Influence of B-ring modifications on proton affinity, transmembrane anion transport and anti-cancer properties of synthetic prodigiosenes†

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Prodigiosin is the parent compound of the tripyrrolic natural products known as the prodigiosenes. Some of these natural products and their synthetic analogs show anti-cancer, immunosuppressive and antimicrobial actions, amongst other biological activities. One mechanism put forth to explain their biological activity is that since prodigiosenes are typically protonated at physiological pH they can alter intracellular pH via HCl co-transport (or Cl⁻/OH⁻ exchange) across cell membranes. In this study we synthesized a series of prodigiosene analogs with different O-aryl substituents attached to the B-ring of the tripyrrolic skeleton. NMR studies showed that these analogs can exist as a mixture of two stable α and β conformers in acidic solution, and that both conformers can bind anions in solution. We found that the electronic nature of the O-aryl substituent on the B-ring influences the rate at which these prodigiosenes catalyze transmembrane anion transport, i.e. the prodigiosenes with the higher pKₐ had greater Cl⁻/NO₃⁻ exchange rates. Four of the synthetic prodigiosenes were tested for their in vitro anti-cancer activities in the NCI60 human tumour panel. Despite their promising in vitro anti-cancer activity (GI₅₀ values ranging from 18 to 74 nM), there was no evidence that this activity is influenced by the extent of protonation of these synthetic prodigiosenes.

Introduction

In the last decade various compounds that catalyze transmembrane transport of anions have been identified.1 Much of this activity has been driven by supramolecular chemists interested in systems that work at the water-lipid interface.2 Challenges in transmembrane anion transport are to extract an anion from water and then move it across a hydrophobic membrane. Such transport is facilitated by (1) amphiphilic “carriers” that bind anions and diffuse through the membrane or (2) compounds that self-assemble into channels that span the membrane and provide a low-energy pathway.1,3 Efforts in this area have led to a better understanding of the factors that control anion transport. Thus, non-covalent interactions like hydrogen bonding, electrostatic effects, anion-π and halogen bonds have been used to effect anion transport.5 Recent work has also clarified how ligand conformation,4 flexibility,5 and lipophilicity6 can be used to catalyze transmembrane anion transport. Another important factor promoting this research is the possibility that anion transporters might be developed into therapeutics for diseases, like cystic fibrosis, that are caused by defective transport of Cl⁻.7

Some synthetic transporters have been developed after considering how nature catalyzes the process. For example, peptides that contain domains of the Cl⁻ channel can transport anions across lipid membranes,8 while modification of natural amphiphiles like cholic acid has provided potent anion carriers.9 The tripyrrolic prodigiosenes are known to catalyze transmembrane anion transport.10 The parent compound, prodigiosin (1), is a natural product isolated from Serratia marcescens (Fig. 1) that has much biological potential11 as it is able to permeabilize cell membranes, alter intracellular pH and trigger apoptosis.12 Early studies indicated that prodigiosin is too cytotoxic for healthy cells and, for this reason, it was not initially pursued as a drug candidate.13 There has been a renewed interest in prodigiosenes since some synthetic analogs have promising anti-cancer,14 immunosuppressive,11b,15 anti-malarial,16 and antimicrobial17 activities at concentrations below where they are cytotoxic to healthy cells.

