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Evaluation of bioaugmentation and biostimulation effects on the treatment of refinery oily sludge using 2^n full factorial design

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Bioremediation approaches for the treatment of oily sludge from a refinery were evaluated using a 2^3 factorial design. The three strategies tested were bioaugmentation with indigenous microbial consortia (MO) isolated from oily sludge, biostimulation with nutrients (NP) and biostimulation with the surfactant Triton X-100 (TX). Eight experimental runs were conducted in triplicate with factor settings \pm (high/low) as per the 2^3 design. The main effects and the effects of various interactions of the factors on oil degradation and microbial growth in suspension were evaluated during a 30 day study. Multifactor ANOVA could reveal the significant effects while the normal order score approach failed in this scenario. The main effect of biostimulation with nutrients in the form of nitrate and phosphate, as well as biostimulation with Triton X-100, was positive and significant when both oil degradation and microbial growth in suspension were chosen as the response variables. However, the main effect of bioaugmentation was only significant for oil degradation but was insignificant for microbial growth at a 90% confidence level. The MO–NP binary interaction and the MO–NP–TX ternary interactions were positive and significant, indicating the synergistic effect of these strategies on oil degradation and microbial growth. All other binary interactions were found to be insignificant.

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Environmental impact

Proper management of oily sludge is essential for preventing soil and groundwater contamination. In this study, a 2^3 full factorial design is used to evaluate the impact of various bioremediation strategies for oily sludge decontamination, *i.e.*, bioaugmentation with indigenous microorganisms (MO), biostimulation with nutrients (NP) and biostimulation with surfactants (TX). This design reveals effective strategies while minimizing the experimental runs. Batch biodegradation studies were conducted over a period of 30 days. The effects were computed based on the 2^3 design and statistically significant effects were identified based on ANOVA. The main effects of the nutrients, surfactant and bioaugmentation on oil degradation were all positive and significant. Significant synergistic effects among the various strategies were also observed.

1. Introduction

Oily sludge is generated from petroleum refineries in huge quantities as a byproduct of various processes and operations. A massive amount of oily sludge is also disposed off from the bottom of crude oil storage tanks during cleaning and maintenance. The handling and disposal of such huge volumes of waste is a major challenge. Oily sludge is a complex mixture of petroleum hydrocarbons and other solids, including heavy metals, that are carcinogenic and potent immunotoxicants. Apart from physicochemical treatment technologies, bioremediation is a clean, environmentally friendly treatment technology that can be used for the degradation of oily sludge generated in oil refineries.^{1–3}

Although they have been successfully applied, landfarming approaches have certain limitations in terms of their large space and time requirements and air pollution due to the emission of volatile organic compounds. In contrast, slurry phase degradation can provide rapid and extensive degradation of oil by enhancing mass transfer rates and promoting interaction among microorganisms, pollutants and nutrients.^{4–6}

Two approaches can be used to enhance the bioremediation process *i.e.*, bioaugmentation with a native or tailored microbial consortium and biostimulation with nutrients and surfactants in controlled batch slurry systems.^{4,7–11} However, the effect of such approaches is not beneficial in all scenarios. Possible reasons for this could be site specific features such as soil or sludge type, the distribution of contaminated hydrocarbons and the presence/distribution of indigenous microorganisms capable of degrading the contaminants.^{8,12,13}

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A mixed consortium of microorganisms may exhibit various modes of hydrocarbon uptake, such as: uptake of soluble hydrocarbons, uptake of emulsified forms or direct interfacial uptake facilitated by the development of hydrophobic cell surfaces. Bioaugmentation with mixed microbial consortia is preferred due to their wide metabolic networks, through which they can easily assimilate the complex hydrocarbons in oily sludge or oil contaminated soils.^{9,14–18} Moreover, the enrichment of indigenous microorganisms and bioaugmentation with these enriched microorganisms which are well adapted to the contaminated environment has been recommended by various researchers.^{10,15} Biostimulation with surfactants may increase the bioavailability of hydrocarbons by emulsification and also by altering the microbial cell surface properties so as to increase the interaction between the microbes and hydrocarbon contaminants.^{2,11,12,17–20} Often oil contaminated soil or oily sludge is found to have a much higher carbon content in comparison to nitrogen (N) and phosphorous (P). An adequate supply of N and P is essential for microbial growth and contaminant degradation. Moreover, during the course of natural attenuation, nutrient depletion may lead to a reduction in the indigenous microbial population. Thus, biostimulation with nutrients in the form of N and P is often found to induce contaminant degradation by the native microbial population, while the activity of bioaugmented cultures is also enhanced.^{2,11,12,21}

Laboratory feasibility tests for bioremediation are essential to determine the potential of indigenous microbes to degrade pollutants and to evaluate strategies for optimizing the rate and extent of degradation before pilot/full scale design of *in situ* or *ex situ* treatment schemes. Various studies focusing on comparative treatment studies of different bioremediation strategies make use of statistical tools to evaluate the significant differences between treatments.^{3,11,12} In this study a 2ⁿ full factorial design was used to investigate the effect of various factors so as to identify an appropriate treatment scheme for bioremediation of oily sludge from a refinery. This design can be used for screening a number of independent factors while minimizing the number of experimental runs. Statistical analysis of the results can provide useful information on bioremediation strategies. Three factors likely to influence biodegradation of oily sludge in laboratory batch systems were identified as bioaugmentation with indigenous microorganisms, biostimulation with nutrients and biostimulation with surfactants. Two different response variables were chosen, *i.e.*, oil degradation in the system and microbial growth in suspension after 30 days. A key objective was to determine the main effect of each of these factors and the effect of interactions between various factors so as to reveal possible synergism/antagonism among the three strategies. Although these strategies are commonly employed for oily sludge bioremediation, synergistic/antagonistic interactions among these strategies have not been explored by other researchers. The effects were quantified based on the 2³ design and the significance of these effects was determined based on analysis of variance (ANOVA).

