



Cite this: *Org. Biomol. Chem.*, 2014, **12**, 6470

A “click” chemistry constructed affinity system for 2-oxoglutaric acid receptors and binding proteins†

Yang Wang,^{a,b} Zeinab Assaf,^{a,c} Xinjun Liu,^b Fabio Ziarelli,^d Amel Latifi,^c Otmane Lamrabet,^c Gilles Quéléver,^a Fanqi Qu,^b Cheng-Cai Zhang^c and Ling Peng^{*a}

An ingenious and specific affinity resin designed to capture the 2-oxoglutaric acid (2-OG) binding proteins was constructed by appending a 2-OG tag to the solid resin *via* a Cu-catalyzed Huisgen “click” reaction. The so-obtained affinity resin was able to recognize, retain and separate the established 2-OG binding protein NtcA in both the pure form and crude cellular extract, thus constituting a valuable means of searching for novel 2-OG receptors with a view to exploring the signalling pathways of 2-OG, a key Krebs cycle intermediate with unprecedented signalling functions.

Received 15th May 2014,
Accepted 30th June 2014

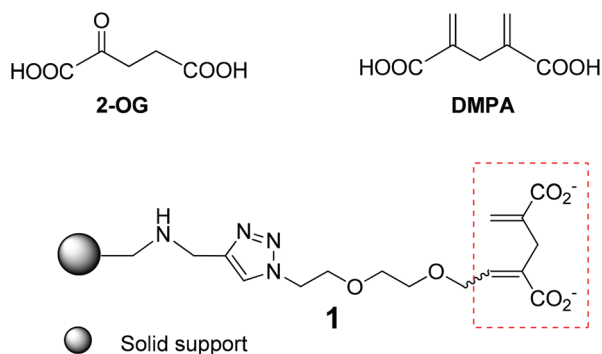
DOI: 10.1039/c4ob01005a

www.rsc.org/obc

Introduction

2-Oxoglutaric acid (2-OG) (Scheme 1) is a strategically important intermediate of the Krebs cycle since it constitutes the precursor and the carbon skeleton for nitrogen assimilation, leading to the synthesis of various biomolecules and the production of cellular energy. In addition to its canonical roles, 2-OG has recently regained considerable attention as a key signalling molecule^{1–5} in different organisms as it plays important roles in various signalling pathways such as regulation of

the balance in carbon and nitrogen metabolism,^{6–8} epigenetic regulation mediated by 2-OG dependent oxygenases,⁹ generation of the “onco-metabolite” 2-hydroxyglutarate *via* misregulated metabolism in various cancers, *etc.*¹⁰ The signalling pathways of 2-OG are rather complex and remain largely unexplored because of the difficulties faced in identifying the corresponding 2-OG receptors or 2-OG binding proteins. The prerequisite therefore in achieving the ultimate goal of better understanding the signalling role of 2-OG is finding a simple and effective way of searching and identifying potential 2-OG receptors. To this end, affinity chromatography appears to be the method of choice by virtue of its high specificity, easy manipulation and simple scale-up.¹¹ In this method, a competent tag bearing 2-OG mimics shall be immobilized onto the solid support *via* an appropriate spacer arm, and the resulting affinity resin (Fig. 1) is expected to capture the 2-OG binding proteins *via* specific interactions between the 2-OG binding site of the proteins and the 2-OG mimic attached on the resin. An elution step using 2-OG containing eluent would then



Scheme 1 Structure of 2-OG, the 2-OG analogue DMPA and the affinity resin **1** with DMPA as a 2-OG tag on the solid support.

^aAix-Marseille Université and CNRS, Centre Interdisciplinaire de Nanoscience de Marseille, UMR 7325, 13288 Marseille, France. E-mail: ling.peng@univ-amu.fr

^bCollege of Chemistry and Molecular Sciences, Wuhan University, P. R. China

^cAix-Marseille Université and CNRS, Laboratoire de Chimie Bactérienne, UMR 7283, Marseille, France

^dAix-Marseille Université and CNRS, Spectropôle, FR 1739 Marseille, France

† Electronic supplementary information (ESI) available: Schemes S1–S3, Fig. S1–S3, synthesis of **2**, **3** and **6**, ¹H and ¹³C NMR spectra of compounds **2**–**5**, IR and ¹³C HRMAS NMR spectra of resins **1**, **6**, **7**. See DOI: 10.1039/c4ob01005a

