

Carbon monoxide – physiology, detection and controlled release

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Carbon monoxide (CO) is increasingly recognized as a cell-signalling molecule akin to nitric oxide (NO). CO has attracted particular attention as a potential therapeutic agent because of its reported anti-hypertensive, anti-inflammatory and cell-protective effects. We discuss recent progress in identifying new effector systems and elucidating the mechanisms of action of CO on, e.g., ion channels, as well as the design of novel methods to monitor CO in cellular environments. We also report on recent developments in the area of CO-releasing molecules (CORMs) and materials for controlled CO application. Novel triggers for CO release, metal carbonyls and degradation mechanisms of CORMs are highlighted. In addition, potential formulations of CORMs for targeted CO release are discussed.

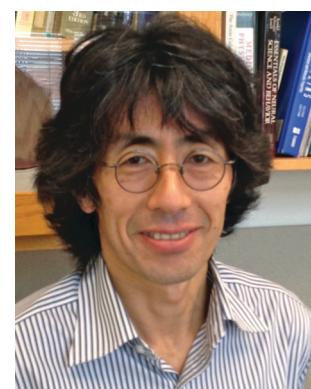
A. Introduction

Carbon monoxide (CO), a colour- and odourless gas, is typically produced when carbon-containing compounds are only partially oxidized, such as in combustion engines. At high concentrations, CO is toxic to humans and animals. It binds to haemoglobin about 200-times more strongly than O₂, and the resulting carboxyhaemoglobin is thus no longer available for the oxygen transport in the body. With 50% of human haemoglobin occupied by CO, seizures and coma may result,



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sometimes with fatal consequences. The major cause of its toxicity, however, seems to originate from its distribution in the tissue where it affects various targets.¹ However, despite this toxicity, CO is increasingly appreciated as a cell-signalling molecule akin to nitric oxide (NO).² For example, CO relaxes smooth muscles at low concentrations and lowers the blood pressure.³

The major endogenous source of CO in the body is breakdown of haem. Physiological degradation of haem occurs in a tightly controlled manner involving the enzyme haem oxygenase (HO; EC 1.14.99.3), which cleaves the porphyrin IX in the presence of NADP and molecular oxygen, resulting in the

primary or first-order haem degradation products (HDPs), consisting of CO, Fe²⁺ and biliverdin IX (Fig. 1).⁴ Biliverdin reductase catalyzes the next degradation step in the presence of NADPH and H⁺, yielding the second-order metabolite bilirubin IX, which is then excreted in bile and urine. The bile pigments biliverdin (green) and bilirubin (yellow) are easily recognized in haematomas, which change colour over time as haem degradation proceeds. CO production following this route amounts to about 16 $\mu\text{mol h}^{-1}$ per human body.⁵

The molecular action of CO, however, is to be considered a local event because of its limited bioavailability and the dependence of its production on haem oxygenases. In humans, there

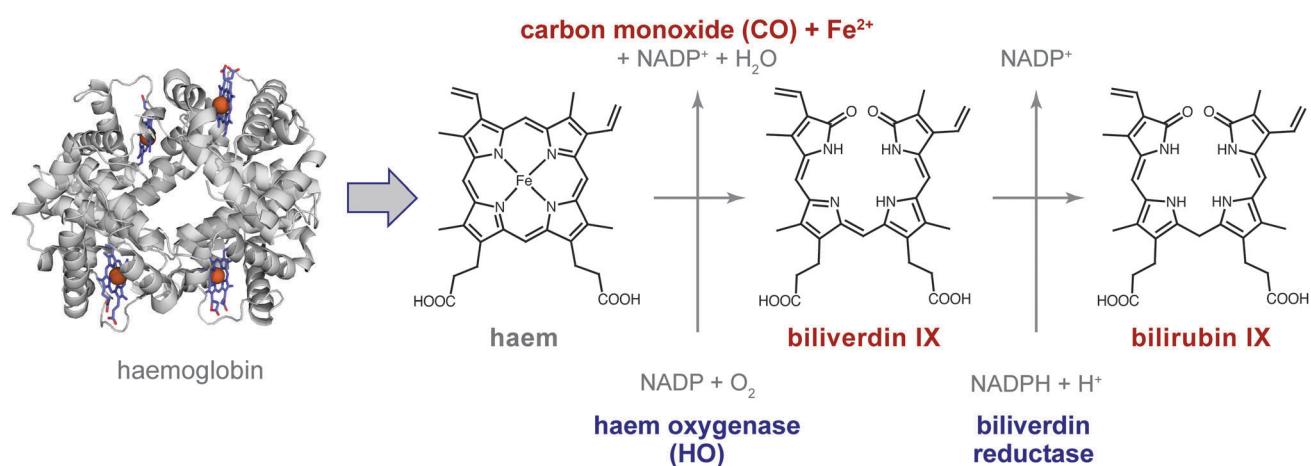


Fig. 1 Haem degradation. Haem (Fe²⁺-protoporphyrin IX), released from haemoglobin (left), is degraded by the aid of haem oxygenase to carbon monoxide (CO), ferrous ions (Fe²⁺) and biliverdin IX. A subsequent step, catalysed by biliverdin reductase, yields bilirubin IX.



Matthias Westerhausen

in 1994. From 1996 to 2004 he was professor at the Ludwig-Maximilians University (LMU) in Munich where he was also vice-president from 2001 to 2004. Since 2004 he has been teaching and researching at the Friedrich Schiller University Jena, Germany. His main interests include topics such as the organic chemistry and catalytic application of d^0 metals with the focus on heavy alkaline earth metals, cooperative effects in heterobimetallic complexes. He is also enjoying the interdisciplinary research on haem degradation products and CO-releasing molecules within the DFG research unit FOR 1738.



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are two isoforms of this enzyme. The expression of HO1 is inducible, triggered by the presence of free haem, thus signalling the need for haem degradation. In contrast, HO2 is constitutively expressed. Since the activity of HOs always results in the clearance of haem and the production of CO, in many cases it is not unambiguously clear whether the removal of haem or the release of the haem degradation products, such as CO, or both, are the primary physiologically relevant end results. In any case, ample evidence has accumulated to demonstrate that haem catabolism and the endogenous production of CO serve a wide array of physiological functions. For example, induction of HO1 in the brain improves outcome after cerebral ischemia^{6,7} and HO2 was shown to be neuroprotective during intracerebral haemorrhage,⁸ suggesting that either the clearance of haem or the production of CO (or both) is beneficial. This becomes particularly clear in cerebral malaria, a multifactorial disease induced by cerebral accumulation of haemoglobin *via* *Plasmodium* infection, claiming more than 1 million lives annually, most of them being children: HO1 and CO were shown to be beneficial, HO1 because it removes free haem and CO because it binds to haemoglobin and, therefore, inhibits haem release.^{9,10}

CO has attracted particular attention as a potential therapeutic agent because CO is suggested to have anti-hypertensive, anti-inflammatory and cell-protective effects.^{3,11–14} For example, inhalation of CO gas under controlled conditions alleviates symptoms of human pulmonary hypertension,¹⁵ presumably by interacting with the smooth muscle signalling proteins such as guanylyl cyclase and potassium channels. CO inhalation also appears to protect vital organs, including the brain, heart, lung, and liver, during ischemia/hypoxia and organ transplantation,^{16–19} although the underlying mechanisms remain unknown. Consistent with the postulated beneficial role of CO, higher expression of HO leads to a better outcome for patients after a septic shock.²⁰

However, the practical clinical use of CO gas is currently hampered severely. Owing to the relatively low solubility of CO in water (about 1 mM), its partitioning to body fluids and target tissues is rather limited. To reach appreciable concentration inside the body, high concentrations of CO gas would need to be inhaled. Furthermore, the potential interaction of CO with various molecules in physiological environments complicates a targeted administration as well as research on its physiological functions.³ Therefore, various methods have been devised to deliver CO precisely to the target locations without off-target effect. CO-releasing molecules (CORMs) that can be targeted to specific sites in the body and locally liberate CO are thus urgently needed for research and possibly for clinical applications. Furthermore, in most cases reported beneficial effects of CO on organ function or whole organisms are not understood in molecular detail, specifically lacking insight into the molecular target sites and mechanisms by which CO interacts with biomolecules to alter their function. Here we will therefore review recent advances in studying physiological effector sites of CO as well as novel approaches for the synthesis and use of CO-releasing molecules and materials. Both aspects suffer from

a common limitation, namely elucidation of when and where in the body CO is present to exert its activity. Therefore, we will also address new promising methods for the monitoring of CO in living cells.

Several excellent review articles have been published on topics of the physiology and controlled release of CO.^{21,22} A recent tutorial review discusses features for developing drug molecules for therapy with CO.²³ Two novel fluorescent probes for monitoring CO in living cells have been introduced.²⁴ In addition, emerging concepts on the anti-inflammatory actions of CORMs have been shown.^{3,12} A perspective on CORMs was written by Mann.^{25,26} Reviews have been published on photo-activated CORMs (photoCORMs).^{27–29} Potential photoactivated metallopharmaceuticals, such as active molecules and supported drugs, have been featured in this journal.³⁰

B. Effector systems

It is conventionally thought that transduction of CO by biological molecules requires a cofactor. While some prokaryotic oxygenases and oxidases without any prosthetic group or metal cofactor are capable of interacting with O₂,^{31,32} for CO the presence of reduced iron (Fe²⁺), typically haem iron, is considered to be essential ("haem-based CO sensors").² Numerous haem-containing proteins are present in a typical cell,³³ rendering the number of potential direct and indirect effectors of CO or CO-sensitive components exceedingly large. The potential effectors include cell-signalling enzymes such as transcription factors, some of which take part in regulation of circadian rhythm, cystathione β -synthase involved in H₂S production, guanylyl cyclase and ion channel proteins.^{21,22,33} It is not surprising then that experimental application of exogenous CO (see Section A) has been reported to induce multitudes of effects. Selected putative CO effectors – focusing on those in mammalian systems – are discussed below to highlight diverse mechanisms of regulation by CO. Other examples, such as ion channel regulation in mammalian cells, are found in studies of Wilkinson and Kemp³⁴ and Peers.³⁵ For more discussion of CO-sensitive microbial proteins including the CO-sensing transcription factor from *Rhodospirillum rubrum* (CooA) and CO dehydrogenase (CODH), readers are referred to Roberts *et al.*,³⁶ Gullotta *et al.*,²² and Boer *et al.*³⁷

Neuronal PAS domain 2 (NPAS2) transcription factor

Circadian control of cell function involves multiple transcription factors.³⁸ Among them, NPAS2, expressed in the brain, contains a basic helix-loop-helix (bHLH) DNA binding domain and two PAS (Per-Arnt-Sim; PAS-A and PAS-B) domains, and each PAS domain is capable of binding haem.³⁹ NPAS2 forms a heterodimer complex with another protein and binds to its target DNA sequence. An *in vitro* study suggests that the dimer formation and the DNA binding activity of NPAS2 with haem bound is inhibited by μ M levels of CO.³⁹ Raman spectroscopy measurements indicate CO impairs the histidine-mediated ligation of the reduced haem



iron.⁴⁰ How the altered haem-ligation inhibits the dimer formation and DNA binding is not known.

Guanylyl cyclase, nitric oxide synthase and CO

The enzyme soluble guanylyl cyclase (sGC; EC 4.6.1.2) is a haem-containing heterodimeric complex (typically $\alpha 1/\beta 1$) that catalyzes conversion of GTP to cGMP, an intracellular messenger that initiates a variety of physiological responses.^{41–43} Each subunit possesses multiple domains, from the N to C termini: an H-NOX domain (haem-nitric oxide/oxygen binding domain), a PAS domain, a CC (coiled-coiled) domain and a CAT (catalytic) domain.⁴³ The activity of sGC under reducing conditions is markedly enhanced, more than 100-fold, by the gaseous messenger NO^{44,45} and this regulation accounts for the well-known vasodilatory influence of NO.⁴⁶ CO has been reported to increase the sGC activity albeit much less effectively than NO,^{45,47} approx. 3- to 4-fold.^{44,45} It may be noted that such an increase in the enzymatic activity by CO was not observed in a separate study.⁴⁸ O₂, present at a higher concentration than NO, in contrast, has no effect on sGC.^{47,49} The postulated CO-mediated increase in the sGC activity may contribute in part to the vasodilatory action of low concentrations of CO.^{50,51} The structural and physicochemical bases of the interactions of NO and CO with sGC have been studied.⁵² The H-NOX domain of each β subunit of sGC may contain a haem whose propionate groups are surrounded by the conserved sequence motif Tyr-X-Ser-X-Arg where X is any amino acid (Fig. 2A).⁵³ The reduced haem iron centre is coordinated by a His residue. This results in a pentacoordination mode. A single NO molecule binds to the distal side of the haem iron (at least transiently), creating a hexacoordinated haem (Fig. 2B). The fully activated state of sGC is pentacoordinated⁴⁴ in which the original proximal His ligand is displaced.⁵² Like NO, a CO molecule also binds to the distal side of the haem iron (Fig. 2C). The atomic structural changes induced by binding of NO or CO are subtle (Fig. 2); a very slight pivoting of the haem cofactor (<1 Å) has been suggested to account for the enhanced enzymatic activity.⁵² In addition, a change in the hydrogen bonding network around the haem cofactor has been proposed to account for the contrasting effects of NO and O₂ on sGC.⁴⁹ Additional insights into the conformational changes induced by NO and CO in sGC may be

inferred from the results using 3-(5'-hydroxymethyl-2'-furyl)-1-benzylindazole (YC-1).^{45,54} YC-1 allosterically increases the sGC activity; in particular, in the presence of YC-1, CO is capable of markedly increasing the catalytic activity essentially to the same level as that by NO.^{45,55} It has been hypothesized that YC-1 binds to the H-NOX domain and alters the interaction between the H-NOX and PAS domains.⁵⁶ A naturally-occurring compound that acts like YC-1 has not been reported. Albeit much less effective than NO, activation of sGC by CO has potential to cause an increase in cGMP concentration, which in turn activates cGMP-dependent protein kinase (PKG). PKG could then influence a plethora of signalling effectors – hence CO may exert numerous but indirect downstream effects.