As depicted in Fig. 1 protonated prodigiosin 1-H⁺ has a binding pocket for anions, using hydrogen bonds and electrostatic interactions for anion coordination. The resulting amphiphilic neutral salt 1-H⁺Cl⁻ can then readily diffuse
through the cell membrane. We have previously shown that prodigiosin 1·H⁺Cl⁻ catalyzes the transmembrane exchange of anions such as Cl⁻, NO₃⁻ and HCO₃⁻. Prodigiosenes also alter intracellular pH, presumably through catalyzing co-transport of H⁺Cl⁻ or the exchange of Cl⁻/OH⁻ across cell membranes. This ability to change intracellular pH is one potential mechanism by which prodigiosenes trigger apoptosis. Other possible modes of actions include stranded-DNA cleavage and protein kinase inhibition. However, some analogs without an OMe group demonstrated reasonable anti-cancer activity. For example, PNU-156804 5, with an OMe group, exhibited nM IC₅₀ values against leukemia and melanoma cells, and the synthetic analog 6 was active against lung cancer cell lines. In this study we prepared novel derivatives substituted with different –OAr groups on the B-ring, alongside common features at the A and C-ring, so as to probe the influence of electronic effects upon pKₐ values, transmembrane anion exchange rates and anti-cancer activity. This is, to our knowledge, the first report of prodigiosenes substituted with –OAr groups on the B-ring. We first required a reliable method to obtain the O-substituted pyrrolinones 8 (Scheme 1), as this moiety would become the B-ring of the ultimate prodigiosenes. Following a patented procedure the –OBN pyrrolinone 8a was successfully obtained from the –OME pyrrolinone 7 (Scheme 1). However, using phenol as a nucleophile instead of benzyl alcohol failed to give pyrrolinone 8b. Attempts using sodium phenoxyde were also unsuccessful (Conditions B).

To accomplish the key substitution with phenolates, we decided to use the –OTs pyrrolinone 9, bearing a competent leaving group (Scheme 2). By varying the nature and amount of the base and solvent, as well as the concentration of the pyrrolinone, we found a suitable method for the formation of prodigiosene analogs.

**Synthesis of B-ring modified prodigiosenes**

There are few reports regarding synthetic modification of the prodigiosene B-ring. Moreover, the impact of B-ring modifications on anti-cancer activity seems mixed. Indeed, the desmethoxy analog 4 (Fig. 3) was relatively inactive as an anti-cancer agent, as compared to the parent prodigiosin (1), suggesting that the OMe group is essential for anti-cancer activity. However, some analogs without an OMe group demonstrated reasonable anti-cancer activity. For example, PNU-156804 5, with an OMe group, exhibited nM IC₅₀ values against leukemia and melanoma cells, and the synthetic analog 6 was active against lung cancer cell lines. In this study we prepared novel derivatives substituted with different –OAr groups on the B-ring, alongside common features at the A and C-ring, so as to probe the influence of electronic effects upon pKₐ values, transmembrane anion exchange rates and anti-cancer activity. This is, to our knowledge, the first report of prodigiosenes substituted with –OAr groups on the B-ring.

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12-a condensation of aldehyde C-rings of the ultimate prodigiosene targets, Scheme 3). Aldol N-deprotection of the drawing or an electron-donating substituent. Acid-catalyzed regardless of whether the phenol contained an electron-withdrawing groups (when the O group in apparently, the presence of a strong electron-withdrawing group in hand we then prepared dipyrrinones $8j$ in excellent yields.

With the B-ring precursors $8$ in hand we then prepared dipyrrolines $12a-h$ (intermediates that contain the B- and C-rings of the ultimate prodigiosine targets, Scheme 3). Aldol condensation of aldehyde $11$ with pyrrolinones $8b-j$, promoted by TMSOTf and Et$_3$N, proceeded in good yield, except when the O-aryl group was substituted with strong electron-withdrawing groups ($8i$ R = Ph-pCN and $8j$ R = Ph-pNO$_2$). Apparently, the presence of a strong electron-withdrawing group in $8i$ and $8j$ makes the ether prone to hydrolysis or substitution, as we always detected the corresponding phenol in the crude mixture (TLC) after the reaction. Bromination of isomers $8i$ and $8j$ proceeds in moderate yields (47–62%), regardless of whether the phenol contained an electron-withdrawing or an electron-donating substituent. Acid-catalyzed deprotection of the N-Boc group then gave the pyrrolinones $8b$–$8j$ in excellent yields.

