

Cite this: *Anal. Methods*, 2015, 7, 8597

A preliminary investigation to group disparate batches of licit and illicit diazepam tablets using differential scanning calorimetry

S. Bibi,^a D. H. Bremner,^b M. Macdougall-Heasman,^b R. Reid,^a K. Simpson,^c A. Tough,^a S. Waddell,^a I. J. Stewart^b and K. H. Matthews^{*a}

Increasing numbers of illicit and unlicensed medicines are in general circulation and regularly seized by the police and other regulatory authorities. Forensic identification of seized tablets tends to focus on visual appearance and chromatographic identification of the contained drug. This process is relatively time consuming and places a strain on forensic laboratories. It was therefore of interest to investigate the possible application of differential scanning calorimetry (DSC) as a fast and efficient tool to facilitate the identification of contained drug/s and associated tablet excipients. Sixteen different cases (Cases A to P) of diazepam tablets obtained from Police Scotland were characterised based on visual appearance (colour and manufacturers' logos), physical attributes (size, weight and hardness), drug type, drug quantity (HPLC) and thermal properties (DSC). Raw DSC data was further processed using principal component analysis (PCA) as an objective assessment of the thermograms obtained with a view to statistical grouping of different cases. Cases J/K, M/O and L/P could be paired on visual appearance and Cases B/C/E/G and J/K/L/P on tablet hardness (17–23 and 80–89 N respectively). HPLC indicated that 75% of the cases examined contained diazepam but less than half of these contained the recognised amount (10 mg); Cases B/E/L/P contained phenazepam and J/K contained etizolam. The thermal signatures of individual tablets provided by DSC produced qualitative information about both drugs and excipients, indicating lactose in Cases D/F/H/I/J/K/M/N/O and Emcompress™ in B/E/L/P. In particular, DSC coupled with PCA provided confident groupings of A/C/G, B/E/L/P and H/I/J/K, and specific pairings of B/E, L/P and F/N.

Received 2nd July 2015
Accepted 18th August 2015

DOI: 10.1039/c5ay01171d

www.rsc.org/methods

1 Introduction

It is recognised that although the prevalence of prescribed dose benzodiazepines is generally declining, that of illicit (non-prescribed) use has been rising steeply since the 1980s and now presents a major health problem.^{1,2} Global estimates of non-medical use of prescription drugs are generally not available despite such use remaining a major public health concern but the misuse or non-medical use of benzodiazepines remains high and, at times, higher than that of many illicit (non-licensed) substances.³ Polydrug users, particularly heroin addicts and those on methadone medication, use benzodiazepines to enhance the effect of the opioids.^{4,5} Benzodiazepines are also often cited among other substances reported in both fatal and non-fatal overdose cases concerning opioid users. In Scotland, the misuse of benzodiazepines has been directly linked, or been a contributor to, drug related deaths, with

benzodiazepines noted in 28% of the fatalities reported during 2013 and diazepam specifically in 20% of all deaths.⁶ This is slightly down on the previous year's statistics of 34% for 'any benzodiazepine' and 27% for diazepam.²

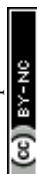
Control of these drugs in the UK is compromised by the increase in the amount of diazepam, for example, purchased from online pharmacies outside the European Union, mainly China and India, where the counterfeit trade appears to be sophisticated.³ Although medicinal solutions of this anxiolytic are legitimately available for oral and intravenous injection; as are tablets containing 2, 5 and 10 mg of the active,⁷ the 10 mg tablets are most commonly abused.

Apart from the necessary qualitative and quantitative analyses of drug type and content, other important tablet characteristics such as appearance (colour and identifying marks) and physical attributes such as size, shape, weight and hardness, can all contribute to forensic classification. Licitly produced tablets (tablets licensed in the UK) would be expected to conform to recognised national pharmacopoeial standards whereas illicitly produced tablets would not necessarily be subject to the same rigorous rules. Previous work using advanced chromatographic and spectroscopic methods, e.g. GC-MS, HPLC and SEM-EDXA, indicated that apart from a lack of control in drug quality,

^aSchool of Pharmacy and Life Sciences, Robert Gordon University, Riverside East, Garthdee Road, Aberdeen AB10 7GJ, UK. E-mail: k.h.matthews@rgu.ac.uk

^bSchool of Science, Engineering and Technology, Abertay University, Bell Street, Dundee DD1 1HG, UK

^cSpecialist Crime Division, Police Scotland, 173 Pitt Street, Glasgow G2 4JS, UK



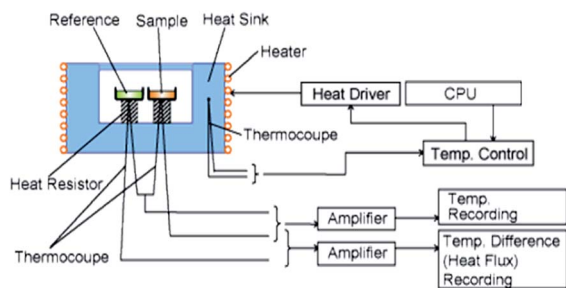


Fig. 1 Schematic diagram of a heat flux differential scanning calorimeter (DSC). Courtesy of <http://Hitachi-Hightec.com>.

quantity and even drug type, diazepam tablets being sold illicitly often bear a variety of genuine manufacturers' logos that serve to confuse the issue of forensic identification.⁸ In these preliminary studies, 39% of the individually seized batches contained the expected quantity of diazepam while 5% contained a lesser amount. More worryingly, 32% contained a much higher than expected level of diazepam with one batch containing 48 mg of the active ingredient *i.e.* almost five-times the recommended dose. Surprisingly, 24% of the batches tested were found to contain the much more potent and unlicensed (in the UK) phenazepam and the similarly unclassified thienotriazolodiazepine, etizolam.

