



Cite this: *Chem. Commun.*, 2015, 51, 15858

Received 26th June 2015,
Accepted 4th September 2015

DOI: 10.1039/c5cc05258k

www.rsc.org/chemcomm

Interfacial thiol–isocyanate reactions for functional nanocarriers: a facile route towards tunable morphologies and hydrophilic payload encapsulation†

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Functional nanocarriers were synthesized using an *in situ* inverse miniemulsion polymerization employing thiol–isocyanate reactions at the droplet interface to encapsulate hydrophilic payloads. The morphology of the nanocarriers is conveniently tunable by varying the reaction conditions and the dispersions are easily transferable to the aqueous phase.

One of the major challenges in developing nanocarriers is their design towards the ability to encapsulate hydrophilic substances. The encapsulation of hydrophilic compounds is highly valuable but at the same time also a very demanding task¹ especially when the final purpose of the carriers is envisaged for an aqueous environment (*e.g.* the fields of drug delivery or bio-imaging to name a few).

Of the several heterophase polymerization techniques, miniemulsion has taken a fortified position as a versatile synthesis technique, allowing for effective encapsulation of both hydrophobic as well as hydrophilic compounds.^{2,3} It was previously shown that hydrophilic substances can be successfully encapsulated by using a nanoprecipitation process or by interfacial reactions such as step growth, radical or anionic polymerization in inverse miniemulsion.^{4,5}

The versatility of the thiol functionality to participate in different very efficient chemical reactions has seen tremendous focus recently for the generation of materials with interesting physical properties. In this regard, the thiol–ene click reaction⁶ has attracted significant interest owing to its high selectivity, efficiency and ability to use mild reaction conditions. Thiol–ene chemistry has – among other applications – been widely employed to obtain networks with tunable network properties.^{7,8} Lately, particles/capsules have also been reported based on this

type of chemistry.^{9–11} Another thiol reaction that holds high potential, but has not been fully explored despite the fact that it is known since the late 1950s,¹² is the thiol–isocyanate (thiol–NCO) conjugation; this reaction has to date remained largely unexploited especially for synthesizing nanomaterials. The nucleophilic addition of thiols to isocyanates yields thiourethane linkages. In the presence of a base catalyst, this equimolar reaction is facilitated through the generation of a strong nucleophilic thiolate ion and an electron deficient carbonyl carbon at the isocyanate. This reaction proceeds on a fast time scale (faster than classical urethane formation), at ambient temperature and affords high conversions. The reaction mechanism is given in Fig. 1. The thiourethane (–NH–CO–S–) functionality is a sulphur analogue of urethane (–NH–CO–O–) and the former's incorporation into the polymer chain is known to introduce interesting mechanical and thermal properties.^{13,14} Also, the high refractive index values of polythiourethanes make the latter highly appealing for optical and coating applications as compared to their polyurethane counterparts.^{15,16} Shin *et al.* made ternary networks using thiol–isocyanate–ene sequential/simultaneous click reactions where an

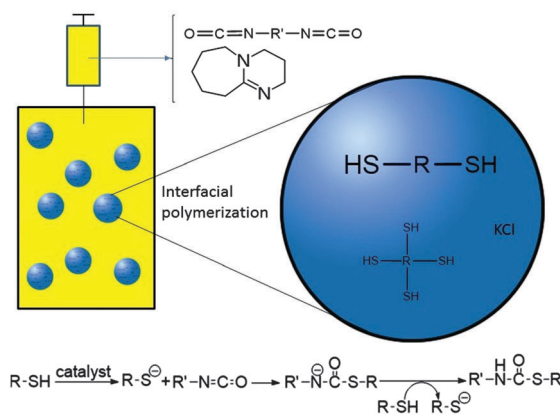


Fig. 1 Schematic representation of a base catalyzed interfacial polymerization reaction between diisocyanate and (tetra-)dithiol monomers in an inverse miniemulsion.

