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Simultaneous cellulose conversion and hydrogen production assisted by cellulose decomposition under UV-light photocatalysis†

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Photocatalytic conversion of cellulose to sugars and carbon dioxide with simultaneous production of hydrogen assisted by cellulose decomposition under UV or solar light irradiation was achieved upon immobilization of cellulose onto a TiO₂ photocatalyst. This approach enables production of hydrogen from water without using valuable sacrificial agents, and provides the possibility for recovering sugars as liquid fuels.

Economic and efficient methods for production of hydrogen, especially those that can generate hydrogen utilizing renewable energy resources (*e.g.*, water, solar and biomass), are currently the subject of extensive research. Solar hydrogen production from water splitting *via* a photocatalytic or photoelectrochemical process is considered as one of the potentially promising methods.¹ Although photocatalytic hydrogen production from water splitting has been demonstrated a few decades ago,² photocatalysts that efficiently generate hydrogen from pure water (without sacrificial agents) have not been extensively reported.^{3–5} In most cases, the photocatalytic hydrogen production from water is performed with an excess of electron donors such as alcohols, sugars and organic acids/amines.^{6–10} A more practical and economical case for hydrogen production, however, would not involve the use of such valuable chemicals as the electron donors.¹¹

Lignocellulosic material as the most abundant component of biomass is being considered as a promising renewable resource for sustainable hydrogen production. Cellulosic biomass can be converted into hydrogen and other valuable products through several thermochemical processes such as steam gasification, fast pyrolysis and hydrolysis by acids or enzymes.¹² Because of the rigid intra- and inter-molecular hydrogen bonds in cellulose, the conversion of cellulose to platform molecules (*e.g.* sugars) for preparation of liquid

fuels and high-value chemicals normally requires extremely harsh reaction conditions (*e.g.* high temperature and/or high pressure).^{13,14} On the other hand, the separation and purification of the valuable bio-products from the reaction medium is rather difficult, which is still an issue to be resolved for the practical application.

If cellulose could be photocatalytically decomposed into the platform molecules, the valuable bio-products for liquid fuels production can be *in situ* harvested with the application of appropriate reactor engineering. In the meantime, it is possible to obtain hydrogen during the cellulose decomposition process, by utilizing the cellulose and the as-generated intermediates as the sacrificial electron donors. The hydrogen production assisted by the cellulose decomposition may also be more practical and feasible compared to the photocatalytic water splitting.¹⁵ Only a few examples of the photocatalytic cellulose degradation have been reported,^{16–18} but the degradation of cellulose with the simultaneous generation of hydrogen is limited. In this work, we report the development of a relatively simple process for immobilizing cellulose onto a solid photocatalyst (*e.g.* TiO₂). Upon UV light irradiation, the simultaneous hydrogen production from pure water and efficient cellulose conversion to carbon dioxide *via* the formation of sugar intermediates was achieved as shown in Scheme S1 (ESI†). This approach has, hence, opened a new way of solar driven hydrogen production from water accompanied with the conversion of cellulose to valuable platform molecules.

Firstly, a cellulose colloidal aqueous suspension was prepared by two-step ball-milling (details see ESI†) of crystalline α -cellulose powder (Sigma-Aldrich, size *ca.* 10–1000 μm estimated from SEM image and particle size analysis (Fig. S1 and S2, ESI†)). The mechanochemical pre-treatment significantly cuts down the microcrystalline cellulose into sub-micron sized (*ca.* 0.1–3 μm) cellulose particles (Fig. S3, ESI†), which could be easily suspended in water (Fig. S4, ESI†). Upon the dispersion of platinized TiO₂ (P25, BET surface area $\sim 50 \text{ m}^2 \text{ g}^{-1}$, particle size $\sim 20\text{--}30 \text{ nm}$, loading of 0.5 wt% platinum for H₂ production, hereafter denoted as TiO₂(Pt)) photocatalyst nanoparticles (0.1 g) into a 250 ml aqueous solution filled with certain amount of the cellulose suspension stock (8 ml, 20 ml, 40 ml and 80 ml) by sonication

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and stirring, the cellulose@TiO₂(Pt) composite powder was obtained by filtering the mixed colloidal suspension. An excess of water was used to wash out the trace amounts of soluble sugars (glucose oligomers produced from ball-milling) that possibly remained on the mixed composite powder. Finally, the cellulose immobilized TiO₂(Pt) powder was obtained after drying the powder in air under room temperature, and denoted as TC1, TC2, TC3, and TC4 for the samples prepared from 8 ml, 20 ml, 40 ml and 80 ml cellulose suspension, respectively.

