

A two-directional approach to pyrrolizidines: total syntheses and biological evaluation of alkaloid *cis*-223B and (\pm)-xenovenine†

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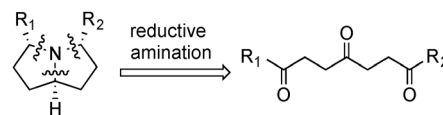
Total syntheses of alkaloid *cis*-223B and xenovenine are reported in 3 and 4 steps respectively using a two-directional synthesis/triple reductive amination strategy, and their neurotoxic properties assessed.

Pyrrolizidine alkaloids (PAs) are widely present in nature. In plants they are produced as a defence mechanism against insect herbivores, where more than 650 PAs and PA *N*-oxides have been isolated from 6000 different species (REF). This class of alkaloids is also present in moths, ants and poisonous frogs. The occurrence in anuran skin was first reported in 1993 for a Bufonid (*Melanophryniscus*) toad.¹ To date, 26 3,5-PAs including stereoisomers have been assigned to the class. These compounds display a broad range of biological activity, including glucosidase inhibition and anti-HIV potency, and can also play a necrotic and allergic role against living organisms, as they are used as chemical defence by batrachians.² There are cases where the same alkaloids have been isolated in myrmicine ants, raising the hypothesis that the presence of some alkaloids in frogs could be caused by their dietary source in this case, illustrated by alkaloid *cis*-223H.¹

In order to quickly access this class of molecules, we proposed a simple bio-inspired approach through a triple reductive amination. Starting from a triketone, we would form in one step 2 rings and 3 carbon–nitrogen bonds (Scheme 1). We surmised that the folded structure of the bicyclic system would enable us to attain the required stereochemistry by thermodynamic control.

Although there is no direct biomimetic proof of the supposed synthetic pathway for these pyrrolizidines, we inferred a similar mechanism to the one observed for alkaloid biosynthesis in ants and ladybirds.³ It has been particularly well documented in the case of coccinelline and precoccinelline.⁴

To explore our bio-inspired triple reductive amination strategy, two natural product targets were selected: the symmetrical alkaloid *cis* 223B **1**, and non-symmetrical (\pm)-xenovenine **2** (Fig. 1).



Scheme 1 Retrosynthetic analysis.

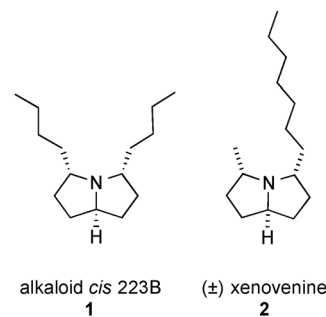


Fig. 1 Structures of alkaloid *cis*-223B and xenovenine.

We surmised that adaptation of our previous methods in two-directional synthesis⁵ would be highly suited to the simple synthesis of the triketone precursors.

Alkaloid *cis* 223B contains two stereogenic centres. Its central planar symmetry provides a *meso* configuration. **1** was identified by GC-MS for the first time in 1989 in bufonid toads *Melanophryniscus stelzneri* (Argentina) and the frog *Oophaga pumilio* (Panama) but never was isolated.^{1,6} The first and only synthesis of alkaloid *cis* 223B was reported by our group in 2009: the latter was obtained in 7 steps and 43% overall yield.⁷ A synthesis of the *trans* configuration was later reported by Toyooka.⁸ (\pm)-Xenovenine was isolated in 1980 from the venom of the ant species *Solenopsis xenoveneum* by Jones and co-workers.⁹ Structurally, this molecule is formed of a pyrrolizidine core with a 3,5-unsymmetrical substitution. Various syntheses of **2** have been published between 1986 and 2000.¹⁰ The biological properties of either compound have not been reported.

