

Switchable fluorescence by click reaction of a novel azidocarbazole dye†

Cite this: *RSC Adv.*, 2014, 4, 11528Anna Hörner,^{‡a} Daniel Volz,^{‡ad} Tobias Hagendorn,^a Daniel Fűrnis,^a Lutz Greb,^a Franziska Röncke,^c Martin Nieger,^b Ute Schepers^c and Stefan Bräse^{*ac}Received 25th December 2013
Accepted 31st January 2014

DOI: 10.1039/c3ra47964a

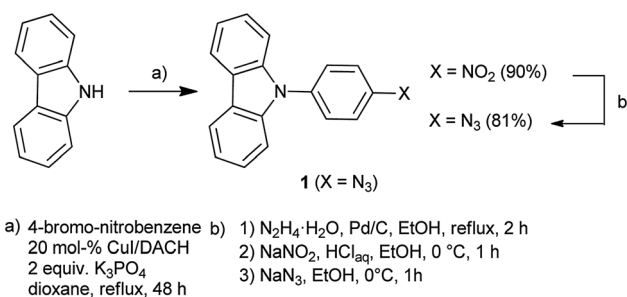
www.rsc.org/advances

Imaging is – even these days – still restricted to of a few classes of robust dyes. A demand for switchable tags led us to the design of a new class of pre-fluorophores. We achieved this by using a non-fluorescent *N*-(4-azidophenyl)-carbazole tag which turns fluorescent by click reaction with alkynes and cyclooctynes. The spectral properties of the labelled dyes were investigated. Our results suggest that a twisted internal charge transfer (TICT) transition is responsible for the emission. DFT calculations and single-crystal X-ray diffraction of selected examples support this explanation. The feasibility of the new dyes for biological application has also been tested *via* confocal microscopy.

Introduction

Carbazole moieties are a structural key motif in both dyes and luminophores. Pigments like Violet 23 or Hidrotin Blue 2R are used *e.g.* for the staining of clothing and strongly benefit from carbazole units in terms of photostability and high absorbance.^{1,2} Many fluorescent substances featuring carbazole subunits are known.^{3–6} A strong, deep blue luminescence is characteristic for most carbazole derivatives, making them broadly used in luminescent devices such as organic light-emitting diodes.^{7–9} However, because of the π - π^* and n - π^* transitions occurring upon excitation of the carbazole core, Stokes shifts for carbazole-substituted compounds are usually small. This hampers the discrimination between excitation and emission when using them as dyes in fluorescence microscopy and biological applications.

To resolve this issue, we attempted to design a new fluorescent label, combining the beneficial properties of carbazole such as high photostability and extinction, with the large Stokes shift of a charge-transfer (CT) system. CT systems typically consist of a donor-moiety, which may be excited, and a matching acceptor-unit, to which the electron density is shifted.

Scheme 1 Synthetic route to *N*-(4-azido-phenyl)-carbazole 1.

Typically, this leads to broad, ill-structured spectra as well as large Stokes shifts. In our case, the carbazole unit acts as a donor-unit, while aryl-substituted 1,2,3-triazoles are used as acceptors (Schemes 1–3).

Here, we report the synthesis and the photophysical properties of new carbazole dyes. Single-crystal X-ray diffraction revealed the molecular structure, while DFT- and TD-DFT calculations provided valuable insights into the photophysical mechanism leading to light-emission and a large Stokes shift. The dyes themselves are formed by Click-functionalization of a non-fluorescent precursor, *N*-(4-azidophenyl)-carbazole 1, which is non-fluorescent due to vibronic quenching by the free azido-group. This enables an easy discrimination of labelled material and traces of remaining, non-reacted precursor, *e.g.* in fluorescence microscopy.

Synthesis of precursor 1 and the fluorescent dyes

The synthesis of the precursor 1 is summarized in Scheme 1. After an Ullmann-type coupling of a straight carbazole with

^aDepartment of Chemistry, Karlsruhe Institute of Technology, Fritz-Haber-Weg 6, 76131 Karlsruhe, Germany. E-mail: stefan.braese@kit.edu

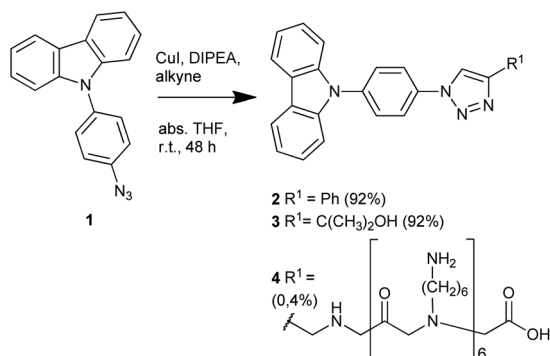
^bDepartment of Chemistry, University of Helsinki, P.O. Box 55 FIN-00014 University of Helsinki, Finland. E-mail: martin.nieger@helsinki.fi

