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The chemistry and biological activity of the Hyacinthaceae



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The chemistry and biological activity of the Hyacinthaceae

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This review reflects a (bracketed)† family and subfamily taxonomic arrangement for the Hyacinthaceae *sensu* APGII.² A subsequent arrangement (APGIII)³ based on further molecular analyses is not here adopted, where the Hyacinthaceae was hypothetically included in an expanded circumscription of the Asparagaceae. In that the group of plants here dealt with are monophyletic,⁴ the systematic approach adopted for delimiting and naming family, subfamily or tribal level classifications ultimately has little bearing in relation to this review. We have adopted largely the phylogenies and generic concepts of Speta,⁵ Pfosser and Speta,⁴ and

subsequent workers (Martínez-Azorín *et al.*)⁶ who have considered molecular, morphological and biogeographic characters in their circumscriptions. Ali Syed *et al.*⁷ have provided an excellent overview of alternative and often controversial systematic arrangements, of primary and secondary centres of geographic diversity, and the distribution of subfamilies and tribes at a global level. It should be noted that such is the state of flux of generic circumscription in the Hyacinthaceae that when providing a subfamilial classification for this unit (Asparagaceae subfamily Scilloideae *sensu* APGIII) Chase *et al.*⁸ did not list genera names, which they otherwise offered with most other expanded Asparagalean families they treated. Given the past nomenclatural instability of the Hyacinthaceae, the names of plants under which the original phytochemical and pharmacological work was conducted often differs radically from the names accepted currently. For this reason we here provide both current and reported names, with species authors, to make clear this link. The literally reported (and sometimes incorrectly spelt) taxon names and species authors (if given) are provided in parentheses. The subfamilies are discussed in turn, starting with the Hyacinthoideae as most work has been conducted on this, the largest, subfamily. The Urgineoideae and finally the Ornithogaloideae follow. No reported investigations into the Oziroëoideae appear in the literature. Within each subfamily, those species receiving a substantial amount of attention will be discussed first, followed by the genera grouped by region (Europe and the Mediterranean, Middle and Far East and North Africa, southern Africa and finally, in the case of the Urgineoideae, India).

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† 55 families under APGII came to be known as 'bracketed families'. These were optional segregates of families that could be circumscribed in a larger sense.



Plants have been considered on a regional basis as in many cases the secondary metabolites found reflect a strong geographic pattern. Any early work on a genus is discussed before later work.

1 The Hyacinthoideae

The subfamily Hyacinthoideae comprises more than 400 species in thirty-eight genera.^{5,7} The Hyacinthoideae are distributed in sub-Saharan Africa, India, Madagascar, eastern Asia, the Mediterranean region and Eurasia. Sub-Saharan Africa appears to be the centre of origin for the subfamily, with a secondary centre of diversification and radiation being the Mediterranean region.⁷ Twenty genera of the Hyacinthoideae have been investigated, with the bulbs receiving most attention. The exception is *Hyacinthus orientalis* L., the flowers of which have been extensively examined. Homoisoﬂavanones and spirocyclic nortriterpenoids are widespread throughout the subfamily with chalcones, xanthenes, ﬂavonoids, alkaloids, stilbenoids, chromones, norlignans and benzopyranones also being reported.



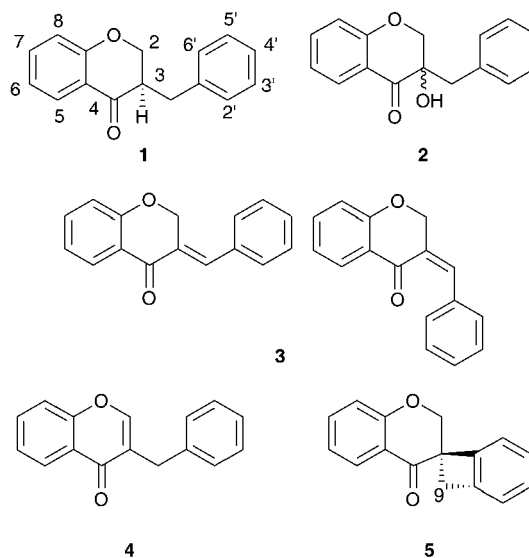
Dulcie Mulholland leads the Natural Products Research Group and is Head of the Department of Chemistry at the University of Surrey. She undertook her PhD at the University of Natal, Durban, South Africa, under the supervision of David Taylor on the Chemistry of the Meliaceae. For the last 20 years she has collaborated with Neil Crouch investigating the phytochemistry of the Hyacinthaceae of southern Africa.



Sianne Schwikkard obtained her PhD from the University of Kwazulu-Natal, South Africa in 1998 (under the supervision of Professor Dulcie Mulholland). This was followed by post-doctoral studies at Virginia Tech with Professor David Kingston and further post-doctoral studies at the Rand Academic University with Professor Fanie van Heerden. After spending 18 months working for Sasol Ltd,

Sianne relocated to the United Kingdom and following a post-doctoral position, under Professor Keith Jones at Kingston University, has continued to teach in a part-time capacity at Kingston University in addition to her position as a part-time research associate at the University of Surrey.

Abegaz *et al.*⁹ and du Toit *et al.*¹⁰ have reviewed the phytochemistry, biological activity, biosynthesis and synthesis of the homoisoﬂavanones. The homoisoﬂavanones can be divided into five categories, the 3-benzyl-4-chromanones **1**, the 3-hydroxy-3-benzyl-4-chromanones **2**, the 3-benzylidene-4-chromanones (*E* or *Z*) **3**, the 3-benzyl-chrom-2-en-4-ones **4** and the scillascillins **5**. The spirocyclic nortriterpenoids are generally of the lanosterol-type, showing a wide range of substitution patterns. The genera receiving the most attention, *Eucomis* L'Hér., *Muscari* Mill. and *Ledebouria* Roth. will be discussed first, followed by the lesser-studied Eurasian and Mediterranean genera *Leopoldia* Parl., *Scilla* L., *Barnardia* Lindl., *Othocallis* Salisb., *Oncostema* Raf., *Hyacinthus* L., *Bellevalia* Lapeyr., *Autonoe* (Webb & Berthel.) Speta and *Hyacinthoides* Medik. Lastly, the southern African genera *Drimiopsis* Lindl. & Paxton, *Resnova* van der Merwe, *Veltheimia* Gled., *Merwillia* Speta, *Lachenalia* J.Jacq. ex Murray,



Neil Crouch PhD (Natal) has worked as an ethnobotanist for the South African National Biodiversity Institute (SANBI) for nearly 20 years, largely on Zulu traditional medicine. He networks SANBI in natural products, pharmacological, horticultural, conservation, and Indigenous Knowledge Systems cataloguing programmes and is accordingly involved in state-sponsored bioprospecting consortia that develop new drugs for malaria and tuberculosis. As an Honorary Professor in the School of Chemistry and Physics at the University of Kwazulu-Natal, he explores his strong interest in the chemistry, traditional use and biosystematics of the Hyacinthaceae, a plant family with a major centre of diversity in southern Africa.

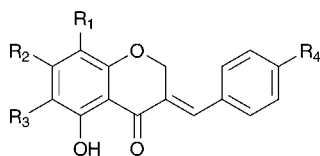
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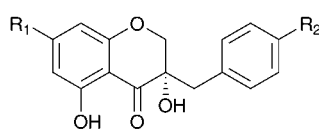
Pseudoprospero Speta, *Schizocarphus* van der Merwe and *Spetaea* Wetschnig & Pfosser are dealt with.

The genus *Eucomis* is widespread throughout eastern and southern Africa. It is used by traditional healers to treat a wide range of complaints, including rheumatism, as an anti-inflammatory and as a treatment for mental disease. Homoisoflavanones and triterpenoid glycosides have been isolated from the bulbs of many of the species investigated, with the homoisoflavanones usually accumulating in the waxy layer between the storage leaves of the bulb and the terpenoids in the bulb tissue.¹¹ Early work on *Eucomis bicolor* Baker resulted in the isolation of the yellow crystalline needles of eucomin **6** and the colourless hexagonal plates of eucomol **7**.¹² The configuration at C-3 of eucomol **7** was determined by CD experiments and confirmed by X-ray analysis of the *p*-bromophenacyl derivative.¹³ Four additional homoisoflavanoids have been isolated from the bulbs of *E. bicolor*, (**8–11**)^{14,15} followed a number of years later by the identification of seven triterpenoid oligosaccharides (**12–18**).^{16,17} Scillasaponin A **12** has a modified spirocyclic side chain at C-17 and C-23 and has been found to inhibit cyclic AMP phosphodiesterase (IC₅₀ = 11.5 × 10⁻⁵ M).¹⁶ Compounds **13**, **14** and **16** were found to be toxic to HeLa cells at concentrations of both 50 μg ml⁻¹ and 5 μg ml⁻¹.¹⁷ A later investigation of *E. bicolor* resulted in the isolation of twelve compounds from the dichloromethane extract and ten from the methanol extract. The dichloromethane extract yielded the seven known compounds (**7**, **10**, **19**, **20**, **21**, **22** and **23**) and five novel triterpenoids (**24** [the aglycone of **25**], **26**, **27**, **28** and **29**). The methanol extract was found to contain the known homoisoflavanoid **30** as well as nine novel lanosterol glycosides (**31–39**).¹⁸

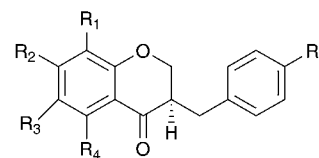
Eucomis autumnalis (Mill.) Chitt. (*syn. Eucomis undulata* Aiton) bulbs are used in the Eastern Cape of South Africa to treat cancer,¹⁹ and are used by Zulu traditional healers as an emetic and enema to treat fevers, as well as for the treatment of skin infections, biliousness, and urinary and respiratory infections. It is also used as a charm against witchcraft.²⁰ Phytochemical investigations of the bulbs of *E. autumnalis* have resulted in the isolation of three homoisoflavanones (**40–42**),^{14,21,22} two dibenzo- α -pyrones (**43–44**),²³ two spirocyclic nortriterpenoids (**20** and **45**) and an acid, compound **46**.²⁴ The structures assigned to compounds **43** and **44** were subsequently revised and reported to be the corresponding xanthenes.²⁵ The proposed structure for eucosterol **20** was confirmed by the X-ray structure of the *p*-bromobenzenesulphonate derivative.²⁶ The extensive use of *E. autumnalis* by traditional healers for fever and the treatment of skin conditions, in particular wounds, boils and open sores, has led to extensive investigations into the anti-inflammatory and antibacterial effects of extracts of the leaves, bulbs and roots. The inhibition of prostaglandin synthesis was assessed in terms of the inhibition of the cyclooxygenase enzymes COX-1 and COX-2. COX-1 is found in normal cells and serves to produce substances that protect the stomach and kidneys (prostanoids). While both enzymes are involved in the inflammatory process, inhibition of COX-1 is associated with gastro-intestinal and kidney related side effects. As such, a higher selectivity for COX-2 inhibition is preferred.²⁷ Initial testing of the leaves, bulbs and roots of *E. autumnalis* against COX-1 showed good activity from all extracts (leaf extract IC₅₀ = 15 μg ml⁻¹, root extract 27 μg ml⁻¹ and bulb extract 72 μg ml⁻¹).^{28,29} A comparison of the COX-1 and COX-2



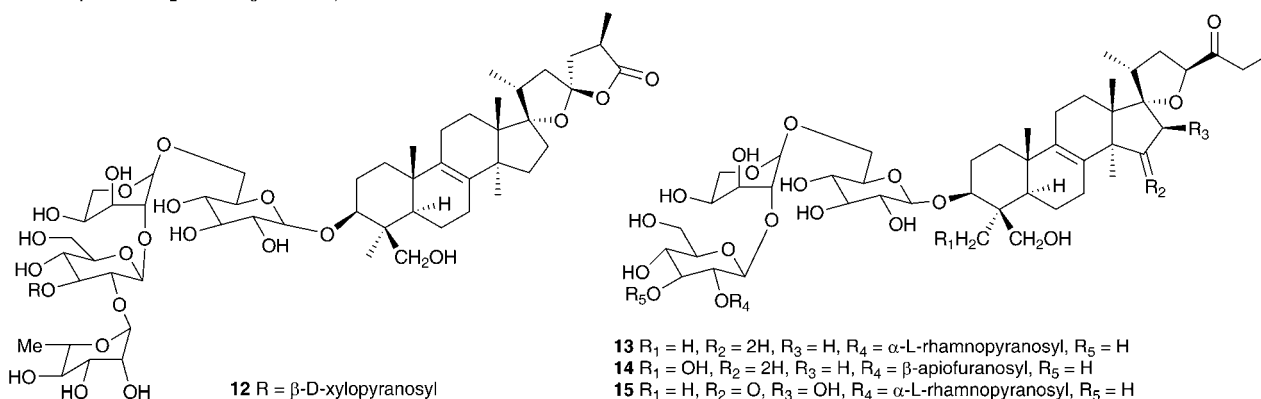
- 6** R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe
8 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OMe
40 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OMe
41 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH
47 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OH
49 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH
53 R₁ = OMe, R₂ = OH, R₃ = OH, R₄ = OH



- 7** R₁ = OH, R₂ = OMe
9 R₁ = OMe, R₂ = OMe
57 R₁ = OH, R₂ = OH



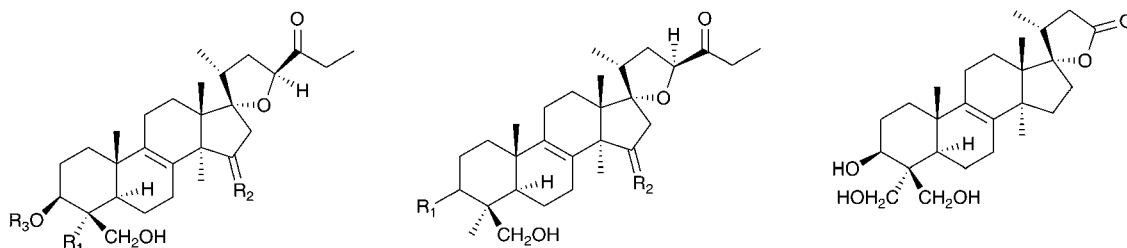
- 10** R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe
11 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OMe
19 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe
30 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH
42 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OH
48 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH



12 R = β -D-xylopyranosyl

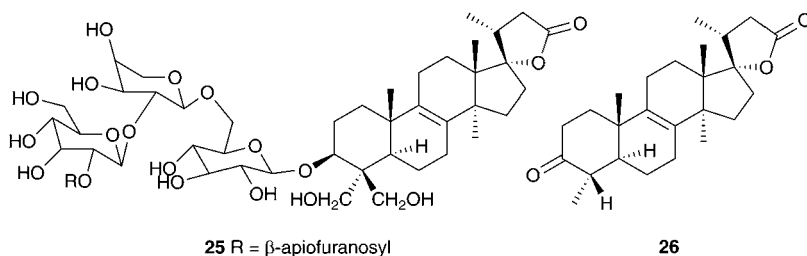
- 13** R₁ = H, R₂ = 2H, R₃ = H, R₄ = α -L-rhamnopyranosyl, R₅ = H
14 R₁ = OH, R₂ = 2H, R₃ = H, R₄ = β -apiofuranosyl, R₅ = H
15 R₁ = H, R₂ = O, R₃ = OH, R₄ = α -L-rhamnopyranosyl, R₅ = H
16 R₁ = OH, R₂ = 2H, R₃ = H, R₄ = α -L-rhamnopyranosyl, R₅ = α -D-xylopyranosyl
17 R₁ = H, R₂ = O, R₃ = H, R₄ = α -L-rhamnopyranosyl, R₅ = α -D-xylopyranosyl
18 R₁ = H, R₂ = O, R₃ = OH, R₄ = α -L-rhamnopyranosyl, R₅ = α -D-xylopyranosyl





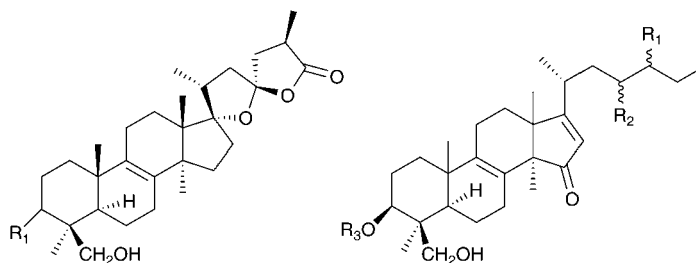
- 20 R₁ = CH₃, R₂ = O, R₃ = H
 21 R₁ = CH₂OH, R₂ = 2H, R₃ = H
 22 R₁ = CH₃, R₂ = 2H, R₃ = H
 31 R₁ = CH₃, R₂ = O, R₃ = β-D-glucopyranoside
 54 R₁ = CH₂OH, R₂ = O, R₃ = H

- 23 R₁ = =O, R₂ = 2H
 32 R₁ = β-O-β-D-glucopyranoside, R₂ = 2H
 35 R₁ = β-O-β-D-arabinopyranosyl-β-D-glucopyranoside, R₂ = O
 36 R₁ = β-O-β-D-arabinopyranosyl-β-D-glucopyranoside, R₂ = 2H
 39 R₁ = β-D-arabinopyranosyl-(1"→6')-[β-D-xylopyranosyl-(1"→3')]-β-D-glucopyranoside, R₂ = O
 63 R₁ = β-OH, R₂ = β-OH



25 R = β-apiofuranosyl

26



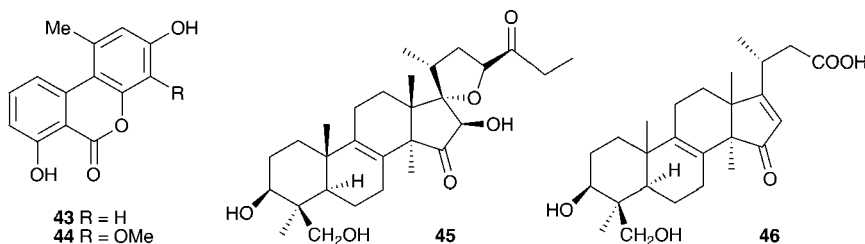
- 27 R₁ = =O
 33 R₁ = β-O-β-D-glucopyranoside
 38 R₁ = β-O-β-D-arabinopyranosyl-β-D-glucopyranoside
 28 R₁ = OH, R₂ = OH, R₃ = H
 29 R₁ = =O, R₂ = OH, R₃ = H
 34 R₁ = =O, R₂ = OH, R₃ = β-D-glucopyranoside
 37 R₁ = =O, R₂ = OH, R₃ = β-D-arabinopyranosyl-β-D-glucopyranoside

inhibitory activity of the leaf, root and bulb extracts, showed that the leaf extract inhibited COX-1 to a greater extent while the root and bulb extract showed greater inhibition of COX-2 (COX-2/COX-1 of 0.7 and 0.8 respectively for the root and bulb extracts).^{27,30} There has been some interest in the biological effects of plant lectins obtained from the bulbs of *E. autumnalis*. These non-enzymatic proteins can bind, with high specificity and reversibility, mono- and oligosaccharides. Cyclooxygenase is a membrane-bound glycoprotein and as such could be affected by plant lectins. The partially purified lectin-like protein extract of *E. autumnalis* was found to have very good cyclooxygenase inhibitory activity (COX-1 inhibition of 88%)³¹ as well as good activity against the Gram-positive bacterium *Bacillus subtilis* (MIC of 0.2 mg ml⁻¹).³² Due to the widespread use of the bulbs of *E. autumnalis* by South African traditional healers, extensive work has been carried out on the factors

possibly affecting the effectiveness of the bulb extracts. Environmental factors such as sunlight intensity, temperature and watering as well as age of the bulbs and the effect of cold storage have all been examined.³³ Bulbs lifted and stored at a low temperature (8–10 °C) showed greater COX-1 inhibitory activity.³⁴ The leaves of young plants showed greater COX-1 inhibitory activity than those of older plants, where the greatest activity was associated with bulbs and roots.³⁵

Six homoisoflavanones (19, 40, 47–50)^{14,36} and the optically active 2-hydroxy-2-[(4-hydroxyphenyl)methyl]butanedioic acid 51 have been isolated from the bulbs of *Eucomis comosa* (Houtt.) H. R. Wehrh. (as *Eucomis punctata* L'Herit).³⁷ The bulb wax of *Eucomis comosa* yielded the chroman-4-one 52³⁸ while an examination of the methanol extract of the fresh, air-dried bulbs gave five homoisoflavanones (47, 48, 50 and both the *E* and *Z* form of 6).³⁹ The same study included

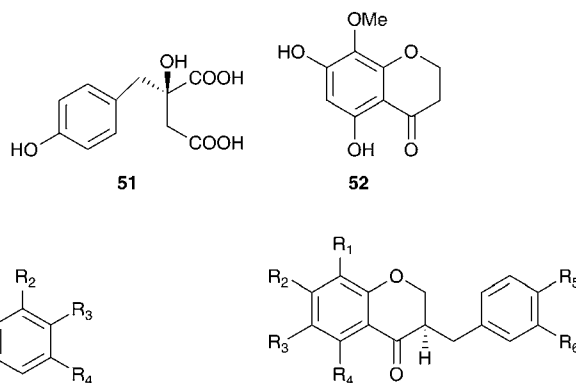




Eucomis pallidiflora Baker subsp. *pole-evansii* (N.E.Br.) Reyneke ex J.C.Manning and *Eucomis schijffii* Reyneke. The bulbs of *Eucomis pallidiflora* subsp. *pole-evansii* contained the homoisoflavanone 53 and the spirocyclic nortriterpenoid 20 while *Eucomis schijffii* yielded two compounds, the spirocyclic nortriterpenoid 54 and scillascillin 55.³⁹ *Eucomis montana* Compton was investigated and the bulbs were found to contain eleven homoisoflavanones (6, 7, 9, 10, 30, 50, 56, 57, 58, 59 and 60).⁴⁰ No inhibitory activity against cyclooxygenase enzymes (COX-1 and COX-2) was found for compounds 48, 53 and 55 isolated from *E. comosa*, *E. pallidiflora* and *E. schijffii* respectively.³⁹ Some antibacterial activity has been observed for homoisoflavanones isolated from *E. comosa* (MIC of 0.52 mM for compound 6 (*E* isomer) and 0.24 mM for compound 47) and *E. schijffii* (MIC of 0.50 mM for compound 55).⁴¹ The dichloromethane and methanol extracts of the whole plant of *Eucomis vandermerwei* I.Verd. yielded four known homoisoflavanones (6, 7, 10 and 56).⁴² Similarly, the whole plant of *Eucomis zambesiaca* Baker was investigated and this resulted

in the isolation and identification of five homoisoflavanones (6, 19, 40, 61 and 62) and three nortriterpenoids (20, 21 and 63).⁴²

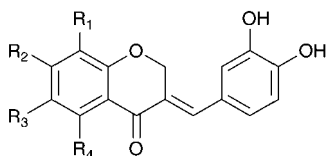
The bulbs of *Muscari racemosum* Mill. were found to contain homoisoflavanones of the 3-benzylidene-4-chromanone-type (64–66).⁴³ Long-range coupling between C-2 and C-9, was indicative of *Z* geometry. These homoisoflavanones have been investigated with respect to their effect on lipid peroxidation *in vitro*.^{44,45} Strong anti-oxidant activity was noted ($IC_{50} = 0.94\text{--}7.98 \mu\text{M}$) and this led to an investigation into their antimutagenic/anticarcinogenic activity. It has been suggested that polyphenolics may be cancer preventative due to their antioxidant properties⁴⁴ and a mixture of these three homoisoflavanones was found to contain significant antimutagenic activity.⁴⁴ Estrogenic activity was found in an ether extract of the bulbs. The extract induced proliferation of MCF7 cells in a dose dependent manner at concentrations up to $5 \mu\text{g ml}^{-1}$. This concentration produced the greatest effect of 181% of the control.⁴⁶



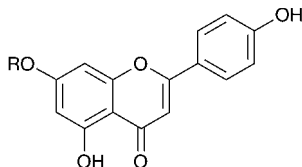
55 R₁ = OH, R₂ = H, R₃-R₄ = O-CH₂-O
60 R₁ = OMe, R₂ = H, R₃ = OMe, R₄ = OH
77 R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OH
79 R₁ = OH, R₂ = H, R₃ = OH, R₄ = OMe
98 R₁ = OH, R₂ = H, R₃ = OMe, R₄ = OH
104 R₁ = OH, R₂ = H, R₃ = OH, R₄ = OMe
105 R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OH
106 R₁ = OMe, R₂ = H, R₃ = OMe, R₄ = OH
107 R₁ = OMe, R₂ = OH, R₃ = OMe, R₄ = H

50 R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe, R₅ = OH, R₆ = H
56 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = OH, R₆ = H
58 R₁ = OH, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OH, R₆ = H
59 R₁ = OMe, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OH, R₆ = H
61 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH, R₆ = H
62 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OH, R₆ = H
70 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OH, R₆ = OH
71 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OH, R₆ = OMe
72 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OMe, R₆ = OH
73 R₁ = OH, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OH
74 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OH
75 R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe, R₅ = OH, R₆ = OH
76 R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe, R₅ = OMe, R₆ = OH
78 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OH
80 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe, R₆ = OH
97 R₁ = OH, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = OH, R₆ = H,





- 64** R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH,
65 R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe,
66 R₁ = OH, R₂ = OMe, R₃ = H, R₄ = OH,

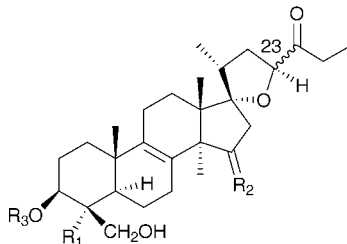


- 67** R = β-D-arabinopyranoside
68 R = β-D-xylopyranoside
69 R = β-D-glucopyranoside

A wide range of compounds has been isolated from the bulbs of *Muscari armeniacum* Leichtlin ex Baker. These include flavonoids (**67–69**),⁴⁷ homoisoflavanones, with an unusual 3',4' substitution pattern in the B ring (**30, 62, 70–80**),^{48,49} spirocyclic nortriterpenoid glycosides (**81–87**)⁵⁰ and polyhydroxylated pyrrolizidine alkaloids (**88–92**).⁵¹ The alkaloids were assessed for their action against glycosidases. Compound **88** was found to be a potent inhibitor of rat intestine lactase (IC₅₀ = 4.4 μM) as well as a moderate inhibitor of α-L-fucosidase (IC₅₀ = 46 μM) and amyloglucosidase (IC₅₀ = 25 μM). Inverting the hydroxyl group at C-1 to give compound **89**, enhanced the

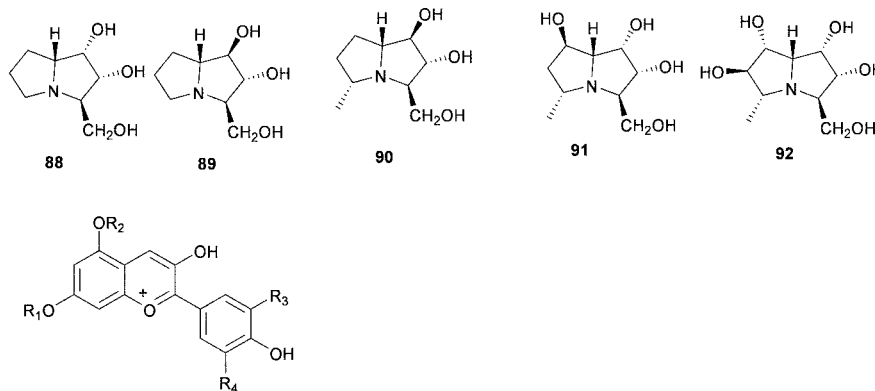
amyloglucosidase inhibition (IC₅₀ = 8.6 μM) but resulted in the loss of the α-L-fucosidase inhibition. Compounds **90** and **91** were less active.⁵¹ Three ribosome-inactivating proteins have been isolated from the bulbs of *M. armeniacum*. It has been proposed that this type of activity is linked to an anti-viral response.⁵² The petals of *M. armeniacum* have yielded four anthocyanins, the new muscarinin A **93** and the known 3-O-β-D-glucopyranoside of delphinidin, petunidin and malvidin (**94–96**).^{53–55} The principle water soluble polysaccharide in the bulbs of *M. armeniacum* (as *Muscari szovitsianum* Baker) was found to be a neutral glucofructan, with glucose and fructose residues in the ratio 26 : 1.^{56,57}

Phytochemical investigations into the bulbs of *Muscari botryoides* (L.) Mill. confirmed a close taxonomic relationship with *Muscari armeniacum*. Similar homoisoflavanones (**19, 30, 58, 60, 77, 79, 97, 98**)⁴⁸ were isolated as well as similar spirocyclic nortriterpenoid glycosides (**86, 87, 99, 100, 101, 102, 103**).⁵⁰ *Muscari neglectum* Guss. ex Ten. was found to contain a similar spread of homoisoflavanones in the bulbs (**30, 62, 80, 104–107**) as well as an interesting scillascillin-type homoisoflavanone with an unusual oxygenation pattern in the B ring (**55**).⁵⁸ Ethanol extracts of the bulbs and leaves of *Muscari neglectum* have been tested for antifungal activity against the wood rots *Postia placenta* and *Trametes versicolor*. Significant reduction in wood weight loss was observed for Turkish oriental beech and Scots pine (weight loss dropped from 35–42% in the untreated wood to 5.4–18.3% in the treated wood).⁵⁹



- 81** (23*R*) R₁ = CH₃ R₂ = O, R₃ = β-D-glucopyranoside
82 (23*R*) R₁ = CH₃ R₂ = O, R₃ = α-L-arabinopyranosyl-(1→3)-α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
83 (23*R*) R₁ = CH₃ R₂ = O, R₃ = β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→3)-α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
84 (23*S*) R₁ = CH₂OH, R₂ = 2H, R₃ = β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
85 (23*S*) R₁ = CH₂OH, R₂ = 2H, R₃ = α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
86 (23*R*) R₁ = CH₃, R₂ = O, R₃ = α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
87 (23*R*) R₁ = CH₃, R₂ = O, R₃ = α-L-arabinofuranosyl-(1→2)-α-L-arabinopyranosyl-(1→3)[rhamnopyranosyl-(1→2)]-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
99 (23*R*) R₁ = CH₃ R₂ = O, R₃ = β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
100 (23*S*) R₁ = CH₂OH, R₂ = 2H, R₃ = α-L-arabinopyranosyl-(1→2)-β-D-galactopyranosyl-(1→3)-[β-D-apiofuranosyl-(1→2)-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
101 (23*S*) R₁ = CH₂OH, R₂ = 2H, R₃ = β-D-glucopyranoside
102 (23*R*) R₁ = CH₂OH, R₂ = 2H, R₃ = α-L-arabinopyranosyl-(1→3)[β-D-apio-D-furanosyl-(1→2)]-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside
103 (23*R*) R₁ = CH₂OH, R₂ = 2H, R₃ = β-D-apio-D-furanosyl-(1→2)]-β-D-glucopyranosyl-(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside

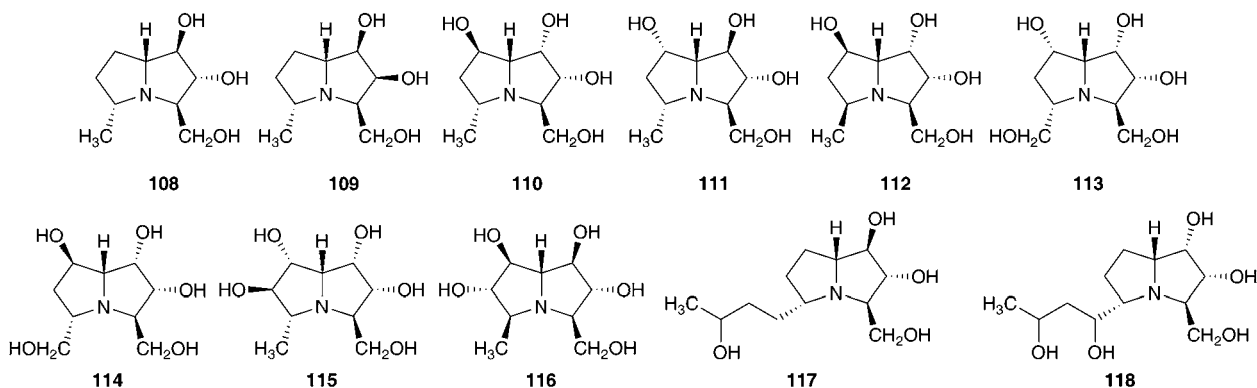


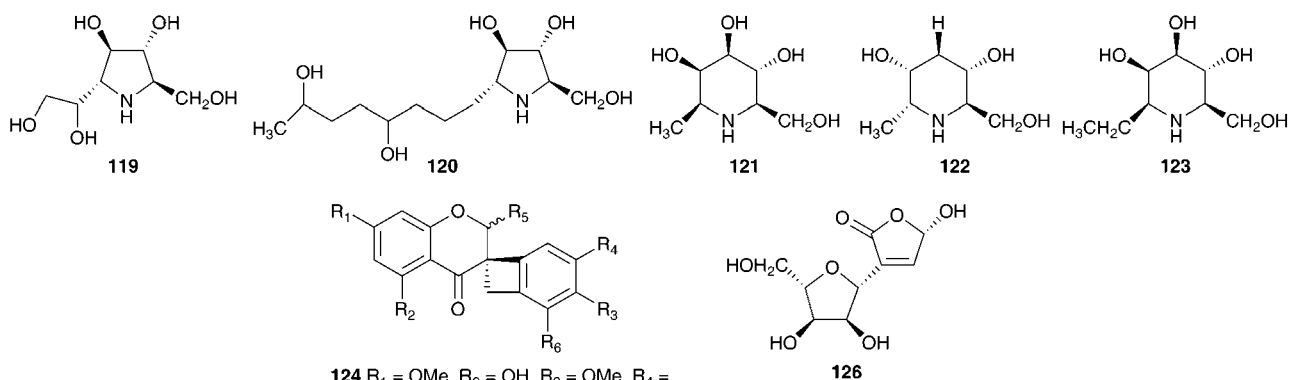


An aqueous ethanol extract of the bulbs of *Ledebouria socialis* (Baker) Jessop (as *Scilla socialis* Baker) was found to contain eleven hyacinthacines (108–118) as well as two pyrrolidines (119–120) and three piperidines (121–123).⁶⁰ Hyacinthacines appear to be restricted to the Hyacinthoideae. These compounds have been found in *Othocallis siberica* (Haw.) Speta (as *Scilla sibirica*), *Oncostema peruviana* (L.) Speta (as *Scilla peruviana*), *Hyacinthoides non-scripta* (L.) Chouard ex Rothm., *Hyacinthoides hispanica* (Mill.) Rothm. (as *Scilla campanulata*), and *Muscari armeniacum*. The hyacinthacines are characterised as 7α-*R*-hydro-1,2-dihydroxy-3-hydroxymethylpyrrolizidines and in general appear to be weak to moderate inhibitors of glycosidases.⁶⁰ The dichloromethane and methanol extracts of the bulbs of *Ledebouria socialis* have yielded five compounds, the scillascillin-type homoisoflavanones 124 and 125, phytol, stigmasterol and the polyhydroxylated difuran derivative, polybotrin 126.⁶¹ The different types of compounds isolated reflect the different solvents used for extraction, with the more polar extracts yielding the more polar polyhydroxy alkaloids and the less polar extracts the homoisoflavanones and sterols.

