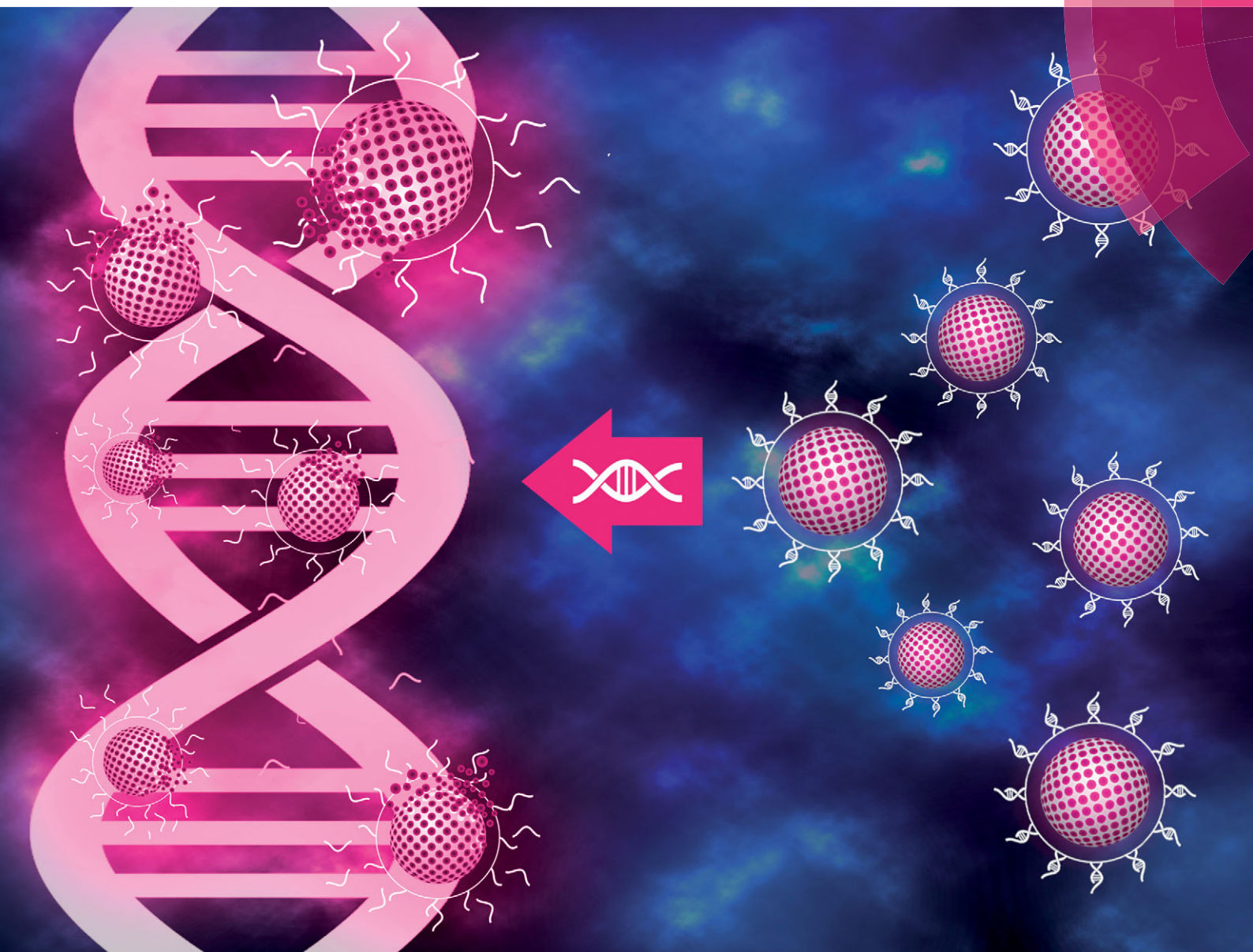


# ChemComm

Chemical Communications

[www.rsc.org/chemcomm](http://www.rsc.org/chemcomm)