^cInstitute of Toxicology and Genetic, Karlsruhe Institute of Technology, Campus North, Hermann-von-Helmholtz-Platz 1, 76344 Eggenstein-Leopoldshafen, Germany. E-mail: ute.schepers@kit.edu

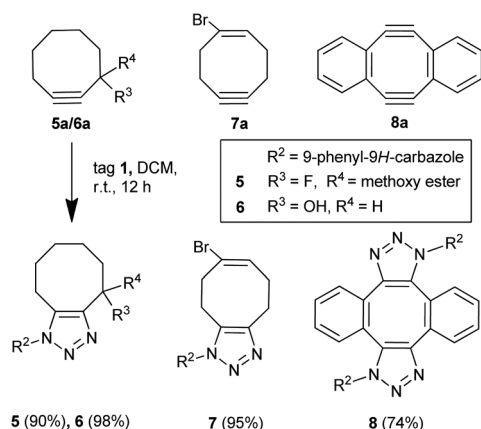
^dCynora GmbH, Hermann-von-Helmholtz-Platz 1, 76344 Eggenstein-Leopoldshafen, Germany

† Electronic supplementary information (ESI) available: Characterisation, spectra, computation method and X-ray crystallographic information files for compounds 2 and 3 (CIF). CCDC 936576 and 936577. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c3ra47964a

‡ Both authors contributed equally.



Scheme 2 Reaction of *N*-(4-azido-phenyl)-carbazole **1** with terminal alkynes.



Scheme 3 Reaction of *N*-(4-azidophenyl)-carbazole **1** with cyclooctynes.

4-bromo-nitrobenzene, the resulting *N*-(4-nitrophenyl)-carbazole is reduced with hydrazine, diazotized with HNO_2 and converted to *N*-(4-azidophenyl)-carbazole **1** by reaction with sodium azide. Further functionalization of non-fluorescent *N*-(4-azidophenyl)-carbazole **1** with terminal alkynes in a Cu(I)-catalysed alkyne-azide cycloaddition (CuAAC) (Scheme 2) as well as with cyclooctynes (Scheme 3) yielded the fluorescent carbazole dyes.^{10,11}

For the CuAAC click reaction, we chose copper(I) iodide in *N,N*-diisopropylethylamine (DIPEA) as the catalyst and several alkyne moieties such as the commercially available phenyl-acetylene as well as 2-methyl-3-butyn-2-ol, yielding the fluorescent dyes **2** and **3**, respectively. High conversion was achieved for both examples (92% yield each) within 4 hours. Furthermore, azide **1** was coupled to a polycationic peptoid containing an alkyne side chain on solid phase in a submonomer approach under the same conditions (see ESI†).^{12,13} This peptoid is a heptameric compound with six aminohexyl- and one propargyl-side chain and was used to investigate the influence of charged moieties on the carbazol fluorescence.^{14,15}

In contrast to the CuAAC reaction, the reaction between azides and cyclooctynes does not require any catalysts at room temperature due to the strained ring structure of cyclic alkyne. This makes the strain-promoted azide-alkyne cycloaddition

(SPAAC)¹⁶ suitable for click reactions in the presence of cells or whole organisms. All reactions depicted in Scheme 3 were carried out by simply dissolving the reactants (azide **1** and one equivalent of cyclooctynes **5a–8a**) in dichloromethane (DCM) and stirring overnight to yield dyes **5–8** in good to very high yields (74–98%).

Single-crystal X-ray diffraction

We were able to characterize molecules **2** and **3** by single-crystal X-ray diffraction (Fig. 1 and ESI†).¹⁷ The most striking observation for both structures was the twisting of all three π -systems, which breaks the conjugation between the three ring systems into electronically independent localized moieties. The smallest angle between the triazole ring and the phenyl ring is 29.9° for component **2** and 38.7° for component **3**. Between the carbazole moiety and the phenyl ring, an angle of 49.4° for component **2** and 66.2° for component **3** has been found. The molecular structures of **2** and **3** have also been used as starting structures of DFT-calculations and to interpret the photo-physical properties (see below).

Absorption spectroscopy

It is tempting, yet illegitimate to interpret a twist of the π -systems in solution from the results found in single-crystals without further evidence. We thus recorded UV-Vis spectra in various solvents. In summary, absorption spectra of both the unreacted precursor **1** and the fluorescent compounds **2** to **8** were quite similar. Carbazole moieties are very strong chromophores, which dominate the UV-Vis spectra; Peaks varied only in a 4 nm range (Fig. 2, Table 1 and ESI†). Molar absorption coefficients were quite high and depended on the solvents (Table 1 and ESI†). Substitutions with aromatic rings and modifications of side groups did not lead to a bathochromic shift or strong changes in the absorption spectra (Fig. 2 and Table 1). This suggested that the twisting of the conjugation found in solid state by X-ray diffraction also occurred in solution. Subtle variations manifested themselves in the differences in height of the shoulders in the normalized absorption spectra. Due to the broken conjugation, the UV-Vis spectra may be

