



Cite this: *Mater. Horiz.*, 2019, 6, 1271

Received 14th December 2018,  
Accepted 29th March 2019

DOI: 10.1039/c8mh01606b

rsc.li/materials-horizons

# Bioinspired interface design modulates pathogen and immunocyte responses in biomaterial-centered infection combination therapy†

Jinhua Li,<sup>†ab</sup> Wei Liu,<sup>‡c</sup> David Kilian,<sup>a</sup> Xianlong Zhang,<sup>\*c</sup> Michael Gelinsky<sup>a</sup> and Paul K. Chu<sup>\*b</sup>

The rise of multidrug-resistant pathogens and the dearth of novel antibiotic development means that breakthrough strategies that go beyond the classical antibiotic mechanism are urgently required to fight this approaching human health cataclysm. Herein, inspired by the metabolism cascade of pathogens and the clearance of infection by the host immune system, a new three-step strategy is reported to achieve combination therapy of biomaterial-centered infections. Band-structure-tunable cobalt doped TiO<sub>2</sub> semiconductor can effectively restrain bacterial biofilm formation and enhance phagocytosing and killing of bacteria by immune cells. Released Co<sup>2+</sup> ions can produce a proinflammatory microenvironment and potentiate the antibacterial capability of immune cells. This design concept can be applied to develop other antibacterial biomaterials, and holds great promise for advanced biomaterial-centered infection therapy.

Nowadays it is increasingly necessary to understand the interaction mechanism between biomaterials and host tissues in a more sophisticated manner. Biomaterial-centered infections involve the interactions among biomaterials, bacteria/biofilms, and host innate immunity in the local environment.<sup>1,2</sup> In this context, it is highly required to investigate the impact of biomaterials on innate immunity more than the mere relationship between bacteria/biofilms and biomaterials. It is believed that the inflammatory response is highly beneficial for the host innate immune defense against bacterial infection; macrophages and neutrophils

## New concepts

An alarming and persistent rise in the antibiotic resistance among many important pathogenic bacterial species poses one of the greatest contemporary challenges to public health. Whereas most of the previous work developing antibacterial biomaterials is solely focused on inhibiting bacterial biofilm formation, herein, we demonstrate a new design concept of “infection combination therapy” for biomaterial-centered bacterial infections. This concept is achieved by three-step tactics: (1) band-structure-tailorable TiO<sub>2</sub> semiconductor through cobalt doping can act as the extracellular electron acceptors to interrupt the bacterial electron transfer (discharge process) and proton transfer (hydrogenation process), jeopardize the energy metabolism causing bacteria starvation, and thereby suppress bacterial biofilm formation (bacteria starvation therapy). (2) Innate immune cells (including macrophages and neutrophils) can effectively phagocytose and kill the biofilm-formation-blocked bacteria using the divide-and-conquer strategy. (3) Released cobalt ions can create a proinflammatory local environment and potentiate the phagocytosing and killing of bacteria by the innate immune cells (infection immunotherapy). This concept of infection combination therapy can contribute to the advanced design of biomaterials for antibacterial treatment. We anticipate that this novel design concept can be extended to developing other antibacterial biomaterials and find a wide range of applications in the advanced therapy of biomaterial-centered bacterial infections.

are important innate immune cells, and play a central role as effector cells in the immune response by initiating and maintaining inflammation.<sup>3</sup> Therefore, strategies that can incorporate, modulate, and even encourage the host immune response could provide a new paradigm for biomaterial-centered infection therapy. This may be manipulated *via* the modification and functionalization of biomaterials.<sup>4,5</sup> For example, in response to varied tissue microenvironment cues, highly plastic macrophages can switch phenotypes and functions to the classically activated pro-inflammatory M1 and alternatively activated anti-inflammatory M2 forms,<sup>6</sup> and the macrophage polarization can be modulated by the physical and chemical cues of biomaterials.<sup>7,8</sup> It has been reported that biofilms can impair host innate immunity through skewing macrophages to the M2 phenotype and inhibiting the inflammatory reaction around the infective site, thereby impairing the clearance of bacteria by immune cells.<sup>9</sup>

<sup>a</sup> Centre for Translational Bone, Joint and Soft Tissue Research, University Hospital and Faculty of Medicine Carl Gustav Carus, Technische Universität Dresden, Fetscherstraße 74, Dresden 01307, Germany

<sup>b</sup> Department of Physics and Department of Materials Science and Engineering, City University of Hong Kong, Tat Chee Avenue, Kowloon, Hong Kong, China. E-mail: paul.chu@cityu.edu.hk

<sup>c</sup> Department of Orthopaedics, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai Jiao Tong University, Shanghai 200233, China. E-mail: dr\_zhangxianlong@163.com

† Electronic supplementary information (ESI) available: Details on the experimental procedures, and supplementary figures (Fig. S1–S34), tables (Tables S1–S5), and references. See DOI: 10.1039/c8mh01606b

‡ These authors contributed equally to this work.



Therefore, modulating the macrophage polarization response to biomaterials may be a promising strategy for eliciting desired outcomes of biomaterial-centered infection therapy. In fact, bacteria are exposed to a rapid and profound host innate immune response characterized by the recruitment of neutrophils and macrophages to sites of infection; these cells are essential for the initiation and execution of the inflammatory response and subsequent resolution of infection.<sup>10</sup> Furthermore, an in-depth understanding of the mechanism underlying the immune response mediated by biomaterials would aid in the development of novel biomaterials that can create a beneficial host immune microenvironment for combating bacterial infection.

Extracellular electron transfer (EET) is the description of the bacterial bioelectrochemical process that electrons are transferred from the cytosol to the cellular exterior.<sup>11,12</sup> Redox-active minerals are abundant in soils and in aquatic or subsurface sediments; they can electrically support bacterial growth by serving as electron sinks for heterotrophy-based respiration.<sup>13,14</sup> Such minerals include the solid oxides/hydroxides of Fe(III) and Mn(IV)<sup>15,16</sup> with relatively high redox potentials as extracellular electron acceptors, including  $\text{Fe}(\text{OH})_3/\text{Fe}^{2+}$  ( $E^\circ = +1.06$  V),  $\text{FeOOH}/\text{Fe}^{2+}$  ( $E^\circ = +0.67$  V),  $\text{Fe}_2\text{O}_3/\text{Fe}^{2+}$  ( $E^\circ = +0.66$  V) and  $\text{MnO}_2/\text{Mn}^{2+}$  ( $E^\circ = +1.29$  V).<sup>17</sup> Inspired by these biological processes, we hypothesize that (i) tuning the band structure of the  $\text{TiO}_2$  semiconductor by cobalt doping can inhibit bacterial biofilm formation through interrupting extracellular electron transfer and proton transfer by acting as extracellular electron acceptors, (ii) innate immune cells *e.g.* macrophages and neutrophils can destroy the biofilm-formation-blocked bacteria using the divide-and-conquer strategy, and (iii) released cobalt ions can reinforce the antibacterial function of innate immune cells when maintaining biocompatibility. We demonstrate herein the capability to modulate the behaviors of bacteria and innate immune cells on cobalt-doped  $\text{TiO}_2$  coating by facile plasma modification, thereby leading to combination therapy for biomaterial-centered infections.

The samples were prepared using a plasma electrolytic oxidation (PEO) apparatus. The preparation parameters are shown in Table S1 (ESI†). The pristine sample was undoped  $\text{TiO}_2$  coating ( $\text{TiO}_2\text{:Co-0}$ ), while a series of Co-doped  $\text{TiO}_2$  coatings ( $\text{TiO}_2\text{:Co-0.5}$ ,  $\text{TiO}_2\text{:Co-1}$ ,  $\text{TiO}_2\text{:Co-2}$ ,  $\text{TiO}_2\text{:Co-4}$ ) were acquired by tuning the Co dopant concentration. Surface morphologies of the samples were determined by scanning electron microscopy (SEM).  $\text{TiO}_2\text{:Co-0}$  had a homogeneous porous coating on its surface with smooth morphology at higher magnification (inset) and Co doping did not sharply change the surface topographies of the coatings (Fig. 1A). Crystal structures of the samples were characterized by X-ray diffraction (Fig. 1B). In  $\text{TiO}_2\text{:Co-0}$ , the coating mainly consisted of anatase  $\text{TiO}_2$  phase (JPCDS No. 21-1272). When performing PEO in Co-containing electrolyte,  $\text{Co}^{2+}$  might occupy the substitutional sites to replace  $\text{Ti}^{4+}$  in the host lattice or enter into the interstitial lattice sites due to the fact that, for the hexacoordinated central metal with octahedron geometry,  $\text{Co}^{2+}$  and  $\text{Ti}^{4+}$  have very similar ionic radii of 79 pm and 74.5 pm, respectively. Hence, X-ray photoelectron spectroscopy (XPS), that

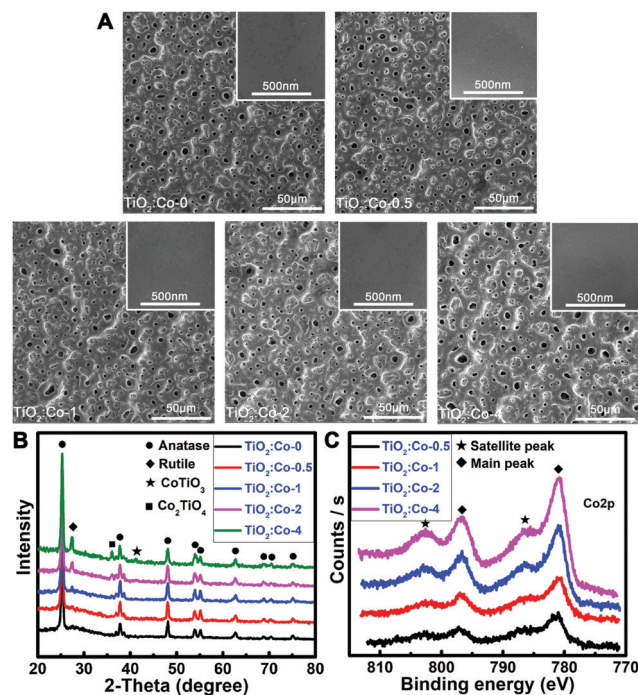


Fig. 1 Surface topographies and compositions of the coating samples. (A) SEM images of the coatings with inserted high resolution images on the top-right. (B) XRD patterns of the coatings. (C) High resolution  $\text{Co}2p$  XPS spectra acquired from the surfaces of Co-doped coatings.