The synthesis of NO from L-arginine by nitric oxide synthases (NOSs), haem-containing proteins, is also subject to regulation by CO and NO;⁵⁷ these gaseous regulators inhibit the catalytic activity, potentially providing a negative feedback mechanism. A structural study suggests that CO binds to the distal side of the haem iron centre near the ligand L-arginine.⁵⁸ While the changes in the overall structure of the enzyme induced by CO binding are again subtle, the presence of a CO near the haem iron centre and the ligand L-arginine is speculated to alter the local hydrogen bonding network, leading to inhibition of the NOS catalytic activity.⁵⁸

CO binding without haem in microbial enzymes

Selected microbial proteins interact with CO in a haem-independent manner.^{37,59} For example, CODH (EC 1.2.99.2), which catalyzes oxidation of CO to CO₂, contains no haem but each subunit in the dimeric enzyme complex forms multiple metalloclusters with copper, nickel, iron, and/or sulphur.³⁷ Crystal structures of CODH suggest that CO interacts with the nickel ion in one of the metalloclusters ("C-cluster").^{60,61} One could speculate that CO binds in particular to iron-sulfur clusters from CODH, which are probably as abundant in cells of higher organism as haem.⁶² Whether or not similar metallocluster-dependent CO binding occurs in mammals has not been determined. Certainly, the identification of molecular entities capable of binding CO and thus functioning as cellular CO buffer systems will be an important task for future studies.

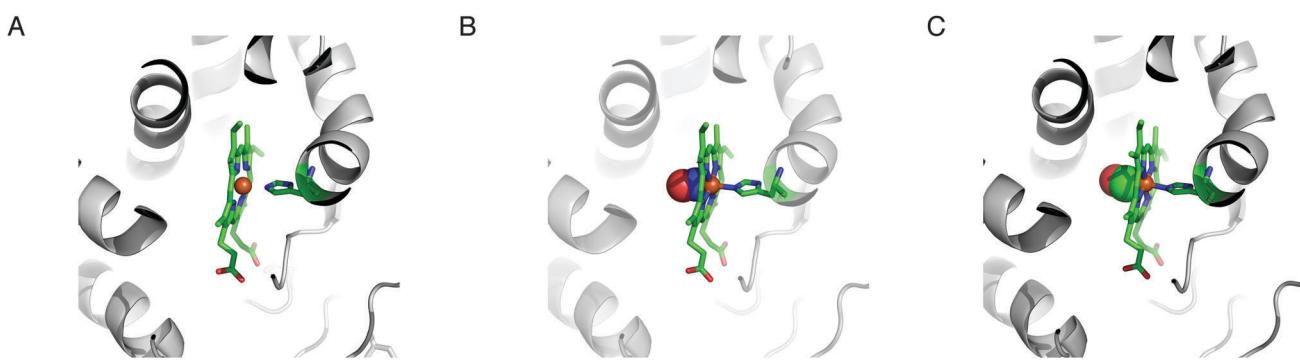


Fig. 2 CO binding to soluble guanylyl cyclase. Haem coordination by soluble guanylyl cyclase (A, PDB ID 2O09) with an unoccupied coordination site, in the presence of NO (B, PDB ID 2O0C), and in the presence of CO (C, PDB ID 2O0G). The images were rendered using MacPyMol v0.99.



Large-conductance Ca^{2+} - and voltage-gated K^+ channels and CO

CO relaxes isolated vascular smooth muscle cells in the absence of endothelial cells.⁶³ A consensus exists that CO (somehow) stimulates large-conductance Ca^{2+} - and voltage-gated K^+ channels in vascular smooth muscle cells. These K^+ channels, which are also known as BK ("big potassium"), maxi K and Slo1 channels, mediate gated fluxes of K^+ ions according to their electrochemical gradient.⁶⁴ Opening of the "gate" of the channel to allow K^+ flux is allosterically facilitated by binding of Ca^{2+} to its Ca^{2+} sensors ("RCK1 Ca^{2+} sensors" and "RCK2 Ca^{2+} sensors") in the cytoplasmic domain and/or by depolarization-mediated activation of its transmembrane voltage-sensor domains (VSDs) (Fig. 3A and B). Greater activation of BK channels helps to stabilize the membrane potential at a negative level, which in turn prevents opening of depolarization-activated Ca^{2+} channels, thereby inhibiting cellular activation. In this manner, stimulation of BK channels in smooth muscle cells by various cellular signalling molecules such as CO exerts a vaso-dilatory influence.^{65–67}

In all likelihood, CO activates BK channels in multiple ways. The evidence is consistent with the idea that phosphorylation of the BK channel by PKG at selected cytoplasmic Ser residues (Ser855 and Ser869⁶⁸ and Ser1072⁶⁹ in human Slo1 AAB65837) increases the overall probability that the channel gate is open. Exactly how phosphorylation of these Ser residues in the cytoplasmic domain alters the energetics of the ion conduction gate in the transmembrane domain located several nanometres away is unclear.^{70–72} Because CO stimulates sGC, albeit weakly, leading to activation of PKG (see above), it is expected that CO promotes PKG-mediated phosphorylation of BK channels and then increases the channel activity. CO is also reported to stimulate another K^+ channel type (TREK1) through activation of the cGMP/PKG pathway.⁷³

Additionally, CO has been postulated to directly alter the BK channel activity in a cell-signalling cascade-independent fashion. Application of CO itself and generation of CO by CORM-2 (Fig. 7) increase the channel open probability in excised membrane patches where the normal intracellular signalling network is disrupted.^{75–79} According to the idea that the interaction with CO requires a reduced cofactor, the sensitivity of the BK channel to CO and CORM application in cell-free patches suggests the protein itself may harbour a tightly bound cofactor. Structural studies show that some ion channel proteins do contain structurally and functionally important Zn^{2+} .^{80,81} An atomic structure of the complete BK channel protein is not yet available. However, the structures of the cytoplasmic domain of the BK channel (see Fig. 3C) expressed in insect cells at resolutions of approx. 3.0 Å show no redox-sensitive cofactor.^{70–72} How could the BK channel apparently without a cofactor respond to CO? This remains an open question.

Some circumstantial clues are available. The enhancement of the channel activity by CO is observed using low Ca^{2+} solutions chelated by mM concentrations of EGTA or EDTA⁷⁷ (but see ref. 78). These chelators have higher affinities for multivalent cations like iron, copper, manganese and cobalt than for calcium. Thus CO stimulates the BK channel activity even when the concentrations of these free multivalent cations are negligible. The CO action was also observed following pretreatment of the channel with the oxidizing agent H_2O_2 ,⁷⁷ solvent exposed oxidation-prone cofactors, if present, may not be essential. However, cyanide at μM levels abolished the response to CO (cyano metal complexes often have critical stability constants $\log K > 30$).⁷⁹ This observation may suggest that the BK channel protein contains a metal cofactor. However, as noted above, such a tightly-bound cofactor is not

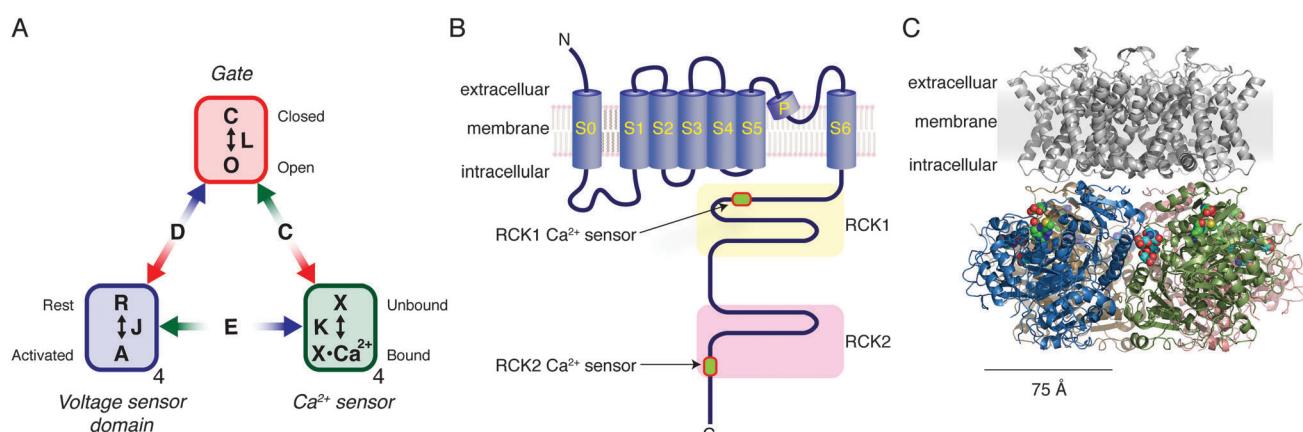


Fig. 3 A model of the Slo1 BK channel function and the structure. (A) Allosteric model of the Slo1 BK channel function.⁷⁴ The ion conduction gate can be closed or open as specified by the equilibrium constant L . Each of the four voltage-sensor domains could be at rest or activated as specified by the equilibrium constant J . Each of the Ca^{2+} sensors could be unbound or bound as specified by the constant K . The model lumps the two Ca^{2+} sensors of each Slo1 subunit into one functional unit. The allosteric interactions are specified by the constants D , C and E . (B) Structural organization of each of the four Slo1 subunits in a functional Slo1 BK channel. Each polypeptide is about 1100 residues long. (C) Probable structure of a functional Slo1 BK channel. The grey transmembrane domain is a homology model based on PDB ID 2R9R and the cytoplasmic domain is from PDB ID 3NAF. In the cytoplasmic domain, each subunit is shown using a different colour. The Ca^{2+} ligand residues are displayed using spheres. The images were rendered using MacPyMol v0.99.

detected in the available structures of the BK channel cytoplasmic domain.^{70–72} If the BK channel contains a cofactor, it may be present in the unstructured areas of the channel protein such as the cytoplasmic RCK1–RCK2 linker segment and the cytoplasmic distal C terminus unresolved in the available structures. It is also possible that the BK channel protein itself does not contain any cofactor but it may intimately associate with a separate cofactor-containing structural component in select cells, perhaps forming a macromolecular complex. The idea that BK channels participate in formation of macromolecular complexes has received some experimental support.^{76,82} Clearly, elucidation of the mechanism of CO sensing by the BK and other channels requires further study. Functional eukaryotic channel complexes including BK channels have been difficult to express and purify in large quantities. This limitation has severely hampered the progress in elucidation of the physicochemical mechanism of CO sensing in ion channel proteins, precluding application of more direct metal assay techniques such as inductively coupled plasma mass spectrometry. A high-resolution structure of the whole-channel complex in a native environment will be also helpful.

The stimulatory effect of CO is diminished by high concentrations of intracellular Ca^{2+} ^{77,78} or low intracellular H^+ .⁸³ Therefore, the conformational changes in the BK channel protein induced by CO may resemble those by Ca^{2+} and H^+ .^{77,83,84} CO increases the gate open probability without requiring activation of the VSDs.⁸³

Chemical modification and mutagenesis studies implicate two specific His residues in the channel's response to CO.^{75,77} Pretreatment of the channel with the imidazole modifier DEPC^{75,77} or mutation of His365 and His394 located near one of the two Ca^{2+} sensors ("RCK1 Ca^{2+} sensor"; Fig. 3B) in the BK channel protein^{70–72,77,79} at low intracellular Ca^{2+} concentrations markedly impairs the channel's response to CO and CORM-2 (50 μM). However, application of high concentrations of CORM-2 (e.g., 300 μM) stimulates the channel even when His365 and His394 are absent,⁷⁹ suggesting that His365 and His394 may preferentially mediate the channel's sensitivity to low concentrations of CO. The exact functional roles played by His365 and His394 are not yet known. The two His residues could somehow directly interact with CO in a novel way or couple the information regarding the interaction of CO with its sensor located elsewhere to the ion conduction gate. In addition to His365 and His394, Cys911 is involved in the channel's response to CO. Cys911 is located near the RCK2 Ca^{2+} sensor (Fig. 3B)^{70–72} and plays an important role in the oxidative sensitivity of the Ca^{2+} sensing of the channel.^{85,86} Mutation of Cys911 diminishes the channel's response to CORM-2; the dose dependence results suggest that the mutation preferentially decreases the efficacy of CO without altering the half-maximal effective concentration.⁷⁹

In summary, the stimulatory effect of CO on the BK channel, not surprisingly, involves multiple amino-acid residues: at least, two His residues (His365 and His394) near the RCK1 Ca^{2+} sensor and a Cys residue (Cys911) near the RCK2 Ca^{2+} sensor. Whether these residues comprise the CO sensor(s) of

the channel or the coupling mechanism connecting the ultimate effector, the gate of the channel, and the CO sensor elsewhere in the channel or even in a different intimately associated protein remains unclear. Histidine and cysteine can complex with metal ions but the atomic structures of the cytoplasmic domain of the channel^{70–72} do not reveal any metal bound to His365, His394 or Cys911.

Local action of CO on BK channels

CO is potentially capable of altering functions of numerous haem-containing enzymes. The uncontrolled and global action of CO would be detrimental to normal cell function. How could CO target specific effectors, such as BK channels? The key concept is the local action of CO. CO is primarily generated by HO1 and HO2 from haem in an oxygen- and NADPH-dependent manner (Fig. 1). Thus, the molecules in the close vicinity of HOs are expected to be more readily influenced by CO. In selected cells, Slo1 channels are reported to exist co-localized with HO2 so that small quantities of CO could have a significant and rapid impact on the channel function without altering other proteins.⁷⁶ Under normoxia, HO2 provides a basal stimulatory influence on nearby BK channels *via* CO. With a decrease in oxygen tension (hypoxia), the CO generation by HO2 is compromised and the stimulatory influence diminishes. In this manner, CO is an important mediator of hypoxia sensing in carotid body glomus cells⁷⁶ although other mechanisms are operative.^{87,88}

A similar mechanism of O_2 -dependent regulation involving CO and haem together is observed in voltage-independent trimeric Na^+ channels⁸⁹ expressed widely in epithelial cells (ENaCs).⁹⁰ Application of haem and NADPH, both of which are required for the HO to produce CO, stimulates the channel. CORM-2 also stimulates the channel activity. In contrast, haem applied under hypoxic conditions inhibits the channel activity. The molecular domain of ENaC responsible for ligation of haem is not known and whether ENaC and HO colocalise has not been established.