2. Materials and methods

2.1. Source of chemicals

The chemical surfactant Triton X-100 was procured from SD Fine Chemicals Pvt. Ltd. (Mumbai, India). The various chemicals used for preparation of mineral media, nutrient broth and bacteriological agar were procured from SD Fine Chemicals Pvt. Ltd., SRL industries Ltd., Merck and Hi Media Pvt. Ltd. The high purity dichloromethane (DCM) used for extraction was obtained from Merck (India).

2.2. Source of oily sludge

The oily sludge was obtained from the weathering pits of a petroleum refinery in Mumbai (India) during August, 2010. The sludge was dewatered and centrifuged under high pressure to recover almost 90% of the crude oil prior to its disposal in the weathering pits. The sludge was collected from the refinery site, dried, sieved, homogenized and stored at 4 °C. The batch biodegradation studies reported here were conducted almost two years after the sludge was collected (October–November, 2012).

2.3. Isolation and enrichment of microorganisms from refinery oily sludge

Laboratory experiments were conducted to isolate and enrich microorganisms from the oily sludge. Enrichment was conducted in 500 mL flasks containing 100 mL mineral medium¹⁷ with 0.5% (w/v) oil extracted from the sludge as the sole substrate. Isolation of pure cultures was carried out both by spread plating and streak plating on nutrient agar plates. The isolates were identified based on 16S rDNA sequencing (Macrogen, Inc., Korea) followed by BLAST analysis (NCBI) and were identified based on the closest match with available sequences in the database. Five pure cultures were combined and used for bioaugmentation in the oily sludge biodegradation studies.

2.4. 2ⁿ full factorial design

A 2³ full factorial design was utilized in which eight runs were conducted at an appropriate setting for each factor. The three factors were chosen as bioaugmentation with indigenous cultures isolated from sludge (MO), biostimulation with nutrients (NP) and biostimulation with Triton X-100 (TX), a chemical surfactant. The effects of the three factors were studied on two response variables *i.e.*, % degradation of total petroleum hydrocarbons (TPH) in sludge over 30 days and increase in viable count in the aqueous phase ($\ln(N/N_0)$, where N_0 and N are viable count in the aqueous phase at 0 and at 30 days). Eight runs were conducted as per the design matrix for the 2³ full factorial design to determine the main effects, and the effects of binary and ternary interactions.²² The level of the three factors (\pm) in each run was controlled as per the design matrix. The normal plot approach and analysis of variance (ANOVA) approach were both used to evaluate the significant main effect and the interaction effects. The ANOVA was performed using STATISTICA ver.8.



2.5. Biodegradation study

A thirty day long biodegradation study was set-up as per the full factorial design matrix. For each run as specified in this matrix, triplicate batches were set-up. Each batch flask (500 mL) contained 10% (w/v) oily sludge in 100 mL mineral media (MM). Biostimulation consisted of the addition of nutrients in the form of nitrate-nitrogen ($\text{NO}_3\text{-N}$) and phosphate-phosphorous ($\text{PO}_4\text{-P}$). The MM already contained some N and P at a baseline level (–) of 222.4 mg L^{-1} N as $\text{NH}_4\text{-N}$ and 198.66 mg L^{-1} P as $\text{PO}_4\text{-P}$ such that the N : P ratio was 1.21 : 1 (mass basis). For nutrient addition (NP+), additional N and P *i.e.*, 70 mg L^{-1} $\text{NO}_3\text{-N}$ and 31.2 mg L^{-1} $\text{PO}_4\text{-P}$ were supplemented in the medium in the forms of KNO_3 and KH_2PO_4 , respectively. A nonionic surfactant, Triton X-100, was also used to study its effect on the extent of oily sludge degradation. Flasks biostimulated with Triton X-100 (TX+) contained the surfactant at twice the critical micelle concentration (CMC). For bioaugmentation (MO+), 5 mL of the 5-member reconstituted consortium adjusted to unit absorbance at 600 nm was added as inoculum. These cultures enriched and isolated from the oily sludge were maintained in the laboratory using 0.5% (w/v) oil extracted from the sludge. The reconstituted consortium was prepared by the addition of each of the strains in equal proportions after growing them up to end of the log phase.

The flasks were sacrificed initially (*i.e.*, day 0) and after 30 days of incubation at 35°C and 150 rpm in a rotary shaker. The sludge slurry contained in the flasks was filtered. The residual sludge collected in the filter paper was further analyzed for the residual oil content at the outset *i.e.*, day 0, and after 30 days. The aqueous suspension obtained was analyzed for pH, viable cell count and total organic carbon (TOC) content. Moreover, liquid-liquid extraction using dichloromethane as a solvent (in a ratio of 1 : 1)¹⁹ was also performed to estimate oil/total petroleum hydrocarbons (TPH) in the aqueous suspension. Residual TPH in the sludge was estimated by soxhlet extraction using dichloromethane as a solvent followed by gravimetric analysis and % degradation of oil was computed based on the zero day values. Standard error (SE) was based on the propagation of error for studies conducted in the triplicate setup. Viable counts in the aqueous phase of the sludge slurry set-up were determined through a standard plate count procedure at 0 and 30 days, and $\ln(N/N_0)$ was determined as a measure of culture growth in suspension.