is a surface-sensitive characterization technique, was utilized to determine the surface chemistry. XPS full spectra and elemental content data (Fig. S1 and Table S2, ESI†) of the coatings reveal that increasing the  $\text{Co}^{2+}$  concentration of the electrolyte can increase the Co doping content in the  $\text{TiO}_2$  coating. Fig. 1C gives the high-resolution  $\text{Co}2p$  XPS spectra and the fitted spectra are shown in Fig. S2 (ESI†). The absence of a metallic  $\text{Co}2p_{3/2}$  peak at 778 eV with  $\Delta = 14.86$  eV indicated the inexistence of Co metal phase. There are clear characteristics in the 2p core level photo-emission spectra that allow unambiguous distinguishing between  $\text{Co}^{2+}$  and  $\text{Co}^{3+}$ . First, the  $\text{Co}^{2+}$ -2p core level spectrum possesses strong shakeup satellite peaks toward higher binding energies, different from the very weak ones of the  $\text{Co}^{3+}$ -2p core level spectrum.<sup>18,19</sup> Second, the splitting of the  $\text{Co}2p_{1/2}$ - $\text{Co}2p_{3/2}$  orbital component  $\Delta$  is usually 16 eV for the high-spin  $\text{Co}^{2+}$ , while it is 15 eV for the low-spin  $\text{Co}^{3+}$ .<sup>18</sup> The existence of strong shakeup satellite peaks at 786.4 eV and 802.6 eV indicated the oxidation state of 2+ of the Co atom in the  $\text{TiO}_2$  coating. Meanwhile, the measured  $\Delta$  (spin-orbit splitting) value from the main peaks at 781.0 eV and 796.9 eV was 15.9 eV, which was accordant with the valence state of 2+. Following the increase of  $\text{Co}^{2+}$  doping level in  $\text{TiO}_2$  coating, new diffraction peaks gradually appeared for the  $\text{CoTiO}_3$  phase (JCPDS No. 29-0516) and  $\text{Co}_2\text{TiO}_4$  phase (JCPDS No. 39-1410). Both anatase  $\text{TiO}_2$  ( $I4_1/amd$ ) and rutile  $\text{TiO}_2$  ( $P4_2/mnm$ ) have a tetragonal crystal structure, but the rutile phase has a smaller lattice parameter (*c*-axis) and unit cell volume.<sup>20</sup> During the formation of new  $\text{CoTiO}_3$  and  $\text{Co}_2\text{TiO}_4$  phases, crystal lattice rearrangement occurred and might impose extrusion stress on the surrounding  $\text{TiO}_2$  unit cell and cause the



shrink of the *c*-axis. As a result, a phase transformation from anatase to rutile might occur upon  $\text{Co}^{\text{II}}\text{Ti}^{\text{IV}}\text{O}_3$  and  $\text{Co}^{\text{II}}_2\text{Ti}^{\text{IV}}\text{O}_4$  phase formation. Meanwhile, increasing amounts of oxygen vacancies were created in the  $\text{TiO}_2$  lattice upon subvalent  $\text{Co}^{2+}$  replacing  $\text{Ti}^{4+}$  sites of the lattice for charge compensation, which also promoted the rutile formation.<sup>21</sup> In fact, new rutile phase (JPCDS No. 21-1276) emerged in the  $\text{TiO}_2$  coating when increasing the  $\text{Co}^{2+}$  dopant content, especially in  $\text{TiO}_2\text{:Co-2}$  and  $\text{TiO}_2\text{:Co-4}$ . In addition, there were no peaks of metallic Co or binary cobalt oxides in the coatings even at a high cobalt content of 6.03 at%. Therefore,  $\text{Co}^{2+}$  largely substituted for the  $\text{Ti}^{4+}$  sites of the  $\text{TiO}_2$  lattice and the phase transformation of anatase to rutile implied that the solubility limit of  $\text{Co}^{2+}$  in  $\text{TiO}_2$  was  $\sim 2$  at%, which was quite consistent with the determined value for  $\text{Co}^{2+}$ -doped  $\text{TiO}_2$  nanopowders.<sup>22</sup> Besides, the fitted Ca2p XPS spectra (Fig. S3, ESI†) included the predominant peaks at 347.1 eV and 350.7 eV belonging to  $\text{Ca}_3(\text{PO}_4)_2$  and the third speak at 347.5 eV assigned to  $\text{CaHPO}_4$ ; accordingly, the fitted P2p XPS peaks at 133.6 eV and 132.8 eV (Fig. S4, ESI†) pertain to  $\text{PO}_4^{3-}$  and  $\text{HPO}_4^{2-}$ , respectively.<sup>23</sup> Hence,  $\text{Co}^{2+}$  was substitutionally doped and homogeneously distributed in the  $\text{TiO}_2$  matrix.

UV-Vis absorption spectra of the samples were determined (Fig. 2A). A strong absorption below 400 nm for  $\text{TiO}_2\text{:Co-0}$  was attributed to the absorption edge of anatase  $\text{TiO}_2$ . Increasing the Co doping content shifted the absorption edge to a longer wavelength range. Meanwhile, there existed a clear broad peak between 500 nm and 700 nm, which resulted from the  $\text{CoTiO}_3$  and  $\text{Co}_2\text{TiO}_4$  phases,<sup>24</sup> and the intensity increased with the

$\text{Co}^{2+}$  dopant content. The homogeneous absorption in the visible light range reflects the homogeneous distribution of the additional energy levels, and the shoulder at  $\sim 530$  nm and band at  $\sim 605$  nm come from respective free excitons and bound excitons.<sup>25</sup> Fig. 2B depicts the  $(\alpha h\nu)^{1/2}$  plot versus  $h\nu$ , and the vertical segments of the spectra were extended to intersect with the  $h\nu$  axis to acquire the band gap ( $E_g$ ) of various samples. The  $E_g$  of  $\text{TiO}_2\text{:Co-0}$  was 3.10 eV, but the  $E_g$  of the Co-doped samples gradually decreased from 2.69 eV of  $\text{TiO}_2\text{:Co-0.5}$  to 1.55 eV of  $\text{TiO}_2\text{:Co-4}$  with increasing Co dopant content. Hexacoordinated  $\text{Co}^{2+}$  and  $\text{Ti}^{4+}$  have very similar ionic radii and  $\text{Co}^{2+}$  doping will produce defects in the  $\text{TiO}_2$  lattice and create additional energy levels (impurity levels or trap levels) between the conduction band (CB) and valence band (VB),<sup>25,26</sup> thereby leading to the narrowed band gap and enhanced visible light absorption with incremental Co doping content. Ultraviolet photoelectron spectroscopy (UPS) was employed to determine the VB maximum and work function (Fig. S5–S9, ESI†). Based on the data of VB maximum and band gap, the CB minimum was acquired (Table S3, ESI†). Fig. 2C summarizes the energy band information, including CB minimum (red), VB maximum (blue) and Fermi level ( $E_F$ , green) with incremental content of Co doping. The CB minimum for  $\text{TiO}_2\text{:Co-0}$ ,  $\text{TiO}_2\text{:Co-0.5}$ ,  $\text{TiO}_2\text{:Co-1}$ ,  $\text{TiO}_2\text{:Co-2}$ , and  $\text{TiO}_2\text{:Co-4}$  was  $-4.36$ ,  $-4.81$ ,  $-5.14$ ,  $-5.63$ , and  $-6.03$  eV, respectively. Table S4 (ESI†) lists the biological redox potentials (BRP) for the redox couples in the electron transfer system of the respiration chain located on the bacterial membrane (ESI†) and they are depicted in Fig. 2D according to the conversion equation

