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## Screen-printed back-to-back electroanalytical sensors: heavy metal ion sensing†

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Screen-printed back-to-back microband electroanalytical sensors are applied to the quantification of lead(II) ions for the first time. In this configuration the electrodes are positioned back-to-back with a common electrical connection to the two working electrodes with the counter and reference electrodes for each connected in the same manner as a normal “traditional” screen-printed sensor. Proof-of-concept is demonstrated for the electroanalytical sensing of lead(II) ions utilising square-wave anodic stripping voltammetry where an increase in the electroanalytical sensitivity is observed by a factor of 5 with the back-to-back microband configuration at a fixed lead(II) ion concentration of  $5 \mu\text{g L}^{-1}$  utilising a deposition potential and time of  $-1.2 \text{ V}$  and 30 seconds respectively, compared to a conventional (single) microband electrode. The back-to-back microband configuration allows for the sensing of lead(II) ions with a linear range from 5 to  $110 \mu\text{g L}^{-1}$  with a limit of detection (based on  $3\sigma$ ) corresponding to  $3.7 \mu\text{g L}^{-1}$ . The back-to-back microband configuration is demonstrated to quantify the levels of lead(II) ions within drinking water corresponding to a level of  $2.8 (\pm 0.3) \mu\text{g L}^{-1}$ . Independent validation was performed using ICP-OES with the levels of lead(II) ions found to correspond to  $2.5 (\pm 0.1) \mu\text{g L}^{-1}$ ; the excellent agreement between the two methods validates the electroanalytical procedure for the quantification of lead(II) ions in drinking water. This back-to-back configuration exhibits an excellent validated analytical performance for the determination of lead(II) ions within drinking water at World Health Organisation levels (limited to  $10 \mu\text{g L}^{-1}$  within drinking water).

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## 1. Introduction

Screen-printed electrochemical derived sensors have revolutionised the field due to their capability to bridge the gap between laboratory experiments with in-field implementation.<sup>1–6</sup> Furthermore the ability to mass produce screen-printed electrodes allows their use as highly reproducible, economic one-shot sensors, alleviating potential memory effects and contamination whilst eradicating the requirement for electrode pre-treatment and preparation, as is often the case for solid electrodes (such as glassy carbon and boron-doped diamond *etc.*) prior to their use.<sup>7–11</sup>

Recently we introduced the concept of the screen-printed back-to-back electrode configuration where both sides of a

plastic substrate are screen-printed upon utilizing the usually redundant back of the screen-printed sensor, converting this “dead-space” into a further electrochemical sensor which results in improvements in the analytical performance;<sup>12</sup> Fig. 1

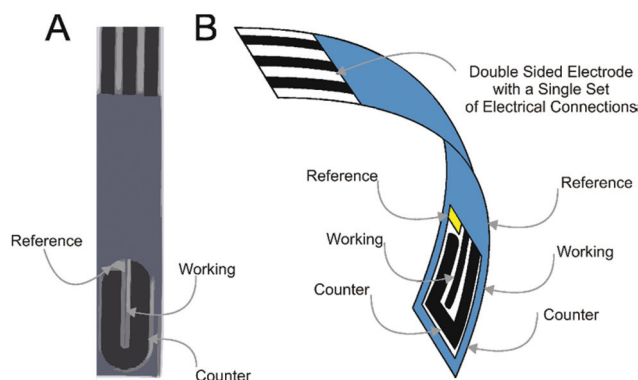


Fig. 1 Optical image of a single side of a back-to-back screen-printed graphite microband (A) and a schematic depiction of the back-to-back configuration demonstrating the electrode lay-out and single point of electrical connection (B).

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depicts the concept where screen-printed microband electrodes are fabricated “back-to-back”.<sup>12</sup>

In this paper the exploration of the screen-printed back-to-back electrode configuration towards the sensing of the heavy metal lead(II) ions is considered for the first time. This target metal ion has received much attention owing to its high toxicity, as its accumulation in the body has serious deleterious effects on humans.<sup>13</sup> In particular, lead strongly affects the mental and physical development of children, and can cause poisoning in adults, inducing severe damage to the liver, brain, kidneys, reproductive system and central nervous system.<sup>13</sup> Since lead(II) ions are naturally found within drinking water, the World Health Organisation (WHO) recommends this is limited to 10  $\mu\text{g L}^{-1}$ .<sup>14</sup> We demonstrate that screen-printed microband electrode back-to-back configurations exhibit analytically useful sensing capabilities with improvements in the electroanalytical sensitivity observed over that of traditionally employed single microband electrodes. Proof-of-concept is demonstrated for the sensing of lead(II) ions at analytically useful levels within model conditions and is demonstrated to successfully quantify lead(II) ions in drinking water which is independently validated with ICP-OES; the analytical utility of the proposed back-to-back microband electrode is demonstrated to quantify lead(II) ions in drinking water at WHO levels.