3. Results and discussion

Oily sludge is a natural material that is often reported to have microbial cultures associated with it. In batch biodegradation studies with this oily refinery sludge, Jasmine and Mukherji²³ have earlier demonstrated that in batch systems where no microbial cultures were added (un-spiked controls), a large variability in oil biodegradation was observed ($44 \pm 10\%$ over 30 days). In contrast, systems containing 0.1% sodium azide added as a biocide showed negligible oil degradation ($6 \pm 4.5\%$ over 30 days). Loss of oil in the un-spiked controls was found to increase progressively with time and this was coupled with an

increase in the culture count in the aqueous phase. In some cases, biodegradation in the un-spiked controls was almost comparable to systems where specific aliphatic and aromatic hydrocarbon degrading *Burkholderia* cultures were added extraneously. The predominant cultures found in the aqueous phase of the un-spiked controls were isolated and identified. The large variability observed in the unspiked controls indicated heterogeneity in the distribution of these microorganisms in the sludge. The strains isolated from oily sludge were identified through 16S rRNA analysis. The predominant cultures were identified as *Microbacter* sp., *Bacillus* sp., *Pseudomonas aeruginosa*, *Acinetobacter baumannii* and *Stenotrophomonas* sp. Each of these strains exhibited good growth when 0.5% (w/v) of the oil extracted from oily sludge was provided as the sole source of carbon and energy (Fig. 1) and the end of log phase was reached within 7 days. Ongoing studies have revealed the remarkable ability of these cultures to utilize hydrocarbons across various groups as a sole substrate, including *n*-alkanes, cycloalkanes, and 2-, 3- and 4-ring PAHs (unpublished results). Although the degradation of hydrocarbons, crude oil and oily sludge by *Pseudomonas* sp., *Acinetobacter* sp., and *Bacillus* sp. is widely reported,^{7,14,24–27} no previous studies have reported the role of *Microbacter* sp. or *Stenotrophomonas* sp. in oily sludge degradation.

In this study, a reconstituted consortium comprised of these strains was used for bioaugmentation. The use of mixed indigenous microbial consortia may be advantageous due to possessing a broader metabolic capacity, synergistic effects and co-metabolic effects.^{7,21} These indigenous microorganisms are expected to be better acclimatized to the oily sludge and may exhibit better tolerance to co-contaminants¹¹ compared to other extraneous oil degrading cultures. Nitrate was supplemented as nitrate-nitrogen in the batches with nutrient supplementation since an excess of ammonium nitrogen is known to be toxic to some microorganisms. An excess of phosphates is also sometimes reported to be toxic to microorganisms, such that microbial degradation may be adversely affected in nutrient supplemented systems. The nonionic surfactant Triton X-100 was used in this study since nonionic surfactants are comparatively less toxic and Triton X-100 has been reported to enhance microbial degradation of oil.^{17,19,20}

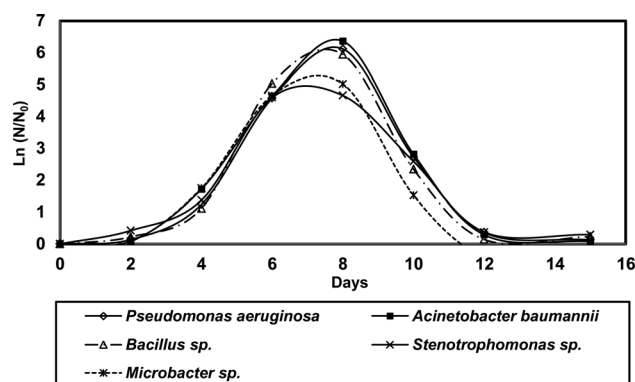


Fig. 1 Growth of pure cultures isolated from oily sludge on 0.5% (w/v) of extracted oil provided as the sole substrate in batch cultures.



The study was designed as per 2^3 factorial design. The conditions prevailing in each run/treatment are illustrated in Table 1. The oily sludge contained 9–10.5% oil on a dry weight basis. Significant variation was observed in the experiments due to the heterogeneous distribution of oil and microorganisms in the sludge, as illustrated by the SE values. TPH associated with the aqueous phase was consistently found to be very low ($0.35 \pm 0.17\%$), hence, % degradation was estimated solely based on TPH determined by soxhlet extraction of the sludge. This study revealed that bioaugmentation with microorganisms and biostimulation with nutrients and surfactants increased the extent of degradation of oil in oily sludge to $57 (\pm 9.3)\%$ over 30 days. In contrast, the individual treatments (*i.e.*, only the addition of microorganisms, only the addition of nutrients or only the addition of surfactants) yielded much lower degradation of oil. Oil degradation over 30 days was only $6.7 \pm 3.1\%$ in the controls where neither bioaugmentation nor biostimulation was performed. Interestingly, for studies conducted with the sludge within a year of sludge collection, TPH degradation in the uninoculated controls was much higher, *i.e.*, $44 \pm 10\%$.²³ Thus, prolonged storage at 4 °C decreased the activity of the indigenous cultures. With the addition of nutrients only, oil degradation with respect to the initial oil content was $32.4 \pm 9.7\%$, whereas the addition of only Triton X-100 increased oil degradation to $39.1 \pm 4.6\%$ over 30 days. Bioaugmentation with the microbial consortium alone caused $20.5 \pm 3.0\%$ oil degradation over 30 days. Further interpretation regarding the impact of using multiple strategies, *i.e.*, the simultaneous addition of nutrients and surfactants, and simultaneous bioaugmentation and biostimulation with nutrients were interpreted using factorial design and ANOVA concepts.