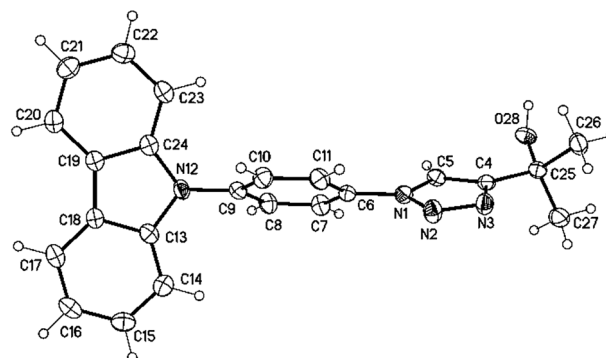


Fig. 1 Molecular structure of **3** (displacement parameters are drawn at 50% probability level).



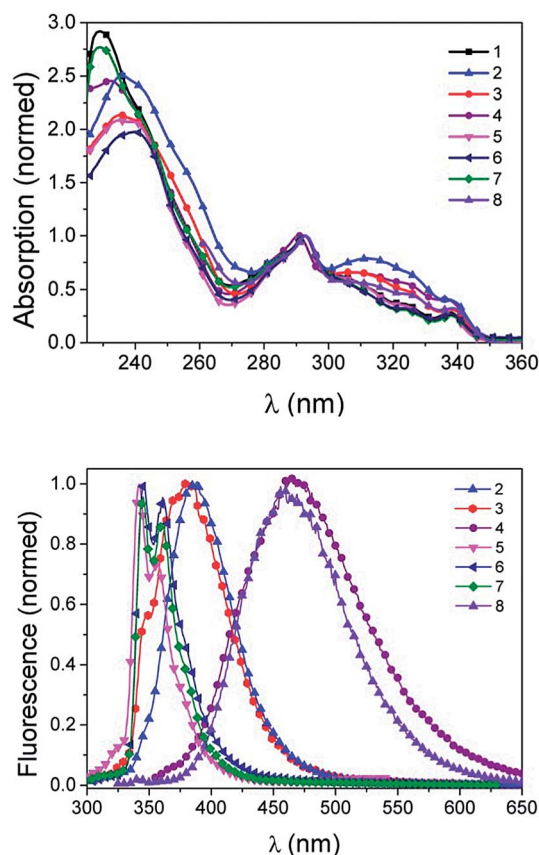


Fig. 2 Absorption (top) and emission (bottom) spectra of compounds 1–7 in DCM, 8 in DMSO. Excitation wavelength $\lambda_{\text{ex}} = 290$ nm.

Table 1 Spectroscopic data for the compounds at room temperature

	λ_{abs} (nm)	ϵ ($\text{M}^{-1} \text{cm}^{-1}$)	λ_{em} (nm)	ϕ	Stokes shift
1 ^b	293	21 300 \pm 600			
2 ^b	292	2400 \pm 2000	387	0.72 \pm 0.051	95
3 ^a	292		492	0.41 \pm 0.03	200
3 ^b	292	15 700 \pm 500	384	0.365 \pm 0.030	92
3 ^c	292	23 500 \pm 800	362	0.468 \pm 0.046	70
3 ^e	291	19 800 \pm 700	424		132
4 ^a	291		434		143
4 ^d	291	4000 \pm 200	467	0.027 \pm 0.001	176
4 ^e	291	3800 \pm 250	431	0.102 \pm 0.005	140
5 ^a	293		415		122
5 ^b	293	1950 \pm 50	342/356	0.311 \pm 0.081	49/63
6 ^a	293		490		197
6 ^b	293	19 600 \pm 900	345/360	0.319 \pm 0.040	52/67
7 ^a	293		492		199
7 ^b	293	12 400 \pm 300	345/360	0.284 \pm 0.058	52/67
7 ^c	291	32 000 \pm 7000	345/362	0.317 \pm 0.036	54/71
7 ^e	291	11 100 \pm 1000	431	0.347 \pm 0.049	140
7 ^g	292		341/357		49/65
7 ^h	291		342/357		51/66
7 ⁱ	291		344/360		53/69
7 ^j	291		345/362		54/71
8 ^a	293		416		123
8 ^f	293	28 000 \pm 3000	459	0.552 \pm 0.078	166

^a Solid, ^b DCM, ^c DMF, ^d H₂O, ^e MeOH, ^f DMSO, ^g Cyclohexane, ^h Diethyl ether, ⁱ Ethyl acetate, ^j DMF.

described as a superposition of the expected spectra of the isolated chromophores (phenyl, triazol, carbazole) with additional bands due to a charge-transfer transition from the carbazole to the aryl-triazole (300–350 nm).