## 2. Experimental

All chemicals used were of analytical grade and were used as received without any further purification and were obtained from Sigma-Aldrich. The solutions were prepared with deionised water of resistivity not less than 18 M $\Omega$  cm. All measurements were performed with a Palmsens (Palm Instruments BV, The Netherlands) potentiostat.

All measurements were conducted using a screen-printed graphite microband three electrode configuration (bSPE) consisting of a carbon-graphite geometric working electrode (100  $\mu\text{m}$  diameter and 20 mm length), carbon-graphite counter electrode and Ag/AgCl reference.<sup>15</sup> Such bSPEs were fabricated in-house with appropriate stencil designs using a microDEK 1760RS screen-printing machine (DEK, Weymouth, UK). A carbon-graphite ink formulation (Product Code: C2000802P2; Gwent Electronic Materials Ltd, UK) was first screen-printed onto a polyester flexible film (Autostat, 250  $\mu\text{m}$  thickness). This layer was cured in a fan oven at 60 degrees for 30 minutes. This layer defines the graphite working electrodes for the configurations, as shown in Fig. 1, which tailors off onto a larger size graphite pad to enable ease of connection to an edge connector.<sup>16</sup> Next a silver/silver chloride reference electrode was included by screen-printing a Ag/AgCl paste (Product Code: C2040308P2; Gwent Electronic Materials Ltd, UK) onto the plastic substrate. Last, a dielectric paste ink (Product Code: D2070423D5; Gwent Electronic Materials Ltd, UK) was printed to cover the connections and define the carbon-graphite working electrode. After curing at 60 degrees for 30 minutes the screen-printed electrode is ready to use.

In this configuration the sensor is printed only upon one side. There are two options to achieve the back-to-back configuration. The first is that the above approach is repeated on the back side of the polyester substrate<sup>12</sup> or two electrode can be taken and placed back-to-back, herein the latter approach is utilised for simplicity. From this point onwards when the electrodes are back-to-back, a superscript “2” is introduced, such that in the case of a single microband electrode (bSPE) in the back-to-back configuration becomes denoted as b<sup>2</sup>SPE. Additional side-by-side experiments were performed with the microband electrodes printed in a side-by-side configuration as demonstrated in ESI Fig. 1.†

Square wave anodic stripping voltammetry (SWASV) was used throughout this work for the determination of lead(II) ions with a deposition potential of  $-1.2$  V. The following buffer solutions were utilised and explored in this work: solution A: contained a combination of 2.50 M ammonium acetate, 1.55 M acetic acid and 0.02 M phenol in ethanol solution;<sup>17</sup> solution B: contained a combination of 2.50 M ammonium acetate and 1.55 M acetic acid;<sup>17</sup> solution C: 0.10 M sodium acetate buffer solution (pH 4.5) and solution D: 0.02 HCl solution (pH 1.7). Drinking water was obtained from a drinking water tap (Manchester City Centre, Manchester, UK) which was run for a minute before a sample being obtained. The sample was then stored at room temperature and used within a day of sampling. Prior to electroanalytical measurements the drinking water samples were simply modified to pH 1.7.

Inductively Coupled Plasma/Optical Emission Spectrometry (ICP-OES) experiments were carried out using a Thermo Scientific DUO iCAP 6300 ICP Spectrometer, exhibiting a relative standard deviation of 2.7%.

## 3. Results and discussion

First the optimised solution characteristics for the anodic stripping voltammetric determination of lead(II) ions are explored. Four solutions of differing compositions (see Experimental section) were utilised using the b<sup>2</sup>SPE with a fixed concentration of 2000  $\mu\text{g L}^{-1}$  of lead(II) ions and a deposition potential and time of  $-1.2$  V and 60 seconds respectively. In this approach the lead(II) ions are accumulated in the form of lead (0) through the application of the negative (deposition) potential at a selected time following which, the potential is swept positive. This process results in the electrochemically deposited lead metal upon the electrode surface to be stripped back to lead(II) ions giving rise to a voltammetric stripping peak, the analytical signal, where the peak height (and area) of the response is proportional to lead(II) ion concentrations.<sup>18</sup> It was found that a distinctive stripping peak is observed at a peak potential of  $\sim -0.60$  V (vs. Ag/AgCl); such responses are typical of that seen within literature.<sup>19–24</sup> The average current density (taken from the peak current/electrode geometric area) for the lead stripping peak was found to correspond to 13.60  $\mu\text{A cm}^{-2}$  (solution A), 417.5  $\mu\text{A cm}^{-2}$  (solution B), 542.6  $\mu\text{A cm}^{-2}$  (solution C) and 1037  $\mu\text{A cm}^{-2}$  (solution D).