The influence of the various factors *i.e.*, bioaugmentation with indigenous cultures isolated from sludge (MO), biostimulation with nutrients (NP) and biostimulation with Triton X-100 (TX) could be determined for both sets of response variables, *i.e.*, oil degradation and culture growth in suspension over 30 days based on the 2^3 factorial design as illustrated in

Fig. 2a and b. The average oil degradation over 30 days across all the experimental runs was 31.3% and average culture growth in suspension ($\ln(N/N_0)$) was 1.02. All the main effects and interaction effects increased the oil degradation over 30 days except for the binary interactions between nutrient and surfactant addition and bioaugmentation and surfactant addition. Surprisingly, the main effect of bioaugmentation was to reduce culture growth in suspension. All the other main effects and binary and ternary interaction effects were positive when culture growth in suspension was used as the response variable.

After calculating the average values of the main effects, the effects of two way interactions and three way interactions were determined, and an attempt was first made to determine if the normal order score approach could highlight the significant effects. However, this approach failed to provide much insight on the significant main and interaction effects as illustrated in Fig. 3a and b. In the normal order score approach, the effects that are random and normally distributed fall on a straight line when effects are plotted against the normal order score. In contrast, significant effects that are not randomly distributed fall off the straight line joining the random effects.²² A straight line passing through the origin could not be fitted for the data illustrated in Fig. 3a and b. However, since each run was conducted in triplicate the significant and insignificant effects could be determined based on ANOVA. The ANOVA approach yields more insight since it reveals information contained in the replicates which is missing in the normal order score approach.

Multifactor ANOVA was performed using STATISTICA and the results are summarized in Table 2. The significance of main and interaction effects may be determined based on the *F*-statistic and *p*-value. Statistically significant main effects of NP and Triton X-100 addition were found for both the measured response variables at a 90% confidence level. However, the main effect of bioaugmentation was significant for TPH degradation but insignificant in the case of microbial growth in suspension.

The addition of nutrients had significant effects on the extent of degradation and also on microbial growth in the aqueous phase. Thus, the nutrients present in the baseline media cause a limitation in the bioremediation of oily sludge over the 30 day duration. Interestingly, most of the two way interactions other than that between bioaugmentation and nutrient addition were insignificant while the three way interaction between all of the three factors was significant. Thus, simultaneous addition of nutrients and surfactants along with bioaugmentation with indigenous cultures may offer a significant benefit in bioremediation of this refinery sludge. Thus, complex interactions affect oil biodegradation and culture growth in the aqueous phase in this oily sludge bioremediation scenario. Various other researchers have utilized the analysis of variance approach for demonstrating the impact of various treatments on the extent of biodegradation and microbial activity.^{3,11,12,21}

From the ANOVA results some variation is found between the significant and insignificant effects for the two response variables chosen, *i.e.*, oil degradation and culture growth in suspension. The main effect of bioaugmentation with

Table 1 Oil degradation and culture growth in suspension over 30 days for various treatments for the oily sludge biodegradation study as per 2^3 design^a

Run	Factors			Response variables			
	NP	TX	MO	% degradation		$\ln(N/N_0)$	
				Mean	SE	Mean	SE
1	–	–	–	6.7	3.1	0.40	0.28
2	+	–	–	32.4	9.7	1.28	0.32
3	–	+	–	39.1	4.6	1.61	0.37
4	+	+	–	27.8	8.8	1.33	0.39
5	–	–	+	20.5	3	0.22	0.32
6	+	–	+	38.2	8.9	0.61	0.31
7	–	+	+	28.0	4.4	0.23	0.06
8	+	+	+	57.8	9.3	2.48	0.49

^a NP: nutrients, TX: Triton X-100, MO: microorganisms, SE: standard error, *N*: viable count of microorganisms at day 30, *N*₀: viable count of microorganisms at day 0.



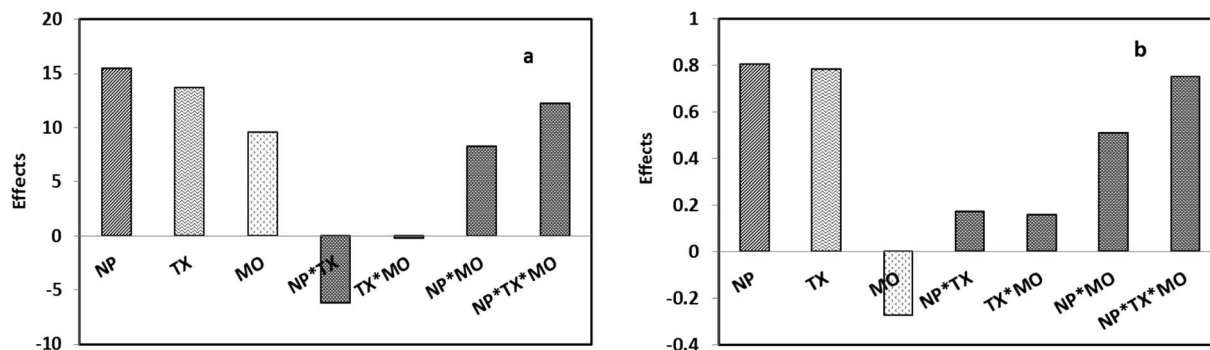


Fig. 2 Magnitude of main effects and binary and ternary interaction effects for the oily sludge biodegradation study as per 2^3 factorial design for (a) oil degradation and (b) culture growth in suspension as the response variable.

microorganisms was to increase oil degradation and this effect was found to be significant.

