


Cite this: *RSC Adv.*, 2025, 15, 1721

# Incredible use of plant-derived bioactives as anticancer agents

Kiran Kangra,<sup>a</sup> Saloni Kakkar,<sup>a</sup> Vineet Mittal,<sup>a</sup> Virender Kumar,<sup>b</sup> Navidha Aggarwal,<sup>c</sup> Hitesh Chopra,<sup>d</sup> Tabarak Malik<sup>id</sup>\*<sup>ef</sup> and Vandana Garg<sup>\*a</sup>

Cancer is a major global concern. Despite considerable advancements in cancer therapy and control, there are still large gaps and requirements for development. In recent years, various naturally occurring anticancer drugs have been derived from natural resources, such as alkaloids, glycosides, terpenes, terpenoids, flavones, and polyphenols. Plant-derived substances exhibit their anticancer potential through antiproliferative activity, cytotoxicity, apoptosis, angiogenesis and cell cycle arrest. Natural compounds can affect the molecular activity of cells through various signaling pathways, like the cell cycle pathway, STAT-3 pathway, PI3K/Akt, and Ras/MAP-kinase pathways. Capsaicin, ouabain, and lycopene show their anticancer potential through the STAT-3 pathway in breast, colorectal, pancreatic, lung, cervical, ovarian and colon cancers. Epigallocatechin gallate and emodin target the JNK protein in skin, breast, and lung cancers, while berberine, evodiamine, lycorine, and astragalin exhibit anticancer activity against breast, liver, prostate, pancreatic and skin cancers and leukemia through the PI3K/Akt and Ras/MAP-kinase pathways. *In vitro/in vivo* investigations revealed that secondary metabolites suppress cancer cells by causing DNA damage and activating apoptosis-inducing enzymes. After a meticulous literature review, the anti-cancer potential, mode of action, and clinical trials of 144 bioactive compounds and their synthetic analogues are included in the present work, which could pave the way for using plant-derived bioactives as anticancer agents.

Received 14th July 2024  
Accepted 31st December 2024

DOI: 10.1039/d4ra05089d

rsc.li/rsc-advances

## 1 Introduction

Among the non-communicable diseases, cancer is the second most life-threatening disease after cardiovascular diseases.<sup>1</sup> It is caused by a combination of genetic factors, environmental stress on cellular activity,<sup>2</sup> obesity, poor diet, excessive alcohol intake, smoking, and vitamin B<sub>12</sub> deficiency.<sup>3</sup> According to Sung *et al.*, transitional cases (64%) would increase significantly more than transitioned cases (32%), reaching 28.4 million cases globally in 2040, with an increase of 47% cases from 2020. Approximately 10 million people would die from cancer in 2024, out of which approximately 19.3 million would be new cases.<sup>4</sup> According to the site of occurrence, there are 131 different types of cancers, including skin cancer, lung cancer, oral cancer, and breast

cancer.<sup>5</sup> It is predicted that there would be 2.3 million more cases of female breast cancer (11.7%), followed by lung (11.4%), colorectal (10.1%), prostate (7.3%), and stomach (5.6%) cancers. Different types of cancers are caused by a variety of variables. Particularly, in the case of skin cancer, ozone depletion, melanin and microbial impact are responsible for its onset.<sup>6</sup> Lung cancer is primarily caused by smoking but can also occur in non-smokers owing to other factors, like exposure to radon gas or secondhand smoke. Prostate cancer affects men and is one of the most common cancers in older men. Colorectal cancer affects the colon or rectum and is more common in older adults. It can be developed due to hereditary reasons or develop sporadically. Breast cancer occurs primarily in women but can also affect men. Thus, the pathophysiology of cancer involves a multitude of genetic, molecular, and environmental factors. Cancer arises from mutations in the DNA of cells that disrupt normal control mechanisms governing cell growth and division. These mutations can be inherited or acquired over time due to exposure to carcinogens such as chemicals, radiation, or viruses.

### 1.1 Pathophysiology of cancer

Cancer is a four-step process involving mutation along with cell's proliferative, survival, invasion, and metastatic capacities. In cancer, the cell's genetic system (DNA) and anti-tumor genes are suppressed by environmental factors or unhealthy diet, smoking, drinking obesity *etc.* Tumor suppressor gene inactivation is

<sup>a</sup>Department of Pharmaceutical Sciences, Maharshi Dayanand University, Rohtak, 124001, India. E-mail: kirankangra90@gmail.com; drsaloni.pharma@mdurohtak.ac.in; drvineet.pharma@mdurohtak.ac.in; drvandana.pharma@mdurohtak.ac.in

<sup>b</sup>College of Pharmacy, Pandit Bhagwat Dayal Sharma University of Health Sciences, Rohtak, 124001, India. E-mail: sachdeva.virender5@gmail.com