ISSN 1359-7345



**COMMUNICATION**

Ramón Martínez-Máñez *et al.*

Oligonucleotide-capped mesoporous silica nanoparticles as  
DNA-responsive dye delivery systems for genomic DNA detection



Cite this: *Chem. Commun.*, 2015, 51, 1414

Received 21st October 2014,  
Accepted 18th November 2014

DOI: 10.1039/c4cc08306g

www.rsc.org/chemcomm

# Oligonucleotide-capped mesoporous silica nanoparticles as DNA-responsive dye delivery systems for genomic DNA detection†

Lluís Pascual,<sup>abc</sup> Isabel Baroja,<sup>abc</sup> Elena Aznar,<sup>ac</sup> Félix Sancenón,<sup>abc</sup>  
M. Dolores Marcos,<sup>abc</sup> Jose Ramón Murguía,<sup>abc</sup> Pedro Amorós,<sup>d</sup> Knut Rurack<sup>e</sup> and  
Ramón Martínez-Mañez<sup>\*abc</sup>

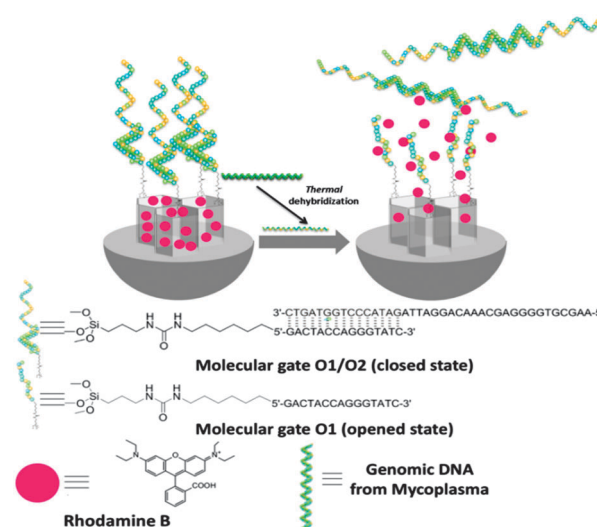
**New hybrid oligonucleotide-capped mesoporous silica nanoparticles able to detect genomic DNA were designed.**

Nanodevices equipped with molecular gates based on mesoporous silica nanoparticles have proven their potential for use in biological and medical applications.<sup>1</sup> So far, capped materials have been mainly used in drug delivery and their use in sensing protocols is still scarce.<sup>2</sup> For sensing, the mesoporous support is loaded with an indicator before the material is capped with a responsive chemistry. Uncapping and cargo delivery is then selectively achieved only in the presence of a target analyte.<sup>3</sup> In this context we are interested in the use of DNA sequences for the design of gated materials and others and we have recently reported the use of oligonucleotides as caps on mesoporous supports in the context of the detection of biomolecules *via* delivery of optical or electro-active probes.<sup>4</sup> However, in this context, the detection of genomic DNA using capped materials is still very rare.

Inspired by these preliminary studies we report herein a new hybrid gated material with a covalently attached double-stranded DNA (dsDNA) sequence that selectively delivers a dye in the presence of genomic DNA. As a proof of concept the detection of *Mycoplasma* genomic DNA was pursued in this work. Detection of bacteria of genus *Mycoplasma* is of interest in diverse areas. *Mycoplasma* contamination is a common problem in cell culture laboratories of research institutions and hospitals.<sup>5</sup> Moreover, several *Mycoplasma*

species are related to the development of certain pathologies such as pneumonia, rheumatoid arthritis and non-gonococcal urogenital diseases.<sup>6</sup>