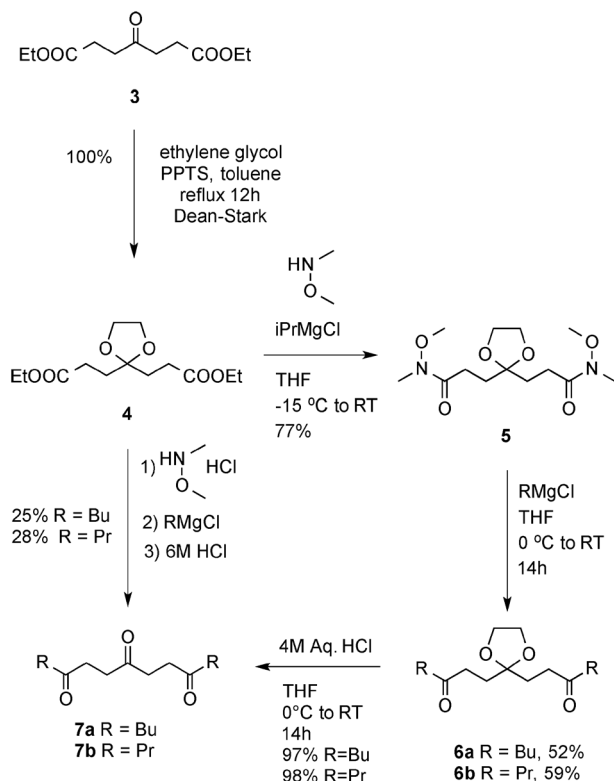
Using a two-directional approach, our initial synthesis started with the acetal protection of diethyl 4-oxopimelate **3** to the diester **4** (Scheme 2). The latter was then converted to Weinreb diamide **5**.

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Scheme 2 Synthesis of cyclisation precursors.

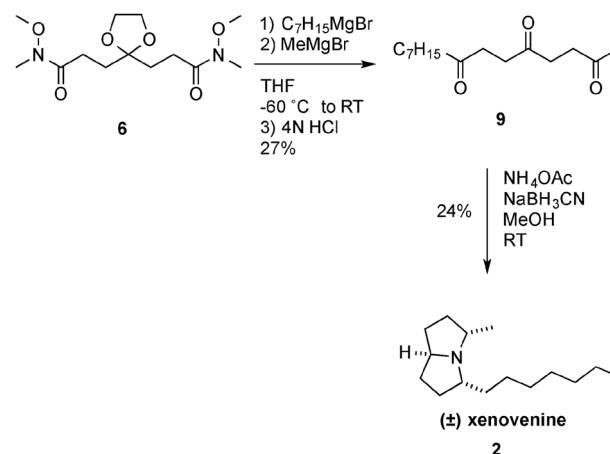
The dialkylation of **5** with butylmagnesium bromide and subsequent deprotection of the central ketone gave access to the key triketone intermediate **7a** in 4 linear steps and 39% yield. The process was then later streamlined by simply allowing the deprotection to occur directly after the alkylation. A further development even allowed access to **7a** directly from **4**, enabling in one pot: diamide formation, dialkylation and acetal deprotection in 25% overall yield.

To convert the triketone precursors **7a** and **7b** to the corresponding alkaloid *cis*-223B and the bis C3-analogue **8**, several reductive amination conditions were attempted (Table 1). Conditions were optimised for the precursor **7b**, with the best result obtained with 2 equivalents of sodium cyanoborohydride and 1.6 equivalents of ammonium acetate in methanol at 23 °C for 14 hours, giving a modest 29% yield of pyrrolizidine **8**. These same conditions gave an improved 40% yield of alkaloid *cis*-223B when applied to the reaction of the longer chain triketone **7a**. This increase in yield is likely down to the reduced volatility of the pyrrolizidine **1** vs. **8**.

Having successfully demonstrated our two-directional approach for the synthesis of alkaloid *cis*-223B and chain-shortened analogue **8**, we next turned our attention to the possibility of accessing non-symmetrical pyrrolizidines. Using a sequential Grignard addition strategy to symmetric protected diamide **6**, with addition of the longer chain first, followed by the methyl group, we were able to gain access to the non-symmetrical triketone **9**. The best isolated yield obtained for **9** after optimisation was 27% (Scheme 3). Our optimised conditions for the triple reductive cyclisation was then successfully applied to triketone **9**, which, upon reaction gave (±)-xenovenine **2** as a single diastereoisomer in 24% yield. Jones reported a related triple reductive amination

Table 1 Optimisation of triple reductive amination of triketones

Entry	Solvent	Reagents/eq.	Temp./time	Product	Yield (%)
1	MeOH	NaBH ₃ CN 2.0 NH ₄ OAc 1.6 KOH 0.2	RT 14 h	8	13
2	MeOH	NaBH ₃ CN 2.0 NH ₄ OAc 1.6	RT 14 h	8	Traces
3	MeOH	NaBH ₃ CN 2.0 NH ₄ Cl 1.6	RT 14 h	8	29
4	THF	NaBH(OAc) ₃ 3.0 NH ₄ OAc 5.0 Cat. acetic acid	60 °C 1 h	8	25
5	EtOH	NH ₃ -EtOH 5.0 Ti(OEt) ₄ 2.5 NaBH ₄ 1.5	RT 14 h	8	7
6	HCO ₂ H	H ₂ NCHO 12.0	170 °C 4 h	8	—
7	MeOH	NaBH ₃ CN 2.0 NH ₄ OAc 1.6	RT 14 h	1	40
8	MeOH	Et ₃ SiH 2.0 NH ₄ OAc 1.6	RT 14 h	8	—



Scheme 3 Synthesis of xenovenine.

in his report of the isolation of xenovenine,¹¹ which gave a mixture of diastereomers. We were therefore particularly pleased to find that our optimised procedure gave the product as a single diastereomer.