Early work by Gunn *et al.*⁶² on some southern African *Ledebouria* species, examined the cardiac effect of *Ledebouria cooperi* (Hook.f.) Jessop (as *Scilla cooperi* Hook and as *Scilla rogersii* Baker) and *Ledebouria ovatifolia* (Baker) Jessop (as *Scilla lanceafolia* Baker). Both species produced a similar action to digitalis when applied to isolated frog and cat hearts as well as *in situ* frog, cat and dog hearts. The extracts obtained were, however, insufficient in quantity to determine minimum lethal doses and the active components were not identified.

The chemical constituents of a further six southern African *Ledebouria* species have been investigated. Two samples of the bulbs of *Ledebouria zebrina* (Baker) S.Venter (as *Scilla zebrina* Baker), collected from two different regions in South Africa (Durban and Blyde River, Mpumalanga) have been investigated and the chemical differences noted. Both contained homoisoflavanones (Durban bulbs compounds 127 and 128 and the Blyde River bulbs compounds 127, 129, 130). In addition to the homisoflavanones, the Blyde River bulbs also contained spirocyclic nortriterpenoids (131, 21), while the Durban bulbs produced a chalcone 132.⁶³ The bulbs of *Ledebouria cooperi*





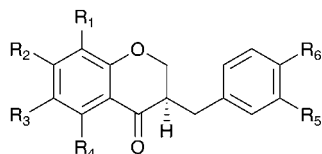
- 124** R₁ = OMe, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OMe
125 R₁ = OMe, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OH
139 R₁ = OMe, R₂ = OAc, R₃-R₄ = O-CH₂-O, R₅ = β-OAc, R₆ = H
140 R₁ = OMe, R₂ = OAc, R₃-R₄ = O-CH₂-O, R₅ = α-OAc, R₆ = H
142 R₁ = OMe, R₂ = OH, R₃-R₄ = O-CH₂-O, R₅ = H, R₆ = H
143 R₁ = OH, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = H
144 R₁ = OH, R₂ = OH, R₃ = OH, R₄ = OMe, R₅ = H, R₆ = H
160 R₁ = OH, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = H
161 R₁ = OMe, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = H

yielded the homoisoflavanones **59** and **62** as well as the eucosterol-type spirocyclic nortriterpenoid **133** and malic acid. *Ledebouria ovatifolia* bulbs were found to contain the chalcones **132** and **134** as well as the homoisoflavanone **30**.^{64,65} *L. ovatifolia* is used by the Zulu to treat gastro-enteritis, influenza and backache. In addition, it has been found to be toxic to sheep.⁶⁶ Extracts of the bulbs were found to have good antibacterial activity against Gram-positive bacteria (MIC of 0.8–12.5 mg ml⁻¹), but poor anti-inflammatory and poor antihelmintic activity.^{66,67} A later investigation of the dichloromethane, ethyl acetate and methanol extracts of the bulbs led to the isolation of twenty-nine compounds, including xanthenes (**135–138**), homoisoflavanones (**7**, **10**, **30**, **50**, **74**, **75**, **139–148**), triterpenoids (**133**, **149–150**), chalcones (**132**, **151**) and simple aromatic compounds (**152–154**), including polybotrin **126**.^{61,68} Sixteen of the above compounds isolated from *L. ovatifolia* were assessed for anti-inflammatory activity, in particular the selective inhibition of the COX-2 enzyme. Compounds **75**, **137** and **146** were found to significantly inhibit COX-1 and COX-2 (percentage activity relative to the non-inhibitor control of 54–56% for COX-1 and 0% for COX-2), while compound **145** showed good selectivity for the inhibition of COX-2 over COX-1 (IC₅₀ for COX-2 of 2.87 μM, with no statistically significant inhibition for COX-1 up to a concentration of 20 μM).^{61,68} During the same study, the norlignan hinokiresinol **155**, previously isolated from *Drimiopsis burkei* Baker,⁶⁹ was synthesised and tested for selective COX-2 inhibitory activity. Significant selective inhibition against COX-2 was observed (percentage activity relative to the non-inhibitor control of 93.8% for COX-1 and 0% for

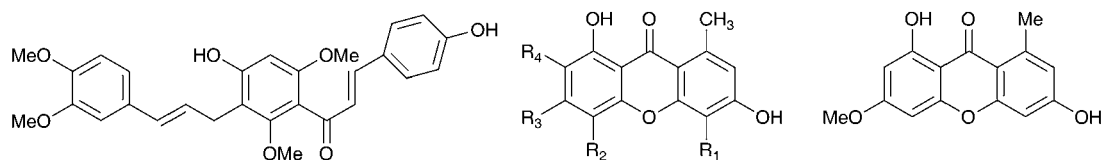
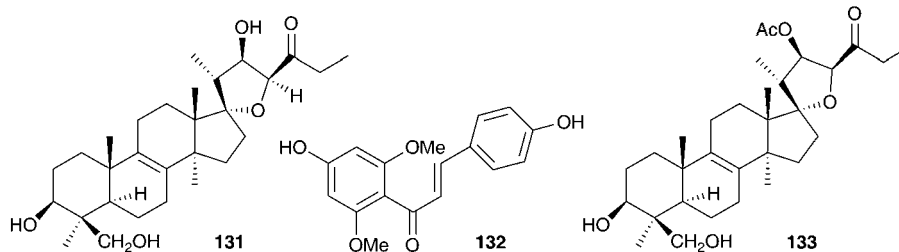
COX-2).^{61,68} *Ledebouria leptophylla* (Baker) S.Venter (as *Ledebouria graminifolia* (Baker) Jessop), sourced in Botswana, is used traditionally to treat gastro-enteritis, backache and coughs as well as a treatment for skin irritations and for dressing wounds. A phytochemical investigation of the bulbs found seven 3-benzyl-4-chromanones (**30**, **62**, **128**, **156–159**), two 3-benzyl-3-hydroxy-4-chromanones (**7**, **57**), two scillascillin-type homisoflavanones (**160–161**) and two xanthenes (**136** and **162**). Cultivated and wild-sourced bulbs contained the same compounds.^{70,71} There was initial concern about the presence of the xanthenes and speculation as to whether they may have come from a fungal or lichen contamination of the wild bulbs. The fact that the cultured bulbs contained the same xanthenes confirmed that they were secondary metabolites of *L. leptophylla*.⁷²

Within South Africa *Ledebouria revoluta* (L.f.) Jessop is extremely widespread. It is used by the Sotho to drive away lightening and with the exception of pregnant women, to treat lumbago. The Kwena and Tswana use it to treat wounds and skin conditions. The bulbs were found to contain homoisoflavanones (**30**, **59**, **61** and **157**).⁷³ *Ledebouria floribunda* (Baker) Jessop is used by traditional healers in the Eastern Cape of South Africa to treat skin disorders, tuberculosis, urinary tract infections and gastro-enteritis. Three homoisoflavanones, with unusual substituents at C-6, -7 and -8 (**163–165**) have been isolated from the bulbs as well as two 7-*O*-diglycosides (**166–167**) and five additional homoisoflavanones (**10**, **30**, **50**, **62**, **168**). These latter seven homoisoflavanones have shown good antioxidant activity against the DPPH radical (IC₅₀ of 31–273 μg ml⁻¹) and in the β-carotene/





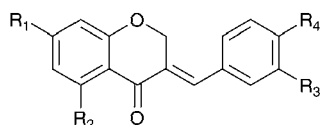
- 127 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OMe, R₅ = OMe, R₆ = OH
 128 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OMe, R₅ = H, R₆ = OH
 129 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OH, R₅ = OMe, R₆ = OH
 130 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe, R₆ = OH
 141 R₁ = OMe, R₂ = OH, R₃ = H, R₄ = OMe, R₅ = H, R₆ = OH
 148 R₁ = H, R₂ = OH, R₃ = H, R₄ = OMe, R₅ = OMe, R₆ = OAc
 156 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = H, R₆ = OH
 157 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = H, R₆ = OMe
 158 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OH
 159 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OH
 166 R₁ = H, R₂ = O-[α-L-rhamnopyranosyl-(1→6)-β-D-glucopyranoside], R₃ = H, R₄ = OH, R₅ = H, R₆ = OMe
 167 R₁ = H, R₂ = O-[α-L-rhamnopyranosyl-(1→6)-β-D-glucopyranoside], R₃ = H, R₄ = OH, R₅ = H, R₆ = OH
 168 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OH



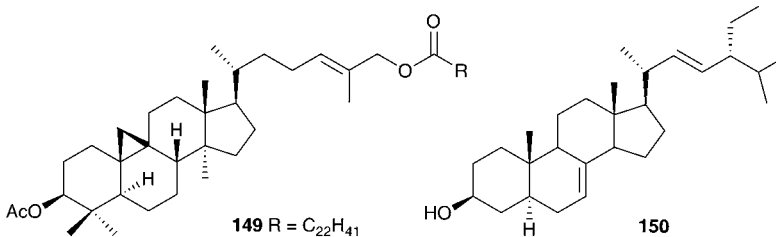
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- 135 R₁ = OMe, R₂ = H, R₃ = OMe, R₄ = OMe
 137 R₁ = H, R₂ = H, R₃ = OH, R₄ = OMe
 138 R₁ = H, R₂ = H, R₃ = OH, R₄ = H
 162 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OMe

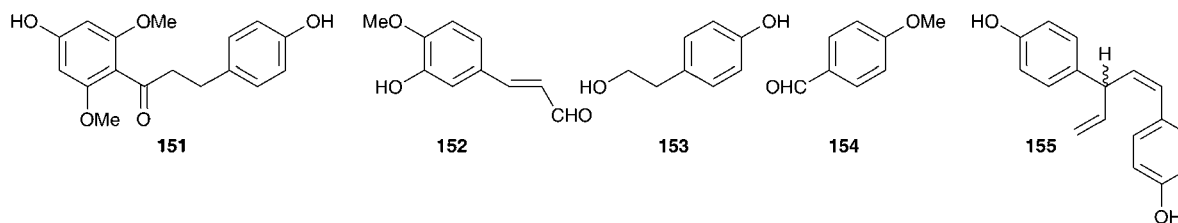
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- 145 R₁ = OH, R₂ = OMe, R₃ = OH, R₄ = OH
 146 R₁ = OH, R₂ = OH, R₃ = OH, R₄ = OH
 147 R₁ = OAc, R₂ = OMe, R₃ = H, R₄ = OAc

149 R = C₂₂H₄₁

150



151

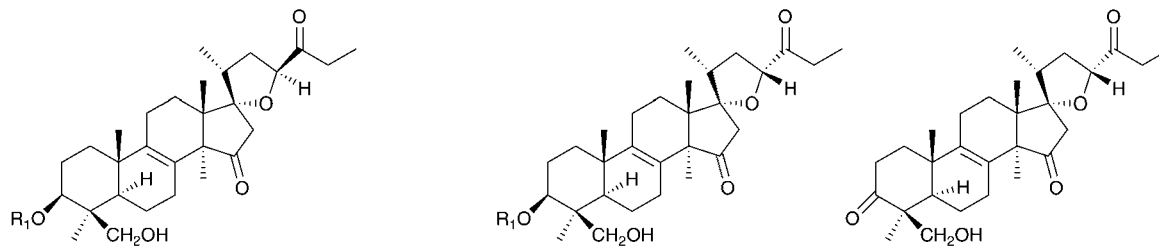
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176 R₁ = β-D-apio-D-furanosyl-(1→2)]-β-D-glucopyranosyl(1→2)-α-L-arabinopyranosyl-(1→6)-β-D-glucopyranoside

177 R₁ = α-L-arabinofuranosyl(1→2)-α-L-arabinopyranosyl(1→3)[α-L-rhamnopyranosyl(1→2)]-β-D-glucopyranosyl(1→2)-α-L-arabinopyranosyl(1→6)-β-D-glucopyranoside
178 R₁ = H

179

homoisoflavanones was determined using circular dichroism and found to be *R* in all cases.⁹⁹ For the scillascillin type, this was further confirmed by X-ray analysis.⁹⁸ The crude extract of the bulbs has been subjected to anti-inflammatory bioassay-directed fractionation using a croton oil-induced mouse ear dermatitis test. The homoisoflavanone rich fraction was found to have high activity and further fractionation resulted in the isolation of **30**, **59**, **61**, **80** and **97**, all of which showed activity, with compound **30** being the most active at 41% inhibition (100 μg ear⁻¹).¹⁰⁰ Two flavonoids have been isolated from the flowers of *L. comosa*, 3-glucoarabinosyl- and 3-rhamnoarabinosyl-3,4,5-trihydroxy-7-alkoxyl flavone.¹⁰¹

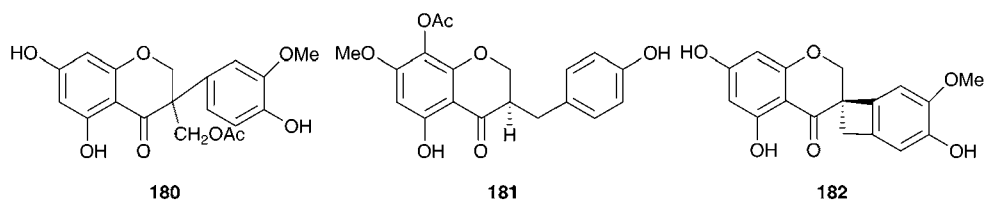
Leopoldia comosa is used in the Basilicata region of southern Italy to treat toothache and headache.¹⁰² One third of the plants used for medicinal purposes in southern Italy are used to treat skin and soft tissue infections. As *Staphylococcus aureus* is a common cause of such infections, the effect of a number of Italian plants, including the bulbs of *Leopoldia comosa*, on the inhibition of the *S. aureus* biofilms has been studied. Significant dose-dependent biofilm inhibition (IC₅₀ = 16 μg ml⁻¹) was noted for *L. comosa*.¹⁰³

Investigations into the chemistry of the bulbs of *Scilla luciliae* (Boiss.) Speta (as *Chionodoxa gigantea* Whittall and as *Chionodoxa luciliae* Boiss.) has yielded a large number of lanosterol-type spirocyclic nortriterpenoids, many of which have been tested for biological activity. Compounds **183–186**^{16,104} and compounds **13** and **187–188**¹⁷ have been isolated from the bulbs of *Scilla luciliae* (as *Chionodoxa gigantea*). In separate investigations, compounds **13**, **184** and **187–190**¹⁷ as well as compounds **189** and **190–197** and eucosterol **20**,¹⁰⁵ compounds **198–204**,¹⁰⁶ **205–207**¹⁰⁷ and **208–209**¹⁰⁸ have been found in the bulbs of this species (as *C. luciliae*). Compounds **183–186** all

showed some inhibitory activity on cyclic AMP phosphodiesterase, with compound **183** giving the best results (IC₅₀ = 11.2 × 10⁻⁵ M).¹⁰⁴ Cytotoxicity against HeLa cells was assessed for **13**, **184**, **187**, **188**, **189**, and **190** with the 15-deoxoeucosterol oligosaccharides (**13**, **187** and **190**) showing good activity down to 5 μg ml⁻¹.¹⁷ Compounds **198–204** as well as **183** and **185** were tested for activity against HSC-2 human oral squamous carcinoma cells. Compounds **183**, **185**, **200**, **201** and **204** all gave good results with LD₅₀ values in the range 10–23 μg ml⁻¹ (etoposide LD₅₀ = 24 μg ml⁻¹, for etoposide resistant HSC cells). It was noted that the 23*R* isomer of **185**, compound **186** did not show any activity.¹⁰⁶ Compounds **208** and **209** showed only weak activity in the same assay (LD₅₀ of 254 and 238 μg ml⁻¹ respectively).¹⁰⁸ In addition to the spirocyclic nortriterpenoids, the bulbs of *Scilla luciliae* (as *Chionodoxa luciliae*) have been found to contain the homoisoflavanones **210–222**.¹⁰⁹

The bulbs and aerial parts of *Scilla bifolia* L. have been studied using HPLC and mass spectrometry. Eighteen polyphenols were identified from extracts of the bulbs and aerial parts (caftaric acid, isoquercetin, rutin, myricetin, fistein, quercetin, patuletin, gentisic acid, caffeic acid, chlorogenic acid, *p*-coumaric acid, ferulic acid, hyperoside, quercitrin, luteolin, kaempferol, apigenin and sinapic acid).¹¹⁰

The methanol and aqueous ethanol extracts of the bulbs of *Autonoe madeirensis* (Menezes) Speta (as *Scilla maderensis* Menezes), a species endemic to the Portuguese archipelago of Madeira, was found to produce cardiac action on a frog heart and although TLC showed compounds with R_f values and colours very similar to the bufadienolides proscillaridin A **223** and scillaren A **224**, typical of the Urgineoideae, the presence of these compounds was not conclusively established.¹¹¹

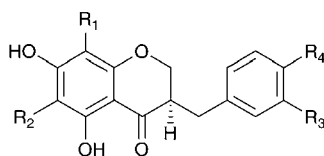
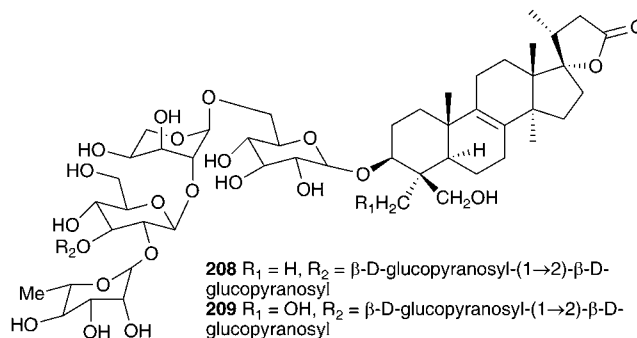


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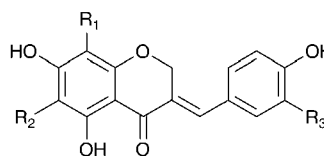
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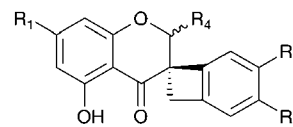




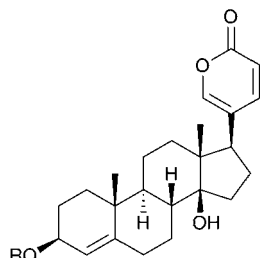
- 210 R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OMe
 211 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OMe
 212 R₁ = OMe, R₂ = H, R₃ = H, R₄ = OH
 213 R₁ = H, R₂ = H, R₃ = OH, R₄ = OMe
 214 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH
 215 R₁ = H, R₂ = H, R₃ = OH, R₄ = OH



- 216 R₁ = OMe, R₂ = H, R₃ = OH
 217 R₁ = H, R₂ = OMe, R₃ = OH
 218 R₁ = OMe, R₂ = H, R₃ = H



- 219 R₁ = OH, R₂-R₃ = O-CH₂-O, R₄ = OH
 220 R₁ = OMe, R₂-R₃ = O-CH₂-O, R₄ = OH
 221 R₁ = OH, R₂-R₃ = O-CH₂-O, R₄ = OH
 222 R₁ = OH, R₂ = OH, R₃ = OMe, R₄ = OH



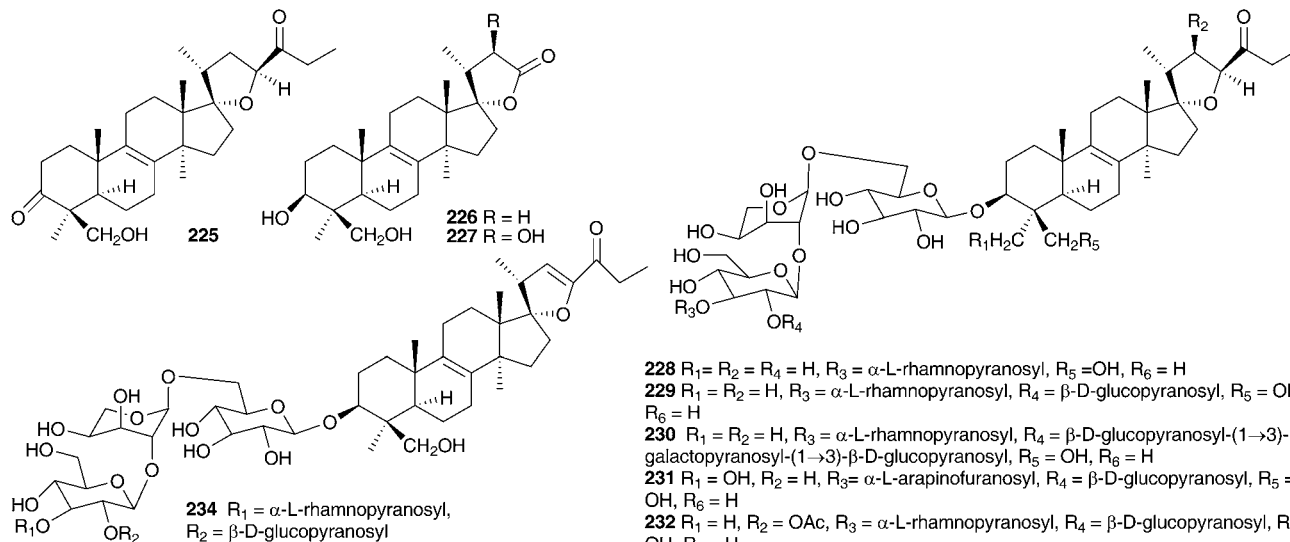
- 223 : R₁ = α-L-rhamnopyranoside
 224 R₁ = α-L-rhamnopyranosyl-(1→4)-β-D-glucopyranoside

Barnardia japonica (Thunb.) Schult. & Schult.f. (as *Scilla scilloides* (Lind.) Druce) has been used for a long time by traditional Chinese healers to treat abscesses and to promote circulation.¹¹² This plant also grows extensively in the wild in Japan and Korea and has been used by traditional healers of the latter country.¹¹³ The root extract has been evaluated for its potential as an antimicrobial agent, as an anti-inflammatory and as an antioxidant.¹¹³ The root extract of *B. japonica* (as *Scilla scilloides*) was found to inhibit the growth of *Staphylococcus aureus*, *Salmonella enteritidis*, *Escherichia coli* and *Candida parapsilosis* down to a concentration of 0.1% of the extract. Antioxidant activity was evaluated by looking at the inhibition of hyaluronidase, an enzyme that initiates the degradation of hyaluronic acid, associated with inflammation. Inhibition was noted at concentrations of root extract of 0.1 (14.8% inhibition) and 1% (48.2% inhibition), but not below. Anti-oxidant activity was assessed by monitoring the oxidation of linoleic acid and good activity was found (anti-oxidative index of 33.2 at a concentration of 1%).¹¹³ Extensive phytochemical investigations of the bulbs of *B. japonica* (as *Scilla scilloides*) resulted in the isolation of spirocyclic nortriterpenes (21, 22, 131, 225, 226 and 227),^{114,115} the related

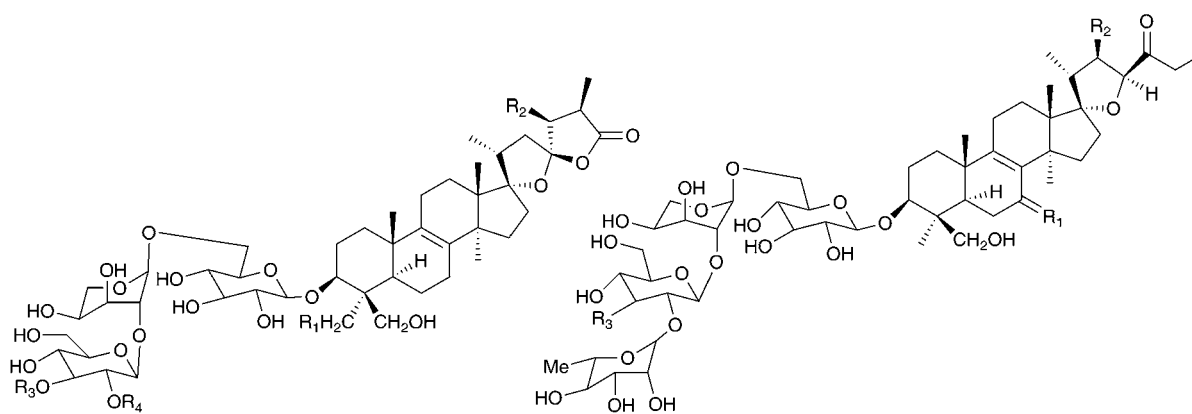
oligoglycosides (187, 228–239 and 241–247)^{112,116–118} and homoisoflavanones (42, 55 and 248).¹¹⁹ Nishida *et al.*¹²⁰ reported the isolation of sixteen compounds from the methanol extract of the bulbs, including three nortriterpenes (21, 22, and 225), a lignan 249, a xanthone 137, two homostilbenes (250 and 251) and nine homoisoflavanones (49, 55, C-3 epimer compound of 62, 182, 252, 253, and 254–156). The configuration at C-3 for 253, 254 and the epimer of compound 62 was unusually found to be *S* by circular dichroism.¹²⁰ Scillascilloside E-3 239 and scillascilloside E-1 237 have shown potent cytotoxicity against a range of human cancer cells (ED₅₀ 1.5–3.0 nM and 1.6–5.9 nM respectively).^{112,117} Patents have been filed covering the use of *B. japonica* (as *Scilla scilloides*) bulb extracts for the treatment of fungal infections¹²¹ and as a treatment for cancer.¹²² A later investigation into the methanol extract of the bulbs resulted in the isolation of two norlanostane-type triterpenoid glycosides (257 and 258), a phenylpropanoid glycoside 259 and two alkaloids (260 and 261).¹²³ The aqueous methanol extract of the bulbs of *B. japonica* (as *Scilla japonica*) produced some cardiac action on an isolated toad heart muscle, although the active components responsible were not identified.¹²⁴

Othocallis siberica (as *Scilla sibirica* Haw.) is native to south-western Russia, the Caucasus and Turkey. The bulbs were found to contain 0.04% glycosides and 0.15% alkaloids, with the alkaloid fraction being able to stop the function of an isolated frog heart. The glycoside fraction showed a strong hypertensive action and was a stimulant to respiration.¹²⁵ The hot water extract of the bulbs produced an acylated glucomannan with D-mannose, D-glucose and an acyl group in a ratio of 7.7 : 1 : 2.4.¹²⁶ As part of their investigations into potential glycosidase inhibitors, Yamashita *et al.* examined the ethanol extract of the bulbs of *O. siberica* (as *Scilla sibirica*). They isolated



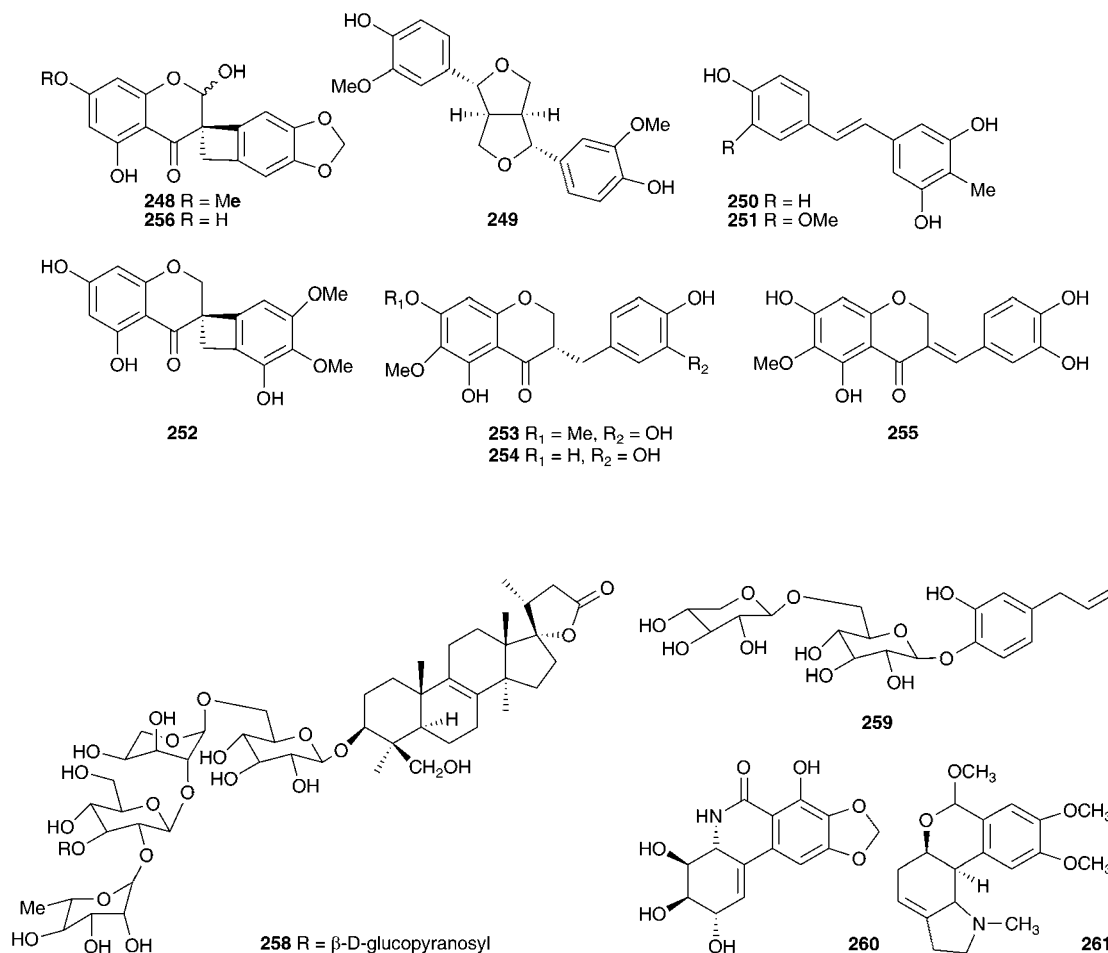


- 236** R₁ = H, R₂ = H, R₃ = α -L-rhamnopyranosyl, R₄ = β -D-glucopyranosyl
244 R₁ = H, R₂ = OH, R₃ = β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
245 R₁ = H, R₂ = OH, R₃ = α -D-xylopyranosyl- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
246 R₁ = OH, R₂ = H, R₃ = β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
247 R₁ = H, R₂ = OH, R₃ = H, R₄ = α -L-rhamnopyranosyl
241 R₁ = O, R₂ = OAc, R₃ = β -D-glucopyranosyl
242 R₁ = 2H, R₂ = H, R₃ = α -D-xylopyranosyl- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl
243 R₁ = 2H, R₂ = H, R₃ = β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl



- 236** R₁ = H, R₂ = H, R₃ = α -L-rhamnopyranosyl, R₄ = β -D-glucopyranosyl
244 R₁ = H, R₂ = OH, R₃ = β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
245 R₁ = H, R₂ = OH, R₃ = α -D-xylopyranosyl- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
246 R₁ = OH, R₂ = H, R₃ = β -D-glucopyranosyl, R₄ = α -L-rhamnopyranosyl
247 R₁ = H, R₂ = OH, R₃ = H, R₄ = α -L-rhamnopyranosyl
241 R₁ = O, R₂ = OAc, R₃ = β -D-glucopyranosyl
242 R₁ = 2H, R₂ = H, R₃ = α -D-xylopyranosyl- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl
243 R₁ = 2H, R₂ = H, R₃ = β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl

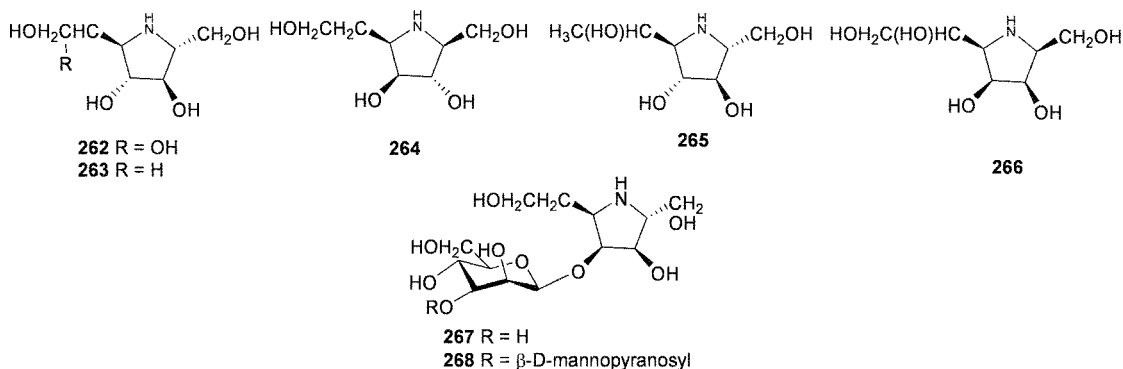


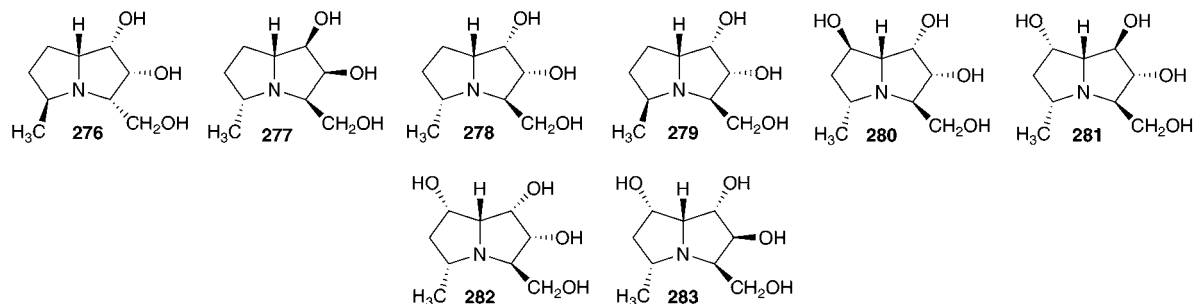
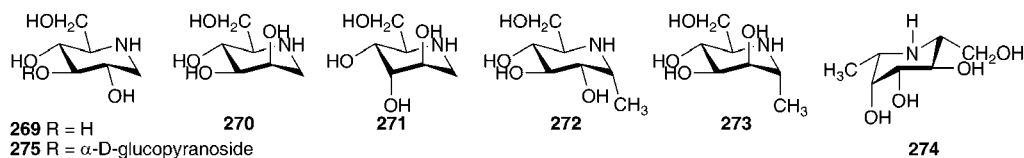


and identified five pyrrolidines (262–266), two pyrrolidine glycosides (267–268), six piperidines (269–274), one piperidine glycoside (275) and eight pyrrolizidines (276–283).¹²⁷ These alkaloids were tested for their inhibitory activity against various glycosidases. Compound 262 was found to be a potent inhibitor of bacterial (*Caldocellum sacchrolyticum*) β-glucosidase and mammalian β-galactosidase (IC₅₀ = 3.2 μg ml⁻¹ and 4.4 μg ml⁻¹ respectively). The loss of the hydroxy group at C-6, to give

compound 263, significantly lowered the activity. Moderate or no activity was found for all other compounds isolated.¹²⁷

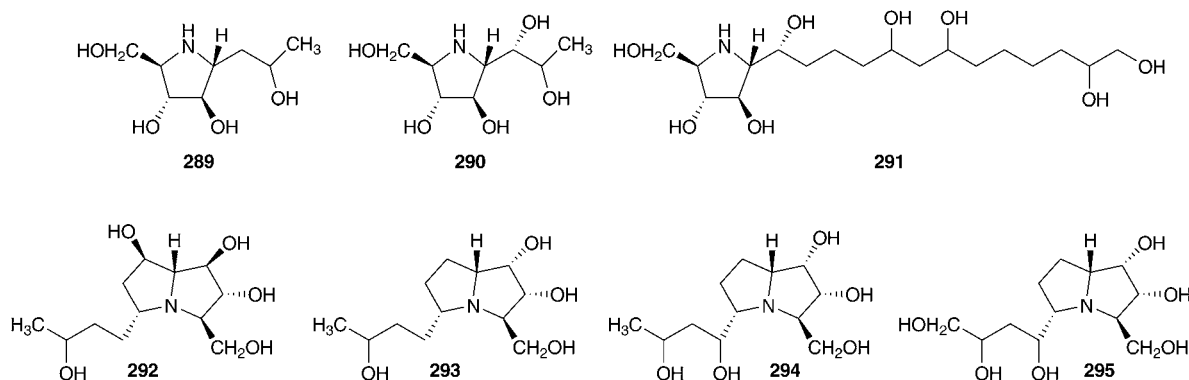
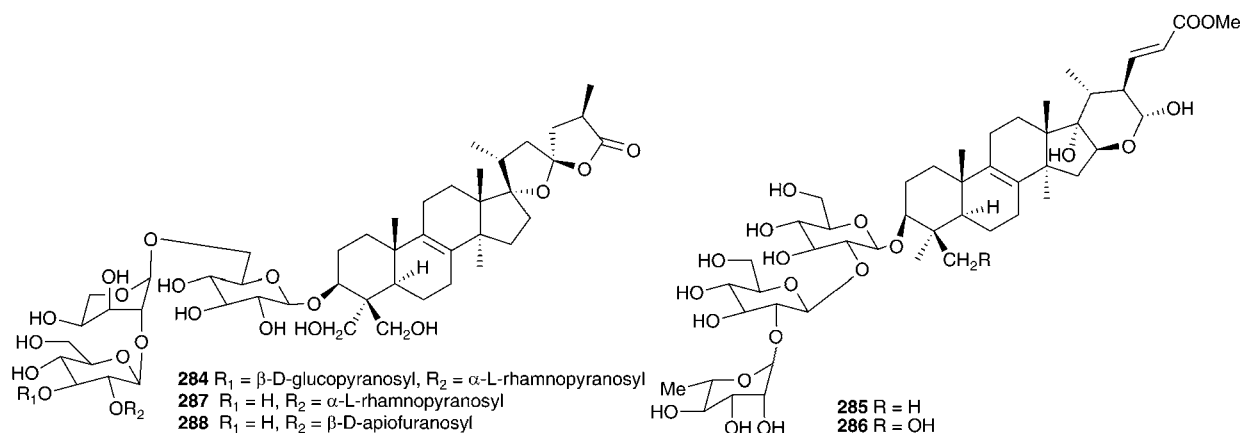
The bulbs of *Oncostema peruviana* (as *Scilla peruviana*) were found to contain rearranged lanosterol glycosides (scillasapinin B 284, peruvianoside A 285, peruvianoside B 286, and compounds 187–188 and 287–288)^{16,17,128,129} as well as a range of polyhydroxylated pyrrolidines (289–291) and pyrrolizidines (292–295).¹³⁰ Moderate inhibitory activity against cyclic AMP





phosphodiesterase was noted for peruvianoside A **285** ($IC_{50} = 23.5 \times 10^{-5}$ M) and scillasaponin B **286** ($IC_{50} = 14.0 \times 10^{-5}$ M),^{16,128} while compound **187**, a 15-deoxoeucoesterol oligosaccharide, was found to be toxic to HeLa cells at a concentration of $5 \mu\text{g ml}^{-1}$.¹⁷ The pyrrolidine **291** was found to be a potent

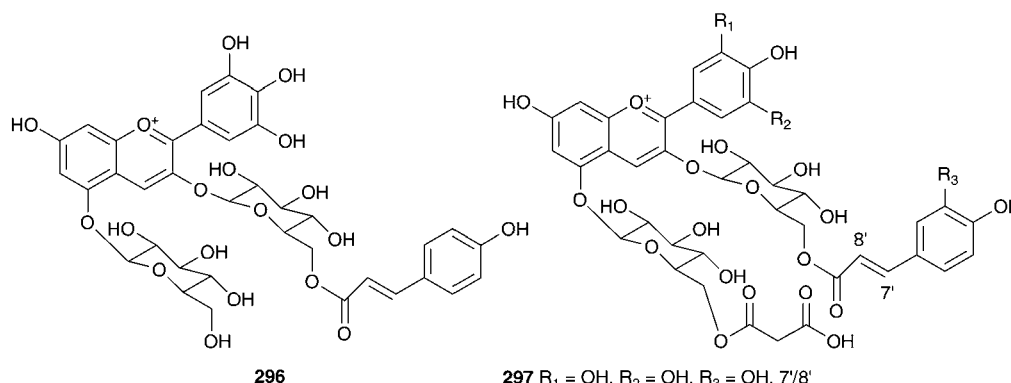
inhibitor of bacterial β -glucosidase ($IC_{50} = 80$ nM), compounds **292** and **293** showed significant inhibition of yeast α -glucosidase ($IC_{50} = 6.6$ and $6.3 \mu\text{M}$ respectively) with compound **293** also showing good inhibition of bacterial β -glucosidase ($IC_{50} = 5.1 \mu\text{M}$).¹³⁰



Phytochemical work on the Hyacinthaceae has primarily focused on the bulbs. The exception to this has been the genus *Hyacinthus*, where most of the work has been done on the flowers. *Hyacinthus orientalis* is used as an ornamental plant and can have blue, pink, yellow or white flowers. The blue flowers of *H. orientalis* have been found to contain a range of acylated anthocyanins (296–302).¹³¹ The anthocyanin content was also studied in the blue flowers produced *in vitro*. The same anthocyanins were produced with some small changes in the percentage composition of the different compounds.¹³² The red flowers of *H. orientalis* likewise produced a range of acylated anthocyanins (303–316) and, as with the blue flowers, the red flowers produced *in vitro* produced the same anthocyanins with some differences in composition.^{133–135} A later investigation of the blue flowers of *H. orientalis* revealed a further seven anthocyanins (317–323) in addition to those isolated previously.¹³⁶ Differences in essential oil content between wild *H. orientalis* flowers and those generated *in vitro* have been studied. Four common components were found, 1-hepten-3-ol, benzyl

alcohol, phenethyl alcohol and cinnamyl alcohol. The main components in the wild flowers were phenethyl alcohol (55% at stage 3 and 48% at stage 4) and cinnamyl alcohol (23% at stage 3 and 29% at stage 4) while the *in vitro* generated flowers produced 75% phenethyl alcohol as the major component.¹³⁷ The bulbs of *H. orientalis* were found to be rich in polyhydroxy alkaloids. α -Homonojirimycin 324 was the major component¹³⁸ with the compounds 325–334 being found in smaller amounts.^{138,139} Compound 332 was found to be a good inhibitor of bacterial β -glucosidase ($IC_{50} = 3.8 \mu M$), mammalian β -galactosidases ($IC_{50} = 4.0 \mu M$ and $4.4 \mu M$) and mammalian trehalases ($5.0 \mu M$ and $2.0 \mu M$), while compound 333 inhibited rice α -glucosidase ($IC_{50} = 2.2 \mu M$) and rat intestinal maltase ($IC_{50} = 2.5 \mu M$) and compound 334 inhibited α -L-fucosidase ($IC_{50} = 50 \mu M$).¹³⁹

The methanol extract of the bulbs of *Bellevalia paradoxa* (Fisch. & C.A.Mey.) Boiss. (as *Muscari paradoxum* (Fisch. et C.A. Mey.) K.Koch) yielded eight 27-norlanostane glycosides (335–342)⁵⁵ and seven tetranorlanostane glycosides (25, 343–348).⁵⁴ All were tested



296

297 R₁ = OH, R₂ = OH, R₃ = OH, 7'/8'

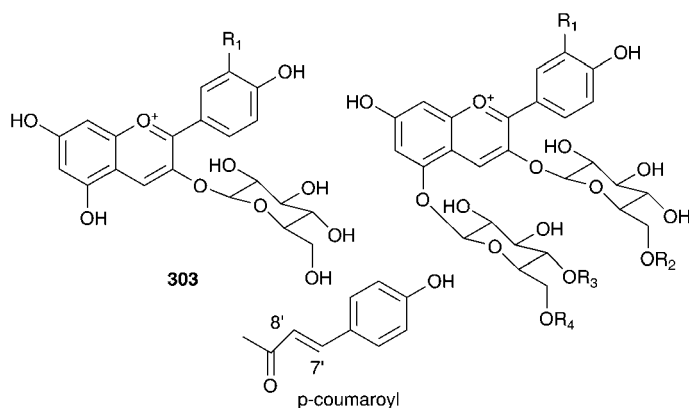
trans

298 R₁ = OH, R₂ = OH, R₃ = H, 7'/8' cis299 R₁ = OH, R₂ = OH, R₃ = H, 7'/8' trans300 R₁ = OMe, R₂ = OH, R₃ = H, 7'/8'

trans

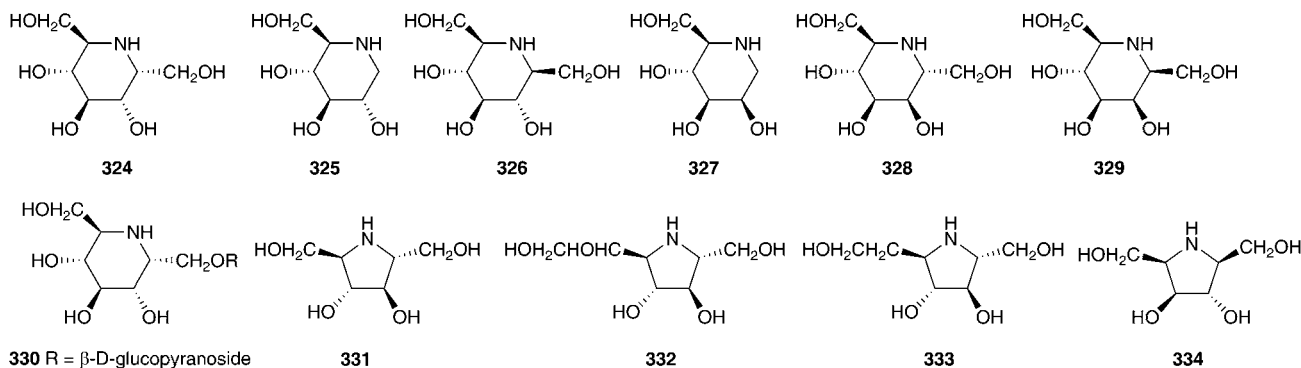
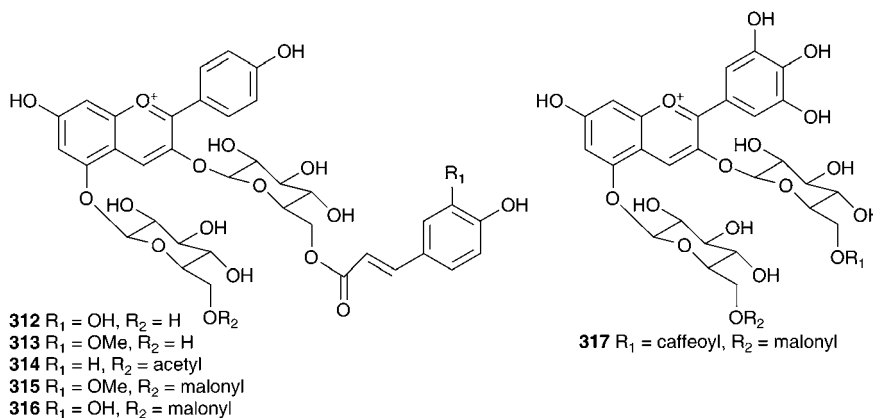
301 R₁ = OH, R₂ = H, R₃ = H, 7'/8' trans302 R₁ = H, R₂ = H, R₃ = H, 7'/8' trans322 R₁ = H, R₂ = OH, R₃ = H, 7'/8' trans323 R₁ = OH, R₂ = OMe, R₃ = H, 7'/8'

trans



303

304 R₁ = H, R₂ = H, R₃ = H, R₄ = H305 R₁ = H, R₂ = p-coumaroyl, R₃ = H, R₄ = H, 7'/8' cis306 R₁ = H, R₂ = p-coumaroyl, R₃ = H, R₄ = H, 7'/8' trans307 R₁ = H, R₂ = H, R₃ = H, R₄ = malonyl308 R₁ = H, R₂ = p-coumaroyl, R₃ = H, R₄ = malonyl, 7'/8' cis309 R₁ = H, R₂ = p-coumaroyl, R₃ = H, R₄ = malonyl, 7'/8' trans310 R₁ = H, R₂ = p-coumaroyl, R₃ = malonyl, R₄ = H, 7'/8' trans311 R₁ = OH, R₂ = p-coumaroyl, R₃ = H, R₄ = malonyl, 7'/8' trans318 R₁ = H, R₂ = caffeoyl, R₃ = H, R₄ = H319 R₁ = H, R₂ = feruloyl, R₃ = H, R₄ = H320 R₁ = H, R₂ = feruloyl, R₃ = H, R₄ = malonyl321 R₁ = H, R₂ = caffeoyl, R₃ = H, R₄ = malonyl



for cytotoxicity against HSC-2 human oral squamous cell carcinoma. Compounds **25** and **343–344** showed no activity while compounds **345–348** gave IC₅₀ values of 6.3–59 $\mu\text{g ml}^{-1}$, with the standard etoposide giving an IC₅₀ of 24 $\mu\text{g ml}^{-1}$. Compounds **335**, **336**, **338**, **340** and **341**, all 23*S* isomers, showed good activity relative to etoposide, but the corresponding 23*R* isomers showed no activity up to a dose of 100 $\mu\text{g ml}^{-1}$. The methanol extract of the bulbs of *Bellevalia romana* (L.) Sweet produced oligoglycosides of the eucosterol-type, four new compounds bellevaliosides A **349**, B **350**, C **351** and D **352** as well as compounds **86**, **87** and **353**, previously isolated from *Muscari* species.¹⁴⁰ In addition to the spirocyclic nortriterpenoids, five homoisoflavanones (**354–358**) were isolated.¹⁴¹ The bulbs of *Autonoe madeirensis* (as *Scilla maderensis* Menezes), have produced a slightly unusual phytochemical profile. An investigation of the ethanol extract of the bulbs resulted in the isolation of a range of 2-hydroxy di- and tricarboxylic acids and esters, *cis*- and *trans*-hydroxycinnamate esters and a flavone diglycoside **359**.¹⁴² A later study of the bulbs found 24*S*-ethyl-5 α -cholesta-7,22*E*-dien-3-ol- β -galactopyranoside **360** to be present.¹⁴³ Dias *et al.* isolated the pyrimidine derivative 2-(4'-aminobenzenamine)-pyrimidine and found it to be an α -adrenoreceptor antagonist.¹⁴⁴

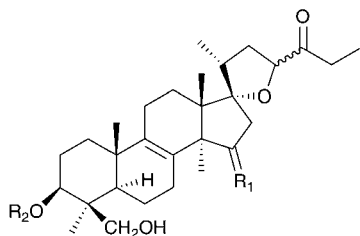
Work on the bulbs of *Hyacinthoides hispanica* (as *Scilla campanulata*) has focused on the isolation and structure determination of a mannose-specific lectin. Crystals have been grown and the structure determined by X-ray diffraction

studies.^{145–149} In addition to the mannose-binding lectin, a fetuin-binding lectin has been isolated and identified.¹⁵⁰ Polyhydroxylated alkaloids have been isolated from the ethanol extract of the bulbs of *H. hispanica* (as *Scilla campanulata*) and tested for glycosidase inhibitory activity. Compounds **361–368** were identified with compound **364** showing good inhibition of *Caldocellum saccharolyticum* β -glucosidase (IC₅₀ = 3.8 μM) and bovine liver β -galactosidase (IC₅₀ = 4.4 μM). Compound **366** was more active than compound **364** against *Caldocellum saccharolyticum* β -glucosidase (IC₅₀ = 0.34 μM), but less so of bovine liver β -galactosidase (IC₅₀ = 24 μM).¹⁵¹

Hyacinthoides non-scripta (bluebells) has long been known to be toxic to livestock. Horses suffer from abdominal pain and dysentery while cattle suffer from lethargy and dullness after consuming the plants. The toxic principles are the polyhydroxy pyrrolidine and pyrrolizidine alkaloids, common to many Hyacinthoideae.^{151,152} Alkaloids **361–367**, also found in *Hyacinthoides hispanica* (as *Scilla campanulata*), have been isolated from the leaves, stalks and immature fruits of the bluebells.^{151,152} A glucomannan has been isolated from the seeds of *H. non-scripta*. It is a linear polymer with *D*-glucopyranose and *D*-mannopyranose residues occurring in the ratio of 1 : 1.3.¹⁵³

Southern African genera that have received the attention of phytochemists are *Drimiopsis*, *Resnova*, *Veltheimia*, *Merwillia*, *Lachenalia*, *Pseudoprospero*, *Schizocarphus* and *Spetaea*. The range of compounds isolated is typical of the subfamily Hyacinthoideae,





335 (23S) $R_1 = 2H$, $R_2 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

336 (23S) $R_1 = O$, $R_2 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

337 (23R) $R_1 = O$, $R_2 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

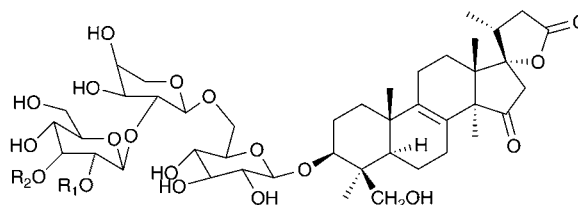
338 (23S) $R_1 = O$, $R_2 = \beta$ -apiofuranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

339 (23R) $R_1 = O$, $R_2 = \beta$ -apiofuranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

340 (23S) $R_1 = 2H$, $R_2 = [\alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)]- $[-\alpha$ -D-xylopyranosyl-(1 \rightarrow 2)- β -arabinopyranosyl-(1 \rightarrow 3)]- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

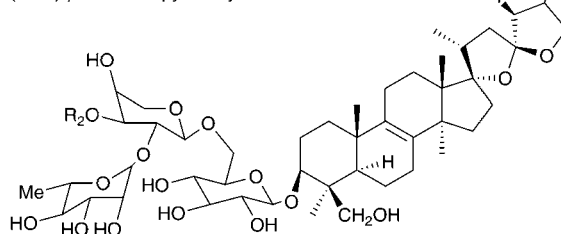
341 (23S) $R_1 = O$, $R_2 = [\alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)]- $[-\alpha$ -D-xylopyranosyl-(1 \rightarrow 2)- β -arabinopyranosyl-(1 \rightarrow 3)]- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside

342 (23R) $R_1 = O$, $R_2 = [\alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)]- $[-\alpha$ -D-xylopyranosyl-(1 \rightarrow 2)- β -arabinopyranosyl-(1 \rightarrow 3)]- β -D-glucopyranosyl-(1 \rightarrow 2)- β -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside



343 $R_1 = \alpha$ -L-rhamnopyranosyl, $R_2 = \beta$ -L-arabinopyranosyl

344 $R_1 = \alpha$ -L-rhamnopyranosyl, $R_2 = \alpha$ -L-arabinofuranyl-(1 \rightarrow 2)- β -L-arabinopyranosyl

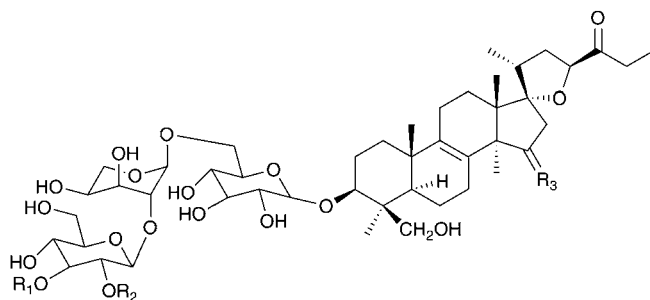


345 $R_1 = H$, $R_2 = \beta$ -L-arabinopyranosyl

346 $R_1 = H$, $R_2 = \alpha$ -L-arabinofuranyl-(1 \rightarrow 2)- β -L-arabinopyranosyl

347 $R_1 = OH$, $R_2 = \alpha$ -L-arabinofuranyl-(1 \rightarrow 2)- β -L-arabinopyranosyl

348 $R_1 = H$, $R_2 = H$



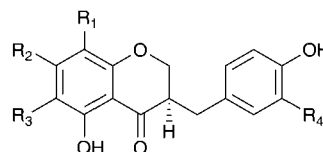
349 $R_1 = \alpha$ -L-rhamnosyl, $R_2 = D$ -apiofuranosyl-(1 \rightarrow 2)-D-xylopyranosyl, $R_3 = O$

350 $R_1 = \alpha$ -L-rhamnosyl, $R_2 = D$ -xylopyranosyl, $R_3 = O$

351 $R_1 = D$ -apiofuranosyl, $R_2 = D$ -apiofuranosyl-(1 \rightarrow 2)-D-xylopyranosyl, $R_3 = O$

352 $R_1 = \alpha$ -L-rhamnosyl, $R_2 = D$ -apiofuranosyl-(1 \rightarrow 2)-D-xylopyranosyl, $R_3 = OH$

353 $R_1 = \alpha$ -L-rhamnosyl, $R_2 = D$ -apiofuranosyl-(1 \rightarrow 2)-L-arabinopyranosyl, $R_3 = O$

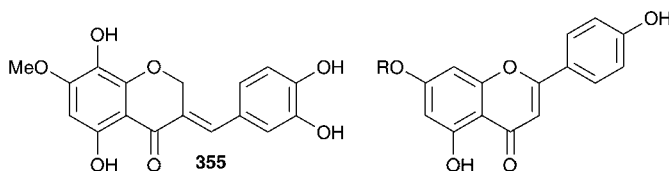


354 $R_1 = OMe$, $R_2 = OMe$, $R_3 = OH$, $R_4 = OH$

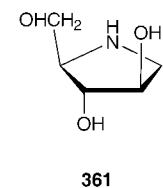
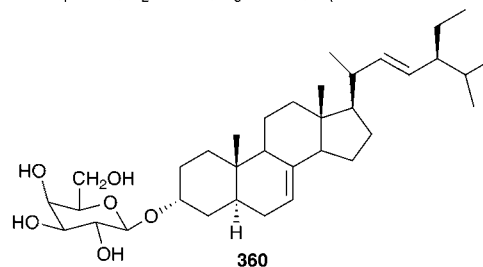
356 $R_1 = H$, $R_2 = H$, $R_3 = H$, $R_4 = OH$

357 $R_1 = OH$, $R_2 = OMe$, $R_3 = H$, $R_4 = H$

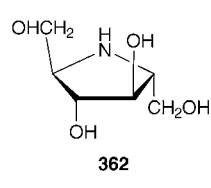
358 $R_1 = OH$, $R_2 = OMe$, $R_3 = OMe$, $R_4 = H$



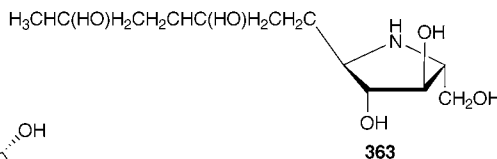
359 $R = \beta$ -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside



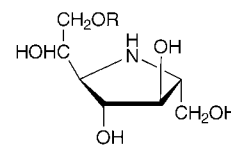
361



362



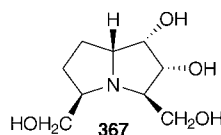
363



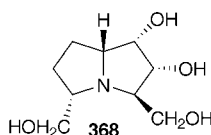
364 $R = H$

365 $R =$ apiofuranoside

366 $R = \beta$ -D-xylopyranoside



367

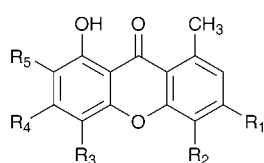


368

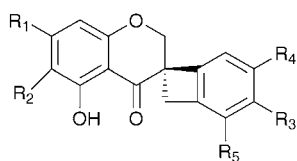


including homoisoflavanones, spirocyclic nortriterpenoids and xanthenes, as well as some more unusual alkaloids. Three species of *Drimiopsis* have been studied, *Drimiopsis maculata* Lindl. & Paxton, *Drimiopsis burkei* and *Drimiopsis barberi* Baker. *Drimiopsis maculata* is a southern African species used by traditional healers for treating a variety of complaints, including stomach complaints in children and the relief of constipation. It has not been found to be toxic to livestock.²⁵ The bulbs of this plant have yielded scillascillin-type homoisoflavanones (compound **106**, previously isolated from *Muscari botryoides* and compound **182**, previously isolated from *M. botryoides* and *M. armeniacum*),¹⁵⁴ xanthenes (drimiopsins A–E, **163**, **369**, **138** and **370–372**),²⁵ three homoisoflavanones (**30**, **59**, **373**) and a norlignan **374**.⁶⁹ *Drimiopsis burkei* is widespread throughout Botswana and Zimbabwe, occurring southward towards the Eastern Cape Province of South Africa.⁶⁹ In Botswana it is known as *thejane* and is used to treat stomach complaints.¹⁵⁵ A sample of this plant collected in KwaZulu-Natal, South Africa was found to contain the norlignan hinokiresinol **155** and a homoisoflavanone **375**,⁶⁹ while a sample collected in Mochudi, Botswana was found to contain the scillascillin-type homoisoflavanones (**248** and **376**), xanthenes (**137**, **138**, **369**, **377–378**), the norlignan hinokiresinol **155** and the

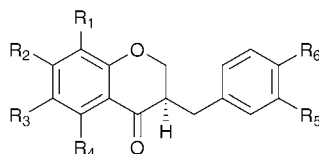
homoisoflavanones (*E*- eucomin **6**, **104**, **128** and **379–382**).¹⁵⁵ *Drimiopsis barberi* is the only genus member to occur in Cameroon, where it is used to treat fever.¹⁵⁶ The bulbs have been found to contain the unusual isoquinoline alkaloids (**383** and **384**) as well as the more expected scillascillin-type homoisoflavanones (**161**, **376**, **385–387**), 3-benzyl-4-chromanones (**61**, **214** and **388**) and *Z*-eucomin **6**. While the configuration at C-3 was not specified, the data compared favourably to known compounds of this type and would appear to be *R*.^{155,156} The homoisoflavanones **144**, **373**, **375**, **387** and **389–391**, were tested against the Gram-positive, methicillin-resistant *Staphylococcus aureus*. Compound **391** was found to have significant inhibitory activity with an MIC value of 0.47 mM. Compounds **389** and **375** showed some activity, but the quantities available were insufficient for conclusive testing.⁴¹ The homoisoflavanone **389**, showed significant vasodilating effects when tested on rat aortic rings (inhibition of the contraction elicited in aorta rings by high K⁺ gave pIC₅₀ of 5.31 M). This result would serve to validate the use of these bulbs for the treatment of conditions involving vasoconstriction.¹⁵⁷ *Resnova humifusa* (Baker) U.Müll.-Doblies & D.Müll.-Doblies, a South African species, earlier placed in *Drimiopsis* by Jessop⁴⁰ was found to contain nine homoisoflavanones (**7**, **10**, **30**, **56–60**, **392**). The same study reported seven