Transmembrane exchange of Cl$^-$ and NO$_3^-$ anions by prodigiosin (1) involves anion binding by the protonated transporter (1-H$^+$). This is followed by diffusion of this neutral complex $1$-H$^+$ across the lipid membrane, and Cl$^-$/NO$_3^-$ anion exchange at the lipid–water interface. The anion binding and transport processes are complicated by the fact that protonated prodigiosenes often exist as two isomers (Scheme 4). This $\alpha/\beta$ conformational equilibrium can vary as a function of ligand structure and environmental conditions. We investigated qualitative aspects of the solution-state conformation of the $\alpha$-Ph derivative $3b$-$H^+$. Our goals were (1) to determine whether $3b$-$H^+$ exists in both $\alpha$ and $\beta$ conformations in solution and (2) to obtain evidence that $3b$-$H^+$ undergoes anion exchange in solution. The complex $3b$-HCl (1 mM) showed one set of signals, consistent with the $\beta$ conformation, when initially dissolved in CDCl$_3$ (see spectra on p. 56 of ESI). However, after being in CDCl$_3$ solution over time the same sample showed two sets of $^1$H NMR signals, in an approximate 3:1 ratio. Based on analogy with previous NMR studies of other prodigiosenes in CDCl$_3$ by Rizzo et al. and by Quesada et al., we assigned the major set of NMR signals to the $\beta$ isomer of $3b$-HCl and the minor set of signals to the $\alpha$ isomer. We next investigated whether both conformers of $3b$-$H^+$ undergo anion binding and exchange in solution. Thus, we titrated methanesulfonic acid (MsOH) into a solution of $3b$-HCl in CDCl$_3$ and followed changes in $^1$H NMR signals for the pyrrolic NH protons ($\delta$ 12–13 ppm). We reasoned that changes in this region of the spectrum would reflect anion binding and exchange between chloride and methanesulfonate anions, since it is the pyrrolic NH atoms that hydrogen bond with the anion. Fig. 4 shows representative spectra from the titration. The NH region of the $^1$H NMR spectrum for the $3b$-HCl salt (Fig. 4a) indicates a mixture of isomers, but the NMR signals were poorly resolved. However, this overlap...
for the pyrrolic NH protons was removed upon the addition of MsOH to a solution of 3b·HCl (Fig. 4b). As shown in Fig. 4b, we observed two well-resolved sets of three pyrrolic NH signals for the 3b-HOMs complex. There are two important findings here. First, the NH chemical shifts change upon addition of the MsOH, suggesting that both the α and β isomers bind anions. Thus, as shown in Fig. 4b chemical shifts for the six NH protons moved upfield when MsOH was added to a solution of 3b·HCl. These changes in NH chemical shifts indicate anion exchange, as Cl$^-$ is replaced by the softer methanesulphonate anion in the binding site of the prodigiosene. Second, the α/β isomer ratio for 3b·HCl and 3b·HOMs does not change significantly with a change in bound anion (Fig. 4c). A 2D NOESY NMR experiment of the 3b-HOMs complex allowed us to assign signals for the three NH protons in the major β isomer and for the three NH protons in the α isomer. The NOE correlation that best supports our assignments can be observed between one of the minor pyrrole NH signals and the signal corresponding to the C2 methyl group of the α isomer (Fig. 5).

Determination of apparent $pK_a$ values for synthetic prodigiosenes

We hypothesized that prodigiosene basicity would correlate with its efficiency as an anion transporter, since the protonated prodigiosene 3b-H$^+$ is the likely transporter. The concept that transporter efficiency can be tuned by modulating the acidity of hydrogen bond donors has been demonstrated with other systems. To learn how B-ring substitution influences the acid-base properties of these synthetic prodigiosenes we used a spectrophotometric method, described by Manderville, to measure the apparent $pK_a$ values for five B-ring analogs: the –OPh derivative (3b-H$^+$), two analogs with electron-donating groups on the OAr ring (3d-H$^+$ R = Ph-pOMe, 3e-H$^+$ R = Ph-pNMMe$_2$), and two analogs with electron-withdrawing groups on the B-ring (3f-H$^+$ R = Ph-pCF$_3$, 3h-H$^+$ R = Ph-pCl).