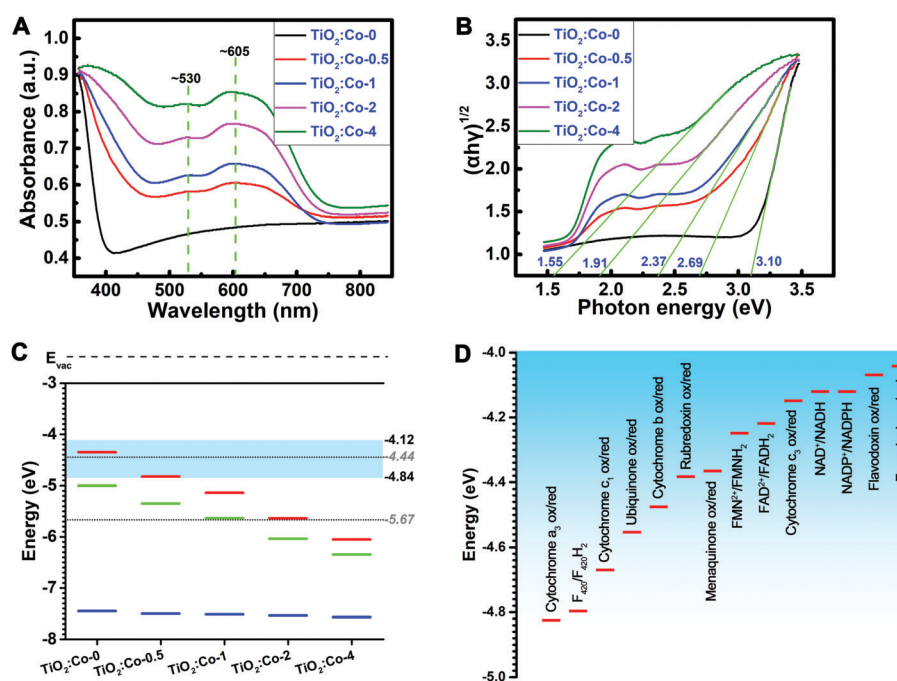


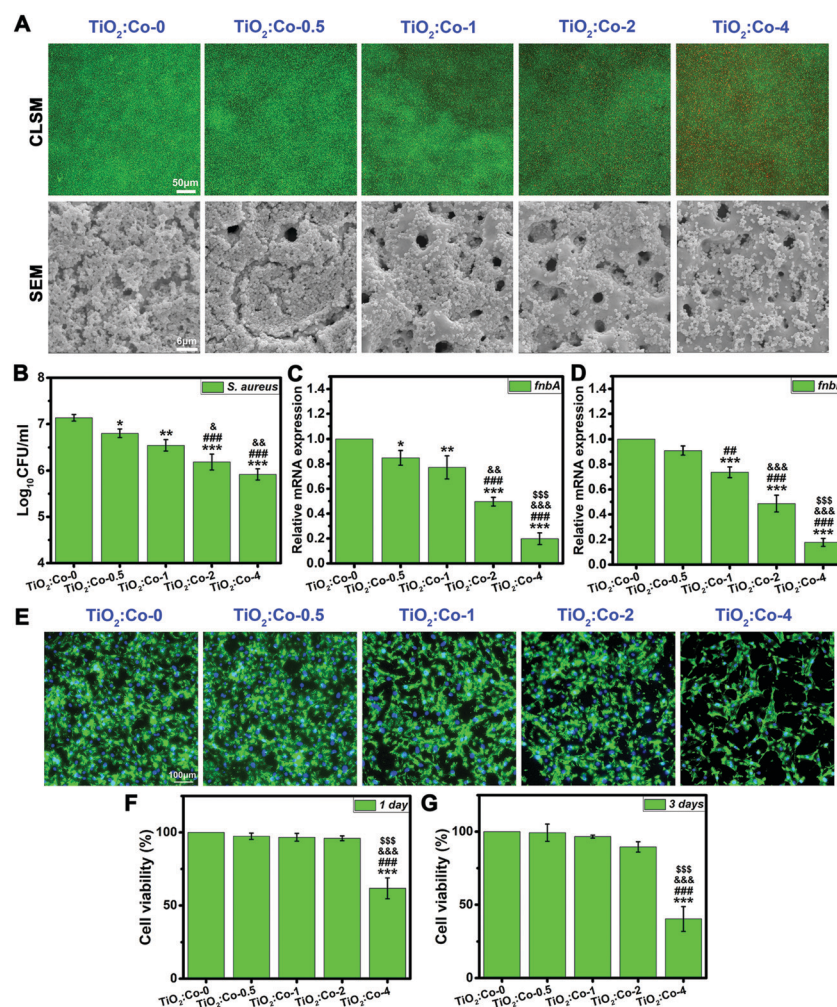
Fig. 2 The analysis of the band structure of the coatings and the redox couples. (A) UV-Vis diffuse reflectance spectra of the coatings. (B) The Kubelka–Munk transformed reflectance spectra from (A). (C) Distribution of the CB minimum ( $E_C$ , red line), Fermi level ( $E_F$ , green line), and VB maximum ( $E_V$ , blue line) of the coatings relative to the standard levels (dotted lines) of hydrogen production ( $4\text{H}^+ + 4\text{e}^- \rightarrow 2\text{H}_2(\text{g})$ ,  $E = -4.44$  eV) and water oxidation ( $2\text{H}_2\text{O} + 4\text{h}^+ \rightarrow \text{O}_2(\text{g}) + 4\text{H}^+$ ,  $E = -5.67$  eV). Please notice that the light blue area shows the range of biological redox potentials (BRP,  $-4.12$  to  $-4.84$  eV) resulting from (D). (D) Demonstration of the sequential redox couples that contribute to the extracellular electron transfer (EET) of bacteria in the BRP range of  $-4.12$  to  $-4.84$  eV.



of standard hydrogen electrode ( $E^\circ$ , V) and absolute potential level ( $E$ , eV) as:  $E^\circ = -4.44 - E$ ,<sup>27</sup> in the range of  $-4.12$  to  $-4.84$  eV. The BRP range is also indicated in Fig. 2C (light blue region). Interestingly, the CB minimum decreased dramatically from overlap of  $\text{TiO}_2\text{:Co-0}$  ( $-4.36$  eV) with the BRP range ( $-4.12$  to  $-4.84$  eV) to levels of  $\text{TiO}_2\text{:Co-2}$  ( $-5.63$  eV) and  $\text{TiO}_2\text{:Co-4}$  ( $-6.03$  eV) far below the BRP minimum ( $-4.84$  eV), which are able to allow the doped samples to accept extracellular electrons from bacterial redox couples, just as served by  $\text{Fe}(\text{OH})_3/\text{Fe}^{2+}$  ( $-5.5$  eV),  $\text{FeOOH}/\text{Fe}^{2+}$  ( $-5.11$  eV),  $\text{Fe}_2\text{O}_3/\text{Fe}^{2+}$  ( $-5.1$  eV), and  $\text{MnO}_2/\text{Mn}^{2+}$  ( $-5.73$  eV).

To test our hypothesis, we first explored the *in vitro* antibacterial performance of doped samples. As shown in Fig. 3A, the viability of adhering MRSA (methicillin-resistant *Staphylococcus aureus*) was visualized by live/dead fluorescence staining after being cultured on samples for 24 h. The green fluorescence intensity for live MRSA sharply decreased with incremental Co

doping content, demonstrating the significant inhibition effect on bacterial biofilm formation; meanwhile, the red fluorescence intensity for dead MRSA increased clearly, thereby indicating the apparent loss of MRSA viability when in contact with doped samples (especially on  $\text{TiO}_2\text{:Co-2}$  and  $\text{TiO}_2\text{:Co-4}$ ). The morphology of adhering MRSA and their biofilm production on samples were further examined by SEM. A large number of MRSA were adhering on  $\text{TiO}_2\text{:Co-0}$  with a continuous dense biofilm totally covering the surface micropores. By contrast, the number of adhering MRSA was significantly reduced on the doped samples upon increasing the Co content, thereby showing the sharply inhibited biofilm formation (especially on  $\text{TiO}_2\text{:Co-2}$  and  $\text{TiO}_2\text{:Co-4}$ ). Hence, the SEM observations agreed well with the live/dead staining results. To quantify the number of adhering MRSA on the samples, the spread plate method was utilized and the average  $\text{Log}_{10}\text{CFU}$  per ml values (Fig. 3B) were in the order of



**Fig. 3** *In vitro* antibacterial effect and biocompatibility. (A) Bacterial viability and biofilm formation on the coatings examined by live/dead fluorescence staining and SEM methods after overnight culture. (B) Bacteria colony-forming units (CFUs) recovered from the coatings. (C and D) The effect of the coatings on the biofilm genes *fnbA* (C) and *fnbB* (D) transcription by qRT-PCR, normalized to the 16S rRNA gene. (E) The fluorescence staining images of the actin cytoskeleton (green) and cell nuclei (blue) of the rBMSCs on the coatings at 24 h post-culture. (F and G) The cellular viability results of the rBMSCs on the coatings at 1 day (F) and 3 days (G) post-culture using CCK-8 assay. Significant differences for all pairwise comparisons are indicated by the symbols over the bars: \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$  vs.  $\text{TiO}_2\text{:Co-0}$ ; ## $P < 0.01$  and ### $P < 0.001$  vs.  $\text{TiO}_2\text{:Co-0.5}$ ;  $^bP < 0.05$ ,  $^{bb}P < 0.01$ , and  $^{bbb}P < 0.001$  vs.  $\text{TiO}_2\text{:Co-1}$ ;  $^{SS}P < 0.001$  vs.  $\text{TiO}_2\text{:Co-2}$ .