These results show that the optimal solution for the electroanalysis of lead(II) ions is solution D (0.02 HCl solution; pH 1.7) which is used herein.

Further optimisation was next performed in order to find the optimal deposition time utilising square wave anodic stripping voltammetry (SWASV) using  $5 \mu\text{g L}^{-1}$  of lead(II) ions in a 0.02 M HCl solution (pH 1.7). This concentration is chosen since as it is below that indicated by the WHO where lead(II) levels within drinking water are recommended to be limited to  $10 \mu\text{g L}^{-1}$ .<sup>25</sup> Optimisation of the deposition times were next explored using both the bSPE and b<sup>2</sup>SPE. It became apparent that the optimum results are obtained when a deposition time of 30 seconds is employed with further deposition times found to result in a plateauing of the observed current density response. Through the application of a deposition time of 30 seconds, the current density is found to correspond to 3.40 and  $17.55 \mu\text{A cm}^{-2}$  utilising the bSPE and b<sup>2</sup>SPE configurations respectively. Such values indicate that the b<sup>2</sup>SPE exhibits a  $\sim 5$  times improvement over that of a single bSPE indicating the advantageous use of the back-to-back configuration.

The analytical efficacy of the b<sup>2</sup>SPE configuration was next explored towards the sensing of lead(II) ions, utilising SWASV and compared to a single microband (bSPE). Fig. 2 depicts the response from additions of lead(II) ions made over the concentration range of  $5\text{--}110 \mu\text{g L}^{-1}$  using both electrode configurations. Analysis of the SWASV profiles in the form of plots of peak height ( $I_H$ ) vs. concentration are found to be linear over the concentration range with the following linear regression: bSPE:  $I_H/\mu\text{A} = 8.00 \times 10^{-3} \mu\text{A} \mu\text{g}^{-1} \text{L}^{-1} + 2.90 \times 10^{-3} \mu\text{A}$ ;  $R^2 = 0.99$ ;  $N = 10$ ; b<sup>2</sup>SPE:  $I_H/\mu\text{A} = 0.06 \mu\text{A} \mu\text{g}^{-1} \text{L}^{-1} + 0.31 \mu\text{A}$ ;  $R^2 = 0.99$ ;  $N = 10$ . Analysis of the current density using both electrode configurations are depicted in Fig. 3 where the b<sup>2</sup>SPE exhibits a greater analytical response over the bSPE towards the detection of lead(II) ions over the concentration range used. The limit of quantification (LOQ;  $10\sigma$ ) was found to correspond to  $5.00 \mu\text{g L}^{-1}$  for both cases, with values for the limit of detection (LOD) ( $3\sigma$ ) for the b<sup>2</sup>SPE showing a  $\sim 3$  times improvement in comparison to the bSPE, with values corresponding to 1.01 and  $3.70 \mu\text{g L}^{-1}$  respectively.

The improvement in the current density through the use of the back-to-back configuration is a key advantage of using this

