Michellamines $A_6$ and $A_7$, and further mono- and dimeric naphthylisoquinoline alkaloids from a Congolese Ancistrocladus liana and their antiausterity activities against pancreatic cancer cells†

Blaise Kimbadi Lombe, Doris Feineis, Virima Mudogo, Reto Brun, Suresh Awale and Gerhard Bringmann

Michellamines $A_6$ (1) and $A_7$ (2) are the first dimers of 5,8'-coupled naphthylisoquinoline alkaloids with cis-configured stereocenters in both tetrahydroisoquinoline subunits. They were isolated from the leaves of a recently discovered, yet unidentified Congolese Ancistrocladus liana that shares some morphological characteristics with Ancistrocladus likoko. Two further new dimeric analogs, michellamines $B_4$ (3) and $B_5$ (4), were obtained, along with two previously likewise unknown monomers, ancistrobonsolines $A_1$ (5) and $A_2$ (6), which, besides one single known other example, are the only naphthylhydroisoquinolines with an $M$-configured biaryl axis and $R$-configuration at C-3. Moreover, five compounds earlier reported from other Ancistrocladus species were identified, ancistroealaine C (7), korupensamines A (8a) and B (8b), and michellamines $A_2$ (9) and E (10). Their complete structural elucidation succeeded due to the fruitful interplay of spectroscopic, chemical, and chiroptical methods. Chemotaxonomically, the stereostructures of the metabolites clearly delineate this Congolese Ancistrocladus liana from all known related species, showing that it might be a new taxon. Ancistrobonsolines $A_1$ (5) and $A_2$ (6) exhibited strong preferential cytotoxicities against human PANC-1 pancreatic cancer cells under nutrient-deprived conditions, without displaying toxicity in normal, nutrient-rich medium. Against cervical HeLa cancer cells, the dimeric alkaloids michellamines $A_6$ (1) and E (10) displayed the highest cytotoxic activities, comparable to that of the standard agent, 5-fluorouracil. Furthermore, ancistrobonsolines $A_1$ (5) and $A_2$ (6) showed weak-to-moderate antiprotozoal activities.

Introduction

Plants belonging to the genus Ancistrocladus Wall. (Ancistrocladaceae) continue to receive much attention, mainly because they produce a class of structurally, biosynthetically, and pharmacologically unusual compounds, the naphthylisoquinoline alkaloids. The genus Ancistrocladus comprises 18 botanically accepted species of tropical lianas, of which four occur in the Democratic Republic of the Congo (DRC), namely, $A. ealaensis$ J. Léonard, $A. congolensis$ J. Léonard, $A. likoko$ J. Léonard, and $A. ileboensis$ G. Heubl, V. Mudogo & G. Bringmann. The number of Congolese Ancistrocladus species may, however, be actually much higher, as there have not yet been thorough and specific botanical campaigns covering the entire Congolese forest, which is so vast and so rich in biodiversity. This assumption is also supported by recent phylogenetic investigations on Ancistrocladus samples collected at different sites in DRC, which hinted at the presence of a potentially new taxon from the region near the village Bon-solrieve, close to the town Mbundaka, in the northwestern part of DRC.

Most recently, likewise close to the village Bon-solrieve, we have discovered another Ancistrocladus liana. Particularly striking were its leaves, which were much larger (about 57 cm long and 12 cm wide) than those of any other Ancistrocladus plant species found nearby. This liana had hooked inflorescences, which, together with the aforementioned large size of...
the leaves, were reminiscent of *A. likoko*, the only known Congolese species with similar – yet different – morphological characteristics. Comparative LC-UV-MS profiling of the leaf alkaloid pattern of this liana and that of an authentic sample of *A. likoko*, however, revealed substantial phytochemical differences between the two samples. This metabolic divergence and the fact that *A. likoko* had so far not been found to occur in that sampling area indicated that the liana might belong to a new, as yet undescribed plant species or subspecies. The isolation and structural elucidation of its alkaloids thus became a rewarding task, not only for the chemotaxonomic characterization of this liana and its possible delineation from other *Ancistrocladus* species, but also for their biological evaluation.

Herein we report on the isolation and structural elucidation of eleven mono- and dimeric naphthylisoquinoline alkaloids from the leaves of this yet unidentified Congolese *Ancistrocladus* liana (Fig. 1). The isolated metabolites comprise six previously unknown compounds: michellamines A<sub>6</sub> (1), A<sub>7</sub> (2), B<sub>4</sub> (3), and B<sub>5</sub> (4), ancistrobonsoline A<sub>1</sub> (5) and its 6-O-methyl derivative, ancistrobonsoline A<sub>2</sub> (6), and five analogs described from earlier work on other *Ancistrocladus* plants: ancistroealaine C (7), korupensamines A (8a) and B (8b),<sup>8</sup> and michellamines A<sub>2</sub> (9)<sup>9</sup> and E (10).<sup>9</sup> Furthermore, we discuss the chemotaxonomic position of this liana relative to other *Ancistrocladus* species. The cytotoxic activities of the isolated compounds against cervical HeLa cancer cells and their antiausterity potencies against PANC-1 human pancreatic cancer cells are also described, as well as the antiprotozoal properties of the new monomeric compounds.

**Results and discussion**

**Isolation and structural elucidation of metabolites**

**Identification of the known compounds 7–10.** Extraction of the air-dried leaves with a mixture of methanol and dichloromethane (1 : 1), followed by concentration under reduced pressure, provided a crude residue, which was then submitted to a cation-exchange column to remove undesired, non-basic metabolites. The resulting alkaloid-enriched fraction was partitioned between water and dichloromethane, and monitored by HPLC (Fig. 2).