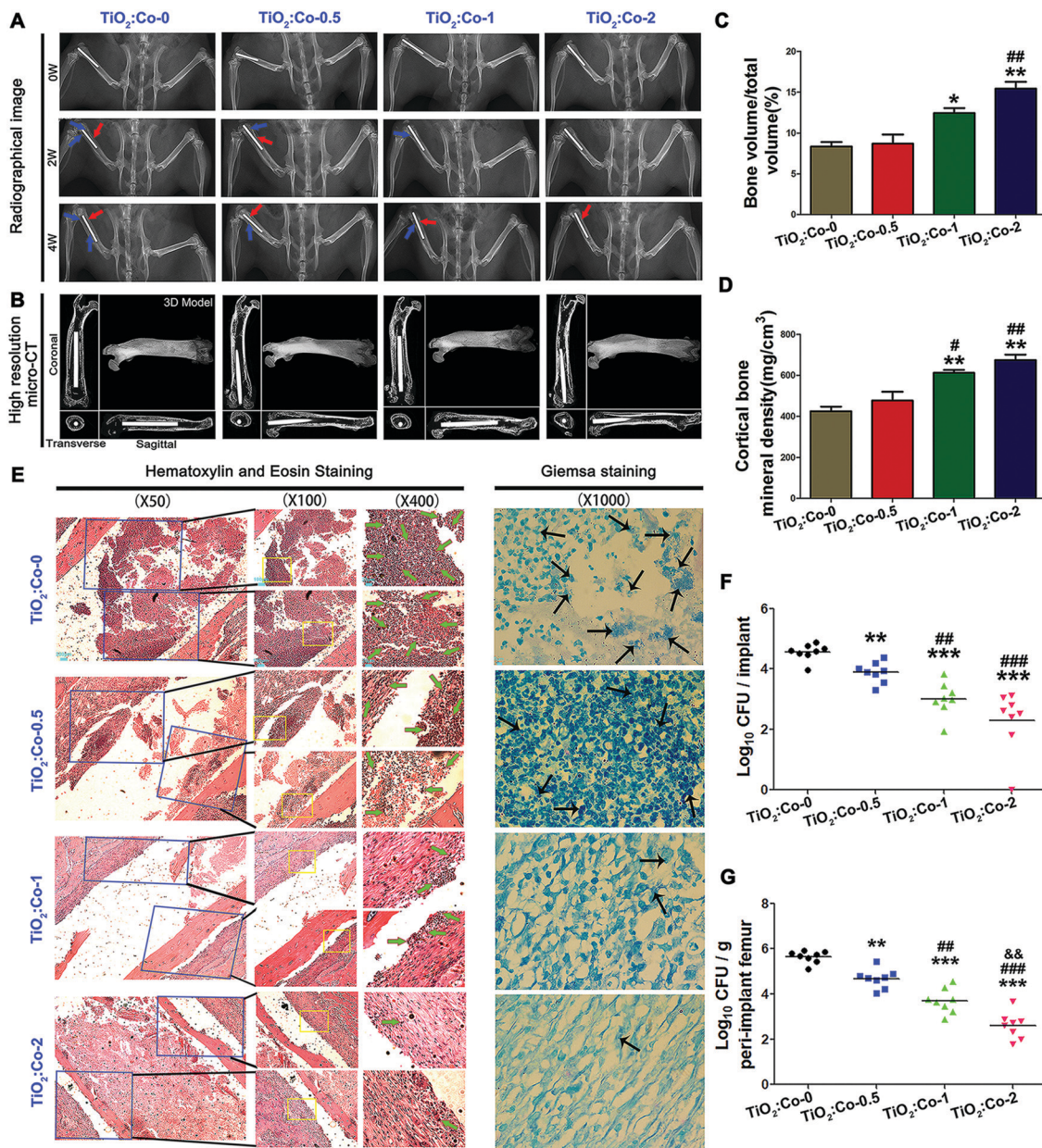


TiO<sub>2</sub>:Co-0 ( $7.13 \pm 0.07$ ) > TiO<sub>2</sub>:Co-0.5 ( $6.80 \pm 0.09$ ) > TiO<sub>2</sub>:Co-1 ( $6.55 \pm 0.12$ ) > TiO<sub>2</sub>:Co-2 ( $6.19 \pm 0.17$ ) > TiO<sub>2</sub>:Co-4 ( $5.92 \pm 0.12$ ). Even at a low Co doping content, TiO<sub>2</sub>:Co-0.5 has shown good anti-biofilm ability compared to TiO<sub>2</sub>:Co-0 ( $P < 0.05$ ). Further increasing the Co content can endow the doped samples with better anti-biofilm outcomes ( $P < 0.01$ ). Noticeably, TiO<sub>2</sub>:Co-2 and TiO<sub>2</sub>:Co-4 exhibited a more powerful anti-biofilm effect when compared with TiO<sub>2</sub>:Co-0 and TiO<sub>2</sub>:Co-0.5 ( $P < 0.001$ ) and TiO<sub>2</sub>:Co-1 ( $P < 0.05$ ). In addition, broad-spectrum biofilm inhibition capability was ascertained by using *Staphylococcus epidermidis* (*S. epidermidis*, Fig. S10–S12, ESI†), *Pseudomonas aeruginosa* (*P. aeruginosa*, Fig. S13–S15, ESI†) and *Escherichia coli* (*E. coli*, Fig. S16–S18, ESI†). We sought to establish whether doped samples could modulate the expression levels of biofilm-related genes to block biofilm formation and development. By virtue of quantitative polymerase chain reaction (qPCR) analysis, the expression levels of *fmbA* and *fmbB* genes participating in MRSA biofilm formation are shown in Fig. 3C and D. TiO<sub>2</sub>:Co-0.5 can effectively inhibit the *fmbA* gene expression ( $P < 0.05$ ), which was further remarkably downregulated on doped samples ( $P < 0.01$ ) with incremental Co content. Regarding TiO<sub>2</sub>:Co-2 and TiO<sub>2</sub>:Co-4, the statistical difference even ascended to a value of  $P < 0.001$  compared with other groups, especially for the *fmbB* gene expression. Similar results were also obtained by analyzing the expression levels of biofilm-related genes *icaA* and *icaR* for *S. epidermidis* (Fig. S19, ESI†). Therefore, it is concluded that the expression inhibition of biofilm-related genes resulted in the blocked biofilm formation on the doped samples. Prior to *in vivo* study, the biocompatibility was evaluated *in vitro* on rat bone mesenchymal stem cells (rBMSCs). Fig. 3E displays the fluorescence staining images of the actin cytoskeleton (green) and cell nuclei (blue) of rBMSCs after 24 h culture on the samples. The rBMSCs can adhere and spread very well with polygonal morphology and abundant actin on TiO<sub>2</sub>:Co-0, TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2; however, apparently fewer rBMSCs can adhere on TiO<sub>2</sub>:Co-4. Fig. 3F and G show the cell viability results using a Cell Counting Kit-8 (CCK-8) assay. At day 1 and 3 of culture, the rBMSCs maintained high viability on TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2 with negligible cytotoxicity. Nevertheless, TiO<sub>2</sub>:Co-4 caused significant toxicity to rBMSCs in comparison with other groups ( $P < 0.001$ ). SEM observations also showed that, apart from TiO<sub>2</sub>:Co-4, the majority of sample surfaces (even micropores) were covered by the spreading rBMSCs (Fig. S20, ESI†). These concordant results indicated the excellent biocompatibility of the doped samples except TiO<sub>2</sub>:Co-4. In accordance with the endosymbiotic theory (symbiogenesis), eukaryotic mitochondrion is an analogue of prokaryote bacteria. However, the electron transport chain of mammalian cells for respiration is located in the intracellular mitochondria. As a result, the electron transport chain is not interrupted and the mammalian cells can survive on the doped samples. In fact, previous work showed that a low content of Co<sup>2+</sup> (<5 at%) incorporation into a mesoporous bioactive glass (MBG) scaffold had no significant cytotoxicity and dramatically promoted angiogenesis and osteogenesis,<sup>28</sup> which supported our work very well since the

Co doping contents of TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1, TiO<sub>2</sub>:Co-2 and TiO<sub>2</sub>:Co-4 were 1.91, 2.45, 4.07 and 6.03 at%, respectively (Table S2, ESI†).

After confirming the favorable biocompatibility of TiO<sub>2</sub>:Co-0, TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2, we next explored their *in vivo* anti-infectious performance. A femoral osteomyelitis model was established on rats using MRSA. Fig. 4A shows the X-ray images of femurs. At week 0 (the 2nd day post surgery), there were no radiographic signs of osteolysis or periosteal reaction detected in each group. Nevertheless, at week 2, we detected the apparent radiographic signs of osseous destruction (blue arrow) and periosteal reaction (red arrow) around TiO<sub>2</sub>:Co-0 and the slight situations for TiO<sub>2</sub>:Co-0.5, while little osteolytic lesions were detected around TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2. At week 4 post surgery, the osteolysis exacerbation and periosteal reaction progress indicated the development of bacterial infection around TiO<sub>2</sub>:Co-0, while other groups especially TiO<sub>2</sub>:Co-2 showed relatively mild osseous destruction or periosteal reaction. Micro-CT analyses were further performed at week 4 post surgery to validate the radiographic results. As seen in Fig. 4B, TiO<sub>2</sub>:Co-0 showed evident periprosthetic osteolysis and porous change throughout the femur. Although TiO<sub>2</sub>:Co-1 displayed slight loosening and femur porous change, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2 demonstrated beneficial osseointegration and cortical integrity. Further supported by the quantitative results of bone volume/total volume (Fig. 4C) and cortical bone mineral density (Fig. 4D), they showed the ascending trend in order of TiO<sub>2</sub>:Co-0, TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2. Compared with TiO<sub>2</sub>:Co-0 and TiO<sub>2</sub>:Co-0.5, TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2 can produce the statistical differences of  $P < 0.05$  and  $P < 0.01$ , respectively. To visualize femur histopathological change and bacterial residue, hematoxylin and eosin (H&E) staining and Giemsa staining were conducted as shown in Fig. 4E. We observed from H&E staining for TiO<sub>2</sub>:Co-0 conspicuous cancellous bone destruction, intra-medullary abscess and inflammatory cell infiltration at the bone-implant interface, as the typical indicators of femur infection; meanwhile, Giemsa staining indicated many bacterial residues in the intramedullary tissue. By contrast, TiO<sub>2</sub>:Co-0.5 displayed slight bone destruction, moderate abscess lesion and less inflammatory cell infiltration as well as decreased bacterial residues. Regarding TiO<sub>2</sub>:Co-1 and especially TiO<sub>2</sub>:Co-2, they can contribute to the mildest bone infection and fewest bacterial residues. Fig. 4F shows the number of bacterial colonies dislodged from implants in the order of TiO<sub>2</sub>:Co-2 < TiO<sub>2</sub>:Co-1 < TiO<sub>2</sub>:Co-0.5 < TiO<sub>2</sub>:Co-0 with a statistical difference among groups of  $P < 0.01$ . For TiO<sub>2</sub>:Co-2, the difference can increase to  $P < 0.001$  versus TiO<sub>2</sub>:Co-0.5 or TiO<sub>2</sub>:Co-0. Therefore, with Co content increase, doped samples can significantly decrease the adhering bacterial burden. We also assayed the bacterial burden from powder bone of each group and obtained a similar trend of CFU number in the femur (Fig. 4G). TiO<sub>2</sub>:Co-2 can even exhibit a statistical difference of  $P < 0.01$  versus TiO<sub>2</sub>:Co-1. Hence, Co doped samples can also exert a powerful inhibition effect on planktonic bacteria in peri-implant femur. Besides, although the mechanical properties of the coating samples decreased slightly after the *in vivo* study,





**Fig. 4** *In vivo* antibacterial effect using a rat osteomyelitis model. (A) Radiological images of the rat femurs at 0, 2, and 4 weeks post surgery. The red arrow and blue arrow show the periosteal reaction and osteolytic lesions, respectively. (B) The micro-CT reconstruction images of the rat femurs at the time of sacrifice. (C and D) The bone volume/total volume (C) and the cortical bone mineral density (D) of the rat femurs. (E) Hematoxylin and eosin staining and Giemsa staining images of the infected femur slices. The green arrow shows the neutrophil infiltration, and the black arrow shows the bacterial residues. (F and G) Bacteria colony-forming units (CFUs) recovered from the explanted implant (F) and from the infected femur (G). Significant differences for all pairwise comparisons are indicated by the symbols over the bars: \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$  vs. TiO<sub>2</sub>:Co-0; # $P < 0.05$ , ## $P < 0.01$ , and ### $P < 0.001$  vs. TiO<sub>2</sub>:Co-0.5; §§ $P < 0.01$  vs. TiO<sub>2</sub>:Co-1.