In contrast, the main effect of bioaugmentation on culture growth in suspension was to reduce culture growth. However this reduction was found to be insignificant based on the ANOVA results. The increase in oil degradation corresponding with a negligible increase in culture growth in suspension may be because of an increase in N_0 due to bioaugmentation causing nutrient deficiency. The enzymes produced by the microorganisms present at high concentrations could have promoted higher oil degradation. This may also be due to the different uptake mechanisms exhibited by the microbes responsible for oil degradation. In such sludges, oil mostly exists as sorbed oil rather than free phase oil. Microorganisms also associate with particulate matter in the sludge and some sorbed hydrocarbons may have been utilized through direct interfacial uptake. Thus, viable count in the aqueous phase is not a true measure of microbial activity in the system. Degradation of oil is a better measure of the activity of oil degraders in this system, although obtaining estimates of degradation is more laborious and time consuming. Biostimulation with nutrients and surfactants was found to cause an increase in the growth of microorganisms in the aqueous phase. Surfactants are known to facilitate micellar solubilization of oil and emulsification of oil and are thus likely

to cause the desorption of oil. Moreover, surfactants are known to alter microbial cell surface hydrophobicity^{19,20} such that the distribution of microorganisms in the system may be altered. The simultaneous addition of nutrients possibly promoted oil degradation by microorganisms suspended in the aqueous phase, thereby leading to higher culture growth in the aqueous phase.

Microbial activity in the aqueous phase led to a change in the color of the aqueous suspension after 30 days. This phenomenon may be indicative of oil/hydrocarbons leaching out over time or possibly due to the accumulation of intermediates formed during microbial degradation of oil. While the oil/hydrocarbon content in the aqueous suspension across the various treatments was negligible both initially and after 30 days, the TOC in the aqueous phase was significantly higher after 30 days (Fig. 4). An increase in TOC was also observed for the un-inoculated controls. The increase in TOC suggests the accumulation of soluble intermediates formed during the biodegradation of oil associated with the sludge. In most cases, the increase in TOC per unit decrease in TPH was found to vary from 4% to 12%, while it was found to be 30% for the un-inoculated controls. Thus, the bioaugmentation and biostimulation strategies used led to more complete biodegradation and comparatively lower accumulation of intermediates.

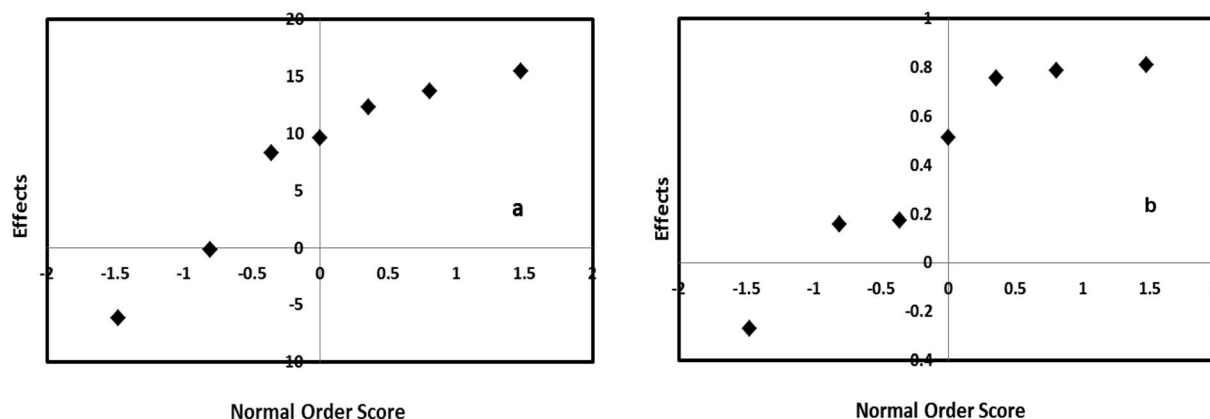


Fig. 3 Normal plot of effects for the 2^3 factorial design with (a) oil degradation and (b) culture growth in suspension over 30 days as response variables.



Table 2 Significant and insignificant effects at a 90% confidence level based on ANOVA^a

Response	Factors	Significant		Factors	Insignificant	
		F-statistics	p-value		F-statistics	p-value
% degradation	NP	10.869	0.005	NP × TX	1.677	0.214
	TX	9.896	0.006	TX × MO	0.012	0.914
	MO	4.227	0.056			
	NP × MO	3.225	0.091			
	NP × TX × MO	7.235	0.016			
ln(N/N ₀)	NP	10.192	0.006	MO	2.442	0.138
	TX	11.853	0.003	NP × TX	1.062	0.318
	NP × MO	4.156	0.058	TX × MO	0.369	0.552
	NP × TX × MO	11.133	0.004			

^a NP: nutrients, TX: Triton X-100, MO: microorganisms, N: viable count of microorganisms at day 30, N₀: viable count of microorganisms at day 0.