As illustrated in the examples discussed earlier, haem is a stable cofactor in numerous proteins, often conferring gas sensitivity to the parent proteins. While the concentration is low, free haem does exist in cells.⁹¹ Free haem at nanomolar levels is capable of acutely and directly regulating the BK channel activity.⁹² The relatively unstructured cytoplasmic RCK1–RCK2 linker region (Fig. 3B) coordinates haem using His and Cys residues.^{92–95} Addition of exogenous free haem to the channel weakens the allosteric coupling between the VSDs and the ion conduction gate.⁹⁶ Consequently, the gate open probability is higher at negative voltages and lower at more positive voltages.⁹⁶ This modulatory action of exogenous free haem is diminished by CO and may represent another mechanism by which CO alters the channel function.⁹³

Voltage-gated Ca^{2+} channels, CO and ROS

Yet another mechanism by which CO influences its effectors is through generation of reactive oxygen species (ROS). CO binds to the mitochondrial complex IV enzyme cytochrome c oxidase



and increases the mitochondrial production of ROS.^{97,98} ROS can in turn alter many proteins including voltage-gated Ca^{2+} channels.⁹⁹

Both NO and CO are excellent ligands for haem in sGC with similar binding behaviours (Fig. 2B and C). The high binding affinities to iron porphyrins have been also used in the detection of NO and CO.¹⁰⁰ However, elucidation of the physiological roles of CO demands for improved analytical methods of CO sensing.^{101–103} The following section addresses recent advances in this rapidly developing field.

C. CO detection

Established methods for CO detection include gas chromatography,¹⁰⁴ laser infrared absorption¹⁰⁵ and electrochemical assays.^{101,106} In addition, colourimetric CO sensing is an important alternative,^{103,107} in particular if the aim is to monitor CO in living cells, organs or whole organisms.

The standard *in vitro* test for CO is the carboxy-myoglobin (Mb-CO) assay, where CO is added to a solution of reduced deoxy-Mb and the formation of Mb-CO is followed *via* absorbance spectroscopy. Optical changes on CO binding in the α or β regions or in the Soret band are detected.¹⁰⁸ Using this method, striking discrepancies in the CO release rates of $(\text{Ru}(\text{CO})_3\text{Cl}_2)_2$ (CORM-2) and $[\text{Ru}(\text{CO})_3\text{Cl}(\text{glycinate})]$ (CORM-3, see Fig. 7) have been reported.¹⁰⁹ Subsequently, it was shown that the reducing agent sodium dithionite is responsible for rapid CO release from Ru-based CORMs in the Mb-CO assay. In addition, dithionite also enhances CO release rates from other CORMs.¹⁰³ To overcome the problem, two novel protocols have been recently introduced. Oxy-haemoglobin¹⁰³ and a modified myoglobin assay¹¹⁰ have been used for reliable determination of CO-release rates from organometallic carbonyl complexes.

It was recently shown that CO can be detected selectively with high sensitivity by the binuclear rhodium complex *cis*- $[\text{Rh}_2(\text{C}_6\text{H}_4\text{PPh}_2)_2(\text{O}_2\text{CCH}_3)_2](\text{HAc})_2$.¹¹¹ It contains two cyclometalated phosphine ligands and undergoes a colour change from violet to orange *via* replacement of an acetic acid (HAc) molecule by CO (Fig. 4). In addition, replacement of the second HAc by CO induces a colour change to yellow.¹¹² A collection of binuclear rhodium complexes displayed high CO selectivity with remarkable detection limits. For example, $[(\text{Rh}_2((m\text{-CH}_3\text{C}_6\text{H}_3)\text{P}(m\text{-CH}_3\text{C}_6\text{H}_4)_2)_2(\text{O}_2\text{CCH}_3)_2](\text{HAc})_2$, adsorbed on silica gel, is capable of CO detection by the “naked eye” at concentrations as low as 0.2 ppm in air.¹¹² Furthermore, the binding of CO in all rhodium complexes was fully reversible.

Unfortunately, the organometallic dirhodium compounds are only soluble in organic solvents, such as chloroform. Thus, these complexes are not useful for the real-time detection of physiological levels of CO inside living cells. In this context, fluorescence sensing and imaging has emerged as one of the most powerful techniques to monitor the concentration, localization and even movements of biomolecules in living systems.^{113,114} A variety of fluorescent probes for other small signalling molecules, such as NO and hydrogen sulfide,¹¹⁵ have



Fig. 4 Colourimetric detection of CO via binuclear rhodium complexes. (A) Basic structure of the complexes ($\text{HAc} = \text{CH}_3\text{CO}_2\text{H}$). (B) The photograph shows $[(\text{Rh}_2((m\text{-CH}_3\text{C}_6\text{H}_3)\text{P}(m\text{-CH}_3\text{C}_6\text{H}_4)_2)_2(\text{O}_2\text{CCH}_3)_2]\cdot(\text{CH}_3\text{CO}_2\text{H})_2$ adsorbed on silica gel in the absence (left) and presence of 8 ppm (middle) and 2000 ppm (right) of CO. Adapted and reprinted with permission from ref. 112. Copyright 2011 American Chemical Society.

been already deployed in biology.¹¹⁶ Recently, the development of fluorescent probes for CO has experienced a boost with a biosensor¹¹⁷ and an organometallic palladium complex probe.¹¹⁸ Although these two approaches appear distinct, the fundamental design strategy is similar. In both cases the strong binding affinity of CO to transition metal ions is exploited.²⁴

The palladium probe is able to detect CO in living cells based on metal-mediated carbonylation chemistry (Fig. 5).¹¹⁸ The cyclopalladated species COP-1 quenches the fluorescence of the borondipyrromethene difluoride (BODIPY) core *via* heavy-atom electronic effects. Upon binding of CO, a carbonylation reaction concomitantly releases $\text{Pd}(0)$ and a BODIPY dye with high fluorescence intensity. A 10-fold fluorescence enhancement was observed only in the presence of CO compared with biologically relevant reactive oxygen, nitrogen and sulphur species. The fluorescence intensity enhancement is concentration dependent with a detection limit of 1 μM of CO. The nontoxic and biocompatible palladium-based probe allows CO monitoring in living cells. However, the response time is about an hour to reach the highest level of fluorescence enhancement.

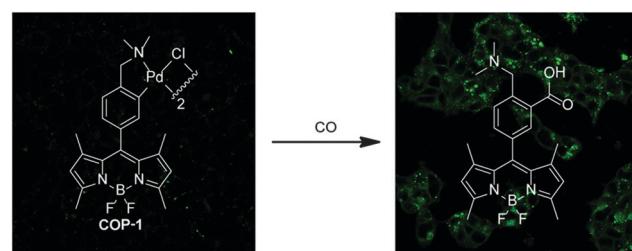


Fig. 5 Confocal microscopy images of the turn-on fluorescent probe for selective CO detection based on palladium-mediated carbonylation reactivity (COP-1). The organometallic probe is capable of detecting CO both in aqueous buffer and in living HEK293T cells with high selectivity. Left: cells incubated with COP-1 for 30 min. Right: cells incubated with 50 μM CORM-3 and 1 μM COP-1. Adapted and reprinted with permission from ref. 118. Copyright 2012 American Chemical Society.



The other approach utilizes the interaction of CO with an iron-containing haem protein as exemplified in the detection mechanism of the fluorescent probe COSer (Fig. 6).¹¹⁷ This biosensor uses a yellow fluorescent protein (YFP) as the fluorescent reporter and CooA, a dimeric CO-sensing haem protein from *Rhodospirillum rubrum*. Upon treatment with 10 μM CO for 10 minutes, the probe exhibits a small two-fold fluorescence intensity enhancement. This observation was attributed to the conformational change of CooA upon binding to CO. The probe showed good selectivity for CO against other relevant haem binding ligands, such as H_2S , GSH, NO, O_2 , CN^- and imidazole. The probe COSer is able to monitor CO fluctuations inside living HeLa cells.

The properties of the genetically encoded probe COSer and small-molecule probe COP-1 have been critically assessed.²⁴ The COP-1 probe displays a larger fluorescence signal enhancement (10-fold) than the genetically encoded fluorescent probe COSer (2-fold) *in vitro*. However, the biosensor COSer is faster than COP-1 in the response time. Another striking difference is given by the reversibility of the sensors. While the interaction of the COSer probe with CO is reversible, that of COP-1 is irreversible. Thus, the COSer is better suited for real-time detection of CO, while COP-1 is more appropriate for monitoring low concentrations of CO because of signal accumulation.

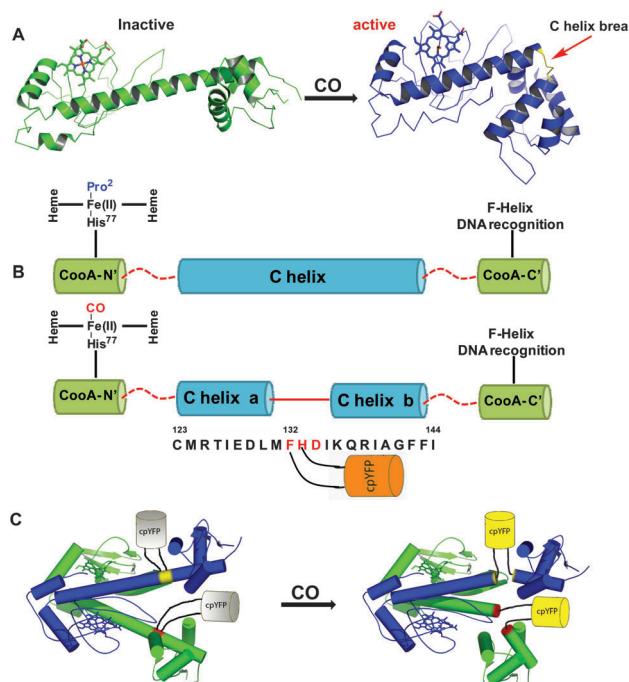


Fig. 6 The biosensor COSer is composed of yellow fluorescent protein (YFP) as the fluorescent reporter and CooA, a dimeric haem protein from *Rhodospirillum rubrum* as the CO recognition unit. (A) Left: inactive monomer CooA. Right: active CO binding to CooA induces a conformational change. (B) Upon CO binding, the C helix of CooA is broken into two parts ("C helix a" and "C helix b") connected by residues 132 to 134. (C) COSer contains YFP inserted by two short linkers between residues 132 and 133 in each CooA monomer. The resulting structural change upon CO binding to CooA induces an increase in fluorescence of the probe. Reprinted with permission from ref. 117. Copyright 2010 Wiley-VCH.

It is also important to note that the irreversible COP-1 might act as a scavenger of CO from biological systems, much like myoglobin (see beginning of Section C), thereby significantly shifting equilibria of CO exchange.

More robust and more sensitive CO sensors are still needed for the investigation of CO-mediated cellular signalling mechanisms and for the evaluation of potential pharmaceutical CO donors designed to treat human diseases. While pharmaceutical NO donors are widely used, the development of methods for delivering CO has not yet led to clinical trials of promising CO-donating compounds.¹¹⁹ Nevertheless, investigations on the potential role of CO gas and CO-releasing molecules as therapeutics are ongoing.^{3,12-14}

D. Controlled release of CO

CO-releasing molecules (CORMs)

The high toxicity of inhaled CO necessitates sophisticated strategies to administer a defined amount of CO as a therapeutic agent at a predetermined location and time. Consequently, carriers of CO that meet specific requirements, such as solubility in aqueous media, low toxicity of these "small" CO-releasing molecules (CORMs) and their degradation products as well as a triggered CO liberation from these compounds, are clearly needed.

Diverse compound classes enable a triggered CO delivery. For example, transition metal-free $\text{Na}_2(\text{H}_3\text{B}-\text{CO}_2)$ (CORM-A1) liberates CO upon acidification.^{120,121} The first protonation step yields $\text{Na}[\text{H}_3\text{B}-\text{CO}_2\text{H}]$ and a second leads to split-off of water and formation of $\text{H}_3\text{B}-\text{CO}$, which rapidly loses CO.¹²² Further, a water-soluble fluorescein analogue 6-hydroxy-3-oxo-3*H*-xanthene-9-carboxylic acid was recently introduced as the first transition metal-free CORM activated by light (photo-CORM) at a wavelength of 500 nm.¹²³ Light-triggered release of a CORM based on micelle-encapsulated unsaturated cyclic α -diketones was also generated. These systems allow the delivery of CO to be monitored by fluorescence imaging techniques.¹²⁴ Organometallic compounds have recently gained significant attention in potential medicinal treatments.¹²⁵ Up to now, metal carbonyl complexes represent preferred reagents to deliver CO because they offer manifold advantageous variations, such as nature and oxidation state of the metal centre (size, charge, Pearson hardness, Lewis acidity, structural diversity), number of carbonyl ligands, nature of coligands (charge, Lewis basicity, π -bonding properties, complex stability) and outer coordination sphere (solubility, Brønsted acidity, amphiphilic character, Fig. 7 and 11).^{23,25,26,126} Current developments also include investigations to enhance the variety of CO-release triggers and to clarify degradation pathways after liberation of CO.