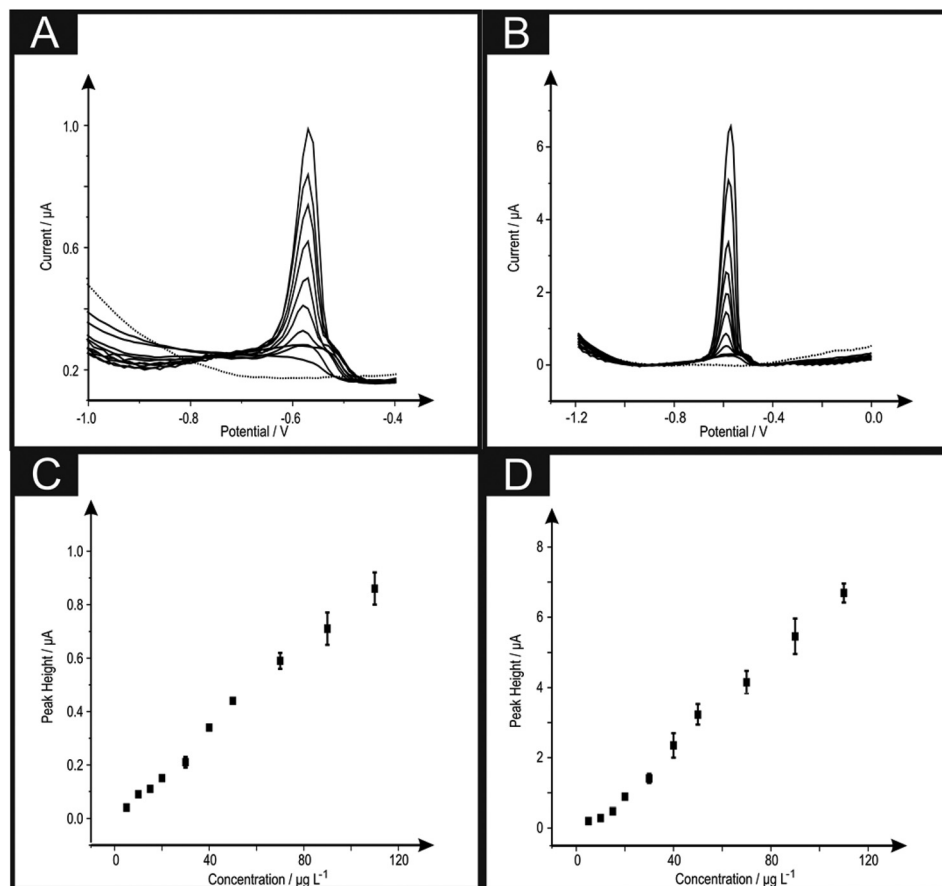
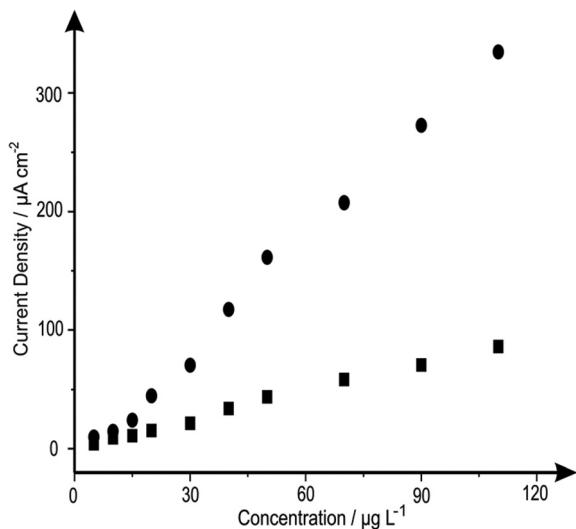


Fig. 2 Square wave voltammograms obtained in a solution of 0.02 M (pH 1.7) HCl in the absence (dotted line) and increasing concentration additions of lead(II) ions ( $5$  to  $110 \mu\text{g L}^{-1}$ ) and corresponding calibration plots over the range studied using bSPEs (A + C) and b<sup>2</sup>SPEs (B + D) respectively. Data presented is an average and error bars from three experiments where for each measurement a new electrode was utilised. Deposition potential and time of  $-1.2$  V and 30 seconds respectively.





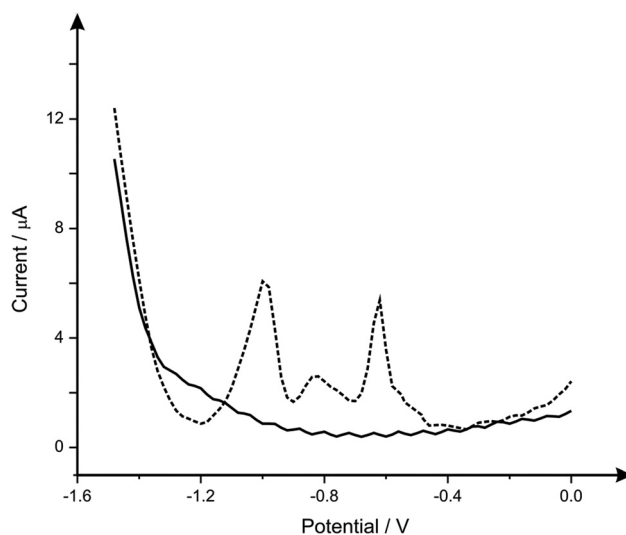
**Fig. 3** Analysis of the data presented in Fig. 2 in terms of plots of current density (of the peak heights) against increasing concentrations of lead(II) ions utilising the bSPEs (squares) and b<sup>2</sup>SPEs (circles). Deposition potential and time:  $-1.2$  V and 30 seconds respectively.

