Streptavidin binding as a model to characterize thiol–ene chemistry-based polyamine surfaces for reversible photonic protein biosensing†

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Biotin- and iminobiotin-bonded surfaces obtained by thiol–ene chemistry and subsequent modification with polyamines were characterized with respect to streptavidin-binding capacity and reversibility for photonic biosensing using X-ray photoelectron spectroscopy and Mach–Zehnder-interferometric sensors. The streptavidin–iminobiotin system was exploited for reversible multilayer deposition and determination of affinity constants on each layer.

Kinetic studies of receptor–protein interactions play a pivotal role in medical research and drug development. Comprehensive analyses of kinetic parameters necessitate measurements for a large variety of receptors and proteins at different levels of concentrations. In order to facilitate these analyses, a surface modification approach is desirable that provides high binding capacity and offers the possibility of repeated measurements on a single regenerative sensor array.

One promising strategy to obtain high binding capacities is the use of a multifunctional polymer grafting layer that increases the density of surface functionalities (e.g. amines) and allows immobilizing large amounts of receptors. The surface grafting of polymers can be performed by non-covalent adsorption.1 However, this causes a collapse of the 3D polymer shape.2 The goal of this work is to extend flat planar 2D surface modifications to voluminous 3D surface layers, which are not broadly utilized although they may be promising strategies to enhance saturation capacities. For biosensing, the globular-shaped dendrimers2 and linear-shaped brush polymers3 are of interest to gain well orientated surface receptors or to form voluminous 3D surface layers, respectively. During the surface grafting process, the primary shape of the polymer has to be preserved and, therefore, covalent immobilization strategies are more promising than non-covalent adsorption methods.2 For the covalent surface grafting, we investigated thiol–ene coupling-based surface modifications with subsequent derivatization of the resulting carboxylic groups using selected polyamine-type polymers. The thiol–ene coupling was chosen owing to its mild reaction conditions, quantitative yields, and its feasibility in the presence of oxygen.4 In addition, it allows a spatially controlled surface modification employing optical lithography or laser writing with feature sizes down to the micron range.5

In this study, we selected polyamidoamines (PAMAM) G1 and G5, as well as polyethyleneimine (PEI, 750 kDa), to compare dendrimers that have significantly different hydrodynamic diameters with a more cost-effective brush polymer. These polyamines were systematically characterized with respect to their chemical composition using X-ray photoelectron spectroscopy (XPS) and with respect to their binding capacity and reversibility using two streptavidin-binding models based on biotin and iminobiotin (see Fig. 1). On the basis of these results, we devised a novel reversible multilayer deposition concept which allows repeated determination of the association constant either at each layer or as an enhanced cumulative signal.