Emission spectroscopy

To evaluate the strongest transition between absorption and emission 3D spectra of compound 7 in ethyl acetate and cyclohexane have been recorded. The result for cyclohexane is given in Fig. 3, while the corresponding plot for ethyl acetate is given in the ESI† For both solvents, 3D-spectra were similar and in agreement with the 2D-measurements (fluorescence over wavelength, see Fig. 2) over a range of excitation wavelengths. The strongest transition was found to occur at an excitation wavelength of 292 ± 1 nm. Excitation with lower energies (345 nm) did not lead to detectable luminescence. We thus define the strong transition at 292 ± 1 nm as the preferred excitation wavelength and refer the Stokes shifts to this transition.

Solvent- and polarity-influences on the emission

Emission spectra of the novel carbazole dyes were highly dependent on the solvent, with varying emission maxima between 341 nm and 492 nm (Table 2, Fig. 4).

However, differences between compounds 2–4 and 5–7 were noted probably due to structural variations. While compounds 2–4 have been reacted with terminal alkynes, compounds 5–8 have been reacted with cyclooctynes and contain bicyclic (5–7) and polycyclic (8) moieties. Stokes shifts were large, ranging from 49 nm to 200 nm and strongly solvent dependent (see Tables 1 and 2). The Lippert–Mataga-Plot, which was plotted for substance 7, also served to rationalize this (see Fig. 4). A linear correlation of the solvent orientation polarisability and the obtained Stokes shift was found (slope of the line and standard deviation in ESI†). The orientation polarisability of the

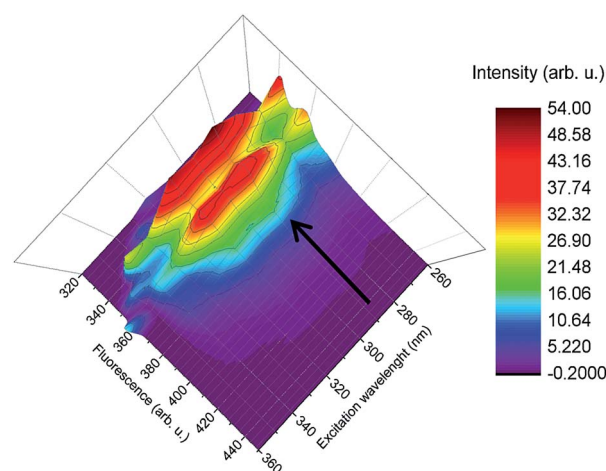


Fig. 3 3D fluorescence spectra of compound 7 in cyclohexane. $\lambda_{\text{ex}} = 260$ –370 nm, excitation steps in 10 nm difference, $\lambda_{\text{em}} = 320$ –450 nm.



Table 2 Solvent dependency for compound 7 in numbers^a

Solvent	Orientation polarizability (Δf)	Stokes shifts [nm]	Stokes shift 1 [10^4 cm^{-1}]	Stokes shift 2 [10^4 cm^{-1}]	Mean [10^4 cm^{-1}]
Cyclohexane	0	49/65	0.49	0.65	0.57
Diethyl ether	0.17	51/66	0.51	0.66	0.585
Ethyl acetate	0.201	53/69	0.53	0.69	0.61
DCM	0.217	52/67	0.52	0.67	0.595
DMF	0.274	54/71	0.54	0.71	0.625
Methanol	0.310	140	1.40		1.40

^a DCM = dichloromethane, DMF = dimethylformamide; calculation of stokes shifts see ESI. Δf numbers for methanol and ethyl acetate, as well as formula according to ref. 18.

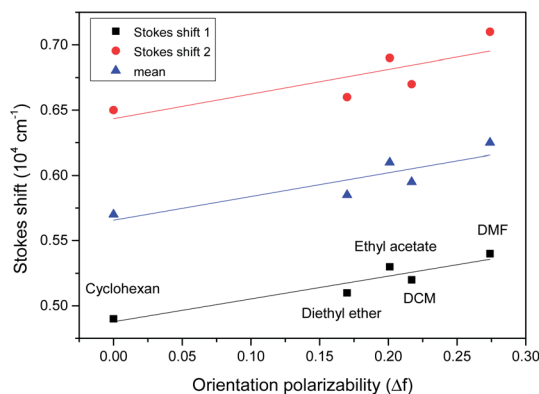
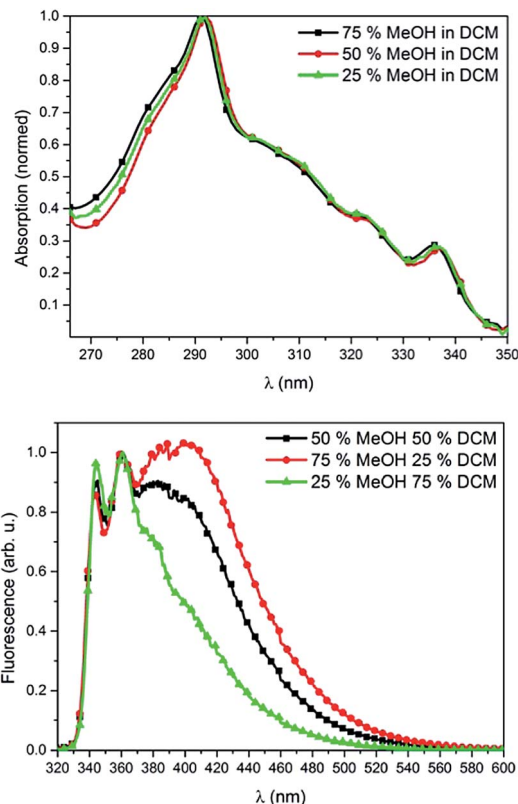


