Protein synthesis in artificial cells: using compartmentalisation for spatial organisation in vesicle bioreactors†

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We have previously developed methods to construct vesicles with defined compartment number, content and size, where compartments were separated from one another via a lipid bilayer.‡ We also used such multi-compartment vesicles as micro-reactors that hosted a multi-step reaction cascade, mimicking a cellular signalling pathway.† This work builds on this: we demonstrate protein synthesis using in vitro transcription and translation (IVTT), using the separation of content to achieve segregation of function. We show for the first time that each compartment can be devoted to the biochemical synthesis of distinct biomolecules—green fluorescent protein (GFP) being synthesised in one compartment, red fluorescent protein (RFP) in another (Fig. 1). In doing so we succeed in using compartmentalisation to introduce a level of spatial organisation of function that has not been previously seen in artificial cells.

For cell-free protein expression we used the PURExpress14 IVTT kit that contained the biochemical components necessary to synthesise proteins in the presence of a plasmid. The reaction mix was made by adding the kit components to plasmids encoding for fluorescent proteins, as well as an added volume to yield a final solution composition of 0.9 M sucrose and 9 mM 1,2-diphytanoyl-sn-glycero-3-phosphocholine (DPhPC) liposomes (see ESI† for experimental details). Two such solutions were made: one containing Dasher-GFP plasmid, the other an RFP plasmid.

Two-compartment vesicles were generated according to previously published methods, using the phase transfer of two water-in-oil droplets (Fig. 1 and ESI† for details).‡,† Briefly, a water/oil column was first made, with a 4.7 mM DPhPC lipid-in-oil solution. This was left to stabilise for two hours, resulting in a densely packed monolayer at the water/oil interface. Two sucrose-loaded lipid-stabilised droplets were then expelled above the same location of the column: one droplet containing the GFP plasmid, the other the RFP plasmid. As these droplets descended through the column under gravity, they made contact with another resulting in a droplet interface bilayer. The droplet pair entered the aqueous phase of the column, they were encapsulated by a second monolayer, encasing both droplets together in a bilayer, and resulting in dual-compartment vesicles with defined

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content in each compartment. Immediately after vesicles were generated, 1.5 M sucrose was injected into the bottom of the aqueous phase to reduce the osmotic imbalance between the vesicle interior and exterior.

Protein synthesis was followed over time by monitoring fluorescence using a FITC filter for GFP and a TRITC filter for RFP at 100 ms and 1000 ms exposures respectively (Fig. 2). Over the course of 50 minutes at 37°C, fluorescence of both compartments was observed to increase concurrently, indicating successful spatially segregated protein synthesis: different regions of the vesicle were specialised to perform distinct biological processes, acting as modules within the artificial cell system. Furthermore, as biological macromolecules are largely impermeable to membranes, the process remained spatially isolated over the course of the experiment.

Vesicles were found to be formed with a lower efficiency than those previously reported, with approximately 12% successful rate of formation ($n = 50$). This was attributed to the presence of a relatively large concentration of macromolecules (estimated at 2.6 mg ml$^{-1}$ by the supplier), which are thought to interfere with the formation of a well-packed monolayer at the water/oil interface of the droplet precursors. Friddin et al. observed the same effect in a droplet interface bilayer system and specifically attributed it to the energy supply fraction of the IVTT kit,15 which destabilised the interface. In a similar manner to our experiments, they (and others)16 counteracting this by adding liposomes inside the droplets to increase the probability of sufficient monolayer coverage.

On the occasions where phase transfer was unsuccessful, vesicles were observed to degrade in one of two ways (discussed in detail elsewhere).12 Either the inner bilayer failed, leading to fusion of the two compartments, or the external bilayer failed whereupon the compartment content was released to the external solution.

The above IVTT experiments had to be carried out in different conditions to those commonly used and recommended, specifically in the presence of sucrose and lipids. To test the effect of...
Finally, in this work we extend the concept of soft-matter compartmentalisation to the field of artificial cells. Compartmentalisation is a recurring feature in chemical, biological, and engineering systems. It refers to the splitting-up of a unit into distinct spatial sub-divisions, each one considered a module designed separately and optimised to perform a designated task, often in different chemical environments. This enables multi-part, higher-order, or emergent features to arise, and leads to more efficient systems. Our approach differs from bulk self-assembly approaches with respect to the greater degree of control over vesicle properties (compartment content, number, and size) and that only one bilayer separates compartments from one another. There is therefore scope for engineering communication routes between compartments, for example by the insertion of transmembrane proteins or DNA origami pores.

Conclusions

We generated two-compartment vesicles, with two ‘genomes’ contained in each compartment. Using compartmentalisation as an engineering principle allowed protein synthesis of distinct proteins to occur in defined regions, and demonstrated the spatial segregation of complex processes in a vesicle-based artificial cell. This has implications in microreactor and medicinal settings, for instance for isolated on-board synthesis multiple therapeutic biomolecules in a cell-like drug-delivery vehicle, and can be considered an enabling technology for bottom up synthetic biology. Finally, the developed system extends the concept of modularisation that is central to synthetic biology. Instead of applying to genetic circuits, here it pertains to modular processes that take place within individual compartments of an artificial cell. This paves the way higher-order cellular characteristics to be introduced in future.

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Notes and references