

## Energy positive domestic wastewater treatment: the roles of anaerobic and phototrophic technologies†

Cite this: *Environ. Sci.: Processes Impacts*, 2014, **16**, 1204

B. D. Shoener, I. M. Bradley, R. D. Cusick and J. S. Guest\*

The negative energy balance of wastewater treatment could be reversed if anaerobic technologies were implemented for organic carbon oxidation and phototrophic technologies were utilized for nutrient recovery. To characterize the potential for energy positive wastewater treatment by anaerobic and phototrophic biotechnologies we performed a comprehensive literature review and analysis, focusing on energy production (as kJ per capita per day and as  $\text{kJ m}^{-3}$  of wastewater treated), energy consumption, and treatment efficacy. Anaerobic technologies included in this review were the anaerobic baffled reactor (ABR), anaerobic membrane bioreactor (AnMBR), anaerobic fluidized bed reactor (AFB), upflow anaerobic sludge blanket (UASB), anaerobic sequencing batch reactor (ASBR), microbial electrolysis cell (MEC), and microbial fuel cell (MFC). Phototrophic technologies included were the high rate algal pond (HRAP), photobioreactor (PBR), stirred tank reactor, waste stabilization pond (WSP), and algal turf scrubber (ATS). Average energy recovery efficiencies for anaerobic technologies ranged from 1.6% (MFC) to 47.5% (ABR). When including typical percent chemical oxygen demand (COD) removals by each technology, this range would equate to roughly 40–1200 kJ per capita per day or  $110\text{--}3300 \text{ kJ m}^{-3}$  of treated wastewater. The average bioenergy feedstock production by phototrophic technologies ranged from 1200–4700 kJ per capita per day or  $3400\text{--}13\,000 \text{ kJ m}^{-3}$  (exceeding anaerobic technologies and, at times, the energetic content of the influent organic carbon), with usable energy production dependent upon downstream conversion to fuels. Energy consumption analysis showed that energy positive anaerobic wastewater treatment by emerging technologies would require significant reductions of parasitic losses from mechanical mixing and gas sparging. Technology targets and critical barriers for energy-producing technologies are identified, and the role of integrated anaerobic and phototrophic bioprocesses in energy positive wastewater management is discussed.

Received 20th December 2013  
Accepted 18th March 2014

DOI: 10.1039/c3em00711a

rsc.li/process-impacts

### Environmental impact

Conventional wastewater treatment processes are energy-intensive and focus on removal of organics and nutrients rather than recovery. Re-envisioning wastewater as a renewable resource may enable energy positive treatment, creating economic incentives for increased access to sustainable sanitation in both developed and developing communities. To this end, anaerobic technologies can recover the chemical energy of organic carbon in wastewater as methane, hydrogen, and electricity. In a complementary fashion, phototrophic technologies could address a major limitation of anaerobic treatment (nutrient removal) while increasing the energetic potential of wastewater resources by leveraging nutrients for biomass growth and organic carbon storage. In this review we evaluate the strengths and weaknesses of these technologies and project a path toward energy positive wastewater treatment.

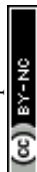
## 1 Introduction

The sanitation industry is facing a confluence of events that are straining utility budgets<sup>1,2</sup> and reducing their ability to provide reliable protection of public health and the aquatic environment.<sup>2</sup> Critical challenges include rapid and localized

population growth and decay;<sup>1,3-5</sup> aging infrastructure;<sup>6</sup> deterioration of surface waters resulting from excess nutrient (N and P) loading;<sup>7-10</sup> and a reliance on expensive, energy-intensive<sup>11,12</sup> treatment processes. These pressures are exacerbated by decreased resilience of ecosystems<sup>13-16</sup> and increased variability in renewable freshwater resources<sup>17-19</sup> resulting from climate change, with current energy-intensive approaches to wastewater treatment (consuming roughly  $0.3\text{--}0.6 \text{ kW h m}^{-3}$  of wastewater treated,<sup>20,21</sup> or 3% of U.S. electricity demand<sup>12</sup>) further contributing to climate change *via* greenhouse gas emissions from electricity production.<sup>22,23</sup> Ultimately, these stressors have

University of Illinois at Urbana-Champaign – Civil & Environmental Engineering, 205 N. Mathews Avenue 3221 Newmark Civil Engineering Lab, Urbana, Illinois 61801, USA. E-mail: jsguest@illinois.edu

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c3em00711a



intensified the need to address the water-energy nexus in wastewater management. Given that upgrades to U.S. infrastructure are expected to cost roughly \$300 billion over the next 20 years,<sup>6</sup> the industry has an unprecedented opportunity to re- envision wastewater streams as resource-rich sanitation media. In particular, treatment strategies enabling nutrient recovery as well as energy recovery and generation should be advanced, enabling resource positive sanitation – the management of wastewater as a renewable resource for nutrient recovery and net energy production<sup>24–26</sup> that can have a net benefit for the environment<sup>27</sup> – to gain traction at a broad scale.

In response to these challenges, a great deal of research has been conducted on alternative wastewater treatment technologies that recover or produce energy during wastewater treatment. Most notably, anaerobic technologies can recover usable energy from organic carbon (typically measured as chemical oxygen demand, COD), and phototrophic technologies can increase the chemical energy of a wastewater through CO<sub>2</sub>

fixation during growth and carbon storage. In addition to the production of bioenergy products such as methane, hydrogen, or electricity,<sup>28–31</sup> anaerobic processes are expected to be less energy intensive than aerobic processes due to a lack of aeration and a reduction in sludge wastage.<sup>32</sup> Although published studies have analyzed the performance of one or a small number of anaerobic system designs,<sup>28,33–37</sup> an in-depth comparison of technologies focusing on their potential in *domestic* wastewater management is still needed. The limited literature on anaerobic treatment of domestic wastewater is partially due to lower methane production, lower COD removal, and higher methane solubility, all of which stem from the fact that domestic wastewater is a relatively dilute resource stream.<sup>28</sup> Ultimately, it is unclear whether the conversion of organic carbon to usable energy will be adequate for energy positive treatment using existing and emerging technologies.<sup>20,38</sup>

Beyond COD, nutrient (nitrogen, N; phosphorus, P) limits are becoming increasingly common across the U.S.<sup>39,40</sup> In



*Brian Shoener is a Jack Kent Cooke Foundation Graduate Scholar and M.S. student in Environmental Engineering at the University of Illinois at Urbana-Champaign. He holds dual B.S. degrees in Civil Engineering and Management for Engineers from Bucknell University. Brian's current research focuses on the advancement of anaerobic membrane bioreactor technology via quantitative sustainable design. More broadly, his interests are in process modeling and improving energy recovery from methanogenic wastewater treatment processes.*

*via quantitative sustainable design. More broadly, his interests are in process modeling and improving energy recovery from methanogenic wastewater treatment processes.*



*Dr Cusick earned his B.S. in Environmental Engineering from the University of California, Riverside, holds an M.S. and Ph.D. in Environmental Engineering from the Pennsylvania State University and is now an Assistant Professor of Environmental Engineering at the University of Illinois at Urbana-Champaign. His research focuses the development of electrochemical technologies for nutrient and energy recovery from wastewater as well as natural and engineered salinity gradients.*

*technologies for nutrient and energy recovery from wastewater as well as natural and engineered salinity gradients.*



*Ian Bradley is a Deuchler Fellow and Ph.D. student in Environmental Engineering at the University of Illinois at Urbana-Champaign. He earned a B.S. degree in Industrial and Enterprise Systems Engineering and a M.S. degree in Civil and Environmental Engineering, both from the University of Illinois. Ian's current research focuses on algal growth for biofuel feedstock production in conjunction with wastewater treatment. He is interested in biological systems and process design for water and sanitation systems in the developed and developing world.*

*with wastewater treatment. He is interested in biological systems and process design for water and sanitation systems in the developed and developing world.*



*Dr Guest is currently an Assistant Professor in the Department of Civil and Environmental Engineering at the University of Illinois at Urbana-Champaign. He holds B.S. and M.S. degrees in Civil Engineering from Bucknell University and Virginia Polytechnic Institute and State University, respectively, and a Ph.D. in Environmental Engineering from the University of Michigan. Dr Guest's interests*

*center on biotechnology development for resource recovery from sanitation media, with particular emphasis on microalgae and methane-generating processes. His group works in developed and developing communities, and he currently serves as Thrust Leader for Sanitation and Resource Recovery for the Safe Global Water Institute.*



addition to high capital costs of wastewater treatment plant (WWTP) upgrades (e.g., \$3.36–3.96 billion for the for plants discharging to the Chesapeake Bay watershed<sup>41</sup>), the addition of biological nutrient removal (BNR) incurs higher operational costs that create or exacerbate financial challenges for utilities.<sup>41</sup> As an alternative to conventional BNR processes that leverage chemotrophic bacteria, phototrophic processes rely on light (typically sunlight) to promote growth and nutrient assimilation. As the phototrophs (including algae and cyanobacteria) take up inorganic carbon and grow, they also take up N and P from the wastewater and achieve nutrient recovery *via* assimilation. Studies have examined the potential for energy production using algae (e.g., ref. 42 and 43) and have even examined the potential for energy positive wastewater treatment,<sup>38</sup> but such studies have been limited to single cultivation technologies. To our knowledge there has not been a comprehensive comparative assessment of existing and emerging phototrophic technologies as tools to enable energy positive domestic wastewater management. In fact, studies focusing on bioenergy feedstock production with phototrophs have been largely disjointed from the wastewater literature, often using high strength synthetic media for cultivation (e.g., ref. 44 and 45). The prospect of using phototrophs for nutrient recovery and/or bioenergy feedstock production holds significant promise, however, and warrants further discussion.

As researchers attempt to balance the potential gains in net energy production with performance and economic trade-offs of each technology, the range of configurations for anaerobic and phototrophic systems continues to grow. To better understand the status and relative potential of each configuration, we undertook a critical review of the literature to characterize the demonstrated energy production by and critical barriers to a range of anaerobic and phototrophic technologies that have the potential to contribute to energy positive wastewater management. Based on available data, we quantified the typical performance of technologies in terms of treatment efficacy and bioenergy (or bioenergy feedstock) production, including the production of methane, hydrogen gas, electricity, biocrude oil, biodiesel, and heat. Seeking a deeper understanding of the potential energy balance of each technology, we also quantified usable energy yield (based on downstream conversion of bioenergy feedstocks) and anticipated energy consumption (based on experimental conditions in published studies). To be considered energy positive, a wastewater treatment scheme was required to produce energy in excess of the energy required to operate the process while also discharging water that meets regulatory standards. Given these constraints, we identify technologies with the greatest potential to enable energy positive carbon, nitrogen, and phosphorus management and present operational and performance targets for anaerobic and phototrophic treatment technologies to improve their net energy balance.

## 2 Anaerobic systems

Anaerobic processes for bioenergy production are most commonly leveraged for industrial wastewater treatment or for

solids management at domestic wastewater treatment facilities.<sup>32</sup> Limited application of these systems stems from a perceived need for high organic loading rates and mesophilic temperatures.<sup>31</sup> However, anaerobic technologies have been demonstrated at psychrophilic temperatures<sup>31,46,47</sup> and have the potential to be applied more broadly for low-strength wastewater treatment.<sup>25</sup> Anaerobic wastewater treatment processes can generally be categorized as suspended growth, sludge blanket, attached growth, membrane-based, or microbial electrochemical systems.<sup>32,48</sup> The first step of COD degradation in anaerobic treatment systems is the fermentation of complex organic matter into long chain volatile fatty acids, carbon dioxide, and hydrogen by acidogenic microorganisms. Long chain fatty acids are then further fragmented into acetic acid and hydrogen. Methane (CH<sub>4</sub>) and hydrogen gas (H<sub>2</sub>) are possible bioenergy products from anaerobic systems. In methane-producing reactors, acetoclastic methanogens ferment acetic acid to methane and carbon dioxide and hydrogenotrophic methanogens convert hydrogen and carbon dioxide to methane.<sup>49</sup> In microbial electrochemical technologies, exoelectrogenic bacteria oxidize acetate to carbon dioxide and produce electrical current transferring electrons to a conductive surface.<sup>50</sup> The anaerobic systems considered in this review are described below.

### 2.1 Suspended growth processes

Suspended growth processes are characterized by complete-mix conditions to prevent biomass from settling and to facilitate contact between the microorganisms and the wastewater. The most common processes include anaerobic sequencing batch reactors (ASBRs), completely-mixed anaerobic digesters, and the anaerobic contact process.<sup>32</sup> Of these, only ASBR had adequate peer-reviewed data (*i.e.*, >5 papers) on the treatment of domestic wastewater.

**2.1.1 Anaerobic sequencing batch reactor (ASBR).** The ASBR progresses through four stages similar to the aerobic sequencing batch reactor: settle, decant, feed, and react.<sup>51</sup> ASBRs often have higher solids residence times (SRTs) compared to continuous flow processes and enable more precise operational parameter (e.g., hydraulic retention time, HRT) control.<sup>52</sup> However, their suitability for the treatment of low-strength wastewaters has been questioned due to low gas production on dilute streams, although intermittent mixing has been suggested to improve gas-liquid separation and to enhance sludge settling.<sup>53</sup>

### 2.2 Sludge blanket processes

Successful operation of anaerobic sludge blanket processes relies on the aggregation of organisms into diversely populated granules capable of settling.<sup>54</sup> The granules form naturally from reactor operation and consist of a mixed population of bacteria and archaea that are able to carry out the overall fermentation and gas production from organic carbon substrates.<sup>55</sup> The gas bubbles produced from methanogenesis help to fluidize the granules, enhancing mass transfer without mixing.<sup>55</sup> Technologies with adequate (>5) peer-reviewed studies included the



upflow anaerobic sludge blanket (UASB) and the anaerobic baffled reactor (ABR).