novel geometry. The reason for the improvement was thought that in the use of the back-to-back design, the electrode area is consequently doubled without resulting in any increase in unwanted capacitive currents, as would be observed if simply a larger electrode area was utilised, with improvements in the analytical performance observed with the analytical sensitivity (gradient of a plot of peak height/analytical signal against concentration) and the corresponding limit-of-detection being reduced.<sup>12</sup> The microband electrodes are advantageous since due to their geometric shape, have an additional contribution from radial diffusion in addition to planar diffusion. This change results in enhanced mass transport of electroactive species to the electrode surface, reduced double-layer capacitance, and less susceptibility to ohmic losses.<sup>18</sup> These characteristics make it possible to perform analysis with enhanced sensitivity on short time scales under time independent conditions. In the case of the back-to-back configuration, we are electrically wiring two microband electrodes back-to-back and hence this induces an additional improvement in the sensitivity of the electrodes performance. Effectively our configuration acts akin to a microband array and this is exemplified earlier in this paper where a five times improvement in the current density is evident for the sensing of lead(II) ions using the back-to-back configuration over that of a single microband electrode. This results in improvements in the analytical performance with a greater sensitivity observed (see Fig. 3) with a lower limit of detection achievable for the case of the b<sup>2</sup>SPE over that of the bSPE.

It is also important to note that as previously mentioned by Metters *et al.*<sup>12</sup> the back-to-back configuration allows for a perfect electrode configuration where diffusion zones do not overlap or interfere with each other which would be the case if two electrodes were wired in unison and placed in the solution

side-by-side (see ESI Fig. 1†). To prove this insight for the sensing of lead(II) ions, a comparison between the side-by-side (see ESI Fig. 1†) and the back-to-back configuration (see Fig. 1) was explored. ESI Fig. 2A† shows comparative SWASV of both configurations at a concentration of lead(II) ions at  $100 \mu\text{g L}^{-1}$ , it is clear that the peak height in the case of the side-by-side configuration is significantly hindered compared to that of the back-to-back. Additionally shown in ESI Fig. 2B† are the resultant calibration curves for the side-by-side configuration over the lead(II) ion concentration range of  $100$ – $598 \mu\text{g L}^{-1}$ , the range of which is limited by this configuration. These results are further verified by the vast difference within the LOD ( $3\sigma$ ) for the side-by-side configuration compared to the back-to-back configuration, with values corresponding to  $31$  and  $1.1 \mu\text{g L}^{-1}$  respectively. Note the high LOD and large error bars are a result of diffusional zones between the working electrodes of the side-by-side overlapping where the microbands electrode deplete the same region of solution.<sup>26–28</sup> In the case of the back-to-back electrodes, diffusional zones will likely never interact which is an advantage of using this novel electrode configuration.

It is important when analysing heavy metals that the response you are obtaining is that of the metal you desire, therefore the analysis of common interferences of lead(II), cadmium(II) and zinc(II) are next considered. Fig. 4 depicts the response obtained from a b<sup>2</sup>SPE sensor, for the simultaneous detection of lead(II), cadmium(II) and zinc(II) where it is clear that three separate peaks are observed at  $-0.6$  V,  $-0.8$  V and  $-1.0$  V respectively. Such responses are in agreement with literature concerning the simultaneous detection of these analytes upon carbon electrode.<sup>29–31</sup> Upon analytical analysis (ESI Fig. 3†) of the cadmium and zinc the lower linear range was utilised to calculate the LOD which was found to correspond



**Fig. 4** Square wave voltammograms obtained using the b<sup>2</sup>SPE within  $0.02$  M (pH  $1.7$ ) HCl (solid line) and the simultaneous detection of lead(II), cadmium(II) and zinc(II) at concentrations of  $8$ ,  $16$  and  $36 \mu\text{g L}^{-1}$  respectively (dashed line). Deposition potential and time of  $-1.5$  V and  $120$  seconds respectively.