Metal carbonyl complexes have been available for many decades;¹²⁷ however, their therapeutic use as CO-donating reagents has become appreciated only recently.¹²⁸ Metal-based CORMs can contain essential trace elements (especially manganese, iron and cobalt) as well as non-physiological

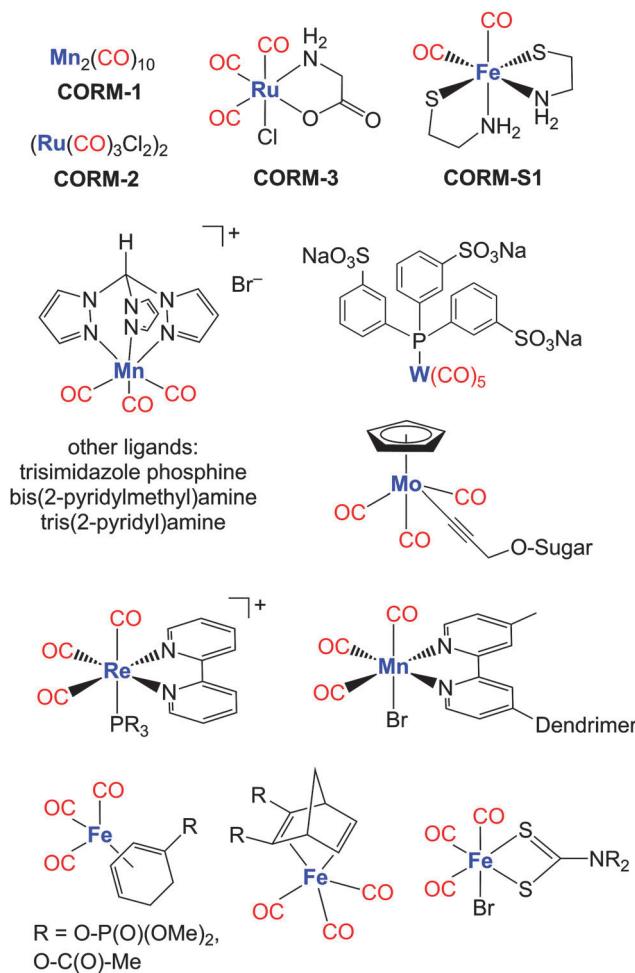


Fig. 7 Selection of metal-based CO-releasing molecules (CORMs).

metals such as ruthenium, tungsten and rhenium.^{125,129} Here, only a selection of very recently investigated CORMs of Cr-, Mn-, Fe-, Mo-, W-, Re- and Ir-containing complexes will be discussed, classified by the three most important mechanisms of CO-release: photoCORMs, solvent-induced ligand exchange on CORMs and enzyme-triggered CORMs (ET-CORMs).^{23,126} CORMs are considered to be possible prodrugs that deliver the signalling molecule CO at the disease site. The vast development of these compounds is based on the first CORM generation consisting of the DMSO- and ethanol-soluble metal carbonyl complexes $\text{Mn}_2(\text{CO})_{10}$ (CORM-1, light-triggered CO release) and CORM-2 (ligand exchange-triggered CO release) as well as water-soluble $(\text{OC})_3\text{RuCl}(\text{O}_2\text{C}-\text{CH}_2-\text{NH}_2)$ (CORM-3, ligand substitution-triggered CO release).¹³

In order to ensure solubility and stability in aqueous media, solvent-separated ions containing non-coordinating anions proved to be advantageous. The recent photoCORM $[(\text{OC})_3\text{Re}\{\text{P}(\text{CH}_2\text{OH})_3\}(\text{bpy})](\text{F}_3\text{CSO}_3)$ was stable and soluble in aerated water and showed no apparent cytotoxicity; irradiation with light initiated the liberation of one CO molecule, which was replaced by a water ligand.¹³⁰ A similar strategy was applied by the group of Mascharak to deliver CO with photoactive manganese(I) complexes of the type $[(\text{OC})_3\text{Mn}(\text{L})]^+$ with L being

a tripodal ligand such as tris(2-pyridyl)amine or bis(2-pyridylmethyl)amine.^{131–133} However, $[(\text{OC})_3\text{Re}\{\text{P}(\text{CH}_2\text{OH})_3\}(\text{bpy})](\text{F}_3\text{CSO}_3)$ bears an outstanding feature: the CORM itself and the inactive CORM (iCORM) products are fluorescent and can be detected by fluorescence microscopy in biological medium.¹³⁰ CORM and iCORM can be discriminated *via* different emission wavelengths.

CO-containing metal anions also prove to be soluble in aqueous media. Iridates(III) of the type $[\text{Cl}_4\text{Ir}(\text{CO})(\text{L})]^-$ with trans-arranged Lewis base L and CO are an impressive example because the nature of L (H₂O, pyridine, 1-methyl-imidazole, 4-dimethylamino-pyridine) influences the back donation of charge into the $\pi^*(\text{CO})$ orbital and the M-CO dissociation energies thus allowing predetermining the CO-release properties.¹³⁴ If the toxicity of the metal comes to the fore, iron-based CORMs seem to be advantageous. This strategy may particularly promising if iron(II) is embedded in a coordination sphere of biogenic ligands also limiting toxic degradation products after CO liberation. Examples of this strategy include photolabile $[(\text{OC})_2\text{Fe}(\text{SCH}_2\text{CH}_2\text{NH}_2)_2]$ (CORM-S1)^{135,136} and $[(\text{OC})_2\text{Fe}\{\text{SCH}_2\text{CH}(\text{CO}_2\text{H})\text{NH}_2\}_2]$ ¹³⁷ with bidentate cysteamine and cysteine ligands, respectively, and *cis*-arranged carbonyl ligands. Homologous $[(\text{OC})_2\text{Ru}(\text{SCH}_2\text{CH}_2\text{NH}_2)_2]$ is not a suitable photoCORM due to the rather short wavelength required for Ru-CO bond activation. However, polypyridyl ruthenium(II) carbonyl complexes allow photoinduced CO liberation.¹³⁸ In addition, Mascharak *et al.* demonstrated the role of ancillary ligands in the capacity of CO photorelease of mono- and dicarbonyl ruthenium(II) complexes with an *N,N,S*-donor ligand.¹³⁹ Nevertheless, the lack of toxicity led to the development of diverse organoiron complexes as suitable photoCORMs.^{27–28,30} A recent example was reported by the group of Kodanko. The stable iron carbonyl complex $[\text{Fe}(\text{CO})(\text{N}_3\text{Py})](\text{ClO}_4)_2$ released CO upon irradiation with 365 nm light and showed photoinitiated growth inhibition of prostate cancer cells.¹⁴⁰ Another group of photoCORMs consists of $\text{Fe}(\text{CO})_3$ fragments bound to π -systems of unsaturated hydrocarbons, such as norbornadiene,¹⁴¹ cyclohexadiene,^{142,143} indenyl¹⁴⁴ and cyclopentadienyl.¹⁴⁵ The substitution patterns of these side-on bound unsaturated hydrocarbons influence solubility in aqueous media and half-lives of CO liberation after irradiation. Photo-activation also initiates CO release from $\text{Mn}(\text{CO})_4$ derivatives with 2-pyridylphenyl ligands.¹⁴⁶ An elegant method to deliver extremely CO-rich molecular metal complexes can be realized by metalodendritic photoCORM. Thus, a metalodendritic photoCORM was built from an organic dendrimer with 2,2'-bipyridyl end groups acting as strong bidentate ligands to multiple $\text{Mn}(\text{CO})_3$ fragments.¹⁴⁷

Light-triggered CO release is probably not suitable in all therapeutic applications and, therefore, other triggers were studied. In chromium complexes of the type $[(\text{OC})_5\text{Cr}(\text{L})]$ with L as halide¹⁴⁸ or aminoesters¹⁴⁹ the rate-determining substitution of L by a solvent molecule (such as water or DMSO) induces the CO release process. Similar CO release mechanisms can be assumed for rhenium(II)-based CORMs also containing varying amounts of bromide anions; an exchange of a bromide by a



water molecule in the vicinity of the metal centre also explains the pH dependence of CO liberation.^{150,151} Further, three iron-based CORMs, $[(\text{PaPy}_3)\text{Fe}(\text{CO})](\text{ClO}_4)$, $[(\text{SBPy}_3)\text{Fe}(\text{CO})](\text{BF}_4)_2$, and $[(\text{Tpmen})\text{Fe}(\text{CO})](\text{ClO}_4)_2$, derived from designed polypyridyl ligands, rapidly release CO upon dissolution and caused vaso-relaxation in a mouse aorta muscle ring preparation.¹⁵² The water soluble $[\text{Fe}_2(\mu\text{-SCH}_2\text{CH}(\text{OH})\text{CH}_2(\text{OH})_2)(\text{CO})_6]$ releases CO *via* substitution by cysteamine with minimal cytotoxicity of the CORM itself on two cell lines QSG-7701 and HepG2.¹⁵³ In addition, Ford *et al.* discussed an oxidative cascade leading to the release of further CO from $\text{Na}_3[\text{W}(\text{CO})_5(\text{tris}(\text{ sulphonatophenyl})\text{phosphine})]$ subsequent to the initial photo-activated CO dissociation (Fig. 7).¹⁵⁴

Closely related to the photoCORMs with a $\text{Fe}(\text{CO})_3$ moiety bound to an unsaturated hydrocarbon, cyclohexadiene iron tricarbonyl complexes also represent enzyme-triggered CO releasing molecules (ET-CORM) if acyloxy side-arms are bound to the side-on bound unsaturated hydrocarbon.^{142,143} This substance class has been well studied for many years.^{155–157} The recently studied use as ET-CORMs is surprisingly convenient. In the first reaction step the ester group is attacked by an esterase leading to a cyclohexadienylalcohol ligand (Fig. 8). Complexes that are modified in such a manner readily decompose under mild oxidative conditions liberating CO. Cytotoxicity and CO-release activity can be tuned by variation of the acyl group of the η^4 -bound acyloxy-cyclohexadiene ligand¹⁵⁸ or by addition of additional substituents at this ligand. CO release from phosphoryloxy-substituted (η^4 -cyclohexadiene) $\text{Fe}(\text{CO})_3$ complexes was induced by a phosphatase and monitored *via* gas chromatography.¹⁵⁹ However, the esterase-triggered oxidative mechanism was monitored with the myoglobin assay under reducing conditions maintained by dithionite. A comparable structural motif, namely a butadiene moiety as part of a six-membered cycle side-on coordinated at a $\text{Fe}(\text{CO})_3$ fragment, stabilises the (η^4 -pyrone)tricarbonyliron(0) complexes that are capable to act as CO transfer reagents for the delivery of controlled amounts of CO.^{160–162} Stronger π -bases, such as a cyclopentadienide anion, push the electroneutral butadiene

base out of the coordination sphere and lead to a η^1 -coordination of the pyrone ligand *via* an oxygen base.¹⁶³

In order to study the mode of action in biological tissues, the detection of CORMs¹⁶⁴ and of liberated CO at the disease site (*i.e.* at the location of CORM degradation) is of utmost importance (see Section B). Therefore, interactions between CORMs and biologically relevant scaffolds were structurally investigated. L-Histidine can act as a tridentate *N,N,O*-ligand at a $\text{Mn}(\text{CO})_3$ fragment.¹⁶⁵ The reaction of a ruthenium-based CORM of the type $[(\text{OC})_3\text{RuL}_3]$ with lysozyme yields the formation of an adduct of five $[\text{Ru}(\text{CO})(\text{H}_2\text{O})_4]^{2+}$ ions with this enzyme, binding to the histidine and aspartate sites; during formation of this complex, the majority of the carbonyl ligands was substituted by water molecules.^{166,167} In this adduct the histidine moiety acts as a monodentate ligand completing the octahedral coordination sphere of the ruthenium ions (Fig. 9).

In light of the potential clinical application of CORMs, the degradation pathway and the nature of the degradation

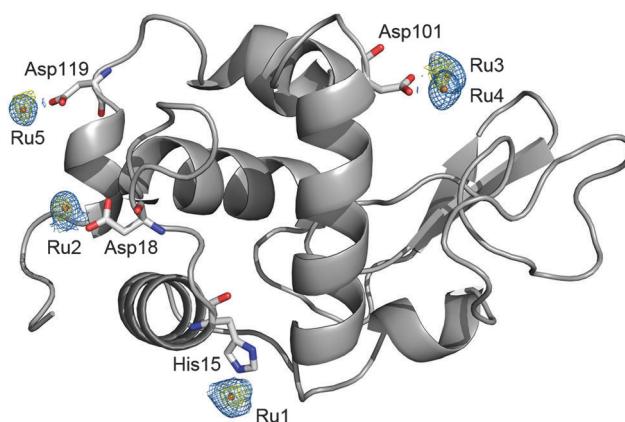


Fig. 9 Structure of hen egg white lysozyme bound to Ru fragments derived from $\text{fac-}[\text{Ru}(\text{CO})_3\text{Cl}_2(1,3\text{-thiazole})]$. Four amino-acid residues (His15, Asp18, Asp101 and Asp119) interact with five ruthenium atoms. Adapted and reprinted with permission from ref. 167. Copyright 2012 Elsevier.

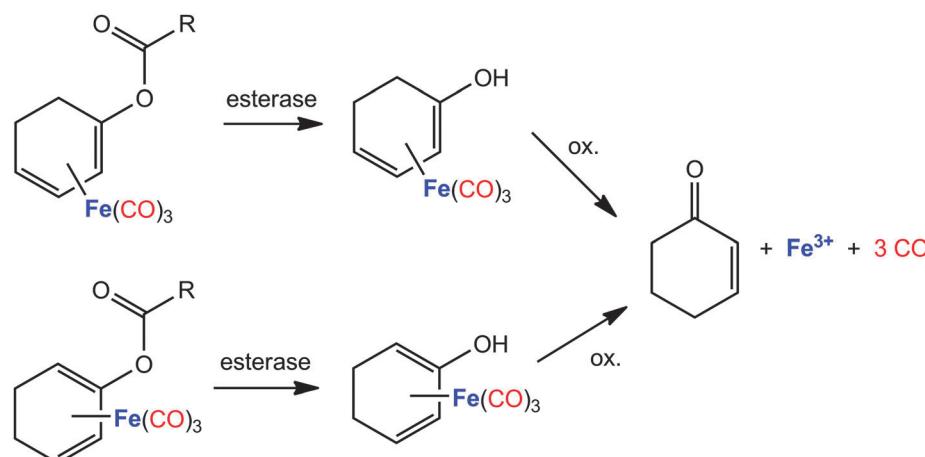


Fig. 8 Degradation and CO liberation from acyloxy-cyclohexadiene tricarbonyliron complexes (enzyme-triggered CORMs, ET-CORMs).^{158,159}

products are of high interest. The first reaction step is the dissociation of at least one CO leaving a vacant coordination site. This remaining metal fragment can either degrade to the metal ions and free coligands as observed for example for $[(\text{MeCN})(\text{OC})\text{Fe}(\text{H}_2\text{NCH}_2\text{CH}_2\text{PPh}_2)_2]^{2+}$ (CORM-P1)¹⁶⁸ or attract another ligand to recomplete the coordination sphere as discussed for the dithiocarbamate (dtc) complexes $[(\text{OC})_4\text{Mn}(\text{dtc})]^{169}$ and $[(\text{OC})_3\text{Fe}(\text{Br})(\text{dtc})]$ binding HPO_4^{2-} , H_2PO_4^- , halide or water at the free coordination site.¹⁷⁰ Further degradation steps may proceed *via* oxidative pathways. Thus, manganese(i)-based mononuclear CORMs were oxidized during step-wise CO release, finally yielding a dinuclear manganese(III) complex with a central Mn–O–Mn unit.¹⁷¹ Further oxidation was observed for CORMs based on heavier transition metals. Thus, the first reaction steps of the degradation of $[(\text{OC})_2\text{ReBr}_4]^{2-}$ (RecORM-1)¹⁷² involve several ligand exchange reactions finally giving the ReO_4^- anion *via* intermediate $[(\text{OC})_2\text{Re}^{\text{II}}(\text{Br})(\text{H}_2\text{O})_3]^+$ and $[(\text{OC})_2\text{Re}^{\text{I}}(\text{Br})(\text{H}_2\text{O})_2(\text{OH})]^-$. After CO release and ligand dissociation, cascades followed by oxidation processes, molybdenum-based CORMs end up as phosphomolybdate $[\text{PMo}_{12}\text{O}_{40}]^{3-}$; an X-ray structure determination showed the adduct formation of this anion with lysozyme *via* a hydrogen-bridge network.¹⁷³