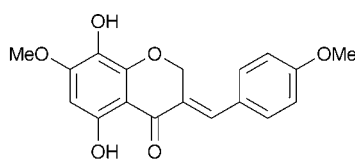
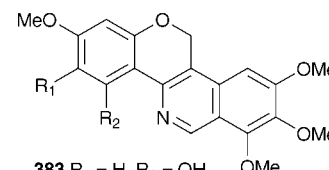
- 369** R₁ = OH, R₂ = H, R₃ = OMe, R₄ = OMe, R₅ = OMe
370 R₁ = OH, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = H
371 R₁ = OH, R₂ = H, R₃ = OMe, R₄ = OH, R₅ = H
372 R₁ = OMe, R₂ = H, R₃ = H, R₄ = OH, R₅ = OMe
377 R₁ = OH, R₂ = H, R₃ = H, R₄ = OMe, R₅ = OMe
378 R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OMe, R₅ = OMe



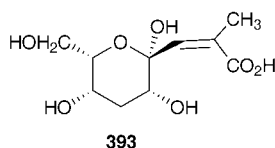
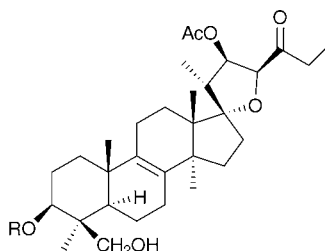
- 376** R₁ = OH, R₂ = H, R₃ = OMe, R₄ = OMe, R₅ = OMe
385 R₁ = OMe, R₂ = H, R₃ = OMe, R₄ = OMe, R₅ = OMe
386 R₁ = OMe, R₂ = H, R₃ = OMe, R₄ = OH, R₅ = OH
387 R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OMe, R₅ = H



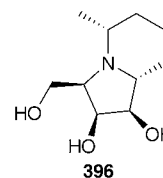
- 373** R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OH, R₅ = H, R₆ = OH
375 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OMe, R₅ = H, R₆ = OH
379 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OH
380 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OH
381 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OH
388 R₁ = H, R₂ = OH, R₃ = OH, R₄ = OH, R₅ = H, R₆ = OH
389 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = H, R₆ = OH
390 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe, R₆ = OH
391 R₁ = OMe, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = H, R₆ = OH
392 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OMe
394 R₁ = OH, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OH

**382**

- 383** R₁ = H, R₂ = OH
384 R₁ = OH, R₂ = H

**393**

- 395** R = β-D-glucopyranosyl-(1→6)-O-α-L-arabinopyranosyl-(1→2)-O-(3-O-β-D-glucopyranosyl-(1→3))-β-D-glucopyranosyl-(1→2)-O-β-D-apio-D-furanoside

**396**

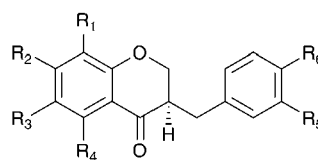
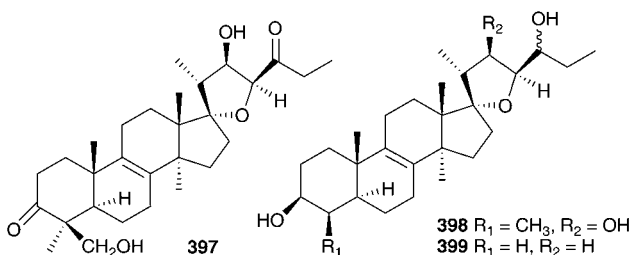
to be common with those found in the South African species *Eucomis montana* (7, 10, 30, 56, 58, 59, 60), which in addition contained compound 8.⁴⁰ A later study on the methanol extract of the bulbs of this plant resulted in the isolation of four homoisoflavanones (50, 58, 59, 61), a chalcone 132 and a tetrahydropyran derivative 393.⁴²

Veltheimia bracteata Harv. ex Baker is indigenous to South Africa. The petroleum ether and diethyl ether extracts of bulbs (as *Veltheimia viridifolia*) grown from seeds in Berlin have yielded two homoisoflavanones (394 and muscomin, 97)¹⁵⁸ as well as a spirocyclic nortriterpenoid 133 and its pentaglycoside 395 from the petrol and chloroform extracts respectively.¹⁵⁹ The homoisoflavanone 394, was found to exhibit good inhibitory activity against phosphodiesterase (PDE).¹⁵⁸ Compound 394 was found to inhibit the two PDE-isoenzymes by 70% and 75% at a concentration of 100 $\mu\text{mol l}^{-1}$.¹⁵⁸ *Veltheimia capensis* (L.) DC., native to the Western Cape, South Africa, has been found to contain an indolizidine alkaloid, steviamine 396. This alkaloid is also present in *Stevia rebaudiana* Bertoni (Asteraceae), but such alkaloids have not yet been isolated from any other Hyacinthaceae taxa.¹⁶⁰

Merwillia plumbea (Lindl.) Speta is widely used by southern African traditional healers. The Zulu use it as a purgative and to facilitate full term labour. The Sotho eat the cooked bulbs as an aperient, use bulb decoctions in enemas to treat internal tumours and for the treatment of lung disease in cattle. Powdered bulbs are rubbed on sprains and fractures by the Southern Sotho, while the Tswana believe that bulbs rubbed on the body will make them resistant to witchcraft. The Swati use a lotion prepared from the bulbs to treat veldsores and boils.^{161,162} Concern for the sustainability of *Merwillia plumbea* has led Ncube *et al.* to include the bulbs of this plant in a study to compare the antibacterial, anticandidal and cyclooxygenase inhibitory activity of the leaves and the, more commonly harvested, bulbs of some popular medicinal plants, harvested during different seasons in the year. Antimicrobial activities between the leaves and bulbs of *Merwillia plumbea* were found to

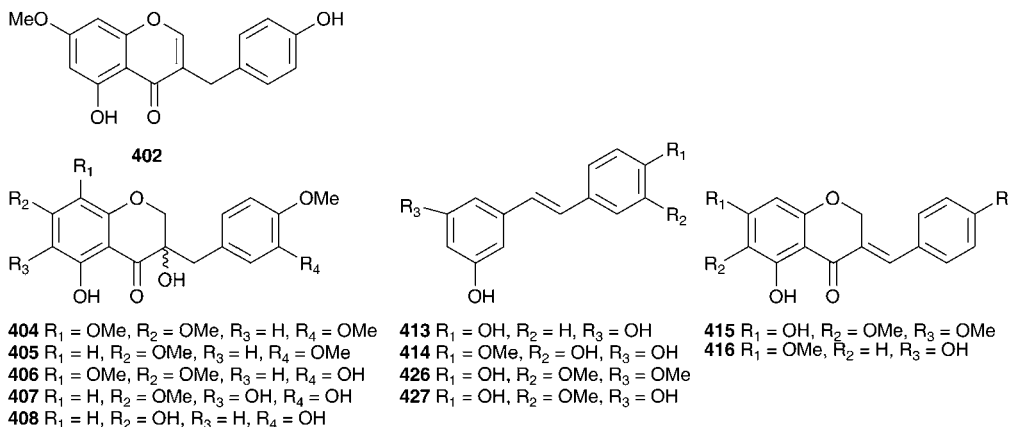
be comparable over all four seasons and as such they concluded that the leaves could be used in many cases as a substitute for the bulbs. Likewise both the leaves and bulbs produced good cyclooxygenase inhibitory activity throughout the year.^{163,164} A further study found that *in vitro* propagation of these plants could, under the right conditions, result in an increase in the flavonoid and gallotannin levels found in the shoots, which would allow for the successful mass propagation of these plants containing elevated levels of the potentially medicinally active components.^{110,161} A phytochemical investigation of the bulbs (as *Scilla natalensis* Planch.) found two homoisoflavanones 62 and 72, seven known eucosterol-type spirocyclic nortriterpenoids and the new spirocyclic nortriterpenoids 397, 398 and 399.¹⁶¹ An extract of the lectin-like proteins from the bulbs of *M. plumbea* (as *Merwillia natalensis* Planch.) have been found to exhibit moderate inhibitory activity (47%) against cyclooxygenase (COX-1).³¹ In a separate study, the homoisoflavanones 62 and 400 have been isolated from *M. plumbea* (as *Scilla natalensis* Planch.).¹⁶² In addition *M. plumbea* (as *S. natalensis*) has been shown to have good *in vitro* activity against the parasite *Schistosoma haematobium* (LC = 0.4 mg ml⁻¹).⁶⁶ Three homoisoflavanones (7, 62, 401) have been isolated from *Merwillia dracomontana* (Hilliard & B.L. Burt) Speta (as *Scilla dracomontana* Hilliard and Burt).¹⁶² A southern African member of the genus *Merwillia*, under its basionym *Scilla kraussii* Baker, was found to contain the homoisoflavanone, 80.¹⁶²

Lachenalia is the largest endemic genus of the Hyacinthaceae in southern Africa. Surprisingly, very little phytochemical work has been done on the genus.¹⁶⁵ A study of the leaves of *Lachenalia unifolia* Jacq. has yielded flavone sulphates (luteolin-3'-sulphate and -7,3'-disulphate, tricetin-3'-sulphate and -7,3'-disulphate, diosmetin-3'-sulphate and -7,3'-disulphate)¹⁶⁶ while the bulbs of *L. rubida* Jacq., collected from the Western Cape, South Africa, were found to contain the homoisoflavanones 402 and 403.¹⁶⁵ The bulbs of *Pseudoprosporo firmifolium* (Baker) Speta have been found to contain five 3-hydroxy-3-benzyl-4-chromanone type



- 400 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = OH, R₆ = OMe
401 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OMe
403 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = H, R₆ = OH
409 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = H, R₆ = OMe
410 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OMe, R₅ = OH, R₆ = OMe
411 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OMe, R₅ = H, R₆ = OH
412 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = H, R₆ = OMe
417 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OMe, R₅ = H, R₆ = OMe
418 R₁ = H, R₂ = OMe, R₃ = H, R₄ = OH, R₅ = OMe, R₆ = OH
419 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OH, R₆ = OMe
420 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OH, R₅ = OMe, R₆ = OH
421 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OH, R₅ = H, R₆ = OH
422 R₁ = H, R₂ = OH, R₃ = H, R₄ = OH, R₅ = OMe, R₆ = OMe
423 R₁ = H, R₂ = OMe, R₃ = OH, R₄ = OMe, R₅ = H, R₆ = OMe
424 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OH
425 R₁ = OH, R₂ = OMe, R₃ = H, R₄ = OMe, R₅ = H, R₆ = OMe
428 R₁ = H, R₂ = OMe, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OMe
429 R₁ = H, R₂ = OH, R₃ = OMe, R₄ = OMe, R₅ = H, R₆ = OMe





homoisoflavanones, all as racemic mixtures (**404–408**) as well as the spirocyclic nortriterpenoid, 15-deoxoeucosterol.¹⁶⁷ A subsequent study on the bulbs of *Pseudoprospero firmifolium* subsp. *natalensis* J.C. Manning confirmed the previous findings for the typical subspecies but, in addition, found the homoisoflavanone **59** in the dichloromethane extract.⁴²

Schizocarphus nervosus (Burch.) van der Merwe is widespread in southern Africa. Although toxic to livestock,¹ it plays an important role in traditional medicine within the region. The bulbs are used by the Zulu to treat dysentery and nervous conditions in children as well as treat rheumatic fever.¹⁶⁸ In Botswana the bulbs are used to enhance fertility and to treat infections.¹⁶⁹ The methanol extract of the bulbs of *S. nervosus* (as *Scilla nervosa* (Burch.) Jessop) has been tested for anti-inflammatory and anti-microbial activity. The crude methanol extract displayed a strong, but short-acting effect on a croton-oil induced contact dermatitis in a mouse ear (66% reduction of oedema after 3 h) and the nonpolar components were active against *Staphylococcus aureus* (IC₅₀ = 1.8 μg ml⁻¹), *Klebsiella pneumonia* (IC₅₀ = 2.0 μg ml⁻¹) and *Candida albicans* (IC₅₀ = 1.0 μg ml⁻¹). Little discrimination was noted between the Gram-positive and Gram-negative bacteria, but some specificity for the yeast was observed.¹⁶⁹ Five homoisoflavanones (**49** and **409–412**) and two stilbenoids (**413**, **414**) have been isolated from the bulbs.¹⁶⁸ A further thirteen homoisoflavanones (**41**, **49** and **415–425**) and three stilbenes (**414** and **426–427**) have been isolated from the bulbs of *S. nervosus* (as *Scilla nervosa* (Burch.) Jessop subsp. *rigidifolia*).^{170,171} Compound **419** was found to be highly active against colon cancer (HT-29 cell line ED₅₀ = 0.88 μg ml⁻¹) and breast cancer (MDA-MB-435 cell line ED₅₀ = 0.42 μg ml⁻¹).¹⁷⁰ The yellow inter-bulbscale deposits of this subspecies were likewise investigated and eight homoisoflavanones were isolated, two of which were novel (**428–429**).¹⁷² There has been some evidence presented to indicate that the bulbs may cause damage to liver cells.¹⁷³ A homoisoflavanone **400** has been isolated from *Spetaea lachenaliiflora* Wetschnig & Pfosser (as *Scilla plumbea* Lindl.). This is a little known species, neither toxic nor known to be used by traditional healers.⁶⁵

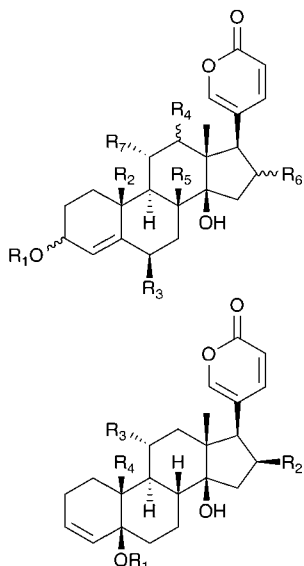
2 The Urgineoideae

The subfamily Urgineoideae, with more than 100 species,⁷ has received considerable taxonomic attention in recent years, with

widely differing opinions subsequently offered on generic circumscriptions. Speta recognised eleven genera,¹⁷⁴ several of which have been studied from both a phytochemical and a pharmacological perspective. More than any other compound type, the bufadienolides characterise the Urgineoideae. Bufadienolides are C-24 steroids possessing an α -pyrone group (doubly unsaturated six-membered lactone ring) at position 17 β . The presence of these highly toxic cardioactive steroids has led to these plants being used by man in antiquity, with the ancient Egyptians reportedly using preparations to treat heart disease.¹⁷⁵ Research into this subfamily was further stimulated by the sometimes fatal toxicity associated with various species, both to stock^{176–178} and to humans.^{179,180} The systematic position of the sea onion, *Charybdis maritima* (L.) Speta, has long been debated.¹⁸¹ The ancient Greeks referred to the plant as *skilla*, the Romans as *scilla* and for much of the early research on this plant, the names *Urginea maritima* or *Scilla maritima* were applied. Subsequent work by Pfosser and Speta¹⁸¹ provided evidence for two genera in the subfamily Urgineoideae in the Mediterranean region, *Urginea* Steinh. and *Charybdis* Speta. Phytochemical investigations to date support this generic distinction.

With much of the early research on *Charybdis maritima* focused on the plant then known as *Scilla maritima*, the trivial names of isolates of this urgineoid accordingly reflect this nomenclatural association. Notably, *Scilla* is a distinct genus currently assigned to the subfamily Hyacinthoideae, and has a very different chemical profile. Kopaczewski isolated a toxic component from the bulbs of *C. maritima* (as *Scilla maritima*) which he termed scillitin.¹⁸² It was found to be a diuretic but the whole extract from the bulbs was found to be more poisonous to guinea pigs and rabbits than scillitin itself.¹⁸³ In contrast, Smith found scillitin to be more toxic to rats than the crude extract.¹⁸⁴ The bufadienolides scillaren A **224**, scilliroside **430**, proscillaridin A **223** and scillaren F **431** have also been isolated from *C. maritima* (reported as *Scilla maritima* and/or *Urginea maritima*).^{185–190} Using HPLC, with proscillaridin A **223** as a reference, Tittel and Wagner showed the possible presence of bufadienolides in a plant identified as *Scilla rubra* L. (here considered synonymous with *C. maritima*).¹⁹¹ Petrol extracts of bulbs of this species (as *Scilla maritima* and *Urginea maritima*) were found to contain fatty acids (lauric acid, myristic acid,





- 223 : $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_7 = \text{H}$
 224 $R_1 = \alpha\text{-L-rhamnopyranosyl-(1}\rightarrow\text{4)-}\beta\text{-D-glucopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_7 = \text{H}$
 430 : $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{CH}_3$, $R_3 = \text{OAc}$, $R_4 = \text{H}$, $R_5 = \text{OH}$, $R_6 = \text{H}$, $R_7 = \text{H}$
 433 : $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_7 = \text{H}$
 435 : $R_1 = \beta\text{-D-glucopyranosyl-(1}\rightarrow\text{4)-}\beta\text{-D-glucopyranosyl-(1}\rightarrow\text{4)-}\alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_7 = \text{H}$
 436 : $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 = \text{H}$, $R_5 = \text{H}$, $R_6 = \text{H}$, $R_4 = \alpha\text{-OH}$, $R_7 = \text{H}$
 437 : $R_1 = \beta\text{-D-glucopyranosyl-(1}\rightarrow\text{4)-}\alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 = \text{H}$, $R_5 = \text{H}$, $R_6 = \text{H}$, $R_4 = \alpha\text{-OH}$
 449 $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CHO}$, $R_3 - R_6 = \text{H}$, $R_7 = \text{OH}$
 488 $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{CHO}$, $R_3 = \text{H}$, $R_4 = \beta\text{-OH}$, $R_5 - R_7 = \text{H}$

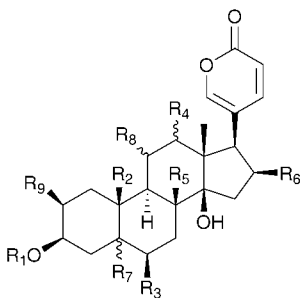
- 431 $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{H}$, $R_3 = \text{H}$, $R_4 = \text{CHO}$
 434 $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{OAc}$, $R_3 = \text{H}$, $R_4 = \text{CHO}$
 478 $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{H}$, $R_3 = \text{OH}$, $R_4 = \text{CHO}$
 493 $R_1 = \beta\text{-D-glucopyranoside}$, $R_2 = \text{H}$, $R_3 = \text{H}$, $R_4 = \text{COOH}$
 494 $R_1 = \text{H}$, $R_2 = \text{H}$, $R_3 = \text{H}$, $R_4 = \text{CHO}$

palmitic acid, palmitoleic acid, stearic acid, oleic acid, linoleic acid and linolenic acid) and sterols (sitosterol, stigmasterol, campesterol and cholesterol).^{192–194}

Investigations carried out in the early 1920s focused primarily on *Charydhis maritima* (squill). Both the extracts of fresh bulbs and dried powdered bulbs showed digitalis-like activity on the heart when tested on frogs and cats.^{195,196} Markwalder¹⁹⁶ found no difference in activity between the fresh and dried bulbs and noted that neither the dried outer leaves nor the heart of the bulb exhibited much activity, while the mature middle layers of the bulb were rich in active components. The structure of scillaren A 224, the principle active component of *C. maritima* (reported as *Urginia maritima*) was elucidated by Stoll *et al.* in 1933.¹⁹⁷ The interesting activity shown by the squill bufadienolides resulted in some comparative studies with digitoxigenin and strophanthidin as well as the effects of combination therapy.^{198,199} Extensive work on the bulbs of *C. maritima* (as *Urginia maritima*) over the following years resulted in the isolation and identification of a large number and variety of bufadienolides. They can be divided into those having a double bond at C-4 (*e.g.*, 224), those having a double bond at C-3 (*e.g.*, 431) and those without either (dihydro C-4) (*e.g.*, 432). Without exception, within *C. maritima* they contain a β -hydroxy

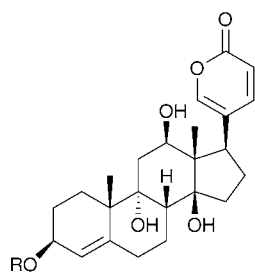
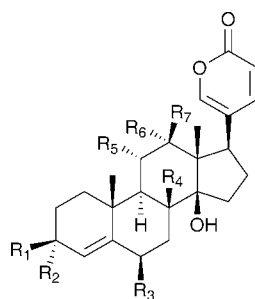
group at C-14, a *trans* junction of rings B and C and a *cis* junction of rings C and D. They have been isolated as the aglycones as well as a variety of glycosides, with scillaren A 224 and proscillaridin A 223 being present in all specimens of *C. maritima* investigated. Samples of the plant from across its Mediterranean, European and Middle Eastern range have been investigated and this would account for the great diversity of bufadienolides isolated.

Von Wartburg and Renz^{200,201} isolated five bufadienolides from *C. maritima* (as *Urginia maritima*), scilliroside 430, scillaren A 224, scillaren F 431, proscillaridin A 223 and scillarenin- β -D-glucopyranoside 433. The structures reported at that time were subsequently revised and the structures shown here reflect that revision.^{202,203} Gorlich²⁰⁴ found proscillaridin A 223, scillaren A 224, scilliglaucoside (scillaren F) 431 and scillicyanoside 434 in crystalline form as well as minor amounts of scillicoeloside (no structure given) and glucoscillaren A 435. Later work on this plant resulted in the isolation of further minor components, scilliphaeoside 436 and glucoscilliphaeoside 437.²⁰⁵ A reinvestigation of the bulbs of *C. maritima* (as *Urginia maritima*) in 1991 by Krenn *et al.*²⁰⁶ resulted in an additional four new bufadienolides, identified as 5 α -4,5-dihydroproscillaridin A 432, 5 α -4,5-dihydroglucoscillaren A 438,



- 432 $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_9 = \text{H}$, $R_7 = \alpha\text{-H}$
 438 $R_1 = \beta\text{-D-glucopyranosyl-(1}\rightarrow\text{4)-}\beta\text{-D-glucopyranosyl-(1}\rightarrow\text{4)-}\alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_9 = \text{H}$, $R_7 = \alpha\text{-H}$
 439 $R_1 = \alpha\text{-L-rhamnopyranosyl-}\beta\text{-D-glucopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_6 = \text{H}$, $R_7 = \beta\text{-H}$, $R_8 = \alpha\text{-OH}$, $R_9 = \text{H}$
 440 $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CHO}$, $R_3 - R_6 = \text{H}$, $R_7 = \alpha\text{-H}$, $R_8 = \text{H}$, $R_9 = \text{H}$
 477 $R_1 = \alpha\text{-L-rhamnopyranosyl-}\beta\text{-D-glucopyranoside}$, $R_2 = \text{CH}_3$, $R_3 - R_6 = \text{H}$, $R_7 = \beta\text{-H}$, $R_8 = \alpha\text{-OAc}$, $R_9 = \text{H}$
 490 $R_1 = \alpha\text{-L-rhamnopyranoside}$, $R_2 = \text{CH}_3$, $R_3 = \text{H}$, $R_4 = \text{H}$, $R_5 = \text{H}$, $R_6 = \text{H}$, $R_7 = \text{H}$, $R_8 = \alpha\text{-OH}$, $R_9 = \text{H}$



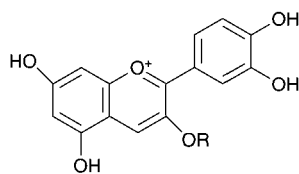
476 R = α -L-rhamnopyranoside479 R₁ = OH, R₂ = OCH₃, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = OH480 R₁ = =O, R₃ = H, R₄ = H, R₅ = OH, R₆ = H, R₇ = H481 R₁ = OH, R₂ = OH, R₃ = H, R₄ = H, R₅ = H, R₆ = OH, R₇ = H482 R₁ = -O- β -D-glucopyranoside, R₂ = H, R₃ = OAc, R₄ = H, R₅ = H, R₆ = H, R₇ = H483 R₁ = -O- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside, R₂ = H, R₃ = OAc, R₄ = H, R₅ = H, R₆ = H, R₇ = H484 R₁ = -O- α -L-rhamnopyranosyl-(1 \rightarrow 4)- α -L-rhamnopyranoside, R₂ = H, R₃ = H, R₄ = OH, R₅ = H, R₆ = H, R₇ = H485 R₁ = -O- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside, R₂ = H, R₃ = OAc, R₄ = OH, R₅ = H, R₆ = H, R₇ = H486 R₁ = -O-3'-OAc- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside, R₂ = H, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = H487 R₁ = -O-2'-OAc- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside, R₂ = H, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = H489 R₁ = -O- α -L-rhamnopyranosyl- α -L-rhamnopyranoside, R₂ = H, R₃ = H, R₄ = H, R₅ = H, R₆ = OH, R₇ = H491 R₁ = -O- α -L-rhamnopyranoside, R₂ = H, R₃ = H, R₄ = OH, R₅ = H, R₆ = H, R₇ = H492 R₁ = -O- β -D-glucopyranoside, R₂ = H, R₃ = H, R₄ = OH, R₅ = H, R₆ = H, R₇ = H

3-O- β -D-glucopyranoside **433**, proscillaren A **223**, scilliphaesidin 3- β -D-glucopyranoside **441**, scilliglucoside **431**, scilliphaoside **436** and 12-*epi*-scilliphaeoside. In addition, the bulbs from Tunisia contained glucoscilliphaeoside **437**, 12-*epi*-glucoscilliphaeoside, 12 β -hydroxyscilliglucosidin 3-O- β -D-glucopyranoside **488**, 12-*epi*-scilliphaeosidin 3-O- β -D-glucopyranoside **443** and 12-*epi*-scilliphaeosidin 3-O- α -L-rhamnopyranosyl- α -L-rhamnopyranoside **489**. Bulbs sourced from Sardegna also contained gamabufotalin 3-O- α -L-rhamnopyranoside **490**, scillirubrosidin 3-O- α -L-rhamnopyranoside **491**, scillirubroside **492**, 12 β -hydroxyscilliroside **446**, 5 α -4,5-dihydroscillirosidin 3-O- α -L-rhamnopyranosyl- β -D-glucopyranoside **475**, deacetylscilliroside **443**, 10-carboxy-5 β -14 β -hydroxybufa-3,20,22-trienolide 5-O- β -D-glucopyranoside **493** and scilliglucogenin **494**.

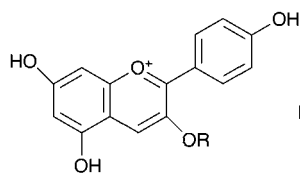
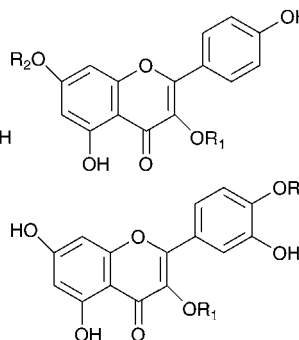
The flavonoid, anthocyanin and sterol content of the bulbs of *C. maritima* (as *Urginia maritima*) have been investigated. The

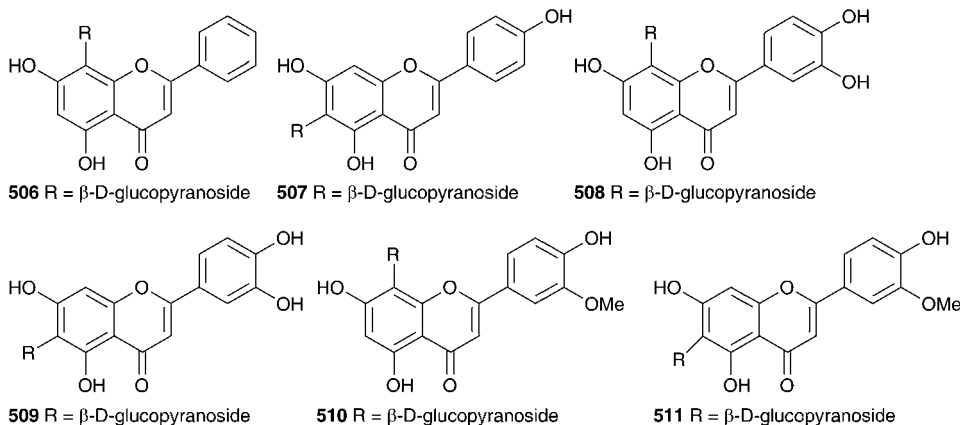
ethanol extract of the bulbs was studied by gas-liquid chromatography and the sterols identified by comparison to known reference compounds. Sitosterol, stigmasterol, campesterol, cholesterol, Δ^5 -avenasterol and Δ^7 -avenasterol were found to be present.^{193,194} Red bulbs of *C. maritima* (as *Urginia maritima*), collected in the Balearic Islands contained cyanidin **495**, cyanidin-3-monoglucoside **496** and pelargonidin-3-monoglucoside **497**, both in the free form and acylated with *p*-coumaric acid.²¹²⁻²¹⁶ The proportion of anthocyanin was found to be higher in tetraploids than in triploids or hexaploids.²¹⁷

The flavonoids isolated included kaempferol-7-O- β -D-glucopyranoside-3-O- β -D-triglucopyranoside **498**, kaempferol-7-O- β -D-glucopyranoside-3-O- α -L-rhamnopyranosyl- β -D-glucopyranoside **499**, kaempferol-7-O- β -D-glucopyranoside-3-O- β -D-diglucopyranoside **500** and kaempferol-7-O- α -L-rhamnopyranoside-3-O- α -L-rhamnosyl- β -D-glucopyranoside **501**.²¹⁸ In addition the



495 R = H

496 R = β -D-glucopyranoside497 R = β -D-glucopyranoside498 R₁ = tri- β -D-glucopyranoside, R₂ = β -D-glucopyranoside499 R₁ = α -L-rhamnopyranosyl- β -D-glucopyranoside, R₂ = H500 R₁ = di- β -D-glucopyranoside, R₂ = H501 R₁ = α -L-rhamnopyranosyl- β -D-glucopyranoside, R₂ = α -L-rhamnopyranoside502 R₁ = tri- β -D-glucopyranoside, R₂ = H502 R₁ = H, R₂ = β -D-glucopyranoside503 R₁ = β -D-glucopyranoside, R₂ = H504 R₁ = H, R₂ = H

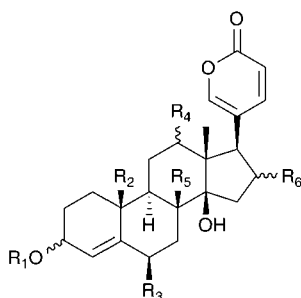


flavonoid O-glycosides **502–505**, have been isolated.²¹⁹ Five C-glycosylflavones have been isolated from the ethyl acetate extract of the bulbs. They have been identified as vitexin **506**, isovitexin **507**, orientin **508**, isoorientin **509**, scoparin **510** and isoscoparin **511**.^{220–223} Fernandez *et al.* identified twenty-five flavonoids in a Spanish sample of *C. maritima* bulbs (as *Urginia maritima*). They followed a parallel series: pelargonidin, kaempferol, cyaniding, quercetin and taxifolin. A complex mixture of both C- and O-glycosides was obtained.²²⁴ A fructan, sinistrin, has been isolated from *C. maritima* (as *Urginia maritima*) and found to be a mixed type β -D-fructan, with (2–1) linked and (2–6) linked unbranched β -D-fructofuranosyl and α -D-glucopyranoside residues.^{225,226}

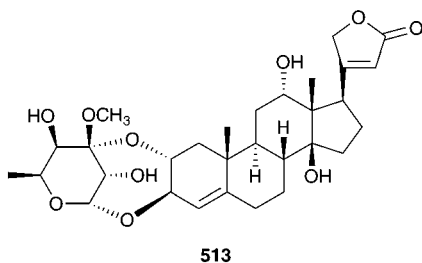
C. maritima (reported as *Urginia maritima*) has been used as a form of rodenticide since the 13th century.²⁰² The toxic component has been found to be scilliroside **430**, which affects the cardiovascular and central nervous systems, resulting in death.²⁰² Scilliroside has an emetic effect on humans, cats, dogs and pigeons, but rats and mice are unable to vomit and as such they die within a few hours of ingesting the bulbs. The flavonoid

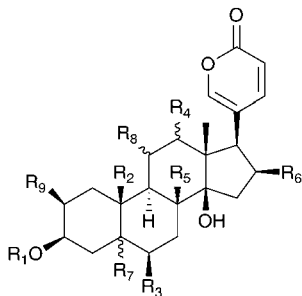
fraction has been found to have a diuretic effect²²⁷ and the ethanolic extract of Spanish squill has shown good insecticidal activity against the stored grain pest *Tribolium castaneum* (red flour beetle).²²⁸ Twelve wild populations of the bulbs were studied with the red bulbs giving 60–90% mortality if applied at 20–30 $\mu\text{g insect}^{-1}$.²²⁸ An extract of the bulbs of *C. maritima* (as *Urginia maritima*) growing in Turkey has been found to exhibit greater toxicity than cisplatin against the A549 non-small cell lung cancer (NSCLC) cell line.²²⁹ Tissue cultures of *C. maritima* (as *Urginia maritima*) grown both in the dark and under light showed no sign of the bufadienolides commonly associated with this species. The cultures grown in the light did though produce anthocyanins.²³⁰