In addition to drug analysis, identification and characterisation of tablet excipients, *i.e.* pharmacologically inert materials that are blended with the pure drug to enable tablet production, was also of interest when classifying tablets. To this end, differential scanning calorimetry (DSC) was investigated for 'thermal profiling' of unknown tablets. DSC is a non-standard technique in the forensic identification of tablets despite being well-established as a key analytical tool in preformulation studies within the pharmaceutical sector.^{9–11} The method involves heating a small amount (<10 mg) of powdered sample, contained in a closed aluminium pan, in a furnace alongside an empty pan and recording the temperature difference between both pans, Fig. 1. Changes to the heat capacity of the sample are indicative of phase changes or chemical reactions within the sample and are characteristic for individual samples. Thermal events encountered typically include dehydration, recrystallization, melting and degradation. For example, diazepam tablets containing the common excipient lactose would be expected to show a signal for the melting of the drug and the dehydration and melting/degradation of lactose.¹² This well-established thermoanalytical technique was applied to the analysis of disparate tablet seizures supplied by Police Scotland and the raw data further processed using principal component analysis (PCA) to evaluate the grouping of individual tablets in terms of thermal profile.

2 Materials and methods

2.1 Materials

Confiscated tablets containing 'diazepam' were kindly donated by Police Scotland (Cases A to P). Licit diazepam tablets (28 × 10 mg) were supplied by Actavis (Barnstaple, UK) and Teva

(Runcorn, UK) and are coded as 'Com 1' and 'Com 2' respectively. Diazepam, temazepam, oxazepam, nitrazepam, clonazepam and phenazepam were supplied by Sigma-Aldrich (Gillingham, UK). Tablet excipients including Maize Starch B.P. (Thornton & Ross, Huddersfield, UK), calcium phosphate dihydrate, *Emcompress*TM (Forum Products, Surrey, UK) and several grades of lactose monohydrate including 'Lactose B.P.' (supplier unknown) – Grade 1; *Granulac*TM 230 (Meggle, Wasserburg, Germany) – Grade 2; *Pharmatose*TM 80 – Grade 5; *Lactopress*TMSD, 150, 175 and 300 μm (DFE Pharma, Goch, Germany) – Grades 4, 3, 6 and 7 respectively – were all analysed by DSC. Tablet lubricants magnesium stearate and stearic acid, and tablet binder, poloxamer 407, were also supplied by Sigma-Aldrich. Polyvinylpyrrolidone, PVP (PVP-K25), another common tablet binder known to be present in commercial products, was also analysed.

2.2 Measurement of physical attributes

2.2.1 Tablet weight and size. All tablets (16 individual cases assigned letters A to P) were weighed on a calibrated analytical balance (Mettler Toledo) and the mean tablet weight and standard deviation calculated. Tablet thickness was determined at the point of maximum depth and tablet diameter was measured in the direction of the score line using a digital calliper (150 cm, 110 DBL series, Stanton, London, UK). The absolute number of tablets available varied from case to case.

2.2.2 Tablet hardness ('crushing strength'). The force required to cause mechanical failure of individual tablets was performed using a THB 28 tablet hardness tester (Erweka, Heusenstamm, Germany). Following careful cleaning of the test platform, individual tablets were placed flat on the platform with the diameter parallel and the score line (if present) perpendicular to the plane of the compressive force. The applied force at which the tablet undergoes catastrophic mechanical failure was recorded and the crushed tablets and associated fragments/powder carefully removed to a clean, dry glass sample bottle. Crushed samples were labelled according to the case code and individually numbered for easy identification prior to analysis by DSC.

2.3 Visual analysis and drug content

2.3.1 Visual analysis. Colour photographs of individual tablets were taken using a Nikon D3200 DSLR camera fitted with a Sigma EX 105 mm macro lens and a 58 mm skylight filter (Jessops, Marlow, England).

2.3.2 Drug content by HPLC. High performance liquid chromatography, HPLC, was used to identify and quantify the drug content of each batch of tablets. A prominence UFLC system with DAD detection in the region 200–400 nm (Shimadzu, Kyoto, Japan) coupled with a C18 Gemini analytical column, 250 × 4.6 mm i.d. and 5 μm sized stationary phase (Phenomenex, Macclesfield, UK) was used. Prior to tablet analysis, calibration was undertaken with the pure benzodiazepine samples. Individual samples (1 mg) were dissolved in acetonitrile (1 mL) and 0.1 M HCl added to a final volume of 5 mL giving a concentration of 0.2 mg mL⁻¹. All solutions were



Table 1 Identification of individual tablets containing details of visible markings (logos), a summary of the key excipients identified by DSC and the diazepam content as determined by HPLC