Dependent on the pH value of the aqueous solutions, the carbonyl ligands can be attacked by hydroxide ions yielding M–C(O)OH moieties (Fig. 10). There is evidence that $[(\text{OC})_3\text{Ru}(\text{Cl})(\text{O}_2\text{CCH}_2\text{NH}_2)]$ acts as a strong acid.¹⁷⁴ Thus, this CORM

binds OH^- from water and the remaining protons lead to a pH value of 3. If the resulting anion is titrated with a base until a nearly neutral pH value of 6 is reached, a doubly charged anion (due to deprotonation of the Ru–CO₂H moiety) or a chloride-free anion (*via* exchange of the chloride ion by a hydroxyl ligand) is formed as shown in the middle row of Fig. 10. In alkaline solution (pH = 10) both reaction patterns are realized yielding the dianion depicted in the bottom row of Fig. 10.¹⁷⁴ The mechanisms of CO release from ruthenium(II)-based CORMs with methoxycarbonyl or ethoxycarbonyl ligands are also known.¹⁷⁵ Whereas in this example the oxidation state of Ru remained 2+, transition metals can adopt several stable oxidation states and the CO release properties may interfere with the redox chemistry of these metals in aqueous solution. Mann *et al.* noted that CORMs might be able to catalyze Fenton-type reactions leading to the formation of ROS.²⁵ It is well-known that the pK_a values of water molecules in the vicinity of metal cations differ significantly from free water molecules easing the formation of metal-bound hydroxide¹⁷⁶ and influencing the redox behaviour of the metals.

Thus far, CO-release trigger, solubility in aqueous media, kinetics of CO liberation, toxicity of the CORMs itself, and their degradation products played the major role in the development of new CORMs. Recent work focused on the targeted delivery by variation of the outer ligand sphere. Thus, peptides can be part of one ligand in order to support targeted delivery to cellular systems.^{177–179} In addition, it can be desirable to develop fast and slow CO releasers to suit diverse therapeutic applications. In special cases it can also be advantageous to immobilize the CORMs in order to ease removal of metal-containing degradation products and/or to control the environment of the metal ions (see the section CO-releasing materials). Future developments need to combine strategies for predetermining CO-release properties and for targeted delivery. It is also necessary to prepare and assess iCORMs independently to investigate their physiological properties after CO release.^{178,180}

The recent review by Romão *et al.* conceptualised elegantly the future CORM design.²³ They proposed a model as a tool to help rationalising the design of metal carbonyl CORMs with the appropriate pharmaceutical properties. As an example, an octahedral geometry with six ligands surrounding the metal centre was shown (Fig. 11). At least one CO ligand coordinates to the metal centre. Thermodynamic and kinetic stability of the complex is provided by chelating ligands and 18 electrons in the valence shell of the central metal atom. All ancillary ligands display an influence on the electronic density, oxidation behaviour and CO release at the metal centre. Thus, the coordination sphere of a given CORM drug influences resistance to plasma proteins and responds on a specific CO release trigger. However, a pharmaceutical CORM needs an appropriate pharmacological profile.²³ It is very important to control solubility in aqueous solutions, cellular internalisation, as well as the pharmacological ADME characteristics, pharmacokinetic profile and targeting to diseased tissues (ADME is an abbreviation in pharmacokinetics and pharmacology for absorption, distribution, metabolism, and excretion, and describes the

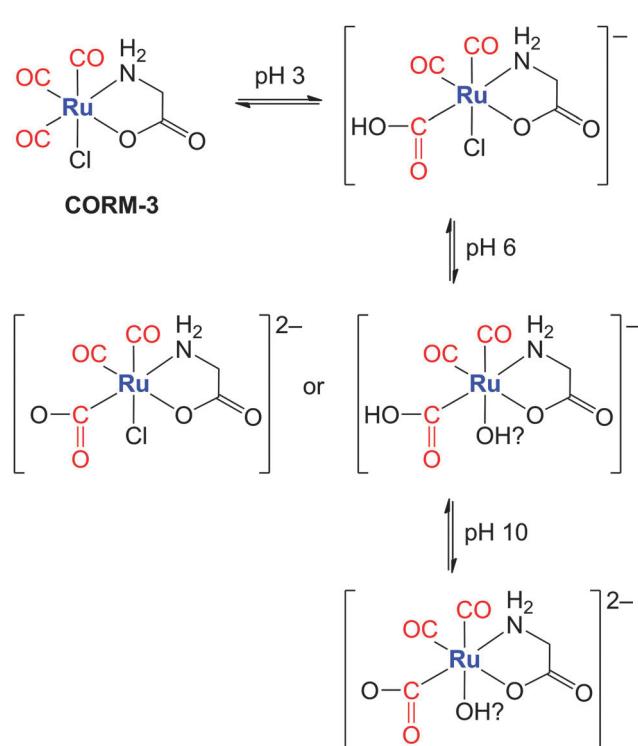


Fig. 10 Proposed pH-dependent interaction of CORM-3 with bases. The attack of a hydroxide ion at a carbonyl ligand results in a hydroxycarbonyl moiety; at higher pH values the acyl group can be deprotonated or the chloride ion substituted by an OH group. The suggested final species combines both possible interaction pathways under alkaline conditions.¹⁷⁴



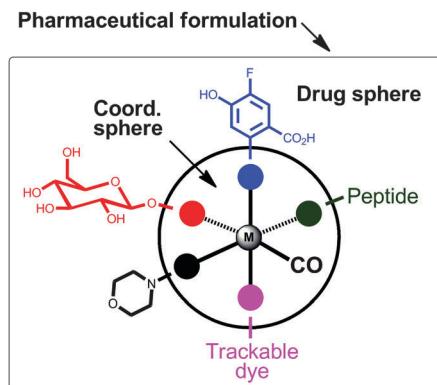


Fig. 11 Conceptual model for the development of pharmaceutical CORMs (adapted from Romão *et al.*).²³

disposition of a pharmaceutical compound within an organism). The resulting “drug sphere” can be obtained by modifying the coordinating ligands at their distal sites (Fig. 11). Further, the pharmaceutical formulation determines which different chemical substances, including the active CORM, are combined to produce a final medicinal product.¹⁸¹ For example, a tablet contains a variety of other substances apart from the drug itself, and studies have to be carried out to ensure that the drug is compatible with these other substances. Extending the model of Romão *et al.*, we postulate five different substituents in the coordination sphere. Carbohydrates and peptides can enhance water solubility,¹⁸² biocompatibility and even biodistribution to certain tissues.^{177,183,184} Morpholino groups may provide an amphiphilic character to the CORM. Solubility, membrane permeation and the pharmacokinetic profile may be controlled by terminal groups, such as amino, carboxylate groups and fluorine moieties. In addition, trackable dyes could help to investigate the metabolism of the CORM *in vitro* and *in vivo*.¹³⁰ Finally, we emphasize that the coordination sphere, drug sphere and the pharmaceutical formulation “will play a decisive role in the generation of novel CORM drugs”.²³

CO-releasing materials (CORMAs)

As previously shown, metal carbonyl complexes are the most appropriate and successful class of (soluble) CORMs; however, it is also important to evaluate their possible shortcomings. In fact, very few pharmaceutical drugs are organometallic compounds,^{125,129} mostly due to side reactivity of metals with biological substances (*e.g.*, nucleophilic and electrophilic side chains of proteins) and the toxicity of many heavy metals. Systemic application of water-soluble CORMs results in distribution throughout the body, which can lead to increased toxicities against healthy tissues. The spatially- and time-controlled release in the tissue still remains a great challenge.¹¹⁹ Moreover, the CO release process inevitably generates a metal–coligand fragment, which can potentially have biological activity. These fragments could be retained in insoluble matrices. Thus, development of solid-storage forms of CO in combination with a specific trigger for the gas release is an

important research goal. In addition, macromolecular and nanoscale carrier systems¹⁸⁵ can be utilized to achieve tissue-specific enrichment and delivery of CORMs.^{30,186}

Hubbell *et al.* developed CO-releasing micelles with reduced diffusion in tissues and better ability to target distal tissue draining sites.¹⁰⁶ The micelles were prepared from triblock copolymers composed of a hydrophilic poly(ethylene glycol) block, a poly(ornithine acrylamide) block bearing $[\text{Ru}(\text{CO})_3\text{Cl}(\text{ornithinate})]$ moieties and a hydrophobic poly(*n*-butylacrylamide) block.¹⁸⁷ CO release from the micelles was induced *via* addition of cysteine. It was slower than that of $[\text{Ru}(\text{CO})_3\text{Cl}(\text{glycinate})]$ (CORM-3, Fig. 7); however, the micelles attenuated successfully the lipopolysaccharide-induced inflammatory response of human monocytes. In addition, the toxicity of $[\text{Ru}(\text{CO})_3\text{Cl}(\text{amino acidate})]$ moieties was significantly reduced by the “stealth” feature of poly(ethylene glycol).¹⁰⁶ $\text{Ru}(\text{CO})_3\text{Cl}(\text{glycinate})$ was also covalently attached to an amphiphilic peptide. The small peptide self-assembled into nanofiber gels and spontaneously released CO with prolonged release kinetics compared with CORM-3.¹⁷⁹

Recently, a novel concept of triggering CORMAs was presented. Biocompatible magnetic iron oxide nanoparticles have been used as carriers for CORMs. In the proof-of-concept study the rate of CO release from $[\text{RuCl}(\text{CO}_3)(\mu\text{-DOPA})]@\text{maghemite}$ nanoparticles was doubled upon exposure to an external alternating magnetic field (31.7 kAm^{-1} , 247 kHz , 25°C , 39.9 mTesla , DOPA = dioxyphenyl-alaninato, Fig. 12).¹⁸⁸

Porous coordination polymers, also known as metal–organic frameworks (MOFs), form structures with very high inner surface areas and ordered pore channels with various sizes.^{189,190} These features make MOFs highly attractive materials for gas-storage, especially for small gaseous molecules, such as H_2 , CH_4 and CO_2 .^{191,192} Very recently, iron-based MOFs have been generated for the loading and delivery of CO.¹⁹³ The materials are rapidly synthesized in the microwave from iron(III)chloride and terephthalic acid and derivatives thereof (Fig. 13). CO loading occurs *via* unsaturated coordination sites shown as empty circles in Fig. 13C. CO coordination was verified by infrared and Mössbauer spectroscopy. This novel type of CORMA shows good biocompatibility and releases CO with $t_{1/2}$ from 38 to 76 min *via* degradation of the material under physiological conditions.

Protocols for the covalent immobilization of photoCORMs²⁸ on nanoparticles have also been established. These nanocarriers have many potential benefits for diagnosing¹⁶⁴ and treating local and metastatic cancer, following the enhanced permeation and retention effect.¹⁸⁵ $[\text{Mn}(\text{CO})_3(\text{tpm})]^+$ (tpm = tris(pyrazolyl)methane) complexes containing alkyne-functionalized tpm ligands were used. These complexes were covalently linked to silicon dioxide nanoparticles and dopable nanodiamonds *via* the copper-catalyzed azide–alkyne 1,3-dipolar cycloaddition (Fig. 14).^{194,195} The myoglobin assay¹⁰³ demonstrated that the CORM-functionalized nanoparticles have photoinducible CO-release properties very similar to the free complexes.^{28,29}

The organometallic *fac*- $\text{Mn}(\text{CO})_3$ fragment was also bound to a methacrylate or methacrylamide polymer backbone *via* bis(pyridylmethyl)amine-type ligands. The resulting $\text{Mn}(\text{CO})_3$ –polymer



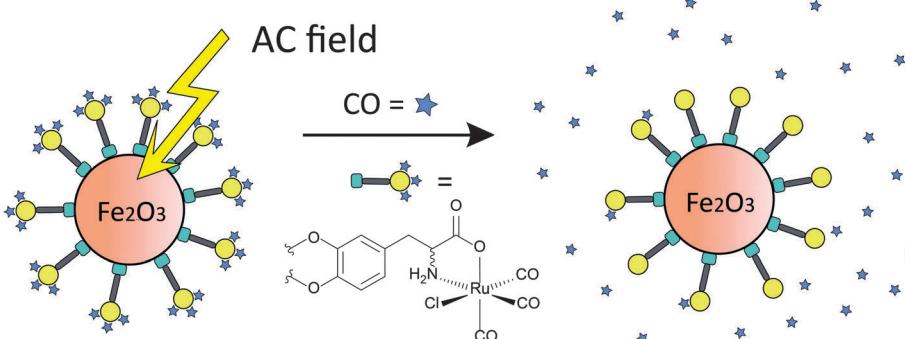


Fig. 12 Schematic presentation of induced CO release from CORM-functionalized iron oxide nanoparticles through an alternating magnetic (AC) field. Tri(carbonyl)-chlorido-dihydroxyphenylalaninato-ruthenium(II) displays an immobilised analogue of CORM-3. Adapted and reprinted with permission from ref. 188. Copyright 2013 RSC Publishing.