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**Fig. 1** Metabolites isolated from an as yet unidentified Congolese *Ancistrocladus* liana, among them six new compounds: michellamines A<sub>6</sub> (1), A<sub>7</sub> (2), B<sub>4</sub> (3), and B<sub>5</sub> (4), ancistrobonsolines A<sub>1</sub> (5) and A<sub>2</sub> (6), and five previously known ones: ancistroealaine C (7), korupensamines A (8a) and B (8b), and michellamines A<sub>2</sub> (9) and E (10). Yellow ellipses on 1 and 2 highlight the combination of two 1,3-cis-configurations, never observed in any other related dimer; for reason of comparison, see the structure of the well-known michellamine A (11) with its 1,3-trans-configurations (underlaid in gray) in both molecular halves – this compound is not produced by the investigated Congolese liana.
Resolution of the water-phase subfraction on a preparative HPLC column permitted isolation of nine pure constituents, obtained as amorphous solids. The fastest-eluting compound (Peak A, Fig. 2a) was found to be ancistroealaine C (7), earlier known from Ancistrocladus ealaensis. The second alkaloid (Peak B) was korupensamine A (8a), while peak C contained two alkaloids overlapping each other, korupensamine B (8b) and michellamine A2 (9), and peak G was michellamine E (10). These four metabolites (8a/b–10) had previously been isolated from other Ancistrocladus species, yet only from Central Africa: The korupensamines A (8a) and B (8b), first known from the Cameroonian plant A. korupensis as well as michellamine E (10), had both also been reported from A. likoko, where they occur as major metabolites. Korupensamine A (8a) is likewise the main naphthylisoquinoline alkaloid in A. congolensis, in which michellamine A2 (9) was discovered before its more recent detection as a minor constituent of A. likoko. The other four remaining compounds from the aqueous subfraction (Peaks D, E, F, and H, Fig. 2a), as well as those from the dichloromethane subfraction (Peaks I and J, Fig. 2b), were as yet unknown.

**Michellamine A6 (1).** The first new metabolite, compound 1 (Peak D, Fig. 2a), had a molecular formula C_{46}H_{48}N_{2}O_{8}, as evidenced from HRESIMS (757.3486, [M + H]+). 1H and 13C NMR measurements showed only a half set of signals, which indicated that this metabolite consisted of two equivalent molecular portions, but left open the question of whether the two halves were homomorphous or enantiomorphous to each other. The latter possibility (i.e., the presence of an achiral meso compound) was excluded by its optical activity, showing that 1 was C_{2}-symmetric.

The 1H NMR spectrum (Table 1) exhibited four aromatic protons. Two of them displayed a meta-coupling, typical of protons at C-1’ and C-3’, while the other two appeared as singlets, which suggested the presence of a symmetric dimer of 5,6’, 5,8’, 7,6’, or 7,8’-coupled naphthylisoquinoline monomers. The aliphatic region, with two three-proton doublets (Me-), 2.27, dd (11.3, 16.9) 33.3, 2.49, dd (11.3, 16.9) 33.4.

**Table 1** 1H and 13C NMR data of michellamine A6 (1), and of ancistrobonsolines A1 (5) and A2 (6)\(^a\)

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\(a\) 1H and 13C NMR data were recorded in methanol-d_{4} (5 in ppm). \(b\) These data are identical for the second molecular half of the C_{2}-symmetric dimer 1.
1 and Me-3), signals for two aryl substituents (a methyl group and a methoxy function), a quartet, a multiplet, and two doublet of doublets, was, however, closely similar to that of the likewise isolated known ancistroealaine C (7), so that a 5,8-coupled molecular framework was expected. In agreement with this assumption, one of the two aromatic singlets was assigned as H-7, from its HMBC correlations with C-6, C-8, and C-9 (Fig. 3a). This showed that the other, remaining singlet had to be at C-7” and, hence, clearly excluded any biaryl linkage involving C-7.

The two remaining possible coupling types (5,6’ or 5,8’) were further discriminated by joint correlative HMBC signals from H-7 and H-1’ to C-8′ (Fig. 3a), which excluded a 5,6’-coupling, where such interactions would not have been observed. Thus, the naphthalene and isoquinoline portions were linked to each other by a 5,8’-axis, and the two monomeric naphthalinoisooquinoline halves were therefore connected via C-6’. These couplings, in particular the 6’,6”-linkage between the naphthalene portions, were further corroborated by the chemical shifts of C-6’ (δc 120.4) and of the surrounding nuclei, C-5’ (δc 152.4) and C-7’ (δc 134.9), which were near-identical to those of other dimers with similar molecular frameworks (e.g., michellamine A (9) δc 120.3 for C-6′, 152.3 for C-5′, and 134.7 for C-7’). The location of the methoxy group at C-4′ was deduced from NOE interactions of its protons with H-3′, in conjunction with joint HMBC correlations from both the O-methyl protons and H-3’ to C-4′ (Fig. 3a).

Based on NOESY interactions of the proton at C-3 and the one at C-1 (Fig. 3a), the relative configuration at the stereocenters C-1 and C-3 was determined to be cis. Ruthenium-mediated oxidative degradation” of this alkaloid led to the R-enantiomer of 3-aminobutyric acid (derived from C-3), as monitored by GC-MSD analysis of the Mosher derivatives. This revealed C-3 to be R-configured and, given the aforementioned relative cis-configuration, C-1 had to be S-configured.

At the naphthalene-isoquinoline biaryl axes, within the 5,8′-coupled monomeric halves, the stereo-array was determined by NOESY interactions of the proton at C-7′ with the equatorial proton at C-4 (δH 2.64 ppm; J = 17.3, 3.3 Hz) and between the axial proton (δH 2.27 ppm; J = 17.3, 12.0 Hz) and the aromatic proton at C-1′. This, in conjunction with the absolute R-configuration at C-3, as depicted in Fig. 3b, indicated P-configuration at the outer axes, revealing these portions to be structurally identical to the co-occurring known ancistroealaine C (7). Hence, the metabolite was found to be the 6’-homocoupling product of 7, with the absolute stereostructure 1 presented in Fig. 1. This was further confirmed by the similarity of the ECD (electronic circular dichroism) spectrum of 1 with that of the co-occurring, likewise P,P-configured michellamine A (9). Owing to this combination of two P-configured outer biaryl axes, the new compound was included in the ‘series A’ of michellamines – which so far comprised michellamines A–A2 – and was thus named michellamine A3. Its 1,3-cis-configurations in both tetrahydroisoquinoline portions, as highlighted in Fig. 1, is a unique structural feature that had not been found in any other similarly coupled naphthylisoquinoline dimers.