Case code	Markings	Excipients detected	Diazepam content (mg per tablet)
Com 1	“C/DC”	Lactose, stearic acid	8.79
Com 2	“10”	Lactose	8.48
A	No markings	Stearic acid, <u>no</u> lactose	7.32
B	“MA/D10”	<i>Emcompress</i> , <u>no</u> lactose	0 (Phenazepam)
C	“MA/D10”	Stearic acid, <u>no</u> lactose	7.15
D	“COX/DC”	Lactose (amorphous)	23.24
E	“MA/D10”	<i>Emcompress</i> , <u>no</u> lactose	0 (Phenazepam)
F	“MSJ” (small tablet)	Lactose	7.58
G	“MA/D10”	Stearic acid, <u>no</u> lactose	6.61
H	“COX/DC”	Lactose (amorphous)	23.38
I	“COX/DC”	Lactose (amorphous)	23.11
J	“MA/D10”	Lactose (amorphous)	0 (Etizolam)
K	“MA/D10”	Lactose (amorphous)	0 (Etizolam)
L	“10”	<i>Emcompress</i> , <u>no</u> lactose	0 (Phenazepam)
M	“MA/D10”	Lactose (amorphous)	9.52
N	“Tensium”	Lactose	9.37
O	“MA/D10”	Lactose (amorphous)	9.73
P	“10”	<i>Emcompress</i> , <u>no</u> lactose	0 (Phenazepam)

stored in a refrigerator at 3 °C to minimise hydrolysis. An injection volume of 20 µL was used. The mobile phase was composed of a phosphate buffered solution (4.8 g of dihydrogen potassium orthophosphate and 1.2 mg of dipotassium hydrogen orthophosphate anhydrous dissolved in 1000 mL distilled water, adjusted to pH 6.0 ± 0.5 with 0.1 M HCl, Eluent A) and HPLC grade acetonitrile, Eluent B (Fisher Scientific, Loughborough, UK).

A randomly selected tablet from each batch was crushed in a mortar and pestle and approximately 20 mg was accurately weighed and transferred to a volumetric flask (10 mL) and acetonitrile (2 mL) added followed by 0.1 M HCl to 10 mL giving a final concentration of 2.0 mg mL⁻¹. Samples for analysis were filtered (1 mL Plastipak syringe and disposable 0.45 µm Whatman syringe filter) prior to injection (20 µL). A mobile phase gradient of 75 : 25 (Eluent A : Eluent B) was changed to 45 : 55 over 31 minutes, held for 4 minutes and returned to 75 : 25 at 35 minutes. Peak integration was undertaken using LC solutions software. Values quoted in Table 1 are the mean of 3 measurements from each case.

2.4 Thermoanalytical analysis by differential scanning calorimetry

2.4.1 Calibration of instrument. Prior to a particular sequence of sample measurements, the DSC was calibrated using an indium standard ($T_m = 156.6$ °C) at the heating rate used (10 °C min⁻¹). All efforts to reduce external contamination of the heating cell and sample pans were undertaken.

2.4.2 Analysis of a selection of benzodiazepines. Pure samples (5–10 mg) of diazepam, temazepam, oxazepam, nitrazepam and clonazepam were accurately weighed (±0.05 mg) placed in non-hermetically sealed, standard aluminium pans

(40 µL) and heated from 20–250 °C under a constant flow rate of nitrogen (50 mL min⁻¹) in a Q100 series DSC (TA Instruments, Newcastle, USA) at a heating rate of 10 °C min⁻¹.

2.4.3 Analysis of a selection of sugars. Pure samples (5–10 mg) of a selection of sugars that may be used as tablet excipients, namely, glucose, fructose, trehalose, sorbitol, mannitol and lactose were placed in standard aluminium pans and heated from 20–250 °C at 10 °C min⁻¹.

2.4.4 Analysis of a selection of tablet excipients. Samples (5–10 mg) of Maize Starch B.P., *Emcompress*TM, poloxamer 407, PVP, stearic acid and magnesium stearate, a common tablet lubricant, were placed in individual, standard aluminium pans and heated from 20–250 °C at 10 °C min⁻¹.

2.4.5 Analysis of a selection of lactose grades. A sample (5–10 mg) of different commercial grades of lactose typically used for tablet manufacture were placed in individual, standard aluminium pans and heated from 20–250 °C at 10 °C min⁻¹.

2.4.6 Analysis of licit and illicit tablets. Tablet samples were crushed and finely ground in a small agate mortar and pestle. Samples of each tablet (5–10 mg) were placed in a standard aluminium pan and heated from 20–250 °C at 10 °C min⁻¹.

In addition to normal processing of DSC data using TA universal analysis software (version 4.3), principal component analysis (PCA) was performed on the raw heat flow data from 25–213 °C using *Unscrambler@ X* (Camo Software AS, Oslo, Norway). The rationale for this was to further objectify the DSC data with a view to producing statistical matches of disparate tablets.