**2.2.1 Upflow anaerobic sludge blanket (UASB).** In a UASB reactor, wastewater enters the reactor and is distributed across the bottom, traveling upward through the sludge blanket.<sup>56</sup> Granular sludge in the reactor allows for high volumetric COD loadings (as compared to other wastewater treatment technologies).<sup>32</sup> To enable better solids capture and to prevent loss of granules, modifications to the basic design have added packing material or a settling tank.<sup>32</sup> UASBs are advantageous because of their simple construction, scalability, and small footprint, though downstream processing is usually necessary to reduce effluent particulate organics.<sup>36</sup>

**2.2.2 Anaerobic baffled reactor (ABR).** The ABR utilizes a sequence of baffles to impede the flow of wastewater as it passes through the reactor.<sup>57</sup> Flow patterns and gas production force sludge in the reactor to rise and settle slowly.<sup>32</sup> Since its inception, the ABR has undergone several modifications in an effort to improve performance, such as changes to baffle design, including a settler in the system, or achieving solids capture using packing material. Advantages of this process include: simplicity of construction and operation, prolonged retention of influent solids, staged operation, and insensitivity to shock loads.<sup>32</sup> Disadvantages include having to construct shallow reactors to accommodate gas and liquid upflow velocities as well as difficulty with distributing the influent flow evenly.<sup>33</sup>

### 2.3 Attached growth processes

Attached growth anaerobic technologies rely on packing material in the reactor to provide surfaces for biofilm formation. The primary characteristics that differentiate reactors within this category are the packing material type and degree of bed expansion.<sup>32</sup> For example, the packed bed and fluidized bed configurations are operated at increasing upflow velocities, with fluidized bed being the highest. Because of the similarity of packed beds to the UASB and the availability of data for the fluidized system, only the anaerobic fluidized bed (AFB) was included.

**2.3.1 Anaerobic fluidized bed (AFB).** AFB reactors are operated at high upflow velocities in order to suspend particulate media such as granular activated carbon (GAC) in the reactor,<sup>20</sup> with wastewater treatment achieved by biofilms attached to the media. While AFBs are particularly effective for low strength wastewaters, the main shortcoming is minimal solids capture.<sup>32</sup> AFBs are therefore more appropriate for wastewater streams with primarily soluble COD.

### 2.4 Membrane-based processes

Membranes have been used in water treatment for over half a century, and are becoming increasingly common in applications ranging from wastewater treatment to desalination.<sup>58</sup> Microfiltration and ultrafiltration membranes are primarily used for particulate removal and can be arranged as flat sheets or hollow fibers.<sup>58</sup> One of the main benefits of using membranes in biological treatment processes is the completely independent control of SRT and HRT; SRT values have been reported as high

as 300 days, where biomass was only removed from the system during sampling.<sup>47</sup>

**2.4.1 Anaerobic membrane bioreactor (AnMBR).** An AnMBR is an anaerobic reactor coupled with membrane filtration.<sup>28</sup> The membrane can be configured as external cross-flow, internal submerged, or external submerged.<sup>34</sup> Inclusion of a membrane allows for robust solids capture while also improving effluent quality over other mainstream anaerobic processes.<sup>32</sup> This increase in quality comes about because of the decoupling of SRT from HRT. Higher SRTs correlate to greater volatile fatty acid (VFA) and soluble COD removals.<sup>32</sup> Additionally, AnMBRs allow for a much smaller footprint by enabling higher solids concentrations in the reactor.

### 2.5 Microbial electrochemical technologies (METs)

METs (also referred to as bioelectrochemical systems, BES) leverage microorganisms capable of extracellular electron transfer<sup>29,59</sup> to produce electrical energy from wastewater. Like all electrochemical technologies, such as fuel cells and batteries, METs are composed of an anode, where electrons for current are generated, and a cathode, where electrons are consumed. In METs, anaerobic bacteria naturally present in most wastewaters oxidize biodegradable organic matter and continuously transfer electrons to the anode.<sup>60</sup> Electrons flow from the anode, through an external circuit, to a cathode, where electrical current is consumed in a reduction reaction.<sup>29</sup> Current production in METs is dependent on the redox potential difference between organic matter oxidation at the anode ( $E_o = -0.32$  V) and current consumption at the cathode.<sup>61</sup> If the anode is more negative than the cathode, as in the case with oxygen reduction ( $E_o = 0.82$  V) in a microbial fuel cell (MFC), electrical current production is spontaneous. If the cathode reaction occurs at a redox potential that is more negative than the anode, such as hydrogen production ( $E_o = -0.414$  V) in a microbial electrolysis cell (MEC), then additional cell voltage must be applied to drive current production in the cell.<sup>29,62</sup> Although a multitude of cathodic reactions (*e.g.*, caustic production and hydrocarbon electro-synthesis<sup>48</sup>) have been paired with anodic oxidation of organic matter, the review will only focus on electricity and hydrogen production in METs.

**2.5.1 Microbial fuel cell (MFC).** The most commonly investigated MFC architecture is a single chamber reactor in which both the bio-anode and oxygen reduction cathode operate in the same solution.<sup>60</sup> The cathode electrode, which acts as the barrier between reactor solution and air, is coated with hydrophobic diffusion layers to allow oxygen transport but prevent water loss.<sup>63</sup> Although a variety of wastewaters have been evaluated for electricity generation,<sup>64</sup> power production has been significantly lower ( $<0.5$  W m<sup>-2</sup> cathode area) than reactors fed synthetic and well buffered solutions (1.0–4.3 W m<sup>-2</sup>) due to low solution conductivity as well the dilute concentrations and complex nature of organic substrates in domestic wastewater.<sup>48,65–67</sup> Additionally, cathodic materials are often expensive due to the need of precious metals (*e.g.*, platinum).<sup>68,69</sup>



**2.5.2 Microbial electrolysis cell (MEC).** MECs produce hydrogen from substrate by coupling a hydrogen evolution electrode to the bio-anode.<sup>30,70–72</sup> Hydrogen is a promising fuel for meeting future energy demand because it only produces water when combusted or oxidized in a fuel cell and has a high energy yield ( $142.35 \text{ kJ g}^{-1}$ ).<sup>73</sup> Since MEC current production is not spontaneous, voltage must be applied to produce hydrogen (0.6–1.2 V in practice<sup>62</sup>). Due to cathode catalyst, electrolyte, and substrate deficiencies, energy consumed by applying voltage can exceed the energy recovered as hydrogen gas.<sup>73,74</sup> Also, to prevent hydrogen losses due to hydrogenotrophic methanogenesis that occur in single chamber architecture,<sup>75,76</sup> a membrane or gas diffusion electrode is required to separate anode and cathode.<sup>74,74</sup>

### 3 Phototrophic systems

Simple, passive phototrophic processes (cultivating algae and/or phototrophic bacteria) such as open ponds are commonly used to treat municipal and agricultural wastewaters.<sup>77</sup> To date, the objective for these technologies tends to be nutrient (and often COD) removal from wastewater, rather than nutrient recovery or bioenergy feedstock production. Alternatively, more capital-intensive systems such as photobioreactors have been studied for phototroph cultivation, but this work has most often focused on bioenergy feedstock cultivation rather than wastewater treatment (e.g., ref. 78 and 79). Both types of systems predominantly operate with suspended cultures in open (e.g., ponds<sup>80</sup>) or closed systems (e.g., photobioreactors<sup>81</sup>) that allow for sunlight penetration and nutrient assimilation to promote growth and carbon storage before biomass is harvested.<sup>82</sup> Alternative systems consist of attached or immobilized phototrophs for easier harvesting.<sup>83</sup> Ultimately, the energetic benefit of phototrophic systems stems from the fact that they can increase the energetic content of wastewater through the conversion of light energy to chemical energy (as organic carbon). In order to evaluate the relative potential of phototrophic technologies in achieving energy positive municipal wastewater treatment, only published studies using actual wastewater as the growth medium have been included in the analysis.

#### 3.1 Suspended systems

Conventional phototrophic systems consist of suspended cultures that are operated in either continuous, batch, or semi-batch mode.<sup>84,85</sup> The most common large-scale phototroph cultivation systems are waste stabilization ponds (WSPs),<sup>77</sup> high rate algal ponds (HRAPs),<sup>80</sup> stirred tank reactors,<sup>86</sup> and tubular photobioreactors (PBRs).<sup>81,87</sup> At laboratory-scale, a wider variety of reactor configurations have been evaluated, including flat panel (a.k.a., flat plate) and annular PBRs,<sup>78</sup> as well as more basic well-mixed systems that are simply lit from overhead (these studies were classified as “Stirred Tank Reactors” for this review).<sup>88</sup>

**3.1.1 High rate algal pond (HRAP).** While open raceway ponds are used commercially for the production of algal

biofuels and health products,<sup>89</sup> a subset of published studies use HRAPs for wastewater treatment (e.g., ref. 80 and 90). HRAPs are open raceway ponds first proposed in the 1950s with the goals of providing improved wastewater treatment over traditional WSPs and algal biomass for potential biofuel applications.<sup>91</sup> Although they have the potential to be a more cost effective solution than PBRs for wastewater treatment,<sup>92,93</sup> HRAPs have relatively low biomass productivity (and thus require larger land areas) as compared to reactor-based technologies.<sup>81</sup>

**3.1.2 Photobioreactor (PBR).** Another widely used technology for cultivating algal biomass is the PBR.<sup>78,94</sup> These closed array systems allow for high biomass productivity as well as axenic growth conditions for monoculture maintenance.<sup>95</sup> Although a range of configurations have been evaluated at the lab-scale,<sup>78,94</sup> larger systems tend to be tubular PBRs due to economies of scale. There are relatively few studies that examine PBRs in conjunction with wastewater treatment, largely because of high costs compared to other treatment technologies<sup>94</sup> and because axenic cultures are generally not targeted for municipal wastewater treatment. Most PBR studies focus on pure species with high lipid productivities and, consequently, higher energy potential and revenue generation.<sup>95–97</sup>

**3.1.3 Stirred tank reactor.** There is extensive literature on phototrophic growth in stirred tank reactors (open, completely mixed reactors lit from overhead). Although published studies using stirred tank reactors cover a range of operational conditions (including various lighting schemes, batch *vs.* continuous *vs.* semi-continuous operation, *etc.*) and a subset have been performed at the pilot-scale<sup>38</sup> the majority of these studies have been at the laboratory-scale (e.g., ref. 98–100). In order to look for general trends in performance of stirred tank reactors, data from these studies have been aggregated to identify performance trends and enable comparisons to larger-scale, more broadly applied technologies (e.g., HRAPs). Any insights gained may be applied to the design of larger-scale batch or sequencing batch reactors for both wastewater treatment and algal biomass production.

**3.1.4 Waste stabilization pond (WSP).** WSPs are the most widely used phototrophic treatment technology.<sup>101</sup> In the U.S. alone there are >7000 WSPs in use, which accounts for over one-third of all centralized treatment systems.<sup>77</sup> During the day, phototrophs in these systems produce dissolved oxygen, which facilitates COD degradation by aerobic heterotrophs<sup>102</sup> and promotes photo-oxidative damage for pathogen removal.<sup>103–105</sup> Although WSPs are often a cost effective solution for wastewater management utilities,<sup>106</sup> they are used almost exclusively in rural areas due to large land requirements.<sup>107</sup> With the exception of early visionary proposals linking wastewater to bioenergy with algae,<sup>91,108–110</sup> WSP literature has focused almost exclusively on wastewater treatment (removal of COD, N, P, heavy metals<sup>111,112</sup>) with little discussion of biomass production or potential biofuel applications. Despite limited literature linking WSPs to bioenergy feedstock cultivation, this technology represents one of the easiest opportunities to transition from an existing energy *neutral/consuming* technology to an energy *producing* process given that algal biomass is already generated.



### 3.2 Attached growth systems

The cost of biomass harvesting (including flocculation, centrifugation, and sedimentation<sup>82</sup>) remains a key barrier to the broad implementation of suspended growth algal systems.<sup>88</sup> Although sedimentation is often the most inexpensive approach, it achieves low (50–90% (ref. 80 and 113)) biomass recoveries and is typically used when low value biomass is being removed from the system.<sup>114</sup> Technologies that seek to achieve high percentages (>95%) of suspended biomass recovery for use as biofuel feedstock would add significantly to the cost of operation.<sup>80,115,116</sup> As an alternative to suspended growth, attached growth systems restrict algal growth to physical structures resulting in aggregated biomass that either sloughs off the structures or can be removed through cleaning.<sup>117</sup> While there are a number of different attached growth systems available (e.g., Algaewheel™ and other industrial solutions<sup>118</sup> as well as various immobilized gel matrices<sup>119</sup>), the data necessary to perform the energetic analysis for most attached growth technologies was lacking. One exception was the algal turf scrubber (ATS), which has been the focus of a number of studies and which reported adequate data for its inclusion in this study.<sup>120,121</sup>

**3.2.1 Algal turf scrubber (ATS).** ATSS consist of long, inclined beds typically constructed of landfill liner that support mixed community biofilms that include cyanobacteria, filamentous periphyton, and epiphytic diatoms.<sup>117,122,123</sup> As water flows down the beds into a concrete sump, nutrients are taken up by the biofilm, supporting microbial activity and reducing the concentrations of nutrients in the effluent.<sup>124</sup> When biomass accumulates, harvesting is often performed by machinery (such as a loader) driven across the bed.<sup>117</sup> Although it is not a common process, there are several private companies operating ATSS on a large scale, notably, Aquafiber Technologies® (7.5 million gallons per day [MGD]) and HydroMentia® (capacity 30 MGD).<sup>118</sup>

## 4 Data analysis

### 4.1 Criteria for inclusion in this study

A comprehensive literature review was conducted on the technologies listed above, with a focus on studies demonstrating treatment of municipal-strength wastewaters (COD <500 g COD per m<sup>3</sup>). For phototrophic systems, comparison studies have found that synthetic wastewater, though displaying comparable nutrient removal rates, generates more biomass than wastewater-based studies<sup>85</sup> and was thus excluded from this review. For methane producing systems, studies using synthetic wastewater with relevant COD concentrations were included because differences in performance (between synthetic and real wastewater) were not readily observed. For METs, all studies with influent COD <500 g COD per m<sup>3</sup> used real wastewaters (i.e., all studies that used synthetic wastewater had COD values above 500 g COD per m<sup>3</sup> and were thus excluded). Once relevant studies were identified, many were excluded from further analysis due to insufficient data that prevented the calculation of energy production normalized to contaminant removal. If a

required value was not explicitly stated but prerequisite values were given, the unknown values were calculated (see Fig. S1 and S2 for inclusion/exclusion decision-making, ESI†). Ultimately, these data were used to report the effluent COD and energy (as kilojoules, kJ) recovered by anaerobic treatment as well as effluent N/P and energy produced by phototrophic technologies (Fig. 1).