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† Electronic supplementary information (ESI) available: Materials, synthesis procedure, characterization methods, sample details, FT-IR spectra, TEM images and EDX spectra of particles, and biocompatibility studies. See DOI: 10.1039/c5cc05258k



amine catalyst was used to trigger the thiol–isocyanate reaction and ultraviolet (UV) light was used to trigger a radical thiol–ene reaction, resulting in a thiourethane/thiol–ene hybrid network.¹⁷ Matsushima *et al.* also used thiol–click chemistry to create thiol–isocyanate–acrylate ternary networks.¹⁸ Previously, thiol–NCO chemistry was used in a modular approach for a rapid and robust fabrication of highly functional, multicomponent polymer brush surfaces¹⁹ and lately for developing self-healing epoxy thermosets.²⁰ Recently, microparticles (40–250 μm) were synthesized using nucleophile-catalyzed thiol–NCO reaction in water employing microfluidics.²¹ With the recent revival of the thiol–NCO reaction towards interesting applications, the potential of this reaction towards designing nanomaterials with functional properties is highly desirable. Though polyaddition reactions between diisocyanates and diols/diamines have been extensively studied for nanocarrier formation, to the best of our knowledge, isocyanate–thiol interfacial reactions have not been reported to date. The latter reaction offers materials with different useful properties as compared to classical urethanes and will hence lead to novel functional nanocarriers.

In this work, for the first time the miniemulsion technique has been used for *in situ* thiol–NCO reactions at the droplet interface for the design of nanocarriers containing thiourethane linkages (Fig. 1). For encapsulation of a hydrophilic payload using inverse miniemulsion, a 1 M KCl solution was used as a model substance. Nanocarriers were formulated at room temperature using 1,8-diazabicycloundec-7-ene (DBU) as a base catalyst and employing different stoichiometric ratios of 1,4-butane-dithiol (BDT) and toluene diisocyanate (TDI) as the respective bi-functional monomers. In order to impart potential post-surface grafting possibilities to the nanocarriers, pentaerythritol tetra-3-mercaptopropionate (PETMP), a multifunctional crosslinker monomer, was also used in the reaction formulations. The detailed synthesis procedure is presented in the ESI.† Briefly, cyclohexane was used as the continuous phase and the dispersed phase employed consists of thiol monomer(s), DMSO, and KCl solution. After pre-emulsification followed by sonication, stable nanodroplets of the dispersed phase were obtained. Subsequently, an additive phase containing the catalyst and the TDI monomer in cyclohexane was slowly introduced into the continuous phase and then the reaction mixture was left for stirring.

As the reaction conditions have a significant effect on the nanocarrier morphology, the influence of the different reaction parameters – the amounts of monomers, the presence of a

crosslinker and a catalyst – on the resulting morphology was studied systematically. Since, the reaction in the absence of a catalyst at room temperature is not feasible, the same reactions were also repeated using elevated temperature (60 $^{\circ}\text{C}$) in order to study the effect of the catalyst on the morphology. For all formulations used, the stability of the resulting dispersions was first checked optically for any visible phase separation and samples were then directly used for further characterization. The colloidal stability, the size and polydispersity index (PDI) of the samples were studied using dynamic light scattering (DLS). The reaction efficiency was studied thermogravimetrically in the form of solid content of the obtained dispersion. Chemical analyses of the insoluble products were performed using high resolution solid state ^{13}C nuclear magnetic resonance (NMR) and Fourier-transform infrared (FT-IR) spectroscopy. As a proof-of-concept for the use of such nanocarriers in biomedical applications, biocompatibility studies were also performed. The dispersion characteristics of the different samples tested are presented in Table 1.

From the hydrodynamic diameters (intensity average), it can be seen that most of the synthesized nanocarriers have a size range between 165 and 225 nm. From the obtained values, a trend between the size and the amount of monomer used in the reaction can be observed; the size of the nanocarriers mostly increases with the increase in the monomer amount. It can also be seen that the polydispersity index (PDI) is higher when crosslinkers were employed for samples prepared using both a base catalyst as well as elevated temperature. The increased size and the PDI observed for the reactions performed in the presence of the crosslinker could be attributed to the structure of the tetrafunctional crosslinker, which is more bulky than the linear bi-functional monomer. The sizes observed here in general are comparable and similar to values reported for polyurethane capsules.²²