The facile “mixing and drying” based immobilization procedure and the possible reactions taking place between the cellulose and TiO₂ nanoparticles are illustrated in Scheme S2 (ESI[†]). It has been well documented that organic compounds with multiple hydroxyl groups (*e.g.* catechol, phenolic resins) can easily bind with TiO₂ nanoparticles *via* their hydroxyl groups and form surface “Ti–ligand” complexes.¹⁹ The cellulose nanoparticles, consisting of poly-hydroxyl groups on linked glucose units, are thus expected to be able to anchor with TiO₂ nanoparticles *via* the surface complexation. The ball-milled cellulose particles have very strong “self-polymerization” ability, which can be evidenced by the fact that the “self-polymerization” induced particle size growth of cellulose particles takes place in aqueous solution. Without mixing with TiO₂ nanoparticles, a white cellulose polymer flake as shown in Fig. S5 (ESI[†]) was obtained after drying the cellulose particles at room temperature. Therefore, the grafting of sub-micron sized cellulose particles onto TiO₂ nanoparticles can be reasonably ascribed to two pathways: condensation of surface hydroxyl groups and formation of intra-molecular H-bonds between the cellulose strands and TiO₂ as shown in Scheme S2 (ESI[†]).

The cellulose coating on the TiO₂(Pt) nanoparticles was initially confirmed by SEM and high resolution TEM (HRTEM) images. Cellulose coating can be clearly distinguished from the cellulose coated samples. A gradually increasing thickness of the cellulose layer on TiO₂(Pt) was observed on increasing the cellulose ratio from TC1 to TC4 (Fig. S6, ESI[†]). The representative TEM image of the TC4 sample (Fig. 1a) shows an amorphous carbon overlayer along the TiO₂(Pt) particles boundary, which is assigned to the cellulose loaded on the TiO₂(Pt) surface. The HRTEM images focusing on a single cellulose coated TiO₂ particles in the Fig. 1a panel clearly show a thin amorphous overlayer (*ca.* 3–7 nm)

corresponding to a few layers of cellulose strands coating on crystallized TiO₂(Pt) (Fig. 1b and c). No such amorphous layer, however, could be found on the bare TiO₂(Pt) particles (Fig. 1d).

The cellulose content on each immobilized sample (TC1 to TC4) was quantified by thermo-gravimetric differential thermal analysis (TG-DTA) and elemental analysis (CHN). Significant weight loss between 200–300 °C was observed due to the oxidation of cellulose in air from the TG-DTA analysis (Fig. S7 and S8, ESI[†]). A gradual increase in weight loss was observed for the samples along with the increased cellulose loading from TC1 to TC4. Trace amounts (<0.5%) of organic impurities on bare TiO₂(Pt) may result from the adsorption of organics during the photodeposition of platinum. The immobilized cellulose contents were determined *ca.* 3.0%, 6.5%, 13.0% and 20.5% according to the weight loss for sample TC1, TC2, TC3 and TC4, respectively. In addition, the cellulose contents of different samples from the CHN analysis were summarized in Table S1 (ESI[†]). Eventhough there are slight derivations of the CHN values and TGA values, the results are generally consistent, indicating the validity of our analysis on the cellulose contents.