The range of bufadienolides found in the bulbs of *Charybdis hesperia* (Webb & Berthel.) Speta (as *Urginea hesperia* Webb & Berthel.) indicates a close relationship to *C. maritima*. Thirteen bufadienolides have been isolated including scillarenin (**512**) and a variety of associated glycosides.²³¹ *Urginea fugax* (Moris) Steinh. is a species which is widespread in the Mediterranean area. In contrast to *C. maritima* and *C. hesperia*, cardenolides



- 512 R₁ = H (β), R₂ = CH₃, R₃ - R₆ = H
 518 R₁ = H, R₂ = CH₃, R₃ = OAc, R₄ = H, R₅ = OH, R₆ = H
 519 R₁ = H, R₂ = CH₃, R₃ = OH, R₄ = H, R₅ = OH, R₆ = H
 520 R₁ = β -D-glucopyranoside, R₂ = CH₃, R₃ = OH, R₄ = β -OH, R₅ = OH, R₆ = H
 522 R₁ = =O, R₂ = CH₃, R₃ = OAc, R₄ = OH, R₅ = OH, R₆ = H
 523 R₁ = =O, R₂ = CH₃, R₃ = H, R₄ = OH, R₅ = OH, R₆ = H
 532 R₁ = H (β), R₂ = CHO, R₃ - R₆ = H
 533 R₁ = =O, R₂ = CHO, R₃ - R₆ = H
 534 R₁ = α -L-rhamnopyranoside, R₂ = CHO, R₃ - R₆ = H
 535 R₁ = β -D-glucopyranoside, R₂ = CHO, R₃ - R₆ = H
 536 R₁ = α -L-rhamnopyranosyl-(1→3)-[β -D-glucopyranosyl(1→4)]- α -L-rhamnopyranoside, R₂ = CH₃, R₃ - R₆ = H
 546 R₁ = H, R₂ = CHO, R₃ - R₅ = H R₆ = OAc
 549 R₁ = α -L-rhamnopyranoside, R₂ = CH₃, R₃ = H, R₄ = OH, R₅ - R₆ = H





- 514** R₁ = α -L-rhamnopyranosido-(1 \rightarrow 4)- β -D-glucopyranoside, R₂ = CH₃, R₃ = H, R₄ = =O, R₅ = H, R₆ = H, R₇ = β -H, R₈ = α -OH, R₉ = H
515 R₁ = α -L-rhamnopyranosyl- β -D-glucopyranoside, R₂ = CH₃, R₃ = OAc, R₄ = β -OH, R₅ = OH, R₆ = H, R₇ = β -H, R₈ = H, R₉ = H
521 R₁ = H, R₂ = CH₃, R₃ = OAc, R₄ = β -OH, R₅ = OH, R₆ = H, R₇ = β -H, R₈ = H, R₉ = H
530 R₁ = H, R₂ = CHO, R₃-R₆ = H, R₇ = β -OH, R₈ = H, R₉ = H
531 R₁ = β -D-glucopyranoside, R₂ = CHO, R₃-R₆ = H, R₇ = β -OH, R₈ = H, R₉ = H
537 R₁ = α -L-rhamnopyranoside, R₂ = CH₃, R₃ = H, R₄ = =O, R₅ = H, R₆ = H, R₇ = H, R₈ = α -OH, R₉ = H
538 R₁ = β -D-digitalopyranoside, R₂ = CHO, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = β -OH, R₈ = H, R₉ = H
539 R₁ = β -D-glucopyranosyl-(1 \rightarrow 4)-O-(2-O-acetyl- α -L-thevetopyranoside), R₂ = CHO, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = β -OH, R₈ = H, R₉ = OAc
542 R₁ = H, R₂ = CH₃, R₃-R₅ = H, R₆ = β -OH, R₇ = β -H, R₈ = H, R₉ = H
543 R₁ = β -D-galactopyranoside, R₂ = CH₃, R₃-R₅ = H, R₆ = β -OH, R₇ = β -H, R₈ = H, R₉ = H
557 R₁ = H, R₂ = CHO, R₃ = H, R₄ = H, R₅ = H, R₆ = OH, R₇ = β -OH, R₈ = H, R₉ = H
558 R₁ = β -D-glucopyranoside, R₂ = CHO, R₃ = H, R₄ = H, R₅ = H, R₆ = OH, R₇ = β -OH, R₈ = H, R₉ = H
559 R₁ = β -D-glucopyranoside, R₂ = CH₃, R₃ = H, R₄ = H, R₅ = H, R₆ = H, R₇ = β -H, R₈ = H, R₉ = H
560 R₁ = 6-deoxy-3-O-methyl- β -L-glucopyranoside, R₂ = CHO, R₃-R₆ = H, R₇ = β -H, R₈ = H, R₉ = H
561 R₁ = β -D-glucopyranosyl-(1 \rightarrow 2)-6-deoxy-3-O-methyl- β -L-glucopyranoside, R₂ = CHO, R₃-R₆ = H, R₇ = β -H, R₈ = H, R₉ = H
562 R₁ = 6-deoxy-3-O-methyl- β -L-glucopyranoside, R₂ = CHO, R₃-R₅ = H, R₆ = =O, R₇ = β -OH, R₈ = H, R₉ = H
563 R₁ = H, R₂ = CHO, R₃-R₆ = H, R₇ = β -H, R₈ = H, R₉ = H

have been isolated from the bulbs.²³² The new cardenolide fugaxin **513** has a rubellin-like carbohydrate linkage between C-3 and C-2, unlike any seen in *C. maritima* and *C. hesperia*, which lends chemotaxonomic support to recognition of both *Urginea* and *Charybdis* in the Mediterranean region.¹⁸¹ Six bufadienolides have been isolated from the bulbs of *Charybdis pancration* (Steinh.) Speta (as *Urginea pancration*), scilliglaucoside **431**, scillirubroside **492**, scilliroside **430**, 5 α -4,5-dihydroscillirosidin 3-O- α -L-thevetopyranosyl- β -D-glucopyranoside **475**, arenobufagin-3 β -O-L-rhamnopyranosyl-4'- β -D-glucopyranoside **514** and the glycoside **515**.^{233,234} *Charybdis aphylla* (Forssk.) Speta (as *Urginea aphylla* (Forssk.) Speta), growing in the northeastern Mediterranean region was found to contain eleven bufadienolides, scillicyanoside **434**, gamabufotalin-3-O- α -L-rhamnopyranoside **490**, scilliglaucoside **431**, gamabufotalin-3-O- α -L-rhamnosyl- β -D-glucopyranoside **439**, proscillaridin A **223**, sciliphaeosidin-3-O- β -D-glucopyranoside **437**, scillarenin-3-O- β -D-glucopyranoside **433**, 16 β -O-acetylgamabufotalin-3-O- α -L-rhamnopyranoside (16 β -O-acetyl **436**), 12 β -hydroxyscilliroside **446** and 5 α -4,5-dihydroxyscillirosidin-3-O- α -L-thevetopyranosyl- β -D-glucopyranosyl- β -D-glucopyranoside (**475** glucoside).²³⁵ The similarity in chemical composition of the bulbs of *Charybdis* from across the European region highlights their close relationship.

As early as 1923, Juritz reported the presence of a glucoside in the toxic South Africa plant *Boosia macrocentra* (Baker) Speta (as *Urginea macrocentra* Baker) with digitalis-like activity and he noted that it could be used as a substitute for squill.²³⁶ This plant has also been implicated in stock mortality and it is used by the Zulu as a vermifuge (as *Drimia macrocentra*).²³⁷ The major component was identified as rubellin **516** by NMR

spectroscopy.²³⁷ In 1927, work by Watt²³⁸ on South African *Sekanama burkei* (Baker) Speta (as *Urginea burkei*), yielded two glucosides, a red compound found to be toxic to animals and having a digitalis-like effect on the heart as well as a diuretic effect, and a white compound which resulted in paralysis of the central nervous system and respiration. Additional work on *S. burkei* (as *Urginea burkei*) and *Urgineopsis modesta* (Baker) Speta (as *Urginea rubella* Baker) in the 1940s and 1950s resulted in the isolation and identification of two cardiac glycosides, transvaalin and rubellin **516** respectively.²³⁹⁻²⁴³ Rubellin **516** was found to behave as a cardiac poison towards rats while transvaalin was found to kill rats primarily by its effect on the central nervous system.^{241,243,244} Rubellin was found to have a digitalis-like action on cats.²⁴⁵ Further work by Louw *et al.*²⁴⁶ on transvaalin led to the conclusion that it was either isomeric with scillaren A **224** or a stable complex of scillaren A **224** and scilliroside **430**. This was based on melting point analysis, toxicity tests and the fact that hydrolysis of transvaalin led to the same products as the hydrolysis of scillaren A, namely scillaridin A **517** and scillabiose (disaccharide). Zoller and Tamm²⁴⁷ and Tschesehe and Hottemann²⁴⁰ all concluded that transvaalin and scillaren A were identical compounds. The structure of rubellin **516** was fully elucidated by Steyn *et al.* in 1986²⁴⁸ using high-field NMR spectroscopy. Later work by Krenn *et al.*²⁴⁹ on *Sekanama sanguinea* (Schinz) Speta (as *Urginea sanguinea* Schinz) yielded eight bufadienolides as well as a steroidal sapogenin and stigmaterol. The first six bufadienolides, namely scillirosidin **518**, deacetylscillirosidin **519**, 12 β -hydroxyscillirosidin **445**, 12 β -hydroxydeacetylscillirosidin **520**, 12 β -hydroxyscilliroside **446** and 5 α -4,5-dihydro-12 β -hydroxyscillirosidin **521** had been isolated previously from other members of the Urgineoideae. Two

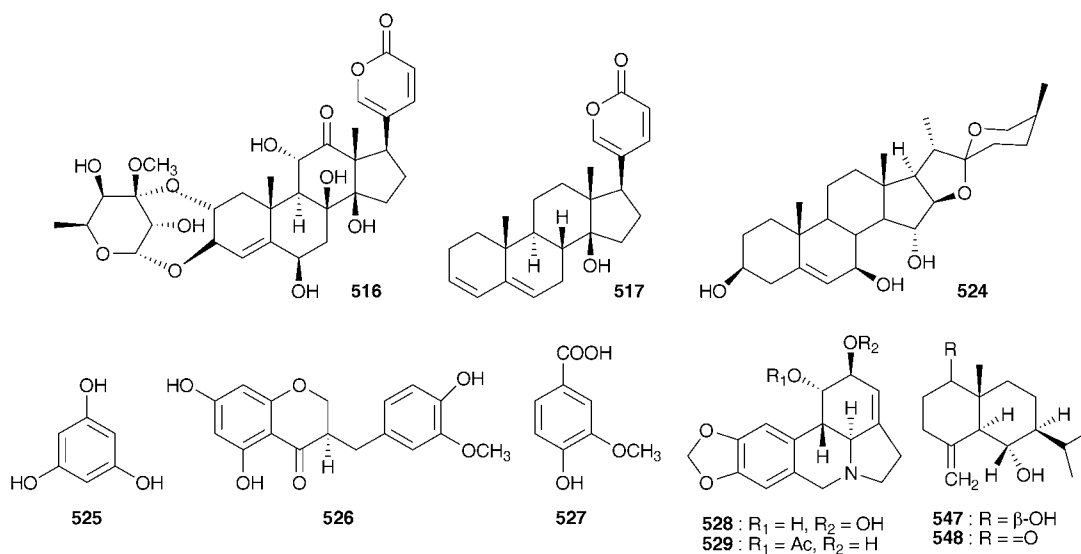


new compounds, 12 β -hydroxyscillirosidin-3-one **522** and 12 β -hydroxyscillirubrosidin-3-one **523** were identified using NMR spectroscopy. The steroidal saponin, 7 β ,15 α -dihydroxyyamogenin **524** was the first such compound to be isolated from a member of the Urgineoideae. Majinda *et al.*²⁵⁰ investigated another specimen of the same plant, and found stigmasterol, phloroglucinol **525**, phloroglucinol 1- β -O-glucopyranoside, scillaren A **224** and the novel 5 α -4,5-dihydroscillaren A. In addition, salicylic acid and 3-hydroxy-4-methylbenzoic acid were isolated. An ethnopharmacological study of *S. sanguinea*, using patient records from Ga-Rankuwa Hospital in South Africa revealed a number of cases of poisoning as a result of ingestion. The amount usually prescribed by traditional healers does not result in adverse side effects, but the patients admitted to the hospital had taken more than the prescribed dose. A brine shrimp assay showed a seasonal variation in toxicity, with the plant being more toxic in February (late Summer) than in September (Spring).²⁵¹ *Sekana nama sanguinea* has traditionally been used for many different conditions, including asthma and it is often taken by pregnant women. A study by Marx *et al.*²⁵² (as *Urginea sanguinea*) showed significant damage to chick embryos after exposure to this plant. They concluded that it should be used with extreme care by traditional healers.²⁵³ Undoubtedly the bufadienolides dominate the chemistry of the Urgineoideae, including the *Sekana nama* species. However, the isolation of a novel homoisoflavanoid from the bulbs of *Sekana nama delagoensis* (Baker) Speta (as *Drimia delagoensis* (Baker) Jessop) indicates that a broader range of secondary metabolites can be expected, if not commonly found.²⁵⁴ The homoisoflavanoid **526** and 3-methoxy-4-hydroxybenzoic acid **527**, were isolated from the dichloromethane and methanol extracts of the bulbs respectively.²⁵⁴

Other sub-Saharan members of the Urgineoideae to receive attention include *Urginavia altissima* (L.f.) Speta (as *Urginea altissima* (L.f.) Baker and *Drimia altissima* (L.f.) Baker), *Fusifilum physodes* (Jacq.) Raf. ex Speta (as *Urginea physodes* (Jacq.) Baker),

Urginavia epigea (R.A.Dyer) Speta (as *Urginea epigea* R.A.Dyer), *Urginea hydenburgensis* R.A.Dyer and *Urginea riparia* Baker. Characteristic bufadienolides have been isolated and identified, many of which include either an aldehyde group at C-10 or a rubellan type carbohydrate group doubly linked to the aglycone at positions 2 α and 3 β . The best known of this group is the widespread *Urginavia altissima* (often referred to as *Drimia altissima* or *Urginea altissima*). Rat populations need to be controlled to limit threats to public health and minimise crop damage. The toxicity of *U. altissima* has been investigated with respect to the field rat, *Arvicanthis abyssinicus*.²⁵⁵ Synthetic rodenticides affect non-target species and can be expensive. The powdered bulb was tested and found to be toxic at levels of 5% of the daily food intake. If the percentage was lower, no mortality was noted. *Urginavia brachystachys* (Baker) Speta has (as *Drimia brachystachys* (Baker) Stedje) reportedly been used as an arrow poison.²⁵⁶

Both bufadienolides and alkaloids have been isolated from *U. altissima*. The alkaloids, lycorine **528** and acetylcaranine **529** have not been found by any subsequent investigators and the identity of the plant investigated by Miyakado and co-workers²⁵⁷ has been called into question as bulbs of *U. altissima* may be easily confused with those of some Amaryllidaceae, especially the genus *Crinum* L.^{258,259} Six bufadienolides have been isolated from *U. altissima* collected in Kenya, all of which have an aldehyde at C-10 (**530–535**).^{260,261} A South African sample of this plant was found to contain 12 β -hydroxyscillirosidin **445**,²⁵⁸ which has been isolated from *S. sanguinea* previously and urginin **536** with an interesting trisaccharide substituent at C-3.²⁵⁸ A specimen from Ethiopia yielded different bufadienolides again, gamabufotalin-3-O- α -L-rhamnopyranoside **490** and arenobufagin-3-O- α -L-rhamnopyranoside **537**.²⁵⁹ It would appear that significant regional differences exist in the type of bufadienolide found in *U. altissima*. *U. altissima* is widely used for the treatment of gout, rheumatism and a number of respiratory conditions, including asthma.²⁵⁸ This plant is also used by Zimbabwean traditional healers to treat skin conditions. It has been found to produce a mild

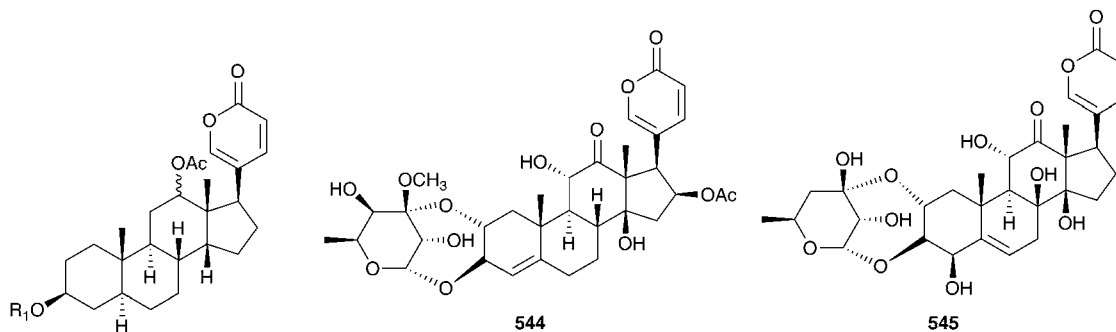


inflammation and itching if rubbed on the skin. This effect has been found to be due to the presence of calcium oxalate raphides in the bulbs and as such the appropriateness of its use as a skin treatment has been questioned.²⁶²

Fusifilum physodes (as *Urginea physodes* (Jacq.) Baker) was found to contain four bufadienolides, physodines A 538, B 539, C 540 and D 541. Two of which, physodines A and B, both having a C-10 aldehyde group, were found to be toxic to weaned guinea pigs (fractions not causing death in 48 h at 200 mg kg⁻¹ were considered non toxic). Physodines C and D were of interest in that they were the first naturally occurring 14-deoxybufadienolides to be isolated.²⁶³ An ethanolic extract of the bulbs has been shown to have antifungal activity.²⁶⁴ The methanol extract of the bulbs of *Fusifilum depressum* (Baker) U.Müll.-Doblies J.S.Tang & D.Müll.-Doblies (as *Drimia depressa* (Baker) Jessop) was found to contain two bufadienolides, 542 and 543.²⁶⁵ The leaves of this plant are toxic to cattle, sheep and rabbits and the plants are used by the Sotho as a good luck charm or to cause harm to one's enemy.²⁶⁵ Novel rubellin type bufadienolides, riparianin 544 and lydenburgenin 545, have been isolated from *Urginea riparia* and *Urginea lydenburgensis* R.A.Dyer respectively.^{237,266} In addition, *Urginea lydenburgensis* was found to contain the novel bufadienolide, scillicyanosidin 546, with an aldehyde group at C-10. Riparianin 544 showed moderate activity against MCF7 (breast), TK10 (renal) and UACC62 (melanoma) cell lines (total growth inhibition of <3.5, <7 and <10.5 ppm respectively). *Urginavia epigea* is used locally to treat backaches or as a soap. The smoke from smouldering bulbs is also used as a treatment for headaches and colds.²⁶⁷ In addition to the buadienolide 517,²⁶⁸ two eudesmane sesquiterpenoids were isolated, 547 and 548. These were the first to be reported from the Hyacinthaceae and due to their occurrence in *U. epigea* but not the oft-confused *U. altissima*, provide species level taxonomic markers.²⁶⁷ The bufadienolide 517, has an unusual 3,5-diene system, readily obtained synthetically, but not commonly isolated directly from plant sources.²⁶⁸ The butanol extract of the bulbs of *U. epigea* was found to have significant molluscicidal activity against the bilharzia-carrying snail, *Bulinus africanus*. This result has raised some concern with regards its use as soap and the potential toxic affect it may have on other aquatic biota.²⁶⁹

An Indian member of the Urgineoideae, known locally as 'jungli-piyaz' or 'bhuikanda' and Indian squill has been used

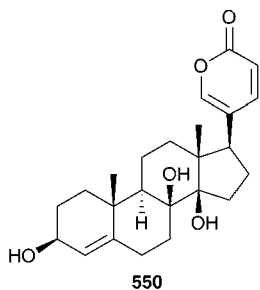
traditionally for stomach complaints, as a cardiac stimulant, a diuretic, as well as to treat asthma, cough or bronchitis and various infections.²⁷⁰ Commercially available samples vary with some plants having a truncated bulb and others a scaly bulb.²⁷¹ For much of the work reported, the plant has been identified as *Urginea indica* (Roxb.) Kunth or *Scilla indica* Baker. The name *Indurgia indica* (Roxb.) Speta is currently accepted. Extracts of the bulbs have shown some activity against *Neisseria gonorrhoeae* infections²⁷² and have had outstanding antimicrobial activity attributed to them in a survey of medicinally used plants in India.²⁷³ The ethanolic extract of the bulbs of *I. indica* (as *Scilla indica* Baker) has been found to have good antioxidant activity (protection against lipid peroxidation induced by ferrous sulphate in a dose-dependant manner, with 49% protection for hydroxyl radical-mediated hydroxylation at a concentration of 250 µg ml⁻¹) as well as to possess cardiac activity on a par with ouabain.^{271,274} Rao and Rangaswami have detected bufadienolides in extracts of the bulbs of *I. indica* (as *Scilla indica* Roxb.) and have isolated scillaren A 224 as the principle active component.^{275,276} Jha and Sen, however, found no trace of scillaren A 224 in a plant identified as *Scilla indica* at the time.²⁷⁷ The principle bufadienolides isolated from this plant (as *Urginia indica* Kunth) by various investigators are scillaren A 224 and proscillaridin A 223,^{275,278-281} with scillaren A being reported as early as 1956. These are also the principle bufadienolides of the European squill *C. maritima*. In small doses, the action of *I. indica* is the same as *C. maritima*, in that it slows the heart beat and increases urine flow and it can successfully be used as a substitute for *C. maritima*.²⁸² Jha and Sen^{278,280} have studied the twelve different cytotypes of *I. indica* (as *Urginea indica*), belonging to four ploidy levels. Scillaren A 224 and proscillaridin A 223 were present in all twelve. Bulbs were investigated through three successive years. The tetraploids showed the highest concentration of these bufadienolides, followed by diploids and then triploids. Significant variation in the bufadienolide concentration was also noted between samples collected in northern and southern India, with the northern samples showing much lower levels of proscillaridin A 223 and scillaren A 224.²⁸³ In addition, two minor components, scilliphaeoside 436 and anhydroscilliphaeosidin, were found to be present in the tetraploid cytotypes and



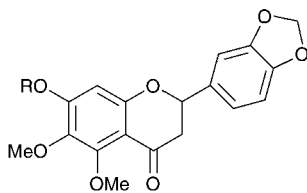
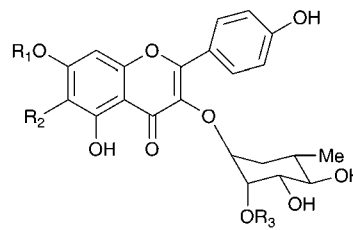
540 R₁ = β-D-xylopyranosyl-(1→4)-O-α-L-rhamnopyranoside

541 R₁ = β-D-glucopyranosyl-(1→3)-O-4-acetyl-α-L-rhamnopyranoside





550

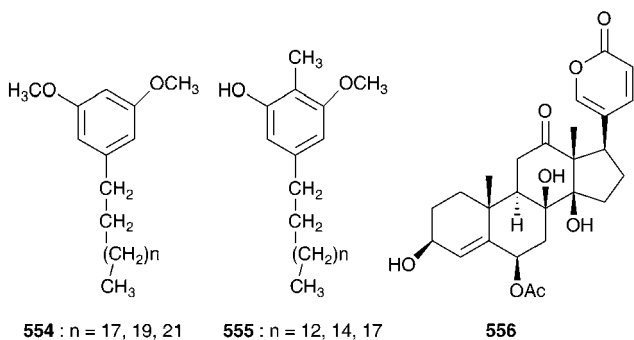
551 R = 6''- β -D-glucopyranosyl- β -D-glucopyranoside552 R₁ = 6-*p*-coumaroyl- β -D-glucopyranoside, R₂ = C-glucopyranoside, R₃ = H
553 R₁ = 6-*p*-coumaroyl- β -D-glucopyranoside, R₂ = -C-glucopyranoside, R₃ = - β -D-glucopyranosyl

scilliphaeoside **436** in the triploids. Likewise the sterol content was investigated with only minor differences noted between the ploidy levels. Stigmasterol predominated, with sitosterol and campesterol being found in the tetraploid cytotypes and some of the triploids respectively.^{284,285} Kopp and Danner²⁸¹ found the bulbs of *I. indica* (as *Urginia indica*) to contain six bufadienolides, scillarenin **512**, proscillaridin A **223**, scillarenin A **224**, scilliphaeosidin 3-*O*- α -L-rhamnopyranoside **549**, scilliglucosidin 3-*O*- α -L-rhamnopyranoside **534** and scilliglucosidin 3-*O*- β -D-glucopyranoside **535**, while Saxena and Chaturvedi isolated 6-desacetoxy scilliroside **550**.²⁸⁶ In addition to the bufadienolides and sterols, Sultana *et al.* have isolated three novel flavonoid glycosides (**551–553**) from bulbs collected in Bangladesh.²⁸⁷

Seasonal variation in the glycosidal content of the bulbs has been observed with the highest such levels found during October at the onset of dormancy. The levels steadily decrease to reach a minimum in June.²⁸⁸ Sterol content has been found to peak during the pre-flowering and pre-vegetative growth stages and it has been suggested that the sterols could be a trigger for the onset of reproductive and vegetative growth in the species.²⁸⁹ Significant antifungal activity has been noted and attributed to the presence of a 29-kDa glycoprotein which inhibited growth of *Fusarium oxysporum* at a concentration of 10 μ g protein well⁻¹.^{290,291} This protein has in addition shown good antitumour activity against the growth of an ascites tumour and mouse mammary carcinoma (tested *in vivo* using 6–8 week old female mice).²⁹² The bulbs of *I. indica* (as *Urginia indica* L.) collected in Nigeria were found to contain six alkylresorcinols of the structure types shown in **554** and **555**.²⁹³ An ethanolic extract of *I. indica* (as *Scilla indica*), has reportedly shown good

inhibition of the Semliki Forest Virus and the active principal found to be a diosgenin saponin.²⁹⁴

The genus *Drimia* Jacq. is closely related to *Urginea*. *Drimia robusta* Baker is widely used in southern Africa by traditional healers. It has been used as a diuretic, as a treatment for feverish colds, to promote the healing of broken bones, to treat diseases of the uterus and to relieve stabbing pains in the chest.^{258,295} Luyt *et al.*²⁹⁵ investigated the biological activity of this plant in order to substantiate its use in traditional medicine. Anti-bacterial activity was found to be present in the ethyl acetate extract of the bulbs. An ethanolic extract of the bulbs showed higher inhibitory activity than indomethacin in the screening for inhibition of cyclooxygenase. This inhibition of prostaglandin synthesis (tested for with the cyclooxygenase assay) could explain the plant's use as a treatment for feverish colds. Further work on the COX-1 inhibition of a water extract of the bulbs showed activity above 70%.²⁶⁴ Evidence for the presence of proscillaridin A **223** was presented: TLC and NMR data were compared with a standard sample. Luyt *et al.*²⁹⁶ then compared bulbs derived from tissue culture with those grown naturally. A standard proscillaridin A **223** was used for comparison and the UV spectra and qualitative TLC results were compared. The results indicated that the bufadienolide content was the same in the *in vitro*-derived plants as in those grown naturally. This is in contrast to the work done on *C. maritima*, where no bufadienolides were found in the tissue cultured bulbs.²³⁰ Phytochemical studies of the bulbs of *D. robusta* yielded eight bufadienolides, 12 β -hydroxyscilliroside **445**, 6 β -acetoxy-3 β ,8 β ,14 β -trihydroxy-12-oxobufa-4,20,22-trienolide **556**, scilliroside **430**, 12 β -hydroxyscilliroside **446**, hellebrigenin-3-*O*- β -glucopyranoside **531**, 16 β -hydroxyhellebrigenin **557**, 16 β -hydroxyhellebrigenin-3-*O*- β -glucopyranoside **558** and 5 β ,16 β -dihydroxybufalin-3-*O*- β -glucopyranoside **559** as well as three common aromatic acids, 4-hydroxy-3-methoxybenzoic acid, 3,4-dihydroxybenzoic acid and *trans*-3-(4'-hydroxyphenyl)-2-propenoic acid.^{258,268,297} The toxicity of *Rhodocodon madagascariensis* Baker, a plant endemic to Madagascar, has been tested against the embryo-larvae of medaka fish (*Oryzias latipes*) and an LD₅₀ of 1 mg ml⁻¹ was established.²⁹⁸



554 : n = 17, 19, 21

555 : n = 12, 14, 17

556



toxic and as early as 1938, the presence of a cardioactive glucoside was reported.²⁹⁹ Katz and Tschesche *et al.* have between them reported the presence of several bufadienolides (bovisides A 560, B, C, D, bovogenin E, bowieasubstanz G, glucobovoside A 561, nabogenin (scilliglaucosidin) 532, bovoruboside 562 and bovogenin A 563) between the years of 1950 and 1959.^{300–314} Structures have been reported for nabogenin 532, bovoruboside 562, bovoside A 560, glucobovoside A 561 and bovogenin A 563, although the structure reported for bovoruboside is not certain. The cardioactive glycosides from *B. volubilis* attracted much attention in the early years of their discovery. The activity was initially found to be in the region of strophanthin,³¹⁵ the action was as rapid and lasting with a cumulative effect, though not as much as digitoxin.^{316–318} Solutions of bovoside A were found to be unstable, due to the aldehyde group at C-19 being vulnerable to oxidation. Reduction of this group to the alcohol (bovosidol A) gave a stable solution which could then be assessed for activity.^{319,320} Material from East Africa (as *Bowiea kilimandscharica* Mildbr.) was initially reported to contain two bufadienolides, kilimanscharogenins A and B. Katz concluded that kilimanscharogenin A was the same compound as bovogenin A 563.³²¹

Chen and Henderson^{322,323} carried out a comparative survey of the cardioactivity of twenty-nine bufadienolides and cardenolides from various sources. Included in the survey were bufadienolides from *B. volubilis*. The tests were carried out on etherised cats by slow continuous intravenous injection and the electrocardiograms were recorded to confirm the digitalis-like action of these compounds. The potency of each compound was determined in terms of lethal dose (LD), the amount (mg kg⁻¹) needed to kill a cat of approximately 2 kg in 30–60 min. The bufadienolides were found to be more potent than the cardenolides, with the glycosides seldom showing more activity than the aglycones. The methyl C-10 analogues were less active than the compounds with an aldehyde group in the C-10 position and 3-epimerisation and 5 α -configuration reduced the activity. A later study by Chen increased the number of cardenolides and bufadienolides to fifty and similar profiles were obtained using the same assay system.³²⁴

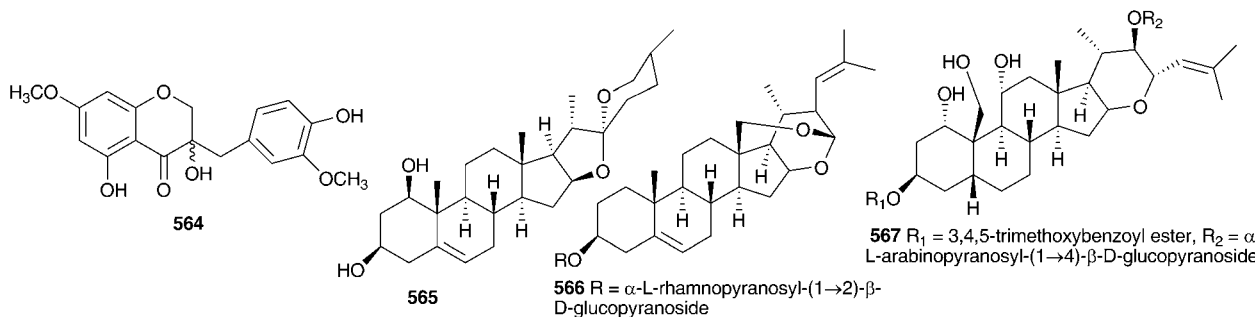
3 The Ornithogaloideae

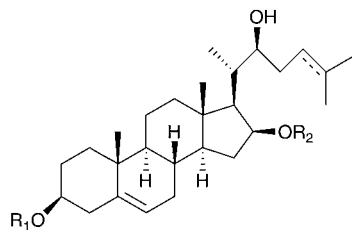
The Ornithogaloideae are widespread throughout Africa, Europe and southwest Asia. This group consists of approximately 280 species,⁵ with a recent phylogenetic analysis by

Martínez-Azorín *et al.*⁶ recognising nineteen monophyletic genera. Phytochemical studies on the Ornithogaloideae have centred on *Ornithogalum* L. and *Galtonia* Decne., primarily driven by the significant biological activity attributed to the compounds isolated. In addition, a small amount of work has been done on *Albuca* L., *Eliokarmos* Raf., *Stellarioides* Medik., *Loncomelos* Raf. and *Honorius* Gray. The types of compounds isolated are strongly linked to the geographic region the plant species derive from, with the African taxa containing primarily cholestane glycosides and spirosterols, and the European taxa cardenolides. Some flavonoids and sterols have been isolated from plants sourced from all regions studied. Additionally, a limited number of homoisoflavanones have been identified from the bulbs.

