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Stereoselective synthesis of γ-hydroxynorvaline through combination of organo- and biocatalysis
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Robert C. Simon, Eduardo Busto, Joerg H. Schrittwieser, Johann H. Sattler, Jörg Pietruszka, Kurt Faber and Wolfgang Kroutil*

An efficient route for the synthesis of all four diastereomers of PMP-protected α-amino-γ-butyrolactone to access γ-hydroxynorvaline was established. The asymmetric key steps comprise an organocatalytic Mannich reaction and an enzymatic ketone reduction. Three reaction steps could be integrated in a one-pot process, using 2-PrOH both as solvent and as reducing agent. The sequential construction of stereogenic centres gave access to each of the four stereoisomers in high yield and with excellent stereocontrol.

Progressive legislative regulations with respect to sustainability, safety and quality improvements have changed the focus of contemporary chemical research to designing more economic processes. In this context, (asymmetric) catalysis has become the method of choice to replace stoichiometric amounts of reagents and auxiliaries, thereby facilitating an efficient and energy saving process. While each of the catalysis sub-fields (metal-, organo- and biocatalysis) has reached an highly cooperative fashion. For example, numerous reports have been published based on the combination of transition metals and catalytically active enzymes ADHs transformed racemic amino-ketoester rac-1, which is readily available by a proline catalysed Mannich reaction. A set of homo- and heterologously expressed alcohol dehydrogenases (ADHs) were assayed on analytical scale (5.6 mg, 20 mM) employing 2-propanol for NAD(P)+ regeneration. Asymmetric reduction of the carbonyl group of amino-ketoester 1 afforded the corresponding diastereomers of alcohol 2, which cyclised spontaneously to the desired 2-amino-lactones 3 (Scheme 1).

In our study we focused initially on the bioreduction of racemic amino-ketoester rac-1, which is readily available by a proline catalysed Mannich reaction. A set of homo- and heterologously expressed alcohol dehydrogenases (ADHs) were assayed on analytical scale (5.6 mg, 20 mM) employing 2-propanol for NAD(P)+ regeneration. Asymmetric reduction of the carbonyl group of amino-ketoester 1 afforded the corresponding diastereomers of alcohol 2, which cyclised spontaneously to the desired 2-amino-lactones 3 (Scheme 1).

A number of ADHs transformed rac-1 into the corresponding α-amino-lactones 3 with high conversion (selected results in Table 1; more details in the ESI†). For example, the (5S)-configured diastereomers (3S,5S)-3 and (3R,5S)-3 were accessible by the enzymes ADH-A15† (entry 1) and evo-1.1.03015c (entry 2), whereas the (5R)-diastereomers (3S,5R)-3 and (3R,5R)-3 were formed utilising evo-1.1.200† (entry 5). The enzymes ADH-T15c (entry 3) and ADH-Ls15d (entry 4) gave excellent results, too, with conversions from natural sources and shows anti-diabetic properties. The widespread occurrence of the γ-butylolactone unit has led to the development of a number of stereoselective syntheses mainly based on organo-catalysis, while only few biocatalytic strategies have been developed. Most of the procedures either require multiple steps, are auxiliaries-assisted, or based on exchiral pool precursors. Hence, access to only one or two out of four stereoisomers is provided. However, as the biological activity of bioactive compounds is related often to one specific unknown stereoisomer, a convenient access to all four diastereomers, e.g. for SAR-studies, is desired.

In this communication, a novel combination of organo- and biocatalysis is reported, which provides access to optically pure α-amino-γ-butyrolactones either in a step-wise fashion or in a cascade reaction.