The photocatalytic conversion of cellulose was carried out in a de-aerated aqueous suspension of cellulose immobilized TiO₂(Pt) samples (TC1–TC4) by suspending the powder samples in pure water under UV-light irradiation. The concurrent production of H₂ and CO₂ in gas phase and the soluble cellulose degradation products in aqueous phase were subsequently monitored. Cellulose immobilized bare TiO₂ (without Pt, denoted as cellulose@TiO₂), mixed cellulose and bare TiO₂ (cellulose + TiO₂), mixed cellulose and TiO₂(Pt) (cellulose + TiO₂(Pt)) and cellulose immobilized TiO₂(Pt) (cellulose@TiO₂(Pt)) samples with the same amounts of cellulose contents were compared for the photocatalytic H₂ and CO₂ production (Fig. 2). The evolved H₂ and CO₂ with the TiO₂(Pt) sample during the UV-Visible light irradiation was employed as control. The measured CO₂ content in the headspace of reactor was mainly due to the reduced solubility of CO₂ in water at elevated temperature (293 K to 313 K) during photoirradiation. This can be evidenced by the fact that a similar CO₂ production profile was observed during the repeated photoirradiation cycles (as shown in the following part). The CO₂ production from the photocatalytic mineralization of trace amount of organic impurities on TiO₂ should be negligible. In the presence of TC1 sample, the conversion of cellulose to CO₂ was accompanied by the notable production of H₂ compared to the control case, whereas the simultaneous production of H₂ and CO₂ was negligible for the other samples compared with the control. In the case of cellulose@TiO₂ sample, the CO₂ production from the cellulose degradation was also found, whereas the cellulose conversion efficiency was much lower than the TC1 sample. The negligible H₂ production, along with the lower CO₂ production efficiency, suggests the platinum co-catalyst is quite important for the simultaneous cellulose oxidation (enhanced e[−]/h⁺ pair separation) and the water reduction (as proton reduction sites). For the physically mixed cellulose and bare TiO₂ sample, there was no H₂ evolution nor CO₂ production from the cellulose conversion, indicating the direct photocatalysis between the cellulose and the TiO₂ was negligible. The activities of

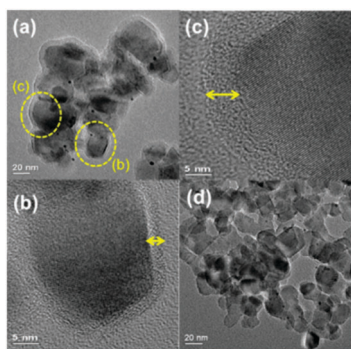


Fig. 1 TEM and HRTEM images of cellulose coated TiO₂ nanoparticles (a–c) and bare TiO₂(Pt) (d). The panel (b and c) are corresponding to selected areas in panel (a).



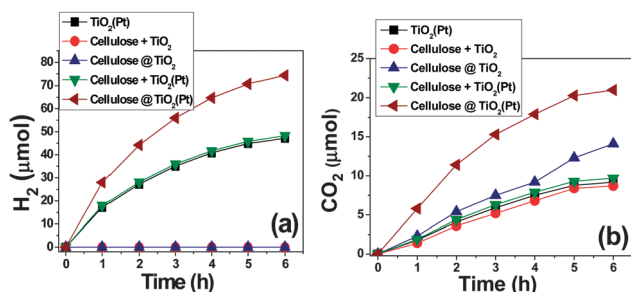


Fig. 2 Time profiles of (a) H_2 production and (b) CO_2 production from the suspension of $\text{TiO}_2(\text{Pt})$, mixed cellulose with TiO_2 and $\text{TiO}_2(\text{Pt})$, and cellulose immobilized TiO_2 and $\text{TiO}_2(\text{Pt})$ samples. Reaction conditions: 15 mg samples suspended in 50 ml de-aerated aqueous solution, UV-Visible light irradiation (iron-halide lamp), reaction temperature (293–313 K).

was not evolved in the photoreactions. All of these results demonstrate that the immobilization of cellulose onto $\text{TiO}_2(\text{Pt})$ is crucial for the efficient conversion of cellulose.

The photocatalytic production of H_2 and CO_2 is expected to gradually reduce as the immobilized cellulose is consumed. The time profiles of H_2 and CO_2 production were measured *via* the repeated cycles of photocatalysis with the cellulose immobilized TiO_2 samples and $\text{TiO}_2(\text{Pt})$ as a control. Each photocatalysis run was performed for 6 h in one day and then the suspension was purged with argon to remove accumulated H_2 and CO_2 before starting the second cycle. The photocatalysis experiment was repeated up to seven cycles until no significant H_2 and CO_2 were produced compared to the control. As shown in Fig. 3a and b, the production of H_2 and CO_2 from the cellulose immobilized TiO_2 samples in the first cycle was significantly higher than that from the control ($\text{TiO}_2(\text{Pt})$). However, the differences for the following cycles gradually decreased because of the depletion of cellulose. In the meantime, the post-tested aqueous solution was analyzed by HPLC after each cycle test. The main soluble

