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Biodegradability of 27 pyrrolidinium, morpholinium, piperidinium, imidazolium and pyridinium ionic liquid cations under aerobic conditions†

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The chemical and thermal stability of ionic liquids (ILs) makes them interesting for a large variety of applications in nearly all areas of the chemical industry. However, this stability is often reflected in their recalcitrance towards biodegradation, which comes with the risk of persistence when they are released into the environment. In this study we carried out a systematic investigation of the biodegradability of pyrrolidinium, morpholinium, piperidinium, imidazolium and pyridinium-based IL cations substituted with different alkyl or functionalised side chains and using halide counterions. We examined their primary degradability by specific analysis and/or their ultimate biodegradability using biochemical oxygen demand tests according to OECD guideline 301F. Biological transformation products were investigated using mass spectrometry. A comparison of the biodegradation potential of these ILs shows that for all five head groups, representatives can be found that are readily or inherently biodegradable, thus permitting the structural design of ILs with a reduced environmental hazard.

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Introduction

Ionic liquids

Ionic liquids (ILs) are chemicals with an outstanding design potential. Their components – cationic and anionic molecular core structures with different side chains – can be combined in innumerable ways to create substances with tailor-made physico-chemical properties important for industrial applications. Most of the compounds possess a very low vapour pressure and non-flammability, which are key properties for improved operational safety in comparison with conventional volatile organic solvents.

Depending on the combination of cations and anions, they can be thermally stable, can be liquid over a wide temperature range and can have a large electrochemical window, which makes them useful for a variety of technical applications in

different fields. Initially used as solvents for synthesis and catalysis in clean technologies, they have been increasingly investigated during the last 15 years.^{1–3} Several industrial processes have been established in which the use of ILs is highly advantageous with respect to the productivity of the processes.^{4,5} Their range of potential applications extends from the already-mentioned usage as solvents and catalysts to media for electrochemical applications and biosensors.^{6–9} ILs have also been considered for utilisation in analytical^{10–13} or supercritical fluid applications,¹⁴ gel production¹⁵ or as pharmaceutical ingredients.^{16,17} Their application as agents for recovery, extraction and purification purposes^{18–20} or as lubricants²¹ and hypergolic rocket fluid²² is also being discussed.

Sustainability considerations

The variety of ILs available for an individual process together with their high operational safety is not only advantageous for the performance and handling of the process, but also for sustainable development.²³ In this context the environmental compatibility of a chemical also plays a major role. The production of substances that simultaneously possess low toxicity and environmental non-persistence is a demanding task for chemists dedicated to sustainable chemistry.²⁴ Nevertheless, its fulfilment appears feasible in the context of the enormous design potential of ILs.^{25–27} The structural aspects that

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influence the toxicity of ILs have already been identified and have been replaced by less toxic arrangements in one application.²⁸

Biodegradable structural elements

Many different studies of ILs for the identification of structural elements which support their overall biological degradation have recently been reviewed.^{3,27,29-31}

The biodegradation potential of ILs ranges widely from very good to very poor. Good primary biodegradability has been observed in ILs with longer alkyl side chains ($>C_6$) at the cationic core, whereas the same head group with shorter side chains is only poorly biodegradable: compare, for example, 1-octyl-3-methyl-1*H*-imidazolium (C_8 mim) with 1-ethyl-3-methyl-1*H*-imidazolium (C_2 mim) combined with halide anions (Cl, Br).^{32,33} Imidazolium-based ILs in general exhibit reduced biodegradability with respect to their core structure,^{32,34} whereas 1-alkyl-3-methylpyridinium-based ILs have an elevated biodegradation potential,^{32,35} and many compounds have been classified as readily biodegradable. Apart from the (bio)degradability of the cation, the behaviour of the anion also has to be taken into account. Anions such as PF_6^- and BF_4^- are sensitive to hydrolytic processes,³⁶ and organic anions such as acetate, and ethyl and octyl sulphate, are readily biodegradable.^{37,38} In contrast, anions such as $N(CN)_2^-$, $B(CN)_4^-$ and $(CF_3SO_2)_2N^-$ are known to be neither hydrolytically degradable nor biodegradable under environmental conditions.^{39,40} The choice of inoculum using pure cultures, such as *Corynebacterium* sp. and *Sphingomonas paucimobilis*, and the presence or absence of degraders in mixed cultures additionally influence the results towards an enhanced biodegradability of the ionic liquid cation.⁴¹⁻⁴³

Thus, a broad biodegradability range of ILs can be created depending (i) on the structural composition with respect to the cationic head group, side chain and counterion, and (ii) on the experimental procedure used for the determination of biodegradability data, more specifically the detection either by a specific analytical procedure or sum parameters, the chosen environmental condition and the choice of microorganisms.

The focus of the study is on the influence of the structural composition and the design potential of ILs.

Missing data

The identification of the biodegradable substructures of ILs has been a mammoth task. Although many commercially available pyridinium and imidazolium-based ILs have been tested, some relevant data for a systematic investigation of other head groups are still missing. To date only *N*-alkyl-*N*-methylmorpholinium and 4-benzyl-4-methylmorpholinium have been examined, head groups that are not readily biodegradable.⁴⁴ A few phosphonium ILs with alkyl and functionalised side chains were also refractory to biological degradation under ready biodegradability test conditions.^{45,46}

As yet, no systematic investigation of the biodegradability of other head groups has been undertaken. We therefore examined a set of 27 ILs with differently substituted head groups –

pyrrolidinium, morpholinium and piperidinium ILs – together with some pyridinium and imidazolium ILs (Tables 1-6). We screened the primary biodegradability of IL cations, tracking the course of degradation using liquid chromatography, determined (where possible) transformation products with mass spectrometry, and investigated the extent of mineralisation of selected ILs by applying the sum parameter biochemical oxygen demand (BOD). Several of the IL cations under investigation were fully degraded and some of them match the criteria for being classified as readily biodegradable. In experiments with a prolonged standard test duration (>28 d) some more compounds underwent biodegradation, showing the importance of microbial adaptation. Even during the degradation of one IL, microbial adaptation occurred as diauxic growth when head groups and side chains were successively biodegraded.

Our systematic studies of the biodegradability of ILs are addressed to the users of ILs in different fields of application to facilitate the selection of environmentally favourable structural elements and hence to contribute to the design of inherently safer ILs.

Materials and methods

Chemicals

All the tested ILs as well as the salts for the mineral salt medium and the standard eluent for ion chromatographic analysis were obtained from Merck KGaA (Darmstadt, Germany). Acetonitrile (HPLC grade) for the ion chromatographic measurements was obtained from VWR International GmbH (Darmstadt, Germany).