3 Results and discussion

3.1 Physical attributes

With the exception of Case F, much smaller in diameter (5.55 ± 0.01 mm) than all the other cases which ranged from 8.03 (A and N)–8.15 mm (G), only Cases C/G and A/N could be confidently distinguished from the rest based on mean diameter and standard deviation. Most notable from consideration

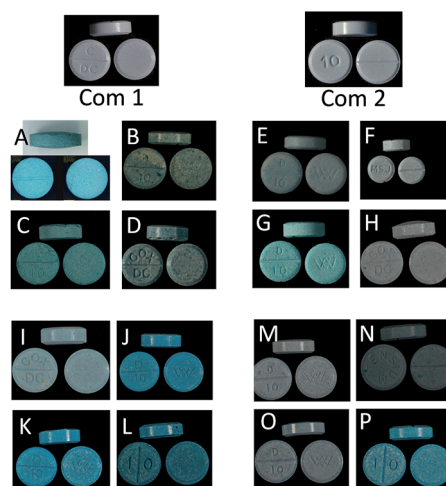


Fig. 2 Photographic images of licit (Com 1 and Com 2) and illicit ‘diazepam’ tablets. Individual Cases A to P are included.



of mean tablet weights was the relatively small standard deviations obtained for the two licit cases, Com 1 and Com 2 (0.0014–0.0017 g). These values should be compared with Cases E and I (0.0133–0.0138 g) respectively which showed the highest deviations in weight. All other cases had relatively narrow deviations from the individual mean tablet weights (0.0016–0.0084 g). For the purposes of comparison of all batches, mean weight was of limited value. Of much more value was comparison of tablet hardness. Clear and significant differences were apparent between Cases B/C/E/G, which were softer (17–23 N) than all the others, the hardest tablets being J/K/L/P (80–89 N).

Measurement of tablet thickness indicated highly variable results ranging from 2.4367 ± 0.0635 mm (F) to 3.5200 ± 0.0872 mm (A). The narrowest standard deviations from the mean (<0.03 mm) were obtained for Cases C/L/M/O/P, Com 1 and Com 2. Based on these physical attributes alone, Cases B/C/E/G and J/K/L/P were grouped with low confidence.

3.2 Visual analysis

Tablet markings indicated that Cases B/C/E/G (“MA/D10”), J/K (“MA/D10”) and L/P (“10”) may be grouped based on logo alone but there remained discrepancies between the absolute font type and character spacing (see Cases E and G – spacing of the numbers “1” and “0” and Cases K and L – different logos), Table 1 and Fig. 2. Based on visual observation of colour alone, only Cases J/K, L/P and M/O showed any resemblance, however, too much variability between colour, hue and distribution of dye substance rendered subjective assessment of little use.

3.3 Drug type and content

Definitive identification of the specific drug and its content in individual tablets was of primary importance for forensic identification. HPLC identified two distinctly different benzodiazepines (diazepam and phenazepam) and the thienotriazolodiazepine, etizolam in particular cases (Table 1). This enabled Cases B/E/L/P to be related based on phenazepam content although the precise quantity per tablet was not determined at this stage in these studies. Identification of etizolam in Cases J and K (quantities unknown) suggested a connection supported by testing of physical attributes, markings and colour (Fig. 2). All other tablets tested contained varying levels of diazepam from 6.61 mg per tablet for Case G to 23.38 mg per tablet for Case H (Table 1). Based on the recognised doses of 2, 5 and 10 mg per tablet, diazepam content of the licit batches (Com 1 and Com 2) were actually less than the expected 10 mg per tablet. The British Pharmacopoeia¹² specifies that for uniformity of (drug) content, not more than one tablet should deviate from 85–115% of the average content of 10 tablets. Clearly, Com 2 is on the lower limit of this specification but would have passed the manufacturer's quality system. This is also true of Cases M, N and O which contain 9.52, 9.37 and 9.73 mg per tablet diazepam respectively. In contrast, Cases A, C, D, F, G, H and I would fail the pharmacopoeial test by a considerable amount. Most worryingly are the dangerously high levels of diazepam in Cases D, H and I of 23.24, 23.38 and

23.11 mg per tablet respectively. It may be safe to assume that based on diazepam content alone, only Cases M, N and O could be construed as licitly produced.

3.4 Differential scanning calorimetry

3.4.1 Benzodiazepines. An overlay of the thermograms for the five different benzodiazepines analysed by DSC (Fig. 3) indicated five distinct and sharply defined endothermic events attributed to the individual melting point of each pure compound. The onset temperatures, in increasing order, were 130.3 °C (diazepam), 158.5 °C (temazepam), 194.3 °C (oxazepam), 227.0 °C (nitrazepam) and 238.4 °C (clonazepam). These can be compared with the literature values of 125–126 °C (diazepam), 119–121 °C (temazepam), 205–206 °C (oxazepam), 224–226 °C (nitrazepam) and 236.5–238.5 °C (clonazepam).¹³

Clearly, there are significant discrepancies in the values of melting point determined in these studies by DSC for temazepam, diazepam and oxazepam compared to the quoted literature values. It should be noted that the quoted values take no account of polymorph or pseudopolymorph (solvate) formation and refer to crystals from specific solvents with melting points determined using traditional laboratory melting point apparatus. The three benzodiazepines showing discrepancies in this study, like most drug substances, are known to exhibit polymorphism.¹⁴ Using DSC, melting for diazepam has been recorded at 132.8 °C,¹⁵ 132 °C¹⁶ and 132.3 °C (unspecified forms);¹⁷ 118 °C (Form V), 120 °C (Form III), 136 °C (Form I-solvate), 134 °C (Form VI-hydrate) and 157 °C (Form VII-solvate).¹⁸ A metastable form of temazepam has been recorded at 159 °C¹⁴ that can be compared with the widely quoted value of 119–121 °C for ‘crystals from cyclohexane’.¹³ The melting of oxazepam (Form IV)¹⁹ is recorded at 195 °C but known to coincide with decomposition rendering the value for the enthalpy of the melt transition quoted in Fig. 3 indeterminate. Although unable to give unambiguous indication of the specific polymorphic form of a substance as a stand alone technique in the absence of definitive X-ray crystallographic data, for the