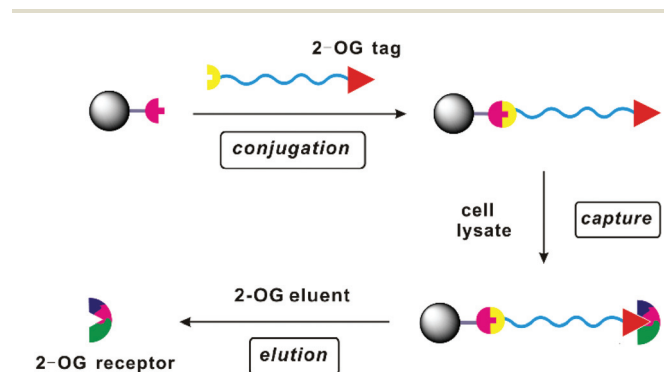


Fig. 1 The principle of using 2-OG affinity chromatography to capture the potential 2-OG receptors.



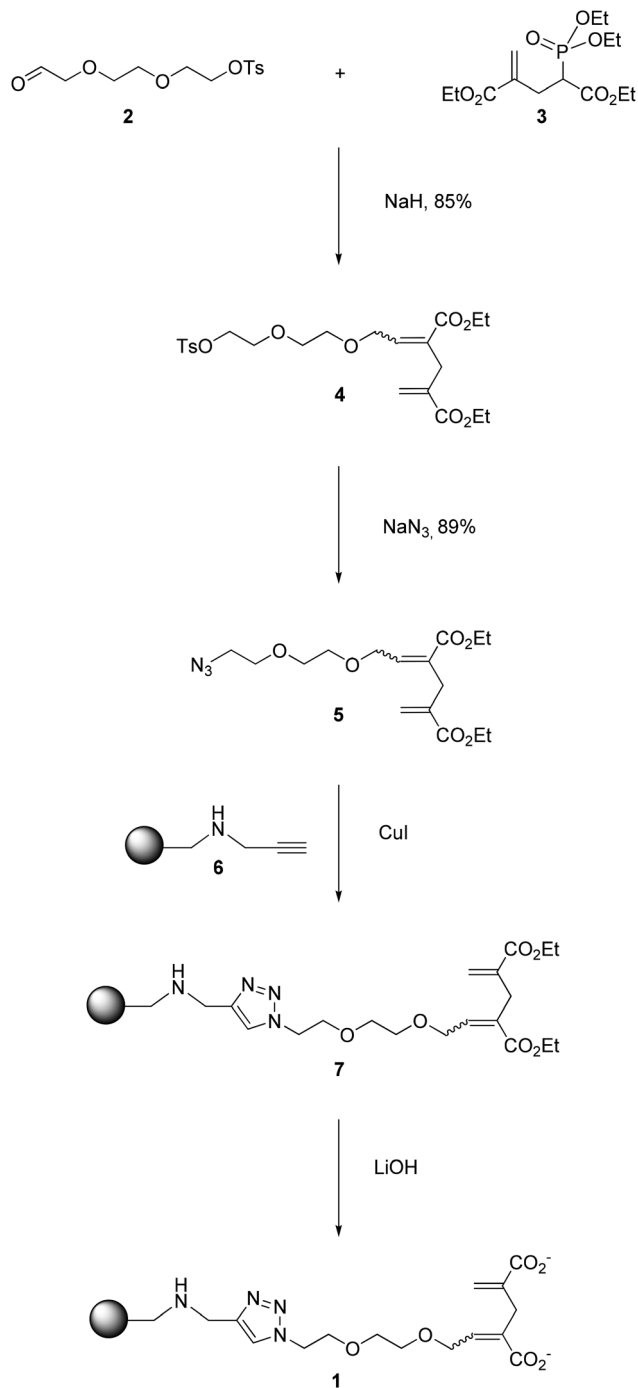
deliver the 2-OG binding proteins enabling their subsequent purification and identification.

The foremost requirement to guarantee a successful affinity chromatographic study is establishing a competent affinity resin with high affinity and specificity, where the apt tag to sense the 2-OG binding proteins is of paramount importance. However, the native 2-OG does not appear to be the tag of choice as the α -keto carboxylic acid entity within 2-OG is rather labile, and ready to undergo different transformations in biological media containing various enzymes. This is certainly the biggest concern during our consideration to devise an appropriate affinity column to capture the 2-OG binding proteins in the crude cell extracts or biological media where many enzymes or proteins may react with the α -keto carboxylic acid entity. Furthermore, 2-OG is a small molecule, and any structural modification with relatively large functional moieties will inevitably modify binding characteristics towards 2-OG receptors.^{4,12–15} Taking all these into consideration, we opted for the nonmetabolisable 2-OG analogue, 2,4-dimethylenepentanedioic acid (DMPA), as the 2-OG tag to construct the affinity resin **1** (Scheme 1) for its structural resemblance to, and resulting ability to mimic, 2-OG in signaling.¹⁴ In addition, it is practical for chemical modification and conjugation of DMPA onto the solid support. For linking the 2-OG tag onto the solid resin, we chose the hydrophilic ethyleneoxy spacer in order to provide a hydrophilic environment allowing the 2-OG tag to search for and bind to the 2-OG binding proteins. To anchor the 2-OG tag onto the resin, we wished to harness the Cu-catalyzed Huisgen reaction,¹⁶ a well acknowledged “click” reaction for affording clean products efficiently and rapidly in high yield under mild conditions, hence allowing a facile and economic solid support synthesis.