Biodegradation tests

1 Manometric respirometry. The manometric respirometry test was performed according to OECD guideline 301F.⁴⁷ The biological oxygen demand (BOD) of the substance was determined for 28 d (but prolonged as soon as a significant increase in oxygen consumption had been detected) using a BOD measurement system (OxiTop®, thermostatically controlled from WTW GmbH, Weilheim, Germany) with two replicates for each test substance. The extent of mineralisation could be inferred from this test: if 60% biodegradation was exceeded within a certain time frame, the compound was classified as “readily biodegradable”.

The activated sludge was taken from the municipal wastewater treatment plant in Delmenhorst (Germany), filtered through grade 1288 filter discs (Sartorius AG, Göttingen) and aerated for ten days to remove organic substances from sewage treatment before inoculation. The medium was prepared with 8.5 mg L^{-1} KH_2PO_4 , 28.5 mg L^{-1} $K_2HPO_4 \cdot 3H_2O$, 33.4 mg L^{-1} $Na_2HPO_4 \cdot 2H_2O$, 0.5 mg L^{-1} NH_4Cl , 36.4 mg L^{-1} $CaCl_2 \cdot 2H_2O$, 22.5 mg L^{-1} $MgSO_4 \cdot 7H_2O$ and 0.25 mg L^{-1} $FeCl_3$ (pH 7.4). An additional 1.16 mg L^{-1} allylthiourea has been added in order to inhibit nitrification. The bacteria suspension was set up using 20% supernatant from the aerated activated sludge and



Table 1 Structural formula and results of pyrrolidinium-based ionic liquid cations obtained from primary and BOD experiments

Cationic head group	Side chain R	Anion	Substance identifier	Classification OECD and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
		Cl ⁻	C ₈ mpyr Cl	Readily biodegradable	100	69 ± 1	This study
		Cl ⁻	C _{3OH} mpyr Cl	Readily biodegradable	100	67 ± 3	This study
		Br ⁻	C _{1COO2} mpyr Br	Inherently, ultimately biodegradable	100	28 d: 34 ± 11 (n = 4) 38 d: 74 ± 0.1 (n = 4)	This study
		Br ⁻	C ₄ mpyr Br	Inherently, ultimately biodegradable	—	28 d: 14 ± 13 42 d: 70 ± 18	This study
		Cl ⁻	C _{1CN} mpyr Cl	Biotic hydrolysis of the cyano group	Qualitative analysis by mass spectrometry	2 ± 2	This study
		I ⁻	C _{2OH} mpyr I	Not readily, but hints of being inherently	28 d: 8 ± 5 43 d: 25 ± 21	28 d: 6 ± 9 (n = 4) 60 d: 42 ± 39 (n = 4)	This study
				Not readily biodegradable	28 d: -2 ± 3 40 d: -3 ± 8	—	This study
			CH ₃ CH ₂ OSO ₃ ⁻	C ₂ mpyr C _{2O} SO ₃	—	—	—

Table 2 Structural formula and results of morpholinium-based ionic liquid cations obtained from primary and BOD experiments

Cationic head group	Side chain R	Anion	Substance identifier	Classification OECD (...biodegradable) and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
		Cl ⁻	C _{3OH} mmor Cl	Inherently	28 d: 12 ± 8	28 d: 31 ± 16 41 d: 59 ± 10	This study
		Cl ⁻	C _{1CN} mmor Cl	Biotic hydrolysis of the cyano group	54 ± 13 & Qualitative analysis by mass spectrometry	0	This study
		I ⁻	C _{2OH} mmor I	Not readily	28 d: 17 ± 1	28 d: -1 ± 1 (n = 4) 60 d: 4 ± 1 (n = 4)	This study
		Cl ⁻	C _{1O2} mmor Cl	Not readily	-3 ± 4	—	This study
		Cl ⁻	C _{2O1} mmor Cl	Not readily	1 ± 4	—	This study
		Br ⁻	C _{2O2} mmor Br	Not readily	0 ± 8	—	This study
		Br ⁻	C ₄ mmor Br	Not readily	-3 ± 4	—	This study

80% mineral salt medium solution. The concentrations of the test substances were chosen according to their expected oxygen demand, which were in a non-inhibitory concentration, usually below 850 $\mu\text{mol L}^{-1}$.⁴⁸ Moreover, blank samples (inoculated media without test substance) and controls (inoculated

media with benzoic acid) were also prepared. In this test a bacteria number of 10^4 cells L^{-1} was applied (determined using a Paddle Tester; Hach Europe, Düsseldorf). Sodium hydroxide was used to absorb the evolved carbon dioxide inside the sample vessels. The vessels were closed with



Table 3 Structural formula and results of piperidinium-based ionic liquid cations obtained from primary and BOD experiments

Cationic head group	Side group R	Anion	Substance identifier	Classification OECD (...biodegradable) and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
		Cl ⁻	C ₃ OHmpip Cl	Inherently and ultimately	—	79 ± 2 ^a	This study
		Cl ⁻	C ₂ OHmpip Cl	Inherently	—	28 d: 29 ± 3 60 d: 85 ± 1	This study
		Cl ⁻	C ₁ CNmpip Cl	Biotic hydrolysis of the cyano group	Qualitative analysis by mass spectrometry	5 ± 2	This study
		Cl ⁻	C ₁₀ O ₂ mpip Cl	Not readily	-2 ± 2	—	This study
		Cl ⁻	C ₂ O ₁ mpip Cl	Not readily	1 ± 4	—	This study
		Cl ⁻	C ₃ mpip (CF ₃ SO ₂) ₂ N	Not readily	—	3 ± 2	This study
		(CF ₃ SO ₂) ₂ N ⁻	C ₃ mpip (CF ₃ SO ₂) ₂ N	Not readily	—	—	This study
		Br ⁻	C ₄ mpip Br	Not readily	—	-4 ± 3	This study
		—	—	—	—	—	—
		—	—	—	—	—	—
		—	—	—	—	—	—

^a Not within a 10-day window.