Michellamine A3 (2). The second new alkaloid, compound 2, corresponding to peak E in Fig. 2a, had a molecular weight 14 units larger than that of 1, matching with the molecular formula C47H50N2O8 (m/z 771.3655, HRESIMS). Its UV spectrum, which was very similar to that of 1, indicated that 2 was a 6’,6”-coupled dimer, too. This was further supported by its 1H NMR spectrum (Table 2), which closely resembled that of the co-isolated michellamine A2 (9), except for the presence of one additional three-proton singlet at 3.04 ppm. In the HSQC experiment, this extra methyl signal correlated with the carbon atom at 41.4 ppm, thus indicating 2 to be an N-methylated michellamine-type dimer.

Further 1D and 2D NMR data revealed that methyl group to be attached to the nitrogen atom in the ‘southeastern’ portion. Key evidence of the exact location of this N-methyl group were the HMBC correlations of its protons with both, C-1 and C-3 (Fig. 4a) and the signals of these methine carbons, which appeared deshielded (62.6 ppm for C-1 and 60.6 for C-3) as compared to those of the N-demethylated congeners – like, for example, in the case of 1 (Tables 1 and 2). This assignment was further supported by the ROESY series [Me-1 ↔ N-Me ↔ Me-3], which, in addition, revealed a 1,3-diaxial relationship of H-1 and H-3 and, thus, a relative cis-configuration at the stereocenters C-1 and C-3 (Fig. 4a). For the stereocenters in the other, ‘northwestern’ part of 2, a relative cis-configuration was observed, too, again evidenced by the ROESY interactions of H-1′ with H-3′, as in 1 (Fig. 3b).

The ruthenium-mediated oxidative degradation of 2 yielded both, R- and S-configured 3-aminobutyric acids, showing that this alkaloid was either 3R,3′S- or 3S,3′R-configured. The simultaneous formation of the N-methylated analog of 3-
aminobutyric acid only as its S-enantiomer, however, clearly indicated that the S-configured stereocenter was located next to the N-methyl group, i.e. at C-3. Thus, this compound was found to be 3S,3'R-configured, which, given the relative cis-configurations in both halves as above, showed that C-1 and C-1’ were R- and S-configured, respectively.

From ROESY interactions between H-4ax and H-7’ and between H-4eq and H-1’ (Fig. 4a), and based on the above-assigned S-configuration at C-3, the configuration at the southeastern axis was determined to be P, thus revealing this portion to correspond to korupensamine D, a known naphthylisoquinoline alkaloid from *A. korupensis* (for its structure, see Fig. S1). At the other, northwestern axis of 2, the ROESY interactions were complementary: H-4”ax with H-1” and H-4”eq with H-7”, which, together with the opposite R-configuration at C-3” established above, indicated that axis to be P-configured, too, as in 1 (Fig. 3b). This metabolite thus had to be a cross-coupling product of anancroelaine C (7) and korupensamine D (Fig. S1), and possessed the absolute structure 2 presented in Fig. 1. It was named michellamine Aₐ, due to its P,P-configured outer biaryl axes, as in 1. It is the second dimer of 5,8'-coupled naphthylisoquinolines with the unusual combination of two 1,3-cis-configured halves.

**Table 2**  
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</table>
| N-Me    | 3.04, s | 41.4 | a The NMR data were recorded in methanol-d₄ (δ in ppm). b This signal was overlapped by that of 2’-Me, but could be deduced to be dd, from the respective 2D NMR cross peaks. c These signals overlapped each other.
as the likewise isolated korupensamine B (a monoprotonated molecule at $|\Delta H-4\rangle$ with $H-1$ assigned to be $|\Delta H-1\rangle$, thus indicating that this metabolite consisted of a naphthyl-1,3-dimethyldihydroisoquinoline subunit. In addition, this dimer had three methoxy functions ($4.11, 4.10,$ and $4.03$ ppm), i.e. one $O$-methyl group more than the other three compounds, 1–3. Comparison of its $^1H$ NMR spectrum with those of 1–3 showed that these four metabolites had one common, nearly perfectly matching half-set of signals, corresponding to the northwestern portions of 1–3, which were equivalent to ancistroealaine C (7). The northwestern portion of 4 was therefore expected to be identical to 7, too, and its southeastern part was assumed to be the naphthyl-1,3-dimethyldihydroisoquinoline moiety, possessing two methoxy groups.

This assumption was corroborated by 1D and 2D NMR data, which led to a molecular architecture similar to that of 3, but with one additional methyl group attached to the oxygen function at C-8 and a double bond between C-1 and the adjacent nitrogen atom in the southeastern portion (Fig. 4c). Key NMR features of the location of the methoxy function at C-8 were the joint HMBC interactions from H-7 and OMe-8 to C-8 and the ROESY series [H-7 $\leftrightarrow$ OMe-8 $\leftrightarrow$ Me-1] (Fig. 4c).

The oxidative degradation procedure delivered only ($R$)-3-aminoxybutyric acid, showing that both, C-3 and C-3‴ were $R$-configured. In the northwestern portion, ROESY interactions at the stereocenters C-1‴ versus C-3‴ and across the naphthalene-isoquinoline linkage were the same as in 1 (Fig. 3b), so that C-1 was $S$-configured and the biaryl axis had the $P$-configuration. In the southeastern portion, the ROESY interaction between H-4ax and H-7 indicated these two spin systems to be on the same side of the molecule, as depicted in Fig. 4c, which evidenced an $M$-configuration at the biaryl axis. Therefore, this metabolite possessed the absolute stereostructure 4, as shown in Fig. 1. It was named michellamine B₃, due to its stereochemical similarity with michellamine B₄ (3). Within a list of now 16 known natural michellamine-type dimers, it is only the third example that possesses a dihydroisoquinoline ring system; the other two analogs with such a structural peculiarity are michellamines F‴ and A₉.

