Interdomain flip-flop motion visualized in flavocytochrome cellobiose dehydrogenase using high-speed atomic force microscopy during catalysis†

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Cellobiose dehydrogenase (CDH) is a dual domain flavocytochrome, which consists of a dehydrogenase (DH) domain containing a flavin adenine dinucleotide and a cytochrome (CYT) domain containing b-type heme. To directly visualize the dynamic domain motion of class-I CDH from Phanerochaete chrysosporium (PcCDH) during catalysis using high-speed atomic force microscopy, the apo-form of PcCDH was anchored to a heme-immobilized flat gold surface that can specifically fix the orientation of the CYT domain. The two domains of CDH are found to be immobile in the absence of cellobiose, whereas the addition of cellobiose triggers an interdomain flip-flop motion involving domain-domain association and dissociation. Our results indicate that dynamic motion of a dual domain enzyme during catalysis induces efficient electron transfer to an external electron acceptor.

Introduction

Cellulose, a major component of plant cell walls, is a promising source of biomass. Conversions of recalcitrant cellulose into a plant-base small molecule are required to realize an ideal biorefinery process. Cellobiose dehydrogenases (CDHs), which are extracellular flavocytochromes, and copper-dependent lytic polysaccharide monooxygenases (LPMOs) have recently been found to contribute to a process of oxidative depolymerization of cellulose. In the process, electrons are donated from cellobiose via CDH to the copper site of LPMO and then activated LPMO catalyzes oxidative cleavage of glycosidic C–H bonds to boost the degradation of cellulose.

Unraveling the mechanism of catalysis of cellobiose oxidation by CDH as an electron donor for LPMO therefore represents a focal point in terms of development of an ideal conversion process of cellulose.

CDH, secreted by filamentous fungi from the phyla of Basidiomycota and Ascomycota, consists of two domains: a dehydrogenase (DH) domain containing flavin adenine dinucleotide (FAD) and a cytochrome (CYT) domain containing b-type heme (iron protoporphyrin IX). The domains are connected by a long flexible linker (Fig. 1a). The DH domain oxidizes the anomeric C1 position of cellobiose, and the two electrons carried by the reduced form of FAD in the DH domain are expected to be sequentially shuttled to heme b in the CYT domain via interdomain electron transfer (IET). The reduced form of CYT then donates the electrons to an external electron acceptor such as LPMO.

Two conformational snapshots of CDH in closed and open states are also determined by X-ray crystal structure analysis of full-length CDH. However, the closed and open structures are obtained from different species, Myriococcus thermophilum (MtCDH) and Neurospora crassa (NcCDH), respectively (Fig. S1†). The next target is to demonstrate the single-molecule dynamic motion of CDH during catalysis.

We thus envisioned capturing the transient interdomain motion of CDH using high-speed atomic force microscopy (HS-AFM) to obtain a fundamental understanding of the structure and function of CDH. HS-AFM has recently been proven to be a powerful imaging technique capable of capturing dynamic
motion of biomolecules temporally at the sub-second timescale and at sub-molecular lateral resolution.\textsuperscript{19–23} We herein report direct visualization of the dynamic motion of class-I CDH from the basidiomycete \textit{Phanerochaete chrysosporium} (PcCDH) by HS-AFM. To anchor and orient CDH on a flat surface for AFM imaging, we applied a technique for reconstitution of hemo-proteins on a heme-immobilized surface via the heme–heme pocket interaction (Fig. 1b).\textsuperscript{24–27} The specific immobilization of the CYT domain of CDH on the flat surface enables us to directly visualize the interdomain motion of catalytically-active PcCDH at a 300 ms temporal resolution.

Results and discussion

To obtain high-resolution AFM images, we used deglycosylated PcCDH, which has higher uniformity in terms of structure. Deglycosylated PcCDH with characteristic absorption maxima at 380, 421, 450, 529, and 570 nm originating from FAD and ferric heme (hemin) was adjusted to pH 1.5 with an acidic solution to remove both cofactors (Fig. S2\textsuperscript{†}). FAD was then incorporated into the resulting apoprotein PcCDH\textsubscript{F+H} to afford the FAD-bound protein, PcCDH\textsubscript{F+H} to catalytically-active PcCDH at a 300 ms temporal resolution.