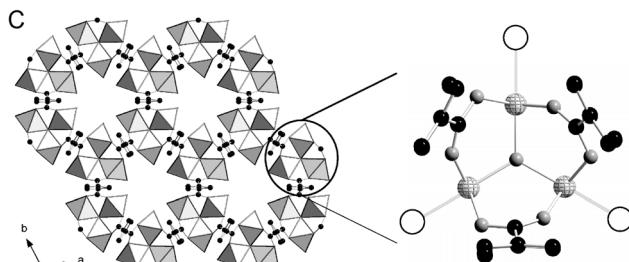
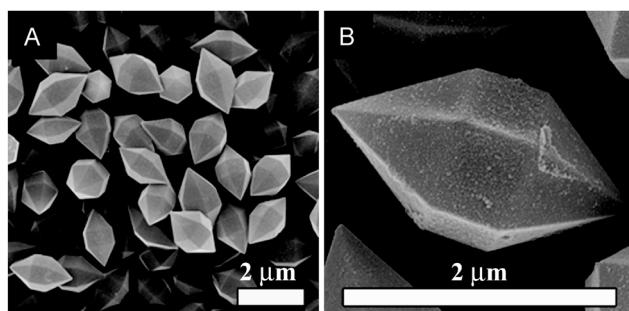


Fig. 13 Metal–organic frameworks (MOFs) with iron for the loading and delivery of CO. (A and B) Scanning electron microscope pictures of crystals of Fe MOF with terephthalic acid. (C) Structure of Fe MOF, as viewed along the c axis. Empty circles represent the coordination sites for CO, iron atoms: light-gray spheres, oxygen atoms: gray spheres, and carbon atoms: black spheres. Adapted and reprinted with permission from ref. 193. Copyright 2013 Wiley-VCH.

conjugates were investigated as photoinducible CO-releasing materials (photoCORMAs).¹⁹⁶ In general, NO- and CO-releasing materials (NORMAs & CORMAs) with NO and CO photodonors can retain toxic metabolites after gas release in the biocompatible polymer matrix.¹⁹⁷ The concept of embedding water-insoluble, photoactive NO metal complexes into nanoparticles¹⁹⁸ and fibrous polymer non-wovens¹⁹⁹ has been transferred to phototriggerable metal carbonyls (Fig. 15). Effective NO or CO release into the surrounding medium is initiated by light stimulation of the high surface area materials. For the generation of NORMAs, novel biscarboxamide rutheniumnitrosyl complexes $\{\text{RuNO}\}^6$ have been synthesised.^{200,201} For a photoCORMA, $\text{Mn}_2(\text{CO})_{10}$ (CORM-1) was used.^{202,203} The metal

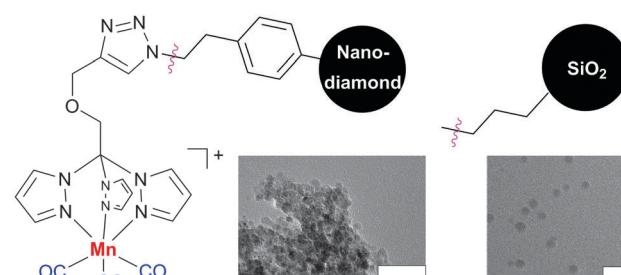


Fig. 14 $[\text{Mn}(\text{CO})_3(\text{tpm})]^+$ (tpm = tris(pyrazolyl)methane) complexes containing alkyne-functionalized tpm ligands covalently attached to dopable nanodiamonds¹⁹⁴ and silica nanoparticles.¹⁹⁵ The corresponding transmission electron microscope pictures are also shown (bars indicate 50 nm). Adapted and reprinted with permission from ref. 194 and 195. Copyright 2011 American Chemical Society and 2012 RSC Publishing.

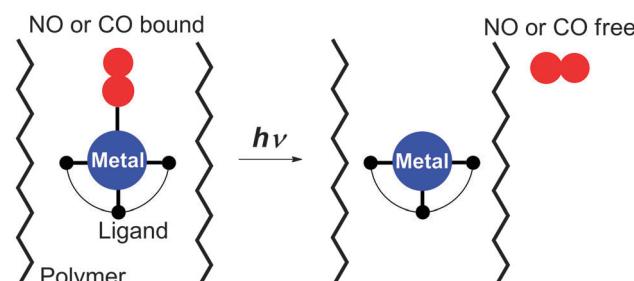
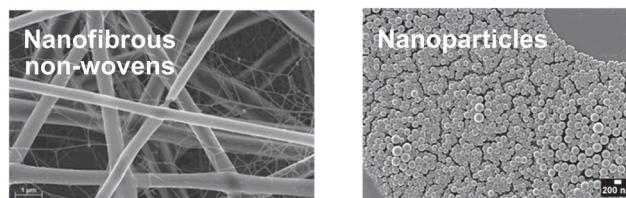


Fig. 15 Concept of embedding water-insoluble, photoactive NO and CO metal complexes into fibrous polymer non-wovens and nanoparticles. The NORMAs and CORMAs are important for the development of safe NO and CO delivering devices for therapeutic purposes; toxic metabolites after gas release are retained in the biocompatible polymer matrix.^{200–203}

complexes were non-covalently embedded into the polymer matrices *via* miniemulsion technique²⁰⁴ or electrospinning.²⁰⁵



Leaching of the metal complexes out of the polymeric matrices into water was negligible due to their water insolubility. Irradiation with $\lambda = 366\text{--}480\text{ nm}$ in water showed a significant phototriggered NO/CO release from the nanoparticles or non-wovens. Cytotoxicity tests of the CORMA with 3T3 mouse fibroblast cells in the dark revealed very low cell death. After illumination, CO bubbled out of the nanofibres thereby eradicating the fibroblast cell culture.^{202,203}

E. CO – where does it go?

The discovery of CO as an endogenous gaseous messenger has triggered intensive research regarding cellular CO signalling and the design of carrier systems that provide a controlled release of CO. Despite substantial progress on various levels, there are quite a number of open questions that need to be addressed, in particular if clinical applications of CO or CO-releasing molecules and materials are envisioned. Some of such open questions are discussed in the following.

For understanding the CO-related physiology it will be mandatory to profile the expression and targeting of haem oxygenases. Because CO signalling is most probably a “local” event, we need to know in detail (a) the localization of HO, (b) the availability of haem and (c) the availability of NADPH. Once released *via* HO activity, what is the fate of the CO molecule? It is often assumed that CO will immediately find the desired target system, but it is not yet clear what CO buffer capacity the cellular cytosol provides and, hence, what is the effective sphere of action of a cytosolic CO molecule? In addition, what is the ultimate destination of CO? How much becomes covalently bound, which fraction only undergoes loose interactions with other molecules, and how much is finally cleared from the body *via* the lungs? Quantitative data are required to facilitate predictions about CO-related physiological processes.

Except for some haemoproteins, our knowledge on the molecular mechanisms by which CO affects protein function is still very much limited. Recent examples showing that CO modulates the function of ion channels, for example, still lack a clear mechanistic insight. Does an action of CO on a protein always require the presence of a haem group or a transition metal or are there other modes of CO-protein interactions feasible?

The necessity of CORMs to target specific disease sites and to release CO at a predetermined time point is obvious. In order to effectively initiate liberation of CO from metal carbonyl complexes, a variety of CO release triggers are required to fulfil boundary conditions such as governed CO release *via* irradiation, enzymatic activation, pH changes, ligand substitution, temperature, redox reactions, and others. Therefore, in some cases it might be beneficial if CORMs are either hydrophobic or amphiphilic in order to have them enter the cells or enrich in the membranes, respectively. In addition, different therapeutic applications might need slow or fast liberation of CO. Specific applications call for particular CORMs with respect to delivery of the carbonyl complexes at the disease site, initiation of CO

release, interference of the tissues with the metal complexes themselves or with their degradation products. The performance of those CORMs can be further adapted with a suitable carrier system (*e.g.* micelle, nanoparticle, fibre, non-woven *etc.*). Smart materials that release CO by a trigger and retain degradation products can reach their target *via* specific interactions of materials and cells. In future investigations, the interaction of these complexes and their degradation products with reactive oxygen species (occasionally causing Fenton-type chemistry), peptides, amino acids and other biological environments deserves particular attention.

Another issue involves inactivated CORM (iCORM) products. Unfortunately, the potential biological activity of such metal-coligand fragments, inevitably remaining after CO release from the metal coordination sphere, has been often neglected. In the future, novel CORMs with biological activity should include a detailed characterisation of corresponding iCORMs.

With the development of specific CORMs it is also necessary to intensify research on real-time detection of physiological levels of CO inside living cells. New CO sensors must be robust, selective and sensitive in the lower μM range. Furthermore, rapid response times and a good signal-to-noise-ratio will be required.

Understanding the CO-related physiology is still in its infancy; the currently available CORMs and CORMAs described are not yet optimized for clinical or experimental applications. Therefore, physiologists, physicians and chemists must collaborate for a better understanding of CO in the body and how to utilize CO as a drug.

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References

- 1 L. R. Goldbaum, R. G. Ramirez and K. B. Absalon, *Aviat., Space Environ. Med.*, 1975, **46**, 1289–1291.
- 2 H. P. Kim, S. W. Ryter and A. M. Choi, *Annu. Rev. Pharmacol. Toxicol.*, 2006, **46**, 411–449.
- 3 R. Motterlini and L. E. Otterbein, *Nat. Rev. Drug Discovery*, 2010, **9**, 728–743.
- 4 M. D. Maines, *FASEB J.*, 1988, **2**, 2557–2568.
- 5 R. F. Coburn, *Ann. N. Y. Acad. Sci.*, 1970, **174**, 11–22.
- 6 E. Aztatzi-Santillán, F. E. Nares-López, B. Márquez-Valadez, P. Aguilera and M. E. Chávez-Cárdenas, *Cent. Nerv. Syst. Agents Med. Chem.*, 2010, **10**, 310–316.



7 E. Zeynalov, Z. A. Shah, R. C. Li and S. Doré, *Neurobiol. Dis.*, 2009, **35**, 264–269.

8 J. Wang, H. Zhuang and S. Doré, *Neurobiol. Dis.*, 2006, **22**, 473–476.

9 A. Ferreira, J. Balla, V. Jeney, G. Balla and M. P. Soares, *J. Mol. Med.*, 2008, **86**, 1097–1111.

10 A. Pamplona, A. Ferreira, J. Balla, V. Jeney, G. Balla, S. Epiphanio, A. Chora, C. D. Rodrigues, I. P. Gregoire, M. Cunha-Rodrigues, S. Portugal, M. P. Soares and M. M. Mota, *Nat. Med.*, 2007, **13**, 703–710.

11 S. W. Ryter and A. M. K. Choi, *Am. J. Respir. Cell Mol. Biol.*, 2009, **41**, 251–260.

12 R. Motterlini, B. Haas and R. Foresti, *Med. Gas Res.*, 2012, **2**, 28.

13 R. Foresti, M. G. Bani-Hani and R. Motterlini, *Intensive Care Med.*, 2008, **34**, 649–658.

14 B. E. Mann and R. Motterlini, *Chem. Commun.*, 2007, 4197–4208.

15 B. S. Zuckerbraun, B. Y. Chin, B. Wegiel, T. R. Billiar, E. Czsimadia, J. Rao, L. Shimoda, E. Ifedigbo, S. Kanno and L. E. Otterbein, *J. Exp. Med.*, 2006, **203**, 2109–2119.

16 J. Kohmoto, A. Nakao, T. Kaizu, A. Tsung, A. Ikeda, K. Tomiyama, T. R. Billiar, A. M. K. Choi, N. Murase and K. R. McCurry, *Surgery*, 2006, **140**, 179–185.

17 G. L. Volti, L. F. Rodella, C. D. Giacomo, R. Rezzani, R. Bianchi, E. Borsani, D. Gazzolo and R. Motterlini, *Nephron Exp. Nephrol.*, 2006, **104**, e135–e139.

18 R. Motterlini, A. Gonzales, R. Foresti, J. E. Clark, C. J. Green and R. M. Winslow, *Circ. Res.*, 1998, **83**, 568–577.

19 A. Nakao, D. J. Kaczorowski, Y. Wang, J. S. Cardinal, B. M. Buchholz, R. Sugimoto, K. Tobita, S. Lee, Y. Toyoda, T. R. Billiar and K. R. McCurry, *J. Heart Lung Transplant.*, 2010, **29**, 544–553.

20 S. Takaki, N. Takeyama, Y. Kajita, T. Yabuki, H. Noguchi, Y. Miki, Y. Inoue, T. Nakagawa and H. Noguchi, *Intensive Care Med.*, 2010, **36**, 42–48.

21 C. A. Piantadosi, *Free Radicals Biol. Med.*, 2008, **45**, 562–569.

22 F. Gullotta, A. d. Masi, M. Coletta and P. Ascenzi, *BioFactors*, 2012, **38**, 1–13.

23 C. C. Romão, W. A. Blattler, J. D. Seixas and G. J. L. Bernardes, *Chem. Soc. Rev.*, 2012, **41**, 3571–3583.

24 L. Yuan, W. Lin, L. Tan, K. Zheng and W. Huang, *Angew. Chem., Int. Ed.*, 2013, **52**, 1628–1630.

25 B. E. Mann, *Organometallics*, 2012, **31**, 5728–5735.

26 B. Mann, in *Medicinal Organometallic Chemistry*, ed. G. Jaouen and N. Metzler-Nolte, Springer, Berlin, Heidelberg, 2010, vol. 32, ch. 10, pp. 247–285.

27 R. D. Rimmer, A. E. Pierri and P. C. Ford, *Coord. Chem. Rev.*, 2012, **256**, 1509–1519.

28 U. Schatzschneider, *Inorg. Chim. Acta*, 2011, **374**, 19–23.

29 U. Schatzschneider, *Eur. J. Inorg. Chem.*, 2010, 1451–1467.

30 D. Crespy, K. Landfester, U. S. Schubert and A. Schiller, *Chem. Commun.*, 2010, **46**, 6651–6662.

31 S. Fetzner and R. A. Steiner, *Appl. Microbiol. Biotechnol.*, 2010, **86**, 791–804.

32 S. Fetzner, *Appl. Microbiol. Biotechnol.*, 2002, **60**, 243–257.

33 H. M. Girvan and A. W. Munro, *J. Biol. Chem.*, 2013, **288**, 13194–13203.

34 W. J. Wilkinson and P. J. Kemp, *J. Physiol.*, 2011, **589**, 3055–3062.

35 C. Peers, *Exp. Physiol.*, 2011, **96**, 836–839.

36 G. P. Roberts, H. Youn and R. L. Kerby, *Microbiol. Mol. Biol. Rev.*, 2004, **68**, 453–473.

37 J. L. Boer, S. B. Mulrooney and R. P. Hausinger, *Arch. Biochem. Biophys.*, 2014, **544**, 142–152.

38 S. M. Siepka, S. H. Yoo, J. Park, C. Lee and J. S. Takahashi, *Cold Spring Harbor Symp. Quant. Biol.*, 2007, **72**, 251–259.