The approach we followed is depicted in Scheme 1. MCM-41 mesoporous silica nanoparticles of *ca.* 100 nm diameter were selected as inorganic supports because of their unique and advantageous properties for the design of capped materials. The MCM-41 support was first loaded with the dye rhodamine B and then the external surface was functionalized with (3-isocyanatopropyl)triethoxysilane, yielding solid **S1-I**. For the gating mechanism two single stranded oligonucleotides were selected; *i.e.* (i) a short DNA sequence, functionalized with an aminohexyl moiety at the 5'-end position (NH<sub>2</sub>-(CH<sub>2</sub>)<sub>6</sub>-5'-GAC TAC CAG GGT ATC-3', **O1**) that could be covalently attached to solid **S1-I** *via* the formation of urea bonds and (ii) a single stranded oligonucleotide (5'-AAG CGT GGG GAG CAA ACA GGA TTA GAT ACC CTG GTA GTC-3', **O2**)



**Scheme 1** Representation of the gated material **S1-O1/O2** capped with a dsDNA. Delivery of the entrapped dye (rhodamine B) is selectively accomplished in the presence of a single strand of *Mycoplasma* genomic DNA.

<sup>a</sup> Centro de Reconocimiento Molecular y Desarrollo Tecnológico (IDM),  
Unidad Mixta Universidad Politécnica de Valencia-Universidad de Valencia, Spain

<sup>b</sup> Departamento de Química, Universidad Politécnica de Valencia,  
Camino de Vera s/n, 46022, Valencia, Spain. E-mail: rmaez@qim.upv.es

<sup>c</sup> CIBER de Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN), Spain

<sup>d</sup> Institut de Ciència dels Materials (ICMUV), Universitat de València,  
P.O. Box 2085, 46071 Valencia, Spain

<sup>e</sup> Fachbereich 1.9 Sensormaterialien, Bundesanstalt für Materialforschung und  
-prüfung (BAM), Richard-Willstätter-Straße 11, 12489 Berlin, Germany

† Electronic supplementary information (ESI) available: Synthesis of **S1-I** and **S1-O1/O2** materials, characterization of the prepared solids, and release experiments with solid **S1-O1/O2**. See DOI: 10.1039/c4cc08306g



which is a highly conserved sequence of the 16S ribosomal subunit of the *Mycoplasma* species genome.

In a typical experiment the two single stranded DNA sequences (**O1** and **O2**) were first hybridized and then covalently attached to the surface of the **S1-I** solid, yielding the final material **S1-O1/O2**. The dsDNA, anchored on the mesoporous support, was expected to be bulky enough to block the pores and inhibit dye delivery. Moreover it was anticipated that the presence of a single strand of *Mycoplasma* genomic DNA would displace **O2** from the solid with the subsequent pore opening and dye release.

MCM-41 nanoparticles were obtained according to reported procedures. Solid **S1-I** was first loaded with rhodamine B in acetonitrile and then an excess of (3-isocyanatopropyl)triethoxysilane was added to the suspension. Solid **S1-I** was collected by centrifugation, washed with acetonitrile and dried (see ESI† for details). For the preparation of the final capped material (*i.e.* **S1-O1/O2**) **O1** and **O2** were first mixed, heated at 90 °C and then gradually cooled to achieve the hybridization of both oligonucleotides. Then the hybridized **O1/O2** sequences were anchored onto the outer surface of the **S1-I** solid through the formation of a urea bond by reaction of the amine group in **O1/O2** and the isocyanatopropyl groups attached on **S1-I** (see ESI† for further details).

The starting MCM-41-like calcined nanoparticles and **S1-I** were characterized by powder X-ray diffraction, TEM, porosimetry, thermogravimetry and elemental analysis (see ESI† for details). Table 1 summarizes the particle diameter, BET specific surface area, pore volumes and pore sizes obtained for the starting nanoparticles and for solid **S1-I**. Moreover, from elemental and thermogravimetric analyses contents of 0.469 and 0.358 mmol g<sup>-1</sup> for isocyanatopropyl moieties and rhodamine B, respectively, were determined in solid **S1-I**. Finally, dynamic light scattering studies in water gave diameters for the MCM-41, **S1-I** and **S1-O1/O2** nanoparticles of 88 ± 2, 114 ± 3 and 154 ± 2 nm, respectively.