Ancistrobonsoline A₁ (5). The dichloromethane subfraction contained two major, nicely resolved metabolites (Fig. 2b). They were obtained as amorphous solids by resolution on a preparative reverse-phase HPLC column. The more polar one (Peak J, Fig. 2b), compound 5, gave an $m/z$ at $392.1843$ [M + H]⁺, corresponding to the molecular formula $C_{24}H_{28}NO_6$. In the $^1H$ NMR spectrum, the presence of two methoxy signals, together with the absence of a quartet around $4.5$ ppm, which typically indicates a proton located at C-1, hinted at a naphthyl-1,3-dimethyldihydroisoquinoline alkaloid, presumably equivalent to the southeastern portion of 4. This was further corroborated by the fact that the signals in the aliphatic region of the $^1H$ NMR spectrum of 5 matched very well with those assigned to the protons of the southeastern half of 4. Detailed analysis of the 1D and 2D NMR spectra of this monomeric alkaloid established it to have the same molecular skeleton as the southeastern portion of 4, yet with an additional proton at C-6‴.

The ROESY interactions across the biaryl axis of 5 were all similar to those in the southeastern portion of 4 (see Fig. 4c), as were also the results of the oxidative degradation. This aforementioned cis- and trans-arrays, established the absolute configurations at C-1‴ and C-1 to be $S$ and $R$, respectively.

From ROESY interactions of H-4‴ax with H-1‴ and of H-4‴er with H-7‴ and in view of the absolute $R$-configuration at C-3‴, the absolute axial configuration in the northwestern half was assigned to be $P$, revealing this portion to be identical to 7, as in 1 (Fig. 3b). In the other, southeastern part, the respective protons interacted in a complementary way: H-4ax with H-7‴ and H-4ε with H-1‴, as shown in Fig. 4b. These correlations, together with the known R-configuration at C-3 (see above), evidenced the southeastern biaryl axis to be $M$-configured, thus showing the corresponding moiety to have the same structure as the likewise isolated korupensamine B (8b) (Fig. 4b). Therefore, compound 3 had to be the 6‴,6‴-cross-coupling product of ancistroealaine C (7) and korupensamine B (8b), with the absolute stereostructure given in Fig. 1. It was named michellamine B₄, in continuation of the series of michellamines with opposite configurations at the two outer biaryl axes (type B).

Michellamine B₃ (4). Compound 4, the slowest-eluting major metabolite of the aqueous fraction (Peak H, Fig. 2a), gave a monoprotonated molecule at m/z 769.3484 in HRESIMS, i.e. 12 units more than that of 3. Its $^1H$ NMR spectrum displayed a full set of signals, with in particular one quartet (instead of two), thus indicating that this metabolite consisted of a naphthyl-1,3-dimethyltetrahydro- and a naphthyl-1,3-dimethyldihydroisoquinoline subunit. This observation raised the question of the constitution and – together with the results of the oxidative degradation – absolute stereostructure given in Fig. 1. It was named michellamine B₄, in continuation of the series of michellamines with absolute stereostructure given in Fig. 1. It was named michellamine B₄, in continuation of the series of michellamines with absolute stereostructure given in Fig. 1.
monomeric alkaloid thus possessed the absolute stereostructure 5, as presented in Fig. 1. It was, consequently, the as yet undescribed enantiomer of the known alkaloid 6,5′-O,O-dimethylancistroealaine A. In agreement with their opposite absolute configurations, their ECD spectra were fully mirror-image like (Fig. 5). Instead of the rational, but long name ent-6,5′-O,O-dimethylancistroealaine A, the new compound was named ancistrobonsoline A₁, after the Congolese village Bonsolervie, where the plant had been collected.

**Ancistrobonsoline A₂ (6).** The sixth new compound discovered during these investigations (Peak I, Fig. 2b) was attributed the molecular formula C₅₂H₄₇NO₄ according to the observation of its mono-protonated molecule at m/z 406.2013 in HRESIMS. Its ¹H NMR spectrum was very similar to that of 5, indicating the presence of a 1,3-dimethyldihydroisoquinoline, yet with the signal of an additional O-methyl group at 3.83 ppm, linked either to O-6 or O-5. NOE interactions of the protons of that O-methyl group with H-7, together with their joint HMBC correlations to C-6, clearly located that methoxy function at C-6.

The absolute configuration at the stereocenter C-3 of 6 was determined to be R by oxidative degradation, which, as in the case of 5, provided (R)-3-amino butyric acid. Across the biaryl axis the specific relationships between the diastereotopic protons at C-4 and the aromatic protons H-1′ and H-7′ were the same as in 5 (Fig. 4c): H-4ax (‘up’) interacted with H-7′ and H-4eq (‘down’) with H-1′, which was consistent with an M-configuration at the 5,8′-axis. This absolute stereochemical assignment was further corroborated by the close similarity of the ECD spectrum of 6 with that of 5. Therefore, the new alkaloid had to possess the absolute stereostructure 6, as presented in Fig. 1, and was, thus, the 6-O-methylated derivative of 5. It was named ancistrobonsoline A₂. With their R-configuration at the stereocenter and M at the biaryl axis, ancistrobonsolines A₁ (5) and A₂ (6) belong to a very small subgroup of naphthyldihydroisoquinolines displaying such stereochemical features, with previously only one single representative, anchistrolikokine F (Fig. S1†).¹²