Based on this rationale, we report here the synthesis, characterization and biological validation of the so-devised affinity resin **1** that can selectively and specifically recognize, retain and separate the 2-OG binding protein NtcA, a known 2-OG receptor involved in regulating the carbon/nitrogen balance and heterocyst differentiation in diazotrophic cyanobacteria.^{4,12–15,17} To our knowledge, this is the first example of an affinity resin using a 2-OG mimic tag validated with a known 2-OG receptor.¹⁸ This affinity system hence holds great promise in searching for 2-OG receptors or 2-OG binding proteins involved in signal transduction and metabolic regulation.

Results and discussion

The construction of the affinity resin **1** was achieved according to the strategy presented in Scheme 2. The precursor of the 2-OG tag featuring the ethyleneoxy spacer (**5** in Scheme 2) was obtained *via* an Horner–Wadsworth–Emmons (HWE) reaction²⁰ by condensing the corresponding aldehyde **2** with the phosphonate **3** in the presence of sodium hydride, affording **4** with an excellent yield of 85% as a mixture of *E/Z* isomers (*E/Z* = 3/1).²¹ Subsequent treatment with NaN₃ delivered the corres-



Scheme 2 Preparation of the affinity resin **1**.

ponding azides **5**, which were further conjugated to the alkyne-bearing resin **6** *via* a Cu-catalyzed Huisgen cycloaddition to yield the resin **7**. Gratifyingly, the Cu-catalyzed “click” reaction proved to be very powerful and rewarding as it proceeded smoothly, and was accomplished rapidly within an hour (Fig. S1†). As shown in the IR spectra (Fig. 2A), the peak at 3289 cm⁻¹ corresponding to the vibration of the C–H bond in the terminal alkyne –C≡C–H moiety of **6** disappeared following the “click” reaction, and a very strong signal was generated



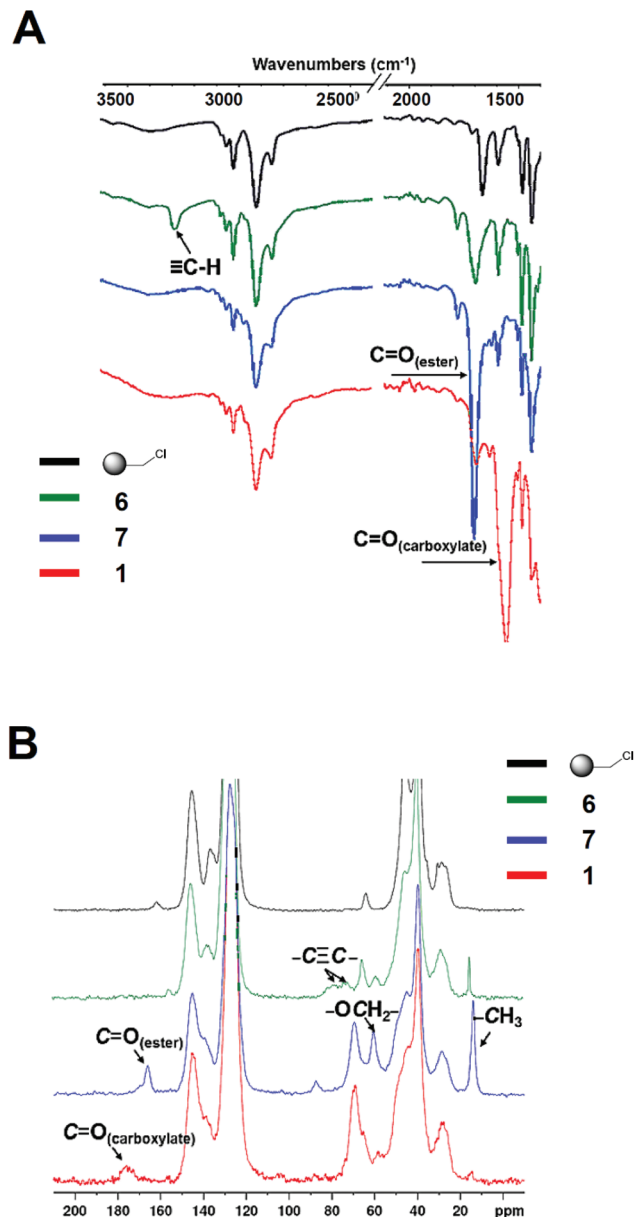


Fig. 2 IR and ¹³C CPMAS NMR spectra of the intermediate products 6 and 7, and the final affinity resin 1.