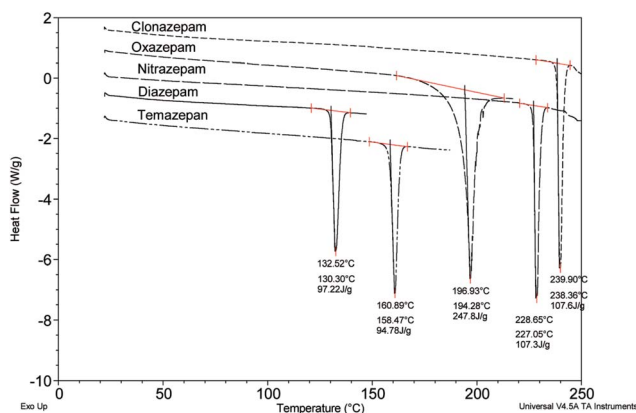


Fig. 3 DSC thermograms of five different benzodiazepines at a heating rate of $10^{\circ}\text{C min}^{-1}$ indicating the individual melting points. Values of T_{onset} and T_{max} and the enthalpy of fusion (J g^{-1}) are indicated for each pure compound.



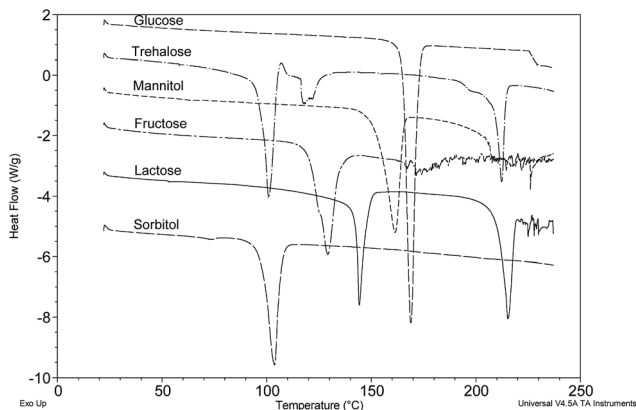


Fig. 4 DSC thermograms of six sugars commonly used as tablet excipients.

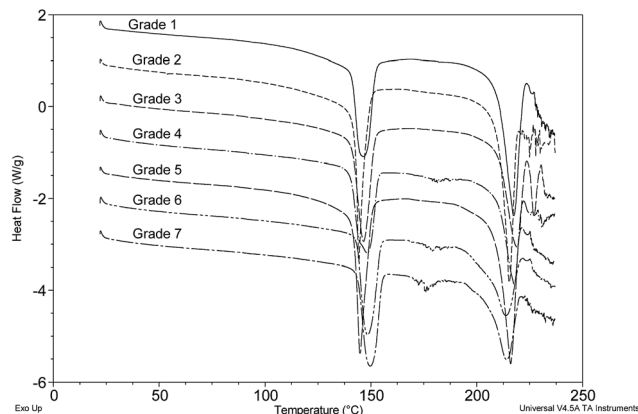


Fig. 5 Grade 1-Lactose B.P. (supplier unknown); Grade 2-Granulac™230; Grade 3-Lactopress™150; Grade 4-Lactopress™MSD; Grade 5-Pharmatose™80; Grade 6-Lactopress™175; Grade 7-Lactopress™300.

purposes of this study, DSC is able to quickly distinguish between the different types of benzodiazepine that may be present in an unknown tablet sample. In these studies, 66% of the tablets tested showed evidence of a sharp melt transition around 130 °C indicative of diazepam, supported by the results of HPLC analysis summarised in the previous section.

3.4.2 Common tablet excipients. Sugars are commonly used as tablet fillers or diluents, lactose being by far the most common.²⁰ Other sugars or sugar alcohols such as glucose, sucrose or sorbitol, and more rarely trehalose or fructose,²¹ can also be used. Samples of each of these sugars were analysed by DSC using a single heating run and presented in Fig. 4. Distinctive thermal signatures were apparent for all samples indicating mainly, dehydration, melting and degradation events. Lactose being the most common tablet excipient was of particular interest and showed a clear dehydration of the α -monohydrate to the α -anhydrous form with an onset temperature of 142.6 °C.^{12,20,21} Melting of the α -anhydrous form was immediately followed by decomposition, indicated by an erratic baseline. No evidence of the recrystallization of amorphous lactose was evident around 167 °C.¹²

Several different pharmaceutical grades of lactose (numbered 1–7—see Section 2.1) were analysed by DSC (Fig. 5) and some minor variation in the peak temperature of 144–150 °C for dehydration of α -lactose monohydrate and observable differences in the peak width were apparent. Grades 4, 6 and 7 showed some evidence of amorphous content (\sim 167 °C) that was much less notable in the other grades tested. As an aside, Grade 3 indicated the probable melting of β -anhydrous lactose at 227 °C.

Other common tablet excipients tested included the binders maize starch, PVP and poloxamer 407, and the lubricants magnesium stearate and stearic acid (known to be present in Com 1). A variety of thermal behaviours were apparent for these four excipients and are shown in Fig. 6.