The response of **S1-O1/O2** was tested in the presence of *Mycoplasma fermentans* genomic DNA by measuring the emission of rhodamine B dye delivered from the pores of the solid. In a typical experiment 0.2 mg of **S1-O1/O2** were suspended in 400 µL of hybridization buffer (20 mM Tris-HCl, 37.5 MgCl<sub>2</sub>, pH 7.5) and separated into 2 aliquots of 200 µL. Both samples were filled to a final volume of 900 µL with hybridization buffer. At the same time a solution of 4000 copies µL<sup>-1</sup> of *Mycoplasma fermentans* quantification standard in water was heated to 95 °C for 5 min in order to dehybridize the double helix structure and then cooled in an ice bath for 3 min. After cooling, 100 µL of the dehybridized genomic solution was added to one of the aliquots whereas 100 µL of water (that was subjected to the same thermal treatment) were added to the other aliquot. Both suspensions were maintained at 25 °C

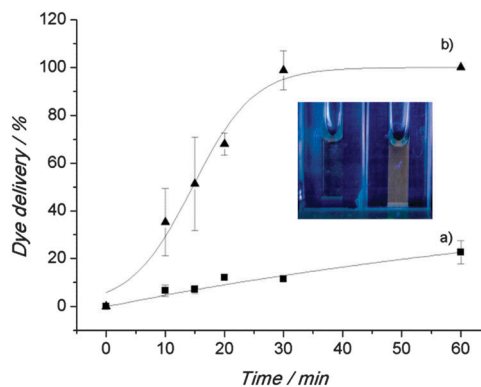


Fig. 1 Release of rhodamine B from solid **S1-O1/O2** in (a) the absence and (b) in the presence of *Mycoplasma fermentans* genomic DNA (4000 copies µL<sup>-1</sup>) in Tris-HCl buffer at pH 7.5. The inset shows the emission changes (under UV lamp, excitation at 254 nm) of the rhodamine B released from solid **S1-O1/O2** in the absence (left) and in the presence (right) of *Mycoplasma fermentans* genomic DNA (4000 copies µL<sup>-1</sup>).

and, at certain times, fractions were taken and centrifuged to eliminate the solid. Cargo release to the solution was then measured by the rhodamine B fluorescence at 575 nm ( $\lambda_{\text{exc}} = 555$  nm). Fig. 1 shows the delivery profile of rhodamine B from solid **S1-O1/O2** in the presence and in the absence of *Mycoplasma fermentans* genomic DNA. As it can be seen, in the absence of the target DNA (Fig. 1, curve a) a poor rhodamine B delivery was observed (less than 15% after 1 h) indicative of a remarkable pore closure. In contrast, in the presence of *Mycoplasma* genomic DNA, the pores were opened due to the displacement of oligonucleotide **O2** from the solid **S1-O1/O2** with the subsequent delivery of the dye (Fig. 1, curve b). Under these conditions *ca.* 90% of the maximum delivery of rhodamine B was observed after 30 min.

The high performance of solid **S1-O1/O2** pointed to the effective anchoring of the **O1/O2** dsDNA on the external surface of the inorganic scaffold through the formation of urea bonds. Additional experimental evidence of the correct grafting arose from the fact that solid **S1-O1/O2** was able to deliver rhodamine B in the presence of urease, due to the enzyme-induced hydrolysis of this bond with the subsequent detachment of the **O1/O2** double strand (see ESI†). Moreover, in the presence of DNase I, a remarkable dye release was also observed indicating the crucial role played by the **O1/O2** sequence in the capping protocol (see ESI†).