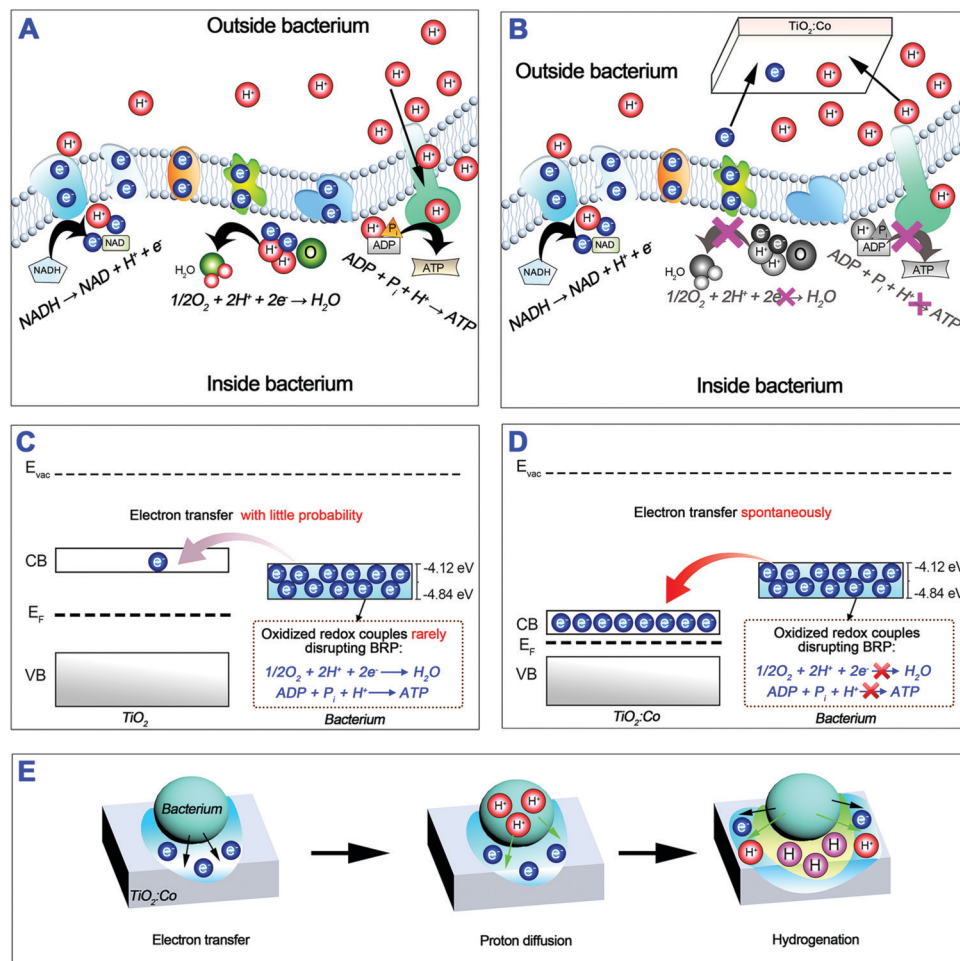
they can maintain good mechanical properties in the *in vivo* environment (Fig. S33 and S34, ESI†).

Encouraged by the excellent *in vivo* anti-infectious outcome, we further tested whether Co doped samples can combat planktonic bacteria *in vitro*. As verified in the ESI†, doped samples have much more inferior inhibition capability on planktonic MRSA (Fig. S21 and S22, ESI†), *S. epidermidis* (Fig. S23 and S24, ESI†), *P. aeruginosa* (Fig. S25 and S26, ESI†) and *E. coli* (Fig. S27 and S28, ESI†). Although a very low concentration of Co

ions can be released from the doped samples (<3 ppm within 8 weeks, Fig. S29, ESI†), the Co element itself has no strong antibacterial property<sup>29–31</sup> and Co ions released from the MBG scaffold (~17.5 ppm within 1 week) did not show any antibacterial effect and had to load ampicillin to kill bacteria.<sup>28</sup> Therefore, some other mechanism should account for the excellent suppressive effect on planktonic bacteria *in vivo*. We thus conclude that the anti-biofilm capability of doped samples is irrelevant to the release of Co ions, but strongly dependent







**Scheme 1** Schematic diagram for elucidating the mechanism of band-structure-tailorable TiO<sub>2</sub> semiconductor coating to thwart bacterial biofilm formation through acting as an extracellular electron acceptor. (A) The interaction of TiO<sub>2</sub> coating with a bacterium, showing that the bacterium can normally implement extracellular electron transfer (EET) and proton transfer for energy metabolism through respiration. (B) The interaction of TiO<sub>2</sub>:Co coating with a bacterium, showing that TiO<sub>2</sub>:Co can serve as an extracellular electron acceptor to interrupt the EET and proton transfer, thereby leading to energy starvation and biofilm formation inhibition. (C and D) The energy band diagrams showing the interactions of TiO<sub>2</sub>-bacterium (C) and TiO<sub>2</sub>:Co-bacterium (D) upon contact. In (C), TiO<sub>2</sub> can rarely disrupt the EET and biological redox potential (BRP). On the contrary, in (D), TiO<sub>2</sub>:Co can spontaneously disrupt the EET and BRP, thereby leading to electron leakage. (E) The proposed hydrogenation process at the TiO<sub>2</sub>:Co-bacterium interface. First, the electrons are transferred from a bacterium and accumulate in the titania layer when the bacterium contacts TiO<sub>2</sub>:Co (electron transfer). Then the protons surrounding the bacterium membrane penetrate into the titania layer to meet the excess electrons (proton diffusion). Therefore, this can finally lead to the generation of a hydrogenated surface titania layer (hydrogenation).

on the tailorable band structure of TiO<sub>2</sub> by Co doping. Scheme 1A illustrates the extracellular electron transfer process along the bacterial membrane respiration chain through a sequence of highly redox-active transmembrane protein complexes (Fig. 2D and Table S4, ESI†). It is reasonable that electron leakage from the electron transfer chain should endanger bacterial energy metabolism and life activity. From the biomimetic viewpoint, Co doped TiO<sub>2</sub> can serve as an extracellular electron acceptor like natural redox-active Fe(III) and Mn(IV) minerals. Upon physical contact with a bacterium, doped TiO<sub>2</sub> can extract electrons from the respiration chain by acting as an electron sink or trap to “short-circuit” the EET chain, as illustrated in Scheme 1B; such extraction is destructive and energetically favorable driven by redox potentials.<sup>32,33</sup> To further clarify this process, we draw the energy band diagram of the interaction of the bacterial EET chain

with undoped TiO<sub>2</sub> (Scheme 1C) and Co doped TiO<sub>2</sub> (Scheme 1D). As a result of the overlap of the CB minimum with the BRP range, undoped TiO<sub>2</sub> can hardly extract electrons from the EET chain and perturb BRP. In contrast, Co doped TiO<sub>2</sub> can spontaneously extract electrons from the EET chain and disrupt BRP, thereby interrupting the EET chain. And as verified herein, the larger the gap between the CB minimum and BRP minimum is, the more powerful the interface electron trap and the EET chain interruption will be. Parallel to the EET process, extracellular proton transfer can generate proton motive force across the bacterial membrane; these protons will re-enter the bacterium interior through adenosine triphosphate (ATP) synthase and the transmembrane proton movement will release energy to synthesize ATP (Scheme 1A).<sup>34</sup> In this context, the interruption of bacterial proton transfer can hinder the ATP synthesis.

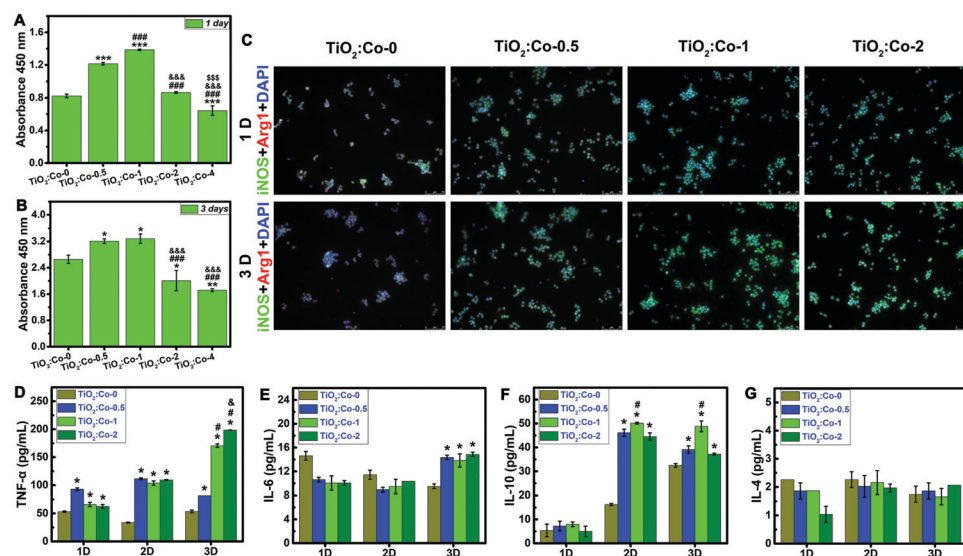




It is well known that the  $\text{TiO}_2$  semiconductor has been widely used for water splitting and hydrogen ( $\text{H}_2$ ) production.<sup>35,36</sup> Hydrogen can strongly influence the structural and electronic properties of a wide range of as-doped host materials including semiconductors, insulators and even aqueous solutions.<sup>37</sup> Considering the fact of the electrostatic attraction between electrons and protons, the extracted electrons by  $\text{TiO}_2\text{:Co}$  from a bacterium will accumulate in the titania layer (electron transfer); such excess electrons will thereafter electrostatically attract the protons surrounding the bacterial membrane especially ATP synthase to penetrate into the titania layer (proton diffusion), thereby generating a hydrogenated surface titania layer (hydrogenation). A recent study showed experimentally and theoretically that a self-hydrogenated shell was produced first on the  $\text{TiO}_2$  surface before forming  $\text{H}_2$  bubbles; this shell was generated through the subsurface diffusion of photo-reduced water protons at the aqueous  $\text{TiO}_2$  interface and can improve  $\text{H}_2$  production by lowering the activation barrier of H–H bond formation.<sup>38</sup> Excess electrons at the aqueous anatase  $\text{TiO}_2(101)$  interface are able to trigger water splitting and trap protons into a stable hydrogenated surface.<sup>39</sup> Interestingly, as shown in Fig. 1B, the designed coating mainly contained anatase  $\text{TiO}_2$  phase with a large percentage of (101) facet. For a spontaneous photoelectrochemical water splitting process, the oxygen and hydrogen reactions must lie between the CBM and VBM of the  $\text{TiO}_2$  semiconductor, *i.e.* having a CBM higher than the  $\text{H}_2/\text{H}_2\text{O}$  level (−4.44 eV) and a VBM lower than the  $\text{H}_2\text{O}/\text{O}_2$  level (−5.67 eV).<sup>35,36</sup> We have indicated the standard levels of  $\text{H}_2$  production and  $\text{H}_2\text{O}$  oxidation in Fig. 2C (dotted lines). Except  $\text{TiO}_2\text{:Co-0}$ , the CBM of doped  $\text{TiO}_2$  is increasingly lower than

the  $\text{H}_2/\text{H}_2\text{O}$  level with incremental Co doping content. Therefore,  $\text{H}_2$  production is theoretically impossible at the bacterium- $\text{TiO}_2\text{:Co}$  interface, and we have verified this by gas chromatography-mass spectrometry analysis. Since hydrogenation can enhance the electrical conductivity of  $\text{TiO}_2\text{:Co}$  coating,<sup>40</sup> this will further facilitate the electron transfer to the adjacent coating and subsequent proton diffusion. The proposed hydrogenation process can be illustrated in Scheme 1E. As a consequence, Co doping can tailor the band structure of  $\text{TiO}_2$  and interrupt the extracellular electron transfer and proton transfer; this will impede bacterial ATP synthesis and energy metabolism, thus contributing to the inhibited biofilm formation on the  $\text{TiO}_2\text{:Co}$  coating.