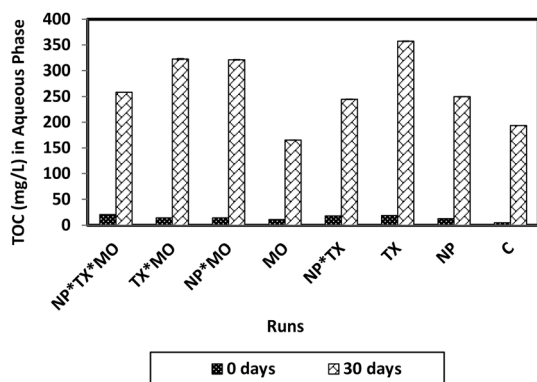


Fig. 4 Variation in TOC in the aqueous phase for the various treatments in the oily sludge biodegradation study as per 2³ design.

The increase in TOC per unit decrease in TPH was least (4%) when all three strategies (bioaugmentation, addition of nutrients and addition of surfactant) were employed simultaneously.

Various researchers have shown the beneficial effects of bioaugmentation on the degradation of oil associated with sludges and soil,^{1,15,28} as observed in this study. Bioaugmentation works best in scenarios where the indigenous population is very low.³ In many scenarios, bioaugmented microorganisms have been found to be unable to adapt to the conditions prevailing in the contaminated environment. Bento *et al.*¹² demonstrated how the presence of indigenous cultures in the soil limited the activity of bioaugmented microorganisms in a diesel contaminated soil system. They were unable to establish a correlation between TPH degradation, various treatments and the activity of diesel degrading microorganisms since the indigenous microbial population degraded diesel more efficiently than the microbial consortium introduced during bioaugmentation. Tahhan *et al.*²⁸ also demonstrated that indigenous microorganisms could degrade hydrocarbons when oily sludge was added to soil without bioaugmentation. However, successive bioaugmentation with the enriched indigenous consortium could improve the TPH removal rates. Alici *et al.*¹⁵ reported that bioaugmentation with tailored microbial consortia could facilitate bioremediation of soil

co-contaminated with diesel and heavy metals and up to 75% removal could be achieved in 42 days. Mariano *et al.*³ reported that various amendments could enhance TPH removal to 45.5% over a period of 55 days; however, bioaugmentation with non-indigenous cultures had no significant beneficial effect on TPH removal. Ayotamuno *et al.*²⁹ reported that bioaugmentation with extraneous microorganisms along with regular mixing and watering resulted in a 63.7–84.5% reduction in TPH over a duration of six weeks. In general, bioaugmentation is reported to be more successful in scenarios where the indigenous microorganisms native to the soil/sludge were enriched and added. Similar findings are revealed in our study. The low oil degradation (in run 1) due to depletion of the indigenous microorganisms originally present in the sludge during prolonged storage of the oily sludge could be partially overcome by bioaugmentation with indigenous microorganisms.

In the present study, addition of nutrients alone or in combination with bioaugmentation could enhance oil/TPH degradation significantly. The impact of nutrient addition on oil biodegradation is reported to vary widely and is possibly dependent on system specific conditions. Gallego *et al.*² reported significant enhancement, such that upon addition of inorganic nitrogen and phosphorous up to 90% degradation of diesel was observed under laboratory conditions. In 12 week long laboratory studies on degradation of petroleum sludge and contaminated soil, Gojic-Cvijovic *et al.*¹¹ demonstrated that the addition of nutrients (NPK) offered a greater beneficial effect compared to surfactant addition when culture growth in the aqueous phase was used as the response variable. Admon *et al.*³⁰ found that the degradation of oily sludge contaminated soil occurred only after application of nutrients in the ratio of C : N : P = 50 : 10 : 1. Liu *et al.*³¹ found that the addition of manure (as nutrients) to oily sludge significantly increased the microbial activity and diversity, TPH in the treated sludge decreased by 58.2% over 365 days of bioremediation in comparison to only 15.6% in the control plot. Machin-Ramirez *et al.*³² demonstrated that addition of commercial fertilizers enhanced the degradation of weathered oily sludge, with removal of 24% of TPH over a duration of 25 days. In contrast, Tahhan *et al.*²⁸ demonstrated the inhibition of oily sludge biodegradation upon addition of nutrients, possibly due to the



higher concentration of nitrogen and phosphorus already present in the sludge.

In addition to nutrients, TPH biodegradation is often limited due to the hydrophobicity and low aqueous solubility of the constituent hydrocarbons. Surfactants may help in increasing the bioavailability of sorbed oil through micellar solubilization and emulsification. However, the use of surfactants in bioremediation experiments has been reported to both stimulate as well as inhibit hydrocarbon degradation, influenced by various chemical properties of the surfactants and its interaction with microorganisms.^{17,19,20,33} In the present study, the main effect of the addition of surfactants was statistically significant both for microbial growth in suspension and oil degradation. The binary interaction of surfactant addition and nutrient addition and that of surfactant addition and bioaugmentation were insignificant for both the response variables chosen. The only significant binary interaction was that of nutrient addition and bioaugmentation with microorganisms, however it was only marginally significant ($0.1 > p\text{-value} > 0.05$). In contrast, the 3-way interaction between surfactant addition, nutrient addition and bioaugmentation was significant even at a 98% confidence level for both the measured response variables. Thus, the simultaneous addition of nutrients, surfactants and indigenous microorganisms had a synergistic effect on both oil degradation and culture growth.