Fig. 4 Lippert–Mataga-plot of compound 7.

environment interacted with the dipole moment of the chromophore. This led to a shift of the emission maxima and consequently to the large stokes shifts.

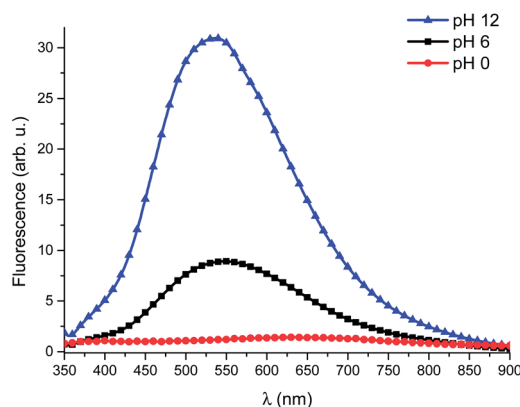
For methanol (MeOH), we noticed significant deviations from the fitted line in the Lippert–Mataga plot. Interestingly, no such effects were found in absorption spectra (Fig. 5). We recorded emission spectra of compound 7 in different mixtures of MeOH and DCM as solvents. In non-protic solvents such as DCM, two small band maxima were found, while an additional, broad band appeared when protic methanol was mixed with the non-protic solvent. When increasing the amount of protic solvent, the broad band became more pronounced.

This behaviour is indicative for the occurrence of different emissive states. The blue-shifted, structured transition could be attributed to carbazole-centered fluorescence, which was apparently not influenced by the surrounding solvents. The second, red-shifted transition was stabilized by protic solvents and was thus more pronounced when moving from pure DCM to pure MeOH. Under aqueous conditions, the carbazole-based emission was fully suppressed in favour of the CT-type band as shown for the water soluble compound 4. The fact that increasing polarity and the presence of protic solvents seem to cause a very large stokes shifts makes these dyes interesting candidates for biological applications.

Fig. 5 Absorption (top) and emission (bottom) spectra of compound 7 in solvent mixtures of MeOH and DCM. Excitation wavelength λ_{ex} = 290 nm.

pH-influences on the emission for compound 4

Compound 4 carried six amino groups and was therefore expected to be influenced by the pH value of the medium due to the introduction of positive charges in close proximity to the emissive chromophore. The absorption and emission spectra have been measured in phosphate buffered saline (PBS) under acidic, neutral and basic conditions (spectra see Fig. 6 and

Fig. 6 Fluorescence spectra of compound 4 in different pH, the intensities are recalculated to the lamp intensities and detection sensitivity. Excitation wavelength λ_{ex} = 290 nm.

ESI[†]). The emission spectra (Fig. 6) were pH sensitive: The band was shifted towards lower energies, when moving to more acidic solvents. This was accompanied by decreased quantum efficiency. Similar to the influences of protic solvents on compound 7, the pH-value did not seem to affect the absorption spectra of compound 4. We thus presume that the positive charge of the ammonium groups influenced the molecules in excited state, which is typical for CT emission. This decreased the intensity and shifted the emission to longer wavelength (normalized spectra see ESI[†]).

Determination of the quantum efficiency and emission decay time

Also measurements of the photoluminescence quantum yield (PLQY)¹⁹ were done in different solvents as well as in the solid state (Table 1). The PLQY values were quite high, despite the large Stokes shift. They mostly varied between 30% and 72%. An exception was compound 4 due to the high interactions with the protonated amino side chains, as stated above (ESI[†]). Using the time-correlated single photon counting (TCSPC) technique, the emission decay time of components 2 and 3 was measured in degassed DCM at room temperature. Both compounds featured a monoexponential decay with $\tau = 3$ and 4 ns, respectively. These lifetimes indicated a fluorescence emission without an involvement of triplet state (T_1). We can exclude delayed fluorescence as the cause for the broad spectra and large Stokes shifts in the solid state.