Table 4 Structural formula and results of pyridinium-based ionic liquid cations obtained from primary and BOD experiments. The data from this study are complemented with data from the literature

Cationic head group	Side chain R	Anion	Substance identifier	Classification OECD (...biodegradable) and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
		I ⁻	C ₂ OHPy I	Inherently and ultimately	100	65 ± 10	This study
		Cl ⁻	C ₂ OHPy Cl	Inherently	—	51 ± 0	This study
		Cl ⁻	C ₁ CNPy Cl	Biotic hydrolysis of the cyano group	Qualitative analysis by mass spectrometry	-2 ± 0	This study
		Cl ⁻	C ₂ Py Cl	Not readily	0	—	33
		Br ⁻	C ₃ Py Br	Not readily	—	1 ± 0	This study
		Br ⁻	C ₄ Py Br	Not readily	—	2	34
		—	—	—	—	—	—
		—	—	—	—	—	—
		—	—	—	—	—	—

gas-tight stoppers and stored in the dark at 20 ± 0.5 °C. The oxygen consumption was determined manometrically. Biodegradation of the test substance was calculated by the biological oxygen uptake (BOD) for the test substance (corrected by the oxygen demand of the blank samples) with respect to the theoretical oxygen demand (ThOD) of the substance and the amount of substance present in the sample. For the calculation of ThOD the cation was considered without including

its inorganic halide anion. The chemical formula can be given as $C_cH_hN_oO_p$ (eqn (1)):

$$\text{ThOD} = \frac{16 \left[2c + \frac{1}{2} (h - 3n) + \frac{5}{2} p - o \right] \frac{\text{mg}}{\text{mg}}}{\text{molecular mass of test substance} \frac{\text{mmol}}{\text{mg}}} \quad (1)$$



Table 5 Structural formula and results of imidazolium-based ionic liquid cations (functionalised side chains and an octyl side chain) obtained from primary and BOD experiments. The data from this study are complemented with data from the literature

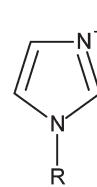
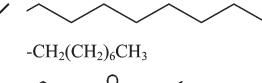
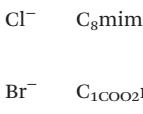
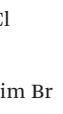
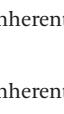
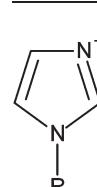
Cationic head group	Side chain R	Anion	Substance identifier	Classification OECD (...biodegradable) and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
	 -CH ₂ (CH ₂) ₆ CH ₃	Cl ⁻	C ₈ mim Cl	Inherently	100	—	33
	 -CH ₂ C(=O)OCH ₂ CH ₃	Br ⁻	C ₁ COO ₂ mim Br	Inherently	—	21	67
	 -CH ₂ CN	Cl ⁻	C ₁ CNmim Cl	Biotic hydrolysis of the cyano group	0	—	33
	 -CH ₂ CN				Qualitative analysis by mass spectrometry	0	This study
	 -CH ₂ CH ₂ OH	I	C ₂ OHmim I	Not readily	0	—	33
	 -CH ₂ CH ₂ CH ₂ OH	Cl ⁻	C ₃ OHmim Cl	Not readily	0	—	33
	 -CH ₂ OCH ₂ CH ₃	Cl ⁻	C ₁ O ₂ mim Cl	Not readily	0	—	33
	 -CH ₂ CH ₂ OCH ₃	Cl ⁻	C ₂ O ₁ mim Cl	Not readily	0	—	33
	 -CH ₂ CH ₂ OCH ₂ CH ₃	Br ⁻	C ₂ O ₂ mim Br	Not readily	0	—	33

Table 6 Structural formula and results of imidazolium-based ionic liquid cations (alkyl side chains) obtained from primary and BOD experiments. The data from this study are complemented with data from the literature

Cationic head group	Side chain R	Anion	Substance identifier	Classification OECD (...biodegradable) and biotic hydrolysis	Primary degradation in % (28 d) (n = 2)	Relative oxygen demand BOD/ThOD in % (28 d) (n = 2)	Reference
	 -CH ₂ CH ₃	Cl ⁻	C ₂ mim Cl	Not readily	0	—	33
	 -CH ₂ CH ₂ CH ₃	PF ₆ ⁻	C ₃ mim PF ₆	Not readily	—	2 ± 1	This study
	 -CH ₂ CH ₂ CH ₂ CH ₃	Cl ⁻	C ₄ mim Cl	Not readily	0	—	33,38,46,68

The reference substance benzoic acid was always degraded within 10 d, consuming around 80% of the ThOD and showing that the test was working reliably.

2 Primary biodegradation. The stock solutions of the test substances were prepared in deionised water. Each test substance was used at a final concentration of 50 $\mu\text{mol L}^{-1}$ in each of the two replicates of the sample vessels. The medium and sample vessels were prepared as described above. Additionally, one sample vessel was prepared for each experimental set up with aniline as positive control at a final concentration of 1.07 mmol L^{-1} (100 mg L^{-1}). Aniline was used as a reference

substance for positive biodegradability, since aniline is known to be biodegradable under the chosen test conditions within 14 d.⁶⁹ In this study aniline was fully biodegraded within ten days (limit of detection $<5 \mu\text{mol L}^{-1}$, data not shown), indicating the general biological activity of the inoculum. The vessels were stirred continuously in the dark with a magnetic bar. To keep the volume constant, evaporated water was frequently replaced with an equal volume of deionised water.

The concentration of the test substance was determined by ion chromatography. The primary biodegradation was then calculated from the concentration of the substance on a



specific day in comparison to that on day zero. The maximum deviation was calculated from the standard deviations of the ion chromatographic measurement according to common error propagation.

Examination of the analyte concentration and metabolites

Sampling and sample preparation for instrumental analysis.

The samples for the specific analysis were taken on the first day and thereafter at short time intervals for 28 d. On each testing day, 6 mL of the sample was centrifuged at approx. 3000g (Labofuge 400R, Heraeus Instruments GmbH, Germany) in 15 mL polypropylene centrifuge tubes (Sarstedt AG & Co., Germany). The samples were stored in a freezer at -20°C until all of the samples had been taken. The final specific analysis of the IL cation was then conducted by ion chromatography.

1 *Ion chromatography.* The ion chromatograph used was a "Metrohm 881 Compact IC pro" equipped with a "Metrohm 850 Conductivity Detector" connected to a "Metrohm 863 Compact Autosampler" (Metrohm AG, Switzerland). The "Metrosep C 4 50" cation exchange column with a "Metrosep C 4 50 Guard" pre-column was kept at 30°C in a column oven. The analytical method was developed on the basis of previous studies.⁴⁹

5 mmol L⁻¹ nitric acid (HNO₃) with 25% acetonitrile (ACN) were used as an eluent. The device was run at a flow rate of 0.9 mL min⁻¹ and the injection volume of each sample was 20 μL . The analytical method was developed to analyse most of the IL cations with one eluent solution. The data were evaluated using standard "Magic Net 1.1" software. An external standard for each substance was run during each measurement sequence to identify the IL peak and to check the consistency of the analytical method. Analytical quality parameters have already been published for other IL cations – these are generally in the range of $<0.33\text{ }\mu\text{M}$ and $<1\text{ }\mu\text{M}$ for the limit of detection and limit of quantification, respectively.⁴⁹

2 *Liquid chromatography-mass spectrometry.* In order to analyse potential degradation products, liquid chromatographic separation combined with mass spectrometric analysis was subsequently undertaken if necessary. The samples were diluted in methanol and deionised water (50 : 50) and analysed using an HP 1100 HPLC coupled to a Bruker Esquire ESI-MS ion trap detector. The chromatograph was run with 55% of 5 mmol L⁻¹ sodium formate pH 3.4 and 45% ACN at a flow rate of 0.5 mL min⁻¹. Column: Multohigh 150 \times 4 mm, 100 Si-5 μ Hilic (CS-Chromatographie Service GmbH, Langerwehe, Germany).