Although synergism/antagonism has not been specifically explored by other researchers, several studies have reported enhanced TPH degradation when surfactant addition is combined with other strategies. Cameotra and Singh³⁴ reported that the effects of the addition of nutrients with bacterial consortia and crude biosurfactant with bacterial consortia resulted in less TPH degradation (91–95%) in comparison to when nutrients and surfactants were added together (98% TPH degradation). Rahman *et al.*²¹ reported that the addition of rhamnolipid biosurfactant increased the degradation of light *n*-alkanes in a scenario where soil mixed with oily sludge was treated. However, simultaneous supplementation with inorganic nutrients and rhamnolipid biosurfactant resulted in more complete degradation.

4. Conclusion

Addition of enriched indigenous microbes, addition of nutrients and addition of surfactant significantly enhanced oil degradation in sludge over and above that observed in the controls. Sludge biodegradation studies using 2³ factorial design with oil degradation and microbial growth over 30 days as response variables revealed that the main effect of nutrient addition and surfactant addition enhanced oil degradation and culture growth in suspension and these effects were significant at a 90% confidence level. However, the main effect of bioaugmentation only enhanced oil degradation but did not increase microbial growth in suspension. This may indicate the possible role of microorganisms attached on sludge solids in degrading sorbed oil. The binary interactions are found to be insignificant except for that between nutrient addition and bioaugmentation, which was found to have a synergistic effect.

No other studies have revealed such a high ternary interaction effect between surfactant addition, nutrient addition and bioaugmentation, indicating significant synergistic interactions among these strategies on oil degradation and microbial growth. Thus, the effect of the various factors is not solely additive. The characteristics of the sludge, the characteristics and composition of the oil in the sludge and the nature of the indigenous microorganisms present possibly affected the results.

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References

- 1 N. Vasudevan and P. Rajaram, Bioremediation of oily sludge contaminated soil, *Environ. Int.*, 2001, **26**, 409–411.
- 2 J. L. R. Gallego, J. Loreda, J. F. Ilasmas, F. Vazquez and J. Sanchez, Bioremediation of diesel contaminated soils: Evaluation of potential *in situ* techniques by study of bacterial degradation, *Biodegradation*, 2001, **12**, 325–335.
- 3 A. P. Mariano, A. P. de. A. G. Kataoka, D. de. F. de. Angelis and D. M. Bonotto, Laboratory study on the biodegradation of diesel oil contaminated soil from a petrol station, *Braz. J. Microbiol.*, 2007, **38**, 346–353.
- 4 M. D. Ferrari, E. Neirotti, C. Albornoz, M. R. Mostazo and M. Cozzo, Biotreatment of hydrocarbons from petroleum tank bottom sludges in soil slurries, *Biotechnol. Lett.*, 1996, **18**, 1241–1246.
- 5 M. S. Kuyukina, I. B. Ivshina, M. I. Ritchova, J. C. Philp, C. J. Cunningham and N. Christofi, Bioremediation of crude oil contaminated soil using slurry-phase biological treatment and landfarming techniques, *Soil Sediment Contam.*, 2003, **12**, 85–89.
- 6 I. V. Robles-Gonzalez, F. Fava and H. M. Poggi-Varaldo, A review on slurry bioreactors for bioremediation of soils and sediments, *Microb. Cell Fact.*, 2008, **7**(5), DOI: 10.1186/1475-2859-7-5.
- 7 S. Mishra, J. Jyot, K. R. Chander and B. Lal, In situ bioremediation potential of a oily sludge – degrading bacterial consortium, *Curr. Microbiol.*, 2001, **43**, 328–335.
- 8 S. E. Fantroussi and S. N. Agathos, Is bioaugmentation a feasible strategy for pollutant removal and site remediation?, *Curr. Opin. Microbiol.*, 2005, **8**, 268–275.
- 9 J. L. R. Gallego, M. J. Garcia-Martinez, J. F. Llamas, C. Bellosch, A. I. Pelaez and J. Sanchez, Biodegradation of oil tank bottom sludge using microbial consortia, *Biodegradation*, 2007, **18**, 269–281.
- 10 R. Hosokawa, M. Nagai, M. Morikawa and H. Okuyama, Autochthonous bioaugmentation and its possible application to oil spills, *World J. Microbiol. Biotechnol.*, 2009, **25**, 1519–1528.