**Chemotaxonomic and biosynthetic significance of the isolated metabolites**

The molecular architectures of the alkaloids thus isolated from this as yet unidentified Ancistrocladus taxon are remarkable in many respects. Firstly, it is striking that all of these compounds belong to the same 5,8′-coupling type and, where dimers are concerned, they are all based on a central 6,6′-axis. Such a specific phenol-oxidative coupling – and, thus strict enzymic assistance – have so far been observed only in A. likoko¹¹,¹²,¹₆

These findings thus reveal a close phylogenetic relationship between the two taxa (in addition to their aforementioned morphological similarities), but delineate one from the other by the fact that the naphthalene moieties of the metabolites produced by the plant from the proximity of Bonsolervie are, all of them, O-methylated only at C-4′ (and C-4″, in the case of dimers), not at C-5′ (nor at C-5″). And their configuration at C-3 (or/and C-3″, for dimeric compounds) is always R, except for the N-methylated isoquinoline portions, where an exclusive S-configuration is observed (see Fig. 1). Such a regioselective O-methylation in the naphthalene subunits and a stereospecificity at C-3 (and/or C-3″) are not known from A. likoko (or from any other related plant). Likewise demarcating these two taxa is the fact that A. likoko is a virtually exclusive producer of monomeric alkaloids (out of 21 metabolites, only two dimers have so far been isolated from this species),¹² whereas the plant investigated here is a rich source of dimeric compounds.

Moreover, the remarkable occurrence of dimers with a symmetric 6′,6″-coupling at the central axis chemotaxonomically delineates this taxon from the other – likewise still unidentified – liana that occurs in the same area.¹⁴ The latter is known to produce a drastically different metabolic pattern, including dimers with unsymmetric, 6′,1″-coupled central axes,¹⁷ with additional oxygen bridges¹⁸ and/or acetal linkages,¹⁹ but so far no symmetric, 6′,6″-coupled ones, not even in traces.

Secondly, it is particularly noteworthy that this liana produces the 1,3-trans-configured korupensamine A (8a) only in small quantities (Peak B, Fig. 2a) and shows no hints at the presence of its dimer, michellamine A (11), not even in traces.²⁰ This is exceptional, since all of the plants known to produce michellamine-type alkaloids, viz. A. korupensis,¹⁶,²⁰ A. congoensis,⁴ and A. likoko,¹³ always contain michellamine A (11) as one of the main dimeric compounds. Moreover, 11 is always accompanied by its monomer, korupensamine A (8a), likewise occurring as a major metabolite in those plants.⁴,¹⁶,²⁰ For this reason, it has even been suggested⁴ that other (minor) dimer analogs differing from michellamine A (11) by the configurations at the stereocenters may originate from the preformed ‘parent’ dimer michellamine A (11), i.e. by modification after the dimerization step. In the case of the metabolites of the plant investigated here, however, one of the main constituents (Peak D, Fig. 2a), michellamine A₆ (1), is twofold 1,3-cis-configured and can be
regarded as a bis-epimer of michellamine A (11) at C-1 and C-1”. Since 1 is accompanied by an unusually large quantity of its – likewise cis-configured – monomer (Peak A, Fig. 2a), anisotroalaine C (= 1-epi-korupensamine A) (7), it seems likely that 1 (as also 2) originates from the correspondingly preformed cis-monomers.

**Biological activities of the isolated metabolites**

**Antiprotozoal properties.** Owing to the pronounced antiprotozoal activities of some naphthylisoquinoline alkaloids, the two new monomers anisotrobonsolines A1 (5) and A2 (6) were tested against the pathogens causing malaria (*Plasmodium falciparum*), human African sleeping sickness (*Trypanosoma brucei rhodesiense*), Chagas’ disease (*T. cruzi*), and visceral leishmaniosis (*Leishmania donovani*). The other isolated new dimeric metabolites were not evaluated, since their analogs had previously been found inactive. Likewise, as shown in Table 3, compounds 5 and 6 exhibited weak to moderate inhibitory properties. Of particular interest is that the results of 5 and 6 showed the OME/OH substitution pattern to play a crucial role for the bioactivities: an O-methylation seems to be favorable for the antiplasmodial and antitypanosomal activities, but disadvantageous for the antileishmanial activity. This finding complements previous results on the antileishmanial activities of related alkaloids, yet with *P* configuration at the axis and *S* at C-3. Likewise interesting are the antiprotozoal activities of 5 and its enantiomer (Table 3), which are – in the case of *T. cruzi* – drastically different from one to another, documenting, once again, the impact of the absolute stereostructure.

**Anti-HeLa potential: cytotoxicity and effects on cell morphology.** As part of our ongoing investigations on the potential of naphthylisoquinoline alkaloids towards different cancer cell lines, the isolated compounds were tested for their cytotoxic activity against HeLa human cervical cancer cells (Table 4). Interestingly, the new twofold 1,3-cis-configured dimeric compounds, michellamines A6 (1) and A7 (2), and the likewise new naphthylidihydroisoquinoline alkaloids, anisotrobonsolines A1 (5) and A2 (6), displayed strong cytotoxic activity, with IC50 values between 14.8 and 21.5 μM. The most potent cytotoxicity (IC50 = 8.8 μM) was displayed by michellamine E (10), which was even more active than the positive control 5-fluorouracil (IC50 = 13.9 μM), an anticancer drug in clinical use.