to 0.3 and 32  $\mu\text{g L}^{-1}$  respectively, which for cadmium is very competitive however this system is not ideal for zinc ion detection compared to many others;<sup>32</sup> clearly the  $\text{b}^2\text{SPE}$  configuration can potentially be used for the simultaneous sensing of lead(II), cadmium(II) and zinc(II), however in this case we solely examine the effect of interference not analytical competency. Returning to the analytical performance of the  $\text{b}^2\text{SPE}$  sensor, its response is benchmarked against the current literature and against the WHO recommended limit, as presented in Table 1. It is clear that the  $\text{b}^2\text{SPE}$  sensor is competitive against other electrochemical configurations. What is of interest is that in the majority of cases, lengthy deposition times are utilised and very few are applied or validated in real samples. It is this we next address.

The detection of lead(II) ions within drinking water was next explored using the  $\text{b}^2\text{SPE}$ ; real sample analysis was undertaken to see if the proposed electroanalytical protocol has any potential interferents. Using SWASV the determination of lead(II) ions within drinking water *via* the standard addition protocol was performed. A typical standard addition plot and voltammograms are shown in Fig. 5. The fitting of the data in Fig. 5 revealed the following linear regression:  $I_{\text{H}}/\mu\text{A} = 0.03 \mu\text{A} \mu\text{g}^{-1} \text{L}^{-1} + 0.08 \mu\text{A}$ ;  $R^2 = 0.99$ ;  $N = 10$ . The concentration of lead(II) ions within the drinking water sample was calculated

from the lower linear range resulting from the standard addition plot and was found to be 2.8 ( $\pm 0.3$ )  $\mu\text{g L}^{-1}$ . The obtained value was compared with Inductively Coupled Plasma-Optical Emission Spectrometry (ICP-OES) (further details can be found in the Experimental section). Results obtained *via* ICP-OES correspond to 2.5 ( $\pm 0.1$ )  $\mu\text{g L}^{-1}$ . The excellent agreement between the proposed electrochemical protocol and independent laboratory analysis indicates the usefulness of the proposed electrochemical sensing protocol.

## 4. Conclusions

We have reported the first example of using the back-to-back electrode configuration towards the sensing of heavy metal ions using SWASV. It is found that in comparison to a single commonly utilised microband electrode that the back-to-back configuration allows a five times improvement in the analytical sensitivity over that of a single microband, and thus create an excellent alternative to that of a standalone microband electrode setup. Proof-of-concept is demonstrated towards the sensing of an unknown concentration of lead(II) ions found within a sample of drinking water, which upon utilisation of the standard addition method presented a concentration

**Table 1** Comparison of the current state-of-the-art electrochemical techniques for the electroanalytical detection of lead(II) ions

| Electrode of choice                               | Limit of detection ( $3\sigma$ )/( $\mu\text{g L}^{-1}$ ) | Linear range/ ( $\mu\text{g L}^{-1}$ ) | Deposition time/s | Comments   | Ref.             |
|---|---|--|-------------------|--|------------------|
| WHO level in drinking water                       | N/A   | 10                                     | N/A               | WHO recommended level of lead in drinking water  | 14               |
| GC  | 18.0  | 100–400                                | 120               | Simultaneous detection of lead(II) and cadmium(II) in a 0.1 M acetate buffer solution using a bismuth film electrode.  | 33               |
|   | 11.0  |  |                   |  |                  |
|   | 2.30  | 20–100                                 | 300               | Simultaneous detection of lead(II) and cadmium(II) with an introduction of $\text{K}_4[\text{FeCN}_6]$ within a 0.1 M acetate buffer solution, using bismuth film electrode.                                 | 34               |
|   | 1.50  |  |                   | Limits of detection revealed to be below that stated by WHO.   |                  |
|   | 0.80  | 5–60                                   | 600               | Simultaneous detection of lead(II) and cadmium(II) using bismuth nanoparticles upon the working electrode, within a tap water solution, results were validated with ICP-MS.                                  | 20               |
| CPE   | 2.30  | 1.5–450                                | 240               | Simultaneous detection of lead(II), cadmium(II) and copper(II) with analytical applications within tap water. Results validated by AAS.  | 35               |
|   | 0.45  | 50–200                                 | 180               | Simultaneous detection of lead(II), cadmium(II) and copper(II) and zinc(II) in HCl.  | 36               |
| Amino-functionalized metal-organic frameworks SPE | 1.04  | 2–70                                   | 300               | Lead(II) detection using a $\text{NH}_2\text{-Cu}_3(\text{BTC})_2$ modified GCE, in a 0.1 M acetate buffer solution.   | 19               |
|   | 0.03  | 0.05–30                                | 300               | Modified porous bismuth SPE demonstrates simultaneous detection of lead(II) and cadmium(II), in real water samples, with the porous electrode offering higher sensitivity due to the increased surface area. | 37               |
|   | 5.00  | 16.8–62.6                              | 120               | <i>In situ</i> modified antimony SPE showing detection of lead(II) in a 0.1 M acetate buffer solution.   | 38               |
|   | 2.00  | 10–100                                 | 120               | Detection of lead(II) within surface waters and validated with ICP-AES using a bismuth film SPE.   | 39               |
|   | 0.91  | 2.5–100                                | 120               | Detection of lead(II) within a solution of HCl using acetamide phosphonic acid self-assembled monolayer on a mesoporous silica modified SPE.   | 40               |
| <b><math>\text{b}^2\text{SPE}</math></b>          | <b>1.10</b>   | <b>5–110</b>                           | <b>30</b>         | <b>Detection of lead(II) within drinking water samples validated against ICP-OES, using an intelligent design which allows for a shorter deposition time compared to current literature.</b>                 | <b>This work</b> |