The next step was to assess the sensitivity of **S1-O1/O2** nanoparticles towards genomic DNA with the determination of the limit of detection. Following a procedure similar to that described above various solutions with different concentrations of *Mycoplasma fermentans* genomic DNA were added to several suspensions of **S1-O1/O2** and the rhodamine B released after 30 min was measured (see ESI†). A correlation between the number of copies of *Mycoplasma* genomic DNA added and the rhodamine B delivered was found in agreement with an uncapping process related to the displacement of **O2** from the **S1-O1/O2** solid. A limit of detection of *ca.* 70 DNA copies µL<sup>-1</sup> was determined (see ESI†). The limit of detection following this simple approach is not far from those reported using

Table 1 Main structural properties calculated from TEM and N<sub>2</sub> sorption analysis

Sample	Diameter particle (nm)	$S_{\text{BET}}$ (m <sup>2</sup> g <sup>-1</sup> )	Pore volume <sup>a</sup> (cm <sup>3</sup> g <sup>-1</sup> )	Pore size <sup>a</sup> (nm)
MCM-41	80 ± 4	1228.2	0.93	2.40
<b>S1-I</b>	80 ± 3	524.9	0.34	2.40

<sup>a</sup> BJH model.





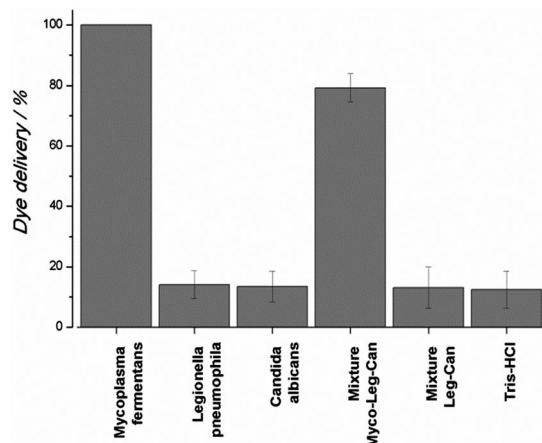


Fig. 2 Release of rhodamine B from solid **S1-O1/O2** in the presence of (from right to left): Tris-HCl buffer, a mixture of *Legionella pneumophila*–*Candida albicans* (Leg–Can) genomic DNA, a mixture of Leg–Can and *Mycoplasma fermentans* (Myco) genomic DNA, Can genomic DNA, Leg genomic DNA and Myco genomic DNA. All DNA were at a concentration of 1000 copies  $\mu\text{L}^{-1}$ .

commercially available PCR *Mycoplasma* detection kits (around  $\text{ca. } 10 \text{ copies } \mu\text{L}^{-1}$ ),<sup>7</sup> suggesting that this procedure for genomic DNA sensing possesses the potential of finding application in medical diagnosis protocols.

In further studies the selectivity in the detection of *Mycoplasma* genomic DNA by the **S1-O1/O2** solid was investigated by carrying out similar experiments in the presence of genomic DNA of the pathogens *Candida albicans* and *Legionella pneumophila* in amounts of 1000 copies  $\mu\text{L}^{-1}$ . The results obtained are shown in Fig. 2. As can be seen there were no significant differences between the blank (dye release in Tris-HCl buffer) and the potentially interfering genomic DNA of *Candida albicans* and *Legionella pneumophila*. In addition, when a mixture of *Candida albicans*–*Legionella pneumophila* (each one at a concentration of 1000 copies  $\mu\text{L}^{-1}$ ) was used no delivery of rhodamine B was observed. Furthermore, when genomic DNA of *Mycoplasma* was present in the mixture a clear delivery of rhodamine B was found pointing to a high degree of selectivity of the **S1-O1/O2** material.

In summary, we have prepared mesoporous silica nanoparticles loaded with rhodamine B and capped with a covalently linked double stranded DNA containing a highly conserved sequence of the 16S ribosomal subunit of the *Mycoplasma* species genome. The nanoparticles were able to deliver the entrapped

dye in the presence of *Mycoplasma fermentans* genomic DNA. A remarkable detection limit of  $\text{ca. } 70 \text{ DNA copies } \mu\text{L}^{-1}$  was found. Moreover the selectivity of the system was assessed, for instance, addition of *Candida albicans* and *Legionella pneumophila* genomic DNA was unable to induce dye release. We believe that DNA-based gated materials could be suitable alternatives to the standard bacteria detection techniques such as PCR and can find application as alternative rapid point-of-care diagnostic systems or in situations and places where sophisticated techniques or equipment are not available.