39 E. M. Dioum, J. Rutter, J. R. Tuckerman, G. Gonzalez, M. A. Gilles-Gonzalez and S. L. McKnight, *Science*, 2002, **298**, 2385–2387.

40 T. Uchida, E. Sato, A. Sato, I. Sagami, T. Shimizu and T. Kitagawa, *J. Biol. Chem.*, 2005, **280**, 21358–21368.

41 T. Ingi, J. Cheng and G. V. Ronnett, *Neuron*, 1996, **16**, 835–842.

42 R. Feil and B. Kemp-Harper, *EMBO Rep.*, 2006, **7**, 149–153.

43 E. R. Derbyshire and M. A. Marletta, *Annu. Rev. Biochem.*, 2012, **81**, 533–559.

44 J. R. Stone and M. A. Marletta, *Biochemistry*, 1994, **33**, 5636–5640.

45 A. Fribe, G. Schultz and D. Koesling, *EMBO J.*, 1996, **15**, 6863–6868.

46 L. J. Ignarro, *Nitric Oxide - Biology and Pathobiology*, Elsevier Inc., London, 2010.

47 E. Martin, V. Berka, E. Bogatenkova, F. Murad and A. L. Tsai, *J. Biol. Chem.*, 2006, **281**, 27836–27845.

48 J. N. Burstyn, A. E. Yu, E. A. Dierks, B. K. Hawkins and J. H. Dawson, *Biochemistry*, 1995, **34**, 5896–5903.

49 E. M. Boon, S. H. Huang and M. A. Marletta, *Nat. Chem. Biol.*, 2005, **1**, 53–59.

50 E. Dubuis, M. Potier, R. Wang and C. Vandier, *Cardiovasc. Res.*, 2005, **65**, 751–761.

51 K. Decaluwe, B. Pauwels, S. Verpoest and J. V. d. Voorde, *Eur. J. Pharmacol.*, 2012, **674**, 370–377.

52 X. Ma, N. Sayed, A. Beuve and F. van den Akker, *EMBO J.*, 2007, **26**, 578–588.

53 P. M. Schmidt, C. Rothkegel, F. Wunder, H. Schroder and J. P. Stasch, *Eur. J. Pharmacol.*, 2005, **513**, 67–74.

54 F. Ko, C. Wu, S. Kuo, F. Lee and C. Teng, *Blood*, 1994, **84**, 4226–4233.

55 E. Martin, Y. C. Lee and F. Murad, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 12938–12942.

56 R. Purohit, A. Weichsel and W. R. Montfort, *Protein Sci.*, 2013, **22**, 1439–1444.

57 K. A. White and M. A. Marletta, *Biochemistry*, 1992, **31**, 6627–6631.

58 H. Li, J. Igarashi, J. Jamal, W. Yang and T. L. Poulos, *JBIC, J. Biol. Inorg. Chem.*, 2006, **11**, 753–768.

59 S. W. Ragsdale, *J. Inorg. Biochem.*, 2007, **101**, 1657–1666.

60 Y. Kung, T. I. Doukov, J. Seravalli, S. W. Ragsdale and C. L. Drennan, *Biochemistry*, 2009, **48**, 7432–7440.

61 J. H. Jeoung and H. Dobbek, *J. Am. Chem. Soc.*, 2009, **131**, 9922–9923.

62 *Anorganische Chemie*, ed. R. Steudel, De Gruyter, Berlin, 2012.

63 H. Lin and J. J. McGrath, *Drug Chem. Toxicol.*, 1988, **11**, 371–385.

64 T. Hoshi, A. Pantazis and R. Olcese, *Physiology*, 2013, **28**, 172–189.

65 M. T. Nelson and J. M. Quayle, *Am. J. Physiol.*, 1995, **268**, C799–C822.

66 R. Brenner, G. J. Perez, A. D. Bonev, D. M. Eckman, J. C. Kosek, S. W. Wiler, A. J. Patterson, M. T. Nelson and R. W. Aldrich, *Nature*, 2000, **407**, 870–876.

67 A. J. Patterson, J. Henrie-Olson and R. Brenner, *Trends Cardiovasc. Med.*, 2002, **12**, 78–82.

68 M. Nara, P. D. Dhulipala, G. J. Ji, U. R. Kamasani, Y. X. Wang, S. Matalon and M. I. Kotlikoff, *Am. J. Physiol.: Cell Physiol.*, 2000, **279**, C1938–C1945.

69 M. Fukao, H. S. Mason, F. C. Britton, J. L. Kenyon, B. Horowitz and K. D. Keef, *J. Biol. Chem.*, 1999, **274**, 10927–10935.

70 Y. Wu, Y. Yang, S. Ye and Y. Jiang, *Nature*, 2010, **466**, 393–397.

71 P. Yuan, M. D. Leonetti, A. R. Pico, Y. Hsiung and R. Mackinnon, *Science*, 2010, **329**, 182–186.

72 P. Yuan, M. D. Leonetti, Y. Hsiung and R. MacKinnon, *Nature*, 2012, **481**, 94–97.

73 M. L. Dallas, J. L. Scragg and C. Peers, *Neuroreport*, 2008, **19**, 345–348.

74 F. T. Horrigan and R. W. Aldrich, *J. Gen. Physiol.*, 2002, **120**, 267–305.

75 R. Wang and L. Wu, *J. Biol. Chem.*, 1997, **272**, 8222–8226.

76 S. E. Williams, P. Wootton, H. S. Mason, J. Bould, D. E. Iles, D. Riccardi, C. Peers and P. J. Kemp, *Science*, 2004, **306**, 2093–2097.

77 S. Hou, R. Xu, S. H. Heinemann and T. Hoshi, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 4039–4043.

78 S. E. Williams, S. P. Brazier, N. Baban, V. Telezhkin, C. T. Muller, D. Riccardi and P. J. Kemp, *Pflugers Arch.*, 2008, **456**, 561–572.

79 V. Telezhkin, S. P. Brazier, R. Mears, C. T. Muller, D. Riccardi and P. J. Kemp, *Pflugers Arch.*, 2011, **461**, 665–675.

80 K. A. Bixby, M. H. Nanao, N. V. Shen, A. Kreusch, H. Bellamy, P. J. Pfaffinger and S. Choe, *Nat. Struct. Biol.*, 1999, **6**, 38–43.

81 G. Wang, C. Strang, P. J. Pfaffinger and M. Covarrubias, *J. Biol. Chem.*, 2007, **282**, 13637–13647.

82 G. Liu, J. Shi, L. Yang, L. Cao, S. M. Park, J. Cui and S. O. Marx, *EMBO J.*, 2004, **23**, 2196–2205.

83 S. Hou, R. Xu, S. H. Heinemann and T. Hoshi, *Nat. Struct. Mol. Biol.*, 2008, **15**, 403–410.

84 S. Hou, F. T. Horrigan, R. Xu, S. H. Heinemann and T. Hoshi, *Channels*, 2009, **3**, 249–258.

85 X. D. Tang, M. L. Garcia, S. H. Heinemann and T. Hoshi, *Nat. Struct. Mol. Biol.*, 2004, **11**, 171–178.

86 G. Zhang, R. Xu, S. H. Heinemann and T. Hoshi, *Biochem. Biophys. Res. Commun.*, 2006, **342**, 1389–1395.



87 Y. J. Peng, J. Nanduri, G. Raghuraman, D. Souvannakitti, M. M. Gadalla, G. K. Kumar, S. H. Snyder and N. R. Prabhakar, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 10719–10724.

88 N. R. Prabhakar, *J. Physiol.*, 2013, **591**, 2245–2257.

89 S. Wang, S. Publicover and Y. Gu, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 2957–2962.

90 A. L. Eastwood and M. B. Goodman, *Physiology*, 2012, **27**, 282–290.

91 P. Ponka, *Am. J. Med. Sci.*, 1999, **318**, 241–256.

92 X. D. Tang, R. Xu, M. F. Reynolds, M. L. Garcia, S. H. Heinemann and T. Hoshi, *Nature*, 2003, **425**, 531–535.

93 J. H. Jaggar, A. Li, H. Parfenova, J. Liu, E. S. Umstot, A. M. Dopico and C. W. Leffler, *Circ. Res.*, 2005, **97**, 805–812.

94 L. Yi, J. T. Morgan and S. W. Ragsdale, *J. Biol. Chem.*, 2010, **285**, 20117–20127.

95 T. Kuhl, N. Sahoo, M. Nikolajski, B. Schlott, S. H. Heinemann and D. Imhof, *ChemBioChem*, 2011, **12**, 2846–2855.

96 F. T. Horrigan, S. H. Heinemann and T. Hoshi, *J. Gen. Physiol.*, 2005, **126**, 7–21.

97 J. R. Alonso, F. Cardellach, S. Lopez, J. Casademont and O. Miro, *Pharmacol. Toxicol.*, 2003, **93**, 142–146.

98 Y. K. Choi, E. D. Por, Y. G. Kwon and Y. M. Kim, *Oxid. Med. Cell. Longevity*, 2012, 794237.

99 M. L. Dallas, J. L. Scragg and C. Peers, *Adv. Exp. Med. Biol.*, 2009, **648**, 89–95.

100 E. M. Hetrick and M. H. Schoenfisch, *Annu. Rev. Anal. Chem.*, 2009, **2**, 409–433.

101 S. S. Park, J. Kim and Y. Lee, *Anal. Chem.*, 2012, **84**, 1792–1796.

102 R. A. Hunter, W. L. Storm, P. N. Coneski and M. H. Schoenfisch, *Anal. Chem.*, 2013, **85**, 1957–1963.

103 S. McLean, B. E. Mann and R. K. Poole, *Anal. Biochem.*, 2012, **427**, 36–40.

104 G. S. Marks, H. J. Vreman, B. E. McLaughlin, J. F. Brien and K. Nakatsu, *Antioxid. Redox Signaling*, 2002, **4**, 271–277.

105 Y. Morimoto, W. Durante, D. G. Lancaster, J. Klattenhoff and F. K. Tittel, *Am. J. Physiol.: Heart Circ. Physiol.*, 2001, **280**, H483.

106 U. Hasegawa, A. J. van der Vlies, E. Simeoni, C. Wandrey and J. A. Hubbell, *J. Am. Chem. Soc.*, 2010, **132**, 18273–18280.

107 J.-M. Barbe, G. Canard, S. Brandes and R. Guilard, *Chem.-Eur. J.*, 2007, **13**, 2118–2129.

108 K. S. Davidge, G. Sanguinetti, C. H. Yee, A. G. Cox, C. W. McLeod, C. E. Monk, B. E. Mann, R. Motterlini and R. K. Poole, *J. Biol. Chem.*, 2009, **284**, 4516–4524.

109 M. Desnard, R. Foresti, D. Morin, M. Dagouassat, A. Berdeaux, E. Denamur, S. H. Crook, B. E. Mann, D. Scapens, P. Montravers, J. Boczkowski and R. Motterlini, *Antioxid. Redox Signaling*, 2012, **16**, 153–163.

110 A. J. Atkin, J. M. Lynam, B. E. Moulton, P. Sawle, R. Motterlini, N. M. Boyle, M. T. Pryce and I. J. S. Fairlamb, *Dalton Trans.*, 2011, **40**, 5755–5761.

111 J. Esteban, J. Ros-Lis, R. Martínez-Máñez, M. Marcos, M. Moragues, J. Soto and F. Sancenón, *Angew. Chem., Int. Ed.*, 2010, **49**, 4934–4937.

112 M. E. Moragues, J. Esteban, J. V. Ros-Lis, R. Martínez-Máñez, M. D. Marcos, M. Martínez, J. Soto and F. Sancenón, *J. Am. Chem. Soc.*, 2011, **133**, 15762–15772.

113 J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, Springer, New York, 2006.

114 B. Wang and E. V. Anslyn, *Chemosensors - Principles, Strategies, and Applications*, John Wiley & Sons, Hoboken, New Jersey, 2011.

115 V. S. Lin and C. J. Chang, *Curr. Opin. Chem. Biol.*, 2012, **16**, 595–601.

116 N. Kumar, V. Bhalla and M. Kumar, *Coord. Chem. Rev.*, 2013, **257**, 2335–2347.

117 J. Wang, J. Karpus, B. S. Zhao, Z. Luo, P. R. Chen and C. He, *Angew. Chem., Int. Ed.*, 2012, **51**, 9652–9656.

118 B. W. Michel, A. R. Lippert and C. J. Chang, *J. Am. Chem. Soc.*, 2012, **134**, 15668–15671.

119 A. R. Marques, L. Kromer, D. J. Gallo, N. Penacho, S. S. Rodrigues, J. D. Seixas, G. J. L. Bernardes, P. M. Reis, S. L. Otterbein, R. A. Ruggieri, A. S. G. Gonçalves, A. M. L. Gonçalves, M. N. D. Matos, I. Bento, L. E. Otterbein, W. A. Blättler and C. C. Romão, *Organometallics*, 2012, **31**, 5810–5822.

120 R. Motterlini, P. Sawle, J. Hammad, S. Bains, R. Alberto, R. Foresti and C. J. Green, *FASEB J.*, 2005, **19**, 284–286.

121 T. S. Pitchumony, B. Spingler, R. Motterlini and R. Alberto, *Chimia*, 2008, **62**, 277–279.

122 T. S. Pitchumony, B. Spingler, R. Motterlini and R. Alberto, *Org. Biomol. Chem.*, 2010, **8**, 4849–4854.

123 L. A. P. Antony, T. Slanina, P. Šebej, T. Šolomek and P. Klán, *Org. Lett.*, 2013, **15**, 4552–4555.

124 P. Peng, C. Wang, Z. Shi, V. K. Johns, L. Ma, J. Oyer, A. Copik, R. Igarashi and Y. Liao, *Org. Biomol. Chem.*, 2013, **11**, 6671–6674.

125 G. Gasser and N. Metzler-Nolte, *Curr. Opin. Chem. Biol.*, 2012, **16**, 84–91.

126 F. Zobi, *Future Med. Chem.*, 2013, **5**, 175–188.

127 W. A. Herrmann, *J. Organomet. Chem.*, 1990, **383**, 21–44.

128 R. Motterlini, J. E. Clark, R. Foresti, P. Sarathchandra, B. E. Mann and C. J. Green, *Circ. Res.*, 2002, **90**, e17–e24.