The genus *Albuca* L. is widespread in southern Africa, with some species occurring in the Arabian Peninsula. The bulbs of *Albuca fastigiata* Dryand. have been investigated and only one compound was isolated, a homoisoflavanoid, 564.³²⁵ *Albuca fastigiata* is found on the East coast of South Africa and is used by the Zulu as a protective charm and to treat malicious food poisoning.³²⁵ Another southern African species, *Albuca setosa* Jacq., has been tested for anti-inflammatory activity in a carrageenan-induced rat paw edema assay.³²⁶ This plant is used by traditional healers to treat wounds, among other conditions, and was found to significantly inhibit carrageenan-induced paw edema for a 3-hour challenge (43–55% over 3 h at 150 mg kg⁻¹ and 68–85% for 300 mg kg⁻¹). *Albuca setosa* has also been used by traditional healers in the Nkonkobe municipality in South Africa for the treatment of diabetes.³²⁷ As part of a study on the steroidal saponins of various monocot families, Okanishi *et al.*³²⁸ found *Albuca nelsonii* N.E.Br. to contain ruscogenin 565. This was identified by comparison of melting points, mixed melting points, optical rotations, infrared and mass spectra with an authentic sample.

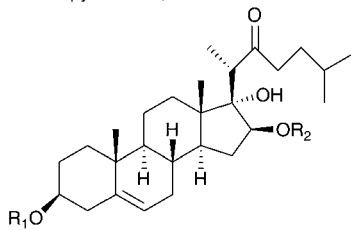
Galtonia candicans (Baker) Decne. is native to South Africa, and was investigated by the laboratories of Kuroda and Mimaki^{329–333} who used bioassay-directed fractionation to isolate the active components of the bulbs. The methanol extract was found to be active against leukemia HL-60 cells (IC₅₀ = 0.017 μ M). This was further partitioned between n-butanol and water, with the activity being retained in the n-butanol fraction (IC₅₀ = 0.0056 μ M). This was found to contain the hexacyclic rearranged cholestane diglycoside, candicanside A 566, the polyoxygenated 5 β -cholestane diglycoside galtonioside A 567, six cholestane





568 R₁ = β-D-glucopyranoside, R₂ = 3-O-acetyl-α-L-rhamnopyranoside

569 R₁ = β-D-glucopyranoside, R₂ = 3-O-acetyl-α-L-rhamnopyranoside, Δ²⁴



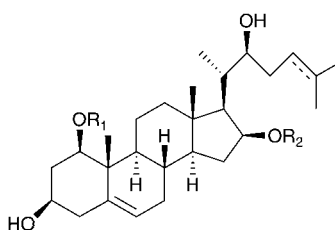
575 R₁ = β-D-glucopyranoside, R₂ = (2-O-(E)-cinnamoyl-β-D-xylopyranosyl)-(1→3)-2-O-acetyl-α-L-arabinopyranoside

576 R₁ = β-D-glucopyranoside, R₂ = (2-O-3,4-dimethoxybenzoyl-β-D-xylopyranosyl)-(1→3)-2-O-acetyl-α-L-arabinopyranoside

577 R₁ = β-D-glucopyranoside, R₂ = β-D-xylopyranosyl-(1→3)-2-O-acetyl-α-L-arabinopyranoside

578 R₁ = β-D-glucopyranoside, R₂ = α-L-arabinopyranoside

580 R₁ = H, R₂ = α-L-arabinopyranoside



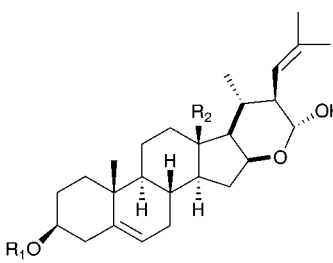
570 R₁ = β-D-glucopyranoside, R₂ = β-D-glucopyranoside

571 R₁ = β-D-glucopyranosyl-(1→6)-β-D-glucopyranoside, R₂ = β-D-glucopyranoside, Δ²⁴

572 R₁ = β-D-glucopyranosyl-(1→6)-β-D-glucopyranoside, R₂ = β-D-glucopyranoside

573 R₁ = α-L-rhamnopyranoside, R₂ = β-D-glucopyranoside

574 R₁ = β-D-glucopyranoside, R₂ = β-D-glucopyranoside



579 R₁ = α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranoside, R₂ = CH₂OH

bisdesmosides **568–573** and six cholestane glycosides **574–579**.^{329–333} Candicanoside A **566** and galtonioside A **567** were found to have potent activity against leukemia HL-60 cells (IC₅₀ = 0.032 μM and 0.057 μM respectively) and further testing on a 38 cell line assay showed differential cytotoxicities against breast cancer, CNS cancer and lung cancer lines, while colon cancer, ovarian cancer and stomach cancer cell lines were resistant to them.^{330,331} Compound **569** showed moderate activity against leukemia HL-60 cell lines (IC₅₀ = 6.8 μM)³³² and compounds **575–578** showed good activity using the same assay (IC₅₀ = 0.00012 μM, 0.00048 μM, 0.0024 μM and 0.053 μM respectively).³³³ Tang and Yu have achieved the total synthesis of candicanoside A **566**, using a 27-step synthesis.^{334,335} Another southern African species of *Galtonia* that has received some attention is *Galtonia princeps* (Baker) Decne.³³⁶ A methanol extract of the bulbs yielded the cholestane derivative **580**. Both the bulbs and aerial leaves of *Galtonia viridiflora* L. Verdc. were found to contain monoglycosides of caffeic acid, principally rhamnose and glucose.³³⁷

Work on the bulbs of *Galtonia saundersiae* (Baker) Mart.-Azorín, M.B. Crespo & Juan (as *Ornithogalum saundersiae*), a native of the East coast of South Africa, began in the early 1990s with the isolation of three acylated cholestane glycosides **581–583**. All three were examined for their inhibitory effect on cyclic AMP phosphodiesterase. Compounds **582** and **583** showed good activity (IC₅₀ = 0.00025 μM and 0.00020 μM respectively), suggesting that the presence of a benzoyl group attached to the sugar moiety is essential for activity.³³⁸ Six additional cholestane

glycosides **584–589** were isolated and tested for their inhibitory activity on cyclic AMP phosphodiesterase. Compounds **585** and **588**, with the acyl group at the C-3 hydroxyl position of the rhamnose group, showed good activity (IC₅₀ = 9.9 × 10⁻⁵ μM and 10.9 × 10⁻⁵ μM respectively), as did compound **589** (40% inhibition at 0.08 mg ml⁻¹), while the other compounds were inactive.^{339,340} Compound **590**, isolated from the bulbs of *G. saundersiae* (as *Ornithogalum saundersiae*), showed significant inhibition (IC₅₀ = 3.1 μM) of the proliferation of peripheral blood lymphocytes produced by a patient with chronic renal failure, suggesting potential as an immunosuppressive agent. Rearranged cholestane glycosides (**591–599**) with a six-membered hemiacetal ring E, between C-16 and C-23, have been isolated. The conformation of ring E was determined through molecular mechanics and molecular dynamics calculations.³⁴¹ These compounds were tested for anti-tumour activity, primarily against leukaemia HL-60 cells and MOLT-4 cells. Compounds **591** (IC₅₀ = 21.0 nM and 18.0 nM), **592** (IC₅₀ = 9.2 nM and 3.2 nM) and **597–599** (IC₅₀ = 0.019, 0.063 and 0.052 μM for HL-60 cells) showed significant cytotoxic activity, primarily through the induction of apoptosis. These results would indicate that the presence of an aromatic ester group at the glycoside moiety is essential for good activity.^{342–348} Further investigation of the methanol extract of the bulbs resulted in the isolated of the related cholestane glycoside **600**.³⁴⁹ In addition to the rearranged cholestane glycosides mentioned above, ten new (**575, 601–609**) and three previously isolated (**567–569**) cholestane glycosides, with a glycosidic moiety at C-16 have been



(called OSW1) has been the subject of much synthetic work. The full synthesis of the compound itself has been accomplished, and a large number of analogues have been produced for structure–activity relationship studies. Studies indicate that OSW1 acts by damaging the mitochondria and inducing apoptosis through an increase of cytosolic calcium and thus the activation of calcium dependent apoptosis. It has been demonstrated that OSW1 is active against some cells that are resistant to conventional anti-cancer agents, in particular fludarabine-resistant chronic lymphocytic leukemia cells.³⁵⁹ In addition to the work on the anti-tumour activity of *G. saundersiae*, the ethanol extract of the whole dried plant has been tested for its protective effects against acute hepatic failure. The authors concluded that it did provide some benefit, primarily through suppressing oxidative stress, lipid peroxidation and apoptosis of hepatocytes, as well as reducing inflammation.^{360,361}

Species from the genus *Ornithogalum* have received substantial attention. While early work on the genus began in the 1950s on the European members of the genus, later work has included species from Turkey and Iran.

Work on the European species of *Ornithogalum* was initially centred on *Ornithogalum umbellatum* L. (common Star of Bethlehem). European species of *Ornithogalum* are characterised by the presence of cardenolides, with some flavonoids also identified. Extracts of the bulbs of *O. umbellatum* were found to have a digitalis-like effect on frog, rabbit and cat hearts. In all cases, a positive inotropic action was noted, decreased conduction and finally systolic standstill.^{362–364} A clinical trial found that when tablets of *O. umbellatum* were substituted for digitoxin there was a loss of activity and fluid retention increased. When the tablets were coated to protect them from stomach acid, the digitalis-like action was re-established but at only half the expected strength. It was concluded that only half the active components were being absorbed. When compared to digitoxin, the drug showed less slowing of the heart, a greater diuretic effect and less gastrointestinal nausea.³⁶⁵ The principal active component of *O. umbellatum* was found to be the cardenolide convallatoxin **610**.³⁶⁶ The cytotoxicity of convallatoxin **610** was studied. It was found to have an IC_{50} of $002 \mu\text{g ml}^{-1}$ when assayed against Eagle's KB strain of human carcinoma and structure–activity studies showed that the glycosidic portion was essential for activity and that the β -hydroxy groups at C-5 and C-14 and the aldehyde at C-10 significantly enhanced the activity.³⁶⁷ The convallatoxin **610** content in the bulbs can be increased by defloration of the plant.³⁶⁸ In total, sixteen cardenolides (**610–626**) have been isolated from the bulbs of *O. umbellatum* L.^{366,369–371} Later phytochemical investigations of the bulbs resulted in the isolation and identification of two flavonoids (**627** and **628**)³⁷² while the four flavonoids (**627–630**) were isolated from the leaves.^{372,373} Compounds **629** and **630** were also isolated from the leaves of *Ornithogalum algeriense* Boiss. and *Ornithogalum kochii* Parl.³⁷³ Sabudak and Oyman reported the presence of 3-O-(2'-methoxy-4'-(2-pentenyl))-phenylsitosol in the bulbs of *O. umbellatum*³⁷⁴ and a study of the roots colonized by the fungus *Glomus intraradices* found a complex range of apocarotenoids.³⁷⁵ A range of cardenolides have been isolated from the species *Loncomelos magnum* (Krasch. & Schischk.) Speta

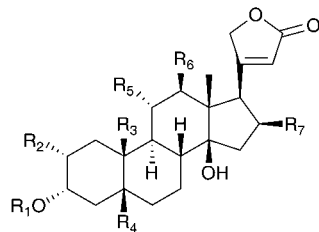
(as *Ornithogalum magnum*) (**610**, **614**, **631–635**),^{376–382} *Honorius nutans* (L.) Gray (as *Ornithogalum nutans*) (**610**, **631**, **632**, **636–664**),^{383,384} *Ornithogalum gussonei* Ten. (**631**, **635**, **665**)³⁸⁵ and *Honorius boucheanum* (Kunth) Holub (as *Ornithogalum boucheanum*) (**666–673**).³⁸⁶ The aerial parts of *O. gussonei* yielded two flavonoids saponaretin **674** and saponarin **675**.³⁸⁷ A terpene glucoside (**676**) has been isolated from *Ornithogalum montanum* Cyr.³⁸⁸ and the main water soluble polysaccharide of the bulbs of *Ornithogalum ponticum* (Zahar.) Speta was found to be a neutral glucofructan with a glucose : fructose ratio of 30 : 1.³⁸⁹

A comparative study on the Bulgarian species *Ornithogalum nanum* Sibth. & Sm., *O. gussonei*, *Loncomelos narbonense* (L.) Raf. (as *O. narbonense*), and *O. montanum* Cyr. resulted in cardenolides (**677–681**) being found in the bulbs of *O. nanum* and *O. gussonei* but not in the bulbs of the other two species.³⁹⁰

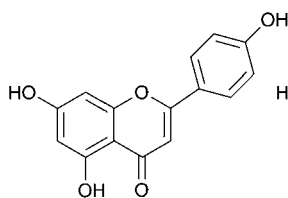
The leaves and bulbs of the Turkish species *Ornithogalum alpigenum* Stapf. have been tested for antioxidant activity, anti-microbial activity and as free radical scavengers. The leaf extracts were found to be good free radical scavengers (90.9%) and the bulb extracts (methanol, ethanol, acetone and benzene) showed variable activity against *Candida albicans*, *Bacillus subtilis* and *Bacillus cereus*. The highest antioxidant activity was found in the methanol extract of the bulbs (88.1% on a β -carotene-linoleic acid system) and the lowest in the benzene extract of the bulbs.³⁹¹ The Iranian species *Ornithogalum procerum* Stapf. and *Ornithogalum cuspidatum* Bertol. have received some attention. The aerial parts of *O. cuspidatum* are used in Iran as food additives as well as an anti-irritant and relaxant to soothe the throat when suffering from a dry cough. The essential oils in the leaves, flowers and bulbs have been determined by GC-MS. The flowers and bulbs contained primarily saturated hydrocarbons, while the leaves contained oxygenated hydrocarbons and terpenoid compounds.³⁹² In addition, a GS-MS analysis of the methanol extract of the bulbs has allowed for the identification of thirteen phytosterols (cholesterol, 3,5-didehydrostigmastan-6,22-diene, 4,4-dimethyl-5 α -cholest-7-en-3-one, 4 β -methylcholesterol, 5-cholestene-3 β ,7 β -diol, campesterol, cholest-4-ene-3,6-dione, stigmast-4-en-3-one, stigmasta-3,5-dien-7-one, stigmasterol, Δ^5 -ergosterol, β -sitosol and γ -sitosol).³⁹³ A GC-MS analysis of the aerial parts and bulbs of *O. procerum* resulted in the identification of twenty-three essential oils in the aerial parts and four polyesterol-type compounds in the bulbs.³⁹⁴ *Ornithogalum sintensii* Freyn, another Iranian species, has been tested for antioxidant activity. A higher phenolic and flavonoid content in the aerial parts of the plant relative to the bulbs, accounted for the greater antioxidant activity in those extracts (IC_{50} for DPPH radical scavenging activity of 368 and 669 $\mu\text{g ml}^{-1}$ for leaves and bulbs respectively).³⁹⁵

Eliokarmos thyrsoides (Jacq.) Raf. is found in southwestern South Africa. Although not reportedly used by traditional healers it has been strongly linked to stock poisoning.¹ Accordingly, the bulbs have been the subject of many phytochemical investigations. An initial investigation of the methanol extract of the bulbs in 1992 (as *Ornithogalum thyrsoides*) resulted in the isolation of four cholestane bisdesmosides (**682–685**), with **685** showing some inhibitory activity on cyclic AMP phosphodiesterase ($IC_{50} = 15.3 \times 10^{-5}$ M).³⁹⁶ Following the discovery that the methanol extract of the bulbs showed potent

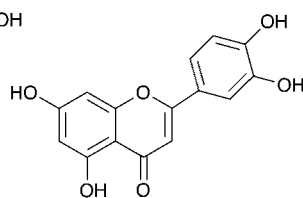




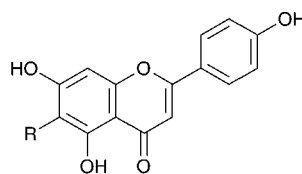
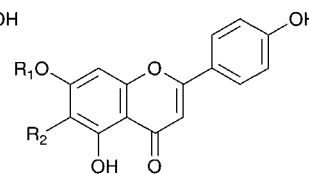
- 610 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 611 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- α -L-fucopyranoside, $R_2 = H$, $R_3 = HO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 612 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_2OH$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 613 $R_1 = 6$ -deoxy- β -D-gulopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 614 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 615 $R_1 = \alpha$ -L-rhamnopyranosyl- β -D-glucopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 616 $R_1 = 6$ -deoxy- β -D-allopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 617 $R_1 = \beta$ -D-allopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 618 $R_1 = 3$ -acetyl- β -digitopyranosyl- β -D-glucopyranosyl- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 619 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 620 $R_1 = 6$ -deoxy- β -D-allopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 621 $R_1 = 6$ -deoxy- β -D-gulopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 622 $R_1 = 6$ -deoxy- β -D-allopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 623 $R_1 = \beta$ -D-allopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 624 $R_1 = 6$ -deoxy- α -L-glucopyranose, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 625 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 626 $R_1 = \beta$ -D-ribose, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 631 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 632 $R_1 = \alpha$ -L-rhamnopyranosyl- β -D-glucopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 633 $R_1 = \alpha$ -L-arabinopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 642 $R_1 = 6'$ -deoxy- β -D-allopyranosyl-4'- β -D-xylopyranosyl-3''- β -D-apiofuranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 643 $R_1 = \alpha$ -L-rhamnopyranosyl-4'- β -D-apiofuranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = OH$
 644 $R_1 = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-3''- β -D-apiofuranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = OAc$
 645 $R_1 = 2'$ -desoxy- β -D-allopyranosyl-4'- β -D-xylopyranosyl-3''- β -D-apiofuranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = OAc$
 646 $R_1 = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-3''- β -D-apiofuranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = OAc$
 653 $R_1 = 6$ -desoxy- α -L-glucopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 654 $R_1 = \alpha$ -L-rhamnopyranosyl- β -D-glucopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 655 $R_1 = \beta$ -D-allopyranosyl- β -D-xyloside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 656 $R_1 = \beta$ -D-digitoxopyranosyl-4'-glucopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = OH$, $R_5 = H$, $R_6 = H$, $R_7 = H$
 657 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 658 $R_1 = 6$ -desoxy- α -L-glucopyranoside, $R_2 = H$, $R_3 = CHO$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 659 $R_1 = \alpha$ -L-rhamnopyranosyl-4'- β -D-apiofuranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 660 $R_1 = \alpha$ -L-rhamnopyranosyl-4'- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 661 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = H$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 662 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 663 $R_1 = \beta$ -D-glucopyranoside, $R_2 = H$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$
 664 $R_1 = 2$ -desoxy- β -D-allopyranoside, $R_2 = OH$, $R_3 = CH_3$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = H$

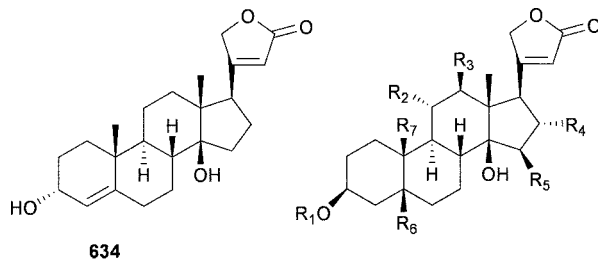


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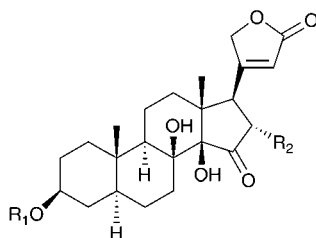
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629 $R = \beta$ -D-glucopyranoside630 $R_1 = \beta$ -D-glucopyranoside, $R_2 = \beta$ -D-glucopyranosyl- β -D-glucopyranoside

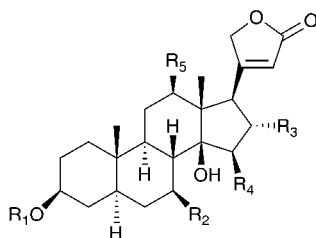


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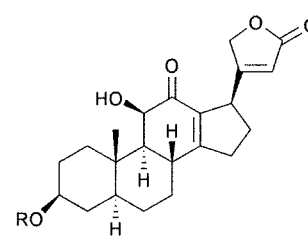
- 635** $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = \beta$ -OH, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
665 $R_1 = H$, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
666 $R_1 = 6'$ -deoxy- β -D-allopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
667 $R_1 = 6'$ -deoxy- β -D-allopyranosyl- β -D-xylopyranosyl- β -D-apiofuranoside, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
668 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = OH$, $R_5 = OH$, $R_6 = H$, $R_7 = CH_3$
669 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- β -D-apiofuranoside, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
670 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = OH$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
671 $R_1 = \alpha$ -L-rhamnopyranosyl- β -D-apiofuranoside, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
672 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
673 $R_1 = 6'$ -deoxy- β -D-allopyranosyl- β -D-xylopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = H$, $R_7 = CH_3$
677 $R_1 = H$, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = CH_3$
678 $R_1 = H$, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = CHO$
679 $R_1 = H$, $R_2 = OH$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = CH_3$
680 $R_1 = H$, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = CH_2OH$
681 $R_1 = 6$ -deoxy- β -D-allopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = H$, $R_6 = OH$, $R_7 = CH_3$



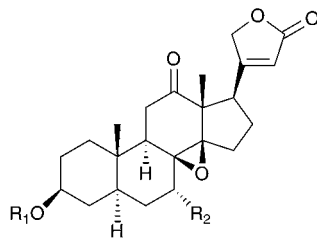
- 637** $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = OH$
638 $R_1 = 3'$ -O-acetyl- β -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = OH$
639 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- β -D-apiofuranoside, $R_2 = H$
640 $R_1 = 3'$ -O-acetyl- β -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$
641 $R_1 = \beta$ -D-digitoxopyranosyl- β -D-xylopyranosyl- α -L-rhamnopyranoside, $R_2 = H$



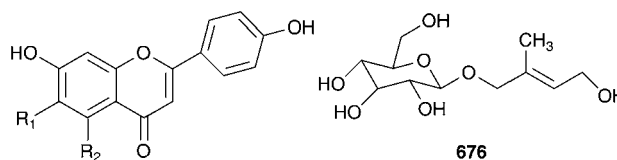
- 636** $R_1 = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside, $R_2 = OH$, $R_3 = OH$, $R_4 = OH$, $R_5 = H$
652 $R_1 = 3'$ -O-acetyl- β -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside, $R_2 = H$, $R_3 = H$, $R_4 = H$, $R_5 = OH$



- 647** $R = 3'$ -O-acetyl- β -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside
648 $R = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside
650 $R = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside



- 649** $R_1 = 3'$ -O-acetyl- β -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside, $R_2 = H$
651 $R_1 = \beta$ -D-digitoxopyranosyl-4'- β -D-xylopyranosyl-4''- α -L-rhamnopyranoside, $R_2 = OH$



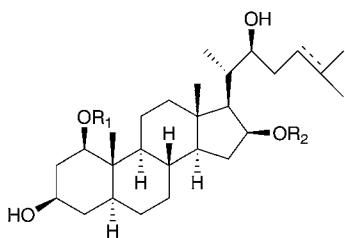
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- 674** $R_1 = \beta$ -D-glucopyranoside, $R_2 = OH$
675 $R_1 = OH$, $R_2 = \beta$ -D-glucopyranoside

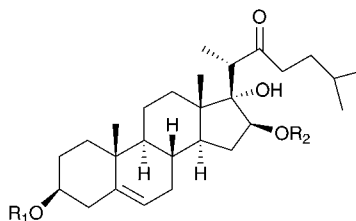


cytotoxicity against leukemia HL-60 cells ($IC_{50} = 0.79 \mu\text{g ml}^{-1}$), fractionation of this extract resulted in the isolation of twelve bisdesmosidic cholestane glycosides (577–578, 686–694),³⁹⁷ seven cholestane glycosides (695–701) and nine spirocyclic glycosides (702–710).^{398,399} The bisdesmosidic cholestane glycosides with an aromatic acyl group at the C-16 diglycoside were found to be the most cytotoxic (686 and 576) with IC_{50} values of 0.00016 and 0.00013 $\mu\text{g ml}^{-1}$ respectively. Compounds 687 and 696, the deacyl derivatives of compounds 688 and 689 and compounds 691 and 692 respectively did not show any activity at all.⁴⁰⁰ Only moderate activity against leukemia HL-60 cells was found for the spirocyclic glycosides (IC_{50} of 1.6–5.3 $\mu\text{g ml}^{-1}$).⁴⁰¹ An arabinoglucuronomannoglycan has been isolated from the leaves of *E. thyrsoides* (as *O. thyrsoides*).⁴⁰² South African bulbs of *Stellarioides longibracteata* (Jacq.) Speta are

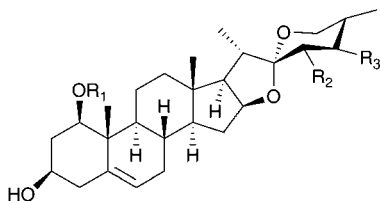
used extensively by the Zulu as an anti-inflammatory, and regionally to treat diabetes. A phytochemical investigation of the bulbs (as *Ornithogalum longibracteatum* Jacq.) revealed one major compound, the homoisoflavanone 711.⁴⁰³ The water extract of the bulbs produced a significant increase in glucose utilisation in Chang liver cells.⁴⁰⁴ *Stellarioides longibracteata* was introduced to China many years ago and it is used in Chinese folk medicine to treat cancer, as an antimicrobial, anti-inflammatory and to treat hepatitis and parotitis.⁴⁰⁵ This species was known earlier as *Ornithogalum caudatum* Ait. and as such many papers and in particular patents, have been filed under that name. Spirosterols, caudaside A 712 and hecogenin 713 have been isolated as well as β -sitosterol⁴⁰⁶ and three homoisoflavanones (714–716).⁴⁰⁷ Compounds 714–716 were tested for anti-tumour activity against P388 (mouse leukemia) and A-549



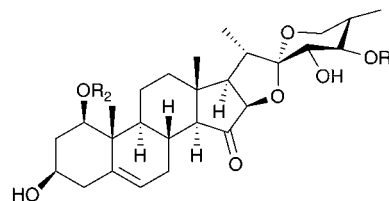
- 682 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \beta$ -D-glucopyranoside
 683 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = 6$ -O-acetyl- β -D-glucopyranoside
 684 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \beta$ -D-glucopyranoside, Δ^{24}
 685 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = 6$ -O-acetyl- β -D-glucopyranoside, Δ^{24}
 695 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \beta$ -apiofuranosyl-6-O-acetyl- β -D-glucopyranoside
 696 $R_1 = \beta$ -D-glucopyranoside, $R_2 = \beta$ -D-glucopyranoside
 697 $R_1 = 6$ -O-3,4,5-trimethoxybenzoyl- β -D-glucopyranoside, $R_2 = \beta$ -D-glucopyranoside
 698 $R_1 = 6$ -O-3,4,5-trimethoxybenzoyl- β -D-glucopyranoside, $R_2 = \beta$ -D-glucopyranoside, Δ^{24}
 699 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \alpha$ -L-rhamnopyranoside, Δ^5
 700 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \alpha$ -L-rhamnopyranoside, Δ^{24}
 701 $R_1 = \alpha$ -L-rhamnopyranoside, $R_2 = \alpha$ -L-rhamnopyranoside, Δ^5, Δ^{24}

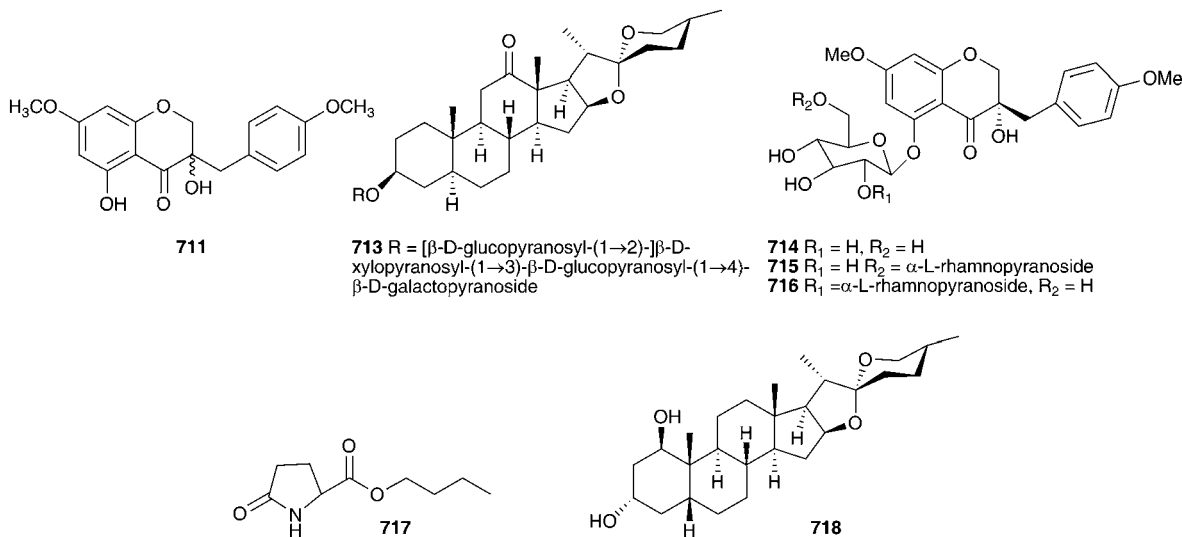


- 686 $R_1 = \beta$ -D-glucopyranoside, $R_2 = 2$ -O-3,4,5-trimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 687 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = \beta$ -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 688 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 2$ -O-3,4-dimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 689 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 2$ -O-3,4,5-trimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 690 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = \beta$ -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 691 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 2$ -O-3,4-dimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 692 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 2$ -O-3,4,5-trimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 693 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 2$ -O-4-hydroxy-3-methoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside
 694 $R_1 = \beta$ -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside, $R_2 = 3$ -O-3,4-dimethoxybenzoyl- β -D-xylopyranosyl-2-O-acetyl-(1 \rightarrow 3)- α -L-arabinopyranoside



- 702 $R_1 = \beta$ -D-glucopyranoside, $R_2 = \text{H}$, $R_3 = \text{H}$
 703 $R_1 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside, $R_2 = \text{H}$, $R_3 = \text{H}$
 704 $R_1 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)-[β -D-xylopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside], $R_2 = \text{H}$, $R_3 = \text{H}$
 705 $R_1 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside, $R_2 = \text{H}$, $R_3 = \text{OH}$
 706 $R_1 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)-[β -D-xylopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside], $R_2 = \text{H}$, $R_3 = \text{OH}$
 712 $R_1 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)-[β -D-xylopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside], $R_2 = \text{OH}$, $R_3 = \text{OH}$
- 707 $R_1 = \text{H}$, $R_2 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside
 708 $R_1 = 6$ -deoxy- β -D-gulopyranoside, $R_2 = \alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside
 709 $R_1 = 6$ -deoxy- β -D-gulopyranoside, $R_2 = [\alpha$ -L-rhamnopyranosyl-(1 \rightarrow 2)-O-[β -D-xylopyranosyl-(1 \rightarrow 3)]]- α -L-arabinopyranoside
 710 $R_1 = 6$ -deoxy- β -D-gulopyranoside, $R_2 = [2,3,4$ -tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 2)-O-[β -D-xylopyranosyl-(1 \rightarrow 3)]]- α -L-arabinopyranoside





(human pulmonary adenocarcinoma) but no activities were noted. Later investigations of the bulbs of *S. longibracteata* (as *O. caudatum*) resulted in the isolation of n-butyl pyroglutamate 717, 26 additional known flavonoids, sterols and acids⁴⁰⁸ and a monoterpene lactone (6-hydroxy-4,4,7 α -trimethyl-5,6,7 α -tetrahydro-4H-benzofuran-2-one).⁴⁰⁹ A number of patents have been filed in China and Russia outlining methods for the extraction of *S. longibracteata* (as *O. caudatum*) and use as a treatment for cancer,^{410,411} cholecystitis and cholecystitis-related diseases⁴¹² and for the isolation of the flavonoid components for use as an anti-inflammatory and analgesic.^{413,414} Extracts from *S. longibracteata* (as *O. caudatum*) have shown some benefit in the treatment of liver conditions.^{415,416} Four water soluble polysaccharide fractions have been extracted from *S. longibracteata* (as *O. caudatum*) and have been found to have potential as antitumour agents.^{417–419} Further work on the anti-cancer effects of this species has shown inhibition of MDA-MB-231 breast cancer cells in a dose and time dependant manner.⁴²⁰ The bulbs of *Stellarioides tenuifolia* (Redoute) Speta (as *Ornithogalum saundersiae*) yielded a crystalline steroidal saponin, 718, with an unusual 5 β configuration.⁴²¹ This unusual configuration at C-5, together with the 1 β ,3 α hydroxyl group configuration, results in a molecule with a shape that is able to undergo self-recognition, similar to single-strand DNA, and thus form a supramolecular zipper.