### 4.2 Energetic analysis

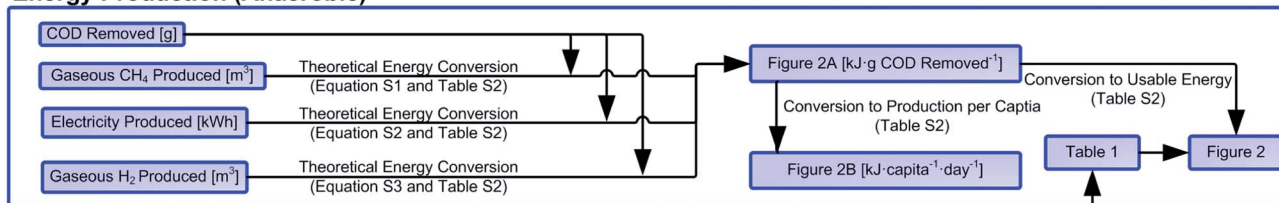
**4.2.1 Anaerobic technologies.** Anaerobic technologies recovered energy either in the form of methane (mol CH<sub>4</sub> per g COD), hydrogen gas (mol H<sub>2</sub> per g COD), or electricity (kJ per g COD). In order to compare the data objectively, each was normalized to kJ recovered per g COD removed by converting each energy source to kJ using standard conversion factors based on energetic content: 803 kJ per mol CH<sub>4</sub>,<sup>32</sup> 286 kJ per mol H<sub>2</sub>,<sup>125</sup> and by converting electricity (reported in kW h) to kJ by multiplying by 3600 s h<sup>-1</sup> (eqn (S1)–(S3)†). Results for each technology were compared on the basis of per capita and per m<sup>3</sup> of wastewater treated using the conversions discussed in Section 4.3.

**4.2.2 Phototrophic technologies.** Phototrophic technology data were compiled from articles that reported both biomass generated and nutrient (N and/or P) removal. Biomass was either reported as total, maximum, or average VSS (g m<sup>-3</sup>), as productivity (g per m<sup>3</sup> per day), or as aerial productivity (g per m<sup>2</sup> per day). The SRT, experiment length, and reactor volume were leveraged to convert all numbers to an average daily productivity (g per m<sup>3</sup> per day). Biomass productivities were then normalized by the average nutrient removal per day (g per m<sup>3</sup> per day) to achieve g biomass produced per g nutrient removed from the treatment system. To convert biomass productivity to energetic potential, reported VSS were converted to units of COD (see Table S2, ESI†). Two scenarios were considered using macromolecule content (lipids/carbohydrates/proteins, L/C/P) within typical ranges from the literature:<sup>126–128</sup> a low COD/VSS ratio of 1.47 g COD per g VSS (assuming 10/40/50% lipids/carbohydrates/proteins, L/C/P<sup>86,98,126–128</sup>) and a high COD/VSS ratio of 1.84 g COD per g VSS (assuming a ratio of 30/20/50% L/C/P<sup>86,126–128</sup>). COD calculations were performed assuming lipids could be represented as stearic acid (C<sub>18</sub>H<sub>36</sub>O<sub>2</sub>), carbohydrates as glucose (C<sub>6</sub>H<sub>12</sub>O<sub>6</sub>), and proteins as C<sub>16</sub>H<sub>24</sub>O<sub>5</sub>N<sub>4</sub>.<sup>129</sup> Although higher COD/VSS ratios would be possible if higher lipid content were achieved (e.g., 70% lipids<sup>81</sup>), the ratios used here represent a reasonable range of expected compositions<sup>86,98</sup> to avoid overly optimistic ratios that would artificially increase calculated energy yield. Although it has been reported that some species can obtain greater than 80% lipids by dry biomass weight,<sup>130,131</sup> mixed algal wastewater cultures routinely see far less lipid accumulation, with an average around 10%.<sup>86,98</sup> Once biomass productivities were converted to COD, the energetic potential of the biomass was then calculated using a theoretical value of 13.9 kJ per g COD.<sup>132</sup> Results for each technology were compared on a per capita basis as described in Section 4.3.

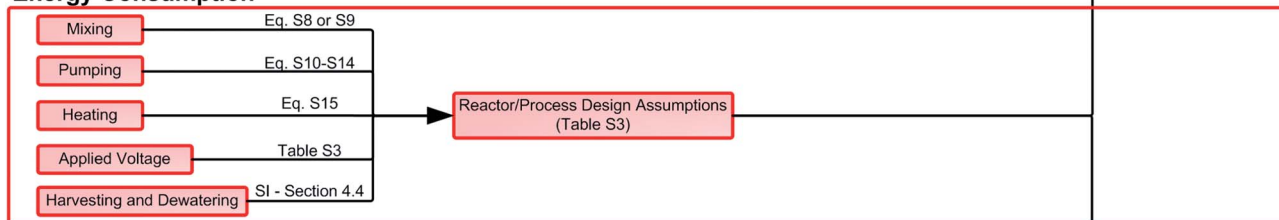
**4.2.3 Conversion to usable energy.** Although the energetic content of treatment system products may provide insight into



## Energy Production (Anaerobic)



## Energy Consumption



## Energy Production (Phototrophic)

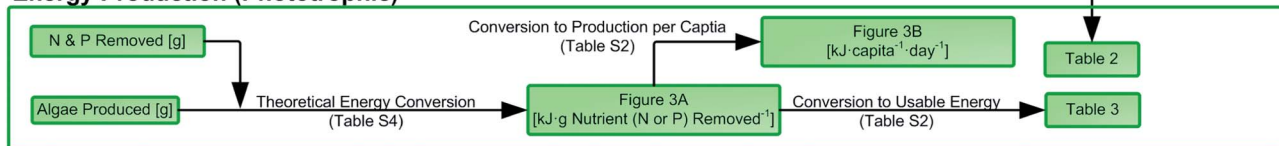


Fig. 1 Flow chart showing the process of data acquisition and analysis used in the manuscript along with conversion factors and their location in the ESI.†

the fundamental limitations of a given technology, the question regarding the feasibility of energy positive treatment can only be answered by determining the *usable* energy (e.g., electricity, heat, liquid fuel) provided by each treatment system. For anaerobic systems, the outputs include methane, hydrogen, and electricity. Given that the predominant form of energy consumed by treatment plants is electricity, methane and hydrogen were converted to electricity in a fuel cell at a 42.3% conversion efficiency.<sup>133</sup>

In order to predict the production of usable energy from phototrophic biomass, the energy yield from four different conversion processes – hydrothermal liquefaction (HTL), transesterification, anaerobic digestion, and combustion – were also calculated. Although anaerobic digestion has a long history in the conversion of algal biomass to methane,<sup>108,109,134</sup> direct combustion of algal biomass has been proposed as more energetically favorable than converting biomass to any bio-fuel.<sup>38,135</sup> For the conversion of phototrophic biomass into liquid fuels, both transesterification and HTL were considered, with HTL representing an emerging process of interest to the algae-to-biodiesel community.<sup>136,137</sup> HTL has been applied to wastewater-grown biomass (e.g., ref. 138 and 139), although energy balances have identified biomass harvesting and dewatering as key barriers to achieving energy positive systems.<sup>38</sup> The list of assumptions and values used for these calculations can be found in Table S2 (ESI).†

**4.2.4 Energy consumption.** An estimation of energy consumption for each technology was included in order to evaluate the feasibility of net energy positive wastewater treatment. However, the published studies analyzed did not include energy consumption data with the exception of Sturm and Lamer 2011.<sup>38</sup> In order to quantify energy consumption of each

process, the energy demand from various activities (e.g., pumping, mechanical mixing, gas sparging, etc.)<sup>32,140</sup> was estimated using standard design equations and the published range of design and operational parameters (see ESI Section S4† for a detailed explanation).

### 4.3 Unit conversions and efficiency calculations

Data were normalized and reported in one of four ways: as energy per gram of pollutant removed, energy per capita, energy per cubic meter of wastewater treated, and as a percent of energetic potential recovered. Energetic data normalized to pollutant removal (kJ per gCOD, g N, or g P) was calculated directly from the published data sets included in the review. These data (in units of kJ per g pollutant removed) were then normalized to per capita values by multiplying (i) by the average percent removal of that pollutant by a given technology, and (ii) by the average daily per capita production of that pollutant (180 g COD; 13 g N; 2.1 g P<sup>141</sup>). Next, energy productions were also reported per cubic meter of wastewater treated by assuming a wastewater production rate of 0.36 m<sup>3</sup> per person per day resulting in a wastewater composition of 500 g COD per m<sup>3</sup>, 36 g N per m<sup>3</sup>, 5.8 g P per m<sup>3</sup>. For efficiency calculations (e.g., percent of chemical energy recovered), COD was assumed to contain roughly 13.9 kJ per g COD,<sup>132</sup> resulting in an influent energetic content of 7000 kJ m<sup>-3</sup>. This conversion factor is lower than more recent values reported in the literature (17.7–28.7 kJ per g COD<sup>142</sup>), but was used throughout the manuscript to provide a consistent framework for energy conversions. All energy values (in units of kJ) represent the energetic content of produced fuel (methane, hydrogen, or electricity) for anaerobic systems or produced biomass for phototrophic systems, unless otherwise



noted (fuels are converted to electricity; biomass is converted to heat, methane, biodiesel, and biocrude oil).

## 5 Results and discussion

In the review of the peer-reviewed literature a total of 225 anaerobic and 86 phototrophic papers were screened and assessed according to the inclusion criteria (Fig. S1 and S2†). Of the papers reporting on anaerobic technologies, only 32 met the necessary criteria for energetic and treatment analysis with a total of 122 experimental data sets (*i.e.*, if the study reported multiple experimental conditions or replicates, all that met the inclusion criteria were included in this review). Published data on phototrophic technologies were less complete, with only 23 papers meeting the necessary criteria for treatment analysis with a total of 33 and 58 data sets for N and P removal, respectively. Of these papers, 13 had the necessary biomass productivity for energy analysis, resulting in 21 and 25 experimental data sets for energy production per g nutrient (N or P) removed (across 37 independent data sets). Furthermore, 9 of these 37 datasets were excluded because they reported greater than 50 g or 225 g of algal biomass grown per g N or g P removed, respectively, which was deemed to be outside the likely range of feasible biochemical compositions. Finally, WSPs were excluded from the energetic analysis due to a lack of biomass productivity data.

### 5.1 Energetic analysis

The energetic analysis began by determining fuel (anaerobic) or bioenergy feedstock (phototrophic) production from each study and the associated caloric content (Section 5.1.1). Energy consumption (Section 5.1.2) of each technology was then estimated based on experimental conditions in published studies and on additional assumptions detailed in Section S4 of the ESI.† An energy balance between consumption and production was then detailed for anaerobic systems to estimate net energy given typical experimental conditions in order to identify key barriers to energy positive treatment (Section 5.2). An energy balance was excluded for phototrophic technologies because of the uncertainty associated with downstream conversion to usable fuels, but available data were leveraged to set targets for cultivation and downstream fuel conversion processes (discussed in Section 5.3). Lastly, we examined the dichotomy between emerging (energy production) and traditional (effluent quality) objectives for treatment technologies (Section 6).

**5.1.1 Energy yield.** The average energy recovery by anaerobic systems ranged from 0.48 kJ per g COD (MFC) to 7.3 kJ per g COD (ABR) and was highest for gas producing technologies (Fig. 2a). The average percent energy recovery (as methane, hydrogen, or electricity) from degraded COD by each technology was as follows (from greatest to least; average  $\pm$  standard deviation): ABR ( $47.5 \pm 4.5\%$ ), AnMBR ( $35.4 \pm 26.8\%$ ), AFB ( $33.8 \pm 12.9\%$ ), UASB ( $24.0 \pm 11.4\%$ ), ASBR ( $17.7 \pm 10.1\%$ ), MEC ( $14.3 \pm 14.4\%$ ) and MFC ( $1.6 \pm 1.4\%$ ). When including typical percent COD removals for each technology, this range would equate to roughly 40–1200 kJ per capita per day (Fig. 2b)

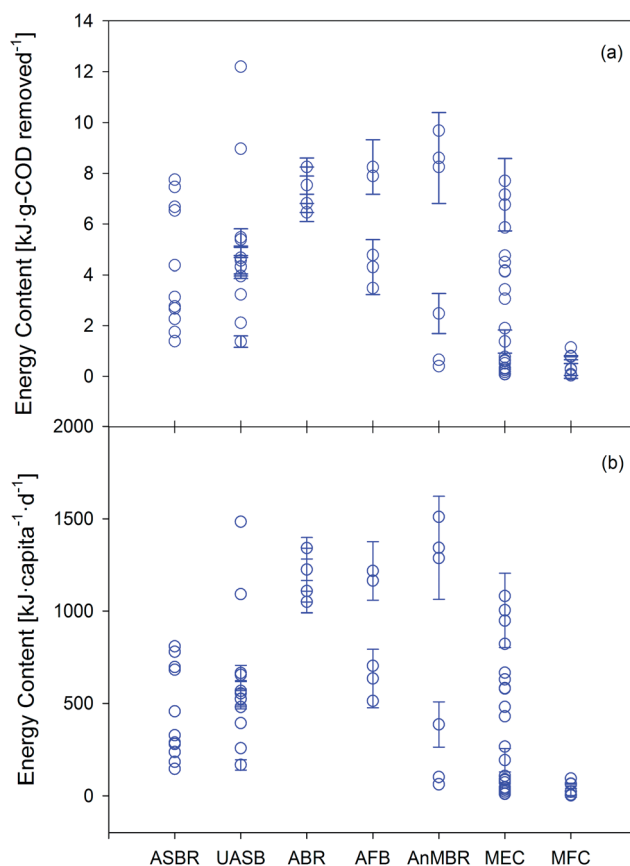


Fig. 2 (a) Energy content [kJ fuel per g COD removed] for each paper studying anaerobic technologies with an influent COD below  $500 \text{ g m}^{-3}$  (for synthetic wastewater) or using actual domestic wastewater. (b) Energy content [kJ fuel per capita per day] determined by multiplying values from (a) by  $180 \text{ g COD per capita per day}$  and by the respective average percent COD removals for each technology (Table 1). All energy products (methane, hydrogen, electricity) are reported as kJ using theoretical unit conversions (see ESI†). Individual points represent distinct experimental data sets, with error bars extending to  $\pm$  standard deviation (if reported).

or  $110\text{--}3300 \text{ kJ m}^{-3}$  of treated wastewater. After conversion of gases to usable electricity in a fuel cell (42.3% efficient), these values represent recoveries of roughly 2–20%. UASBs and AnMBRs had the two highest reported energy recovery data sets (12.2 and 9.7 kJ per g COD degraded, respectively), but AnMBRs also had the greatest variability (standard deviation of 4.3 kJ per g COD). The energy recovery by MECs was statistically different from most of the methane-producing technologies ( $p$ -values  $< 0.024$ ,  $\alpha = 0.05$ ; two-tailed, unpaired  $t$ -test) except ASBRs ( $p$ -value = 0.077), which could not be shown to be statistically different. MFCs did, however, exhibit significantly lower energy production ( $p$ -value = 0.048) with average per capita energy recovery 5–15 fold lower than gas producing technologies, or 2.3–6.5 fold lower after gas conversion to electricity. Although MFC power production from wastewater was limited by substrate conductivity and strength, power densities from single chamber MFCs fed optimized synthetic solutions ( $\sim 1.4 \text{ kJ per g COD}$ )<sup>67</sup> would have still been only 19–55% of the average reported energy recovery rates for methane-producing technologies.

