

A divalent protecting group for benzoxaboroles†

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1-Dimethylamino-8-methylaminonaphthalene is put forth as a protecting group for benzoxaboroles. The ensuing complex is fluorescent, charge-neutral, highly stable under basic conditions, stable to anhydrous acid, and readily cleavable in aqueous acid to return the free benzoxaborole.

Oxaborole heterocycles are boronic acids that are receiving much attention for applications in drug discovery,¹ synthetic methodology,² molecular recognition,³ and biotechnology.⁴ Benzoxaborole (**1**; Fig. 1),⁵ which is characterized by a phenyl ring fused to a five-membered oxaborole, is the most widely employed oxaborole pharmacophore in medicinal chemistry.¹ Compared to phenylboronic acid, the annulated benzylic alcohol in **1** confers high stability,⁶ low pK_a,⁷ and superior polyol-binding (**1** → **2**) under physiological conditions (*i.e.*, water near neutral pH),^{3a,b,f,8} favorable attributes for applications that entail binding to carbohydrates.^{3,4}

The vacant p-orbital on boron—essential for complexation with polyols—can confound multistep synthetic routes and the purification/isolation of derivatives. This issue is compounded further by the commercial availability of countless elaborately functionalized boronic acids but only a small number of simple benzoxaboroles. Our interest in benzoxaboroles for the cellular delivery of chemotherapeutic agents^{4a} and other pharmacological applications drove us to develop a protecting group for **1** that would enhance its synthetic utility.

A variety of successful protecting group strategies have been developed to modulate the undesired reactivity of boronic acids. A common example is a pinacol ester (**3**), which sterically shields the p-orbital from reaction. Similarly, Suginome and coworkers demonstrated the reduced reactivity of boronic acids in a complex with 1,8-diaminonaphthalene (**4**).⁹ An often-employed strategy developed by Molander and coworkers capitalizes on the fluoro-affinity of boron to form a trifluoroboronate salt (**5**).¹⁰ While highly stable, these salts are incompatible with chromatography, limiting their utility in multistep synthetic routes. Burke and coworkers have popularized a trivalent *N*-methyliminodiacetic acid (MIDA)¹¹ ligand for boronic acids that coordinates the vacant p-orbital with a trialkylamine through a dative bond to give a charge-neutral complex (**6**).¹² This complex is broadly compatible with synthetic reagents and chromatographic purification.¹³ The trivalent MIDA group is not, however, suitable for protecting **1**, which can coordinate to only two ligands. Divalent protecting

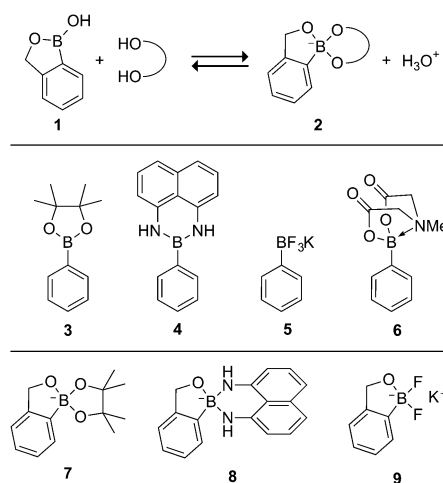


Fig. 1 Structure of free benzoxaborole (**1**), its complexation with a polyol (**2**), and other general structures of protected boronic acids (**3–6**) and their unsuitable complexes (**7–9**) with **1**.

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groups like **3** and **4** are also not appropriate for **1** because they would lead to anionic complexes (**7** and **8**). Likewise, fluoride protection would yield an anionic difluoroborate salt (**9**).

We sought to apply the principle of charge neutrality, as demonstrated by the efficacious MIDA protecting group, to the divalent protection of benzoxaboroles. Inspired by the strong complexes that simple boronic acids form with 1,8-diaminonaphthalene (**4**),⁹ we devised a protecting group based on 1,8-bis(dimethylamino)naphthalene¹⁴ (Proton-sponge®) to generate charge-neutral complexes with **1** that maintain a Lewis basic site to promote acid-mediated cleavage (Scheme 1). We found that complexes **11a–e** were fluorescent, stable to basic conditions, moderately stable to anhydrous acid, and readily deprotected under aqueous acidic conditions.