products were determined as cellobiose, glucose and formic acid according to the HPLC analysis of standard sugars and acids, while the other products such as cellobextrins, arabinose, erythrose and gluconic acid were negligible (Fig. S9 and S10, ESI†). Fig. 3c briefly summarizes the main soluble products that remaining in the reactor from the TC4 sample (as a representative) after each cycle test. The accumulated amount of glucose in each cycle was almost constant, whereas the cellobiose was gradually reduced in the first three cycles and disappeared from the fourth cycle. Only trace amounts of formic acid were detected in the first three cycles (Fig. S10, ESI†), while the concentration of formic acid gradually increased during the following cycles. Without the light irradiation, the production of gaseous products and organic intermediates were negligible, implying that the total products were generated due to the photocatalytic decomposition of cellulose and its degradation intermediates. Natural sunlight as light source for simultaneous H_2 production and cellulose conversion was also tested outdoors. As shown in Fig. 3d, the production rates of H_2 and CO_2 for TC4 were much higher than those of pure $\text{TiO}_2(\text{Pt})$, and they relatively remained constant during the three cycle tests. However, due to the weak solar irradiation intensity, the efficiencies of H_2 and CO_2 production for TC4 at outdoors were much lower than those obtained with iron-halide lamp source indoors. Without UV-light irradiation, no H_2 was evolved for the TC4 sample. (Fig. S11, ESI†) The solid samples after the repeated photocatalysis tests were further analyzed by SEM and TG-DTA. All the cellulose loaded TiO_2 samples expose a smooth and clear surface from the SEM images (Fig. S12, ESI†), and no significant amounts of cellulose were found by TG-DTA analysis (Fig. S13, ESI†), which demonstrated that all the cellulose loaded onto $\text{TiO}_2(\text{Pt})$ samples was nearly completely decomposed.

Table 1 summarizes the results of the photocatalytic generation of gaseous products (H_2 and CO_2) and liquid products (glucose and formic acid) after seven repeated cycles. The total accumulated H_2 and CO_2 produced in seven cycles after subtracting those produced from pure $\text{TiO}_2(\text{Pt})$ were 35.8, 72.5, 122.5, 195.2 μmol (H_2) and 16.7, 32.0, 53.8, 85.3 μmol (CO_2) for TC1, TC2, TC3 and TC4, respectively. In addition, the accumulated amounts of glucose (0.3–0.7 μmol) and formic acid (0.5–1.1 μmol) in aqueous phase were observed for all the cellulose immobilized samples. The total amounts of gaseous and liquid products were increased with more cellulose loading. The H_2 and CO_2 yields after seven cycles were *ca.* 80–90% and *ca.* 70–80% respectively, based on the assumption that complete conversion of glucose unit ($\text{C}_6\text{H}_{10}\text{O}_5$) on cellulose to H_2 and CO_2 .²⁰ The efficiencies of H_2 generation were slightly higher than those of CO_2 evolution, because the *in situ* generated intermediates might not be fully mineralized into CO_2 . This is also consistent with the results that the observed CO_2/H_2 ratios were *ca.* 0.45 for all the samples, which were slightly smaller than the theoretical value of 0.5.²⁰ Furthermore, increasing the immobilized cellulose ratio reduced the cellulose conversion efficiency, as the TC4 sample showed the lowest H_2 and CO_2 yields. This was also demonstrated by the comparison of total C-balance, which takes account of all generated C-containing products against the initial cellulose content. The TC1 sample exhibited the highest C-balance (93.4%), whereas the TC4 sample had the lowest one (75.1%).

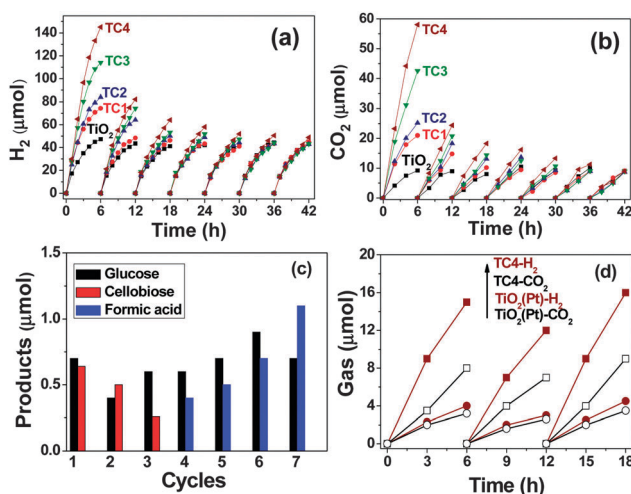


Fig. 3 Time profiles of (a) H_2 production and (b) CO_2 production from the suspension of $\text{TiO}_2(\text{Pt})$ and cellulose immobilized $\text{TiO}_2(\text{Pt})$ samples on repeated cycling. (c) HPLC analysis results on the production of cellobiose, glucose and formic acid from the TC4 sample during repeated cycles. (d) H_2 and CO_2 production from natural sunlight irradiation.