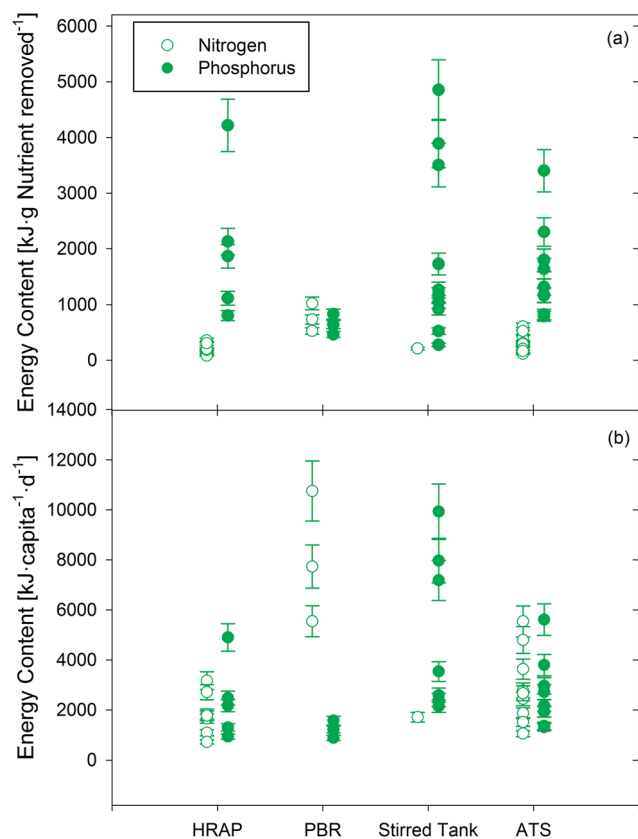




**Table 1** Average and range of percent COD or nutrient (N or P) removal for each technology used in Fig. 2 and 3

Technology	Average percent removal (min, max)	
	COD	
ASBR	58.1 (33, 91)	
UASB	67.6 (54, 85)	
ABR	90.3 (88.7, 92.5)	
AFB	82 (72, 89.7)	
AnMBR	86.7 (82, 90)	
MEC	78 (33.7, 96.7)	
MFC	45.5 (19, 83)	
	N	P
HRAP	67.1 (36, 87.2)	52.1 (32, 72.9)
PBR	78.5 (68, 89.7)	93.2 (85, 99)
Stirred tank	62.3	78.2 (7, 100)
ATS	70.5 (18.1, 90.7)	78.6 (58.3, 95.7)

Although discussions linking energy and nutrients in wastewater are generally focused on potential fertilizer offsets from nutrient recovery (e.g., ref. 26), the average energetic content of cultivated phototrophic biomass across all technologies (kJ per capita per day) was 130–510% of the energy saved from offsetting fertilizer production (assuming 100% nutrient recovery and 69 kJ per g N (ref. 143) and 7.6 kJ per g P (ref. 143) for synthetic fertilizer production). The average bioenergy feedstock production by phototrophic systems ranged from 210 kJ per g N (HRAP) to 760 kJ per g N (PBR), and 640 kJ per g P (PBR) to 2500 kJ per g P (stirred tank) (see Fig. 3a and Section S3.2 in ESI† for energy production values for each technology). On a per capita basis, the average energy production for each technology was as follows (if both N and P data were available in a given data set, the lesser prediction of biomass production based on per capita N and P was used): stirred tank reactor ( $4700 \pm 3200$  kJ per capita per day), ATS ( $2300 \pm 1100$  kJ per capita per day), HRAP ( $1800 \pm 860$  kJ per capita per day), and PBR ( $1200 \pm 340$  kJ per capita per day) (Fig. 3b). This range equates to  $3400\text{--}13\,000$  kJ  $\text{m}^{-3}$  of treated wastewater, or 280–400% of the potential recovery from methane-producing anaerobic technologies. As expected, the ratio of energy yield to P uptake was 2–14 $\times$  higher as compared to N uptake (except for PBR data). One explanation for the low energy potential per gram of P in PBRs is the low N : P removal ratio reported by the majority PBR studies (all of which had influent N : P ratios less than 1). For HRAP, stirred tank reactors, and ATS systems, the ratio of N to P mass uptake was roughly  $7.5 \pm 3.0$  (average  $\pm$  standard deviation). These values are higher than typical assumptions of biochemical composition of microalgae using an N : P mass ratio of 4.5 : 1 (N : P molar ratio of 10 : 1; e.g., ref. 144), but within the range that algae can adapt their N : P ratio (reported mass ratios range from 2.3–45 : 1 ref. 145). In the case of PBR experiments, data analysis was limited to two wastewater-relevant studies with adequate data. Additionally, it is possible that low ratios of biomass production per P removed were partially the result of alternate mechanisms (other than growth) including luxury uptake of P (microalgae have been shown to accumulate up to  $\sim 3\times$  normal cellular P<sup>146</sup>) and P



**Fig. 3** (a) Energy potential of phototrophic technologies [kJ algal biomass per g nutrient removed] showing relative bioenergy feedstock production based on nutrient removed (N or P). (b) Energy content [kJ algal biomass per capita per day] determined by multiplying values from (a) by 13 g N per capita per day or 2.1 g P per capita per day and by the respective average percent N and P removals for each technology (Table 1). Energy products are reported as kJ using theoretical unit conversions (see ESI†). Individual points represent distinct experimental data sets, with error bars extending from high to low COD/VSS assumptions (discussed in Section 4.2.2).

adsorption to cell surfaces.<sup>147</sup> Additional sources of feedstock production variability may have included carbon limitation, reactor and process design, and/or differences in lighting efficiencies. Nutrient rich phototrophic systems are often carbon limited owing to a C : N molar ratio typically less than cell requirements ( $\sim 3 : 1$  in typical wastewater vs.  $6 : 1$  cellular).<sup>32,148,149</sup>

**5.1.2 Energy consumption of anaerobic and phototrophic technologies.** While the goal of this manuscript was to examine the potential for full-scale anaerobic and phototrophic technologies to achieve energy positive treatment, pilot- and full-scale data in the peer-reviewed literature was severely limited requiring the inclusion of laboratory-scale data. Recognizing that a full-scale plant would be operated differently (and likely more efficiently) than its laboratory-scale counterpart, analysis of laboratory-scale data may still offer meaningful insights to the major barriers to full-scale implementation that each technology faces. Therefore, the energy consumption analysis that follows is not argued to be a perfectly accurate representation of full-scale energy consumption, but rather a starting point for a



discussion of how design and operational decisions will influence the ability of anaerobic and phototrophic technologies to achieve energy positive wastewater management.

Estimated energy consumption was highly variable across technologies (Tables 2 and 3), due to the wide range of experimental conditions and operational controls (e.g., fouling prevention and mitigation). All systems were assumed to be gravity fed, with energy consumption resulting from any additional operational requirements (assumptions detailed in Section S4 of ESI†). Since UASB, ABR, and MFC generally operate as passive systems, the operational energy consumption for these anaerobic technologies was minimal (assuming the reactor was not heated). Studies included in this analysis operated between 10 and 35 °C, but it should be noted the decision to heat would require roughly 4200 kJ m<sup>-3</sup> per 1 °C increase above influent wastewater temperature; the magnitude of this energy demand underscores the importance of operation at ambient temperatures and the importance of developing anaerobic technologies capable of psychrophilic operation.

Although mechanical mixing and applied voltage result in appreciable energy consumption for ASBR and MEC,

**Table 2** Ranges of energy consumption for anaerobic technologies based on experimental conditions in examined literature (kJ m<sup>-3</sup> wastewater treated)

Technology	Mixing	Pumping	Heating	Applied voltage
ASBR	4800–9400 <sup>a</sup>	28–31 <sup>b</sup>	4200 <sup>f</sup>	—
UASB	—	—	—	—
ABR	—	—	—	—
AnMBR	42 000–58 000 <sup>c</sup>	36–120 <sup>d</sup>	—	—
AFB	—	55–130 <sup>e</sup>	—	—
MEC	—	—	—	2800–7900
MFC	—	—	—	—

<sup>a</sup> Mechanical mixing (Section S4.1 and Table S3). <sup>b</sup> Effluent pumping (Section S4.5 and Table S3). <sup>c</sup> Biogas sparging (Section S4.2 and Table S3). <sup>d</sup> Permeate pumping (Section S4.5 and Table S3). <sup>e</sup> Recirculation pumping (Section S4.5 and Table S3). <sup>f</sup> Energy required for each increase in °C (not included in final energy balance) (Section S4.6 and Table S3).

**Table 3** Ranges of energy consumption for phototrophic technologies based on experimental conditions in examined literature (kJ m<sup>-3</sup> wastewater treated)

Technology	Mixing	Pumping	Harvesting <sup>g</sup>
HRAP	3.2–9.6 <sup>b</sup>	—	34–170
PBR	6300–13 000 <sup>c</sup>	55–58 <sup>d</sup>	—
Stirred tank	770–3100 <sup>e</sup>	28–31 <sup>f</sup>	—
WSP	—	—	—
ATS	—	—	—

<sup>a</sup> Low value is coagulation–flocculation with belt press filter for dewatering, high value is gravity settling with centrifugation (Section S4.4 and Table S3). <sup>b</sup> Paddlewheel mixing (Section S4.3 and Table S3). <sup>c</sup> Aeration (Section S4.2 and Table S3). <sup>d</sup> Influent lift pump (Section S4.5 and Table S3). <sup>e</sup> Mechanical mixing (Section S4.1 and Table S3). <sup>f</sup> Effluent pumping (Section S4.5 and Table S3). <sup>g</sup> Although minimal energy would be required for the physical harvesting of algae from ATS, it was not estimated due to lack of available data.

respectively, the largest source of energy consumption among anaerobic technologies was gas sparging to manage membrane fouling in AnMBR. Estimates of energy consumption from biogas sparging were based on published rates from 0.67 (ref. 150) to 0.93 (ref. 47) L<sub>Gas</sub> L<sub>Reactor</sub><sup>-1</sup> min<sup>-1</sup>, which represent very high rates of gas addition to reactors. Scale-up and more targeted gas scouring techniques can certainly reduce the gas flow demand,<sup>151</sup> but the use of alternative approaches to fouling mitigation and prevention may be even more energetically favorable. In particular, external cross-flow AnMBR configurations<sup>34</sup> or staged reactors with media for biofilm attachment<sup>152</sup> may offer distinct advantages over submerged reactors, so long as the operational conditions are scalable and they mitigate fouling with less energy-intensive methods than gas scouring.

For phototrophic technologies, PBRs, which also rely on gas sparging for mixing, had the highest energy consumption. Typical sparging rates in PBRs are often 0.1–0.3 L<sub>Gas</sub> L<sub>Reactor</sub><sup>-1</sup> min<sup>-1</sup>,<sup>78,153</sup> with actual rates in the field dependent upon biomass characteristics and tendency to aggregate. Ultimately, however, mixing requirements for algal systems are less than many chemotrophic systems due to decreased cell aggregation and a higher sensitivity of algal cells to shear forces.<sup>154</sup> Passive systems such as WSP and ATS consume almost no energy during operation, as is also the case with HRAPs (which require very few paddlewheels per hectare). These systems require much larger land areas,<sup>86</sup> however, resulting in a distinct trade-off between aerial productivity and energy consumption during cultivation.

## 5.2 Energy balance & treatment efficacy of anaerobic technologies

**5.2.1 Usable energy balance for anaerobic processes.** Given that treatment processes are generally powered by electricity, the caloric content of the gaseous products from anaerobic processes were converted to electricity and compared with consumption (Fig. 4). MFCs were the only technology evaluated that can directly produce electricity. The estimated electricity recovery from methane and hydrogen was assumed to be 42.3% for conversion of methane or hydrogen using fuel cells.<sup>133</sup> A significant amount of energy (nearly 60%) is lost in the conversion of alternative fuels to electricity used directly by treatment plants, which further limits the potential for energy neutral operation. The red boxes in Fig. 4 represent energy consumption normalized to g COD removed, excluding energy from heating. These values were calculated by converting the data from Table 2 (energy consumption per m<sup>3</sup> treated) to a COD removal basis using average COD removal efficiencies (Table 1) and an assumed influent concentration of 500 g COD per m<sup>3</sup>. The energy demand for heating (included in Table 3) was excluded from the energy balance because many of the studies operated at ambient temperatures, and no trend was observed between operating temperature and energy recovery or production (data not shown).

