research efforts is the separation of released glucose from the IL-containing hydrolysate in an economical manner. For this purpose, the use of alumina column chromatography,187 liquid–liquid extraction using organic boronate carriers,188 and combined filtration and electrodialysis procedures189 has been proposed. In spite of many remaining challenges, one-pot hydrolysis appears as an interesting and relevant alternative to the previously suggested regeneration pathway for using ILs in lignocellulose saccharification.

Optimization of cellulase cocktails for use in ionic liquid solutions and on ionic liquid-pretreated substrates

Most commercial cellulase cocktails for total enzymatic hydrolysis of lignocellulose have fairly low tolerance towards biomass-dissolving ILs. Therefore, some research groups have optimized new cellulase mixtures for better performance in IL matrices. In the work of Park et al., an optimized cellulase cocktail (“Therm”) consisting of thermophilic enzymes retained over 50% of its activity in 20% (v/v) [EMIM]AcO at 70 °C, as compared to aqueous conditions.166 Engel et al. optimized a cellulase cocktail for the hydrolysis of regenerated α-cellulose in the presence of 10% (v/v) [DMIM]DMP based on inactivation data acquired for the cellulases T. reesei Cel7A and Cel7B and A. niger β-glucosidase in this IL.155 In several articles whole cell cultures have been studied under conditions similar to those in the hydrolysis of IL-pretreated biomass for developing IL-tolerant enzyme systems.190–192 In addition to the need for IL-tolerant cellulase cocktails, the increased amorphous character of regenerated cellulose substrates emphasizes the role of endoglucanases in total hydrolysis, as they apparently have much larger target regions in the substrate.156 Specific cellulases, such as the thermophilic Cel5A from Thermotoga
maritima, have been subjected to random mutation experiments aimed at increasing their activity on especially IL-pre-treated switchgrass.193 Also more complex cocktails with various cellulase and hemicellulase activities have been optimized for the hydrolysis of IL-regenerated lignocellulose.82

Analysis of hydrolysis products in ionic liquid solutions

The presence of IL in enzymatic hydrolysates has in several studies been found challenging for saccharide analysis.185,194,195 Accurate and high-throughput analytics are, however, needed for following hydrolysis kinetics and formed saccharide product distributions. Typically, the yields of released saccharides are quantified by spectrophotometric assays such as the 3,5-dinitrosalicicylic acid (DNS)196 or para-hydroxybenzoic acid hydrazide (PAHBH)197 assays, which have the benefit of allowing rapid analysis of reducing saccharides, but suffer from drawbacks such as not being able to discriminate between oligomers of different lengths and types. For more accurate analyses of saccharide compositions in liquid samples, high performance liquid chromatography (HPLC), gas chromatography (GC) and capillary electrophoresis (CE) techniques are used. In chromatography, the presence of IL has been reported to be problematic as the columns only tolerate low amounts of salts.194 ILs may also give rise to broad peaks which overlap with the saccharide peaks in the chromatograms and the retention times varied with sample composition.194 ILs have been used as components in the background electrolyte in CE when analysing saccharides, but with IL concentrations higher than 20–30 mM, baseline fluctuations were observed.198

The presence of IL appears to interfere with the DNS assay at least in high concentrations (over 40%, v/v),185 although it has been shown by measuring the saccharide standard curves in IL solutions that the IL does not affect analysis results when the samples contain 0–20% (v/v) [EMIM]AcO.144 ILs can cause background absorption interference due to its own colour and apparently also interfere with the colour forming reaction by reacting with the DNS reagent, although it is not clear whether it is the IL itself or some impurity in it that causes the additional colour.152 ILs have also been reported to interfere with other spectroscopic techniques, such as circular dichroism (CD), used for studying conformational changes in the enzyme structure.158

Sugar derivatization with aromatic amines has been used in several studies as a means to increase the saccharide detectability in the presence of ILs in HPLC or CE.32,47,185 The increased selectivity of the detection is highly beneficial in reducing IL interference. Both the spectrophotometric assays and many of the derivatization methods are based on reactions with the reducing end of the saccharides. It has, however, been shown that the imidazolium cations in some ILs form carbenes which react with the reducing ends of the saccharides.199 If this reaction takes place to a significant extent, it will alter the analysis results. This reaction has not been studied in further detail in IL-containing hydrolysates. It is surprising how little the influence of ILs on saccharide analysis has been commented on in the literature, taking into account the large number of articles in which IL-containing hydrolysates have been analysed.

Summary and outlook

It is just over a decade since cellulose dissolution in ILs was first reported rapidly followed by the observation that cellulases are inactivated in ILs. In the recent literature, a more detailed picture has been drawn about the mechanisms and reasons for the apparent cellulase inactivation, but several pieces are still missing from the puzzle. Deeper insights into the molecular scale mechanisms of cellulase inactivation in cellulose-dissolving ILs are still needed in order to be able to design fully compatible systems in which cellulases are truly active in cellulose hydrolysis in the presence of significant amounts of IL. Several groups have reported impressive results on the discovery and development of IL-tolerant cellulases. It should be mentioned that cellulase activity has been studied in other cellulose solvents, such as NMMO200 and deep eutectic solvents (DES),201 which are currently gaining considerable interest as an extension to the IL family. It appears that some of these alternative cellulose solvents display higher enzyme-compatibility than the cellulose-dissolving ILs.

The most important challenges in applying ILs in lignocellulose pretreatment for the biotechnological production of biofuels and -chemicals from lignocellulose are now in finding economical solutions to IL recycling and studying their effects on health and the environment, which so far has been much overlooked. Techno-economic evaluations have shown that co-product streams need to be obtained from the hydrolysis step in addition to the glucose main product for an IL-based biorefinery to be economically viable.59 Many lignocellulose-hydrolysing enzymes have recently been reported to have high IL-tolerance, but so far the general cellulolytic activity of these enzymes has scarcely been compared. Having high IL-tolerance is not sufficient for application in industrial processes, if the enzyme itself has low catalytic activity, or the enzymes are difficult to produce with a similar cost efficiency as the traditional cellulase preparations. A logical next step in the development of IL-tolerant glycosyl hydrolases is to increase their general activity while maintaining their tolerance to IL, and in the long run, to find good production hosts. In addition to the impressive advances made in finding IL-tolerant cellulases for the one-pot hydrolysis, advances have also been made in developing the IL pretreatment technology, e.g. in introducing pretreatment methods with very high biomass loadings and using aqueous ILs instead of dry ILs. The recent advances in this field of biomass processing are highly promising for the future design of biorefinery processes with IL technology.
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Notes and references

Critical Review