Table 1 Summarized results on the photocatalytic conversion of cellulose immobilized onto TiO₂ after seven photocatalytic cycles

Sample	Cellulose fraction (wt%)	(C ₆ O ₅ H ₁₀) unit content (μmol)	Products (gas phase) ^a		Products (aqueous phase) ^b					CO ₂ /H ₂ ^d	C-balance ^e (%)
			H ₂ (μmol)	CO ₂ (μmol)	C ₆ H ₁₂ O ₆ (μmol)	HCOOH (μmol)	H ₂ yield ^c (%)	CO ₂ yield ^c (%)			
TC1	3.0	3.3	35.8	16.7	0.3	Trace	89.5	83.5	0.47	93.4	
TC2	6.5	6.6	72.5	32.0	0.3	0.5	91.7	81.0	0.44	86.6	
TC3	13.0	12.1	122.5	53.8	0.5	0.7	84.5	74.2	0.44	79.2	
TC4	20.5	20.1	195.2	85.3	0.7	1.1	81.2	71.0	0.43	75.1	

Reaction conditions: 15 mg sample in 50 ml H₂O, UV light irradiation, reaction time: 6 h for each cycle, 7 cycles performed in total, reaction temperature, 293–313 K. ^a Total accumulated H₂ and CO₂ contents in 7 cycles subtract those generated from TiO₂ sample (control test). ^b Aqueous solution after photo-irradiation analyzed by HPLC. ^c Calculated based on the complete conversion of glucose unit (C₆O₅H₁₀) to H₂ and CO₂ (C₆O₅H₁₀ + 7H₂O = 12H₂ + 6CO₂). ^d Theoretical molar ratio of CO₂/H₂ is 0.5. ^e [(CO₂) + 6(C₆H₁₂O₆) + [HCOOH]]/[C₆O₅H₁₀].

Given that all the cellulose was decomposed, the lower C-balance suggests that more unknown soluble products (e.g. organic acids or sugars with C_n < 3) other than the standard sugar molecules detected by our HPLC were present in the aqueous solution.

Considering that the oxidation potentials of organic carbohydrates are more negative than the potential of water oxidation,²¹ thus it is more likely that the surface-bound cellulose were directly decomposed by the valence band holes, while the generated soluble intermediates in water could be further oxidized by holes or OH radicals. Eventually, all the organic products should be completely mineralized to CO₂ by the TiO₂ photocatalysis. However, the complete mineralization of the cellulose was not achieved and gradually reduced with the increased cellulose loading, which is probably due to the following reasons. Firstly, the anoxic conditions reduced the mineralization efficiency, as trace amounts of sugars and formic acid were still present after seven cycles. In addition, some unknown soluble products might be difficult to be further photo-oxidized because of the diffusion limit of free OH radicals in water.

The pathway of cellulose decomposition is supposed to be quite complex, as only trace amounts of cellobiose and glucose along with the formic acid, were found as the soluble intermediates in our work. Owing to the labile characteristics of the intermediates, the identification of each intermediate was rather difficult and a lot of *in situ* characterization tools are required. Nevertheless, the surface bound cellulose and the soluble products generated during the whole process could be used as the sacrificial agents to enhance the H₂ yields. The evolved H₂ might stem from both the water reduction and the decomposition of cellulose degradation products (e.g. formic acid), since H₂ production from photocatalytic degradation of formic acid has been proposed.²²

Because of the simplicity of the immobilizing process and the well-known stability of titania, the reproducibility of catalyst would not be an issue in this approach. The optimization of catalysts and reaction conditions, analysis of liquid products, degradation pathway of cellulose and long-term stability test will be further studied in detail. In summary, simultaneous H₂ production from photocatalytic water reduction and cellulose

conversion was achieved. This approach provides an opportunity for harvesting valuable sugars *via* the *in situ* separation of sugars before undergoing further oxidation, which would require the development of appropriate reactor engineering. It has also demonstrated the feasibility of recovering hydrogen fuel from pure water without using valuable sacrificial agents and simultaneous conversion of cellulose into platform molecules by photocatalysis.

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