Although all anaerobic technologies were capable of recovering energy, only four appear to be immediately capable of net energy positive operation: UASB, ABR, AFB, and MFC. It was assumed that three technologies (UASB, ABR, and MFC) could



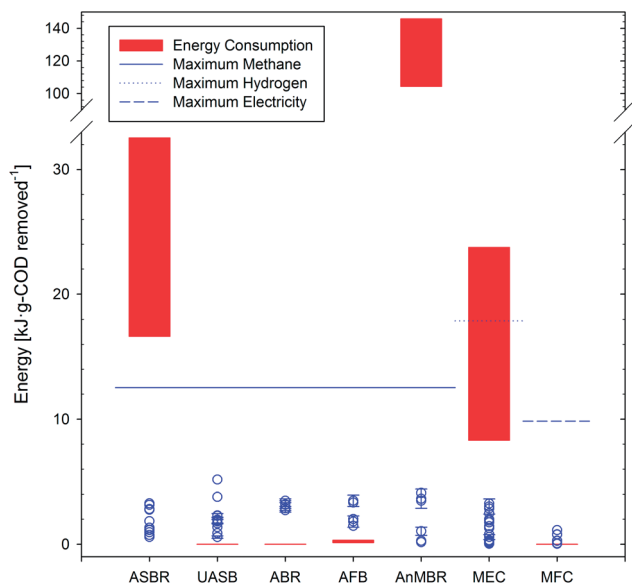


Fig. 4 Energy recovery, consumption, and theoretical maximum energy yield for each technology. Blue circles represent energy production per gram of COD removed in experimental data sets from the literature. Red boxes – indicating the range of energy consumed that needs to be overcome for energy positive treatment (excluding heating requirements of the wastewater) – were calculated based on volumetric energy requirements (Table 2) coupled with typical COD removal of each technology (Table 1) and an assumed influent of 500 g COD per m<sup>3</sup>. Blue horizontal lines show the maximum energy that can be generated for methane (solid), hydrogen (dotted), and electricity (dashed) based on thermodynamics (calculations shown in the Section S3 of ESI†).

be operated as passive systems with no significant operational energy. For the remaining four technologies, energy consumption demands were exacerbated by COD removal efficiencies below 100% (e.g., ASBRs – 58.1% COD removal efficiency – require 1.7 m<sup>3</sup> for every 500 g COD degraded). With ASBR energy recovery ranging from 1.4–7.7 kJ per g COD, the energy balance was hindered by the energy intensity of mechanical mixing (17–33 kJ per g COD) and, to a lesser extent, effluent pumping (0.10–0.11 kJ per g COD). To achieve energy neutrality, the energy for mixing must be drastically reduced. While AnMBRs achieved some of the highest energy recovery values (up to 86.7% with 0.4–9.7 kJ per g COD), continuous biogas sparging for mixing as well as to prevent and mitigate membrane fouling led to significant energy consumption (100–145 kJ per g COD). The gas flow rate needed for sparging would have to decrease by more than an order of magnitude to about 0.03 L<sub>Gas</sub> L<sub>Reactor</sub><sup>-1</sup> min<sup>-1</sup> (with no increase in TMP) or alternative strategies for fouling management would have to be developed in order for energy neutrality to be achieved. Alternatively, the divide between energy recovery and consumption could be narrowed if methane recovery from the effluent were improved (on the order of 30–50% of produced methane may be lost to the effluent<sup>47,155</sup>). MEC energy consumption is a function of applied voltage and current production, and could be improved by developing cost effective low over-potential hydrogen evolution catalysts. However, operating any catalyst in

wastewater will likely limit the kinetics of proton reduction. MECs have the highest energy recovery potential (based on the thermodynamics of hydrogen, methane, and electricity production; see Fig. 4 and Section S3.1, ESI†) and recovered more energy than MFCs, but the energy consumed by applying a voltage make MECs less energetically favorable than MFCs for low strength wastewater treatment.

### 5.2.2 Efficacy of anaerobic technologies for COD removal.

To replace energy intensive aerobic processes, anaerobic technologies must balance energy production with efficient COD removal. Although limited data was available in many cases, a review of the literature revealed that COD removal was highest (>85%) in systems that included physical separation of biomass from the effluent (AnMBRs; Fig. 5d) or leveraged attached growth (AFBs; Fig. 5c). The variability of COD removal in METs was the highest (Fig. 5e), which can be partially attributed to reactor operation. Continuous flow METs, for example, achieved lower COD removals than batch-fed reactors. Comparison with energy consumption data shows that trade-offs between net energy balance and effluent quality do exist in some cases (e.g., AnMBR), but alternative configurations may be able to achieve a high quality effluent under net energy positive operation (AFB). Since the carbon energy density of domestic wastewater is low, innovative solid-liquid separation methods will be needed to meet traditional treatment objectives and achieve energy positive COD removal.

## 5.3 Energy balance & treatment efficacy of phototrophic technologies

### 5.3.1 Usable energy balance of phototrophic processes.

In the conversion of phototrophic biomass to usable energy, HTL achieves the highest energy output followed by anaerobic digestion, combustion, and transesterification (maximum values in that order, Table 4; details of assumptions in Section S4.8 of the ESI†). Although biocrude and biodiesel have a higher energetic potential per mass of fuel (33.2 and 37.2 MJ per kg fuel for biocrude oil or biodiesel, respectively)<sup>38,138</sup> compared to direct combustion (14.2–21.4 MJ per kg fuel, where the dried biomass is the fuel),<sup>156,157</sup> conversion technologies suffer from low conversion efficiencies from algal biomass to fuel (HTL 25–54%; transesterification 10–30%) resulting in appreciable energetic losses as undesirable byproducts (e.g., the solid and aqueous fractions of HTL).<sup>158</sup> However, liquid fuels offer distinct advantages as compared to electricity and heat (e.g., liquid fuels can be easily stored and transported), and there may be circumstances under which energetic losses are adequately offset by the convenience or life cycle impacts of liquid biofuel production. Among liquid fuels, HTL is able to yield oil from not only the neutral lipids, but also other macromolecules to achieve an appreciable increase in energy yield (Table 4). Given that phototrophic biomass cultivated in wastewater is often observed to have low levels of neutral lipids (~10% of dry weight),<sup>38</sup> HTL may have significant potential for wastewater-derived phototrophic biomass. In terms of feedstock preparation for downstream conversion, HTL and anaerobic digestion can both process biomass in a wet state,<sup>159</sup> rather than requiring



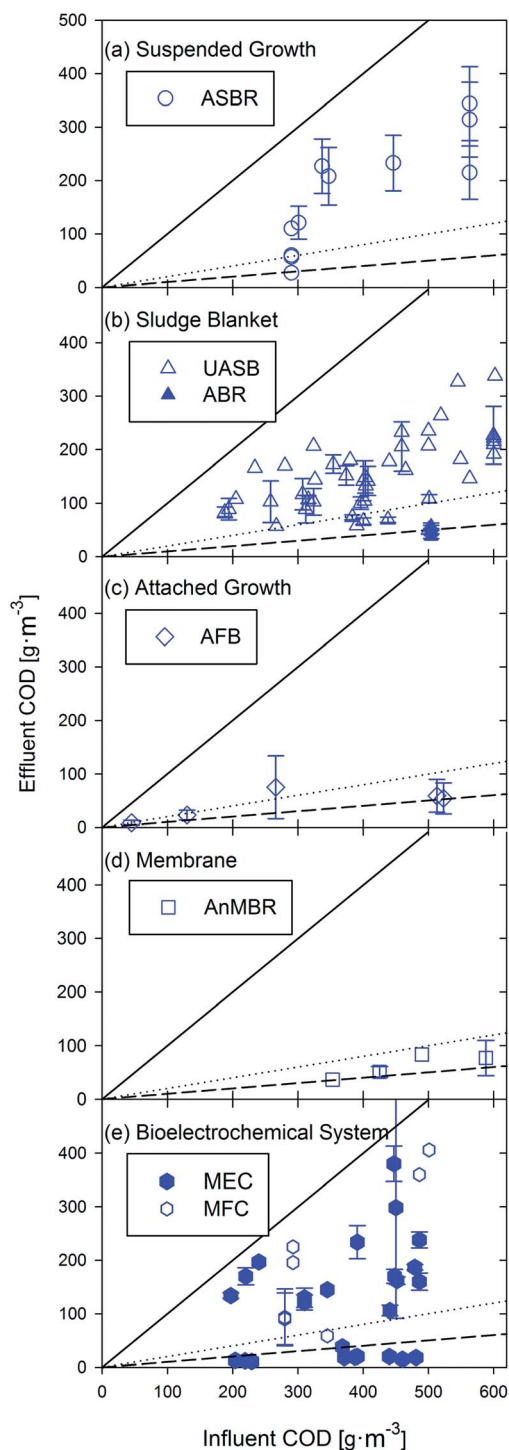


Fig. 5 Influent vs. effluent COD [ $\text{g}\cdot\text{m}^{-3}$ ] from anaerobic treatment technologies treating real and synthetic wastewaters with influent COD concentrations  $<500\text{ g}\cdot\text{m}^{-3}$ . Points and error bars represent averages  $\pm$  standard deviations from experimental data sets. Plots (a)–(e) are separated by technology type (suspended growth, sludge blanket, etc.). The solid line is no COD removal (i.e., 0% removal), the dotted line is 80% removal, and the dashed line is 90% removal.

drying that can demand energy equivalent to the energetic content of the biomass itself.<sup>160</sup> Anaerobic digestion, in particular, is an accessible technology that is well proven at large

scales (it is currently in operation at 1300 WWTPs in the U.S. alone<sup>133</sup>) and has been clearly demonstrated to enable nutrient recycling to agriculture.<sup>161,162</sup> A key challenge for the integration of phototrophic biomass into digestion processes, however, is maintaining an appropriate C : N ratio.<sup>163</sup>

As expected from the consumption data, PBRs and stirred tank reactors face the greatest difficulty in becoming energy neutral or positive, while HRAPs, WSPs, and ATSS may be energy positive once harvesting and low conversion efficiencies have been overcome.<sup>164</sup> Although harvesting is often cited as one of the most critical energy challenges to meet,<sup>165</sup> it is clear that mechanical mixing and aeration must also be reduced if these technologies are to be energy positive. Energy consumption for each cultivation technology (shown in Table 3 [ $\text{kJ}\cdot\text{m}^{-3}$ ] and Table S5† [ $\text{kJ}$  per  $\text{g}$  nutrient removed]) was highest for PBRs (230–470  $\text{kJ}$  per  $\text{g}$  N and 1200–2400  $\text{kJ}$  per  $\text{g}$  P) and stirred tank reactors (40–150  $\text{kJ}$  per  $\text{g}$  N and 180–730  $\text{kJ}$  per  $\text{g}$  P), while HRAPs had relatively low energy demand (2–7  $\text{kJ}$  per  $\text{g}$  N and 10–60  $\text{kJ}$  per  $\text{g}$  P) and WSPs and ATSS required no appreciable energy input during operation. When compared to the energy yield of various conversion technologies (Table 4), energy production by PBRs and stirred tank reactors may have potentially favorable energy balances depending on the conversion process and variability of biomass generation within each technology. HRAPs and ATSS, however, are far more likely to be energy positive across the range of biomass yields due to minimal operational energy demands (Table 3). Although data was not available to estimate energy yield from WSPs, they would also have the potential to achieve energy positive treatment if energy efficient biomass harvesting can be achieved.

It is important to note that the energy consumption calculated for this study does not include the energy needed for the conversion process itself. There is a large degree of uncertainty associated with these technologies, some of which (like HTL) have yet to be implemented on a large scale for phototrophic biomass. However, there is still room for these technologies to be net energy positive when incorporating conversion energy demand. For example, PBRs obtained a maximum energy yield of  $\sim 580\text{ kJ}$  per  $\text{g}$  N for anaerobic digestion and HTL. With a cultivation energy demand of 230–470  $\text{kJ}$  per  $\text{g}$  N (Table S5†), there is still 110–350  $\text{kJ}$  per  $\text{g}$  N that can be used for driving the conversion process.

**5.3.2 Efficacy of phototrophic technologies for N and P removal and recovery.** To replace chemotrophic nutrient removal processes, phototrophic technologies must achieve efficient N and P removal below permit levels. A review of the literature revealed that the highest levels of N removal (average 78.5%) were achieved in PBRs. Although HRAPs and ATSS had similar maximum values of removal (87.2% and 90.7%, respectively, compared to 89.7% for PBR), they had higher variability in performance (Table 1 and Fig. 6a). PBRs also achieved the highest consistent levels of percent P removal (Table 1 and Fig. 6b). Additionally, although experimental conditions varied greatly across studies, PBRs have been demonstrated to achieve effluent concentrations below  $3\text{ g N}$  per  $\text{m}^3$  and both PBRs and stirred tanks achieved effluent P levels below  $0.3\text{ g P}$  per  $\text{m}^3$  (a subset of ATSS studies also demonstrated



Table 4 Energy yield (kJ fuel per g nutrient removed) for phototrophic cultivation technologies and select conversion processes<sup>a,b</sup>

Technology	Nutrient	HTL	Anaerobic digestion	Transesterification	Combustion
HRAP	N	75–160	32–160	34–100	90–130
	P	730–1600	320–1500	330–980	880–1300
PBR	N	270–590	120–580	120–370	330–500
	P	230–500	100–490	100–310	280–420
Stirred tank <sup>c</sup>	P	900–1900	400–1900	400–1200	1100–1600
ATS	N	110–240	47–230	49–150	130–200
	P	580–1300	250–1200	260–790	700–1100

<sup>a</sup> Calculations and assumptions can be found in Table S4. <sup>b</sup> WSP could not be included due to lack of available biomass productivity data. <sup>c</sup> Data was not available to estimate kJ fuel per g N removed.