Briefly, magnesium stearate indicated a broad melting range of 110–125 °C typical of commercial samples;²¹ stearic acid displayed a well-defined endotherm with an onset of 68.6 °C and a weaker endotherm with onset 55.71 °C, the latter event

being attributed to <10% (w/w) palmitic acid content which appears to be common;¹² poloxamer 407 (a synthetic co-poly-ether) displayed a sharp melt at 53.3 °C and maize starch produced a very broad displacement of the base line between 20 and 160 °C known to be associated with the desorption of water as a function of increasing temperature during measurement. PVP displayed a largely non-descript thermogram showing a small drift in the baseline (heat capacity) most probably associated with desorption of water as for maize starch. No other thermal events were detected to 200 °C.

3.4.3 Licit and illicit 'diazepam' tablets. With the benefit of the DSC data presented in Fig. 3–6, analysis of all the available tablets, including the two licit commercial types (Com 1 and Com 2) quickly indicated that eleven of the eighteen test cases, including the licit tablets, contained lactose as filler/diluent. Of these eleven tablets, all but three showed clear signs of amorphous lactose as evidenced by a significant exothermic event around 167 °C attributed to recrystallization of amorphous lactose to the α -anhydrous crystalline state (Fig. 7). Both licit tablets and Case N did not show evidence of amorphous lactose.

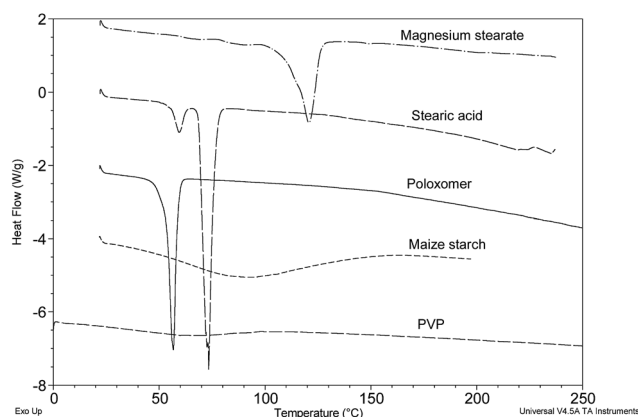


Fig. 6 DSC thermograms of five common tablet excipients for illustrative purposes.



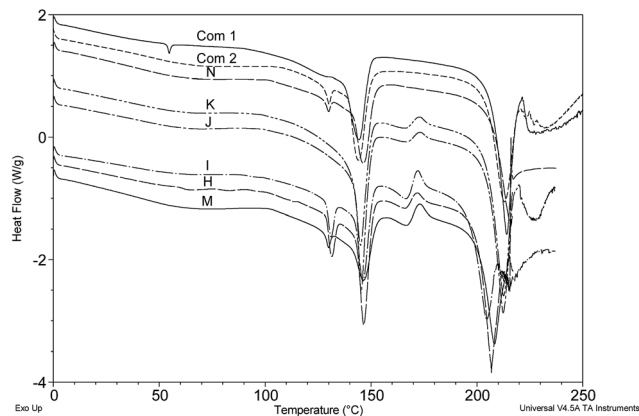


Fig. 7 DSC thermograms of seven selected cases all containing lactose as the bulk excipient and including Com 1 and Com 2 for comparison. Note the apparent absence of amorphous lactose content in both licit samples and Case N.

The presence of diazepam was clearly resolved in Com 2 and to a lesser extent in Com 1. It should be noted that any interaction between drug and excipient/s and/or changes to the degree of drug crystallinity as a result of processing conditions can affect the resolution of the drug melting endotherm. The fact that diazepam is less clearly resolved in Com 1 may be attributable to any of these factors. Both licit tablets, Com 1 and Com 2, contained 8.79 and 8.48 mg of diazepam per tablet respectively, as measured by HPLC (Table 1). Com 1 gave a weak but relatively sharp signal corresponding to the melting of stearic acid, a tablet lubricant, as might have been expected from consideration of the manufacturer's labelling. Of the remaining ten cases containing diazepam, the high levels of drug measured for Cases H, I (Fig. 7) and D (see Fig. 10) indicated a more intense signal for diazepam compared with all the other diazepam-containing tablets. This indication was corroborated by the diazepam contents as determined by HPLC of 23.24, 23.38 and 23.11 mg per tablet respectively (Table 1).

Cases A, C and G showed no sign of lactose content, giving a fairly broad and (as yet) uncharacterised endothermic event

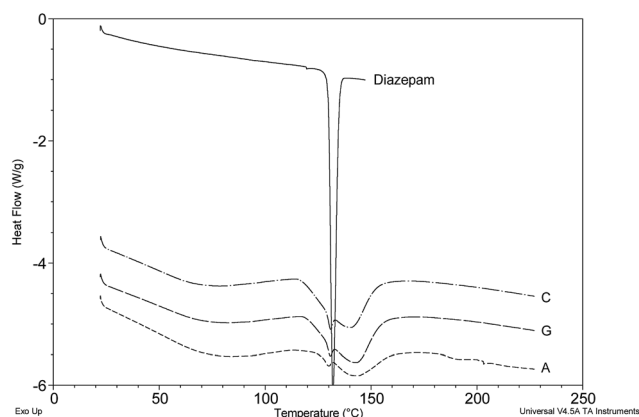


Fig. 8 DSC thermograms of Cases A, C and G containing diazepam and an (as yet) unidentified excipient.