A review of steroidal glycosides isolated from various monocot families (including *Galtonia saundersiae*, as *Ornithogalum saundersiae*) and their biological activities has been published as well as a review of cholestane glycosides from *G. saundersiae*, *G. candicans* and *Eliokarmos thyrsoides* (as *Ornithogalum thyrsoides*).^{422,423}

4 The Oziroëoideae

The genus *Oziroë* Raf. (*syn. Fortunatia* Macbr.), is widespread in South America and can be found in Peru, Chile, Bolivia, Paraguay and north to central Argentina.⁴²⁴ No phytochemical investigations have been recorded on these plant species.

5 Conclusions

Research into the chemistry of the Hyacinthaceae has spanned over 90 years and resulted in the isolation of over 700 compounds. The need to understand the chemistry was largely fuelled by both the toxicity of the plants to stock and the significant ethnomedicinal applications associated with representative species. In addition to the isolation of many compounds of medicinal significance, the phytochemical studies have shed light on the classification of many of the species. A thorough understanding of the chemistry and pharmacology of these important plant species allows for the potential discovery of new medical applications and further aids the conservation of those taxa threatened with unsustainable utilisation by indicating alternatives with similarly active constituents.

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7 References

- 1 T. S. Pohl, N. R. Crouch and D. A. Mulholland, *Curr. Org. Chem.*, 2000, **4**, 1287–1324.
- 2 Angiosperm Phylogeny Group, *Bot. J. Linn. Soc.*, 2003, **141**, 399–436.
- 3 Angiosperm Phylogeny Group, *Bot. J. Linn. Soc.*, 2009, **161**, 105–121.
- 4 M. Pfosser and F. Speta, *Ann. Mo. Bot. Gard.*, 1999, **86**, 852–875.



- 5 F. Speta, *The Families and Genera of Vascular Plants*, Springer, Berlin, 1998, vol. 3, pp. 261–285.
- 6 M. Martínez-Azorín, B. Crespo Manuel, A. Juan and M. F. Fay, *Ann. Bot.*, 2011, **107**, 1–37.
- 7 S. Ali Syed, Y. Yu, M. Pfosser and W. Wetschnig, *Ann. Bot.*, 2012, **109**, 95–107.
- 8 M. W. Chase, J. L. Reveal and M. F. Fay, *Bot. J. Linn. Soc.*, 2009, **161**, 132–136.
- 9 B. M. Abegaz, J. Mutanyatta and M. Nindi, *Nat. Prod. Commun.*, 2007, **2**, 475–498.
- 10 K. du Toit, S. E. Drewes and J. Bodenstern, *Nat. Prod. Res.*, 2010, **24**, 457–490.
- 11 W. Heller and C. H. Tamm, *Fortschr. Chem. Org. Naturst.*, 1979, **40**, 106–136.
- 12 P. Boehler and C. H. Tamm, *Tetrahedron Lett.*, 1967, **8**, 3479–3483.
- 13 H. P. Weber, *Helv. Chim. Acta*, 1977, **60**, 1288–1392.
- 14 C. H. Tamm, *Arzneim. Forsch.*, 1972, **22**, 1776–1784.
- 15 W. Heller, P. Andermatt, W. A. Schaad and C. H. Tamm, *Helv. Chim. Acta*, 1976, **59**, 2048–2058.
- 16 Y. Mimaki, K. Ori, S. Kubo, Y. Sashida, T. Nikaido, L. G. Song and T. Ohmoto, *Chem. Lett.*, 1992, 1863–1866.
- 17 Y. Mimaki, H. Nishino, K. Ori, M. Kuroda, T. Matsui and Y. Sashida, *Chem. Pharm. Bull.*, 1994, **42**, 327–332.
- 18 J. K. Sihra, PhD Thesis, University of Surrey, 2012.
- 19 S. Koduru, D. S. Grierson and A. J. Afolayan, *Curr. Sci.*, 2007, **92**, 906–908.
- 20 R. Williams, N. R. Crouch and G. Pettit, *ISBA Bulletin*, 1996, **44**, 26–31.
- 21 W. T. L. Sidwell and C. H. Tamm, *Tetrahedron Lett.*, 1970, **11**, 475–478.
- 22 L. Farkas, A. Gottsegen, M. Nogradi and J. Strelisky, *Tetrahedron*, 1971, **27**, 5049–5056.
- 23 W. T. L. Sidwell, H. Fritz and C. H. Tamm, *Helv. Chim. Acta*, 1971, **54**, 207–215.
- 24 R. Ziegler and C. H. Tamm, *Helv. Chim. Acta*, 1976, **59**, 1997–2011.
- 25 D. A. Mulholland, C. Koorbanally, N. R. Crouch and P. Sandor, *J. Nat. Prod.*, 2004, **67**, 1726–1728.
- 26 W. T. L. Sidwell, C. H. Tamm, R. Ziegler, J. Finer and J. Clardy, *J. Am. Chem. Soc.*, 1975, **97**, 3518–3519.
- 27 J. L. S. Taylor and J. van Staden, *S. Afr. J. Bot.*, 2002, **68**, 80–85.
- 28 A. K. Jager, A. Hutchings and J. van Staden, *J. Ethnopharmacol.*, 1996, **52**, 95–100.
- 29 J. L. S. Taylor and J. van Staden, *J. Ethnopharmacol.*, 2001, **75**, 257–265.
- 30 A. K. Jager and J. van Staden, *Phytochem. Rev.*, 2005, **4**, 39–46.
- 31 M. Gaidamashvili and J. van Staden, *S. Afr. J. Bot.*, 2006, **72**, 661–663.
- 32 M. Gaidamashvili and J. van Staden, *J. Ethnopharmacol.*, 2002, **80**, 131–135.
- 33 A. R. Ndhllala, R. B. Mulaudzi, M. G. Kulkarni and J. Van Staden, *S. Afr. J. Bot.*, 2012, **79**, 1–8.
- 34 J. L. S. Taylor and J. van Staden, *S. Afr. J. Bot.*, 2002, **68**, 157–162.
- 35 J. L. S. Taylor and J. Van Staden, *Plant Growth Regul.*, 2001, **34**, 39–47.
- 36 R. E. Finckh and C. H. Tamm, *Experientia*, 1970, **26**, 472–473.
- 37 W. Heller and C. H. Tamm, *Helv. Chim. Acta*, 1974, **57**, 1766–1784.
- 38 W. Heller and C. H. Tamm, *Helv. Chim. Acta*, 1978, **61**, 1257–1261.
- 39 N. A. Koorbanally, N. R. Crouch, A. Langois, K. Du Toit, D. A. Mulholland and S. E. Drewes, *S. Afr. J. Bot.*, 2006, **72**, 428–433.
- 40 N. A. Koorbanally, N. R. Crouch, A. Harilal, B. Pillay and D. A. Mulholland, *Biochem. Syst. Ecol.*, 2006, **34**, 114–118.
- 41 K. Du Toit, E. E. Elgorashi, S. F. Malan, D. A. Mulholland, S. E. Drewes and J. Van Staden, *S. Afr. J. Bot.*, 2007, **73**, 236–241.
- 42 J. K. Sihra, PhD Thesis, University of Surrey, Guildford, UK, 2012.
- 43 I. Masterova, V. Suchy, D. Uhrin, K. Ubik, Z. Grancaiova and B. Bobovnický, *Phytochemistry*, 1991, **30**, 713–714.
- 44 E. Miadokova, I. Masterova, V. Vlckova, V. Duhova and J. Toth, *J. Ethnopharmacol.*, 2002, **81**, 381–386.
- 45 I. Juránek, V. Suchy, D. Stara, I. Masterova and Z. Grancaiova, *Die Pharmazie*, 1993, **48**, 310–311.
- 46 M. Urbancikova, I. Masterova and J. Toth, *Fitoterapia*, 2002, **73**, 724–726.
- 47 M. Ellnain-Wojtaszek, Z. Kowalewski and L. Skrzypczakowa, *Ann. Pharm. (Poznan)*, 1978, **13**, 171–175.
- 48 M. Adinolfi, M. M. Corsaro, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *Phytochemistry*, 1986, **26**, 285–290.
- 49 Z. Grancaiova, I. Materova and V. Suchy, *Fitoterapia*, 1992, **63**, 380.
- 50 M. Adinolfi, G. Barone, M. M. Corsaro, L. Mangoni, R. Lanzetta and M. Parrilli, *Can. J. Chem.*, 1988, **66**, 2787–2793.
- 51 N. Asano, H. Kuroi, K. Ikeda, H. Kizu, Y. Kameda, A. Kato, I. Adachi, A. A. Watson, R. J. Nash and G. W. J. Fleet, *Tetrahedron: Asymmetry*, 2000, **11**, 1–8.
- 52 F. J. Arias, P. Antolin, C. de Torre, B. Barriuso, R. Iglesias, M. A. Rojo, J. Miguel Ferreras, E. Benvenuto, E. Mendez and T. Girbes, *Int. J. Biochem. Cell Biol.*, 2003, **35**, 61–78.
- 53 K. Yoshida, H. Aoki, K. Kameda and T. Kondo, *ITE Lett. Batteries, New Technol. Med.*, 2002, **3**, 35–38.
- 54 K. Ori, M. Koroda, Y. Mimaki, H. Sakagami and Y. Sashida, *Phytochemistry*, 2003, **64**, 1351–1359.
- 55 M. Kuroda, Y. Mimaki, K. Ori, H. Sakagami and Y. Sashida, *J. Nat. Prod.*, 2004, **67**, 2099–2103.
- 56 V. V. Barbakadze, I. L. Targamadze and A. I. Usov, *Bioorg. Khim.*, 1996, **22**, 441–445.
- 57 V. Barbakadze and A. Usov, *Bull. Georgian Natl. Acad. Sci.*, 2001, **163**, 484–487.
- 58 G. Barone, M. M. Corsaro, R. Lanzetta and M. Parrilli, *Phytochemistry*, 1988, **27**, 921–923.
- 59 O. Goktas, E. Ozen, E. Baysal, R. Mammadov and M. H. Alma, *Wood Res. (Bratislava, Slovakia)*, 2010, **55**, 53–62.



- 60 A. Kato, N. Kato, I. Adachi, J. Hollinshead, G. W. J. Fleet, C. Kuriyama, K. Ikeda, N. Asano and R. J. Nash, *J. Nat. Prod.*, 2007, **70**, 993–997.
- 61 C. P. Waller, PhD Thesis, University of Surrey, Guildford, UK, 2012.
- 62 J. W. C. Gunn, M. Goldberg and J. H. Ferguson, *Trans. R. Soc. S. Afr.*, 1924, **12**, 1–3.
- 63 D. A. Mulholland, N. R. Crouch, C. Koorbanally, N. Moodley and T. Pohl, *Biochem. Syst. Ecol.*, 2006, **34**, 251–255.
- 64 T. Pohl, PhD Thesis, University of Natal, South Africa, 2002.
- 65 T. Pohl, C. Koorbanally, N. R. Crouch and D. A. Mulholland, *Biochem. Syst. Ecol.*, 2001, **29**, 857–860.
- 66 S. G. Sparg, J. van Staden and A. K. Jager, *J. Ethnopharmacol.*, 2002, **80**, 95–101.
- 67 L. V. Buwa and J. van Staden, *J. Ethnopharmacol.*, 2006, **103**, 139–142.
- 68 C. P. Waller, A. E. Thumser, M. K. Langat, N. R. Crouch and D. A. Mulholland, *Phytochemistry*, 2013, in press.
- 69 C. Koorbanally, D. A. Mulholland and N. R. Crouch, *Biochem. Syst. Ecol.*, 2006, **34**, 588–592.
- 70 J. Mutanyatta, G. Matapa Bhagali, D. D. Shushu and M. B. Abegaz, *Phytochemistry*, 2003, **62**, 797–804.
- 71 D. D. Shushu, J. Mutanyatta and B. M. Abegaz, *J. Herbs, Spices Med. Plants*, 2005, **11**, 97–107.
- 72 B. M. Abegaz, *Phytochem. Rev.*, 2002, **1**, 299–310.
- 73 N. Moodley, N. R. Crouch, D. A. Mulholland, D. Slade and D. Ferreira, *S. Afr. J. Bot.*, 2006, **72**, 517–520.
- 74 I. Calvo Maria, *Fitoterapia*, 2009, **80**, 394–398.
- 75 I. Calvo Maria, *Fitoterapia*, 2009, **80**, 96–101.
- 76 M. Parrilli, R. Lanzetta, V. Dovinola, M. Adinolfi and L. Mangoni, *Can. J. Chem.*, 1981, **59**, 2261–2265.
- 77 J. Lietava, *J. Ethnopharmacol.*, 1992, **35**, 263–266.
- 78 A. Pieroni, V. Janiak, C. M. Durr, S. Ludeke, E. Trachsel and M. Heinrich, *Phytother. Res.*, 2002, **16**, 467–473.
- 79 M. R. Loizzo, R. Tundis, F. Menichini, A. Pugliese, M. Bonesi, U. Solimene and F. Menichini, *Int. J. Food Sci. Nutr.*, 2010, **61**, 780–791.
- 80 R. M. Mamedov and O. M. Gektash, *Khim. Rastit. Syr'ya*, 2011, 141–144.
- 81 M. Parrilli, M. Adinolfi, V. Dovinola and L. Mangoni, *Gazz. Chim. Ital.*, 1979, **109**, 391–393.
- 82 M. Parrilli, M. Adinolfi, V. Dovinola and L. Mangoni, *Rend. Accad. Sci. Fis. Mat., Naples*, 1979, **45**, 527–532.
- 83 M. Parrilli, M. Adinolfi and L. Mangoni, *Gazz. Chim. Ital.*, 1979, **109**, 611–613.
- 84 M. Parrilli, R. Lanzetta, M. Adinolfi and L. Mangoni, *Tetrahedron*, 1980, **36**, 3591–3596.
- 85 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, M. Parrilli and L. Mangoni, *Rend. Accad. Sci. Fis. Mat., Naples*, 1983, **50**, 295–296.
- 86 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *J. Nat. Prod.*, 1983, **46**, 559–562.
- 87 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *J. Nat. Prod.*, 1984, **47**, 721–723.
- 88 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *J. Nat. Prod.*, 1984, **47**, 544–546.
- 89 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *J. Nat. Prod.*, 1984, **47**, 100–105.
- 90 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, M. Parrilli and L. Mangoni, *Rend. Accad. Sci. Fis. Mat., Naples*, 1983, **50**, 293–294.
- 91 M. Adinolfi, G. Barone, M. M. Corsaro, R. Lanzetta, L. Mangoni and M. Parrilli, *Can. J. Chem.*, 1987, **65**, 2317–2326.
- 92 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *Can. J. Chem.*, 1983, **61**, 2633–2637.
- 93 M. Adinolfi, G. Barone, R. Lanzetta, G. Laonigro, L. Mangoni and M. Parrilli, *Can. J. Chem.*, 1984, **62**, 1223–1226.
- 94 R. Lanzetta, G. Laonigro, M. Parrilli and E. Breitmaier, *Can. J. Chem.*, 1984, **62**, 2874–2878.
- 95 G. Barone, M. Belardini, R. Lanzetta, G. Laonigro and M. Parrilli, *Rend. Accad. Sci. Fis. Mat., Naples*, 1983, **50**, 297–298.
- 96 M. Adinolfi, G. Barone, M. Belardini, R. Lanzetta, G. Laonigro and M. Parrilli, *Phytochemistry*, 1984, **23**, 2091–2093.
- 97 M. Adinolfi, G. Barone, M. Belardini, R. Lanzetta, G. Laonigro and M. Parrilli, *Phytochemistry*, 1985, **24**, 2423–2426.
- 98 M. Adinolfi, G. Barone, F. Giordano, R. Lanzetta and M. Parrilli, *Tetrahedron*, 1990, **46**, 6565–6574.
- 99 M. Adinolfi, G. Barone, M. M. Corsaro, L. Mangoni, R. Lanzetta and M. Parrilli, *Tetrahedron*, 1988, **44**, 4981–4988.
- 100 R. Della Loggia, P. Del Negro, A. Tubaro, G. Barone and M. Parrilli, *Planta Med.*, 1989, **55**, 587–588.
- 101 S. H. Hilal, M. M. Shabana and A. S. Ibrahim, *Egypt. J. Pharm. Sci.*, 1985, **24**, 67–74.
- 102 M. Guarrera Paolo, G. Salerno and G. Caneva, *J. Ethnopharmacol.*, 2005, **99**, 367–378.
- 103 C. L. Quave, L. R. W. Plano, T. Pantuso and B. C. Bennett, *J. Ethnopharmacol.*, 2008, **118**, 418–428.
- 104 Y. Mimaki, S. Kubo, Y. Kinoshita, Y. Sashida, L. G. Song, T. Nikaido and T. Ohmoto, *Phytochemistry*, 1993, **34**, 791–797.
- 105 G. Barone, M. M. Corsaro, R. Lanzetta, L. Mangoni and M. Parrilli, *Phytochemistry*, 1993, **33**, 431–436.
- 106 K. Ori, M. Kuroda, Y. Mimaki, H. Sakagami and Y. Sashida, *Chem. Pharm. Bull.*, 2003, **51**, 92–95.
- 107 M. Adinolfi, M. M. Corsaro, R. Lanzetta, A. Mancino, L. Mangoni and M. Parrilli, *Phytochemistry*, 1993, **34**, 773–778.
- 108 M. Kuroda, Y. Mimaki, K. Ori, H. Koshino, T. Nukada, H. Sakagami and Y. Sashida, *Tetrahedron*, 2002, **58**, 6735–6740.
- 109 M. M. Corsaro, R. Lanzetta, A. Mancino and M. Parrilli, *Phytochemistry*, 1992, **31**, 1395–1397.
- 110 P. Baskaran, B. Ncube and J. van Staden, *Plant Growth Regul.*, 2012, **67**, 235–245.
- 111 C. Dias, J. A. Borralho Graca and M. Lurdes Goncalves, *J. Ethnopharmacol.*, 2000, **71**, 487–492.



- 112 S.-M. Lee, H.-K. Chun, C.-H. Lee, B.-S. Min, E.-S. Lee and Y.-H. Kho, *Chem. Pharm. Bull.*, 2002, **50**, 1245–1249.
- 113 E.-J. Yeo, K.-T. Kim, Y. S. Han, S.-Y. Nah and H.-D. Paik, *Food Sci. Biotechnol.*, 2006, **15**, 639–642.
- 114 I. Kouno, N. Noda, Y. Ida, M. Sholichin, K. Miyahara, T. Komori and T. Kawasaki, *Liebigs Ann. Chem.*, 1982, 306–314.
- 115 M. Sholichin, K. Miyahara and T. Kawasaki, *Heterocycles*, 1982, **17**, 251–257.
- 116 M. Sholichin, K. Miyahara and T. Kawasaki, *Chem. Pharm. Bull.*, 1985, **33**, 1756–1759.
- 117 S.-M. Lee, H.-J. Lee, H.-K. Chun, C.-H. Lee, S.-J. Kang, H.-Y. Maeng and Y.-H. Kho, *Saengyak Hakhoechi*, 2001, **32**, 189–192.
- 118 M. Ono, D. Toyohisa, T. Morishita, H. Horita, S. Yasuda, Y. Nishida, T. Tanaka, M. Okawa, J. Kinjo, H. Yoshimitsu and T. Nohara, *Chem. Pharm. Bull.*, 2011, **59**, 1348–1354.
- 119 I. Kouno, T. Komori and T. Kawasaki, *Tetrahedron Lett.*, 1973, **14**, 4569–4572.
- 120 Y. Nishida, M. Eto, H. Miyashita, T. Ikeda, K. Yamaguchi, H. Yoshimitsu, T. Nohara and M. Ono, *Chem. Pharm. Bull.*, 2008, **56**, 1022–1025.
- 121 H. B. Lee and S. M. Lee, *KR Pat.*, 2010-35527, 2010.
- 122 H. G. Jun, Y. H. Ko, C. H. Lee, H. J. Lee and S. M. Lee, *KR Pat.*, 2001-1197, 2001.
- 123 M. T. Ono, Y. Takatsu, T. Ochiai, S. Yasuda, Y. Nishida, T. Tanaka, M. Okawa, J. Kinjo, H. Yoshimitsu and T. Nohara, *Chem. Pharm. Bull.*, 2012, **60**, 1314–1319.
- 124 T. Kurihara and H. Ito, *Annu. Rep. Tohoku Coll. Pharm.*, 1968, 39–47.
- 125 A. I. Ismailov, *Azerb. Med. Zh.*, 1961, **1**, 57–60.
- 126 V. V. Barbakadze, I. L. Targamadze and A. I. Usov, *Bioorg. Khim.*, 1996, **22**, 58–61.
- 127 T. Yamashita, K. Yasuda, H. Kizu, Y. Kameda, A. Watson, R. J. Nash, G. W. J. Fleet and N. Asano, *J. Nat. Prod.*, 2002, **65**, 1875–1881.
- 128 Y. Mimaki, K. Ori, Y. Sashida, T. Nikaido, L. G. Song and T. Ohmoto, *Bull. Chem. Soc. Jpn.*, 1993, **66**, 1182–1186.
- 129 Y. Mimaki, K. Ori, Y. Sashida, T. Nikaido, L. G. Song and T. Ohmoto, *Chem. Lett.*, 1992, 1999–2000.
- 130 N. Asano, K. Ikeda, M. Kasahara, Y. Arai and H. Kizu, *J. Nat. Prod.*, 2004, **67**, 846–850.
- 131 K. Hosokawa, Y. Fukunaga, E. Fukushi and J. Kawabata, *Phytochemistry*, 1995, **38**, 1293–1298.
- 132 K. Hosokawa, Y. Fukunaga, E. Fukushi and J. Kawabata, *Phytochemistry*, 1996, **41**, 1531–1533.
- 133 K. Hosokawa, Y. Fukunaga, E. Fukushi and J. Kawabata, *Phytochemistry*, 1995, **40**, 567–571.
- 134 K. Hosokawa, Y. Fukunaga, E. Fukushi and J. Kawabata, *Phytochemistry*, 1995, **39**, 1437–1441.
- 135 K. Hosokawa, Y. Fukunaga, E. Fukushi and J. Kawabata, *Phytochemistry*, 1996, **42**, 671–672.
- 136 K. Hosokawa, *Recent Res. Dev. Phytochem.*, 1998, **2**, 135–141.
- 137 K. Hosokawa and Y. Fukunaga, *Plant Cell Rep.*, 1995, **14**, 575–579.
- 138 G. C. Kite, C. Sellwood, P. Wilkin and M. S. J. Simmonds, *Biochem. Syst. Ecol.*, 1998, **26**, 357–359.
- 139 N. Asano, A. Kato, M. Miyauchi, H. Kizu, Y. Kameda, A. A. Watson, R. J. Nash and G. W. J. Fleet, *J. Nat. Prod.*, 1998, **61**, 625–628.
- 140 M. Adinolfi, G. Barone, L. Dellagrecia, L. Mangoni, R. Lanzetta and M. Parrilli, *Gazz. Chim. Ital.*, 1990, **120**, 427–433.
- 141 M. Adinolfi, T. Aquila, G. Barone, R. Lanzetta and M. Parrilli, *Phytochemistry*, 1989, **28**, 3244–3246.
- 142 C. Dias, M. Dias, C. Borges, M. A. Almoester Ferreira, A. Paulo and J. Nascimento, *J. Mass Spectrom.*, 2003, **38**, 1240–1244.
- 143 A. Paulo, C. Dias, M. L. Jimeno, C. Borges and J. Nascimento, *Fitoterapia*, 2005, **76**, 765–767.
- 144 C. Dias, A. Paulo, J. Nascimento, P. Houghton, J. E. Hawkes and M. L. Goncalves, *Planta Med.*, 2003, **69**, 1060–1062.
- 145 S. D. Wood, A. K. Allen, L. M. Wright and C. D. Reynolds, *Lectins: Biol., Biochem., Clin. Biochem.*, 1997, **11**, 86–90.
- 146 S. D. Wood, L. M. Wright, C. D. Reynolds, P. J. Rizkallah, A. K. Allen, W. J. Peumans and E. J. M. Van Damme, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 1999, **55**, 1264–1272.
- 147 L. M. Wright, P. J. Rizkallah, S. D. Wood and C. D. Reynolds, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 1998, **54**, 665–667.
- 148 L. M. Wright, S. D. Wood, C. D. Reynolds, P. J. Rizkallah and A. K. Allen, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 1998, **54**, 90–92.
- 149 L. M. Wright, S. D. Wood, C. D. Reynolds, P. J. Rizkallah, W. J. Peumans, E. J. M. Van Damme and A. K. Allen, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 1996, **52**, 1021–1023.
- 150 L. M. Wright, C. D. Reynolds, P. J. Rizkallah, A. K. Allen, E. J. Van Damme, M. J. Donovan and W. J. Peumans, *FEBS Lett.*, 2000, **468**, 19–22.
- 151 A. Kato, I. Adachi, M. Miyauchi, K. Ikeda, T. Komae, H. Kizu, Y. Kameda, A. A. Watson, R. J. Nash, M. R. Wormald, G. W. J. Fleet and N. Asano, *Carbohydr. Res.*, 1999, **316**, 95–103.
- 152 A. A. Watson, R. J. Nash, M. R. Wormald, D. J. Harvey, S. Dealler, E. Lees, N. Asano, H. Kizu, A. Kato, R. C. Griffiths, A. J. Cairns and G. W. J. Fleet, *Phytochemistry*, 1997, **46**, 255–259.
- 153 J. L. Thompson and J. K. N. Jones, *Can. J. Chem.*, 1964, **42**, 1088–1091.
- 154 C. Koorbanally, N. R. Crouch and D. A. Mulholland, *Biochem. Syst. Ecol.*, 2001, **29**, 539–541.
- 155 D. Ngamga, J. Bipa, P. Lebatha, C. Hiza, J. Mutanyatta, M. Bezabih, P. Tane and B. M. Abegaz, *Nat. Prod. Commun.*, 2008, **3**, 769–777.
- 156 D. Ngamga, P. Tane, M. Bezabih and B. M. Abegaz, *Planta Med.*, 2006, **72**, 1009–1009.
- 157 F. Fusi, A. Ferrara, C. Koorbanally, N. R. Crouch, D. A. Mulholland and G. Sgaragli, *J. Pharm. Pharmacol.*, 2008, **60**, 489–497.
- 158 G. Amschler, A. W. Frahm, A. Hatzelmann, U. Kilian, D. Mueller-Doblies and U. Mueller-Doblies, *Planta Med.*, 1996, **62**, 534–539.
- 159 G. Amschler, A. W. Frahm, D. Muller-Doblies and U. Muller-Doblies, *Phytochemistry*, 1998, **47**, 429–436.



- 160 A. Michalik, J. Hollinshead, L. Jones, G. W. J. Fleet, C.-Y. Yu, X.-G. Hu, R. van Well, G. Horne, F. X. Wilson, A. Kato, S. F. Jenkinson and R. J. Nash, *Phytochem. Lett.*, 2010, **3**, 136–138.
- 161 N. Moodley, D. A. Mulholland and N. R. Crouch, *J. Nat. Prod.*, 2004, **67**, 918–920.
- 162 N. R. Crouch, V. Bangani and D. A. Mulholland, *Phytochemistry*, 1999, **51**, 943–946.
- 163 B. Ncube, J. F. Finnie and J. van Staden, *S. Afr. J. Bot.*, 2011, **77**, 387–396.
- 164 B. Ncube, J. F. Finnie and J. van Staden, *S. Afr. J. Bot.*, 2012, **78**, 246–251.
- 165 A. Langlois, D. A. Mulholland, G. D. Duncan, N. R. Crouch and T. J. Edwards, *Biochem. Syst. Ecol.*, 2005, **33**, 961–966.
- 166 C. A. Williams, J. B. Harborne and T. S. Crosby, *Phytochemistry*, 1976, **15**, 349–350.
- 167 C. Koorbanally, S. Sewjee, D. A. Mulholland, N. R. Crouch and A. Dold, *Phytochemistry*, 2007, **68**, 2753–2756.
- 168 V. Bangani, N. R. Crouch and D. A. Mulholland, *Phytochemistry*, 1999, **51**, 947–951.
- 169 K. du Toit, A. Kweyama and J. Bodenstein, *S. Afr. J. Sci.*, 2011, **107**, 96–100.
- 170 B. M. Abegaz, *Phytochem. Rev.*, 2002, **1**, 299–310.
- 171 A. Silayo, B. T. Ngadjui and B. M. Abegaz, *Phytochemistry*, 1999, **52**, 947–955.
- 172 S. O. Famuyiwa, K. F. Sichilongo, S. O. Yeboah and B. M. Abegaz, *Phytochem. Lett.*, 2012, **5**, 591–595.
- 173 P. Pillay, A. Phulukdaree, A. A. Chuturgoon, K. du Toit and J. Bodenstein, *J. Ethnopharmacol.*, 2012.
- 174 F. Speta, *Phyton (Horn, Austria)*, 1998, **38**, 1–141.
- 175 L. Krenn and B. Kopp, *Phytochemistry*, 1998, **48**, 1–29.
- 176 S. J. van der Walt, *Onderstepoort J. Vet. Res.*, 1944, **20**, 75–83.
- 177 N. Sapeika, *S. Afr. Pharm. J.*, 1951, **1**, 28–29.
- 178 D. G. Steyn, 15th Annual Report of the Director of Veterinary Services, 1929, pp. 777–803.
- 179 C. F. Juritz, *12th Annual Meeting of the South African Association for the Advancement of Science*, 1914, 109–150.
- 180 A. Hutchings and S. E. Terblanche, *S. Afr. Med. J.*, 1989, **75**, 41–96.
- 181 M. F. Pfosser and F. Speta, *Plant Syst. Evol.*, 2004, **246**, 245–263.
- 182 W. Kopaczewski, *C. R. Hebd. Seances Acad. Sci.*, 1914, **158**, 1520–1522.
- 183 W. Kopaczewski, *Biochem. Z.*, 1914, **66**, 501–508.
- 184 F. W. Smith, *Analyst*, 1921, **46**, 178–179.
- 185 O. Krestynova-Telupilova and F. Santavy, *Ceska Slov. Farm.*, 1957, **6**, 207–209.
- 186 K. J. Kraus, E. Mutschler and H. Rochelmeyer, *Arzneim. Forsch.*, 1969, **19**, 322–328.
- 187 K. J. Kraus, E. Mutschler and H. Rochelmeyer, *Pharm. Ztg.*, 1969, **114**, 417–419.
- 188 F. Kaczmarek and I. Zygmunt, *Herba Pol.*, 1974, **20**, 330–338.
- 189 E. Gliozheni and T. Bisha, *Bul. Shkencave Nat.*, 1972, **26**, 43–50.
- 190 C. H. Boehringer, *FR Pat.* 1967-123308, 1967.
- 191 G. Tittel and H. Wagner, *Planta Med.*, 1980, **39**, 125–134.
- 192 M. Couladi and A. Loukis, *Fitoterapia*, 1987, **58**, 57–58.
- 193 A. Aliaga, M. Fernandez, F. A. Vega and C. Dios, *Acta Pol. Pharm.*, 1987, **44**, 560–562.
- 194 M. Fernandez, M. Oses and C. Dios, *An. R. Acad. Farm.*, 1987, **53**, 292–295.
- 195 K. Okushima, *Arch. Exp. Pathol. Pharmacol.*, 1922, **95**, 258–266.
- 196 J. Markwalder, *Klin. Wochenschr.*, 1922, **1**, 212–215.
- 197 A. Stoll, E. Suter, W. Kreis, B. B. Bussemaker and A. Hofmann, *Helv. Chim. Acta*, 1933, **16**, 703–733.
- 198 W. Anders and G. Zeise, *Med. Klin.*, 1952, **47**, 1061–1063.
- 199 H. Haas, *Med. Klin.*, 1967, **62**, 121–126.
- 200 A. von Wartburg, *Helv. Chim. Acta*, 1966, **49**, 30–42.
- 201 A. von Wartburg and J. Renz, *Helv. Chim. Acta*, 1959.
- 202 A. J. Verbiscar, J. Patel, T. F. Banigan and R. A. Schatz, *J. Agric. Food Chem.*, 1986, **34**, 973–979.
- 203 H. Lichti, P. Niklaus and A. von Wartburg, *Helv. Chim. Acta*, 1973, **56**, 2083–2087.
- 204 B. Gorlich, *Arzneim. Forsch.*, 1960, **10**, 770–774.
- 205 A. von Wartburg, M. Kuhn and K. Huber, *Helv. Chim. Acta*, 1968, **51**, 1317–1328.
- 206 L. Krenn, R. Ferth, W. Robien and B. Kopp, *Planta Med.*, 1991, **57**, 560–565.
- 207 B. Kopp, L. Krenn, M. Draxler, A. Hoyer, R. Terkola, P. Vallaster and W. Robien, *Phytochemistry*, 1996, **42**, 513–522.
- 208 L. Krenn, B. Kopp, S. Steurer and M. Schubert-Zsilavec, *J. Nat. Prod.*, 1996, **59**, 612–613.
- 209 L. Krenn, M. Jelovina and B. Kopp, *Fitoterapia*, 2000, **71**, 126–129.
- 210 M. Iizuka, T. Warashina and T. Noro, *Chem. Pharm. Bull.*, 2001, **49**, 282–286.
- 211 L. Krenn, B. Kopp, A. Deim, W. Robien and W. Kubelka, *Planta Med.*, 1994, **60**, 63–69.
- 212 I. Garcia-Jalon, F. A. Vega, M. Fernandez and T. Arrupe, *Cienc. Ind. Farm.*, 1974, **6**, 43–52.
- 213 I. Garcia-Jalon, F. A. Vega, M. Fernandez and L. Martinez Valls, *Cienc. Ind. Farm.*, 1973, **5**, 260–269.
- 214 F. A. Vega, C. Martin and I. Garcia-Jalon, *An. R. Acad. Farm.*, 1969, **35**, 291–296.
- 215 F. A. Vega, I. Garcia-Jalon, M. Fernandez and J. Renedo, *Phytochemistry*, 1972, **11**, 2896.
- 216 F. A. Vega, C. Martin, M. Fernandez and I. Garcia-Jalon, *Galenica Acta*, 1970, **23**, 3–48; (43–44–45), 79–96.
- 217 F. A. Vega and M. Fernandez, *Naturwissenschaften*, 1964, **51**, 483–484.
- 218 M. Fernandez, F. A. Vega, T. Arrupe and J. Renedo, *Sci. Pharm.*, 1976, **44**, 307–314.
- 219 M. Fernandez, F. A. Vega, T. Arrupe and J. Renedo, *Phytochemistry*, 1972, **11**, 1534.
- 220 M. Couladis, E. Verykokidou-Vitsaropoulou and S. Philianos, *Fitoterapia*, 1993, **64**, 92.
- 221 F. A. Vega, *An. R. Acad. Farm.*, 1976, **42**, 81–94.
- 222 M. Fernandez, J. Renedo, T. Arrupe and F. A. Vega, *Phytochemistry*, 1975, **14**, 586–586.
- 223 M. Fernandez, J. Renedo, T. Arrupe and F. A. Vega, *Ciencia & Industria Farmaceutica*, 1974, **6**, 386–390.