Michellamine E (10) was, therefore, investigated for its effects on cell morphology and apoptosis using two distinct staining methods, the Hoechst 33342 staining and the ethidium bromide–acridine orange (EB–AO) double staining assay. In the Hoechst 33342 staining, the dye penetrates through the cell membrane and intercalates with DNA and emits blue fluorescence. As shown in Fig. 6a, untreated HeLa cells (the control) displayed regular cell morphology with the intact nuclei. However, treatment with 12.5 μM of 10 induced nuclear fragmentation, suggestive of cells undergoing apoptosis, as indicated by fragmented nuclei exemplarily shown by white arrows in Fig. 6a. The EB–AO assay, on the other hand, allows visualizing the cellular morphology as well as the distinction between the live and dead cells. Acidine orange (AO) is a cell-membrane permeable dye emitting bright-green fluorescence in live cells, and ethidium bromide (EB) penetrates only the membrane of dying or dead cells staining them red. As shown in Fig. 6b, untreated HeLa cells (the control) displayed intact, regular cell morphology with exclusive bright green fluorescence in AO–EB

**Table 3** Antiprotozoal activities of the new naphthylidihydroisoquinoline alkaloids 5 and 6, and the previously reported data of the enantiomer of 5 (ref. 15)

<table>
<thead>
<tr>
<th>Compound</th>
<th>P. falciparum</th>
<th>T. brucei rhodesiense STIB900</th>
<th>T. cruzi Tulahuen C2C4 Lac Z</th>
<th>L. donovani axenic amastigotes MHOM/ET/67/L82</th>
<th>Cytotoxicity (L6 cells)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>K1</td>
<td>NF54</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>2.7</td>
<td>2.1</td>
<td>44.4</td>
<td>109.3</td>
<td>87.9</td>
</tr>
<tr>
<td>ent-5</td>
<td>5.4</td>
<td>n. d</td>
<td>24.8</td>
<td>16.3</td>
<td>43.4</td>
</tr>
<tr>
<td>6</td>
<td>1.8</td>
<td>2.4</td>
<td>12.6</td>
<td>80.1</td>
<td>246.6</td>
</tr>
<tr>
<td>Standard</td>
<td>0.31 b</td>
<td>0.01 b</td>
<td>0.04</td>
<td>5.69 b</td>
<td>1.08 e</td>
</tr>
</tbody>
</table>

*Values reported earlier (see, ref. 15).* a Chloroquine. b Melarsoprol. c Benzimidazole. * Miltefosine. † Podophyllotoxin.
staining. Treatment of the tumor cells with 12.5 μM of 10, however, disrupted the cellular integrity leading to rounding of the cell membrane, membrane rupture, and disintegration of cellular contents resulting in an increased population of red EB fluorescence of dead cells (Fig. 6b).

**Antiausterity activities: preferential cytotoxicity against PANC-1 human cancer cell line and effects on cell morphology.** Following the antiausterity strategy as recently developed,30–32 the isolated naphthylisoquinoline alkaloids 1–10 were tested against PANC-1 pancreatic cancer cells. Pancreatic tumors are hypovascular (limited blood vessels) in nature and are, consequently, constantly exposed to nutrient and oxygen starvation in their microenvironment. However, pancreatic cancer cells show a remarkable tolerance for nutrition starvation, allowing them to adapt and survive in such inadequate supply of nutrients.30 Discovery of anticancer agents by targeting this tolerance of nutrition starvation is the key approach of antiausterity strategy in anticancer drug discovery. The new dimer michellamine A7 (2) and its known analogs michellamines A2 (9) and E (10) exhibited significant preferential cytotoxicities against PANC-1 cells in a concentration-dependent manner (Fig. S2†). Their PC50 values (i.e. the concentration at which 50% of the cells are preferentially killed under nutrient-deprived conditions, without cytotoxicity in normal, nutrient-rich medium) ranged from 18.9 to 24.3 μM, but were surpassed by the even higher activities of the new monomeric compounds ancistrobonsolines A1 (5) and A2 (6), with PC50 values of 7.5 and 12.1 μM, respectively (Table 4).

The effects of ancistrobonsolines A1 (5) and A2 (6), as representatives of the compounds showing potent antiausterity activities, on the cell morphology and apoptosis were further investigated by the ethidium bromide (EB)–acridine orange (AO) double-staining fluorescence assay as described above. As shown in Fig. 7A and A’, the untreated PANC-1 cells, serving as the control, emitted the typical bright-green color, characteristic of living cells, exhibiting an intact cellular morphology. This, however, changed significantly when the cells were treated with 12.5 μM of 5 and 6, as revealed by the yellow fluorescence resulting from the overlapping of light emitted by AO (green) and EB (red) (Fig. 7B’ and D’). With 25 μM, both compounds

![Fig. 6 Morphological changes of HeLa human cervical cancer cells induced by 12.5 μM of michellamine E (10) in comparison to untreated control: (a) Hoechst 33342 nuclei staining and (b) acridine orange (AO)–ethidium bromide (EB) staining.](image)

![Fig. 7 Morphological changes of PANC-1 human pancreatic cancer cells induced by the new compounds ancistrobonsolines A1 (5) and A2 (6) in comparison to the untreated ones (control).](image)
induced a dramatic morphological alteration and disintegration of cellular organelles, leading to total cell death as illustrated by the virtually exclusive red stain in Fig. 7(C’ and E’). All these results suggest that naphthylisoquinoline alkaloids are promising lead structures for anticancer drug development.

**Experimental section**

**General experimental procedures**

UV/Vis spectra were measured with a Shimadzu UV-1800 spectrophotometer, IR spectra with a Jasco FTIR-4600 type A spectrometer, and optical rotations with a Jasco P-1020 polarimeter.

ECD spectra were obtained on a J-715 spectropolarimeter (Jasco) at room temperature, using a standard cell (0.02 cm) and spectrophotometric-grade methanol, and are reported in Δε values (cm² mol⁻¹) at the given wavelength λ (nm). GC-MSD analyses were performed on a GCMS-QP 2010SE (Shimadzu).

**Plant material**

*Ancistrocladus* plant material, morphologically related to those of *A. likoko*, were collected in August 2015 (Global Positioning System coordinates: 0°06.572S, 018°20.146E, 315 ± 22 m) by one of us (B. K. L.) in the vicinity of the village Bonsonville, which is located at about 20 km southeast of the town of Mbandaka in the Democratic Republic of the Congo. A voucher specimen (No. 105) has been deposited at the Herbarium Bringmann, Institute of Organic Chemistry, University of Würzburg.