In solution, protonated prodigiosenes are pink with absorbance maxima above 500 nm. The free-base absorbs below 500 nm. Fig. 6 shows representative pH-dependent spectra for 3d and 3f in 1:1 acetonitrile–water at 25 °C. Absorption maxima for the free-base of the Ph-pOMe analog 3d ($\lambda_{\text{max}}$ = 484 nm) and its protonated form 3d·H$^+$ ($\lambda_{\text{max}}$ = 541 nm) are well separated (Fig. 6a). The Ph-pCF$_3$ derivative showed similar properties, with $\lambda_{\text{max}}$ = 450 nm for the free-base 3f and $\lambda_{\text{max}}$ = 545 nm for 3f·H$^+$ (Fig. 6b). Furthermore, the ionization states of 3d·H$^+$ and 3f·H$^+$ change smoothly as a function of pH, both revealing isosbestic points. The apparent $pK_a$ values, which correspond to values for the equilibrium mixture of α and β isomers, for these B-ring prodigiosenes were determined from plots of the log(ionization ratio) vs. pH.

For this α isomer, the C-ring NH proton forms an intramolecular hydrogen bond with the –OPh group on the B-ring (Scheme 4). Presumably, this hydrogen-bonded NH proton exchanges much more slowly with residual water in the CDCl$_3$ solvent and, therefore, shows a NOE correlation with its neighbouring C2-methyl group.

These NMR titrations allowed us to conclude that (1) both the major β and minor α isomer of 3b-H$^+$ bind and exchange anions and (2) Cl$^-$ interacts more strongly than OMs$^-$ with 3b-H$^+$. We next determined the apparent $pK_a$ values for some O-aryl analogs with the intent to correlate their ease of protonation with their rates of transmembrane anion transport.
Table 1, analogs with electron-donating groups on the B-ring, namely 3d·H⁺ (R = Ph-pOMe, pKₐ = 7.3) and 3e·H⁺ (R = Ph-pNMe₂, pKₐ = 7.4) are weaker acids than are analogs with electron-withdrawing groups, 3f·H⁺ (R = Ph-pCF₃, pKₐ = 6.7) and 3h·H⁺ (R = Ph-pCl, pKₐ = 6.8). This means that the free-basicity of 3d (R = Ph-pOMe) and 3e (R = Ph-pNMe₂) are more basic than 3f (R = Ph-pCF₃) and 3h (R = Ph-pCl). The increased basicity of 3d and 3e means that a greater percentage of these analogs would be protonated at physiological pH, which should enhance their ability to extract anions from water and transport these bound anions as neutral complexes 3·HA across lipid membranes.

Correlation between anion transport rates and transporter basicity

Having established that B-ring substitution influences the acid-base properties of OAr analogs 3b, 3d–f and 3h, we next compared the ability of these synthetic prodigiosines to catalyze the exchange of anions across phospholipid vesicles. Transmembrane transport was evaluated with a liposome model that uses the chloride-sensitive dye, lucigenin (LG), to monitor anion exchange.30 This assay works because LG fluorescence is quenched by Cl⁻ but not by NO₃⁻. Thus, egg-yolk phosphatidylcholine (EYPC) liposomes loaded with 1.0 mM LG and NaCl were suspended in a solution (pH 7.43) containing extravesicular NaNO₃. Upon the addition of our synthetic prodigiosines 3 the fluorescence of the intravesicular LG increases, indicating Cl⁻ efflux and NO₃⁻ influx. Fig. 7 shows representative fluorescence curves, plotted as a function of time, after addition of analogs 3b, 3d–f and 3h. We quantified transport rates by (a) determining the initial rate of change of the relative fluorescence of the intravesicular LG and (b) estimating the half-life of the exchange reaction (Table 1). The data in Fig. 7 and Table 1 contain important findings. First, these synthetic prodigiosines, especially 3d and 3e, are potent anion transporters and relatively low transporter loadings are needed to observe significant Cl⁻ efflux from the liposomes. For example, we used only 0.0008 mol% of transporter relative to EYPC lipid (1 transporter for 125 000 lipid molecules or about 2.5 transporters per liposome) to generate the data in Fig. 7. This is one of the lowest transporter loadings reported to date.5 Second, substitution of the B-ring with an OAr group does not perturb the ability of these analogs to catalyze anion exchange. Indeed, the electron-rich 3d·H⁺ (R = Ph-pOMe) is qualitatively similar to the natural product prodigiosin 1·H⁺ in terms of the efficiency at which it exchanges Cl⁻ and NO₃⁻ anions across the EYPC bilayer (see ESI Fig. S6†) Most importantly, compounds substituted with electron-donating groups (3d and 3e) are better anion transporters than analogs containing electron-withdrawing groups (3f and 3h). These results indicate that the catalytic efficiency of transmembrane anion exchange correlates with how much of the prodigiosine is in its protonated form 3·HA.31