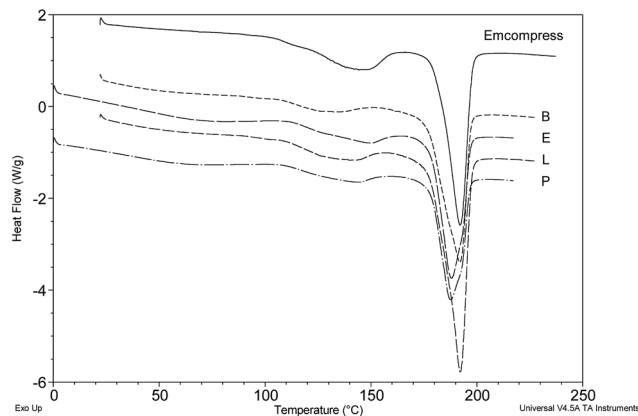


Fig. 9 DSC thermograms of Cases B, E, L and P containing Emcompress™ as the bulk excipient.

from 115–150 °C that included a clear signature peak for diazepam (Fig. 8). Cases B, E, L and P similarly showed no evidence for lactose but were clearly composed of the alternative tablet filler/diluent known as 'Emcompress', a trade name for calcium hydrogen phosphate dihydrate.²¹ HPLC indicated that these particular tablets contained the unlicensed phenazepam (results not shown) a potent benzodiazepine that has only recently been classified in the UK and is known to be around five-times more potent than diazepam.^{22,23} Two polymorphic forms of phenazepam have recently been reported with melting points of 207–212 °C (β -form) and 226–227 °C.²⁴ No evidence of either of these transitions was apparent in the DSC results presented (Fig. 9). Possible reasons could be attributed to the proximity of the main Emcompress dehydration event causing major perturbations within the sample pan and a resultant loss of a distinct signal for the drug, or the drug content being too low for detection. A combination of these factors may also be involved.

It was concluded that DSC is a fast and effective method of tablet analysis able to reveal both crude and subtle differences between disparate batches of tablets. This capability is

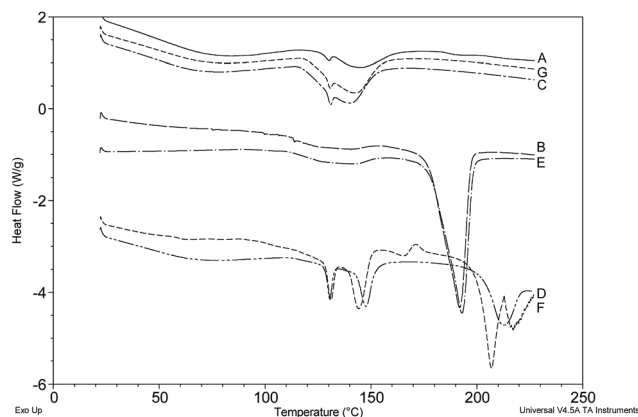


Fig. 10 DSC thermograms of a selection of cases grouped by bulk excipient. Note the significant differences between Cases D and F which both contain lactose.



illustrated in Fig. 10 where seven separate cases can be quickly grouped into three distinct sets based on the bulk excipients *i.e.* A/C/G (unknown excipient), B/E (Emcompress) and D/F (lactose). Comparison of Cases D and F indicates significant differences in the lactose ‘signature’ that suggest tablets of different origin and this is supported by the appearance of distinctly different tablet logos and markings (Fig. 2).

3.4.4 Principal component analysis (PCA) of DSC data. PCA is a widely used chemometric technique that reduces the dimensionality of multivariate data. In essence, a matrix of the covariances of each of the variables (heat flow and temperature) is produced and the eigenvectors are determined. The scalar magnitude of each eigenvector is reduced to unit length and labelled as a principal component (PC) which represents a “latent axis” of variability within the dataset. The PCs are ranked according to their eigenvalues, the largest of which holds the greatest variability. This is an unsupervised technique and existence of a correlation between samples is highlighted by clustering in a scores plot. It was undertaken on all DSC data to remove subjectivity of observation. PCA of DSC data on all tablets included in this study is represented by the scores plot, Fig. 11. Cases B/E and L/P are far-separated from all other cases, the stated pairings being consistent with tablet hardness, weight, visual analysis and HPLC (phenazepam). Another clear grouping based on DSC alone is Cases H/I/J/K, the pairing of J and K corroborated by hardness, visual analysis and GC-MS (etizolam). Cases H and I were closely related based on visual analysis and drug content by HPLC (diazepam). Of the remainder of the cases investigated, including the two known commercial samples, Cases A, C and G did not contain lactose or Emcompress (bulk excipient as yet unknown) but contained similar quantities of diazepam. In addition to this clear grouping, C/G and A/N could be paired on mean tablet diameter and deviation, however, only Cases C and G shared the markings “MA/D10” with A and N showing no markings and “Tensium” respectively.

Finally, the bulk of the cases containing lactose as the main excipient were closely grouped by PCA as might be expected and discrepancies relating to the relative magnitude of diazepam

content (height of the diazepam melt transition relative to the dehydration of lactose) and the conspicuous signature associated with the presence of amorphous lactose, were distinguishing features readily revealed by DSC that aided further correlation of different cases.