DFT calculations

Density functional theory (DFT) geometry calculations for compounds 2, 3 and 7 were performed on B3-LYP/def2-TZVP level^{20,21} using D3 dispersion corrections.²² Comparison of the calculated and measured bond lengths and angles (see ESI[†]) showed a good agreement for the applied functional. Representative frontier orbitals (HOMO and LUMO) for compound 3 are depicted in Fig. 7 (see ESI[†] for compounds 2 and 7). For all calculated compounds, the highest occupied molecular orbital (HOMO) is mainly localized at the carbazole moiety, whereas the lowest unoccupied molecular orbital (LUMO) is localized on the triazole and bridging phenyl ring. As seen for the ground state HOMO–LUMO energies in Table 3, HOMO energies remain basically unchanged (around -5.7 eV) with varying substituents at the triazole, whereas the LUMO energies are affected. Hence, it appears that major differences in the observed emission maxima of compounds 2, 3 and 7 may result from variations in the LUMO energies.

On the basis of these results, the type of electronic transition that led to the lowest excited states may be classified as being largely based on intramolecular charge-transfer character. This assumption was further supplied by gas phase time-dependent density functional theory (TD-DFT) calculations at the same level, which shows 99% participation of HOMO \rightarrow LUMO for the S_0 to S_1 vertical singlet excitation. However, it is known that exact DFT calculations of charge-transfer (CT) excitations are very difficult. Since all excitations show a distinct CT character,

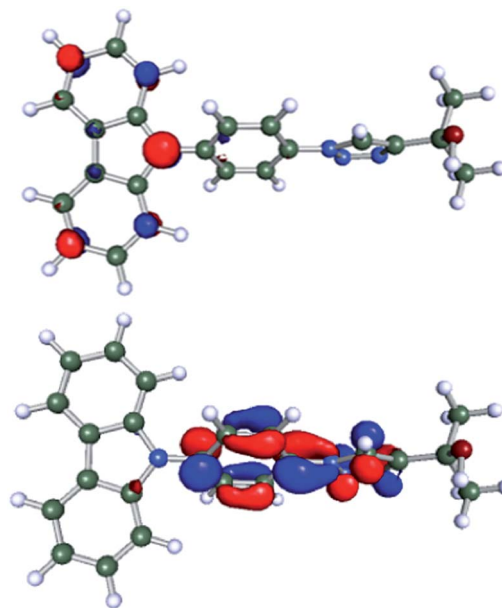


Fig. 7 HOMO (left) and LUMO (right) frontier orbital plots of compound 3.

the excited states and transition energies are not described quantitatively by simple application of TD-DFT. As found for compound 3 (Table 3), the energies of HOMO and LUMO are just slightly affected by varying solvent polarity (conductor-like screening model, COSMO).²³

Twisted internal charge transfer mechanism

All results indicates that a twisted internal charge transfer (TICT) as described by Wang *et al.*²⁴ is responsible for the emission of our new dyes. We believe that the strong polarity dependence of the Stokes shift originates from solvent dependent slow reorientations of the molecule dipole in the excited states. This is in good agreement with all photophysical measurements, the TCSPC measurements, X-ray results and DFT calculations. The reason for this is the polarity-dependent twist of the three connected π -systems of the carbazole, phenyl and triazole moieties.

Biological application

To verify that the specific conditions in biological media lead to a twisting conformation that allows for efficient light detection, we tested compounds 2 and 7 for their applicability in cell culture. Eventually, 10^4 human cervix carcinoma (HeLa) cells were treated with 30 μ M of 2 and 7, respectively.

Table 3 B3LYP/def2-TZVP HOMO and LUMO energies (in eV) at the ground state, left: of 2, 3, 7 (gas phase); right: of 3 with different solvents (COSMO)