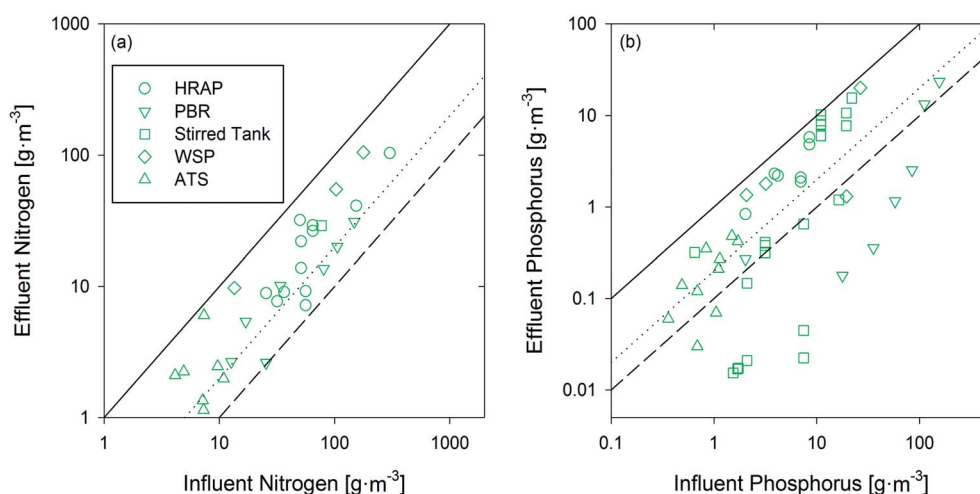


Fig. 6 Influent vs. effluent (a) total N concentrations [g N per m<sup>3</sup>] and (b) total P concentrations [g P per m<sup>3</sup>] for suspended and attached growth systems on a log–log scale. The solid line identifies no nutrient removal, the dotted line 80% removal, and the dashed line 90% removal.

effluent P concentrations below 0.3 g P per m<sup>3</sup>, but these studies had influent P concentrations below 1 g P per m<sup>3</sup>.

When compared to energy consumption data, it can be seen that the technologies that require more energy (PBRs, stirred tank reactors) tend to perform better in meeting traditional treatment objectives such as N and P removal from wastewater. They also generate more biomass and more energy per gram nutrient removed (Fig. 3) with which to offset this energy consumption. Balancing increased nutrient removal and biomass yields (and thus, energy production) with higher energy demands will be a key challenge in the design and development of energy positive phototrophic systems.

## 6 Navigating a path to energy positive wastewater management

A striking conclusion of this review was that phototrophic processes have the potential to produce 280–400% of the amount of energy as anaerobic processes on a per m<sup>3</sup> basis, given existing pollutant removal efficiencies and downstream conversion technologies. The energy recovery by anaerobic technologies reported in this manuscript (2–47%) assumes an

energetic content for COD of 13.9 kJ per g COD, which has recently been found to be a low estimation.<sup>142</sup> A higher energetic content would further reduce anaerobic energy recovery efficiency, whereas cultivating algae on nutrients and converting to fuels could exceed the original energetic content of the influent wastewater. Additionally, the use of nutrients for phototrophic cultivation may result in 130–510% of the energy production as would be offset by the use of nutrients for fertilization. An unfortunate finding of this review was the lack of adequate data to enable a coordinated analysis of both energy production and wastewater treatment. Of the 311 papers screened in the initial literature search, 82% could not be included because they did not measure or report adequate data. From the available data, it is clear that the potential exists for energy positive wastewater treatment and that both anaerobic and phototrophic may play a role. However, there are several critical barriers that must be overcome. Anaerobic processes must balance reduced energy consumption with increased treatment efficacy and fuel recovery, and we must develop a deeper understanding of phototrophic bioprocesses to enable process optimization. To this end, we examine the implications of this work and propose areas for future research.



## 6.1 Implications of this work

This review examines the potential of various biotechnologies to directly treat domestic wastewater with a positive operating energy balance. For anaerobic technologies, influent COD is an important determinant of fuel production; higher COD concentrations lead to more energy recovery and less energy consumption (per gram of COD degraded). Since freshwater serves as a carrier for human waste in developed countries, domestic wastewater is often dilute, limiting the amount of energy that can be recovered during secondary treatment. For phototrophic technologies, a similar relationship exists between influent N and P concentrations and biomass yields. Despite limited energy recovery and production values, replacing energy intensive COD and nutrient removal processes could enable treatment plants that have already established solids digestion and on-site electricity generation to achieve energy positive operation.

At the forefront of energy-conscious wastewater treatment with aerobic COD removal and BNR, an activated sludge WWTP in Strass, Austria has achieved energy self-sufficiency by implementing a high rate aerobic process, anammox treatment of nutrient rich side streams, and on-site electricity generation from biogas generated by solids digestion. A published COD mass balance and energy analysis of the plant indicated that 75% of the COD entering the plant is fed to a digester (61% primary and high rate solids and 14% waste solids from biological nutrient removal) and 36% is converted to biogas.<sup>166</sup> The aerobic BNR process, in which 31% of the influent COD and 80% of the N is removed, accounted for 45% of energy consumption at the plant. The Strass WWTP COD mass balance was used to simulate the energetic potential of replacing the existing aerobic processes with anaerobic and phototrophic wastewater treatment. If the BNR process was replaced with an ABR to remove COD and a HRAP to remove nitrogen, total plant biogas production could potentially increase by 39% and energy recovery from COD could reach 41% (Section S5 of ESI†). The energetic content of biomass produced in the HRAP during N removal (2200 kJ per capita per day) was estimated to be more than twice as much as recovered biogas (1020 kJ per capita per day). If PBRs were employed rather than HRAPs, the estimated biomass energy content alone (7800 kJ per capita per day, assuming N-limited growth) could be more than three times the caloric energy content of wastewater entering the plant (2500 kJ per capita per day). More broadly, combined anaerobic and phototrophic processes could reduce energy demand and achieve energy recovery and production on the order of 5.0–9.2 kW h m<sup>-3</sup> (using higher values for UASBs and PBRs) – well above the whole-plant energy demand of conventional WWTPs (0.3–0.6 (ref. 20 and 21)).

Though achieving energy and resource positive treatment in developed countries is an important goal for future treatment, far more urgent is the need to deploy sanitation infrastructure in developing and underdeveloped communities, where an estimated 2.5 billion people lack access to improved sanitation.<sup>167</sup> Even in cases where individuals have access to bathroom facilities and collection systems, it is estimated that globally 1.5

billion people connected to sewerage infrastructure have their wastewater discharged without treatment.<sup>168</sup> In developing communities in tropical regions, mainstream anaerobic treatment of domestic wastewater has been shown to be a viable means of achieving treatment goals while simultaneously producing biogas.<sup>169</sup> This biogas, if utilized properly, could be an invaluable resource providing a consistent supply of electricity. In developing countries, effluent from anaerobic treatment processes (*e.g.*, UASB) can be fed to WSP for further treatment.<sup>170,171</sup> The data analysis presented in this review indicates that converting WSPs to HRAPs is a path toward more meaningful energy production from wastewater management. Ultimately, one of the greatest opportunities to advance wastewater treatment in developing communities is to recover resources that make wastewater management energy positive and financially viable.

## 6.2 Future research needs – anaerobic technologies

The experimental results compiled in this review clearly show that energy recovery in the form of methane gas is significantly higher than energy recovery by MECs and MFCs. While methane-producing technologies do not require electrodes or applied voltage to generate fuel, converting biogas to electricity requires expensive auxiliary equipment (*i.e.*, gas conditioning, storage, prime movers or fuel cells) and is currently only feasible at high flow wastewater treatment facilities (>30 MGD).<sup>133</sup> Of the more than 1300 treatment plants that employ anaerobic digestion for solids management in the U.S., only 364 are sites generate enough biogas to make combined heat and power (CHP) financially viable, of which 104 currently generate electricity from biogas.<sup>133</sup> Primary anaerobic treatment would make CHP accessible to smaller WWTPs, but it remains to be seen at what scale economic feasibility could be reached.

Though methane is relatively insoluble in water (Henry's constant = 776 bar L mol<sup>-1</sup>), loss of dissolved methane in the wastewater effluent continues to be a critical challenge for anaerobic processes.<sup>172,173</sup> This loss of fuel removes much of the potential for anaerobic processes to be energy positive, especially since energy savings from psychrophilic operation are in tension with increased energy losses due to higher methane solubility at lower temperatures.<sup>47</sup> Finding alternative methods to recover dissolved methane without excessive energy input (*i.e.*, using an amount of energy less than the amount recovered) will be pivotal to achieve energy positive treatment with AnMBR.

In terms of energy recovery, MFC bioelectricity was significantly lower than gaseous products. However, when fuel conversion to electricity was considered, the discrepancy between MFCs and gas-producing technologies was less substantial, indicating that MFCs may be a favorable option for distributed electricity production from wastewater. To capitalize on this potential, research efforts should focus on anode and passive-air cathode fabrication without the use of expensive materials as well as evaluation of power production from source separated waste streams. METs can also be designed to operate in concert with methane-producing processes to enhance treatment efficiency and recover nutrients. Allocating a portion



of soluble organic energy to produce electrical current with MET electrodes could be leveraged toward electrolytic pH adjustment to volatilize and concentrate ammonia<sup>174–176</sup> or recover N and P as struvite.<sup>177,178</sup> Ionic current produced by MET could also be used to polarize capacitor electrodes and remove charge molecules such as nutrients and minerals from wastewater.<sup>179</sup>

### 6.3 Future research needs – phototrophic technologies

Although the predominant focus of nutrient research in the wastewater field has been on improving the efficiency of BNR by chemotrophic bacteria, the energetic potential of phototrophic processes warrants further development of these processes for energy positive nutrient management. In particular, more highly engineered systems that minimize footprint (like PBRs and stirred tank reactors) may have potential in advancing nutrient removal initiatives while also increasing the energy independence of treatment facilities. A critical challenge in achieving reliable and resilient phototrophic treatment systems, however, is a lack of understanding of how process design and operational decisions influence effluent quality, biomass productivity, and biochemical composition.<sup>144</sup> Developing a deeper understanding of mixed community phototrophic biotechnology in the context of wastewater treatment will require long-term experimentation with real wastewaters under natural light (or simulated natural light) conditions with diurnal cycles. Targeted experimentation and modeling may enable process optimization, but a priority should be to determine how complex models will need to be to enable reliable predictions of performance across climates and wastewaters.<sup>180,181</sup>

Harvesting and downstream processing to usable fuels are also opportunities for technology advancement, including research furthering the development of processing technologies that do not require complete drying of biomass prior to processing: anaerobic digestion and HTL hold particularly high potential in this regard. In addition to fundamental advancements to HTL and the management of waste products,<sup>182</sup> a critical challenge is to link process design decisions with downstream processing to usable energy. Without a mechanistic understanding of the links among cultivation decisions, biochemical composition, harvesting, and processing to fuel, any attempts at process optimization are likely to result in trade-offs that may be obscured by energetic impacts of design and operational modifications.

## 7 Conclusion

The pursuit of energy positive domestic wastewater treatment is a necessity due to both the financial costs and the broader environmental impacts incurred by energy consumption. Beyond economic and environmental drawbacks, energy intensive treatment processes may also be infeasible for developing communities that may even lack the energy infrastructure to reliably treat wastewater aerobically. Based on the results of this review, it is clear that WWTPs can be net energy producers, especially if phototrophic technologies are leveraged to increase

the energetic potential of wastewater through inorganic carbon fixation. In the search for energetically favorable technologies, however, there is a critical point to be made: we should not compromise traditional sanitary engineering objectives for wastewater treatment systems (*i.e.*, effluent quality) to achieve energy positive performance, but rather seek to develop technologies that achieve equivalent or superior effluent quality by leveraging biological, chemical, and physical processes whose treatment efficacy is not in direct tension with their energy balance. Therefore, we should seek to advance technologies that have synergies between effluent quality and energy production, such as anaerobic and phototrophic technologies where every gram of pollutant removed increases the potential energy yield from the system.

## Acknowledgements

This work was partially funded by the King Abdullah University of Science and Technology (KAUST) Academic Partnership Program (UIERA 2012-06291), and by the Center of Advanced Materials for the Purification of Water with Systems (Water-CAMPWS) under NSF Agreement Number CTS-0120978. The authors would like to acknowledge the Jack Kent Cooke Foundation for partial funding for B.D. Shoener, and the Safe Global Water Institute (SGWI) at the University of Illinois at Urbana-Champaign (UIUC) for partial funding for I.M. Bradley. We would also like to thank Cheng Zhong (UIUC) and Anthony Greiner (Hazen and Sawyer) for their assistance in energy consumption estimation, Shijie (Moses) Leow (UIUC) for discussions on hydrothermal liquefaction, and the anonymous reviewers for their constructive comments that strengthened the manuscript.

## References

- 1 N. Carey, *Reuters*, 2013.
- 2 Water Infrastructure Network, *Clean and Safe Water for the 21st Century: A Renewed National Commitment to Water and Wastewater Infrastructure*, Water Infrastructure Network, 2001.
- 3 J. Kotkin, *Forbes*, 2013.
- 4 W. H. Frey, *A Big City Growth Revival?*, Brook. Inst., 2013.
- 5 US Census Bureau, 2013.
- 6 ASCE, *2013 Report Card for America's Infrastructure*, 2013.
- 7 H. W. Paerl, L. M. Valdes, A. R. Joyner, M. F. Piehler and M. E. Lebo, *Environ. Sci. Technol.*, 2004, **38**, 3068–3073.
- 8 D. J. Conley, H. W. Paerl, R. W. Howarth, D. F. Boesch, S. P. Seitzinger, K. E. Havens, C. Lancelot and G. E. Likens, *Science*, 2009, **323**, 1014–1015.
- 9 W. M. Kemp, W. R. Boynton, J. E. Adolf, D. F. Boesch, W. C. Boicourt, G. Brush, J. C. Cornwell, T. R. Fisher, P. M. Glibert, J. D. Hagy, L. W. Harding, E. D. Houde, D. G. Kimmel, W. D. Miller, R. I. E. Newell, M. R. Roman, E. M. Smith and J. C. Stevenson, *Mar. Ecol.: Prog. Ser.*, 2005, **303**, 1–29.
- 10 N. N. Rabalais, R. E. Turner and W. J. Wiseman Jr, *Annu. Rev. Ecol. Syst.*, 2002, **33**, 235–263.