GC: glassy carbon; CPE: carbon paste electrode; SPE: screen-printed electrode; AAS: atomic absorption spectroscopy.



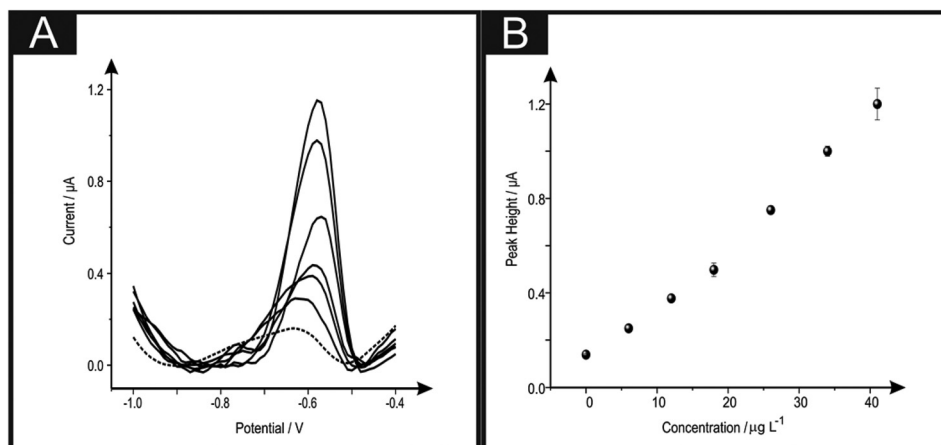


Fig. 5 Square wave voltammograms obtained using different b<sup>2</sup>SPEs (A) in a drinking water sample in the absence (dashed line) and the result of increasing additions of lead(II) ions (solid lines). Standard addition plot (B) for the lead(II) ions within drinking water with corresponding errors bars. (*N* = 3) A new electrode is used for each addition. Deposition potential and time of  $-1.2$  V and 30 seconds respectively.

which upon independently verification with ICP-OES, was found to be the same. This novel screen-printed back-to-back electrode configuration shows extreme promise for the generic sensing of heavy metal ions.