In vitro anti-cancer properties

The in vitro activity of some of the B-ring analogs was evaluated at the National Cancer Institute (NCI, http://dtp.cancer.gov) against the standard NCI60 panel of 60 human cell lines.32 Prodigiosenes selected for testing were the benzyl (3a) and phenyl (3b) analogs and derivatives with an electron-donating (3d R = Ph-pOMe) and an electron-withdrawing group (3f R = Ph-pCl). Activities against the NCI60 panel are reported in Table 2 as mean values averaged over all 60 cell lines; these activities are (1) the half maximal growth inhibition concentration, GI₅₀, (2) the total growth inhibition concentration, TGI, and (3) the half maximal lethal concentration, LC₅₀. All four synthetic prodigiosenes exhibited GI₅₀ values in the nM range, similar in magnitude to 1, indicating that replacement of the B-ring OMe by an O-aroyl group is not universally detrimental to anti-cancer activity. They also show

Table 1  pKₐ values and transmembrane anion exchange rates for some synthetic prodigiosenes with different –OAr groups on the B-ring. Transport experiments were repeated in triplicate and all traces reported in Fig. 7 are the average of those 3 trials.

<table>
<thead>
<tr>
<th>3·HA</th>
<th>pKₐ</th>
<th>Initial rate for Cl⁻/NO₃⁻ exchange (s⁻¹)</th>
<th>t₁/₂ for Cl⁻ efflux (s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>3d (Ph-pOMe)</td>
<td>7.3 ± 0.1</td>
<td>15.1 ± 0.2</td>
<td>65 ± 2</td>
</tr>
<tr>
<td>3e (Ph-pNMe₂)</td>
<td>7.4 ± 0.1</td>
<td>10.8 ± 0.3</td>
<td>87 ± 3</td>
</tr>
<tr>
<td>3b (R = Ph)</td>
<td>7.2 ± 0.1</td>
<td>10.8 ± 0.3</td>
<td>90 ± 5</td>
</tr>
<tr>
<td>3h (Ph-pCl)</td>
<td>6.8 ± 0.1</td>
<td>3.4 ± 0.1</td>
<td>168 ± 3</td>
</tr>
<tr>
<td>3f (Ph-pCF₃)</td>
<td>6.7 ± 0.1</td>
<td>1.0 ± 0.1</td>
<td>287 ± 2</td>
</tr>
</tbody>
</table>

Fig. 7 Anion exchange assay for analogs 3b, 3d–f and 3h. Change in fluorescence of lucigenin (LG) observed in EYPC liposomes at 25 °C. The data was collected using 0.0008 mol% of transporter compounds relative to EYPC lipid. Liposomes containing 1 mM lucigenin, 20 mM HEPES buffer and 100 mM NaCl buffer were suspended in 20 mM HEPES and 100 mM NaNO₃ solution (pH 7.43); at t = 30 s, prodigiosene transporter was added and the fluorescence monitored (λex = 372 nm, λem = 504 nm); at t = 660 s, addition of Triton-X; traces shown are an average of three trials.
LC50 values in the μM range that could indicate a lower toxicity compared to the natural compound 1 (LC50 = 300 nM). The phenyl analog 3b was the most active of the derivatives with a mean GI50 value of 18 nM. A more detailed analysis in Fig. 8 shows that 3b was also typically the most active of the four synthetic prodigiosenes against all 60 cell lines.