around 1707 cm⁻¹, relating to the carbonyl group of the ester functions of 7. The successful conversion of 6 to 7 was further affirmed by ¹³C cross polarization magic angle spinning (CPMAS) NMR, an excellent non-destructive method for the *in situ* analysis of the solid phase reaction.^{22–24} As illustrated in Fig. 2B, the disappearance of the signals associated with the alkynyl moiety at 70–80 ppm in 6 was accompanied by the emergence of the signals at 14 and 60 ppm corresponding to the ethyl groups within the ester functions of 7. Collectively, these results indicate a rapid and complete conversion of 6 to deliver 7 via “click” chemistry. Subsequent alkaline hydrolysis of 7 furnished the final resin 1, which was successfully characterized by both IR and CPMAS NMR. As we can see in Fig. 2A, the hydrolysis of the ester groups in 7 to give the corres-

ponding carboxylate functions was highlighted by the signal of the carbonyl group shifting from 1707 cm⁻¹ (characteristic of the ester groups in 7) to 1658 cm⁻¹ (characteristic of the carboxylate functions in 1). This was further confirmed with the results obtained by ¹³C CPMAS NMR. As clearly depicted in Fig. 2B, the signal concerning the ester carbonyl groups in 7 wore off at 166 ppm, and the new broad peak corresponding to the carboxylate carbonyl groups in 1 emerged at 175 ppm. Moreover, the signals relating to the ethyl moieties in the ester groups of 7 disappeared at 14 and 60 ppm, further stressing the successful hydrolysis of 7 to afford the resin 1.

To verify that the above-obtained affinity resin 1 could indeed recognize and bind 2-OG binding proteins, we examined its interaction with NtcA, an established 2-OG receptor regulating nitrogen metabolism and responsible for heterocyst formation in cyanobacteria.^{4,17} Bovine serum albumin (BSA) was used as a negative control of non-2-OG binding protein. No retention of BSA on resin 1 was observed (data not shown), whereas 1 could effectively retain NtcA on the column, as indicated by the revelation of NtcA in the solution before and after passing through the affinity column composed of 1 (Fig. 3A, lanes 2, 3, 4 and 5). The retained NtcA on the affinity column was then effectively eluted out using a solution containing 2-OG (Fig. 3A, lane 6), authenticating that the retention of NtcA on the resin 1 is specific *via* the explicit interaction between NtcA and the 2-OG tag featured on 1.

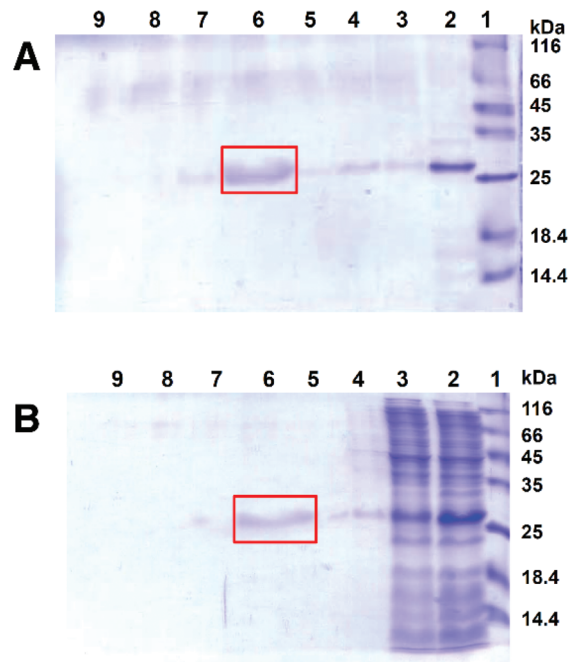


Fig. 3 Protein revelation of resin 1 capturing and eluting NtcA from (A) a pure NtcA solution and (B) *E. coli* cell lysate. Lane 1: protein molecular weight marker; lane 2: NtcA or *E. coli* cell lysate as a reference; lane 3: the supernatant from the incubation buffer; lanes 4 and 5: elution with PBS; lane 6: elution with 2-OG solution (0.5 M) in PBS buffer; lane 7: elution with PBS; lane 8: elution with NaCl solution; lane 9: elution with PBS. The pH of PBS and 2-OG elution was controlled at pH = 7.5.