	2	3	7	3 (CH ₂ Cl ₂)	3 (MeOH)
HOMO	−5.72	−5.71	−5.76	−5.70	−5.70
LUMO	−1.62	−1.52	−1.38	−1.42	−1.41



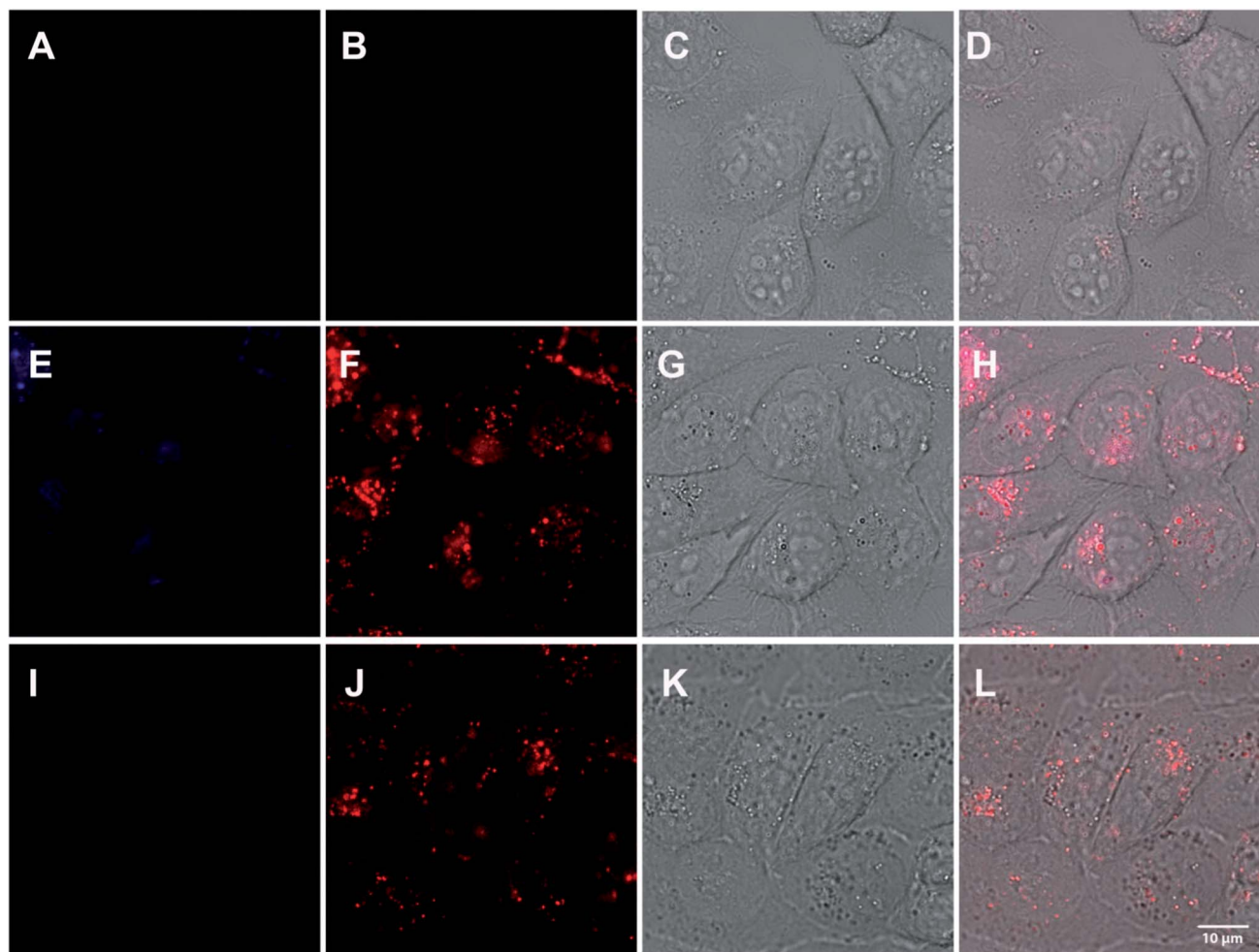


Fig. 8 10^4 HeLa cells were incubated with 30 μM of **2** (E–H) or **7** (I–L) in DMEM (20 mM stock solution in DMSO, freshly diluted 666 \times in DMEM) at 37 $^\circ\text{C}$ for 24 h and subsequently subjected to fluorescent confocal microscopy using a Leica SP5-TCS (DMI6000) inverse microscope. Objective: HCX PL APO CS 63.0 \times 1.20 WATER UV. Excitation wavelength: 351 nm. 1st column (A, E and I): Emission bandwidth: 417–468 nm, 2nd column (B, F and J): emission bandwidth: 518–724 nm, 3rd column (C, G and K): Brightfield, 4th column (D, H and L): Merge. A–D: Control cells, only DMSO, E–H: Compound **2**, I–L: Compound **7**.

Fluorescence confocal microscopy showed that in cells both compounds revealed a strong fluorescence with a bathochromic shift of the measured emission maxima towards 450–550 nm, when excited with a UV laser at 351 nm. Both compounds **2** and **7** showed a strong accumulation within the endosomal-lysosomal system (Fig. 8), which suggests an endocytotic uptake. A λ -scan resulted in maximum fluorescence emission at about 481 nm for **2** and 543 nm for **7**. While the bathochromic shift of compound **2** is less pronounced, the shift in **7** is in accordance with the fluorescence measurements in protic solvents with increasing polarity (Fig. 5).

As the pH of the endocytic pathway is decreasing from the plasmamembrane towards the lysosomal compartment (from 7.4 to 4.4), the red-shifted transition of compound **7** is shifted towards lower energies when moving to more acidic (and thus, more protic) compartments. This is even more pronounced in cells than in solution. Although the excitation wavelength is not optimal, the strong fluorescence emission of the compound promises an application in biological systems.