Following a previous report,¹⁵ 1,8-bis(dimethylamino)naphthalene was mono-demethylated to give 1-dimethylamino-8-methylaminonaphthalene (**10**) in one step of >90% yield without chromatography. After azeotropic removal of water, **1a–e** and **10** readily formed complexes **11a–e** (Scheme 1). Excess **10** was necessary for high yields, but unreacted **10** was re-isolated quantitatively by chromatography (Fig. 2B, inset). Formation of the complex generated a tetrahedral boron center (**1c** → **11c** ¹¹B NMR δ 33.2 sp² → δ 9.1 sp³)¹¹ that was stereogenic—the benzylic protons of **1** became nonequivalent (Fig. 2A) and served as signature of complex formation. The complex was characterized with X-ray crystallography (Fig. 2C).

Next, we investigated the generality of our protecting group design. First, we investigated a small library of compounds similar to **10** for protection of **1a** (see ESI†). Only **10**, however, led to complexes that were stable during chromatography. α -Amino acids (which resemble divalent versions of MIDA) are known to form stable complexes with dialkyl boranes (*i.e.*, borinic acids).¹⁶ We found these complexes to be too fragile for effective protection of **1**. From these data, we conclude that the stability of complexes **11a–e** stems from both the rigid structure imposed by the diaminonaphthalene ring and the strongly donating nitrogen ligands. The ease of purification also relies on coordinating the vacant p-orbital through a dative bond supplied by the dimethyl amino ligand to create a charge-neutral complex. Finally, we note that **11a–e** are highly fluorescent (**11a**, $\Phi_F = 0.45$, Fig. 2B and inset) allowing for easy tracking of product derivatives (**11–18**) during multi-step synthesis and purification using a standard long-wave (365 nm) bench-top lamp.¹⁷ Complex **11a** was subjected to a screen of conditions to determine its stability (Table 1, representative time points, see ESI† for kinetic traces and further discussion). In general, **11a** is deprotected readily under aqueous acidic

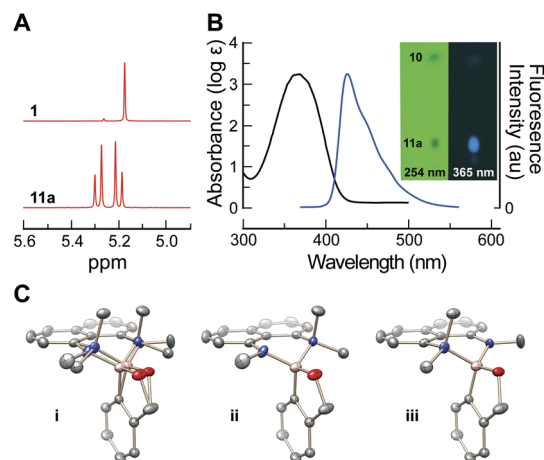


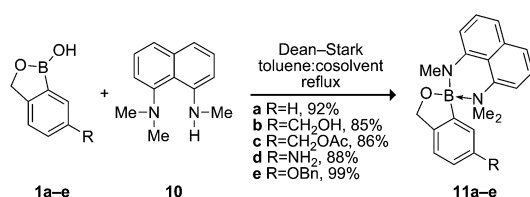
Fig. 2 Characterization of complex **11a**. (A) ¹H NMR spectra of **1a** and **11a** showing the splitting of benzylic proton signals in **1** into two diastereotopic doublets. (B) Absorbance (black) and fluorescence (blue) spectra of **11a** in CHCl₃. Inset: silica gel thin-layer chromatography plate (0.5% v/v MeOH in DCM, *R_f* = 0.7 separation between **10** and **11a**) illuminated under short (254 nm) and long (365 nm) wavelength light. (C) X-ray crystal structure of racemic **11a** with anisotropic thermal ellipsoids set at 50% probability and hydrogen atoms omitted. (i) **11a** showing both components of positional disorder. (ii) and (iii) Disorder components (enantiomers) separated for clarity.

Table 1 Screening of the stability of **11a** under various conditions^a

Entry	Conditions	% Cleaved	Time (h)
1	0.5 M HCl in (THF–H ₂ O, 1 : 1) ^b	94	4
2	0.5 M TFA in (THF–H ₂ O, 1 : 1) ^b	99	4
3	0.5 M AcOH in (THF–H ₂ O, 1 : 1) ^b	90	4
4	0.5 M TFA in DCM ^b	82	4
5	0.5 M BF ₃ OEt ₂ in DCM ^b	43	4
6	4 M HCl in dioxane ^b	30	4
7	0.1 M pH = 7 PBS in THF (1 : 1) ^c	0	36
8	0.5 M NaOH in (THF–H ₂ O, 1 : 1) ^c	0	72
9	20% v/v piperidine ^c in CH ₂ Cl ₂	0	72
10	20% v/v DIEA ^c in CH ₂ Cl ₂	0	72
11	0.5 M LiAlH ₄ in THF ^{b,c}	0	2

^a Average of two experiments at 0.1 M **11a** with 4-bromo-2,6-dimethylaniline as an internal standard. ^b Analyzed with LC-MS.