PAs are known to be toxic to humans and other mammals¹² as well as insects.¹³ Most studies have focussed on PAs isolated from plants, and information about animal derived PAs is limited. The main toxic activity of plant PAs appears to be hepatotoxicity, caused indirectly by pyrrole metabolites.¹¹ Other alkaloids isolated from anurans are known to display neurotoxic activity by interacting directly with specific ion channels within the nervous system. One recognised target is the nicotinic acetylcholine receptor.¹⁴ Binding studies have shown that twelve of thirteen plant PAs tested do not bind competitively to neuronal-nAChR,¹⁵ but a number of PAs isolated from anuran skin



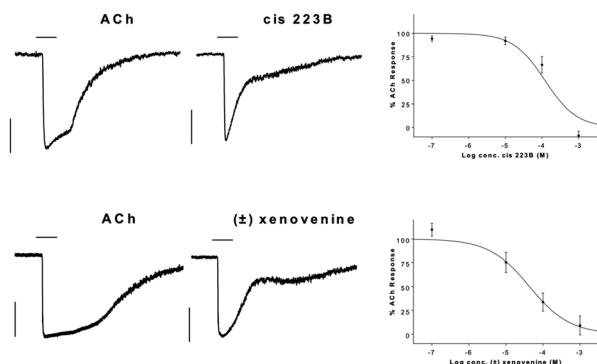


Fig. 2 Inhibition of ACh (10 μ M) induced currents in TE671 cells expressing human muscle-type nAChR by (a) *cis*-223B and (b) (\pm)-xenovenine at V_h = -75 mV. Typical traces showing ACh elicited currents (left) and when co-applied with pyrrolizidines (100 μ M) (centre). Vertical bars represent 200 pA and horizontal bars show 1 s perfusion period. Concentration–inhibition plots fitted with non-linear regression curves. The current at 1 s after onset (the point at which antagonism is greatest) was normalised against the preceding ACh only response at the same time point. Values represent mean \pm SEM for 5–12 cells.

display high affinity for *Torpedo* muscle-type nAChR.¹⁶ Spiropyrrrolizidines are non-competitive antagonists of nAChR which display selectivity for neuronal over muscle-type receptors.¹⁷ Here we show inhibition of acetylcholine induced nAChR currents by *cis*-223B and (\pm)-xenovenine using patch-clamp of TE671 cells expressing human muscle-type nAChR.¹⁸ The results shown in Fig. 2 demonstrates clear, concentration dependent inhibition of the steady state current by both *cis*-223B and (\pm)-xenovenine, with IC_{50} values of 120 μ M and 43.2 μ M respectively. Both compounds clearly inhibit the nicotinic acetylcholine response at high concentrations but the IC_{50} values generated are modest when compared with other alkaloids isolated from anuran skin. Spiropyrrrolizidines are known to show selectivity for neuronal over muscle-type nAChR,¹⁶ and it is possible that this is also true of these compounds. Further investigation into the selectivity of these compounds for neuronal-nAChR and different sub-types thereof would reveal their potential as therapeutic leads for nAChR aberrations such as slow channel myasthenic syndrome.

In conclusion we have applied a bio-inspired cascade reductive amination methodology to the synthesis of alkaloid *cis*-223B **1** (3 steps, 10% yield) and C3-analogue **8** (3 steps, 8% yield). This method was further developed to access unsymmetrical pyrrolizidine alkaloids, demonstrated by the diastereoselective synthesis of (\pm)-xenovenine **2** in 4 steps and 5% overall yield. Despite the relatively modest overall yields (partially due to volatility of the products), these synthetic pathways remain by far the shortest syntheses described for **1** and **2**, and demonstrate the overall effectiveness of the combined two-directional synthesis-tandem reaction strategy.¹⁹ The overall strategy should be applicable to a wide range of 3,5-disubstituted pyrrolizidines.

Furthermore, electrophysiology assays performed on nAChR from human muscle cells enabled us to determine the IC_{50} s of both natural products, confirming for the first time the suspected neurotoxic properties of these natural products.

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