4 Conclusions

DSC is a fast and effective method for generating the thermal profile of unclassified tablets that can be used in conjunction with other characterisation methods to definitively highlight any similarities between individual batches of tablets. This ability makes it invaluable as a method for the forensic identification of tablets.

Acknowledgements

The authors would like to thank *Police Scotland* for supplying the diazepam tablets and Mr Iain Tough (Forensic Laboratory, *RGU*) for expert advice in forensic imaging.

References

- 1 Medical Healthcare Regulatory Agency. 2013, Man sentenced to 44 months imprisonment after £1.6 million worth of unlicensed and Class C medicines seized by MHRA.
- 2 *Drug-related deaths in Scotland in 2013*, Report by the National Records of Scotland, August 2014.
- 3 *World Drug Report 2013*, United Nation Office of Drugs and Crime, United Nations, June 2013.
- 4 C. E. Ashton, *Benzodiazepine abuse. Drugs and dependence*, Harwood, London, 2002.
- 5 J. D. Jones, S. Mogali and S. D. Comer, *Drug Alcohol Depend.*, 2012, **125**, 8–18.
- 6 *Drug-related deaths in Scotland in 2012*, Report by the National Records of Scotland, August 2013.
- 7 *British National Formulary*, Pharmaceutical Press, 2014, **66**, p. 225.
- 8 M. Macdougall-Heasman, I. J. Stewart, D. H. Bremner, A. Tough and K. Simpson, *Poster presented at Home Office Science and Engineering Conference on ‘Tackling Drug and Alcohol Abuse’*, London, 16th September 2014.
- 9 P. Mura, A. Manderioli, G. Bramanti, S. Furlanetto and S. Pinzauti, *Int. J. Pharm.*, 1995, **119**, 71–79.
- 10 L. Muggetti, M. Zurlo, A. Martini, P. Civaroli and R. de Ponti, *Int. J. Pharm.*, 1996, **136**, 81–88.
- 11 G. van den Mooter, P. Augustijns, N. Bleton and R. Kinget, *Int. J. Pharm.*, 1998, **164**, 67–80.
- 12 A. Gombás, P. Szabó-Révész, M. Kata, G. Regdon Jr and I. Erős, *J. Therm. Anal. Calorim.*, 2002, **68**, 503–510.
- 13 *Merck Index, An encyclopedia of chemicals, drugs and biologicals*, ed. S. Budvari, Merck & Co., Inc., New Jersey, U.S.A., 13th edn, 2001.
- 14 D. Giron, *Thermochim. Acta*, 1995, **248**, 1–59.
- 15 D. Tita, A. Fuluius, B. Tita, G. Vlase, B. Vlase and N. Doca, *Rev. Roum. Chim.*, 2010, **55**, 1047–1052.

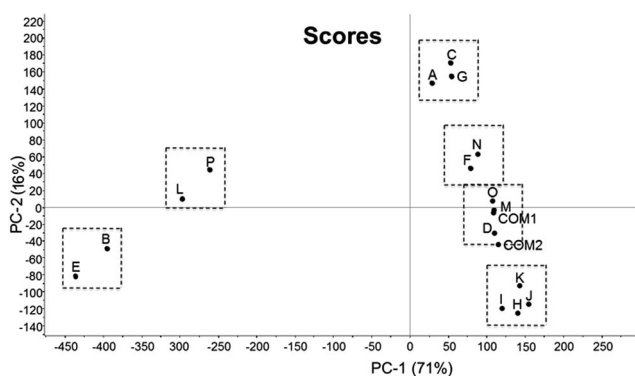


Fig. 11 Scores plot for all cases examined (A to P) indicating unambiguous groupings of different cases. Note that B, E, L and P can be further grouped into pairs and that A/C/G and H/I/J/K from distinct groups.



- 16 D. R. Lide, *Handbook of Chemistry and Physics*, CRC Press, 88th edn, 2007, pp. 3–516.
- 17 O. Chitvanich, B. Sirithunyalug, S. Okonogi, S. Piyamongkol and J. Sirithunyalug, *J. Nat. Sci.*, 2009, **8**, 175–188.
- 18 J. R. Rao, K. N. Rao, S. R. Krishna, G. A. Rani, J. Madhavi, R. C. Kanaiyalal, N. L. Rao and D. Debashish, *International Pat.*, WO 201004692920, 2010.
- 19 J. Masse, O. Hourri, P. Bernier and M. Ribet, *Thermochim. Acta*, 1993, **216**, 191–212.
- 20 G. Alderborn, Tablets and compaction, in *Aulton's Pharmaceutics – the design and manufacture of medicines*, ed. M. E. Aulton and M.G. Taylor, Churchill-Livingstone, 4th edn, 2013, ch. 30, p. 514.
- 21 H. A. Kibbe, *Handbook of Pharmaceutical Excipients*, Pharmaceutical Press, 3rd edn, London, 2010.
- 22 L. Iverson, *Letter to the Minister of State for Crime Prevention and ASB Reduction*, Advisory Council on the Misuse of Drugs (ACMD), UK Government, 2011.
- 23 UK Government, Explanatory memorandum to the Misuse of Drugs Act 1971 (Amendment) Order 2012, no. 1390, National Archives.
- 24 Y. N. Morozov, A. Y. Utekhina, V. P. Shabatin, V. V. Chernyshev and G. B. Sergeev, *Russ. J. Gen. Chem.*, 2014, **84**, 1010–1017.