^c Analyzed with ¹H NMR spectroscopy.



Scheme 1 Protection of benzoxaborole derivatives.

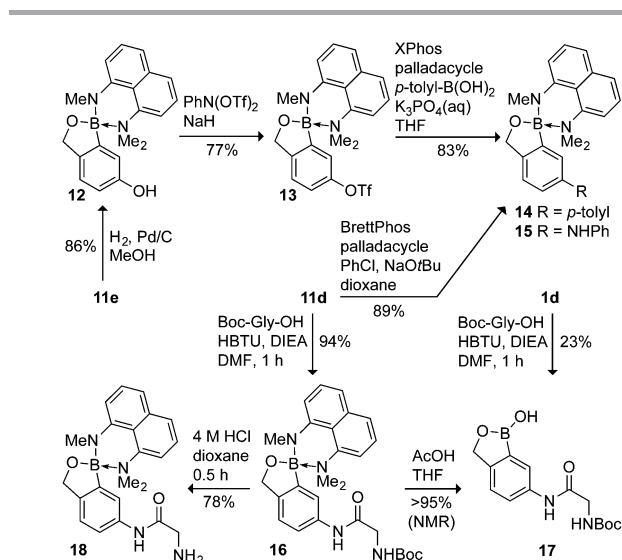
conditions (entries 1–3), but exhibits modest (*i.e.*, kinetic) stability under anhydrous acid (entries 4–6) (Table 1). This sensitivity to acid affirms an aspect of our design—protonation of the methylamino group leads to weaker binding and allows for hydrolytic cleavage. In contrast, **11a** is highly stable under basic conditions (entries 7–10). This reactivity indicates that protecting group **10** is well suited for solid-phase peptide synthesis,^{3c} in which amino acids are coupled under basic conditions and subsequently cleaved from a solid-support using acid. Finally, the complex tolerated strong reducing reagents, such as LiAlH₄ (entry 11). We do note that oxidative conditions were not compatible with **11b** and lead to complex mixtures. This sensitivity is not surprising due to the electron-rich nature of the diaminonaphthalene moiety, which is prone to oxidation.¹⁸



To explore further the stability of the protecting group, we evaluated **11b**, **11d**, and **11e** under a series of synthetic transformations (Scheme 2 and 3, see ESI† for **11b**). As expected, the complex was able to tolerate reducing conditions such as H_2/Pd (**11e** → **12**). Conversion of **12** to triflate **13** and subsequent Suzuki–Miyaura coupling using an XPhos palladacyclic pre-catalyst,¹⁹ provided **14** in high yield. Notably, **10** prevented reaction at the benzoxaborole center.²⁰ Recent data suggest that Suzuki–Miyaura reactions necessitate a vacant boron p-orbital,²¹ validating our protecting group design.²²

The base stability of the complex allowed for Buchwald–Hartwig C–N cross-coupling of the protected boron center to give **15** from **11d**.²³ As noted above, protecting group **10** is compatible with organic amine bases associated with peptide bond formation reagents to provide **16** in high yield (94%). In comparison, a similar peptide coupling reaction attempted with unprotected **1d** provided only a 23% yield of the amide **18**. Notably, in both the Pd-catalyzed and peptide-coupling reactions, the methylamino groups in **11d** are unreactive. Finally, evincing the utility of the protecting group under anhydrous acid, selective deprotection of the amino group in **16** with HCl in dioxane gave **17** in good yield (78%), while the benzoxaborole moiety could be deprotected selectively with aqueous acetic acid to return **18** (Scheme 2).

In conclusion, benzoxaborole **1** has become a privileged entity in medicinal chemistry and for carbohydrate recognition. Its continued development will rely on the efficiency of its derivatization. We have demonstrated the stability and utility of **1** and other benzoxaboroles when protected with **10**. Protecting group **10** occupies the vacant p-orbital on boron while creating charge neutrality in the final product. Its complexes are formed readily and are compatible with synthetic reagents and separation/purification techniques employed commonly in multi-step syntheses to enable or improve the efficiency of manipulating benzoxaborole.



Scheme 2 Synthetic evaluation of protected benzoxaborole derivatives; yields are isolated but not optimized.

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