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## References

- J. P. Metters, R. O. Kadara and C. E. Banks, *Analyst*, 2011, **136**, 1067.
- J. P. Hart and S. A. Wring, *Electroanalysis*, 1994, **6**, 617.
- M. Li, Y. Li, D. Li and Y. Long, *Anal. Chim. Acta*, 2011, **734**, 31.
- A. S. Kumar and J. Zen, *Electroanalysis*, 2002, **14**, 671.
- K. C. Honeychurch and J. P. Hart, *Trends Anal. Chem.*, 2003, **22**, 456.
- C. W. Foster, J. P. Metters and C. E. Banks, *Electroanalysis*, 2013, **25**, 2275–2282.
- C. W. Foster, J. P. Metters, D. K. Kampouris and C. E. Banks, *Electroanalysis*, 2014, **26**, 262–274.
- J. P. Metters, R. O. Kadara and C. E. Banks, *Sens. Actuators, B*, 2012, **169**, 136–143.
- R. O. Kadara, N. Jenkinson and C. E. Banks, *Electrochem. Commun.*, 2009, **11**, 1377–1380.
- F. Tan, J. P. Metters and C. E. Banks, *Sens. Actuators, B*, 2013, **181**, 454–462.
- J. P. Metters, R. O. Kadara and C. E. Banks, *Analyst*, 2012, **137**, 896.
- J. P. Metters, E. P. Randviir and C. E. Banks, *Analyst*, 2014, **139**, 5339–5349.
- R. A. Goyer, *Toxicology of metals - biochemical aspects*, Springer-Verlag, New York, 1995.
- WHO, *Guidance for Drinking Water Quality, Recommendations*, Geneva, 2nd edn, 1993, vol. 1.
- C. W. Foster, J. P. Metters, D. K. Kampouris and C. E. Banks, *Electroanalysis*, 2014, **26**, 262.
- F. E. Galdino, C. W. Foster, J. A. Bonacin and C. E. Banks, *Anal. Methods*, 2015, **7**, 1208–1214.
- E. Ghali and M. Girgis, *Metall. Trans. B*, 1985, **16**, 489–496.
- R. G. Compton and C. E. Banks, *Understanding Voltammetry*, Imperial College Press, 2007.
- Y. Wang, H. L. Ge, Y. C. Wu, G. Q. Ye, H. H. Chen and X. Y. Hu, *Talanta*, 2014, **129**, 100–105.
- D. Yang, L. Wang, Z. L. Chen, M. Megharaj and R. Naidu, *Microchim. Acta*, 2014, **181**, 1199–1206.
- V. B. dos Santos, E. L. Fava, N. S. de Miranda Curi, R. C. Faria and O. Fatibello-Filho, *Talanta*, 2014, **126**, 82–90.
- N. Wang and X. D. Dong, *Anal. Lett.*, 2008, **41**, 1267–1278.
- Y. Wu, N. B. Li and H. Q. Luo, *Sens. Actuators, B*, 2008, **133**, 677–681.
- D. Li, J. Jia and J. Wang, *Talanta*, 2010, **83**, 332–336.
- World Health Organisation, *Exposure to Lead: A major public health concern*, Public Health and Environment, WHO Document Production Services, Geneva, 2010.
- S. J. Hood, R. O. Kadara, D. K. Kampouris and C. E. Banks, *Analyst*, 2010, **135**, 76–79.
- S. J. Hood, D. K. Kampouris, R. O. Kadara, N. Jenkinson, F. J. del Campo, F. X. Munoz and C. E. Banks, *Analyst*, 2009, **134**, 2301–2305.
- O. Ordeig, J. del Campo, F. X. Muñoz, C. E. Banks and R. G. Compton, *Electroanalysis*, 2007, **19**, 1973–1986.
- J. F. van Staden and M. C. Matoetoe, *Anal. Chim. Acta*, 2000, **411**, 201–207.



- 30 J. Gardiner and M. J. Stiff, *Water Res.*, 1975, **9**, 517–523.
- 31 I. Rutyna and M. Korolczuk, *Sens. Actuators, B*, 2014, **204**, 136–141.
- 32 M. Á. G. Rico, M. Olivares-Marín and E. P. Gil, *Talanta*, 2009, **80**, 631–635.
- 33 J. Saturno, D. Valera, H. Carrero and L. Fernandez, *Sens. Actuators, B*, 2011, **159**, 92–96.
- 34 G.-H. Hwang, W.-K. Han, J.-S. Park and S.-G. Kang, *Sens. Actuators, B*, 2008, **135**, 309–316.
- 35 H. Lin, M. Li and D. Mihailovič, *Electrochim. Acta*, 2015, **154**, 184–189.
- 36 H. Devnani, D. S. Rajawat and S. P. Satsangee, *Proc. Natl. Acad. Sci., India, Sect. A*, 2014, **84**, 361–370.
- 37 C. Chen, X. Niu, Y. Chai, H. Zhao and M. Lan, *Sens. Actuators, B*, 2013, **178**, 339–342.
- 38 V. Sosa, C. Barceló, N. Serrano, C. Ariño, J. M. Díaz-Cruz and M. Esteban, *Anal. Chim. Acta*, 2015, **855**, 34–40.
- 39 H.-L. Fang, H.-X. Zheng, M.-Y. Ou, Q. Meng, D.-H. Fan and W. Wang, *Sens. Actuators, B*, 2011, **153**, 369–372.
- 40 W. Yantasee, L. A. Deibler, G. E. Fryxell, C. Timchalk and Y. Lin, *Electrochem. Commun.*, 2005, **7**, 1170–1176.