We then carried out real-time imaging of the domain motion of PcCDH immobilized on the gold surface during the catalytic reaction using the HS-AFM technique. In the absence of cellobiose, immobilized PcCDH is imaged as a round-shaped object with a height of \textasciitilde6–8 nm (Movie S1\textsuperscript{†}). Surprisingly, when cellulose is added to the sample solution at the final concentration of 150 \textmu M, these motionless objects of PcCDH show a significant change in terms of motion (Fig. 2 and Movie S4\textsuperscript{†}). Each PcCDH contains a motionless part, which would be

domain of PcCDH\textsubscript{F+H} to afford the reconstituted protein, PcCDH\textsubscript{F+H}, in ca. 55\% yield (Fig. S4\textsuperscript{†}). The reconstituted protein with both FAD and heme was found to retain the inherent activity of PcCDH. The activity of PcCDH\textsubscript{F+H} toward cellobiose oxidation was determined by reduction of dichloroindolindophenol (DCIP) or ferricytochrome c as an oxidant according to the literature.\textsuperscript{29,30} (Fig. S5\textsuperscript{†}). The reduction of DCIP is mainly dependent on the redox reaction of the DH domain, whereas the reduction of cytochrome c depends mainly on the IET from FAD of the DH domain to heme in the CYT domain. PcCDH\textsubscript{F+H} completely lacks activity for reduction of both electron acceptors. PcCDH\textsubscript{F+H} has activity for DCIP reduction and does not exhibit cytochrome c reduction activity due to the absence of the heme cofactor in the CYT domain. The reconstituted protein, PcCDH\textsubscript{F+H}, with both cofactors, has activity similar to PcCDH in both assays. These results indicate that the step-by-step insertion of FAD and heme converts PcCDH\textsubscript{F+H} to catalytically active PcCDH.

To construct PcCDH\textsubscript{F+H} on a flat surface suitable for HS-AFM measurements, PcCDH\textsubscript{F+H} was immobilized on a heme-modified gold surface (heme/Au) (Fig. S6 and S7\textsuperscript{†}). We confirm the anchoring of PcCDH\textsubscript{F+H} onto heme/Au by visualizing the process of protein reconstitution using HS-AFM (Fig. S8\textsuperscript{†}). We first observed objects with a height of ca. 3 nm on heme/Au, which is consistent with the length of the 11-amino-1-decanethiol-linked heme (Fig. S7\textsuperscript{†}). After addition of PcCDH\textsubscript{F+H}, larger spherical objects with a height of 6–8 nm were observed on the surface (Fig. S8\textsuperscript{†}). The immobilization of PcCDH\textsubscript{F+H} on the heme/Au surface was also confirmed by quartz crystal microbalance (QCM) measurements (Fig. S9\textsuperscript{†}). The frequency decay of QCM was not observed upon addition of PcCDH with the native heme cofactor to heme/Au. PcCDH\textsubscript{F+H} with a vacant heme pocket showed a clear frequency decay. This finding indicates that PcCDH\textsubscript{F+H} is incorporated via the specific interaction between the heme molecule on the gold surface and the heme pocket of the CYT domain, leading to oriented anchoring of the CYT domain of PcCDH.

We tested the activity of PcCDH anchored on the surface in the presence of cellobiose. After the PcCDH@heme/Au substrate was immersed in a 50 \textmu M cellobiose solution in NaOAc buffer (2 mM, pH 4.5), cellobiose and cellulbio-1,5-lactone in the solution were analyzed by ESI-TOF MS. The conversion of cellobiose into cellulbio-1,5-lactone was confirmed under aerobic conditions, thereby indicating that PcCDH linked on the gold surface exhibits the inherent enzymatic activity using O\textsubscript{2} as an electron acceptor.
anchored at the same place on the surface, and a neighboring object moving rapidly around the motionless part. Considering that the CYT domain is anchored to the surface via the heme molecule, the moving part is identified as the DH domain of PcCDH. In addition, we observed a change in the height of PcCDH using a standard AFM arrangement (Fig. S10†). The average height of 6.6 ± 1.0 nm for PcCDH in the absence of cellobiose was changed to 9.1 ± 0.9 nm in the presence of cellobiose. This is consistent with the heights estimated from the X-ray crystal structure of the closed state (ca. 6 nm) and the open state (ca. 10 nm) of PcCDH. By contrast, the changes in the height profile were not detected by HS-AFM measurements. The DH domain, which is linked via a flexible linker, moves rapidly in the open state, and thus the height of the domain in rapid motion cannot be precisely measured using HS-AFM. The tip vibration would force the moving DH domain to remain near the surface in the fast scan mode.