Conclusions

In this study, we present a new class of fluorescent carbazole dyes. We demonstrate that the non-fluorescent *N*-(4-azidophenyl)-carbazole moiety becomes fluorescent by click reaction with an alkyne or cyclooctyne. This specific property allows fluorescent control in samples after click reaction, for example in biological samples or material science. The spectral properties show that absorption spectra are not changed after click reaction with various substrates. The fluorescence of the resulting dyes results from a twisted internal charge transfer mechanism, which has been verified by detailed photophysical experiments, single-crystal X-ray diffraction and DFT-calculations. This leads to a high solvent dependency accompanied by large Stokes shifts from 49–200 nm. Most samples have a high quantum yield between 30 and 72%. The feasibility of the new dyes has been demonstrated in a first series of confocal microscopy experiments.



Acknowledgements

We acknowledge the Carl-Zeiss-Stiftung (A. H. and D. F.), Karlsruhe School of Optics and Photonics (KSOP) (A. H. and D. K. K.) and the Landesgraduiertenförderung Baden-Württemberg (T. H. and L. G.) for and financial support. We thank cynora GmbH for support. The work was further supported by the Helmholtz programme Biointerface (U. S. and S. B.). We also acknowledge the working group of Dr L. Fruk at the DFG-Center for Functional Nanostructures, Karlsruhe Institute of Technology for using their infrastructure.

Notes and references

- 1 Y. F. Lan and J. J. Lin, *Dyes Pigm.*, 2011, **90**, 21–27.
- 2 R. Levinson, P. Berdahl and H. Akbari, *Sol. Energy Mater. Sol. Cells*, 2005, **89**, 351–389.
- 3 W. Maes, T. H. Ngo, G. Rong, A. S. Starukhin, M. M. Kruk and W. Dehaen, *Eur. J. Org. Chem.*, 2010, 2576–2586.
- 4 M. Tavasli, T. N. Moore, Y. H. Zheng, M. R. Bryce, M. A. Fox, G. C. Griffiths, V. Jankus, H. A. Al-Attar and A. P. Monkman, *J. Mater. Chem.*, 2012, **22**, 6419–6428.
- 5 A. C. Grimsdale, K. L. Chan, R. E. Martin, P. G. Jokisz and A. B. Holmes, *Chem. Rev.*, 2009, **109**, 897–1091.
- 6 G. Moad, M. Chen, M. Haussler, A. Postma, E. Rizzardo and S. H. Thang, *Polym. Chem.*, 2011, **2**, 492–519.
- 7 S. Y. Takizawa, V. A. Montes and P. Anzenbacher, *Chem. Mater.*, 2009, **21**, 2452–2458.
- 8 Y. Heischkel and H. W. Schmidt, *Macromol. Chem. Phys.*, 1998, **199**, 869–880.
- 9 D. Volz, T. Baumann, H. Flügge, M. Mydlak, T. Grab, M. Bächle, C. Barner-Kowollik and S. Bräse, *J. Mater. Chem.*, 2012, **22**, 20786–20790.
- 10 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem.*, 2002, **41**, 2596–2599.
- 11 C. W. Tornøe, C. Christensen and M. Meldal, *J. Org. Chem.*, 2002, **67**, 3057–3064.
- 12 H. Jang, A. Fafarman, J. M. Holub and K. Kirshenbaum, *Org. Lett.*, 2005, **7**, 1951–1954.
- 13 R. N. Zuckermann, J. M. Kerr, S. B. H. Kent and W. H. Moos, *J. Am. Chem. Soc.*, 1992, **114**, 10646–10647.
- 14 D. K. Kölmel, D. Fürniss, S. Susanto, A. Lauer, C. Grabher, S. Bräse and U. Schepers, *Pharmaceuticals*, 2012, **5**, 1265–1281.
- 15 T. Schröder, K. Schmitz, N. Niemeier, T. S. Balaban, H. F. Krug, U. Schepers and S. Bräse, *Bioconjugate Chem.*, 2007, **18**, 342–354.
- 16 N. J. Agard, J. A. Prescher and C. R. Bertozzi, *J. Am. Chem. Soc.*, 2004, **126**, 15046–15047.
- 17 Structure solution and refinement program SHELXL-97: G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2008, **64**, 112–122.
- 18 J. R. Lakowicz, *Principles of fluorescence spectroscopy*, Springer, New York, 2nd edn., 1999, p. 189, table 6.2.
- 19 S. Fery-Forgues and D. Lavabre, *J. Chem. Educ.*, 1999, **76**, 1260–1264.
- 20 C. T. Lee, W. T. Yang and R. G. Parr, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1988, **37**, 785–789.
- 21 F. Weigend and R. Ahlrichs, *Phys. Chem. Chem. Phys.*, 2005, **7**, 3297–3305.
- 22 S. Grimme, J. Antony, S. Ehrlich and H. Krieg, *J. Chem. Phys.*, 2010, 132.
- 23 A. Klamt and G. Schüürmann, *J. Chem. Soc., Perkin Trans. 2*, 1993, 799–805.
- 24 J. Wang, Y. Wang, T. Taniguchi, S. Yamaguchi and S. Irle, *J. Phys. Chem. A*, 2012, **116**, 1151–1158.