Mass spectra for cations were acquired in the positive ion mode in the scan range of m/z^+ 50–200. The ESI source conditions were set with a drying gas flow rate of 11 L min⁻¹, a drying gas temperature of 350°C and the nebulizer at 70 psi. Generally, all samples after the biodegradation experiments have been investigated with LC-MS to identify (if applicable) transformation products.

3 *Interpretation of the data.* The following effects that can lead to a false interpretation of the data were excluded:

(1) Excluding false positive results: the declaration of an IL as biodegradable although it is not may be due to the adsorption of the analyte on the bacteria and glass vessels, as well as to systematic deviations of the analytical test system. However, adsorption effects would only be observed within the first few days⁵⁰ and often in samples with a high organic load. As we used a negligible amount of sewage sludge flocs in the filtered test solutions, no adsorption effects were observed in our experiments. Systematic deviations in the analytical measurements were reduced by adjusting the results so as to relate them to the external standards run with each analytical sequence. Substances detected as being primarily biodegradable were further examined for their full biodegradability to confirm the results of the first run.

(2) Excluding false negative results: apart from the stringent test conditions, inhibition effects can also lead to a false classification of the data as "non-primarily biodegradable". However, these can be excluded at the concentrations used in the experiments. In a recent study the inhibitory potential towards activated sludge organisms of a wide structural variety of ILs was found to be low with inhibitory concentrations usually higher than 1500 μM .⁴⁸ At a concentration of 50 μM in the primary biodegradation tests and below 850 $\mu\text{mol L}^{-1}$ in BOD tests, inhibitory effects are therefore unlikely to be observed.

Results

The primary biodegradation of the test substances under aerobic conditions was monitored for 28 d. Specific analysis by ion chromatography (IC) using a conductivity detector yielded information on the concentration of the IL cation at a specific time of the experiment. If an IL cation was not degraded we assumed that it was "not readily biodegradable" in accordance with OECD 301. If primary biodegradation was observed, we used WTW OxiTop® devices measuring the biochemical oxygen demand (BOD) to determine the full mineralisation of the IL. In this full mineralisation test a compound was classified as readily biodegradable if it exhibited $>60\%$ degradation in relation to the theoretical oxygen demand (ThOD) within 28 d and an exponential growth phase of 10 d. In this study this result was attributable solely to the cation, because most of the IL anions investigated contained inorganic moieties ("halides") (Tables 1–6) that were irrelevant to biodegradation tests based on the measurement of oxidisable carbon in the molecule. The same held true for PF₆⁻ and the non-biodegradable anion (CF₃SO₂)₂N⁻.

The results of the primary biodegradation and full mineralisation experiments are given in Tables 1–6. For some ILs the standard test duration of 28 d was prolonged (in total up to 60 d) to investigate their long-term behaviour.

The results from primary degradability, the degradation curves derived from BOD measurements as well as more



detailed information on the used classification are given in the ESI† file.

Biodegradability of head groups

1 Pyrrolidinium compounds. In primary and mineralisation experiments the biodegradation rate of pyrrolidinium compounds varied between 0 and 100% and was dependent on the substituted side chain (Table 1). No biodegradation was found for the ethyl ($C_2\text{mpyr}$ $C_{2\text{O}}\text{SO}_3$) derivative, whereas $C_{3\text{OH}}\text{mpyr}$ Cl and $C_8\text{mpyr}$ Cl could be classified as readily biodegradable, exhibiting >60% degradation within 28 d. $C_{1\text{COO}2}\text{mpyr}$ Br was completely primarily degraded within one week, but although 20% of the ThOD was attained after just 5 d, no further degradation took place until the end of the standard experiment (28 d). The test procedure was prolonged, however, and after 38 d the whole compound was fully mineralised (74%) (Fig. 1).

The MS measurements of samples taken from the BOD experiment showed that after 5 and 28 d no parent compound could be detected any more, but that a degradation product with $m/z^+ = 144$ and the corresponding sodium and potassium adducts ($m/z^+ = 144 + 23 - \text{H}^+ = 166$ and $m/z^+ = 144 + 39 - \text{H}^+ = 182$) were formed (data not shown). This suggests that the ester bond was enzymatically hydrolysed (in abiotic samples the compound remained stable; data not shown) and $C_{1\text{COOH}}\text{mpyr}$ ($m/z^+ = 144$) was released and fully mineralised after an additional adaptation period of 20 d (Fig. 1). Degradation rates below 25% and strong deviations between replicates were observed for $C_{2\text{OH}}\text{mpyr}$ I and $C_4\text{mpyr}$ Br; however, prolonging the test to 60 d enabled both compounds to be mineralised. According to OECD, $C_{1\text{COO}2}\text{mpyr}$ Br and $C_4\text{mpyr}$ Br could be classified as “not readily” but “inherently biodegradable”, *i.e.* the chemicals have potential for biodegradation under aerobic conditions. For $C_{2\text{OH}}\text{mpyr}$ I the deviations between multiple runs and experiments were strong even after prolonging the test duration (around 40%) so that this

compound can only be classified as “not readily”, but with hints of being inherently biodegradable. Primary degradation, but no BOD was observed for $C_{1\text{CN}}\text{mpyr}$ Cl; similar findings have been noted for other head groups with a cyanomethyl side chain (Tables 1–6) and will be presented in a separate paragraph.

2 Morphinium compounds. None of the investigated morpholinium compounds showed significant primary biodegradation apart from the $C_{3\text{OH}}\text{mmor}$ and the $C_{1\text{CN}}\text{mmor}$ cation (Table 2). At the end of the primary biodegradation with $C_{3\text{OH}}\text{mmor}$ a slight decrease in concentration ($12 \pm 8\%$) was observed, so an additional BOD experiment of prolonged duration was performed. In these experiments oxygen consumption began after 25 d and reached levels from 20 to 42% biodegradation (deviation between replicates). After 41 d total biodegradation of $59 \pm 10\%$ degradation was observed, leading to the classification “inherently biodegradable”.