<sup>c</sup>MM College of Pharmacy, Maharishi Markandeshwar (Deemed to be University), Mullana, Ambala 133207, Haryana, India

<sup>d</sup>Department of Biosciences, Saveetha School of Engineering, Saveetha Institute of Medical and Technical Sciences, Chennai 602105, Tamil Nadu, India

<sup>e</sup>Department of Biomedical Sciences, Jimma University, Jimma, Ethiopia. E-mail: tabarak.malik@ju.edu.et

<sup>f</sup>Division of Research & Development, Lovely Professional University, Phagwara, Punjab-144411, India



a natural physiological reaction of the organism and the cancer develops when this reaction becomes pathologic.<sup>7</sup> Except for histological types, almost all cancers share basic pathogenesis. From extensive research, it is evident that the genetic system is involved in the development of malignant tumors. Due to inhibition of angiogenesis and alteration of cells.<sup>8</sup>

The initial stage in the progression of cancer involves the occurrence of a mutation and the subsequent formation of a tumor. This process occurs when a genetic alteration triggers a mutation within a cell, leading to the growth of tumor cells. Following this, the mutation induces cell proliferation and the advancement of the tumor as the mutated cells rapidly multiply and divide, ultimately becoming dominant within the tumor cell population.<sup>9</sup> Subsequently, clonal selection occurs among the proliferating cells, resulting in the generation of a new clone of rapidly growing cells with distinct characteristics. This step is repeated throughout the development of the tumor. Finally, metastasis occurs, wherein cancer cells detach from the primary tumor and travel through either the bloodstream or the lymphatic system to distant areas of the body.<sup>10</sup> Consequently, these cells continue to multiply in the new locations, ultimately giving rise to new tumors composed of cells that bear resemblance to the original tissue. The propensity of tumors to metastasis is a major factor in the lethality of some malignancies, such as pancreatic and uveal cancers.<sup>11</sup> The basic pathophysiology of cancer is described in Fig. 1.

## 2 Review methodology

A concise summary of the methodology employed in this review is shown in Fig. 2. A comprehensive search was conducted using various search engines such as PubMed, Google Scholar,

ScienceDirect, Scopus, Web of Science, and Chemical Abstracts. The search utilized different keywords including “anti-cancer”, “phytochemical”, “plant bioactive”, “clinical trials”, “mechanism of action”, and more. Irrelevant, duplicate, and incomplete data were excluded, while the literature pertaining to the *in vitro* or *in vivo* anticancer potential of plant-based bioactives was included by studying the 1600–1700 review and research article. Additionally, this review focused on articles that described the mechanism of action and clinical trial data for the anti-cancer potential of herbal compounds. The present article primarily reviews data published within the past decade.

## 3 Role of traditional plants and derived bioactives in cancer

Plants are used to cure many ailments and natural or plant-based medications are preferred by 60–70 percent of the

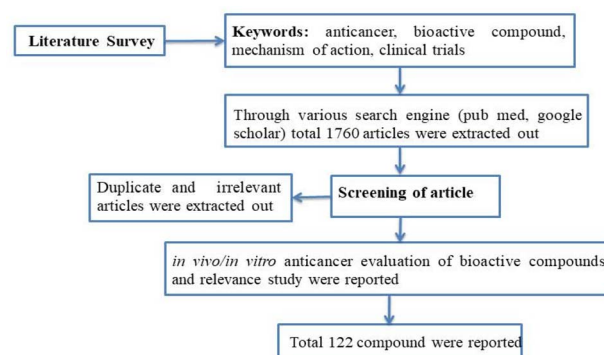


Fig. 2 Flow chart summarizing the review methodology.



Fig. 1 Flow-chart indicating the basic pathophysiology of different types of cancers.