We further tested the robustness of the affinity resin **1** in identifying and capturing the 2-OG receptor NtcA from cell lysate. To this end, we assessed the ability of resin **1** to fish out the NtcA protein from the total cellular extract of an *E. coli* strain which over-expresses NtcA. We found that **1** could indeed selectively and specifically anchor NtcA, and then release it when using 2-OG elution (Fig. 3B). Altogether, these results demonstrate the robust specificity of the affinity resin **1** and its potential use in detecting and searching for 2-OG receptors.

Conclusions

In summary, we have successfully established the affinity resin **1** for specific recognition, capture and release of the 2-OG receptor NtcA *via* affinity chromatography. This resin features an affinity tag bearing a 2-OG mimic, which was appended to the solid support resin *via* a hydrophilic linker. Cu-catalyzed “click” chemistry was implemented to conjugate the tag to the resin, and proved to be very efficient and rewarding. The so-obtained 2-OG resin could not only retain the 2-OG receptor NtcA in its pure form, but also identify and capture NtcA from the cell extract containing a mixture of various macromolecules, before allowing its release from the column with an eluent containing 2-OG. Altogether, these results illustrate the power and value of this resin in specifically recognizing, sufficiently binding and effectively releasing the 2-OG receptor NtcA. Consequently, this resin constitutes a novel useful means of searching for new 2-OG receptors with a view to investigate the signaling pathways of 2-OG and complete our understanding of its signaling roles. We are actively pursuing in this direction.

Experimental section

General methods

All the reactions were carried out under argon. Monotosylated triethylene glycol was synthesized according to the literature.²⁵ The detailed synthesis of compounds **2** and **3** and resin **6** is described in the ESI.† Anhydrous 1,2-dimethoxyethane (DME) was distilled in the presence of sodium-benzophenone. Anhydrous CH₂Cl₂ and DMSO were prepared by distillation in the presence of CaH₂. 85% NaH was prepared by washing 60% NaH in mineral oil with petroleum ether and drying in a vacuum. The Merrifield resin (loading capacity: 3.1 mmol g⁻¹) was purchased from Iris Biotech (Germany). All the other reagents were purchased from Sigma-Aldrich (China) or Acros Organics (China) without any further purification. Silica gel (200–300 mesh) used for flash chromatography was purchased from Qing Dao Hai Yang Chemical Industry Co. (China). ¹H NMR and ¹³C NMR spectra were recorded at 300 MHz and 75 or 150 MHz respectively on Varian Mercury-VX300 and VX600 spectrometers. Chemical shifts are reported in parts per million (ppm) with TMS as an internal reference. The

³¹P NMR spectrum was recorded at 121 MHz on a Varian Mercury-VX300 spectrometer. Chemical shifts were reported relatively to 85% H₃PO₄ as the external standard. The solid-state ¹³C cross polarization magic angle spinning (CPMAS) NMR spectra were recorded on a Bruker Avance-400 MHz NMR spectrometer. HRMS were recorded using Waters Micromass GCT Premier and QStar Elite Mass spectrometers. IR spectra were recorded using a Bruker Alfa IR spectrometer. The microwave reaction was performed in a CEM Discover SP microwave reactor.