Two questions that we sought to answer were would the prodigiosene basicity (1) correlate with its ability to catalyze transmembrane anion exchange and (2) correlate with its anti-cancer activity. The data presented herein suggest that the ease of protonation controls the transmembrane anion exchange. However, despite their significant in vitro anti-cancer activity, there is no evidence that those anti-cancer properties universally correlate with, or are caused by, the acid-base properties of the prodigiosene. Thus, the most basic prodigiosene 3d and the least basic prodigiosene 3h had similar mean GI50 values, 74 and 62 nM respectively, and both analogs were less potent than the phenyl analog 3b (GI50 = 18 nM).

Table 2  
In vitro activity of some prodigiosenes over the NCI60 cancer cell lines. Values are a mean of two experiments (http://dtp.cancer.gov).

<table>
<thead>
<tr>
<th></th>
<th>1</th>
<th>3a (Bn)</th>
<th>3b (Ph)</th>
<th>3d (Ph-pOCH3)</th>
<th>3h (Ph-pCl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GI50 (nM)</td>
<td>14</td>
<td>44</td>
<td>18</td>
<td>74</td>
<td>62</td>
</tr>
<tr>
<td>TGI (μM)</td>
<td>2.1</td>
<td>0.6</td>
<td>0.2</td>
<td>0.8</td>
<td>1.9</td>
</tr>
<tr>
<td>LC50 (μM)</td>
<td>0.3</td>
<td>4.8</td>
<td>2.5</td>
<td>7.2</td>
<td>14.4</td>
</tr>
</tbody>
</table>

a GI50 = half maximal growth inhibition concentration. b TGI = total growth inhibition concentration. c LC50 = half maximal lethal concentration.

Conclusions

Prodigiosenes, both naturally occurring and synthetic analogs, are outstanding catalysts for transport of anions across lipid membranes. Being able to control the efficiency of this transmembrane transport of anions would be valuable for various reasons, such as the potential development of therapeutics or anion sensors. In this study we synthesized some unique prodigiosene analogs with different OAr substituents attached to the B-ring of the tripyrrolic skeleton. We measured the apparent pKₐ values of five B-ring analogs, namely the O-phenyl derivative (3b·H⁺), two O-aryl analogs with electron-donating groups and two O-aryl analogs with electron-withdrawing groups. These five derivatives were then tested for their ability to catalyze transmembrane exchange of Cl⁻ and NO₃⁻ anions in EYPC liposomes. The data indicate that the efficiency of anion transport can indeed be modulated by the electronic nature of the O-aryl substituent on the B-ring. Thus, more basic analogs 3d (R = Ph-pOMe, pKₐ = 7.3) and 3e (R = Ph-pNMe₂, pKₐ = 7.2) showed significantly greater Cl⁻/NO₃⁻ exchange rates than did the less basic analogs 3h (R = Ph-pCl, pKₐ = 6.8) and 3f (R = Ph-pCF₃, pKₐ = 6.7). This suggests that the key factor controlling anion transport for this family of synthetic analogs, and presumably for the natural product prodigiosin, is the protonation of the prodigiosene to give an amphiphilic cation 3·H⁺. This cation extracts anions from water to give a neutral complex 3·HA that then diffuses rapidly across the lipid membranes. In addition to the synthetic and mechanistic work we also evaluated the anti-cancer activity of four synthetic prodigiosenes (NCI60 human tumour panel).
All four analogs showed potent anti-cancer activities, with mean GI_50 values in the nM range, indicating that substitution of the B-ring with OAr groups can give potent anti-cancer agents. Despite their promising in vitro anti-cancer activity there was no evidence that this activity is due to the ease of protonation of these synthetic prodigiosines.

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Notes and references


31 Transmembrane anion transport by tambjamine alkaloids, relatives of the prodigiosenes, can be influenced by transport-lipophilicity (ref. 6a). Therefore we calculated values for log P, the logarithm of the octanol-water partition coefficient, for the synthetic transporters (see ESI†). Importantly, the analog with the highest Cl\(^{-}/\)NO\(_3\) exchange rates, 3d-H\(^{+}\) (R = OMe, log P = 4.9) is significantly less lipophilic than the slowest transporter in this family 3h-H\(^{+}\) (R = CF\(_3\), log P = 5.8).