The experimental solid content here refers to the dispersion solid content containing only particulates, excluding any large aggregates/bulk materials formed during reaction. The theoretical solid content refers to the calculated dispersion solid content upon full conversion of all monomers added to the reaction. The solid contents measured are also in accordance with literature values reported for polyurethane capsules.²² The solid contents are higher when the monomer amounts are increased. This trend is as expected as higher amounts of starting reactants lead to an increased product formation. The overall success of the thiol–NCO reaction is determined by the presence of the thiourethane groups in the product. In Fig. 2, the NMR data obtained for selected

Table 1 Size, PDI, and solid content for nanocarriers synthesized

Sample ^a	Dispersed phase monomer(s)	Additive phase	Size (nm)/PDI	Expt. solid content (%)	Theo. solid content (%)
1	2 mmol BDT	2 mmol TDI, DBU	173/0.04	5.4	7.1
2	4 mmol BDT	4 mmol TDI, DBU	193/0.12	7.4	11.9
3	4 mmol BDT, 0.1 mmol PETMP	4.2 mmol TDI, DBU	200/0.14	11.8	13.7
4	4 mmol BDT, 0.2 mmol PETMP	4.4 mmol TDI, DBU	182/0.26	11.9	14.3
5	2 mmol BDT	2 mmol TDI	168/0.09	5.3	7.2
6	4 mmol BDT	4 mmol TDI	188/0.12	8.0	12.9
7	4 mmol BDT, 0.1 mmol PETMP	4.2 mmol TDI	221/0.19	13.1	13.6
8	4 mmol BDT, 0.2 mmol PETMP	4.4 mmol TDI	202/0.24	12.8	14.3

^a Samples 1–4 were prepared using a catalyst at room temperature, while, samples 5–8 were obtained at 60 $^{\circ}\text{C}$ (for details see the ESI).



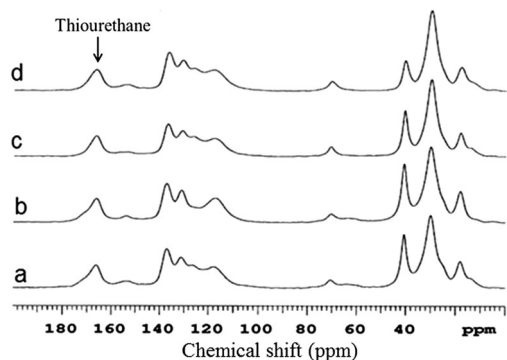


Fig. 2 High resolution solid state ^{13}C NMR data showing the presence of thiourethane for samples 4 (a), 8 (b), 2 (c) and 6 (d) (see Table 1).

samples are depicted. Aliphatic carbons are located in the range between 15 and 45 ppm. The methyl group originating from the diisocyanate is assigned to the peak at 18 ppm. The shoulders at 26 ppm and 64 ppm originate for the crosslinker. The peak at 70 ppm represents the alcohol and the ether functionalities of the surfactant; the latter is a block copolymer consisting of polyhydroxystearic acid and polyethylene glycol moieties.^{23,24} The peaks ranging from 140 ppm to 110 ppm originate from the aromatic ring of the diisocyanate used for polymerization. The small peak at 154 ppm corresponds to urea-urethane.²⁵ The formation of urea can be attributed to the reaction of isocyanate with amine groups formed in the first place by the hydrolysis of the highly reactive isocyanate with ambient water. Such an observation has been reported in the case of formation of polyurethane nanocapsules before.²⁶ Urethane formation is also possible due to the alcohol functionality present in the surfactant (see Fig. S6 in the ESI†). Which of the two reactions is predominating as side reaction is purely based on NMR is difficult to verify. Regardless, a well-defined intense peak at 166 ppm confirms the successful formation of thiourethane.²⁷ FT-IR analysis might aid in exact structure elucidation and spectra for the same selected samples as in Fig. 2 are presented in Fig. 3. The full spectral range for all samples in Table 1 (Fig. S7, ESI†), monomers (Fig. S2–S4, ESI†) and products from an analogue bulk reaction (Fig. S5, ESI†) are presented in the ESI†

The disappearance of the functional groups stemming from the monomers is depicted in Fig. S7 in the ESI† by the disappearance of