Synthesis of 4. To a mixture of 85% NaH (229 mg, 8.11 mmol) in freshly distilled DME (10.0 mL) was added a solution of **3** (2.73 g, 8.11 mmol) in freshly distilled DME (10.0 mL) over a period of 10 min. The reaction mixture was stirred at room temperature for 3 h until the disappearance of powdered NaH. Then, a solution of **2** (1.63 g, 5.40 mmol) in freshly distilled DME (10.0 mL) was added to the reaction mixture over a period of 10 min at –50 °C. The resulting reaction mixture was allowed to warm up to room temperature and stirred for 2 h before quenching by addition of a saturated NH₄Cl (20.0 mL), and then followed by extraction with ethyl acetate (3 × 20.0 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered and concentrated under reduced pressure. The crude residue was purified by flash chromatography using a gradient of petroleum ether–ethyl acetate (6 : 1–4 : 1, v/v), yielding **4** as a colorless oil (2.22 g, 85%, *E/Z* ≈ 3/1). ¹H NMR (300 MHz, CDCl₃) δ 7.80 (d, *J* = 8.10 Hz, 2H, *Ph*CH₃), 7.35 (d, *J* = 8.40 Hz, 2H, –*Ph*CH₃), 6.98 (t, *J* = 5.70 Hz, 1H, *E*: C=CHCH₂), 6.23 (s, 1H, *Z*: C=CHH), 6.19–6.14 (m, 2H, *E*: C=CHH, *Z*: C=CHCH₂), 5.55 (s, 1H, *Z*: C=CHH), 5.49 (s, 1H, *E*: C=CHH), 4.46 (d, *J* = 4.80 Hz, 1H, *Z*: C=CHCH₂), 4.23–4.14 (m, 8H, C=CHCH₂, CH₂OTs, OCH₂CH₃), 3.70 (t, *J* = 4.80 Hz, CH₂CH₂OTs), 3.61–3.56 (m, 4H, OCH₂), 3.31 (s, 2H, *E*: CH₂), 3.29 (s, 2H, *Z*: CH₂), 2.45 (s, 3H, –PhCH₃), 1.32–1.25 (m, 6H, –CH₂CH₃); ¹³C NMR (150 MHz, CDCl₃) δ 166.9, 166.8, 145.2, 144.0, 141.0, 138.6, 137.5, 133.2, 130.6, 130.2, 129.3, 128.2, 126.7, 125.6, 70.9–68.2, 61.2, 60.9, 35.3, 29.3, 21.9, 14.5; HRMS cauld for C₂₃H₃₂O₉Sn⁺ 507.1659, found 507.1659.

Synthesis of 5. To a solution of **4** (2.18 g, 4.50 mmol) in CH₃CN (50.0 mL) was added NaN₃ (1.47 g, 22.5 mmol). The reaction mixture was refluxed for 10 h, and then was allowed to cool down to room temperature and filtered. The so obtained organic phase was concentrated under reduced pressure, and the crude residue was purified by flash chromatography using a gradient of petroleum ether–ethyl acetate (6 : 1–4 : 1, v/v), yielding pale yellow oil **5** as a mixture of *E/Z* isomers (863 mg, 54%, *E/Z* ≈ 3/1). ¹H NMR (300 MHz, CDCl₃) δ 7.00 (t, *J* = 5.90 Hz, 1H, *E*: C=CHCH₂), 6.23 (s, 1H, *Z*: C=CHH), 6.19 (s, 2H, *E*: C=CHH, *Z*: C=CHCH₂), 5.54 (s, 1H, *Z*: C=CHH), 5.48 (s, 1H, *E*: C=CHH), 4.49 (d, *J* = 4.80 Hz, 1H, *Z*: C=CHCH₂), 4.25–4.14 (m, 6H, C=CHCH₂, OCH₂CH₃), 3.70–3.63 (m, 6H, –OCH₂), 3.41 (t, *J* = 4.95 Hz, CH₂N₃), 3.32 (s, 2H, *E*: CH₂), 3.29 (s, 2H, *Z*: CH₂), 1.32–1.24 (m, 6H, –CH₂CH₃); ¹³C NMR (150 MHz, CDCl₃) δ 166.6, 166.5, 143.8, 140.8, 138.3, 137.2, 130.3, 129.0, 126.4, 125.2, 70.6–69.8, 68.0, 60.8,



60.6, 50.6, 35.0, 29.0, 14.2; IR (cm⁻¹) ν 2106.1 (-N₃); HRMS cacl for C₁₆H₂₆N₃O₆⁺ 356.1816, found 356.1815.

Synthesis of 7. To a mixture of **6** (150 mg, 0.465 mmol) in 3.00 mL THF and *N,N*-diisopropylethylamine (2 : 1, v/v) were added **5** (196 mg, 0.552 mmol) and CuI (5.00 mg, 0.0263 mmol). The reaction mixture was stirred at 35 °C for 24 h, and then allowed to cool down to room temperature and filtered. The obtained resin was washed successively with THF, water, saturated EDTA solution, water, MeOH, CH₂Cl₂ and diethyl ether (30.0 mL for each) and dried in a vacuum, yielding 160 mg desired resin **7** as a pale green powder. IR (cm⁻¹) ν 1707 (C=O); ¹³C CPMAS NMR δ 166.0 (C=O), 60.6 (OCH₂CH₃), 14.0 (OCH₂CH₃).