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Fig. 1 Surface modification steps of crystalline silicon (cSi) and amorphous hydrogenated silicon (a-Si:H): (1) activation by thiol–ene coupling; (2) EDC reaction for the covalent binding of PAMAM G1, G5 and PEI. The resultant sensor surfaces were characterized by XPS and streptavidin biosensing. Total sensor preparation time is ~ 6 h.
As a testing platform, we employed silicon-based photonic sensors, which have been demonstrated to be an attractive means to perform kinetic studies. The sensing principle relies on optical wave guiding due to total internal reflection. A part of the optical field of the guided light—the so-called evanescent field—penetrates into the surrounding media and decays exponentially. Typical penetration depths of the evanescent field are in the range of 60–150 nm for light with a wavelength of 1.3 μm. Binding of the target molecule to the functionalized waveguide surface locally changes the refractive index, which induces a change of the phase velocity of the guided light. We use amorphous hydrogenated silicon (a-Si:H) based Mach–Zehnder interferometric (MZI) sensors (see Fig. S1, ESI†) to translate the change of the phase velocity into a modulation of the optical output power. From this sinusoidal optical output an accumulated phase shift (ΔΦ) is calculated, which correlates with the amount of target molecules captured on the surface. The silicon-based photonic biosensors can be made of different materials (e.g. a-Si:H, crystalline silicon, silicon nitride). One prominent method for functionalizing these materials is surface modification using self-assembled functional silane layers. In this study, we performed this silanization using 3-mercaptopropyltrimethoxysilane (MPTMS) on two specific types of silicon—hydrogenated amorphous silicon and undoped crystalline (100) silicon (cSi). The optimized silanization procedures for these substrates resulted in significantly higher surface densities of reactive sulfhydryl groups for a-Si:H (Table S1, ESI†). This preliminary study emphasizes the need for a subsequent surface modification procedure that ensures equally high surface concentrations of functional groups irrespective of the substrate. We investigated whether thiol–ene coupling combined with the selected polyamines can provide an equalization of the functional group density on a-Si:H and cSi. To achieve the covalent binding of the selected multifunctional polymers, the stable MPTMS-derivatized silicon surfaces were activated by photo-induced thiol–ene coupling. This radical addition reaction between surface-bound thiol groups and an activated double bond between the ethylenediamine moiety and PEI (see Fig. 1). To analyse the polyamine surfaces with respect to binding capacity, reversibility, and kinetic behaviour, measurements on a-Si:H MZI sensors were performed using a streptavidin-binding model based on biotin and iminobiotin. The biotin–streptavidin interaction is one of the strongest known in nature with a dissociation constant of $K_D = 10^{-15}$ M, which under mild conditions prevents reversibility of binding. The analogue iminobiotin has a weaker and pH-dependent association to streptavidin due to its guanidinium moiety in the bicyclic ring replacing the urea functionality. Owing to these binding characteristics, biotin was used to perform the protein binding capacity, and iminobiotin to perform

Table 1: XPS (Mg Kα, 1253.6 eV, take-off angle θ = 0°) data including the atomic composition, reactive primary amino group concentration and modification thickness d of EDA, PAMAM G1 and G5, PEI surfaces

<table>
<thead>
<tr>
<th>Atomic composition (%)</th>
<th>EDA</th>
<th>PAMAM G1</th>
<th>PAMAM G5</th>
<th>PEI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Si2p</td>
<td>37.7</td>
<td>49.0</td>
<td>33.8</td>
<td>39.5</td>
</tr>
<tr>
<td>O1s</td>
<td>37.0</td>
<td>35.8</td>
<td>32.3</td>
<td>32.1</td>
</tr>
<tr>
<td>C1s</td>
<td>13.9</td>
<td>12.5</td>
<td>20.8</td>
<td>24.0</td>
</tr>
<tr>
<td>N1s</td>
<td>11.5</td>
<td>2.7</td>
<td>13.1</td>
<td>4.3</td>
</tr>
<tr>
<td>R-NH$_2$ (%)</td>
<td>1.4</td>
<td>0.9</td>
<td>2.2</td>
<td>2.4</td>
</tr>
<tr>
<td>d (nm)</td>
<td>0.46</td>
<td>0.33</td>
<td>0.91</td>
<td>1.09</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1.09</td>
<td>3.9</td>
<td>3.8</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>3.8</td>
<td>6.6</td>
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<tr>
<td></td>
<td></td>
<td></td>
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<td>6.5</td>
</tr>
</tbody>
</table>

$\lambda$ is calculated using $d = -\lambda \ln(I/I_0) \cos \theta$, where $I$ and $I_0$ are the Si2p peak intensities of the functionalized and non-modified surfaces, respectively, measured for a-Si:H and cSi. From the XPS measurement results summarized in Table 1, it can be concluded that PAMAM G5 and PEI provide the best conditions for protein sensing in terms of modification thickness and primary amino group concentration.