Due to the excellent repression effect on planktonic bacteria *in vivo* and the inconsistency *in vitro*, we then sought to assess whether innate immune cells (macrophages and neutrophils) could be involved in eradicating bacteria. Fig. 5A and B shows the cellular viability results of RAW 264.7 macrophages using CCK-8 assay. At day 1 and 3 of culture, RAW 264.7 sustained higher viability on  $\text{TiO}_2\text{:Co-0.5}$  and  $\text{TiO}_2\text{:Co-1}$  than  $\text{TiO}_2\text{:Co-0}$ , and  $\text{TiO}_2\text{:Co-2}$  had no significant cytotoxicity. However,  $\text{TiO}_2\text{:Co-4}$  exerted apparent cytotoxicity to RAW 264.7 at day 1 ( $P < 0.001$ ) and day 3 ( $P < 0.01$ ). Fig. 5C indicates the labelling of the activated macrophages using immunofluorescence staining (M1, green for iNOS positive; M2, red for Arg1 positive). Incremental activated M1 macrophages were observed in  $\text{TiO}_2\text{:Co}$  coating at day 1 and 3, and the proportion of M1 macrophages increased with Co doping content (Fig. 5C and Fig. S30, S31, ESI†), revealing that released  $\text{Co}^{2+}$  ions can boost the activation of M1 macrophages in a dose-dependent manner.<sup>41</sup>



**Fig. 5** Cell viability, activation, and cytokine secretion of the RAW264.7 macrophages cultured on the coatings. (A and B) The cellular viability results of the RAW264.7 macrophages on the coatings at 1 day (A) and 3 days (B) post-culture using CCK-8 assay. (C) The immunofluorescent staining images of the RAW 264.7 macrophages scratched on the coatings at 1 and 3 days post-culture. The green fluorescence shows iNOS positive for M1 phenotype macrophages, the red fluorescence shows Arg1 positive for M2 phenotype macrophages, and the blue fluorescence shows cell nuclei. (D–G) The concentrations of the cytokines TNF- $\alpha$  (D), IL-6 (E), IL-10 (F), and IL-4 (G) secreted by the RAW 264.7 macrophages on the coatings using ELISA at 1, 2, and 3 days post-culture. Significant differences for all pairwise comparisons are indicated by the symbols over the bars: \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$  vs.  $\text{TiO}_2\text{:Co-0}$ ; # $P < 0.05$  and ### $P < 0.001$  vs.  $\text{TiO}_2\text{:Co-0.5}$ ;  $^{\circ}P < 0.05$  and  $^{\circ\circ\circ}P < 0.001$  vs.  $\text{TiO}_2\text{:Co-1}$ ;  $^{\circ\circ\circ}P < 0.001$  vs.  $\text{TiO}_2\text{:Co-2}$ .



Fig. 5D–G give the concentrations of TNF- $\alpha$ , IL-6, IL-10, and IL-4 cytokines in the supernatants using ELISA at day 1, 2, and 3. Overall, the secretion of inflammatory cytokines (TNF- $\alpha$  and IL-6) augmented with Co<sup>2+</sup> release of TiO<sub>2</sub>:Co coating, consistent with the immunofluorescence staining results. Interestingly, the secretion of antiinflammatory cytokines (IL-10) also augmented on TiO<sub>2</sub>:Co coating, which can antagonize proinflammatory cytokines and prevent uncontrolled inflammatory response.<sup>42</sup> Therefore, overall TiO<sub>2</sub>:Co coating can trigger a local inflammatory response. Highly plastic macrophages can respond to external stimuli and display the classically activated proinflammatory M1 and alternatively activated antiinflammatory M2 phenotypes.<sup>43</sup> M1 macrophages can secrete inflammatory cytokines to combat bacterial infection. However, the biofilms formed at the infection site can induce macrophages polarizing to M2 phenotype, thereby leading to an immunosuppressive environment, impairing phagocyte influx, and restraining bacterial clearance.<sup>9,44</sup> In this context, developing an immunomodulatory biomaterial surface that can activate M1 macrophages could be promising for biomaterial-centered infection therapy.

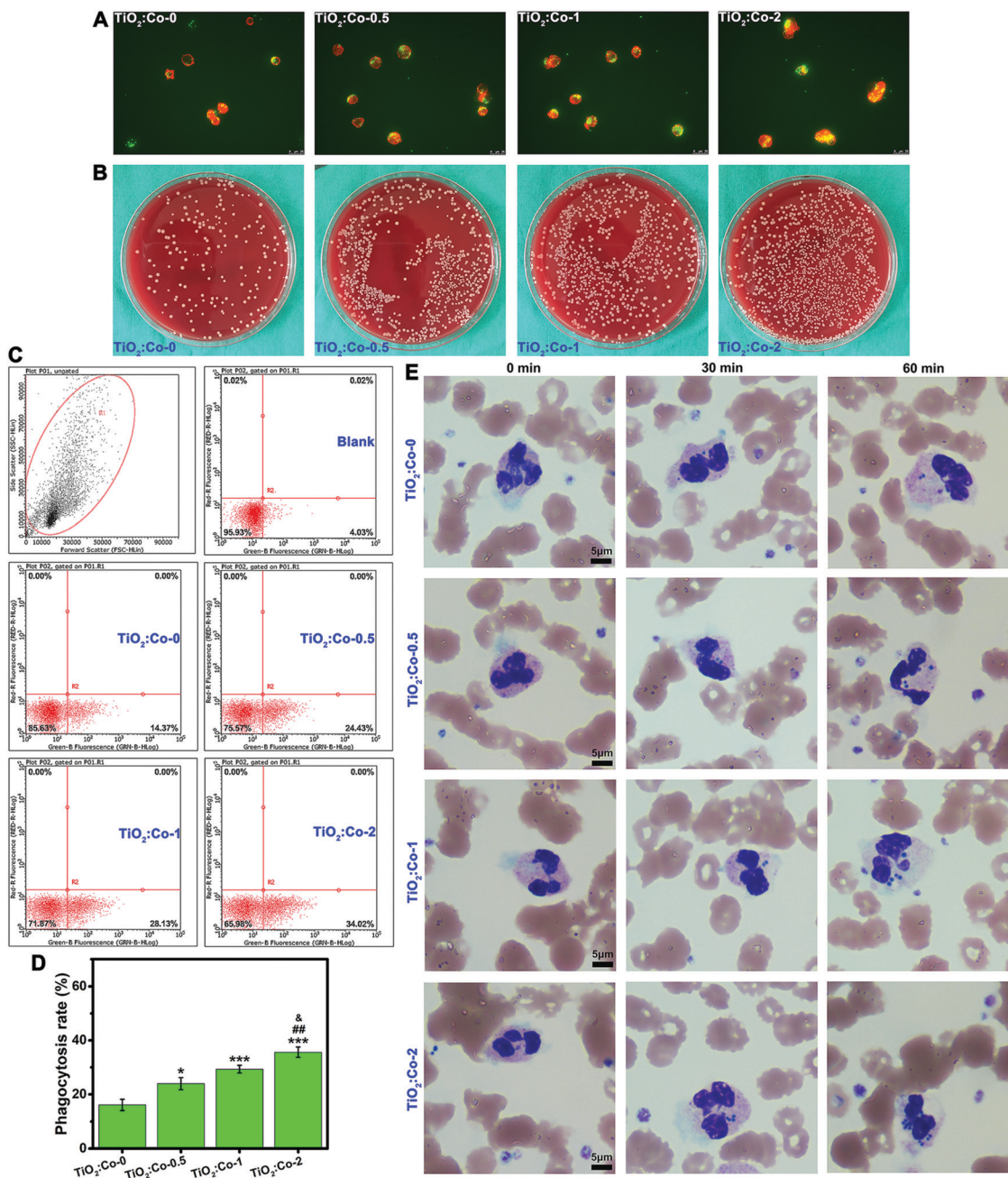
To test if creating such an inflammatory microenvironment could be beneficial for bacterial clearance, we first evaluated the phagocytosis of MRSA by sample-conditioned macrophages. Fig. 6A and Fig. S32 (ESI†) show the fluorescence staining images of phagocytosed MRSA (green) in macrophages (red for cytoskeleton), indicating that released Co<sup>2+</sup> ions from TiO<sub>2</sub>:Co coating can facilitate more phagocytosed MRSA in macrophages in a dose-dependent manner. Fig. 6B gives the representative SPM images of CFU of phagocytosed MRSA in macrophages, revealing the following order: TiO<sub>2</sub>:Co-0 < TiO<sub>2</sub>:Co-0.5 < TiO<sub>2</sub>:Co-1 < TiO<sub>2</sub>:Co-2. Fig. 6C depicts the MRSA phagocytosis by sample-conditioned macrophages using flow cytometry, and the phagocytosis rates of MRSA by macrophages were in the following order: TiO<sub>2</sub>:Co-0 (16.12  $\pm$  2.03%) < TiO<sub>2</sub>:Co-0.5 (23.98  $\pm$  2.24%) < TiO<sub>2</sub>:Co-1 (29.33  $\pm$  1.42%) < TiO<sub>2</sub>:Co-2 (35.63  $\pm$  1.90%). Therefore, we have demonstrated that TiO<sub>2</sub>:Co coating can skew the majority of macrophages polarizing to M1 phenotype, and enhance the phagocytic ability of macrophages on MRSA. We further assessed the phagocytosis of MRSA in human whole blood by polymorphonuclear leukocytes (*i.e.* neutrophils), the major white blood cells playing an important role in innate immunity.<sup>3</sup> Fig. 6E shows that the number of phagocytosed MRSA in neutrophils increased gradually with the incubation period; furthermore, at each time point, TiO<sub>2</sub>:Co coating contributed in a dose-dependent manner to the stronger phagocytic activity of neutrophils with a larger number of engulfed MRSA. This was in accordance with the MRSA phagocytosis by macrophages. Therefore, it is clear that TiO<sub>2</sub>:Co coating can reinforce the phagocytosis of MRSA by neutrophils in human blood. When phagocytes sense a local infection, they emigrate from blood vessels and migrate actively through tissue to the infection site in a well-controlled fashion. Once arrived, phagocytes avidly phagocytose bacteria and kill them intracellularly by releasing reactive oxygen species (ROS) and bactericidal products contained within granules.<sup>45</sup> In the meantime, bacteria produce toxins *e.g.* leukocidin to combat phagocytes.<sup>3</sup> Therefore, an increase