3 Piperidinium compounds. Among the piperidinium compounds tested only $C_{3\text{OH}}\text{mpip}$ Cl and $C_{2\text{OH}}\text{mpip}$ Cl were fully mineralised (Table 3). Pip13OH Cl was degraded up to $79 \pm 2\%$ after 28 d. The 60% pass level was exceeded, but not within a ten day window. The compound was thus classified as “inherently, ultimately biodegradable” rather than “readily biodegradable”. The biodegradation of $C_{2\text{OH}}\text{mpip}$ Cl did not exceed the 60% pass level after 28 d, but after a prolongation of the test duration to 60 d more than 80% degradation was observed, leading to the classification “inherently biodegradable”.

4 Pyridinium compounds. The *N*-alkylpyridinium cations ($C_2\text{py}$, $C_3\text{py}$ and $C_4\text{py}$) did not exhibit significant levels of biodegradation, whereas the hydroxylated compounds ($C_{2\text{OH}}\text{py}$ and $C_{3\text{OH}}\text{py}$) showed good biodegradation rates of $65 \pm 10\%$ and $51 \pm 0\%$, respectively, permitting classification of “inherently biodegradable” (Table 4).

5 Imidazolium compounds. The corresponding imidazolium-based ILs with similar side chains as used for the experiments described above have already been examined by different working groups (Table 5). In this study the biodegradability of imidazolium cations with propyl side chains was investigated additionally, but no biodegradation was observed for this compound (Table 6).

6 Degradation of the cyanomethyl side chain. Regardless of the head group, compounds with a cyanomethyl side chain ($C_{1\text{CN}}\text{mpyr}$ Cl, $C_{1\text{CN}}\text{mmor}$ Cl, $C_{1\text{CN}}\text{mpip}$ Cl, $C_{1\text{CN}}\text{py}$ Cl, and $C_{1\text{CN}}\text{mim}$ Cl) exhibited primary degradation within 28 d, but no oxygen consumption in BOD measurements. LC-MS analysis of the 28 d samples revealed the occurrence of several mass-to-charge ratios suggesting the hydrolysis of the CN group. The corresponding carboxamide ($m/z^+ = 159$) and carboxylic acid detected as sodium ($m/z^+ = 182$) and potassium adducts ($m/z^+ = 198$) were observed in the mass spectrum (Fig. 2).

The methylated cation $\text{R}^+-\text{CH}_2-\text{C}(=\text{O})\text{O}-\text{CH}_3$ (Fig. 2) is formed by an esterification reaction of the free acid and with the methanol solvent under analytical conditions (pH 3.5). The same detection pattern was observed for all analogous IL

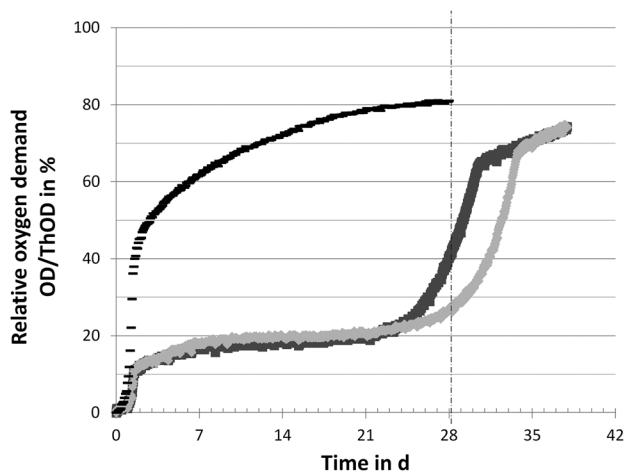


Fig. 1 Biodegradation curves (duplicates in grey: ■ and ◆) derived from BOD measurements for Pyr11COO₂ Br. The black line (—) represents the reference compound benzoic acid.



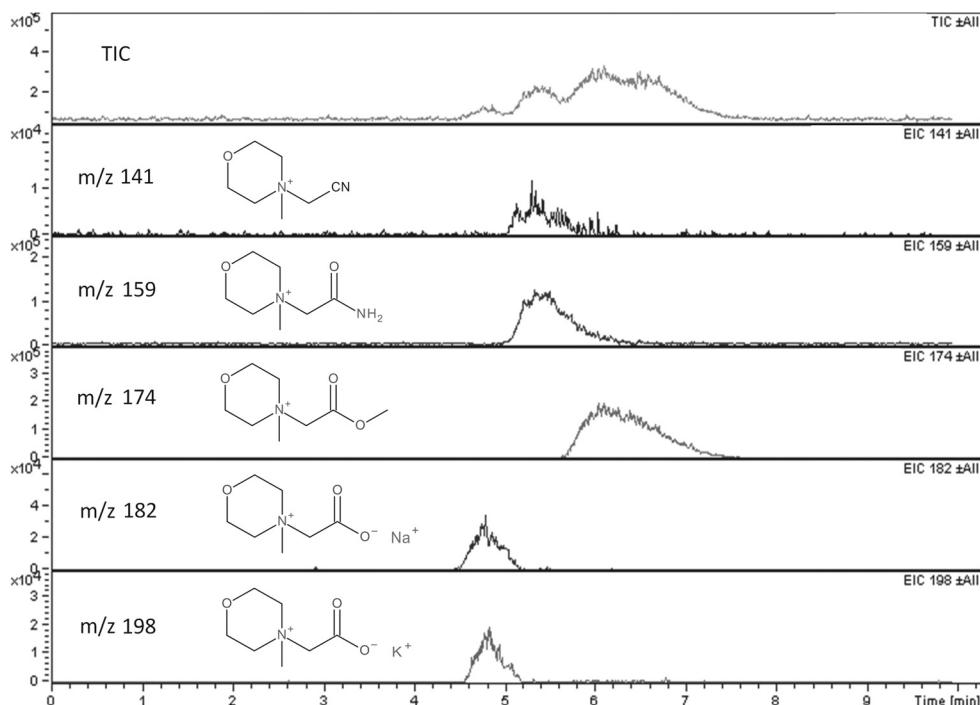


Fig. 2 The total ion current (TIC) over time from LC-MS measurements of the 28 d sample from the BOD biodegradation experiment on $C_{1CN}mmpyCl$. The extracted ions presumably belong to the molecular structures shown.

cations: $C_{1CN}py$, $C_{1CN}mpyr$, $C_{1CN}mmpy$, $C_{1CN}mim$ and $C_{1CN}mpip$. In the standard solutions no such transformation products could be found, indicating that microorganisms are involved in the cleavage. In the case of $C_{1CN}mpyr$ the corresponding acid ($C_{1COOH}mpyr$) formed was the same transformation product as that obtained in the experiments with $C_{1COO_2}mpyr$. $C_{1COO_2}mpyr$ was found to be fully mineralised in a prolonged biodegradation test.