Synthesis of 1. To a mixture of **7** (50.0 mg, 0.155 mmol) in 1.50 mL THF was added 1 M LiOH (1.50 mL). The reaction mixture was stirred at 35 °C for 24 h, and then allowed to cool down to room temperature and filtered to obtain the resin. The obtained resin was washed successively with THF, water, saturated NaCl solution, MeOH, CH₂Cl₂ and diethyl ether (30.0 mL for each) and dried in a vacuum, yielding 40 mg resin **1** as a yellow powder. IR (cm⁻¹): ν 1707 (C=O); ¹³C CPMAS NMR δ 175.9 (C=O).

¹³C CPMAS NMR. The solid-state ¹³C CPMAS NMR spectra were obtained on a Bruker Avance-400 MHz NMR spectrometer operating at a ¹³C resonance frequency of 106 MHz using a commercial Bruker double-bearing probe. About 30 mg of each sample were placed in zirconium dioxide rotors of 4 mm outer diameter and spun at a magic angle spinning rate of 10 kHz. The cross polarization (CP) technique²² was applied with a ramped ¹H-pulse starting at 100% power and decreasing until 50% during the contact time (2 ms) in order to circumvent Hartmann–Hahn mismatches.²³ To improve the resolution, a dipolar decoupling TPPM15 pulse sequence was applied during the acquisition time. To obtain a good signal-to-noise ratio in the ¹³C CPMAS experiment 6000 scans were accumulated using a delay of 2.5 s. The ¹³C chemical shifts were referenced to tetramethylsilane and calibrated with the glycine carbonyl signal, set at 176.5 ppm.

Production and purification of the NtcA recombinant protein. A DNA fragment corresponding to the entire coding region of *ntcA* (alr4392) was amplified by performing PCR using the *ntcA* forward primer 5'-CATATG-ATGATCGTGA CACAAGATAA3' (NdeI site underlined) and the *ntcA* reverse primer 5'-GAGCTC-AGTGAAGTGT CTGCTGAGAG T3' (SacI site underlined). The PCR product was cloned into the pET28 vector (Novagen, USA). A clone confirmed by DNA sequencing was transformed into the BLDE3 *E. coli* strain (Novagen, USA). The recombinant clones obtained were grown in LB rich medium supplemented with Kanamycin (50 μ g mL⁻¹) and glucose (0.2%) with an OD at 600 nm of 0.3–0.4. The culture was then washed twice with LB medium to eliminate the glucose. The expression of the recombinant protein was then induced by adding 1 mM isopropyl- β -D-thiogalactoside (IPTG) for 4 h. The recombinant proteins were purified using Hitrap columns as recommended by Pharmacia. Imidazole was removed from purified proteins using PD10 columns (Pharma-

cia, Sweden). Proteins were concentrated on Vivaspin columns and used for subsequent analyses. Proteins were fractionated by SDS-PAGE (12.5% gel) and stained using the SeeBand procedure (Euromedex, France).

Preparation of the *E. coli* cell lysate. A BL21DE3 culture expressing the *ntcA-6his* recombinant gene was prepared as explained above. After induction of the synthesis of the recombinant protein, the obtained pellet was resuspended in 2 mL of PBS. 20 μ L of the protease inhibitor (Roche, France) was added. After disrupting the cell by ultrasonication (3 \times 15 min), the lysate was centrifuged for 10 min at 12 300 rpm. The supernatant was loaded onto the resin **1** for affinity chromatography.

Affinity chromatography. Affinity resin **1** (20 mg) was pre-equilibrated by washing with PBS (4 \times 1 mL) and 0.5 M EDTA solution (6 \times 1 mL), and then again with PBS (4 \times 1 mL). After pre-equilibration, NtcA (10 μ g/45 μ L) or *E. coli* cell lysate (200 μ L) was added to resin **1**, followed by incubation and agitation (gentle agitation every 5 min) at room temperature for 3 h. Resin **1** was then washed twice with PBS (45 μ L). The elution was realized with 2-OG (45 μ L, 0.5 M). Repeated incubation and agitation (gentle agitation every 5 min) were performed at room temperature for 3 h; resin **1** was washed again with PBS (45 μ L), regenerated with NaCl (45 μ L, 1 M), and washed for the last time with PBS (1 mL). The regenerated resin was stored at -20 °C. The presence of the protein at each step of the chromatography was assessed by SDS-PAGE as explained above.

Acknowledgements

We are grateful for the financial support from the “ProKrebs” project of the ANR program of “Physique, Chimie et du Vivant”, Région PACA, CNRS, Wuhan University and Aix-Marseille Université. Y.W. and X.L. are supported by the overseas Ph.D. fellowship from China Scholarship Council. We thank Dr Emily Witty for English correction.