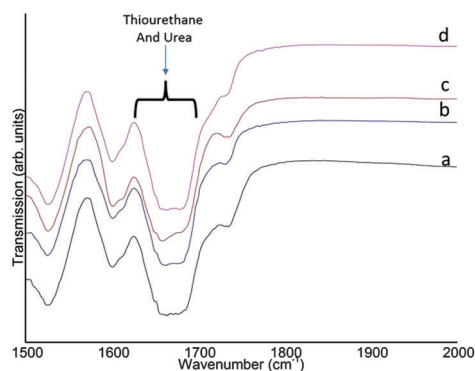


Fig. 3 Transmission FT-IR spectra showing the presence of thiourethanes for samples 4 (a), sample 8 (b), sample 2 (c) and sample 6 (d).

the characteristic isocyanate and thiol peaks (2280 cm^{-1} to 2270 cm^{-1} and 2556 cm^{-1} to 2400 cm^{-1} , respectively).¹⁹ The presence of the thiourethane peak at $\sim 1680\text{ cm}^{-1}$ in Fig. 3 indicates the successful reaction between the diisocyanate and the thiol moieties.^{27,28} It can be seen that the thiourethane peaks are broadened due to the presence of urea at $\sim 1640\text{ cm}^{-1}$.^{26,29} The urea is formed from the reaction between isocyanate end groups and water as indicated above. Since the reaction is performed under equimolar conditions and the signal of the diisocyanate monomers is no longer present in the FT-IR data, it can be assumed that the excess thiol groups present in the reaction medium have been oxidized to form disulphides.³⁰ Consequently, the presence of urethane at $\sim 1700\text{ cm}^{-1}$ ^{26,28,29} could not be ascertained by the FT-IR data. This is likely due to only trace amounts of urethane being formed owing to the small amount of surfactants used (see Fig. S6 in the ESI†). In addition, as the amines are more reactive, the consumption of isocyanates for urea formation might also affect the urethane formation.²⁷ Therefore, the signal at 154 ppm in the NMR spectrum is in all likelihood mainly due to the formation of urea rather than urethane moieties. The NMR results together with the FT-IR data thus clarify the presence of thiourethane-urea linkages in the samples.

The nanocarrier morphology was studied using transmission electron microscopy (TEM). The size and size distribution (qualitatively) observed in the TEM images (Fig. 4) for all samples are in agreement with the size and polydispersity measured using DLS. As it can be seen in Fig. 4a in the case of base catalysed reactions, sample 1 (without a crosslinker) gave a homogeneous bulk nanoparticle morphology. While sample 4 (Fig. 4b) (with a crosslinker) clearly afforded a core-shell nanocapsule morphology (for sample 3 see Fig. S8 in the ESI†). The reaction using elevated temperatures conducted in the absence of the catalyst and without a crosslinker (sample 6) yielded nanoparticles (Fig. 4c). In stark contrast to samples 3 and 4, in Fig. 4d (sample 8) the addition of the crosslinker to the elevated temperature reaction also resulted in nanoparticles

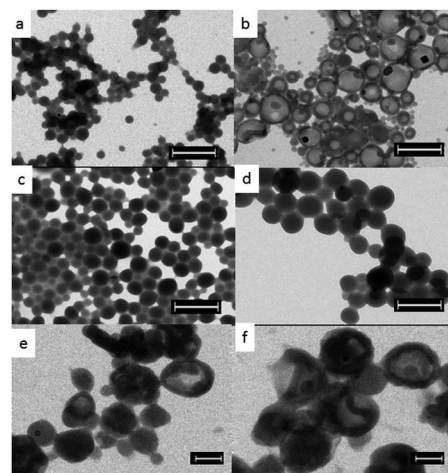


Fig. 4 TEM images of samples 1 (a), 4 (b), 6 (c) and 8 (d) from the organic phase (scale bar 500 nm). Images (a) and (b) are of samples employing base catalyst, while images (c) and (d) are of samples prepared at elevated temperature. Images e and f (scale bar 200 nm) are of samples 3 and 4, respectively, after redispersion in water.



(for sample 7 see Fig. S9 in the ESI†). In the case of nanocapsules the average shell thicknesses are 37.2 ± 7 nm and 51 ± 8 nm for samples 3 and 4 respectively. With increasing amount of cross-linkers the shell-thickness increases and also the size distribution (see Fig. S10 in the ESI†) becomes relatively broader reflecting the trend in DLS results. The successful encapsulation of KCl solution was confirmed by the presence of dark spots (salt crystals) within the capsules in the TEM images and was also confirmed by energy-dispersive X-ray spectroscopy (EDX) (see Fig. S11 in the ESI†).