**Extraction and isolation**

Air-dried leaves (200 g) were ground and repeatedly extracted by maceration with a neutral mixture of MeOH – CH₂Cl₂ (1 : 1, 2 L) with mechanical shaking (160 RPM). After three cycles of 24 h each, the filtrates were evaporated to dryness and the marc was extracted with an acidified (HCl) mixture of MeOH – CH₂Cl₂ (1 : 1, pH = 2–3, 2 L), again with mechanical shaking, for 48 h. The acidified extract was neutralized with NaOH and mixed with the neutral one, after the similarity between the two extracts was established by analytical HPLC. The resulting total crude residue was submitted to a cation-exchange column (Amberlyst-15 from Fluka, Ø 3 cm) to remove undesired, non-basic metabolites. The obtained alkaloid-enriched fraction was partitioned between water and dichloromethane, giving, after evaporation, 800 mg of an aqueous subfraction and 30 mg of dichloromethane subfraction. The polar subfraction was dissolved in methanol and resolved by preparative HPLC, using the following gradient: A/B: 0 min 10% B; 25 min 25% B; 30 min 28% B; 31 min 100% B; 34 min 100% B; 35 min 10% B, in which A = 90% H₂O + 0.05% trifluoroacetic acid and B = 90% MeCN + 0.05% trifluoroacetic acid. From this subfraction, eight metabolites were obtained, including the alkaloids 1-3 and 6-10. Resolution of the lipophilic subfraction by preparative HPLC yielded compounds 4 and 5.

**Michellamine A₈ (1).** [α]D 30: −35 (c = 0.01, MeOH); UV (MeOH) λmax nm: 200, 229, 263, 302, 311, 316, 331, 333, 344; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₂₃₀ +29.1, Δε₃₄₇ +15.2, Δε₃₅₀ −11.8, Δε₃₃₁ +5.1; IR (ATM): νmax cm⁻¹: 3329, 1672, 1600, 1418, 1359, 1249, 1181, 1138, 1071, 836, 799, 722; HRESIMS: m/z 757.3486 [M + H]⁺ (calcd for C₄₆H₄₉N₂O₈: 757.3484). For 1H and 13C NMR data, see Table 1.

**Michellamine A₆ (2).** [α]D 30: −41 (c = 0.08, MeOH); UV (MeOH) λmax nm: 203, 229, 263, 302, 311, 316, 331, 333, 345; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₂₃₀ +26.5, Δε₃₄₃ +23.3, Δε₃₃₀ −14.3, Δε₃₄₀ +4.8; IR (ATM): νmax cm⁻¹: 3329, 1672, 1600, 1418, 1359, 1249, 1181, 1138, 1071, 836, 799, 722; HRESIMS: m/z 771.3655 [M + H]⁺ (calcd for C₄₇H₅₁N₂O₇: 771.3640). For 1H and 13C NMR data, see Table 2.

**Michellamine B₄ (3).** [α]D 30: −25 (c = 0.03, MeOH); UV (MeOH) λmax nm: 201, 229, 264, 302, 311, 330, 333, 343; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₂₆₇ +2.0, Δε₂₃₀ −5.7, Δε₃₁₆ +1.0, Δε₃₃₃ −0.4; IR (ATM): νmax cm⁻¹: 3335, 1672, 1617, 1418, 1360, 1251, 1200, 1137, 1072, 837, 800, 722; HRESIMS: m/z 757.3475 [M + H]⁺ (calcd for C₄₆H₄₉N₂O₇: 757.3483). For 1H and 13C NMR data, see Table 2.

**Michellamine B₅ (4).** [α]D 30: −32 (c = 0.008, MeOH); UV (MeOH) λmax nm: 204, 231, 266, 301, 311, 317, 332, 345; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₂₃₀ −22.0, Δε₂₃₉ −8.0, Δε₃₁₉ −10.1, Δε₂₆₂ +14.1, Δε₃₄₀ −9.0, Δε₃₃₂ −6.7, Δε₃₃₀ −11.1; IR (ATM): νmax cm⁻¹: 3354, 1670, 1625, 1577, 1408, 1354, 1326, 1262, 1199, 1132, 1071, 957, 832, 798, 720; HRESIMS: m/z 769.3484 [M + H]⁺ (calcd for C₄₅H₄₀N₂O₆: 769.3483). For 1H and 13C NMR data, see Table 2.

**Acirobonosoline A₈ (5).** [α]D 30: +75 (c = 0.01, MeOH); UV (MeOH) λmax nm: 336, 325, 315, 312, 326; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₃₂₁ −10.1, Δε₂₃₈ +5.7, Δε₂₄₄ −2.9, Δε₂₆₉ +0.6, Δε₂₈₁ +0.5, Δε₁₃₀ +2.8; IR (ATM): νmax cm⁻¹: 3382, 1671, 1625, 1575, 1400, 1352, 1326, 1297, 1264, 1198, 1132, 1084, 956, 832, 799, 720; HRESIMS: m/z 392.1843 [M + H]⁺ (calcd for C₂₄H₂₆NO₄: 392.1856). For 1H and 13C NMR data, see Table 1.

**Acirobonosoline A₆ (6).** [α]D 30: +56 (c = 0.009, MeOH); UV (MeOH) λmax nm: 336, 325, 315, 313, 237; ECD (MeOH) cm² mol⁻¹: Δε max cm⁻¹: Δε₂₃₁ −12.1, Δε₂₃₉ +4.7, Δε₂₄₂ −1.9, Δε₂₆₂ +1.6, Δε₂₈₆ +0.6, Δε₃₃₄ +2.6; IR (ATM): νmax cm⁻¹: 3384, 1670, 1626, 1577, 1457, 1263, 1200, 1130, 1084, 1052, 956, 834, 798, 719; HRESIMS: m/z 406.2013 [M + H]⁺ (calcd for C₂₅H₂₆NO₄: 406.2013). For 1H and 13C NMR data, see Table 1.