To clarify the evidence that the cellobiose molecule triggers the domain motion, the degree of the motion in PcCDH was analyzed at different concentrations of cellobiose (Movies S2 to S5†). The conformational dynamics of PcCDH were quantified using their two-dimensional correlation coefficient (CC) averaged through 300 ms × 250 frames for each molecule. The decrease in the CC value represents the difference between the two sequential images of the focused enzyme in the movie. In the absence of cellobiose, the CC value of PcCDH is distributed around 0.97, suggesting the immobility of PcCDH in the absence of the substrate. The CC values are gradually shifted from 0.96 to 0.88 as the concentration of cellobiose increases from 50 to 200 μM (Fig. 3). The decrease in the CC values indicates that the domain motion in PcCDH becomes more intense. Furthermore, when cellobiose is removed from the observation cell by exchanging the buffer, the CC value shifts to 0.97, which is similar to that observed in the absence of cellobiose (Fig. S11 and Movie S6†). The lower shift of the CC distribution suggests the acceleration of the structural changes.
between the closed and the open states in PccDHs. Therefore, the degree of the motion in PccDH anchored on the gold surface increases in a cellubiose-dose-dependent manner, indicating that cellubiose binding followed by the reaction initiates the dynamic motion of the two domains.

In the oxidative depolymerization of cellulose, the DH domain of CDH is known to accept two electrons from cellubiose and then reduces the CYT domain through one-electron IET. The catalytic cycle of CDH through a ping-pong mechanism involving five steps has been proposed as shown in Fig. 4. PccDH is expected to be in the closed or open state when the heme is oxidized or reduced, respectively. The HS-AFM study for PccDH in situ gives us sequential views of the domain motions between the closed and open states during catalysis involving the cycle between the oxidation and reduction of the heme. The movement of the DH domain of PccDH is centered on the CYT domain during the cycle of the closed–open–closed states. The DH domain apparently moves back to similar positions approximately within 1 s, as estimated from the sequential HS-AFM images. The previous study using the small-angle X-ray scattering analysis indicates that in the presence of the inhibitor molecules, the conformational population of the open state increases relative to that of the closed state in both MtCDH and NcCDH. The small angle neutron scattering study revealed that glycosylated NcCDHIIIA, which has a C-terminal carbohydrate binding module, has an extended conformation in the fully oxidized state. In the present study, we can directly observe single-molecule domain motion of deglycosylated PccDH triggered by binding of cellubiose. Therefore, the flip-flop domain motions of CDH appear to be strongly related to shuttling of electrons from cellubiose to a terminal electron acceptor.

Conclusions

In this work, our data provide the first demonstration that PccDH can be immobilized on a flat gold surface via the CYT domain with a controlled orientation via the heme–heme pocket interaction and can still remain catalytically active toward cellubiose oxidation. In addition, this strategy enables the two domains of anchored PccDH to retain a degree of freedom in terms of interdomain motions required for the catalysis. As a result, we have directly visualized the interdomain flip-flop motions of PccDH triggered by the binding of cellubiose in situ using the HS-AFM technique. This observation implies that the flip-flop motion of the two domains in PccDH regulates the electron transfer process to the terminal electron acceptor, thereby harmonizing the entire electron relay through the enzymes in the oxidative process of cellulose depolymerization in vivo. The present method visualizing the domain motion of CDH on the flat surface by HS-AFM should be useful for investigating the dynamic motion of other multi-domain redox-active heme enzymes such as P450BM3. Our approach demonstrates that HS-AFM will provide novel insight into understanding the dynamic mechanism of enzyme catalysis including electron transfer as well as structural changes coupled with chemical reactions and photoactivation.

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References