Discussion

Structure-biodegradability relationships

Most of the degradable compounds are among the pyrrolidinium cations. When substituted with hydroxyl-containing side chains and when enough time is allowed, full mineralisation (side chain and core) takes place. The biodegradability of 1-alkyl-1-methylpyrrolidinium compounds increased with lengthening side chain, changing from “not readily biodegradable” (ethyl) to “inherently biodegradable” (butyl) and “readily biodegradable” (octyl). A similar trend has been reported for 1-alkyl-3-methylimidazolium and 1-alkyl-3-methylpyridinium cations.^{32,33} Unfortunately, the influence of the side chain could not be investigated for the other non-aromatic head groups, since these compounds with an octyl side chain were unavailable.

Good biodegradation rates of IL compounds with substituted hydroxyl side chains were not only obtained for pyrrolidinium cations, but also for morpholinium, piperidinium and pyridinium ones. Several of these compounds were

fully mineralised, especially when a hydroxypropyl or even a hydroxyethyl residue was attached. The improved biodegradability of compounds containing alcohol groups – a potential target of enzymatic degradation – has been reported for several other substance classes²⁶ for instance, imidazolium²⁹ and ammonium-based ILs.⁵¹ Apart from the above-mentioned hydroxylated compounds, none of the other investigated morpholinium, piperidinium and pyridinium compounds were mineralised. In comparison with the other head groups the imidazolium group exhibited the lowest biodegradation potential. None of the investigated functionalised methyl imidazolium compounds were degraded and only C_8mimCl was primarily degraded. The core proved to be recalcitrant towards biodegradation in former studies and only the side chain was degraded *via* β -oxidation, yielding 1-(3-carboxyethyl)-3-methylimidazolium.³³

In this study one odd-numbered alkyl side chain was investigated to check whether β -oxidation then leads to products such as 1-(3-carboxypropyl)-3-methylimidazolium that might be further degraded (full mineralisation of the ring). This hypothesis was not confirmed and the propyl compound showed no biodegradation potential within 28 d.

With respect to the side chains a microorganism enabled primary degradation of the cyanomethyl side chain, and the identified transformation products suggest enzymatic hydrolysis *via* nitrile degrading enzymes such as nitrilases or nitrile hydratases together with amidase. These enzymes are commonly applied as catalysts in organic synthesis for the hydrolysis of nitrile groups in the pharmaceutical industry and for bioremediation purposes, amongst others.⁵²⁻⁵⁵ Recently, IL



anions such as $\text{N}(\text{CN})_2^-$, $\text{C}(\text{CN})_3^-$ and $\text{B}(\text{CN})_4^-$ were hydrolysed to their corresponding amides with isolated nitrile hydratase.⁵⁶

Variability and adaptation time

The data presented in this study show some inconsistencies in comparison with the literature data, and in some cases there were evident discrepancies between our independently performed experiments (e.g. $\text{C}_{2\text{OH}}\text{mpyr}$ I and $\text{C}_{3\text{OH}}\text{mpyr}$ Cl). Although the rapidly biodegradable reference compound benzoic acid was always fully degraded within the first week, the biodegradation of the chosen xenobiotics seems to be more dependent on the biological variability. In particular, the nature of the inoculum in terms of spatial and temporal variations is a very variable factor in the assessment of biodegradability.⁵⁷ Hence, during every experiment and within each test vessel a unique microorganism community is present, which may result in completely different degradation results, especially under the stringent test conditions of the OECD guideline.⁵⁸ For example, we found no primary degradation of the $\text{C}_{1\text{CN}}\text{mim}$ cation in our previous study,³³ but this did occur in the present study. The same holds true for observations made for pyridinium compounds. In studies performed independently by Docherty *et al.*⁵⁹ and Pham *et al.*,⁶⁰ vastly different biodegradation rates and different degradation pathways were found for one IL cation (*N*-butyl-3-methylpyridinium). In the first study there was no biodegradation within 43 d,³² but after re-examination⁵⁹ by the same group, mineralisation was nearly complete (88% within 41 d); this was corroborated in primary biodegradation studies by Pham *et al.*⁶⁰ In the present study *N*-butylpyridinium, for example, was investigated and no biodegradation was observed, but whether a different substitution pattern of the pyridinium core or the microorganisms was responsible for degradation/recalcitrance is a question that remains unanswered. The results obtained for $\text{C}_{1\text{COO}_2}\text{mpyr}$ suggest that the microbial composition changes during the test (Fig. 1). In the first step the ester is degraded enzymatically, after which there is a long lag phase of around 20 d, followed in the second step by additional exponential growth and another lag phase. It can be assumed that other microorganisms or enzymes are involved in the degradation of the side chain and the subsequent decomposition of the core. Such a symbiosis of different microorganisms from a test community has been described for the biodegradation of aliphatic and aromatic hydrocarbons.⁶¹ As a consequence of different substrate availabilities, the subsequent activation of enzymes resulting in the diauxic growth of microorganisms has been successfully modeled for different sugars and organic anions.⁶²

For several compounds ($\text{C}_{1\text{COO}_2}\text{mpyr}$ Br, $\text{C}_{2\text{OH}}\text{mpyr}$ I, C_4mpyr Br, $\text{C}_{2\text{OH}}\text{mpip}$ Cl, $\text{C}_{3\text{OH}}\text{mmor}$ Cl) the prolonged test duration leads to an increased biodegradation rate. Long-term exposure of microorganisms to ILs is presumably associated with adaptation processes, such as the induction of specific enzymes, genetic mutation or horizontal gene transfer, which enhance the degradative capacity of the entire community or

change the population in terms of the selective growth of certain strains.⁶³ Such long-term adaptation processes and an increased biodegradation potential (including the complete degradation of the imidazolium ring) were recently observed for C_8mim , too.⁶⁴ Moreover, cultures of axenic bacterial strains, such as *Corynebacterium* sp. and *Sphingomonas paucimobilis*, or isolated enzymes have successfully enhanced the biodegradability of certain IL cations^{41,42} or IL anions,⁵⁶ and a wide range of IL structures can be generally targeted for microbial degradation.