**Known alkaloids isolated,** 7-10. The likewise isolated monomeric dimeric alkaloids, acirolealaine C (7), korupensamines A (8a) and B (8b), and michellamines A₈ (9) and E (10) were found to be identical in their spectroscopic, physical, and/or chromatographic properties with data previously reported.7-10
Ruthenium-mediated oxidative degradation

Following a procedure described earlier,14 ca. 0.8 mg of the compounds 1–10 were submitted to a ruthenium(m)-catalyzed periodate oxidation, followed by derivatization of the resulting amino acids with MeOH/HCl and (R)-x-methoxy-x-trifluoromethylphenylacetyl chloride [(R)-MTPA-Cl, prepared from (S)-MTPA] and analyzed by GC-MSD.

Antiprotozoal activities

The assessment of the antiprotozoal properties of the compounds 5 and 6 against Plasmodium falciparum (NF54 and K1), Trypanosoma cruzi (Tulahuen C24 with the Lac Z gene incorporated) amastigotes in mouse macrophages, Trypanosoma brucei rhodesiense (STIB900) bloodstream stages, and Leishmania donovani (MHOM/ET/67/L82) axenic amastigotes and the testing of the cytotoxicity against mammalian host cells (rat skeletal myoblast L6 cells) were done in vitro as previously described.15

Cytotoxicity assay

The cytotoxicity assays against HeLa cell line (RCB0007, Tsukuba, Japan) were carried out as described previously.24 In brief, exponentially growing cells were harvested and plated in 96-well plates (2 × 10^3 per well) in DMEM and allowed to attach for 24 h in the humidified CO_2 incubator at 37 °C. The cells were then washed with phosphate-buffered saline (PBS) followed by the addition of serially diluted test samples in DMEM. For each concentration, three replications were performed. After 72 h of incubation, the cells were washed twice with PBS, and 100 μL of DMEM containing 10% WST-8 cell counting kit (Dojindo) solution were added. After 3 h of incubation, the absorbance at 450 nm was measured on an EnSpire Multimode plate reader (PerkinElmer, Waltham, MA, USA). Cell viability was calculated from the mean values from three wells using the following equation:

\[
\text{Cell viability (\%) = \frac{[\text{Abs}_{\text{test sample}} - \text{Abs}_{\text{blank}}]}{[\text{Abs}_{\text{control}} - \text{Abs}_{\text{blank}}]} \times 100\%}
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Antiausterity assay

The human pancreatic cancer PANC-1 (RBRC-RCB2095, Tsukuba, Japan) cell line was purchased from the Riken BRC cell bank and maintained in the standard DMEM with 10% FBS supplement under a humidified atmosphere of 5% CO_2 in the incubator at 37 °C. For the antiausterity evaluation, exponentially growing cells were seeded in 96-well plates (1.5 × 10^4 per well) in DMEM and incubated for 24 h for the cell attachment. After this incubation time, the cells were washed twice with PBS, the medium was changed to serially diluted test samples in both nutrient-rich medium (DMEM) and nutrient-deprived medium (NDM) with a control and a blank in each test plate. The composition of the NDM was as follows: 0.1 mg L^{-1} Fe(NO_3)_3 (9H_2O), 265 mg L^{-1} CaCl_2 (2H_2O), 400 mg L^{-1} KCl, 200 mg L^{-1} MgSO_4 (7H_2O), 6400 mg L^{-1} NaCl, 700 mg L^{-1} NaHCO_3, 125 mg L^{-1} Na_2HPO_4, 15 mg L^{-1} phenol red, 25 mM L^{-1} HEPES buffer (pH 7.4), and MEM vitamin solution (Life Technologies, Inc., Rockville, MD, USA); the final pH was adjusted to 7.4 with 10% aqueous NaHCO_3. After 24 h of incubation with the respective test compound in DMEM and NDM, the cells were washed twice with PBS and replaced by 100 μL of DMEM containing 10% WST-8 cell counting kit solution. After 3 h of incubation, cell viability was measured and calculated as described above.

Morphological assessment of cancer cells

For studies on morphological changes, HeLa and PANC-1 cells were seeded in 24-well plates (1 × 10^5) and incubated in a humidified CO_2 incubator for 24 h for the cell attachment. The cells were then washed twice with PBS and treated with vehicle control or test compounds in DMEM for HeLa cells; and vehicle control or test compounds in NDM for PANC-1 cells, and incubated for 24 h. For nuclei staining, two drops of the NucBlue® ready probe (Hoechst 33342) was directly added to the cells in full media and was incubated for further 25 min at the end of the experiment. For cellular morphology, 10 μL of EB/AO reagent (dye mixture: 100 μg mL^{-1} AO and 100 μg mL^{-1} EB in PBS) was added to each test well and further incubated for 10 min. The cellular images were captured in the fluorescent and phase contrast modes, using an EVOS FL digital microscope (20× objectives).

Conclusions

In summary, this paper describes the discovery of the first twofold 1,3-cis-configured michellamelline-type dimers, 1 and 2, and of four other mono- and dimeric naphthylisoquinoline alkaloids, 3–6, in the leaves of an as yet unidentified Congolese Ancistrocladus liana related to A. likoko. These new compounds, obtained along with five previously described analogs, 7–10, chemotaxonomically delineate this Congolese plant from all known related taxa, thus suggesting that it is probably a new species. Further taxonomic investigations, including DNA analysis, are planned. Some of the isolated metabolites have shown attractive cytotoxicities against the HeLa cell line and very good antiausterity activities against PANC-1 human pancreatic cancer cells. The weak-to-moderate antiprotozoal properties of the new monomeric metabolites 5 and 6 highlight the consequence of the stereochemical features on the biological activities and provide valuable information for the ongoing SAR investigations.

Conflicts of interest

There are no conflicts to declare.

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Notes and references