To analyse the polyamine surfaces with respect to binding capacity, reversibility, and kinetic behaviour, measurements on a-Si:H MZI sensors were performed using a streptavidin-binding model based on biotin and iminobiotin. The biotin–streptavidin interaction is one of the strongest known in nature with a dissociation constant of $K_D = 10^{-15}$ M, which under mild conditions prevents reversibility of binding. The analogue iminobiotin has a weaker and pH-dependent association to streptavidin due to its guanidinium moiety in the bicyclic ring replacing the urea functionality. Owing to these binding characteristics, biotin was used to perform the protein binding capacity, and iminobiotin to perform
reversible measurements. For this purpose, the polyamines were (iminobiotinylated using NHS derivatives (S6, ESI†). Before the measurements, the (iminobiotinylated sensor surfaces were rinsed with bovine serum albumin (100 μg ml⁻¹ in 15 mM PBS, 10 min) to block low energy binding sites and suppress non-specific interactions on the surfaces. The MZI sensor measurements were performed as fast screening experiments at high streptavidin concentration (1666 pmol ml⁻¹) to obtain maximum signal output in reasonable time. Physiological conditions (PBS 15 mM, pH 7.5) were used to comply with biosensing requirements. The biotinylated sensors show an increased response from in reasonable time. Physiological conditions (PBS 15 mM, pH 7.5) (Fig. 2). After about 1500 s the streptavidin solution was replaced by a plain buffer solution. As expected, no dissociation of streptavidin occurs on biotinylated surfaces. In contrast, reversible binding of streptavidin was possible on the iminobiotinylated polyamine-modified MZI sensors. Measurements using these sensors were performed by a sequence of four consecutive binding (with 1666 pmol ml⁻¹ streptavidin solution), dissociation (with buffer rinsing) and regeneration experiments. Complete regeneration of the surface was realized by rinsing with 3.5 mM hydrochloric acid (5 min). The signal increases linearly from the first to the fifth bilayer (Fig. S8, ESI†), which indicates a concentration dependent growth behaviour and provides an enhancement of the sensor response. To determine the association constant, the sensor response obtained at a certain number of bilayer stacks was plotted against the different streptavidin concentrations (see Fig. 3A). The association constants were then calculated using the Langmuir isotherm:

\[
S = S_{\text{max}}C/(1/K_A + C)
\]

where \(S\) is the actual and \(S_{\text{max}}\) is the maximum signal at infinitely high concentrations in one particular bilayer stack, \(C\) is the streptavidin concentration in the feed solution, and \(K_A\) is the association constant. The determination of the association constant shows that the streptavidin affinity to the imino-biotinylated surface decreases from the first \((K_A = 2.2 \times 10^6 \text{ M}^{-1})\) to the third \((K_A = 5.7 \times 10^5 \text{ M}^{-1})\) bilayer stack and then stays almost constant (Fig. 3B) (Table S5, ESI†). This behaviour indicates that the STA–PEI-IB bilayer stack deposition leads to a change of the surface-binding properties (e.g. sterical arrangement or concentration of iminobiotin) until a self-assembled equilibrium is obtained. In the literature, streptavidin–iminobiotin association constants were determined in buffer solutions (pH 7.5) by Green¹² and Raphael et al.¹³ with \(K_A\) values of 1.26 × 10⁸ M⁻¹ and 1.58 × 10⁷ M⁻¹, respectively.

In conclusion, the characterization of the thiol–ene chemistry based multifunctional polyamine surfaces clearly documented the advantages of PEI modifications for protein sensing. The PEI polymer is not as highly ordered as the PAMAM G5 dendrimer, but leads to higher surface amino group concentrations and
protein loading capacities. Furthermore, the properties of iminobiotinylated PEI surfaces can be utilized for reversible biosensing, as shown in this study. The reversible streptavidin--iminobiotinylated PEI chain bilayer stack deposition at different streptavidin concentrations allowed the determination of the association constant after each bilayer deposition on a single iminobiotinylated photonic sensor. The determined association constants are in good agreement with the values in the literature. These results clearly demonstrate the suitability of reversible bilayer stack deposition for the repeated determination of binding constants on photonic sensors at enhanced sensor signals.

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Notes and references