Financial support from the Spanish Government (Project MAT2012-38429-C04) and the Generalitat Valencia (Project PROMETEOII/2014/047) is gratefully acknowledged. Ll. P. is grateful to the Universidad Polit cnica de Valencia for his grant.

## Notes and references

- (a) A. Agostini, L. Mondrag n, A. Bernardos, R. Mart nez-M  nez, M. D. Marcos, F. Sancen n, J. Soto, A. Costero, C. Manguan-Garc a, R. Perona, M. Moreno-Torres, R. Aparicio-Sanchis and J. R. Murgu a, *Angew. Chem., Int. Ed.*, 2012, **51**, 10556–10560; (b) Q. Zhang, X. Wang, P.-Z. Li, K. T. Nguyen, X.-J. Wang, Z. Luo, H. Zhang, N. S. Tan and Y. Zhao, *Adv. Funct. Mater.*, 2014, **24**, 2450–2461; (c) C. Chen, J. Geng, F. Pu, X. Yang, J. Ren and X. Qu, *Angew. Chem., Int. Ed.*, 2011, **50**, 882–886; (d) L. Zhou, Z. Chen, K. Dong, M. Yin, J. Ren and X. Qu, *Adv. Mater.*, 2014, **26**, 2424–2430.
- (a) A. Agostini, L. Mondrag n, L. Pascual, E. Aznar, C. Coll, R. Mart nez-M  nez, F. Sancen n, J. Soto, M. D. Marcos, P. Amor s, A. M. Costero, M. Parra and S. Gil, *Langmuir*, 2012, **28**, 14766–14776; (b) C. Coll, A. Bernardos, R. Mart nez-M  nez and F. Sancen n, *Acc. Chem. Res.*, 2013, **46**, 339–349; (c) E. Climent, R. Mart nez-M  nez, A. Maquieira, F. Sancen n, M. D. Marcos, E. M. Brun, J. Soto and P. Amor s, *ChemistryOpen*, 2012, **1**, 251–259.
- (a) M. Oroval, E. Climent, C. Coll, R. Eritja, A. Avino, M. D. Marcos, F. Sancen n, R. Mart nez-M  nez and P. Amor s, *Chem. Commun.*, 2013, **49**, 5480–5482; (b) M. Chen, C. Huang, C. He, W. Zhu, Y. Xu and Y. Lu, *Chem. Commun.*, 2012, **48**, 9522–9524.
- (a) E. Climent, R. Mart nez-M  nez, F. Sancen n, M. D. Marcos, J. Soto, A. Maquieira and P. Amor s, *Angew. Chem., Int. Ed.*, 2010, **49**, 7281–7283; (b) E. Climent, L. Mondrag n, R. Mart nez-M  nez, F. Sancen n, M. D. Marcos, J. R. Murgu a, P. Amor s, K. Rurack and E. P rez-Pay , *Angew. Chem., Int. Ed.*, 2013, **52**, 8938–8942; (c) Z. Zhang, D. Balogh, F. Wang, S. Y. Sung, R. Nechushtai and I. Willner, *ACS Nano*, 2013, **7**, 8455–8468; (d) L. Wu, J. Ren and Q. Xiaogang, *Nucleic Acids Res.*, 2014, DOI: 10.1093/nar/gku858.
- H. Drexler and C. Uphoff, *Cytotechnology*, 2002, **39**, 75–90.
- (a) L. Matas Andreu, S. Molinos Ab s, G. Fern ndez Rivas, V. Gonz lez Soler and V. Ausina Ruiz, *Enferm. Infecc. Microbiol. Cl n.*, 2006, **24**, 19–23; (b) V. Ausina, *Infecciones causadas por micoplasmas*, *Medicina Interna*, Elsevier Espa a, 2004, 15th edn, pp. 2363–2365.
- (a) <http://www.lifetechnologies.com>; (b) <http://www.sigmaaldrich.com>; (c) <http://www.lgcstandards-atcc.org> (all websites accessed on 01/10/2014).