in the capability to phagocytose or kill bacteria will contribute to a reduced risk of infection. Typically, phagocytes show the normal respiratory burst and produce the bactericidal agents of superoxide ( $\bullet\text{O}_2^-$ ) and hydrogen peroxide ( $\text{H}_2\text{O}_2$ ).<sup>46</sup> Cobalt is a redox-active element and the standard redox potential ( $E^\circ$ ) of Co<sup>3+</sup>/Co<sup>2+</sup> is 1.808 V. Considering the standard redox potential of  $\text{H}_2\text{O}_2/\bullet\text{OH}$  (0.38 V), according to the relationship between the thermodynamic function and redox potential:  $\Delta G = -nFE = -nF [E^\circ(\text{Co}^{3+}/\text{Co}^{2+}) - E^\circ(\text{H}_2\text{O}_2/\bullet\text{OH})]$ , where  $\Delta G$ ,  $F$  and  $E$  are the free energy change, Faraday constant and redox potential, respectively; obviously,  $\Delta G$  is < 0, which confirms the spontaneity that Co<sup>2+</sup> can elicit the generation of highly toxic hydroxyl radicals ( $\bullet\text{OH}$ ) *via* the Fenton-like reaction:  $\text{Co}^{2+} + \text{H}_2\text{O}_2 \rightarrow \text{Co}^{3+} + \text{OH}^- + \bullet\text{OH}$ . This is similar to the Fenton reaction of ferrous ions ( $\text{Fe}^{2+}$ ) with  $E^\circ(\text{Fe}^{3+}/\text{Fe}^{2+}) = 0.771$  V to induce  $\bullet\text{OH}$  generation.<sup>47</sup> As a result, the generated  $\bullet\text{O}_2^-$  and  $\bullet\text{OH}$  radicals can enhance the ability of phagocytes to kill bacteria.

Bacteria form biofilms to resist leukocytes due to a lack of penetration of leukocytes into bacterial biofilms and a reduced capability of phagocytes to kill biofilm-encased bacteria. A mature biofilm has a dense polymeric matrix that is difficult to engulf by phagocytes, which leads to the ‘frustrated phagocytosis’.<sup>9,48</sup> The biofilms can skew the host innate immune response from a proinflammatory bactericidal response to an antiinflammatory profibrotic response,<sup>9</sup> and shift macrophage polarization from the M1 to M2 phenotype.<sup>49</sup> Therefore, using the biofilm formation strategy, bacteria can evade host innate immunity. We have validated that TiO<sub>2</sub>:Co coating can block the formation of bacterial biofilms. It is thus reasonable to speculate that phagocytes (*e.g.* macrophages, neutrophils) can phagocytose and kill the biofilm-formation-blocked bacteria using a divide-and-conquer strategy because in this situation biofilms cannot well protect bacteria from phagocytes. When bacterial infection occurs, innate immune cells, particularly macrophages, will be first recruited to the site of infection and polarize to the proinflammatory M1 phenotype, secrete cytokines to boost inflammation reaction, and then phagocytose and kill bacteria.<sup>50</sup> We have verified that the majority of macrophages in the TiO<sub>2</sub>:Co coating can be activated and polarize to the M1 phenotype, and then secrete a high level of inflammatory cytokines to create a proinflammatory microenvironment; the TiO<sub>2</sub>:Co coating can facilitate the MRSA phagocytosis by sample-conditioned macrophages and neutrophils. Therefore, such an immunomodulatory ability of the TiO<sub>2</sub>:Co coating can enhance the capability of phagocytes to phagocytose and kill bacteria, and contribute to the superior antiinfective function. This immunoregulatory antibacterial strategy can be called infection immunotherapy. In fact, cancer immunotherapy has been widely used to inhibit tumor growth.<sup>47,51</sup> The promoted local inflammatory response can recruit more immune cells to combat bacteria; the enhanced phagocytosis of bacteria and release of proinflammatory cytokines by phagocytes can jointly orchestrate local innate immunity and following adaptive immunity to fight infection.<sup>52,53</sup>

On the other hand, it was observed *in vivo* that TiO<sub>2</sub>:Co-1 and TiO<sub>2</sub>:Co-2 showed beneficial osseointegration. As an essential





**Fig. 6** Phagocytosis of bacteria by the sample-conditioned RAW264.7 macrophages and polymorphonuclear leukocytes (neutrophils). (A) The fluorescence staining images of phagocytosis of bacteria by the sample-conditioned RAW264.7 macrophages. The green fluorescence shows phagocytosed bacteria, and the red fluorescence shows macrophages (cytoskeleton). (B) Representative images of phagocytosed bacteria colony-forming units (CFUs) recovered from the RAW264.7 macrophages. (C) The flow cytometry analysis of phagocytosis of labelled bacteria by the sample-conditioned RAW264.7 macrophages. (D) Quantitative results of phagocytosis rate of the RAW264.7 macrophages. (E) Wright–Giemsa staining images of phagocytosis of bacteria in human whole blood using a bacteria-blood-biomaterial co-culture model. The round blue granules show bacteria. Significant differences for all pairwise comparisons are indicated by the symbols over the bars: \* $P < 0.05$  and \*\*\* $P < 0.001$  vs.  $\text{TiO}_2\text{:Co-0}$ ; ## $P < 0.01$  vs.  $\text{TiO}_2\text{:Co-0.5}$ ;  $^bP < 0.05$  vs.  $\text{TiO}_2\text{:Co-1}$ .

element in human physiology, cobalt has been verified to promote osteogenesis and angiogenesis/vascularization.<sup>28,54</sup> Hypoxia-inducible factor (HIF) plays a central role in the cellular adaptation to low oxygen availability. In normoxia, HIF- $\alpha$  is targeted for rapid degradation upon prolyl hydroxylation by HIF prolyl-hydroxylases (PHDs).<sup>55</sup> PHDs need oxygen and act

as primary oxygen sensors in the cell repertoire. Reduced oxygen levels will impair PHDs and stabilize HIF- $\alpha$ .<sup>56,57</sup> Under hypoxic conditions, the hydroxylation of HIF- $\alpha$  can be decreased and lead to the transactivation and induction of HIF- $\alpha$  target genes,<sup>58,59</sup> thereby in favour of neovascularization and bone regeneration in wound healing responses. As a result, inhibition of PHDs can





stabilize HIF- $\alpha$  in bone and wound tissues, enhance neovascularization, and further promote bone repair and wound healing. It has been suggested that the activation of HIF forms the base for ischemic tolerance.<sup>60–62</sup> Cobalt chloride (CoCl<sub>2</sub>) is a hypoxia mimetic agent possessing the capability to upregulate the HIF level by inhibiting PHDs (*i.e.* inhibiting the prolyl hydroxylation of HIF),<sup>57,59</sup> thus having the ability to enhance vascularization and bone formation. Cobalt can induce ischemic tolerance in a variety of tissues and cells, and thereby endow tissues with resistance to subsequent ischemia/hypoxia through HIF activation.<sup>60,63</sup> The hypoxia will increase the expression of pro-angiogenic factors (*e.g.* VEGF) by BMSCs that are able to improve angiogenesis/vascularization in ischemic tissues<sup>64,65</sup> and osteogenesis.<sup>66</sup> Therefore, the observed favorable osseointegration *in vivo* may be attributed to the effect of released Co on the inhibition of PHDs.

## Conclusions

In summary, we devised a novel three-step strategy for biomaterial-centered infection combination therapy based on Co doped TiO<sub>2</sub> semiconductor. The first step is that Co doping can tailor the band structure of TiO<sub>2</sub> to serve as extracellular electron acceptors; in this way TiO<sub>2</sub>:Co can interrupt extracellular electron transfer and proton transfer, and bacterial energy metabolism, thereby able to block bacterial biofilm formation. Secondly, innate immune cells *e.g.* macrophages and neutrophils can adopt divide-and-conquer tactics to annihilate the biofilm-formation-blocked bacteria. In the meantime, TiO<sub>2</sub>:Co can release Co<sup>2+</sup> ions to create a proinflammatory microenvironment, and potentiate the ability of phagocytes to phagocytose and kill bacteria, thus leading to the enhanced antibacterial capability of innate immune cells. We anticipate that this design concept of infection combination therapy can be applied to develop other advanced antibacterial biomaterials and find wide applications in antibacterial research.

## Experimental section

Details on the experimental procedures are provided in the ESI.†

## Conflicts of interest

The authors declare no conflict of interest.

## Acknowledgements

Dr Jinhua Li gratefully acknowledges the funding support from the Alexander von Humboldt Stiftung/Foundation. This work was financially supported by Hong Kong Research Grant Council General Research Funds (CityU 11205617), City University of Hong Kong Donation Research Grant (9220061), and the National Natural Science Foundation of China (81472109).