- 224 M. Fernandez, F. A. Vega, T. Arrupe and J. Renedo, *Galenica Acta*, 1971, **24**, 45–57.
- 225 T. Spies, W. Praznik, A. Hofinger, F. Altmann, E. Nitsch and R. Wutka, *Carbohydr. Res.*, 1992, **235**, 221–230.
- 226 W. Praznik and T. Spies, *Carbohydr. Res.*, 1993, **243**, 91–97.
- 227 D. Fos, J. Giraldez, M. J. Renedo, M. Fernandez and F. A. Vega, *Cienc. Ind. Farm.*, 1979, **11**, 141–146.
- 228 M. J. Pascual-Villalobos and M. Fernandez, *Ind. Crops Prod.*, 1999, **10**, 115–120.
- 229 H. Bozcuk, M. Ozdogan, O. Aykurt, F. Topcuoglu, H. Ozturk, D. Ekinci, A. Karadeniz, A. Mutlu and D. Burgucu, *Turk. J. Med. Sci.*, 2011, **41**, 101–108.
- 230 S. E. Shyr and E. J. Staba, *Planta Med.*, 1976, **29**, 86–90.
- 231 L. Krenn, M. Jambrits and B. Kopp, *Planta Med.*, 1988, **54**, 227–232.
- 232 L. Krenn, A. Hufner, A. Kastenhuber and F. Speta, *Phytochemistry*, 2004, **65**, 2881–2884.
- 233 B. Kopp, M. Unterluggauer, W. Robien and W. Kubelka, *Planta Med.*, 1990, **56**, 193–197.
- 234 L. Krenn, M. Bamberger and B. Kopp, *Planta Med.*, 1992, **58**, 284–285.
- 235 L. Krenn, B. Kopp, E. Griesmayer-Camus and W. Kubelka, *Sci. Pharm.*, 1992, **60**, 65–72.
- 236 C. F. Juritz, *Chem. News J. Ind. Sci.*, 1923, **126**, 69–70.
- 237 N. Moodley, N. R. Crouch and D. A. Mulholland, *Phytochemistry*, 2007, **68**, 2415–2419.
- 238 J. M. Watt, *Arch. Exp. Pathol. Pharmacol.*, 1927, **120**, 65–76.
- 239 P. G. J. Louw, *Onderstepoort J. Vet. Res.*, 1953, **26**, 141–148.
- 240 R. Tschesche and K. H. Hottemann, *Chem. Ber.*, 1953, **86**, 392–395.
- 241 N. Sapeika, *S. Afr. J. Med. Sci.*, 1944, **9**, 31–32.
- 242 P. G. J. Louw, *Onderstepoort J. Vet. Res.*, 1949, **22**, 313–319.
- 243 P. G. J. Louw, *Nature*, 1949, **163**, 30–31.
- 244 N. Sapeika, *S. Afr. Med. J.*, 1950, **24**, 912–913.
- 245 B. Isaacson, *S. Afr. Med. J.*, 1950, **24**, 901–912.
- 246 P. G. J. Louw, *Onderstepoort J. Vet. Res.*, 1952, **25**, 123–133.
- 247 P. Zoller and C. H. Tamm, *Helv. Chim. Acta*, 1953, **36**, 1744–1756.
- 248 P. S. Steyn, F. R. van Heerden and R. Vleggaar, *S. Afr. J. Chem.*, 1986, **39**, 143–146.
- 249 L. Krenn, B. Kopp, M. Bamberger, E. Brustmann and W. Kubelka, *Nat. Prod. Lett.*, 1993, **3**, 139–143.
- 250 R. R. T. Majinda, R. D. Waigh and P. G. Waterman, *Planta Med.*, 1997, **63**, 188–190.
- 251 G. N. Foukaridis, E. Osuch, L. Mathibe and P. Tsipa, *J. Ethnopharmacol.*, 1995, **49**, 77–79.
- 252 J. Marx, E. Pretorius and M. J. Bester, *J. Ethnopharmacol.*, 2006, **104**, 315–321.
- 253 J. Marx, E. Pretorius, W. J. Espag and M. J. Bester, *Environ. Toxicol. Pharmacol.*, 2005, **20**, 26–34.
- 254 C. Koorbanally, D. A. Mulholland and N. R. Crouch, *Biochem. Syst. Ecol.*, 2005, **33**, 743–748.
- 255 M. Teshome, H. Kassa and K. Charles, *Ethiop. J. Health Dev.*, 2010, **24**, 175–179.
- 256 H. D. Neuwinger, *African Ethnobotany: Poisons and Drugs*, Chapman and Hall, London, 1994.
- 257 M. Miyakado, T. Kato, N. Ohno and K. Koshimizu, *Phytochemistry*, 1975, **14**, 2717.
- 258 T. Pohl, C. Koorbanally, N. R. Crouch and D. A. Mulholland, *Phytochemistry*, 2001, **58**, 557–561.
- 259 E. Dagne, W. Mammo, M. Alemu and I. Casser, *Bull. Chem. Soc. Ethiop.*, 1994, **8**, 85–89.
- 260 K. Shimada, E. Umezawa, T. Nambara and S. M. Kupchan, *Chem. Pharm. Bull.*, 1979, **27**, 3111–3114.
- 261 H. Lichti and A. von Wartburg, *Helv. Chim. Acta*, 1960, **43**, 1666–1681.
- 262 A. L. Cogne, A. Marston, S. Mavi and K. Hostettmann, *J. Ethnopharmacol.*, 2001, **75**, 51–53.
- 263 F. R. Van Heerden, R. Vleggaar and L. A. P. Anderson, *S. Afr. J. Chem.*, 1988, **41**, 145–151.
- 264 A. R. Ndhlala, J. F. Finnie and J. van Staden, *J. Ethnopharmacol.*, 2011, **133**, 663–674.
- 265 N. R. Crouch, A. Langlois and D. A. Mulholland, *Phytochemistry*, 2007, **68**, 1731–1734.
- 266 N. R. Crouch, K. du Toit, D. A. Mulholland and S. E. Drewes, *Phytochemistry*, 2006, **67**, 2140–2145.
- 267 C. Koorbanally, D. A. Mulholland and N. R. Crouch, *Biochem. Syst. Ecol.*, 2005, **33**, 295–299.
- 268 N. A. Koorbanally, C. Koorbanally, A. Harilal, D. A. Mulholland and N. R. Crouch, *Phytochemistry*, 2004, **65**, 3069–3073.
- 269 O. O. G. Amusan, J. D. Msonthi and L. P. Makhubu, *Fitoterapia*, 1997, **68**, 185–186.
- 270 S. Abbas, S. Bashir, A. Khan, H. Mehmood Malik and H. Gilani Anwarul, *Phytother. Res.*, 2012, **26**, 704–708.
- 271 Y. B. Tripathi, A. V. Singh and G. P. Dubey, *Current Science*, 2001, **80**, 1267–1269.
- 272 P. Shokeen, M. Bala and V. Tandon, *Int. J. Antimicrob. Agents*, 2009, **33**, 86–91.
- 273 H. N. Thatoi, S. K. Panda, S. K. Rath and S. K. Dutta, *Asian J. Plant Sci.*, 2008, **7**, 260–267.
- 274 S. Rangaswami and S. S. Subramanian, *J. Sci. Ind. Res.*, 1955, **14B**, 131.
- 275 S. Rangaswami and S. S. Subramanian, *J. Sci. Ind. Res.*, 1956, **15C**, 80–81.
- 276 R. V. Rao and S. Rangaswami, *Tetrahedron Lett.*, 1967, **8**, 4563–4565.
- 277 S. Jha and S. Sen, *Curr. Sci.*, 1980, **49**, 273–274.
- 278 S. Jha and S. Sen, *Planta Med.*, 1983, **47**, 43–45.
- 279 S. Rangaswami and S. S. Subramanian, *J. Sci. Ind. Res.*, 1954, **13B**, 150.
- 280 S. Jha and S. Sen, *Phytochemistry*, 1981, **20**, 524–526.
- 281 B. Kopp and M. Danner, *Sci. Pharm.*, 1983, **51**, 227–237.
- 282 D. B. Deb and S. Dasgupta, *Q. J. Crude Drug Res.*, 1976, **14**, 49–60.
- 283 S. Jha and S. Sen, *Nucleus (Calcutta)*, 1982, **25**, 190–192.
- 284 S. Jha and S. Sen, *Indian J. Exp. Biol.*, 1980, **18**, 291–292.
- 285 S. Jha and S. Sen, *Phytochemistry*, 1981, **20**, 1442–1443.
- 286 V. K. Saxena and P. K. Chaturvedi, *J. Inst. Chem. (India)*, 1992, **64**, 35–37.



- 287 N. Sultana, K. Akter, N. Nahar, M. S. H. Khan, M. Mosihuzzaman, H. Sohrab Md and K. Krohn, *Nat. Prod. Res.*, 2010, **24**, 1018–1026.
- 288 M. M. Patil and S. G. Torne, *Curr. Sci.*, 1980, **49**, 276–277.
- 289 S. Jha and S. Sen, *Acta Bot. Indica*, 1981, **9**, 258–264.
- 290 S. R. Shenoy, M. N. S. Kameshwari, S. Swaminathan and M. N. Gupta, *Biotechnol. Prog.*, 2006, **22**, 631–637.
- 291 A. V. Deepak, G. Thippeswamy, M. N. Shivakameshwari and P. Salimath Bharathi, *Biochem. Biophys. Res. Commun.*, 2003, **311**, 735–742.
- 292 A. V. Deepak and P. Salimath Bharathi, *Biochimie*, 2006, **88**, 297–307.
- 293 H. Mikail, P. Magiatis and A. Skaltsounis, *Planta Med.*, 2010, **76**, 1242–1242.
- 294 M. M. Abid Ali Khan, S. N. H. Zaidi, S. A. Musanna and N. Singh, *Asian J. Exp. Biol. Sci.*, 2012, **3**, 476–480.
- 295 R. P. Luyt, A. K. Jager and J. van Staden, *S. Afr. J. Bot.*, 1999, **65**, 443–445.
- 296 R. P. Luyt, A. K. Jager and J. van Staden, *S. Afr. J. Bot.*, 1999, **65**, 443–445.
- 297 L. Krenn, V. Stapf and B. Kopp, *Sci. Pharm.*, 2000, **68**, 421–427.
- 298 L. Rakotobe, M. Berkal, H. Huet, C. Djediat, V. Jeannoda, L. Mambu, F. Crespeau and M. Edery, *Food Chem. Toxicol.*, 2009, **47**, 2289–2293.
- 299 R. Jaretzky and F. Scheermesser, *DE Patent* 1935-J52167, 1935.
- 300 A. Katz, *Pharm. Acta Helv.*, 1954, **29**, 77–80.
- 301 A. Katz, *Experientia*, 1956, **12**, 285–286.
- 302 A. Katz, *Helv. Chim. Acta*, 1950, **33**, 1420–1428.
- 303 A. Katz, *Helv. Chim. Acta*, 1953, **36**, 1417–1423.
- 304 A. Katz, *Helv. Chim. Acta*, 1953, **36**, 1344–1352.
- 305 A. Katz, *Helv. Chim. Acta*, 1954, **37**, 833–836.
- 306 A. Katz, *Helv. Chim. Acta*, 1954, **37**, 451–454.
- 307 A. Katz, *Helv. Chim. Acta*, 1955, **38**, 1565–1572.
- 308 A. Katz, *Helv. Chim. Acta*, 1957, **40**, 831–846.
- 309 A. Katz, *Helv. Chim. Acta*, 1958, **41**, 1399–1404.
- 310 R. Tschesche and U. Dolberg, *Chem. Ber.*, 1957, **90**, 2378–2382.
- 311 R. Tschesche and U. Dolberg, *Chem. Ber.*, 1958, **91**, 2512–2523.
- 312 R. Tschesche, H. W. Sarau and K. Sellhorn, *Chem. Ber.*, 1955, **88**, 1612–1619.
- 313 R. Tschesche and K. Sellhorn, *Chem. Ber.*, 1953, **86**, 54–62.
- 314 R. Tschesche, S. Goenechea and G. Snatzke, *Justus Liebigs Ann. Chem.*, 1964, **674**, 176–184.
- 315 G. Schenck and F. Rattinger, *US Patent* 1951-21868, 1951.
- 316 M. A. Angarskaya, P. I. Bezruk and D. A. Tkachenko, *Farmakol. i Toksikol., Sb., (Kiev Zdorov'c)*, 1964, **1**, 20–25.
- 317 F. Hildebrandt and E. Paas, *Naunyn-Schmiedeberg's Arch. Pharmakol. Exp. Pathol.*, 1953, **220**, 482–489.
- 318 F. Hildebrandt and W. Reinartz, *Naunyn-Schmiedeberg's Arch. Pharmakol. Exp. Pathol.*, 1953, **220**, 290–294.
- 319 A. Katz, *CH Patent* 324078, 1957.
- 320 A. Katz, *CH Patent* 330660, 1958.
- 321 A. Katz, *Pharm. Acta Helv.*, 1954, **29**, 369–378.
- 322 K. K. Chen and F. G. Henderson, *J. Pharmacol.*, 1951, **103**, 420.
- 323 K. K. Chen and F. G. Henderson, *J. Med. Chem.*, 1961, **3**, 111–127.
- 324 K. K. Chen, *J. Med. Chem.*, 1970, **13**, 1029–1034.
- 325 C. Koorbanally, D. A. Mulholland and N. R. Crouch, *Biochem. Syst. Ecol.*, 2005, **33**, 545–549.
- 326 E. J. Ndebia, E. Umapathy, B. N. Nkeh-Chungag and J. E. Iputo, *J. Med. Plants Res.*, 2011, **5**, 4658–4664.
- 327 S. O. Oyedemi, G. Bradley and A. J. Afolayan, *J. Med. Plants Res.*, 2009, **3**, 1040–1044.
- 328 T. Okanishi, A. Akahori, F. Yasuda, Y. Takeuchi and T. Iwao, *Chem. Pharm. Bull.*, 1975, **23**, 575–579.
- 329 M. Kuroda, Y. Mimaki, Y. Sashida, T. Yamori and T. Tsuruo, *Tennen Yuki Kagobutsu Toronkai Koen Yoshishu*, 1999, **41**, 529–534.
- 330 Y. Mimaki, M. Kuroda, Y. Sashida, T. Yamori and T. Tsuruo, *Helv. Chim. Acta*, 2000, **83**, 2698–2704.
- 331 M. Kuroda, Y. Mimaki, Y. Sashida, T. Yamori and T. Tsuruo, *Tetrahedron Lett.*, 2000, **41**, 251–255.
- 332 Y. Mimaki, M. Kuroda, A. Yokosuka and Y. Sashida, *J. Nat. Prod.*, 2001, **64**, 1069–1072.
- 333 M. Kuroda, Y. Mimaki, A. Yokosuka and Y. Sashida, *Chem. Pharm. Bull.*, 2001, **49**, 1042–1046.
- 334 P. Tang and B. Yu, *Angew. Chem., Int. Ed.*, 2007, **46**, 2527–2530.
- 335 P. Tang and B. Yu, *Eur. J. Org. Chem.*, 2009, 259–269.
- 336 K. du Toit, PhD Thesis, University of Natal, South Africa, 2004.
- 337 F. Klar, *Planta Med.*, 2011, **77**, 1274–1274.
- 338 S. Kubo, Y. Mimaki, M. Terao, Y. Sashida, T. Nikaido and T. Ohmoto, *Phytochemistry*, 1992, **31**, 3969–3973.
- 339 S. Kubo, Y. Mimaki, Y. Sashida, T. Nikaido and T. Ohmoto, *Chem. Pharm. Bull.*, 1992, **40**, 2469–2472.
- 340 M. Kuroda, Y. Mimaki, Y. Sashida, T. Nikaido and T. Ohmoto, *Tetrahedron Lett.*, 1993, **34**, 6073–6076.
- 341 M. Kuroda, Y. Mimaki, Y. Sashida, T. Hirano, K. Oka and A. Dobashi, *Chem. Pharm. Bull.*, 1995, **43**, 1257–1259.
- 342 M. Kuroda, Y. Mimaki and Y. Sashida, *Phytochemistry*, 1999, **52**, 435–443.
- 343 T. Hirano, K. Oka, Y. Mimaki, M. Kuroda and Y. Sashida, *Life Sci.*, 1996, **58**, 789–798.
- 344 M. Kuroda, Y. Mimaki, Y. Sashida, T. Hirano, K. Oka, A. Dobashi, H.-Y. Li and N. Harada, *Tetrahedron*, 1997, **53**, 11549–11562.
- 345 Y. Mimaki and Y. Sashida, *Adv. Exp. Med. Biol.*, 1996, **404**, 101–110.
- 346 Y. Mimaki, M. Kuroda, A. Kameyama, Y. Sashida, T. Hirano, K. Oka and A. Dobashi, *Bioorg. Med. Chem. Lett.*, 1996, **6**, 2635–2638.
- 347 Y. Mimaki, M. Kuroda, Y. Sashida, T. Hirano, K. Oka, A. Dobashi, H. Koshino and J. Uzawa, *Tetrahedron Lett.*, 1996, **37**, 1245–1248.
- 348 Y. Sashida, *Stud. Plant Sci.*, 1999, **6**, 201–211.
- 349 M. Kuroda, Y. Mimaki, H. Koshino and Y. Sashida, *Heterocycles*, 2002, **56**, 531–536.



- 350 Y. Mimaki, M. Kuroda, A. Kameyama, Y. Sashida, T. Hirano, K. Oka, K. Koike and T. Nikaido, *Biosci., Biotechnol., Biochem.*, 1996, **60**, 1049–1050.
- 351 Y. Mimaki, M. Kuroda, A. Kameyama, Y. Sashida, T. Hirano, K. Oka, R. Maekawa, T. Wada, K. Sugita and J. A. Beutler, *Bioorg. Med. Chem. Lett.*, 1997, **7**, 633–636.
- 352 M. Kuroda, Y. Mimaki and Y. Sashida, *Phytochemistry*, 1999, **52**, 445–452.
- 353 M. Kuroda, Y. Mimaki, A. Yokosuka, Y. Sashida and J. A. Beutler, *J. Nat. Prod.*, 2001, **64**, 88–91.
- 354 Y. Sashita, K. Oka, T. Hirano, Y. Mimaki, A. Kuroda, A. Fujii and Y. Myata, *JP Patent* 1996-109329, 1996.
- 355 Y. Sashita, K. Oka, T. Hirano, Y. Mimaki, A. Kuroda, A. Fujii and Y. Myata, *JP Patent* 1996-107254, 1996.
- 356 Y. Sashita, K. Oka, T. Hirano, Y. Mimaki, A. Kuroda, A. Fujii and Y. Myata, *JP Patent* 1996-107255, 1996.
- 357 Y. Hui, Z. Yang and Q. Ge, *CN Patent* 2006-10147244, 2006.
- 358 Y. Hui, Z. Yang, C. Yu, Q. Ge and L. Bao, *CN Patent* 2006-10116980, 2006.
- 359 Y. Zhou, C. Garcia-Prieto, D. A. Carney, R.-H. Xu, H. Pelicano, Y. Kang, W. Yu, C. Lou, S. Kondo, J. Liu, D. M. Harris, Z. Estrov, M. J. Keating, Z. Jin and P. Huang, *J. Natl. Cancer Inst.*, 2005, **97**, 1781–1785.
- 360 W. Ying, Y.-L. Wu, X.-C. Feng, L.-H. Lian, Y.-Z. Jiang and J.-X. Nan, *J. Ethnopharmacol.*, 2010, **132**, 450–455.
- 361 Y. Wan, Y.-L. Wu, L.-H. Lian and J.-X. Nan, *Chin. J. Nat. Med.*, 2012, **10**, 0177–0184.
- 362 R. A. Waud, *J. Pharmacol. Exp. Ther.*, 1951, **101**, 36–36.
- 363 R. A. Waud and J. Boyd, *J. Pharmacol. Exp. Ther.*, 1954, **111**, 147–151.
- 364 R. A. Waud and J. Boyd, *Rev. Can. Biol.*, 1952, **11**, 88.
- 365 A. Vogelsang, *Can. Med. Assoc. J.*, 1955, **73**, 295–296.
- 366 H. Mrozik, R. A. Waud, O. Schindler and T. Reichstein, *Helv. Chim. Acta*, 1959, **42**, 683–696.
- 367 R. B. Kelly, E. G. Daniels and L. B. Spaulding, *J. Med. Chem.*, 1965, **8**, 547–548.
- 368 F. E. Hart and G. R. Paterson, *Can. J. Pharm. Sci.*, 1968, **3**, 15–18.
- 369 J. A. Smith and G. R. Paterson, *J. Pharm. Pharmacol.*, 1967, **19**, 221–225.
- 370 R. Ferth and B. Kopp, *Die Pharmazie*, 1992, **47**, 626–629.
- 371 Y. Buchvarov, M. Pargov and K. Akhtardzhiev, *Farmatsiya (Sofia, Bulg.)*, 1976, **26**, 31–34.
- 372 O. Gasic, V. Simanek, V. Lukic, D. Walterova, S. Kevresan, V. Hanus and B. Pal, *Zb. Matice Srp. Prir. Nauke*, 1989, **76**, 21–26.
- 373 O. Azzoui, R. L. Braemer and M. Paris, *Biochem. Syst. Ecol.*, 1989, **17**, 449–450.
- 374 T. Sabudak and U. Oyman, *Turk. J. Chem.*, 2002, **26**, 453–455.
- 375 W. Schliemann, J. Schmidt, M. Nitz, V. Wray, T. Fester and D. Strack, *Phytochemistry*, 2006, **67**, 1196–1205.
- 376 N. F. Komissarenko, *Khim. Prir. Soedin.*, 1971, **7**, 38–40.
- 377 N. F. Komissarenko, *Postep Dziedzinie Leku Rosl., Pr. Ref. Dosw. Wygloszone Symp.*, 1972, 67–71.
- 378 I. G. Zoz and N. A. Chernykh, *Farm. Zh. (Kiev)*, 1969, **24**, 77–78.
- 379 N. F. Komissarenko, *Khim. Prir. Soedin.*, 1969, **5**, 46–48.
- 380 N. F. Komissarenko, *Khim. Prir. Soedin.*, 1965, 156–160.
- 381 N. F. Komissarenko, *Khim. Prir. Soedin.*, 1969, **5**, 46–48.
- 382 N. F. Komissarenko, *Khim. Prir. Soedin.*, 1972, 397–398.
- 383 R. Ferth, A. Baumann, W. Robien and B. Kopp, *Z. Naturforsch., B: J. Chem. Sci.*, 1992, **47**, 1459–1467.
- 384 R. Ferth, A. Baumann, W. Robien and B. Kopp, *Z. Naturforsch., B: J. Chem. Sci.*, 1992, **47**, 1444–1458.
- 385 N. F. Komissarenko and P. E. Krivenchuk, *Khim. Prir. Soedin.*, 1974, **10**, 257.
- 386 U. Ghannamy, B. Kopp, W. Robien and W. Kubelka, *Planta Med.*, 1987, **53**, 172–178.
- 387 V. A. Bandyukova, *Chem. Nat. Compd.*, 1979, **15**, 639.
- 388 M. Nicoletti, L. Tomassini and S. Foddai, *Planta Med.*, 1992, **58**, 472.
- 389 V. V. Barbakadze, E. P. Kemertelidze, H. E. Dekanosidze and A. I. Usov, *Bioorg. Khim.*, 1993, **19**, 912–916.
- 390 Y. Buchvarov, E. Balabanova-Radonova, I. Radenkova and S. Vankov, *Farmatsiya (Sofia, Bulg.)*, 1984, **34**, 33–37.
- 391 A. A. Makasci, R. Mammadov, O. Dusen and H. I. Isik, *J. Med. Plants Res.*, 2010, **4**, 1637–1642.
- 392 E. Nazifi, A. Movafeghi, H. Nazemiyeh, S. Asnaashari, S. Bamdad Moghadam and A. Delazar, *Ulum-i Daroei*, 2010, **16**, 37–44.
- 393 A. Delazar, E. Nazifi, A. Movafeghi, H. Nazemiyeh, S. Hemmati, L. Nahar and S. D. Sarker, *Bol. Latinoam. Caribe Plant. Med. Aromat.*, 2010, **9**, 87–92.
- 394 A. Delazar, E. Nazifi, A. Movafeghi, L. Nahar, H. Nazemiyeh, S. B. Moghadam, S. Asnaashari and S. D. Sarker, *Daru, J. Fac. Pharm., Tehran Univ. Med. Sci.*, 2009, **17**, 33–36.
- 395 M. A. Ebrahimzadeh, S. M. Nabavi, S. F. Nabavi and B. Eslami, *Trop. J. Pharm. Res.*, 2010, **9**, 141–148.
- 396 S. Kubo, Y. Mimaki, Y. Sashida, T. Nikaido and T. Ohmoto, *Bull. Chem. Soc. Jpn.*, 1992, **65**, 1120–1124.
- 397 M. Kuroda, Y. Mimaki, A. Yokosuka, F. Hasegawa and Y. Sashida, *J. Nat. Prod.*, 2002, **65**, 1417–1423.
- 398 M. Kuroda, Y. Mimaki, K. Ori, H. Sakagami and Y. Sashida, *J. Nat. Prod.*, 2004, **67**, 1690–1696.
- 399 M. Kuroda, K. Ori and Y. Mimaki, *Steroids*, 2006, **71**, 199–205.
- 400 M. Kuroda, Y. Mimaki, A. Yokosuka, F. Hasegawa and Y. Sashida, *J. Nat. Prod.*, 2002, **65**, 1417–1423.
- 401 M. Kuroda, Y. Mimaki, K. Ori, H. Sakagami and Y. Sashida, *J. Nat. Prod.*, 2004, **67**, 1690–1696.
- 402 W. T. Mabusela and A. M. Stephen, *Carbohydr. Res.*, 1990, **207**, 332–335.
- 403 D. A. Mulholland, N. R. Crouch, T. Pohl and E. Ndlovu, *Biochem. Syst. Ecol.*, 2004, **32**, 499–502.
- 404 M. van Huyssteen, P. J. Milner, E. E. Campbell and M. van de Venter, *Afr. Networks Ethnomed.*, 2011, **8**, 150–158.
- 405 Y. Tang, B. Yu, J. Hu, T. Wu and H. Hui, *J. Nat. Prod.*, 2002, **65**, 218–220.
- 406 T. Xu, Y. Xu, D. Liu and D. Xu, *Yaoxue Xuebao*, 2000, **35**, 32–36.
- 407 Y. P. Tang, B. Yu, J. Hu, T. Wu and H. Z. Hui, *J. Nat. Prod.*, 2002, **65**, 218–220.



- 408 Y. Tang, B. Yu, J. Hu, T. Wu and Y. Hui, *J. Chin. Pharm. Sci.*, 2001, **10**, 169–171.
- 409 J. F. Bai, Z. Q. Liu, S. M. Wang, F. R. Song and S. Y. Liu, *Chem. J. Chin. Univ.*, 2005, **26**, 1817–1819.
- 410 Z. Han, *CN Patent* 2001-18727, 2001.
- 411 Z. Han, *CN Patent* 2006-10115717, 2006.
- 412 X. Lu and Z. Han, *CN Patent* 2004-10074137, 2004.
- 413 G. I. Mynkina, *RU Patent* 1999-108770, 1999.
- 414 J. Sun, G. Lu and L. Xiu, *CN Patent* 2010-10177347, 2010.
- 415 X. Jin, G. Cui, C. Qian, X. Du and Y. Wu, *Shizhen Guoyi Guoyao*, 2010, **21**, 124–126.
- 416 C. Qian, X. Jin, Y. Wu and X. Li, *Shizhen Guoyi Guoyao*, 2010, **21**, 636–638.
- 417 L. Shi, J. Li, W.-X. Liu, Y. Wang, Z.-Q. Liu and S.-Y. Liu, *Chem. Res. Chin. Univ.*, 2003, **19**, 286–289.
- 418 R. Chen, S. Li, Z. Liu, H. Dong, Y. Li and S. Yang, *Zhongguo Yaoxue Zazhi (Beijing, China)*, 2011, **46**, 1630–1634.
- 419 R. Chen, F. Meng, Z. Liu, R. Chen and M. Zhang, *Carbohydr. Polym.*, 2010, **80**, 845–851.
- 420 X. Liu and Y.-H. Zhang, *Shizhen Guoyi Guoyao*, 2012, **23**, 1388–1390.
- 421 O. Q. Munro, K. du Toit, S. E. Drewes, N. R. Crouch and D. A. Mulholland, *New J. Chem.*, 2006, **30**, 197–207.
- 422 Y. Sashida, *Stud. Plant Sci.*, 1999, **6**, 201–211.
- 423 Y. Mimaki, *Nat. Prod. Commun.*, 2006, **1**, 247–253.
- 424 E. R. Guaglianone and S. Arroyo-Leuenerger, *Darwiniana*, 2002, **40**, 61–76.