- 11 USEPA, *Evaluation Report: Despite Progress, EPA Needs to Improve Oversight of Wastewater Upgrades in the Chesapeake Bay Watershed*, USEPA, Office of the Inspector General, Washington, D.C., 2008.
- 12 USEPA, *Wastewater Management Fact Sheet: Energy Conservation*, Office of Water, 2006.
- 13 M. Waycott, C. M. Duarte, T. J. B. Carruthers, R. J. Orth, W. C. Dennison, S. Olyarnik, A. Calladine, J. W. Fourqurean, K. L. Heck, A. R. Hughes, G. A. Kendrick, W. J. Kenworthy, F. T. Short and S. L. Williams, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 12377–12381.
- 14 A. S. Mori, T. Furukawa and T. Sasaki, *Biol. Rev.*, 2013, **88**, 349–364.
- 15 G. E. Ponce-Campos, M. S. Moran, A. Huete, Y. Zhang, C. Bresloff, T. E. Huxman, D. Eamus, D. D. Bosch, A. R. Buda, S. A. Gunter, T. H. Scalley, S. G. Kitchen, M. P. McClaran, W. H. McNab, D. S. Montoya, J. A. Morgan, D. P. C. Peters, E. J. Sadler, M. S. Seyfried and P. J. Starks, *Nature*, 2013, **494**, 349–352.
- 16 J. R. Bernhardt and H. M. Leslie, *Annu. Rev. Mar. Sci.*, 2013, **5**, 371–392.
- 17 Z. W. Kundzewicz, L. J. Mata, N. W. Arnell, P. Döll, B. Jimenez, K. Miller, T. Oki, Z. Şen and I. Shiklomanov, *Hydrol. Sci. J.*, 2008, **53**, 3–10.
- 18 T. Oki and S. Kanae, *Science*, 2006, **313**, 1068–1072.
- 19 G. A. Meehl and C. Tebaldi, *Science*, 2004, **305**, 994–997.
- 20 P. L. McCarty, J. Bae and J. Kim, *Environ. Sci. Technol.*, 2011, **45**, 7100–7106.
- 21 USEPA, *Clean Energy Opportunities in Water & Wastewater Treatment Facilities Background and Resources*, 2009.
- 22 K. Mizuta and M. Shimada, *Water Sci. Technol.*, 2010, **62**, 2256–2262.
- 23 R. Gori, L.-M. Jiang, R. Sobhani and D. Rosso, *Water Res.*, 2011, **45**, 5858–5872.
- 24 J. S. Guest, S. J. Skerlos, J. L. Barnard, M. B. Beck, G. T. Daigger, H. Hilger, S. J. Jackson, K. Karvazy, L. Kelly, L. Macpherson, J. R. Mihelcic, A. Pramanik, L. Raskin, M. C. M. Van Loosdrecht, D. Yeh and N. G. Love, *Environ. Sci. Technol.*, 2009, **43**, 6126–6130.
- 25 G. T. Daigger, *Water Environ. Res.*, 2009, **81**, 809–823.
- 26 W. Verstraete, P. Van de Caveye and V. Diamantis, *Bioresour. Technol.*, 2009, **100**, 5537–5545.
- 27 M. B. Beck, F. Jiang, F. Shi, R. V. Walker, O. O. Osidele, Z. Lin, I. Demir and J. W. Hall, *Proc. ICE - Eng. Sustain.*, 2010, **163**, 31–46.
- 28 A. L. Smith, L. B. Stadler, N. G. Love, S. J. Skerlos and L. Raskin, *Bioresour. Technol.*, 2012, **122**, 149–159.
- 29 R. A. Rozendal, A. W. Jeremiassé, H. V. M. Hamelers and C. J. N. Buisman, *Environ. Sci. Technol.*, 2008, **42**, 629–634.
- 30 R. D. Cusick, P. D. Kiely and B. E. Logan, *Int. J. Hydrogen Energy*, 2010, **35**, 8855–8861.
- 31 G. Lettinga, S. Rebac and G. Zeeman, *Trends Biotechnol.*, 2001, **19**, 363–370.
- 32 G. Tchobanoglous, F. L. Burton and H. D. Stensel, *Wastewater Engineering: Treatment and Reuse*, McGraw-Hill Professional, 2003.
- 33 W. P. Barber and D. C. Stuckey, *Water Res.*, 1999, **33**, 1559–1578.
- 34 B.-Q. Liao, J. T. Kraemer and D. M. Bagley, *Crit. Rev. Environ. Sci. Technol.*, 2006, **36**, 489–530.
- 35 I. M. Nasir, T. M. Ghazi and R. Omar, *Appl. Microbiol. Biotechnol.*, 2012, **95**, 321–329.
- 36 L. Seghezze, G. Zeeman, J. B. van Lier, H. V. M. Hamelers and G. Lettinga, *Bioresour. Technol.*, 1998, **65**, 175–190.
- 37 K. Solanki, S. Subramanian and S. Basu, *Bioresour. Technol.*, 2013, **131**, 564–571.
- 38 B. S. M. Sturm and S. L. Lamer, *Appl. Energy*, 2011, **88**, 3499–3506.
- 39 USEPA, *Response to National Resources Defense Council Petition Requesting Secondary Treatment Nutrient Limits and Performance Data*, USEPA, Office of Water, 2012.
- 40 USEPA, *Memorandum: Working in Partnership with States to Address Phosphorus and Nitrogen Pollution through Use of a Framework for State Nutrient Reductions*, USEPA, Office of Water, 2011.
- 41 J. S. Wang, S. P. Hamburg, D. E. Pryor, K. Chandran and G. T. Daigger, *Environ. Sci. Technol.*, 2011, **45**, 6239–6246.
- 42 L. Lardon, A. Hélias, B. Sialve, J.-P. Steyer and O. Bernard, *Environ. Sci. Technol.*, 2009, **43**, 6475–6481.
- 43 L. Batan, J. Quinn, B. Willson and T. Bradley, *Environ. Sci. Technol.*, 2010, **44**, 7975–7980.
- 44 C. Goodson, R. Roth, Z. T. Wang and U. Goodenough, *Eukaryotic Cell*, 2011, **10**, 1592–1606.
- 45 G. O. James, C. H. Hocart, W. Hillier, H. Chen, F. Kordbacheh, G. D. Price and M. A. Djordjevic, *Bioresour. Technol.*, 2011, **102**, 3343–3351.
- 46 A. Larrosa-Guerrero, K. Scott, I. M. Head, F. Mateo, A. Ginesta and C. Godinez, *Fuel*, 2010, **89**, 3985–3994.
- 47 A. L. Smith, S. J. Skerlos and L. Raskin, *Water Res.*, 2013, **47**, 1655–1665.
- 48 B. E. Logan and K. Rabaey, *Science*, 2012, **337**, 686–690.
- 49 D. J. Batstone, J. Keller, I. Angelidaki, S. V. Kalyuzhnyi, S. G. Pavlostathis, A. Rozzi, W. T. M. Sanders, H. Siegrist and V. A. Vavilin, *Water Sci. Technol.*, 2002, **45**, 65–73.
- 50 B. E. Logan, B. Hamelers, R. Rozendal, U. Schröder, J. Keller, S. Freguia, P. Aelterman, W. Verstraete and K. Rabaey, *Environ. Sci. Technol.*, 2006, **40**, 5181–5192.
- 51 R. R. Dague, C. E. Habben and S. R. Pidaparti, *Water Sci. Technol.*, 1992, **26**, 2429–2432.
- 52 A. Sarti, B. S. Fernandes, M. Zaiat and E. Foresti, *Desalination*, 2007, **216**, 174–182.
- 53 M. Zaiat, J. a. D. Rodrigues, S. M. Ratusznei, E. F. M. de Camargo and W. Borzani, *Appl. Microbiol. Biotechnol.*, 2001, **55**, 29–35.
- 54 K. S. Singh and T. Viraraghavan, *J. Ferment. Bioeng.*, 1998, **85**, 609–614.
- 55 B. E. Rittmann and P. L. McCarty, *Environmental biotechnology: principles and applications*, McGraw-Hill, Boston, 2001.
- 56 G. Lettinga, A. F. M. van Velsen, S. W. Hobma, W. de Zeeuw and A. Klapwijk, *Biotechnol. Bioeng.*, 1980, **22**, 699–734.
- 57 A. Bachmann, V. L. Beard and P. L. McCarty, *Water Res.*, 1985, **19**, 99–106.





- 58 D. W. Hendricks, *Fundamentals of water treatment unit processes: physical, chemical, and biological*, CRC Press, Boca Raton, 2011.
- 59 B. E. Logan and J. M. Regan, *Trends Microbiol.*, 2006, **14**, 512–518.
- 60 H. Liu, R. Ramnarayanan and B. E. Logan, *Environ. Sci. Technol.*, 2004, **38**, 2281–2285.
- 61 J. J. Heijnen, in *Encyclopedia of Bioprocess Technology*, John Wiley & Sons, Inc., 2002.
- 62 B. E. Logan, *Microbial Fuel Cells*, John Wiley & Sons, 2008.
- 63 S. Cheng, H. Liu and B. E. Logan, *Electrochem. Commun.*, 2006, **8**, 489–494.
- 64 D. Pant, G. Van Bogaert, L. Diels and K. Vanbroekhoven, *Bioresour. Technol.*, 2010, **101**, 1533–1543.
- 65 S. Hays, F. Zhang and B. E. Logan, *J. Power Sources*, 2011, **196**, 8293–8300.
- 66 R. D. Cusick, Y. Kim and B. E. Logan, *Science*, 2012, **335**, 1474–1477.
- 67 R. D. Cusick, M. Hatzell, F. Zhang and B. E. Logan, *Environ. Sci. Technol.*, 2013, **47**, 14518–14524.
- 68 J. R. Kim, G. C. Premier, F. R. Hawkes, J. Rodríguez, R. M. Dinsdale and A. J. Guwy, *Bioresour. Technol.*, 2010, **101**, 1190–1198.
- 69 F. Zhang, Z. Ge, J. Grimaud, J. Hurst and Z. He, *Environ. Sci. Technol.*, 2013, **47**, 4941–4948.
- 70 D. Call and B. E. Logan, *Environ. Sci. Technol.*, 2008, **42**, 3401–3406.
- 71 J. Ditzig, H. Liu and B. E. Logan, *Int. J. Hydrogen Energy*, 2007, **32**, 2296–2304.
- 72 L. Lu, D. Xing, B. Liu and N. Ren, *Water Res.*, 2012, **46**, 1015–1026.
- 73 A. Escapa, L. Gil-Carrera, V. García and A. Morán, *Bioresour. Technol.*, 2012, **117**, 55–62.
- 74 E. S. Heidrich, J. Dolfing, K. Scott, S. R. Edwards, C. Jones and T. P. Curtis, *Appl. Microbiol. Biotechnol.*, 2013, **97**, 6979–6989.
- 75 R. D. Cusick, B. Bryan, D. S. Parker, M. D. Merrill, M. Mehanna, P. D. Kiely, G. Liu and B. E. Logan, *Appl. Microbiol. Biotechnol.*, 2011, **89**, 2053–2063.
- 76 G. K. Rader and B. E. Logan, *Int. J. Hydrogen Energy*, 2010, **35**, 8848–8854.
- 77 USEPA, *Wastewater Technology Fact Sheet: Facultative Lagoons*, 2002.
- 78 C. Posten, *Eng. Life Sci.*, 2009, **9**, 165–177.
- 79 R. N. Singh and S. Sharma, *Renewable Sustainable Energy Rev.*, 2012, **16**, 2347–2353.
- 80 R. J. Craggs, S. Heubeck, T. J. Lundquist and J. R. Benemann, *Water Sci. Technol.*, 2011, **63**, 660–665.
- 81 Y. Chisti, *Biotechnol. Adv.*, 2007, **25**, 294–306.
- 82 E. Molina Grima, E.-H. Belarbi, F. G. Acién Fernández, A. Robles Medina and Y. Chisti, *Biotechnol. Adv.*, 2003, **20**, 491–515.
- 83 Y. Cohen, *Bioresour. Technol.*, 2001, **77**, 257–274.
- 84 J. C. M. Pires, M. C. M. Alvim-Ferraz, F. G. Martins and M. Simões, *Environ. Sci. Pollut. Res.*, 2013, **20**, 5096–5105.
- 85 A. Ruiz-Marin, L. G. Mendoza-Espinosa and T. Stephenson, *Bioresour. Technol.*, 2010, **101**, 58–64.
- 86 M.-O. P. Fortier and B. S. M. Sturm, *Environ. Sci. Technol.*, 2012, **46**, 11426–11434.
- 87 L. Rodolfi, G. C. Zittelli, N. Bassi, G. Padovani, N. Biondi, G. Bonini and M. R. Tredici, *Biotechnol. Bioeng.*, 2009, **102**, 100–112.
- 88 J. K. Pittman, A. P. Dean and O. Osundeko, *Bioresour. Technol.*, 2011, **102**, 17–25.
- 89 K. L. Terry and L. P. Raymond, *Enzyme Microb. Technol.*, 1985, **7**, 474–487.
- 90 J. B. K. Park and R. J. Craggs, *Water Sci. Technol.*, 2010, **61**, 633–639.
- 91 W. J. Oswald and C. G. Golueke, *Adv. Appl. Microbiol.*, 1960, **2**, 223–262.
- 92 O. Jorquera, A. Kiperstok, E. A. Sales, M. Embiruçu and M. L. Ghirardi, *Bioresour. Technol.*, 2010, **101**, 1406–1413.
- 93 A. L. Stephenson, E. Kazamia, J. S. Dennis, C. J. Howe, S. A. Scott and A. G. Smith, *Energy Fuels*, 2010, **24**, 4062–4077.
- 94 L. Xu, P. J. Weathers, X.-R. Xiong and C.-Z. Liu, *Eng. Life Sci.*, 2009, **9**, 178–189.
- 95 E. Molina Grima, F. G. A. Fernández, F. García Camacho and Y. Chisti, *J. Biotechnol.*, 1999, **70**, 231–247.
- 96 A. P. Carvalho, L. A. Meireles and F. X. Malcata, *Biotechnol. Prog.*, 2006, **22**, 1490–1506.
- 97 O. Pulz, *Appl. Microbiol. Biotechnol.*, 2001, **57**, 287–293.
- 98 J. M. Valigore, P. a. Gostomski, D. G. Wareham and A. D. O'Sullivan, *Water Res.*, 2012, **46**, 2957–2964.
- 99 G. Samori, C. Samori, F. Guerrini and R. Pistocchi, *Water Res.*, 2013, **47**, 791–801.
- 100 N. Renuka, A. Sood, S. K. Ratha, R. Prasanna and A. S. Ahluwalia, *J. Appl. Phycol.*, 2013, **25**, 1529–1537.
- 101 S. M. Oakley, A. Pocasangre, C. Flores, J. Monge and M. Estrada, *Water Sci. Technol.*, 2000, **42**, 51–58.
- 102 L. G. Rich, *Unit Operations of Sanitary Engineering*, J. Wiley & Sons, Incorporated, 1961.
- 103 T. P. Curtis, D. D. Mara and S. A. Silva, *Appl. Environ. Microbiol.*, 1992, **58**, 1335–1343.
- 104 O. C. Romero, A. P. Straub, T. Kohn and T. H. Nguyen, *Environ. Sci. Technol.*, 2011, **45**, 10385–10393.
- 105 T. Kohn and K. L. Nelson, *Environ. Sci. Technol.*, 2007, **41**, 192–197.
- 106 D. Mara, *Design Manual for Waste Stabilization Ponds in India*, Lagoon Technology International Ltd., 1997.
- 107 D. Mara, *Desalin. Water Treat.*, 2009, **4**, 85–88.
- 108 C. G. Golueke and W. J. Oswald, *Sol. Energy*, 1963, **7**, 86–92.
- 109 C. G. Golueke, W. J. Oswald and H. B. Gotaas, *Appl. Microbiol.*, 1957, **5**, 47–55.
- 110 J. R. Benemann, J. C. Weissman, B. L. Koopman and W. J. Oswald, *Nature*, 1977, **268**, 19–23.
- 111 R. J. Craggs, C. C. Tanner, J. P. S. Sukias and R. J. Davies-Colley, *Water Sci. Technol.*, 2003, **48**, 291–297.
- 112 K. L. Abis and D. D. Mara, *Water Sci. Technol.*, 2003, **48**, 1–7.
- 113 R. J. Craggs, R. J. Davies-Colley, C. C. Tanner and J. P. Sukias, *Water Sci. Technol.*, 2003, **48**, 259–267.
- 114 Y. Nurdogan and W. J. Oswald, *Water Sci. Technol.*, 1996, **33**, 229–241.
- 115 J. J. Milledge and S. Heaven, *Rev. Environ. Sci. Biotechnol.*, 2012, **12**, 165–178.