All in all, for the screening purposes of this study, those stringent test conditions make it easier to identify ILs with improved biodegradation potential. For the final determination of persistency, further tests need to be conducted.

Conclusion

A comparison of the biodegradation potential shows that for all the five head groups investigated, compounds can be found that are readily or inherently biodegradable. It is highly likely that chemicals classified as “readily biodegradable” are biodegradable in the environment with just a low risk of persistence, or even none at all. This cannot generally be assumed for inherently biodegradable chemicals, although they do have a reduced risk of being persistent. From the structural design point of view, both the type of head group as well as the substituted side chain influence the degradability of the IL cation. When the same side chain is substituted, pyrrolidinium and pyridinium cores generally display better degradation rates in comparison to piperidinium and morpholinium cations. Imidazolium appears to be the most refractory head group. The general ability of microorganisms to adapt to IL cations again reduces the risk of persistence. The degree of mineralisation, however, also seems to depend strongly on the inoculum and the test/environmental conditions. The underlying science behind IL biodegradation, namely the microbiological processes and the microorganisms involved, is not yet well understood, so detailed investigations are needed. The environmental risk of poorly degradable ILs can be eliminated when these are used in closed systems or can be reduced when residues are removed from waste water by separation techniques such as nanofiltration⁶⁵ or advanced oxidation processes.^{33,66}

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References

- 1 C. Yue, D. Fang, L. Liu and T.-F. Yi, *J. Mol. Liq.*, 2011, **163**, 99–121.
- 2 T. Welton, *Chem. Rev.*, 1999, **99**, 2071–2084.
- 3 M. Petkovic, K. R. Seddon, L. P. N. Rebelo and C. Silva Pereira, *Chem. Soc. Rev.*, 2011, **40**, 1383–1403.
- 4 A. Kokorin, *Ionic liquids: Applications and Perspectives*, InTech, Rijeka, Croatia, 2011.
- 5 N. V. Plechkova and K. R. Seddon, *Chem. Soc. Rev.*, 2008, **37**, 123–150.
- 6 T. Tsuda, K. Kondo, T. Tomioka, Y. Takahashi, H. Matsumoto, S. Kuwabata and C. L. Hussey, *Angew. Chem., Int. Ed.*, 2011, **50**, 1310–1313.
- 7 W. Sun, C. X. Guo, Z. Zhu and C. M. Li, *Electrochem. Commun.*, 2009, **11**, 2105–2108.
- 8 D. Wei and A. Ivaska, *Anal. Chim. Acta*, 2008, **607**, 126–135.
- 9 V. V. Singh, A. K. Nigam, A. Batra, M. Boopathi, B. Singh and R. Vijayaraghavan, *Int. J. Electrochem.*, 2012, **2012**, 1–19.
- 10 Z. Tan, J. Liu and L. Pang, *TrAC, Trends Anal. Chem.*, 2012, **39**, 218–227.
- 11 P. Sun and D. W. Armstrong, *Anal. Chim. Acta*, 2010, **661**, 1–16.
- 12 A. Berthod, M. J. Ruiz-Angel and S. Carda-Broch, *J. Chromatogr., A*, 2008, **1184**, 6–18.
- 13 R. Zhang, N. Li, C. Wang, Y. Bai, R. Ren, S. Gao, W. Yu, T. Zhao and H. Zhang, *Anal. Chim. Acta*, 2011, **704**, 98–109.
- 14 S. Keskin, D. Kayrak-Talay, U. Akman and Ö. Hortaçsu, *J. Supercrit. Fluids*, 2007, **43**, 150–180.
- 15 K. M. S. Meera, R. M. Sankar, S. N. Jaisankar and A. B. Mandal, *Colloids Surf., B Biointerfaces*, 2011, **86**, 292–297.
- 16 M. Moniruzzaman and M. Goto, *J. Chem. Eng. Jpn.*, 2011, **44**, 370–381.
- 17 W. L. Hough, M. Smiglak, H. Rodríguez, R. P. Swatloski, S. K. Spear, D. T. Daly, J. Pernak, J. E. Grisel, R. D. Carliss, M. D. Soutullo, J. H. Davis, Jr. and R. D. Rogers, *New J. Chem.*, 2007, **31**, 1429–1436.
- 18 R. Vijayaraghavan, N. Vedaraman, M. Surianarayanan and D. R. MacFarlane, *Talanta*, 2006, **69**, 1059–1062.
- 19 M. G. Freire, A. F. M. Cláudio, J. M. M. Araújo, J. A. P. Coutinho, I. M. Marrucho, J. N. Canongia Lopes and L. P. N. Rebelo, *Chem. Soc. Rev.*, 2012, **41**, 4966–4995.
- 20 P. J. Carvalho and J. A. P. Coutinho, *Energy Environ. Sci.*, 2011, **4**, 4614–4619.
- 21 F. Zhou, Y. Liang and W. Liu, *Chem. Soc. Rev.*, 2009, **38**, 2590–2599.
- 22 S. Schneider, T. Hawkins, Y. Ahmed, M. Rosander, L. Hudgens and J. Mills, *Angew. Chem., Int. Ed.*, 2011, **50**, 5886–5888.
- 23 M. Deetlefs and K. R. Seddon, *Green Chem.*, 2010, **12**, 17–30.
- 24 P. Anastas and N. Eghbali, *Chem. Soc. Rev.*, 2010, **39**, 301–312.
- 25 B. Jastorff, K. Möller, P. Behrend, U. Bottin-Weber, J. Filser, A. Heimers, B. Ondruschka, J. Ranke, M. Schaefer, H. Schröder, A. Stark, P. Stepnowski, F. Stock, R. Störmann, S. Stolte, U. Welz-Biermann, S. Ziegert and J. Thöming, *Green Chem.*, 2005, **7**, 362–372.
- 26 R. S. Boethling, E. Sommer and D. DiFiore, *Chem. Rev.*, 2007, **107**, 2207–2227.
- 27 J. Ranke, S. Stolte, R. Störmann, J. Arning and B. Jastorff, *Chem. Rev.*, 2007, **107**, 2183–2206.
- 28 N. V. Ignat'ev, U. Welz-Biermann, A. Kucheryna, G. Bissky and H. Willner, *J. Fluorine Chem.*, 2005, **126**, 1150–1159.
- 29 S. Stolte, S. Steudte, A. Igartua and P. Stepnowski, *Curr. Org. Chem.*, 2011, **15**, 1946–1973.
- 30 T. P. T. Pham, C.-W. Cho and Y.-S. Yun, *Water Res.*, 2010, **44**, 352–372.
- 31 D. Coleman and N. Gathergood, *Chem. Soc. Rev.*, 2010, **39**, 600–637.
- 32 K. M. Docherty, J. K. Dixon and C. F. Kulpa Jr., *Biodegradation*, 2007, **18**, 481–493.
- 33 S. Stolte, S. Abdulkarim, J. Arning, A.-K. Blomeyer-Nienstedt, U. Bottin-Weber, M. Matzke, J. Ranke, B. Jastorff and J. Thöming, *Green Chem.*, 2008, **10**, 214–224.
- 34 J. R. Harjani, R. D. Singer, M. T. Garcia and P. J. Scammells, *Green Chem.*, 2009, **11**, 83–90.
- 35 T. P. T. Pham, C.-W. Cho, J. Min and Y.-S. Yun, *J. Biosci. Bioeng.*, 2008, **105**, 425–428.
- 36 R. P. Swatloski, J. D. Holbrey and R. D. Rogers, *Green Chem.*, 2003, **5**, 361–363.
- 37 J. R. Harjani, J. Farrell, M. T. Garcia, R. D. Singer and P. J. Scammells, *Green Chem.*, 2009, **11**, 821–829.
- 38 M. T. Garcia, N. Gathergood and P. J. Scammells, *Green Chem.*, 2005, **7**, 9–14.
- 39 S. Steudte, J. Neumann, U. Bottin-Weber, M. Diedenhofen, J. Arning, P. Stepnowski and S. Stolte, *Green Chem.*, 2012, **14**, 2474–2483.
- 40 J. Neumann, C.-W. Cho, S. Steudte, J. Köser, M. Uerdingen, J. Thöming and S. Stolte, *Green Chem.*, 2012, **14**, 410–418.
- 41 C. Zhang, H. Wang, S. V. Malhotra, C. J. Dodge and A. J. Francis, *Green Chem.*, 2010, **12**, 851–858.
- 42 S. Kumar, W. Ruth, B. Sprenger and U. Kragl, *Chem. Today*, 2006, **24**, 70–72.
- 43 J. B. Wesnigk, M. Keskin, W. Jonas, K. Figge and G. Rheinheimer, in *The Handbook of Environmental Chemistry Vol. 2, Part K: Biodegradation and Persistence*, ed. B. Beek, Springer-Verlag, Berlin, Heidelberg, 2001, vol. 2, pp. 253–290.
- 44 C. Pretti, M. Renzi, S. E. Focardi, A. Giovani, G. Monni, B. Melai, S. Rajamani and C. Chiappe, *Ecotoxicol. Environ. Saf.*, 2011, **74**, 748–753.
- 45 F. Atefi, M. T. Garcia, R. D. Singer and P. J. Scammells, *Green Chem.*, 2009, **11**, 1595–1604.
- 46 A. S. Wells and V. T. Coombe, *Org. Process Res. Dev.*, 2006, **10**, 794–798.