## References

- 1 C. R. Arciola, D. Campoccia and L. Montanaro, *Nat. Rev. Microbiol.*, 2018, **16**, 397–409.
- 2 J. Liao, W. Chen, M. Yang, J. Zhou, Z. Wang, Y. Zhou, C. Ning and H. Yuan, *Mater. Horiz.*, 2018, **5**, 545–552.
- 3 S. D. Kobayashi, K. R. Braughton, A. R. Whitney, J. M. Voyich, T. G. Schwan, J. M. Musser and F. R. DeLeo, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 10948–10953.
- 4 H. J. Busscher, H. C. van der Mei, G. Subbiahdoss, P. C. Jutte, J. J. A. M. van den Dungen, S. A. J. Zaat, M. J. Schultz and D. W. Grainger, *Sci. Transl. Med.*, 2012, **4**, 153rv110.
- 5 R. Sridharan, A. R. Cameron, D. J. Kelly, C. J. Kearney and F. J. O'Brien, *Mater. Today*, 2015, **18**, 313–325.
- 6 A. Mantovani, A. Sica, S. Sozzani, P. Allavena, A. Vecchi and M. Locati, *Trends Immunol.*, 2004, **25**, 677–686.
- 7 D. M. Mosser and J. P. Edwards, *Nat. Rev. Immunol.*, 2008, **8**, 958–969.
- 8 S. Chen, J. A. Jones, Y. Xu, H.-Y. Low, J. M. Anderson and K. W. Leong, *Biomaterials*, 2010, **31**, 3479–3491.
- 9 L. R. Thurlow, M. L. Hanke, T. Fritz, A. Angle, A. Aldrich, S. H. Williams, I. L. Engebretsen, K. W. Bayles, A. R. Horswill and T. Kielian, *J. Immunol.*, 2011, **186**, 6585–6596.
- 10 C. Nathan, *Nature*, 2002, **420**, 846–852.
- 11 S. H. Light, L. Su, R. Rivera-Lugo, J. A. Cornejo, A. Louie, A. T. Iavarone, C. M. Ajo-Franklin and D. A. Portnoy, *Nature*, 2018, **562**, 140–144.
- 12 R. S. Hartshorne, C. L. Reardon, D. Ross, J. Nuester, T. A. Clarke, A. J. Gates, P. C. Mills, J. K. Fredrickson, J. M. Zachara, L. Shi, A. S. Beliaev, M. J. Marshall, M. Tien, S. Brantley, J. N. Butt and D. J. Richardson, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 22169–22174.
- 13 L. Shi, H. Dong, G. Reguera, H. Beyenal, A. Lu, J. Liu, H.-Q. Yu and J. K. Fredrickson, *Nat. Rev. Microbiol.*, 2016, **14**, 651.
- 14 R. K. Thauer, K. Jungermann and K. Decker, *Bacteriol. Rev.*, 1977, **41**, 100–180.
- 15 C. R. Myers and K. H. Nealson, *Science*, 1988, **240**, 1319.
- 16 K. A. Weber, L. A. Achenbach and J. D. Coates, *Nat. Rev. Microbiol.*, 2006, **4**, 752.
- 17 J. M. Zachara, J. K. Fredrickson, S.-M. Li, D. W. Kennedy, S. C. Smith and P. L. Gassman, *Am. Mineral.*, 1998, **83**, 1426–1443.
- 18 M. Oku and Y. Sato, *Appl. Surf. Sci.*, 1992, **55**, 37–41.
- 19 A. C. Tuan, J. D. Bryan, A. B. Pakhomov, V. Shutthanandan, S. Thevuthasan, D. E. McCready, D. Gaspar, M. H. Engelhard, J. W. Rogers, K. Krishnan, D. R. Gamelin and S. A. Chambers, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2004, **70**, 054424.
- 20 D. H. Hanaor and C. Sorrell, *J. Mater. Sci.*, 2011, **46**, 855–874.
- 21 X. H. Wang, J. G. Li, H. Kamiyama, M. Katada, N. Ohashi, Y. Moriyoshi and T. Ishigaki, *J. Am. Chem. Soc.*, 2005, **127**, 10982–10990.
- 22 J.-G. Li, R. Büchel, M. Isobe, T. Mori and T. Ishigaki, *J. Phys. Chem. C*, 2009, **113**, 8009–8015.
- 23 X. Zhu, J. Chen, L. Scheideler, R. Reichl and J. Geis-Gerstorfer, *Biomaterials*, 2004, **25**, 4087–4103.



- 24 M. Iwasaki, M. Hara, H. Kawada, H. Tada and S. Ito, *J. Colloid Interface Sci.*, 2000, **224**, 202–204.
- 25 L. Zhou, S.-y. Zhang, J.-c. Cheng, L.-d. Zhang and Z. Zeng, *Mater. Sci. Eng., B*, 1997, **49**, 117–122.
- 26 S. George, S. Pokhrel, Z. Ji, B. L. Henderson, T. Xia, L. Li, J. I. Zink, A. E. Nel and L. Mädler, *J. Am. Chem. Soc.*, 2011, **133**, 11270–11278.
- 27 Z. Xuming, C. Yu Lim, L. Ru-Shi and T. Din Ping, *Rep. Prog. Phys.*, 2013, **76**, 046401.
- 28 C. Wu, Y. Zhou, W. Fan, P. Han, J. Chang, J. Yuen, M. Zhang and Y. Xiao, *Biomaterials*, 2012, **33**, 2076–2085.
- 29 K. Boldbaatar, K. Dashnyam, J. C. Knowles, H.-H. Lee, J.-H. Lee and H.-W. Kim, *Acta Biomater.*, 2019, **83**, 322–333.
- 30 J. Li, J. Wang, D. Wang, G. Guo, K. W. K. Yeung, X. Zhang and X. Liu, *ACS Appl. Mater. Interfaces*, 2017, **9**, 27475–27490.
- 31 Q. Shi, X. Luo, Z. Huang, A. C. Midgley, B. Wang, R. Liu, D. Zhi, T. Wei, X. Zhou, M. Qiao, J. Zhang, D. Kong and K. Wang, *Acta Biomater.*, 2019, **86**, 465–479.
- 32 Y. Xu and M. A. A. Schoonen, *Am. Mineral.*, 2000, **85**, 543–556.
- 33 M. T. Greiner, M. G. Helander, W.-M. Tang, Z.-B. Wang, J. Qiu and Z.-H. Lu, *Nat. Mater.*, 2012, **11**, 76–81.
- 34 P. Mitchell, *Biol. Rev.*, 1966, **41**, 445–501.
- 35 A. Fujishima and K. Honda, *Nature*, 1972, **238**, 37.
- 36 S. U. M. Khan, M. Al-Shahry and W. B. Ingler, *Science*, 2002, **297**, 2243.
- 37 C. G. Van de Walle and J. Neugebauer, *Nature*, 2003, **423**, 626.
- 38 Y. Lu, W.-J. Yin, K.-L. Peng, K. Wang, Q. Hu, A. Selloni, F.-R. Chen, L.-M. Liu and M.-L. Sui, *Nat. Commun.*, 2018, **9**, 2752.
- 39 S. Selcuk and A. Selloni, *Nat. Mater.*, 2016, **15**, 1107.
- 40 X. Chen, L. Liu, P. Y. Yu and S. S. Mao, *Science*, 2011, 1200448.
- 41 Z. Chen, J. Yuen, R. Crawford, J. Chang, C. Wu and Y. Xiao, *Biomaterials*, 2015, **61**, 126–138.
- 42 A. Eckhardt, T. Harorli, J. Limtanyakul, K.-A. Hiller, C. Bosl, C. Bolay, F.-X. Reichl, G. Schmalz and H. Schweikl, *Biomaterials*, 2009, **30**, 1665–1674.
- 43 Z. Chen, T. Klein, R. Z. Murray, R. Crawford, J. Chang, C. Wu and Y. Xiao, *Mater. Today*, 2016, **19**, 304–321.
- 44 C. E. Heim, D. Vidlak, T. D. Scherr, C. W. Hartman, K. L. Garvin and T. Kielian, *J. Immunol.*, 2015, **194**, 3861–3872.
- 45 P. Friedl and B. Weigelin, *Nat. Immunol.*, 2008, **9**, 960–969.
- 46 B. M. Babior, *J. Clin. Invest.*, 1984, **73**, 599–601.
- 47 S. Zanganeh, G. Hutter, R. Spitler, O. Lenkov, M. Mahmoudi, A. Shaw, J. S. Pajarinen, H. Nejadnik, S. Goodman, M. Moseley, L. M. Coussens and H. E. Daldrup-Link, *Nat. Nanotechnol.*, 2016, **11**, 986–994.
- 48 T. J. Foster, *Nat. Rev. Microbiol.*, 2005, **3**, 948.
- 49 C. M. Gries and T. Kielian, *J. Am. Acad. Orthop. Surg.*, 2017, **25**(suppl. 1), S20–S24.
- 50 S. L. Brandt, N. E. Putnam, J. E. Cassat and C. H. Serezani, *J. Immunol.*, 2018, **200**, 3871.
- 51 X. Zhang, C. Wang, J. Wang, Q. Hu, B. Langworthy, Y. Ye, W. Sun, J. Lin, T. Wang, J. Fine, H. Cheng, G. Dotti, P. Huang and Z. Gu, *Adv. Mater.*, 2018, **30**, 1707112.
- 52 A. Aderem, *J. Infect. Dis.*, 2003, **187**, S340–S345.
- 53 D. M. Underhill and H. S. Goodridge, *Nat. Rev. Immunol.*, 2012, **12**, 492.
- 54 W. Fan, R. Crawford and Y. Xiao, *Biomaterials*, 2010, **31**, 3580–3589.
- 55 E. Berra, E. Benizri, A. Ginouvès, V. Volmat, D. Roux and J. Pouyssegur, *EMBO J.*, 2003, **22**, 4082.
- 56 R. H. Wenger, *FASEB J.*, 2002, **16**, 1151–1162.
- 57 K. S. Hewitson, L. A. McNeill, M. V. Riordan, Y.-M. Tian, A. N. Bullock, R. W. Welford, J. M. Elkins, N. J. Oldham, S. Bhattacharya, J. M. Gleadle, P. J. Ratcliffe, C. W. Pugh and C. J. Schofield, *J. Biol. Chem.*, 2002, **277**, 26351–26355.
- 58 S. M. Jones, A. E. Novak and J. P. Elliott, *Neuroscience*, 2013, **252**, 420–430.
- 59 E. Pacary, H. Legros, S. Valable, P. Duchatelle, M. Lecocq, E. Petit, O. Nicole and M. Bernaudin, *J. Cell Sci.*, 2006, **119**, 2667.
- 60 M. Bergeron, J. M. Gidday, A. Y. Yu, G. L. Semenza, D. M. Ferriero and F. R. Sharp, *Ann. Neurol.*, 2000, **48**, 285–296.
- 61 N. M. Jones and M. Bergeron, *J. Cereb. Blood Flow Metab.*, 2001, **21**, 1105–1114.
- 62 M. Bernaudin, Y. Tang, M. Reilly, E. Petit and F. R. Sharp, *J. Biol. Chem.*, 2002, **277**, 39728–39738.
- 63 N. M. Jones, L. Kardashyan, J. K. Callaway, E. M. Lee and P. M. Beart, *Pediatr. Res.*, 2008, **63**, 620–624.
- 64 X. Hu, S. P. Yu, J. L. Fraser, Z. Lu, M. E. Ogle, J.-A. Wang and L. Wei, *J. Thorac. Cardiovasc. Surg.*, 2008, **135**, 799–808.
- 65 T.-S. Li, K. Hamano, K. Suzuki, H. Ito, N. Zempo and M. Matsuzaki, *Am. J. Physiol.: Heart Circ. Physiol.*, 2002, **283**, H468–H473.
- 66 D. Kaigler, Z. Wang, K. Horger, D. J. Mooney and P. H. Krebsbach, *J. Bone Miner. Res.*, 2006, **21**, 735–744.