- 11 G. D. Gojgic-Cvijovic, J. S. Milic, T. M. Solevic, V. P. Beskoski, M. V. Ilic, L. S. Djokic, T. M. Narancic and M. M. Vrvic, Biodegradation of petroleum sludge and petroleum polluted soil by a bacterial consortium: a laboratory study, *Biodegradation*, 2012, **23**, 1–14.
- 12 F. M. Bento, F. A. O. Camargo, B. C. Okeke and W. T. Frankenberger, Comparative bioremediation of soils contaminated with diesel oil by natural attenuation, biostimulation and bioaugmentation, *Bioresour. Technol.*, 2005, **96**, 1049–1055.
- 13 P. W. G. Liu, L. M. Whang, T. C. Chang, I. C. Tseng, P. T. Pan and S. S. Cheng, Verification of necessity for bioaugmentation lessons from two batch case studies for bioremediation of diesel contaminated soils, *J. Chem. Technol. Biotechnol.*, 2009, **84**, 808–819.
- 14 F. M. Ghazali, R. N. Z. A. Rahman, A. B. Salleh and M. Basri, Biodegradation of hydrocarbons in soil by microbial consortia, *Int. Biodeterior. Biodegrad.*, 2004, **54**, 61–67.
- 15 C. Alici, R. Musella, F. Tasso, C. Ubaldi, S. Manzo, C. Cremisini and A. R. Sprocati, Bioremediation of diesel oil in a co-contaminated soil by bioaugmentation with a microbial formula tailored with native strains selected for heavy metals resistance, *Sci. Total Environ.*, 2009, **407**, 3024–3032.
- 16 E. B. de. Moraes and S. M. Tauk-Tornisielo, Biodegradation of oil refinery residues using mixed culture of microorganisms isolated from a landfarming, *Braz. Arch. Biol. Technol.*, 2009, **52**, 1571–1578.
- 17 G. Mohanty and S. Mukherji, Effect of an emulsifying surfactant on diesel degradation by cultures exhibiting inducible cell surface hydrophobicity, *J. Chem. Technol. Biotechnol.*, 2007, **82**, 1004–1011.
- 18 C. C. Lai, Y. C. Huang, Y. H. Wei and J. S. Chang, Biosurfactant – enhanced removal of total petroleum hydrocarbons from contaminated soils, *J. Hazard. Mater.*, 2009, **167**, 609–614.
- 19 S. Mohanty and S. Mukherji, Alteration in cell surface properties of *Burkholderia* spp. during surfactant aided biodegradation of petroleum hydrocarbons, *Appl. Microbiol. Biotechnol.*, 2012, **94**, 193–204.
- 20 S. Mohanty and S. Mukherji, Surfactant aided biodegradation of NAPLs by *Burkholderia multivorans*: comparison between Triton X-100 and rhamnolipid JBR-515, *Colloids Surf., B: Biointerfaces*, 2013, **102**, 644–652.
- 21 K. S. M. Rahman, T. J. Rahman, Y. Kourkoutas, I. Petsas, R. Marchant and I. M. Banat, Enhanced bioremediation of *n*-alkanes in petroleum sludge using bacterial consortium amended with rhamnolipid and micronutrients, *Bioresour. Technol.*, 2003, **90**, 159–168.
- 22 P. M. Berthouex and L. C. Brown, *Statistics for Environmental Engineers*, 2nd edn, Lewis Publisher, Boca Raton, 2002, pp. 233–259.
- 23 J. Jasmine and S. Mukherji, Sustainable management of tank bottom sludge from refineries: role of bioaugmentation with *n*-alkane and naphthalene degrading *Burkholderia* cultures, *Int. J. Innovat. Sustain. Dev.*, 2013, **3**(1), 9–16.
- 24 N. Arvanitis, E. A. Katsifas, K. I. Chalkou, C. Meintanis and A. D. Karagouni, A refinery sludge deposition site: presence of *nahH* and *alkJ* genes and crude oil biodegradation ability of bacterial isolates, *Biotechnol. Lett.*, 2008, **30**, 2105–2110.
- 25 O. Ward, A. Singh and J. Van Hamme, Accelerated biodegradation of petroleum hydrocarbon waste, *J. Ind. Microbiol. Biotechnol.*, 2003, **30**, 260–270.
- 26 U. J. J. Ijah and S. P. Antai, Removal of Nigerian light crude oil in soil over a 12-month period, *Int. Biodeterior. Biodegrad.*, 2003, **51**, 93–99.
- 27 S. Verma, R. Bhargava and V. Pruthi, Oily sludge degradation by bacteria from Ankleshwar, India, *Int. Biodeterior. Biodegrad.*, 2006, **57**, 207–213.
- 28 R. A. Tahhan, T. G. Ammari, S. J. Goussous and H. I. Al-shdaifat, Enhancing the biodegradation of total petroleum hydrocarbons in oily sludge by a modified bioaugmentation strategy, *Int. Biodeterior. Biodegrad.*, 2011, **65**, 130–134.
- 29 M. J. Ayotamuno, R. N. Okparanma, E. K. Nweneka, S. O. T. Ogaji and S. D. Probert, Bio-remediation of a sludge containing hydrocarbons, *Appl. Energy*, 2007, **84**, 936–943.
- 30 S. Admon, M. Green and Y. Avnimelech, Biodegradation kinetics of hydrocarbons in soil during land treatment of oily sludge, *Biorem. J.*, 2001, **5**, 193–209.
- 31 W. Liu, Y. Luo, Y. Teng, Z. Li and L. Q. Ma, Bioremediation of oily sludge-contaminated soil by stimulating indigenous microbes, *Environ. Geochem. Health*, 2010, **32**, 23–29.
- 32 C. Machín-Ramírez, A. I. Okoh, D. Morales, K. Mayolo-Deloisa, R. Quintero and M. R. Trejo-Hernandez, Slurry-phase biodegradation of weathered oily sludge waste, *Chemosphere*, 2008, **70**, 737–744.
- 33 S. Mohanty, J. Jasmine and S. Mukherji, Practical considerations and challenges involved in surfactant enhanced bioremediation of Oil, *BioMed Res. Int.*, 2013, **2013**, 328608, DOI: 10.1155/2013/328608.
- 34 S. S. Cameotra and P. Singh, Bioremediation of oil sludge using crude biosurfactants, *Int. Biodeterior. Biodegrad.*, 2008, **62**, 274–280.