- 116 N. Uduman, Y. Qi, M. K. Danquah, G. M. Forde and A. Hoadley, *J. Renewable Sustainable Energy*, 2010, **2**, 012701.
- 117 B. T. Higgins and A. Kendall, *J. Ind. Ecol.*, 2012, **16**, 436–447.
- 118 L. Christenson and R. Sims, *Biotechnol. Adv.*, 2011, **29**, 686–702.
- 119 L. E. De-Bashan and Y. Bashan, *Bioresour. Technol.*, 2010, **101**, 1611–1627.
- 120 E. Kebede-westhead, C. Pizarro, W. W. Mulbry and A. C. Wilkie, *J. Phycol.*, 2003, **39**, 1275–1282.
- 121 W. W. Mulbry and A. C. Wilkie, *J. Appl. Phycol.*, 2001, **13**, 301–306.
- 122 W. Mulbry, S. Kondrad, C. Pizarro and E. Kebede-Westhead, *Bioresour. Technol.*, 2008, **99**, 8137–8142.
- 123 R. J. Craggs, *Water Sci. Technol.*, 2001, **44**, 427–433.
- 124 R. J. Craggs, W. H. Adey, K. R. Jenson, M. S. St. John, F. B. Green and W. J. Oswald, *Water Sci. Technol.*, 1996, **33**, 191–198.
- 125 National Academy of Science, Committee on Alternatives and Strategies for Future Hydrogen Production and Use, National Research Council, and National Academy of Engineering, *The Hydrogen Economy: Opportunities, Costs, Barriers, and R&D Needs*, 2004.
- 126 M. R. Brown, S. W. Jeffrey, J. K. Volkman and G. A. Dunstan, *Aquaculture*, 1997, **151**, 315–331.
- 127 A. Richmond and Q. Hu, *Handbook of Microalgal Culture: Applied Phycology and Biotechnology*, Wiley-Blackwell, 2nd edn, 2013.
- 128 A. H. Stouthamer, *Antonie Van Leeuwenhoek Int. J. Gen. Mol. Microbiol.*, 1973, **39**, 545–565.
- 129 C. P. L. Grady Jr, G. T. Daigger, N. G. Love and C. D. M. Filipe, *Biological Wastewater Treatment*, CRC Press, 3 edn, 2012.
- 130 F. B. Metting Jr, *J. Ind. Microbiol.*, 1996, **17**, 477–489.
- 131 P. Spolaore, C. Joannis-Cassan, E. Duran and A. Isambert, *J. Biosci. Bioeng.*, 2006, **101**, 87–96.
- 132 W. F. Owen, *Energy in Wastewater Treatment*, Prentice-Hall, Inc., Englewood Cliffs, 1982.
- 133 USEPA, RDC, ERG, and CHPP, *Opportunities for Combined Heat and Power at Wastewater Treatment Facilities: Market Analysis and Lessons from the Field*, 2011.
- 134 C. G. Golueke and W. J. Oswald, *Appl. Microbiol.*, 1959, **7**, 219–227.
- 135 A. F. Clarens, H. Nassau, E. P. Resurreccion, M. A. White and L. M. Colosi, *Environ. Sci. Technol.*, 2011, **45**, 7554–7560.
- 136 T. M. Brown, P. Duan and P. E. Savage, *Energy Fuels*, 2010, **24**, 3639–3646.
- 137 V. Patil, K.-Q. Tran and H. R. Giselrød, *Int. J. Mol. Sci.*, 2008, **9**, 1188–1195.
- 138 D. R. Vardon, B. K. Sharma, J. Scott, G. Yu, Z. Wang, L. Schideman, Y. Zhang and T. J. Strathmann, *Bioresour. Technol.*, 2011, **102**, 8295–8303.
- 139 G. W. Roberts, M.-O. P. Fortier, B. S. M. Sturm and S. M. Staggs-Williams, *Energy Fuels*, 2013, **27**, 857–867.
- 140 T. D. Reynolds and P. A. Richards, *Unit Operations and Processes in Environmental Engineering*, PWS Publishing Company, 1996.
- 141 G. Tchobanoglous, H. D. Stensel, R. Tsuchihashi and F. L. Burton, *Wastewater Engineering: Treatment and Resource Recovery*, McGraw-Hill Higher Education; McGraw-Hill [distributor], New York; London, 5th edn, 2013.
- 142 E. S. Heidrich, T. P. Curtis and J. Dolfig, *Environ. Sci. Technol.*, 2011, **45**, 827–832.
- 143 C. W. Gellings and K. E. Parmenter, in *Knowledge for Sustainable Development—An Insight into the Encyclopedia of Life Support Systems*, Eloss Publishers, Oxford, UK, 2004, vol. II, pp. 419–450.
- 144 J. S. Guest, M. C. M. van Loosdrecht, S. J. Skerlos and N. G. Love, *Environ. Sci. Technol.*, 2013, **47**, 3258–3267.
- 145 R. J. Geider and J. La Roche, *Eur. J. Phycol.*, 2002, **37**, 1–17.
- 146 N. Powell, A. N. Shilton, S. Pratt and Y. Chisti, *Environ. Sci. Technol.*, 2008, **42**, 5958–5962.
- 147 S. A. Sañudo-Wilhelmy, A. Tovar-Sanchez, F.-X. Fu, D. G. Capone, E. J. Carpenter and D. A. Hutchins, *Nature*, 2004, **432**, 897–901.
- 148 R. Craggs, D. Sutherland and H. Campbell, *J. Appl. Phycol.*, 2012, **24**, 329–337.
- 149 I. Woertz, A. Feffer, T. Lundquist and Y. Nelson, *J. Environ. Eng.*, 2009, **135**, 1115–1122.
- 150 I. Vyrides and D. C. Stuckey, *Water Res.*, 2009, **43**, 933–942.
- 151 I.-S. Chang and S. J. Judd, *Process Biochem.*, 2002, **37**, 915–920.
- 152 R. Yoo, J. Kim, P. L. McCarty and J. Bae, *Bioresour. Technol.*, 2012, **120**, 133–139.
- 153 G. Yu, Y. Li, G. Shen, W. Wang, C. Lin, H. Wu and Z. Chen, *J. Appl. Phycol.*, 2009, **21**, 719–727.
- 154 C. Gudín and D. Chaumont, *Bioresour. Technol.*, 1991, **38**, 145–151.
- 155 J. Kim, K. Kim, H. Ye, E. Lee, C. Shin, P. L. McCarty and J. Bae, *Environ. Sci. Technol.*, 2011, **45**, 576–581.
- 156 A. M. Illman, A. H. Scragg and S. W. Shales, *Enzyme Microb. Technol.*, 2000, **27**, 631–635.
- 157 J. Ma and O. Hemmers, *J. Energy Resour. Technol.*, 2011, **133**, 011801.
- 158 D. R. Vardon, B. K. Sharma, G. V. Blazina, K. Rajagopalan and T. J. Strathmann, *Bioresour. Technol.*, 2012, **109**, 178–187.
- 159 P. E. Savage, *Science*, 2012, **338**, 1039–1040.
- 160 A. A. Peterson, F. Vogel, R. P. Lachance, M. Fröling, M. J. Antal Jr and J. W. Tester, *Energy Environ. Sci.*, 2008, **1**, 32–65.
- 161 A. Hospido, M. Carballa, M. Moreira, F. Omil, J. M. Lema and G. Feijoo, *Water Res.*, 2010, **44**, 3225–3233.
- 162 H. Wang, S. L. Brown, G. N. Magesan, A. H. Slade, M. Quintern, P. W. Clinton and T. W. Payn, *Environ. Sci. Pollut. Res. Int.*, 2008, **15**, 308–317.
- 163 B. Sialve, N. Bernet and O. Bernard, *Biotechnol. Adv.*, 2009, **27**, 409–416.
- 164 USDOE, *National algal biofuels technology roadmap*, Office of Energy Efficiency and Renewable Energy, Biomass Program, 2010.
- 165 J. B. K. Park, R. J. Craggs and a. N. Shilton, *Water Res.*, 2011, **45**, 6637–6649.



- 166 B. Wett, K. Buchauer and C. Fimml, *Energy self-sufficiency as a feasible concept for wastewater treatment systems*, Singapore, 2007, pp. 21–24.
- 167 WHO and UNICEF, *Progress on Sanitation and Drinking-water – 2013 Update, WHO/UNICEF Joint Monitoring Programme for Water Supply and Sanitation Report*, WHO, Office of Publication, France, 2013.
- 168 R. Baum, J. Luh and J. Bartram, *Environ. Sci. Technol.*, 2013, **47**, 1994–2000.
- 169 B. Heffernan, J. B. van Lier and J. van der Lubbe, *Water Sci. Technol.*, 2011, **63**, 100–107.
- 170 F. El Hafiane and B. El Hamouri, *Water Sci. Technol.*, 2005, **51**, 125–132.
- 171 L. Seghezzi, A. P. Trupiano, V. Liberal, P. G. Todd, M. E. Figueroa, M. A. Gutiérrez, A. C. D. S. Wilches, M. Iribarnegaray, R. G. Guerra, A. Arena, C. M. Cuevas, G. Zeeman and G. Lettinga, *Appl. Biochem. Biotechnol.*, 2003, **109**, 167–180.
- 172 W. M. K. R. T. W. Bandara, H. Satoh, M. Sasakawa, Y. Nakahara, M. Takahashi and S. Okabe, *Water Res.*, 2011, **45**, 3533–3540.
- 173 M. M. Benjamin, *Water Chemistry*, Waveland Press, Long Grove, Illinois, 2010.
- 174 P. Kuntke, K. M. Śmiech, H. Bruning, G. Zeeman, M. Saakes, T. H. J. A. Sleutels, H. V. M. Hamelers and C. J. N. Buisman, *Water Res.*, 2012, **46**, 2627–2636.
- 175 P. Kuntke, M. Geleji, H. Bruning, G. Zeeman, H. V. M. Hamelers and C. J. N. Buisman, *Bioresour. Technol.*, 2011, **102**, 4376–4382.
- 176 J. Desloover, A. Abate Woldeyohannis, W. Verstraete, N. Boon and K. Rabaey, *Environ. Sci. Technol.*, 2012, **46**, 12209–12216.
- 177 R. D. Cusick and B. E. Logan, *Bioresour. Technol.*, 2012, **107**, 110–115.
- 178 R. D. Cusick, M. L. Ullery, B. A. Dempsey and B. E. Logan, *Water Res.*, 2014, **54**, 297–306.
- 179 C. Forrestal, P. Xu and Z. Ren, *Energy Environ. Sci.*, 2012, **5**, 7161–7167.
- 180 G. T. Daigger, *Water Sci. Technol.*, 2011, **63**, 516–526.
- 181 K. J. Flynn, *Limnol. Oceanogr.*, 2003, **48**, 2243–2249.
- 182 M. Pham, L. Schideman, J. Scott, N. Rajagopalan and M. J. Plewa, *Environ. Sci. Technol.*, 2013, **47**, 2131–2138.