To test the potential of these nanocarriers for biomedical applications, the synthesized nanocapsules have been transferred from the oil phase to a water phase. The redispersed aqueous dispersions were colloidally stable and the hydrodynamic diameters measured using DLS were 267 nm with a PDI value of 0.186 and 297 nm with a PDI value of 0.177 for samples 3 and 4 respectively. The significant increase in size after redispersion can be attributed to the pronounced hydration of the hydrophilic part of the block copolymeric surfactant in the aqueous phase. Using the TEM studies (Fig. 4e and f), the capsule morphology for samples with different cross-linker amounts can be clearly seen where the shell remains intact and evidently unaffected by the redispersion process. Using the EDX studies (see Fig. S12 in the ESI†), the presence of salt crystals (seen as dark spots within the capsules) was ascertained. Additionally, there was no sign of large aggregates of the nanocarriers (broken capsule debris) in the overview inspection of the TEM grid. Thus, DLS measurements and TEM characterization unambiguously confirm that aqueous dispersion of nanocapsules containing hydrophilic payload can be achieved successfully. Additionally, biocompatibility studies using Alamar blue assay performed on the redispersed samples (see Fig. S13 in the ESI†) clearly indicate that the nanocarriers are fully biocompatible thereby making them excellent candidates for biomedical applications. As a proof-of-concept, doxorubicin, a potent anticancer drug was encapsulated (see Fig. S14 in the ESI†).

For the first time thiourethane-based nanocarriers encapsulating hydrophilic substances have been synthesized *via* the inverse mini-emulsion technique using an *in situ* thiol-NCO reaction at the droplet interface. The presence of thiourethane functionality was confirmed by FT-IR spectroscopy and high resolution ^{13}C solid-state NMR spectroscopy. Also, side reactions leading to urea formation were ascertained by FT-IR. The morphology of the nanocarriers could be conveniently tuned by adjusting the reaction conditions as confirmed by TEM imaging. The presence of catalyst allows for the formation of tunable morphologies depending on the choice of the monomers used (*i.e.* with or without a crosslinker). The tunability of morphology offers designing nanocarriers for the desired purpose. For instance, nanocapsule morphology (with crosslinker) is suited for applications where a large aqueous core to polymer ratio encapsulating a large payload is required and the bulk particle morphology (without crosslinker) for applications envisaging high polymer content. The nanocapsules were also conveniently transferred to an aqueous phase while keeping their shell intact and were also successfully tested for their biocompatibility. With a suitable choice of the monomers, surface functionalization using the versatile thiol and isocyanate groups will be feasible. The shell is currently being modified to impart biodegradability by varying the monomer

choices and to functionalize using post-modification steps (*via* the tetrafunctional thiol moiety). The shells will be further studied for their physical properties and any possible leakage. In concise, we could successfully demonstrate that thiol-NCO reaction at the interface opens new possibilities for designing functional nanocarriers for biomedical applications.

The authors thank BOF funding of Hasselt University and the support from BELSPO *via* IAP Programme (P7/05). The authors are thankful to Prof. M. Van Bael for the access to the DLS device. A.E. is a FWO (Fonds Wetenschappelijk Onderzoek) postdoctoral fellow. M.P. is funded by “Agency for Innovation by Science and Technology” in Flanders (IWT).