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The next step was the synthesis of all four stereoisomers of amino-lactone 3; for this purpose, the biocatalysed reduction of ketoester 1 was optimised using alcohol dehydrogenases ADH-A and evo-1.1.200. Investigating 2-PrOH as a cosolvent revealed that a substrate concentration of 30 mM (7.95 mg mL\(^{-1}\), 10 vol% 2-PrOH) led to completion of the reaction in the case of the (S)-selective ADH-A within 24 h (for details, see ESI\(^t\)). The (R)-selective enzyme evo-1.1.200 transformed even 50 mM (13.3 mg mL\(^{-1}\), 30 vol% 2-PrOH) to completion within 24 h. The optimum pH was determined to be 6.3 in order to keep the partial racemisation of the optically pure Mannich adduct 1 at a minimum (see ESI\(^t\)). In the subsequent semi-preparative scale transformations [56 mg (S)-1] > 99% conversion was achieved; however, ring closure was incomplete, as mainly the optically pure amino-alcohol 2 rather than the lactone 3 was isolated (ca. 50–70%; see ESI\(^t\)). Assuming that the corresponding methyl ester of 1 would undergo faster lactonisation than the ethyl analogue, a transesterification step (HCl–MeOH) was integrated into the reaction sequence. The optimised conditions finally allowed conversion of the pure amino-ester \((\text{R})\)-(3S)-1 (up to > 300 mg), readily accessible by a proline-catalysed Mannich reaction of acetone and imine 4, into the corresponding lactones 3. The \textit{syn-} and \textit{anti-}diastereomers were isolated in excellent yields (78–82%) and with remarkable stereocontrol (de > 98 : 2 and ee > 99% in all cases; see Scheme 2).

As an alternative to the conventional step-by-step synthesis (Scheme 2) two sequential approaches were established. In a first approach, aldimine 4 was formed in 2-PrOH, which also served as a solvent for the subsequent Mannich reaction and as...
the reducing agent for the enzyme-mediated reduction (after dilution with buffer solution). All three steps were accomplished in one pot, after which the transesterification-lactonisation cascade was performed in a separate vessel. The whole sequence afforded the syn-configured lactone (3S,5R)-3 in a remarkable 47% yield with a d.r. of 86:14 in favour of the syn-configuration (Scheme 3a: 5 steps in total, 2 pots, 90% average yield per step). The diminished d.r. in comparison to the previous results can be rationalised by the usage of 2-ProH for the proline-catalysed reaction, which under these conditions afforded (S)-1 with 72% ee.

In a second approach, the Mannich reaction was carried out with the preformed aldimine 4 in acetone as solvent, and amino-ketoester (R)-1 was obtained at prolonged reaction time (16 h) with >99% conversion and in optically pure form (ee > 99%). After evaporation of the solvent, the crude product was subjected to reduction by ADH-A in buffer-2-ProH. The final transesterification-lactonisation cascade furnished the diastereomerically pure amino lactone (3S,5R)-3 in 51% yield with an excellent d.r. of 99:1 (Scheme 3b: 4 steps, 2 pots, 85% average yield per step).

Final PMP-deprotection of the diastereomerically pure α-amino-lactone 3 was achieved employing TCCA (trichloroisocyanuric acid) in MeOH,4b providing the hydrochloride salt of 3 with 72% ee.

Notes and references

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15 ADHs employed during the studies: ADH-A originates from Rhodococcus ruber; see: (a) K. Edegger, C. C. Gruber, T. M. Poesl, S. R. Wallner, I. Lavandera, K. Faber, F. Niehaus, J. Eck, R. Oehrlein, A. Hafner and W. Kroutil, Chem. Commun., 2006, 2402; (b) Eozymes were obtained from Evocatal (origin not stated); (c) ADH-T was obtained from Codexis Inc. (origin not stated); (d) ADH-LS originates from Leifsonia sp.; see K. Inoue, Y. Makino and N. Itoh, Appl. Environ. Microbiol., 2005, 3633; (e) ADH-LB originates from Lactobacillus brevis; see: W. Hummel and B. Riebel, Biotecnol. Lett., 2003, 25, 51.
17 The ring-closure was found to be promoted by the GC injector pretending full conversion to the lactone 3. In an additional control experiment the purified alcohol 2 was subjected to GC and GC-MS analysis, showing only the signal for the lactone 3.