129 N. P. E. Barry and P. J. Sadler, *Chem. Commun.*, 2013, **49**, 5106–5131.

130 A. E. Pierri, A. Pallaoro, G. Wu and P. C. Ford, *J. Am. Chem. Soc.*, 2012, **134**, 18197–18200.

131 M. A. Gonzalez, S. J. Carrington, N. L. Fry, J. L. Martinez and P. K. Mascharak, *Inorg. Chem.*, 2012, **51**, 11930–11940.

132 M. A. Gonzalez, M. A. Yim, S. Cheng, A. Moyes, A. J. Hobbs and P. K. Mascharak, *Inorg. Chem.*, 2011, **51**, 601–608.

133 S. J. Carrington, I. Chakraborty and P. K. Mascharak, *Chem. Commun.*, 2013, **49**, 11254–11256.

134 D. E. Bikiel, E. G. Solyeira, F. D. Salvo, H. M. S. Milagre, M. N. Eberlin, R. S. Corrêa, J. Ellena, D. A. Estrin and F. Doctorovich, *Inorg. Chem.*, 2011, **50**, 2334–2345.

135 R. Kretschmer, G. Gessner, H. Görls, S. H. Heinemann and M. Westerhausen, *J. Inorg. Biochem.*, 2011, **105**, 6–9.

136 V. P. L. Velásquez, T. M. A. Jazzaï, A. Malassa, H. Görls, G. Gessner, S. H. Heinemann and M. Westerhausen, *Eur. J. Inorg. Chem.*, 2012, 1072–1078.

137 L. Hewison, T. R. Johnson, B. E. Mann, A. J. H. M. Meijer, P. Sawle and R. Motterlini, *Dalton Trans.*, 2011, **40**, 8328–8334.

138 C. Bischof, T. Joshi, A. Dimri, L. Spiccia and U. Schatzschneider, *Inorg. Chem.*, 2013, **52**, 9297–9308.

139 M. A. Gonzalez, S. J. Carrington, I. Chakraborty, M. M. Olmstead and P. K. Mascharak, *Inorg. Chem.*, 2013, **52**, 11320–11331.

140 C. S. Jackson, S. Schmitt, Q. P. Dou and J. J. Kodanko, *Inorg. Chem.*, 2011, **50**, 5336–5338.

141 A. J. Atkin, I. J. S. Fairlamb, J. S. Ward and J. M. Lynam, *Organometallics*, 2012, **31**, 5894–5902.

142 S. Romanski, H. Rücker, E. Stamellou, M. Guttentag, J.-M. Neudörfl, R. Alberto, S. Amslinger, B. Yard and H.-G. Schmalz, *Organometallics*, 2012, **31**, 5800–5809.

143 S. Romanski, B. Kraus, U. Schatzschneider, J.-M. Neudörfl, S. Amslinger and H.-G. Schmalz, *Angew. Chem., Int. Ed.*, 2011, **50**, 2392–2396.

144 L. Hewison, S. H. Crook, T. R. Johnson, B. E. Mann, H. Adams, S. E. Plant, P. Sawle and R. Motterlini, *Dalton Trans.*, 2010, **39**, 8967–8975.

145 D. Scapens, H. Adams, T. R. Johnson, B. E. Mann, P. Sawle, R. Aqil, T. Perri and R. Motterlini, *Dalton Trans.*, 2007, 4962–4973.

146 J. S. Ward, J. M. Lynam, J. W. B. Moir, D. E. Sanin, A. P. Mountford and I. J. S. Fairlamb, *Dalton Trans.*, 2012, **41**, 10514–10517.

147 P. Govender, S. Pai, U. Schatzschneider and G. S. Smith, *Inorg. Chem.*, 2013, **52**, 5470–5478.

148 W.-Q. Zhang, A. J. Atkin, R. J. Thatcher, A. C. Whitwood, I. J. S. Fairlamb and J. M. Lynam, *Dalton Trans.*, 2009, 4351–4358.

149 W.-Q. Zhang, A. C. Whitwood, I. J. S. Fairlamb and J. M. Lynam, *Inorg. Chem.*, 2010, **49**, 8941–8952.

150 F. Zobi, A. Degonda, M. C. Schaub and A. Y. Bogdanova, *Inorg. Chem.*, 2010, **49**, 7313–7322.

151 F. Zobi and O. Blacque, *Dalton Trans.*, 2011, **40**, 4994–5001.

152 M. A. Gonzalez, N. L. Fry, R. Burt, R. Davda, A. Hobbs and P. K. Mascharak, *Inorg. Chem.*, 2011, **50**, 3127–3134.

153 L. Long, X. Jiang, X. Wang, Z. Xiao and X. Liu, *Dalton Trans.*, 2013, **42**, 15663–15669.

154 R. D. Rimmer, H. Richter and P. C. Ford, *Inorg. Chem.*, 2010, **49**, 1180–1185.

155 G. R. Stephenson, P. W. Howard, D. A. Owen and A. J. Whitehead, *J. Chem. Soc., Chem. Commun.*, 1991, 641–642.

156 A. Bohac, M. Lettrichova, P. Hrnčiar and M. Hutta, *J. Organomet. Chem.*, 1996, **507**, 23–29.

157 S. Ban, H. Sakurai, Y. Hayashi and K. Narasaka, *Chem. Lett.*, 1997, 699–700.



158 S. Botov, E. Stamellou, S. Romanski, M. Guttentag, R. Alberto, J.-M. Neudorfl, B. Yard and H.-G. Schmalz, *Organometallics*, 2013, **32**, 3587–3594.

159 S. Romanski, B. Kraus, M. Guttentag, W. Schlundt, H. Rücker, A. Adler, J.-M. Neudörfl, R. Alberto, S. Amslinger and H.-G. Schmalz, *Dalton Trans.*, 2012, **41**, 13862–13875.

160 I. J. S. Fairlamb, S. M. Syvänen and A. C. Whitwood, *Synlett*, 2003, 1693–1697.

161 I. J. S. Fairlamb, A.-K. Duhme-Klair, J. M. Lynam, B. E. Moulton, C. T. O'Brien, P. Sawle, J. Hammad and R. Motterlini, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 995–998.

162 P. Sawle, J. Hammad, I. J. S. Fairlamb, B. Moulton, C. T. O'Brien, J. M. Lynam, A. K. Duhme-Klair, R. Foresti and R. Motterlini, *J. Pharmacol. Exp. Ther.*, 2006, **318**, 403–410.

163 I. J. S. Fairlamb, J. M. Lynam, B. E. Moulton, I. E. Taylor, A. K. Duhme-Klair, P. Sawle and R. Motterlini, *Dalton Trans.*, 2007, 3603–3605.

164 K. Meister, J. Niesel, U. Schatzschneider, N. Metzler-Nolte, D. A. Schmidt and M. Havenith, *Angew. Chem., Int. Ed.*, 2010, **49**, 3310–3312.

165 F. Mohr, J. Niesel, U. Schatzschneider and C. W. Lehmann, *Z. Anorg. Allg. Chem.*, 2012, **638**, 543–546.

166 T. Santos-Silva, A. Mukhopadhyay, J. D. Seixas, G. J. L. Bernardes, C. C. Romão and M. J. Romão, *J. Am. Chem. Soc.*, 2011, **133**, 1192–1195.

167 M. F. A. Santos, J. D. Seixas, A. C. Coelho, A. Mukhopadhyay, P. M. Reis, M. J. Romão, C. C. Romão and T. Santos-Silva, *J. Inorg. Biochem.*, 2012, **117**, 285–291.

168 T. M. A. Jazzazi, H. Görls, G. Gessner, S. H. Heinemann and M. Westerhausen, *J. Organomet. Chem.*, 2013, **733**, 63–70.

169 S. V. C. Vummella, D. Branduardi, M. Masetti, M. De Vivo, R. Motterlini and A. Cavalli, *Chem.–Eur. J.*, 2012, **18**, 9267–9275.

170 L. Hewison, S. H. Crook, B. E. Mann, A. J. H. M. Meijer, H. Adams, P. Sawle and R. A. Motterlini, *Organometallics*, 2012, **31**, 5823–5834.

171 H.-M. Berends and P. Kurz, *Inorg. Chim. Acta*, 2012, **380**, 141–147.

172 F. Zobi, O. Blacque, R. A. Jacobs, M. C. Schaub and A. Y. Bogdanova, *Dalton Trans.*, 2012, **41**, 370–378.

173 J. D. Seixas, A. Mukhopadhyay, T. Santos-Silva, L. E. Otterbein, D. J. Gallo, S. S. Rodrigues, B. H. Guerreiro, A. M. L. Goncalves, N. Penacho, A. R. Marques, A. C. Coelho, P. M. Reis, M. J. Romão and C. C. Romão, *Dalton Trans.*, 2013, **42**, 5985–5998.

174 T. R. Johnson, B. E. Mann, I. P. Teasdale, H. Adams, R. Foresti, C. J. Green and R. Motterlini, *Dalton Trans.*, 2007, 1500–1508.

175 L. Oresmaa, H. Tarvainen, K. Machal and M. Haukka, *Dalton Trans.*, 2012, **41**, 11170–11175.

176 G. Parkin, *Chem. Rev.*, 2004, **104**, 699–768.

177 H. Pfeiffer, A. Rojas, J. Niesel and U. Schatzschneider, *Dalton Trans.*, 2009, 4292–4298.

178 H. Pfeiffer, T. Sowik and U. Schatzschneider, *J. Organomet. Chem.*, 2013, **734**, 17–24.

179 J. B. Matson, M. J. Webber, V. K. Tamboli, B. Weber and S. I. Stupp, *Soft Matter*, 2012, **8**, 6689–6692.

180 J. L. Wilson, H. E. Jesse, B. Hughes, V. Lund, K. Naylor, K. S. Davidge, G. M. Cook, B. E. Mann and R. K. Poole, *Antioxid. Redox Signaling*, 2013, **19**, 497–509.

181 G. L. Patrick, *An Introduction to Medicinal Chemistry*, Oxford University Press, 2013.

182 T. Desmet, W. Soetaert, P. Bojarová, V. Kren, L. Dijkhuizen, V. Eastwick-Field and A. Schiller, *Chem.–Eur. J.*, 2012, **18**, 10786–10801.

183 I. Neundorf, J. Hoyer, K. Splith, R. Rennert, H. W. P. N'Dongo and U. Schatzschneider, *Chem. Commun.*, 2008, 5604–5606.

184 K. Splith, I. Neundorf, W. Hu, H. W. P. N'Dongo, V. Vasylyeva, K. Merz and U. Schatzschneider, *Dalton Trans.*, 2010, **39**, 2536–2545.

185 A. Schroeder, D. A. Heller, M. M. Winslow, J. E. Dahlman, G. W. Pratt, R. Langer, T. Jacks and D. G. Anderson, *Nat. Rev. Cancer*, 2012, **12**, 39–50.

186 L. Y. T. Chou, K. Ming and W. C. W. Chan, *Chem. Soc. Rev.*, 2011, **40**, 233–245.

187 F. H. Schacher, P. A. Rupar and I. Manners, *Angew. Chem., Int. Ed.*, 2012, **51**, 7898–7921.

188 P. C. Kunz, H. Meyer, J. Barthel, S. Sollazzo, A. M. Schmidt and C. Janiak, *Chem. Commun.*, 2013, **49**, 4896–4898.

189 T. R. Cook, Y.-R. Zheng and P. J. Stang, *Chem. Rev.*, 2013, **113**, 734–777.

190 M. Eddaoudi, D. B. Moler, H. Li, B. Chen, T. M. Reineke, M. O'Keeffe and O. M. Yaghi, *Acc. Chem. Res.*, 2001, **34**, 319–330.

191 S. K. Henninger, F. Jeremias, H. Kummer and C. Janiak, *Eur. J. Inorg. Chem.*, 2012, 2625–2634.

192 C. Janiak and J. K. Vieth, *New J. Chem.*, 2010, **34**, 2366–2388.

193 M. Ma, H. Noei, B. Mienert, J. Niesel, E. Bill, M. Muhler, R. A. Fischer, Y. Wang, U. Schatzschneider and N. Metzler-Nolte, *Chem.–Eur. J.*, 2013, **19**, 6785–6790.

194 G. Dördelmann, T. Meinhardt, T. Sowik, A. Krueger and U. Schatzschneider, *Chem. Commun.*, 2012, **48**, 11528–11530.

195 G. Dördelmann, H. Pfeiffer, A. Birkner and U. Schatzschneider, *Inorg. Chem.*, 2011, **50**, 4362–4367.

196 N. E. Brückmann, M. Wahl, G. J. Reiß, M. Kohns, W. Wätjen and P. C. Kunz, *Eur. J. Inorg. Chem.*, 2011, 4571–4577.

197 E. Tfouni, F. G. Doro, A. J. Gomes, R. S. d. Silva, G. Metzker, P. G. Z. Benini and D. W. Franco, *Coord. Chem. Rev.*, 2010, **254**, 355–371.

198 D. Bechet, P. Couleaud, C. Frochot, M.-L. Viriot, F. Guillemin and M. Barberi-Heyob, *Trends Biotechnol.*, 2008, **26**, 612–621.

199 K. A. Wold, V. B. Damodaran, L. A. Suazo, R. A. Bowen and M. M. Reynolds, *ACS Appl. Mater. Interfaces*, 2012, **4**, 3022–3030.

200 C. Bohlender, M. Wolfram, H. Goerls, W. Imhof, R. Menzel, A. Baumgaertel, U. S. Schubert, U. Mueller, M. Frigge, M. Schnabelrauch, R. Wyrwa and A. Schiller, *J. Mater. Chem.*, 2012, **22**, 8785–8792.

201 C. Bohlender, K. Landfester, D. Crespy and A. Schiller, *Part. Part. Syst. Charact.*, 2013, **30**, 138–142.

202 R. Wyrwa, M. Schnabelrauch, C. Altmann and A. Schiller, *Kohlenstoffmonoxid freisetzende Materialien und deren Verwendung*, German Pat., DE10 2012 04 132.2, 2012.

203 C. Bohlender, S. Gläser, M. Klein, J. Weisser, S. Thein, U. Neugebauer, J. Popp, R. Wyrwa and A. Schiller, *J. Mater. Chem. B*, 2014, DOI: 10.1039/C3TB21649G.

204 K. Landfester, *Annu. Rev. Mater. Res.*, 2006, **36**, 231–279.

205 A. Greiner and J. Wendorff, *Angew. Chem., Int. Ed.*, 2007, **46**, 5670–5703.