Notes and references

- 1 S. Vrignaud, J. P. Benoit and P. Saulnier, *Biomaterials*, 2011, **32**, 8593–8604.
- 2 K. Landfester, *Angew. Chem., Int. Ed.*, 2009, **48**, 4488–4507.
- 3 A. Ethirajan and K. Landfester, *Chem. – Eur. J.*, 2010, **16**, 9398–9412.
- 4 M. Antonietti and K. Landfester, *Prog. Polym. Sci.*, 2002, **27**, 689–757.
- 5 J. M. Siebert, G. Baier, A. Musyanovych and K. Landfester, *Chem. Commun.*, 2012, **48**, 5470–5472.
- 6 C. E. Hoyle, A. B. Lowe and C. N. Bowman, *Chem. Soc. Rev.*, 2010, **39**, 1355–1387.
- 7 N. Zaquen, B. Wenn, K. Ranieri, J. Vandenberg and T. Junkers, *J. Polym. Sci., Part A: Polym. Chem.*, 2014, **52**, 178–187.
- 8 J. Vandenberg, K. Ranieri and T. Junkers, *Macromol. Chem. Phys.*, 2012, **213**, 2611–2617.
- 9 D. V. Amato, D. N. Amato, A. S. Flynt and D. L. Patton, *Polym. Chem.*, 2015, **6**, 5625–5632.
- 10 C. Fleischmann, J. Gopez, P. Lundberg, H. Ritter, K. L. Killops, C. J. Hawker and D. Klinger, *Polym. Chem.*, 2015, **6**, 2029–2037.
- 11 U. Paiphansiri, G. Baier, A. Kreyes, D. Yiamsawas, K. Koynov, A. Musyanovych and K. Landfester, *Macromol. Chem. Phys.*, 2014, **215**, 2457–2462.
- 12 R. G. Arnold, J. A. Nelson and J. J. Verbanc, *Chem. Rev.*, 1957, **57**, 47–76.
- 13 E. Dyer and D. W. Osborne, *J. Polym. Sci.*, 1960, **47**, 361–371.
- 14 O. D. McNair, D. P. Brent, B. J. Sparks, D. L. Patton and D. A. Savin, *ACS Appl. Mater. Interfaces*, 2014, **6**, 6088–6097.
- 15 A. Kultys, M. Rogulska and S. Pikus, *J. Polym. Sci., Part A: Polym. Chem.*, 2008, **46**, 1770–1782.
- 16 S. S. Naik, J. W. Chan, C. Comer, C. E. Hoyle and D. A. Savin, *Polym. Chem.*, 2011, **2**, 303–305.
- 17 J. Shin, H. Matsushima, C. M. Comer, C. N. Bowman and C. E. Hoyle, *Chem. Mater.*, 2010, **22**, 2616–2625.
- 18 H. Matsushima, J. Shin, C. N. Bowman and C. E. Hoyle, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 3255–3264.
- 19 R. M. Hensarling, S. B. Rahane, A. P. LeBlanc, B. J. Sparks, E. M. White, J. Locklin and D. L. Patton, *Polym. Chem.*, 2011, **2**, 88–90.
- 20 X. K. D. Hillewaere, R. F. A. Teixeira, L.-T. T. Nguyen, J. A. Ramos, H. Rahier and F. E. Du Prez, *Adv. Funct. Mater.*, 2014, **24**, 5575–5583.
- 21 J. Tan, C. Li, H. Li, H. Zhang, J. Gu, B. Zhang, H. Zhang and Q. Zhang, *Polym. Chem.*, 2015, **6**, 4366–4373.
- 22 E.-M. Rosenbauer, M. Wagner, A. Musyanovych and K. Landfester, *Macromolecules*, 2010, **43**, 5083–5093.
- 23 N. Yasarawan and J. S. van Duijneveldt, *Soft Matter*, 2010, **6**, 353–362.
- 24 E. Lovelady, S. D. Kimmins, J. Wu and N. R. Cameron, *Polym. Chem.*, 2011, **2**, 559–562.
- 25 S. M. Clift, J. Griminger, K. Muha, presented in part at the SPI Conference, 1994.
- 26 U. Paiphansiri, J. Dausend, A. Musyanovych, V. Mailänder and K. Landfester, *Macromol. Biosci.*, 2009, **9**, 575–584.
- 27 J. D. Flores, J. Shin, C. E. Hoyle and C. L. McCormick, *Polym. Chem.*, 2010, **1**, 213–220.
- 28 J. F. Smith and E. C. Friedrich, *J. Am. Chem. Soc.*, 1959, **81**, 161–163.
- 29 F. Tiarks, K. Landfester and M. Antonietti, *J. Polym. Sci., Part A: Polym. Chem.*, 2001, **39**, 2520–2524.
- 30 V. Dritsa, K. Pissaridi, E. Koutoulakis, I. Mamarelis, C. Kotoulas and J. Anastassopoulou, *In Vivo*, 2014, **28**, 91–98.