Notes and references

- 1 S. Zhang and D. A. Bryant, *Science*, 2011, **334**, 1551–1553.
- 2 W. He, F. J.-P. Miao, D. C.-H. Lin, R. T. Schwandner, Z. Wang, J. Gao, J.-L. Chen, H. Tian and L. Ling, *Nature*, 2004, **429**, 188–193.
- 3 S. C. Hebert, *Nature*, 2004, **429**, 143–145.
- 4 S. Laurent, H. Chen, S. Bédu, F. Ziarelli, L. Peng and C.-C. Zhang, *Proc. Natl. Acad. Sci. U. S. A.*, 2005, **102**, 9907–9912.
- 5 A. B. Feria Bourrellier, B. Valot, A. Guillot, F. Ambard-Bretteville, J. Vidal and M. Hodges, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 502–507.
- 6 S. Gálvez, M. Lancien and M. Hodges, *Trends Plant Sci.*, 1999, **4**, 484–490.
- 7 M. Hodges, *J. Exp. Bot.*, 2002, **53**, 905–916.
- 8 A. Ninfa and P. Jiang, *Curr. Opin. Microbiol.*, 2005, **8**, 168–173.



- 9 C. Loenarz and C. J. Schofield, *Nat. Chem. Biol.*, 2008, **4**, 152–156.
- 10 R. A. Cairns and T. W. Mak, *Cancer Discovery*, 2013, **3**, 730–741.
- 11 S. Ziegler, V. Pries, C. Hedberg and H. Waldmann, *Angew. Chem., Int. Ed.*, 2013, **52**, 2744–2792.
- 12 H. Chen, S. Laurent, S. Bédu, F. Ziarelli, H.-L. Chen, Y. Cheng, C.-C. Zhang and L. Peng, *Chem. Biol.*, 2006, **13**, 849–856.
- 13 X. Liu, H. Chen, E. Laurini, Y. Wang, V. Dal Col, P. Posocco, F. Ziarelli, M. Fermeglia, C.-C. Zhang, S. Pricl and L. Peng, *Org. Lett.*, 2011, **13**, 2924–2927.
- 14 X. Liu, Y. Wang, E. Laurini, P. Posocco, H. Chen, F. Ziarelli, A. Janicki, F. Qu, M. Fermeglia, S. Pricl, C.-C. Zhang and L. Peng, *Org. Lett.*, 2013, **15**, 4662–4665.
- 15 Y. Wang, X. Liu, E. Laurini, P. Posocco, F. Ziarelli, M. Fermeglia, F. Qu, S. Pricl, C.-C. Zhang and L. Peng, *Org. Biomol. Chem.*, 2014, **12**, 4723–4728.
- 16 M. Meldal and C. W. Tornøe, *Chem. Rev.*, 2008, **108**, 2952–3015.
- 17 M.-X. Zhao, Y.-L. Jiang, Y.-X. He, Y.-F. Chen, Y.-B. Teng, Y. Chen, C.-C. Zhang and C.-Z. Zhou, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 12487–12492.
- 18 Although Schofield *et al.* have established photoreactive affinity systems to probe 2-OG oxygenases,¹⁹ the effective tags that they employed to capture these proteins were based on 8-hydroxyquinoline, a known iron chelator presented in inhibitors of 2-OG oxygenases. Therefore, it is unlikely that the so-obtained affinity systems are generally applicable for all 2-OG binding proteins.
- 19 D. Rotili, M. Altun, A. Kawamura, A. Wolf, R. Fischer, I. K. H. Leung, M. M. Mackeen, Y.-M. Tian, P. J. Ratcliffe, A. Mai, B. M. Kessler and C. J. Schofield, *Chem. Biol.*, 2011, **18**, 642–654.
- 20 W. S. Wadsworth and W. D. Emmons, *J. Am. Chem. Soc.*, 1961, **83**, 1733–1738.
- 21 The HWE reaction often favors the formation of *E* isomers. Since the *E*- and *Z*-isomers of **4** have similar polarity, it is very difficult for us to separate them by column chromatography. We therefore used the mixture of the *E/Z* isomers directly in the subsequent reaction.
- 22 J. Schaefer and E. O. Stejskal, *J. Am. Chem. Soc.*, 1976, **98**, 1031–1032.
- 23 O. B. Peersen, X. Wu, I. Kustanovich and S. O. Smith, *J. Magn. Reson.*, 1993, **104**, 334–339.
- 24 R. L. Cook, C. H. Langford, R. Yamdagni and C. M. Preston, *Anal. Chem.*, 1996, **68**, 3979–3986.
- 25 H. Sashiwa, Y. Shigemasa and R. Roy, *Carbohydr. Polym.*, 2002, **49**, 195–205.