47 Organisation for Economic Co-operation and Development (OECD), *OECD guideline for testing of chemicals 301 - Ready Biodegradability*, 1992.

48 M. Markiewicz, M. Piszora, N. Caicedo, C. Jungnickel and S. Stolte, *Water Res.*, 2013, **47**, 2921–2928.

49 S. Stolte, S. Steudte, A. Markowska, J. Arning, J. Neumann and P. Stepnowski, *Anal. Methods*, 2011, **3**, 919–926.

50 M. Markiewicz, C. Jungnickel, A. Markowska, U. Szczepaniak, M. Paszkiewicz and J. Hupka, *Molecules*, 2009, **14**, 4396–4405.

51 B. Peric, J. Sierra, E. Martí, R. Cruañas, M. A. Garau, J. Arning, U. Bottin-Weber and S. Stolte, *J. Hazard. Mater.*, 2013, **261**, 99–105.

52 A. Banerjee, R. Sharma and U. C. Banerjee, *Appl. Microbiol. Biotechnol.*, 2002, **60**, 33–44.

53 M. Kobayashi and S. Shimizu, *Nat. Biotechnol.*, 1998, **16**, 733–736.

54 P. K. Mascharak, *Coord. Chem. Rev.*, 2002, **225**, 201–214.

55 R. Singh, R. Sharma, N. Tewari and D. S. Rawat, *Chem. Biodiversity*, 2006, **3**, 1279–1287.

56 J. Neumann, M. Pawlik, D. Bryniok, J. Thöming and S. Stolte, *Environ. Sci. Pollut. Res. Int.*, 2013, DOI: 10.1007/s11356-013-2341-2.

57 G. Vázquez-Rodríguez, R. I. Beltrán-Hernández, C. Coronel-Olivares and J.-L. Rols, *Anal. Bioanal. Chem.*, 2011, **401**, 1127–1137.

58 A. K. Goodhead, I. M. Head, J. R. Snape and R. J. Davenport, *Environ. Sci. Pollut. Res. Int.*, 2013, DOI: 10.1007/s11356-013-2064-4.

59 K. M. Docherty, M. V. Joyce, K. J. Kulacki and C. F. Kulpa, *Green Chem.*, 2010, **12**, 701–712.

60 T. P. T. Pham, C.-W. Cho, C.-O. Jeon, Y.-J. Chung, M.-W. Lee and Y.-S. Yun, *Environ. Sci. Technol.*, 2009, **43**, 516–521.

61 L. G. Whyte, L. Bourbonnière and C. W. Greer, *Appl. Environ. Microbiol.*, 1997, **63**.

62 B. W. Brandt, F. D. L. Kelpin, I. M. M. van Leeuwen and S. A. L. M. Kooijman, *Water Res.*, 2004, **38**, 1003–1013.

63 M. Devers, N. Rouard and F. Martin-Laurent, *Environ. Microbiol.*, 2008, **10**, 676–684.

64 M. Markiewicz, S. Stolte, Z. Lustig, J. Łuczak, M. Skup, J. Hupka and C. Jungnickel, *J. Hazard. Mater.*, 2011, **195**, 378–382.

65 J. F. Fernandez, J. Neumann and J. Thöming, *Curr. Org. Chem.*, 2011, **15**, 1992–2014.

66 P. Stepnowski and A. Zaleska, *J. Photochem. Photobiol. A*, 2005, **170**, 45–50.

67 N. Gathergood, M. T. Garcia and P. J. Scammells, *Green Chem.*, 2004, **6**, 166–175.

68 A. Romero, A. Santos, J. Tojo and A. Rodríguez, *J. Hazard. Mater.*, 2008, **151**, 268–273.

69 OECD Guideline for Testing of Chemicals No. 302B - Zahn-Wellens/EMPA Test, 1992.