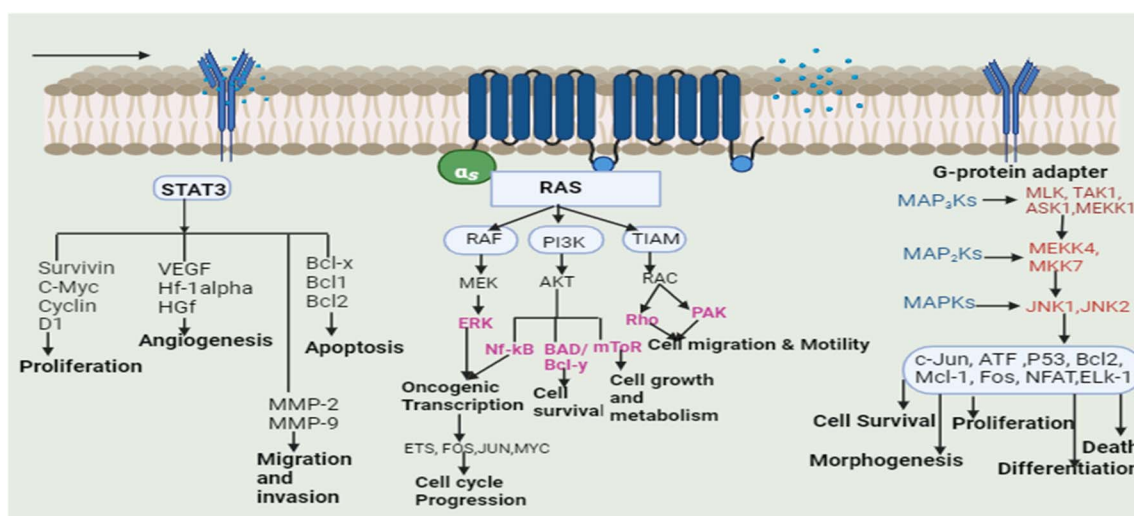
population over synthetic medicines for one reason or the other. These plants may aid the patient resistance to sickness by arbitrating physiological homeostasis and retraining the body tissues.<sup>12</sup> Based on the traditional uses of plants and scientific reports, a lot of research has also been dedicated to the study of plants in order to cure cancer, and several plants have been successfully used in the treatment of cancer.<sup>13</sup> Isolated phytoconstituents from these plants such as vincristine, vinblastine, chlorogenic acid, gingerol, apigenin, catechin, gallic acid, cinnamic acid, and podophyllotoxin, along with their derivatives and analogues, are used for the treatment of cancer by inhibiting several signaling pathways. Different types of tumors have altered cell signaling pathways (cell death pathways: apoptosis and autophagy, embryonic developmental pathways: Notch, Wnt, Hedgehog, Janus kinase pathway, signal transducer and activator of the transcription factor pathway and RAF/MAPK pathway). Cells integrate the signals received from various growth factors and receptors to control different cellular functions, including cell motility, differentiation, architecture, and polarity. Signalling pathways control cellular growth and induce various alterations in various cell types.<sup>14</sup>

The transcription (STAT3) pathway with signal transducers and activators is a major intrinsic pathway in cancer development (Fig. 3). It transmits intracellular signals that are normally generated at cell surface receptors to the nucleus. STAT3 activation involved a number of human tumors, including haematological and solid tumors. The evidence suggests that oncogenic cell transformation activates STAT-3, providing the survival signal. The dysfunctioning of STAT-3 during mammary gland involution demonstrates that it has proapoptotic functions. Functioning STAT-3 can prevent apoptosis in most cells.

These effects are arbitrated by STAT-3-regulated cell survival gene products, *i.e.* Bcl Bcl1, Bcl-2, Survivin, Mcl-1, and cIAP2. Thus, inhibiting the STAT-3 activation can reduce the activity of these gene products, thereby increasing apoptosis.<sup>15</sup> Furthermore, the master protein kinases known as c-Jun N-terminal kinases (JNKs) control a variety of physiological processes, such as inflammatory reactions, morphogenesis, cell proliferation, differentiation, survival, and death. It is becoming clear that persistent JNK activation contributes to cancer development and progression. Further, RAS proteins can interact with other well-known effectors such as phosphatidyl inositol 3-kinases (PI3Ks) *via* the RAF/MAPK pathway (PI3Ks). The interaction of different RAS proteins with PI3Ks could lead to DNA damage, and finally, to tumor development.

In the present study, different bio-actives from various categories that are reported to possess anti-cancer potential against various cell lines and in experimental animals are summarised.

**Alkaloids:** these are the largest group of phytochemicals with a heterocyclic ring structure and at least one nitrogen atom. To distinguish various alkaloids, a categorization based on biosynthetic pathways is commonly used. Alkaloids can be found in all types of plants, although they are most prevalent in the Ranunculaceae, Leguminosae, Papaveraceae, Menispermaceae, and Loganiaceae families.<sup>16</sup> Vinca alkaloids (vincristine, vinblastine, vinorelbine, vindesine, and vinflunine) were the first microtubule-targeting agents (MTAs) and approved for clinical use in hematological and lymphatic neoplasms.<sup>17</sup> Various alkaloids from plant sources and their synthetic analogues with cytotoxicity on different cell lines are reported in Table 1 and the structures of isolated alkaloids are shown in Fig. 4.



**Fig. 3** Signal transducers and activators of the transcription (STAT3) pathway, Ras/Raf/MAPK (mitogen-activated protein kinase) pathway, and JNK (c-Jun N-terminal kinases) pathway in the development of cancer/tumor [JNK (c-Jun N-terminal kinases), signal transducers and activators of transcription (STAT3), vascular endothelial growth factor (VEGF), hepatocyte nuclear factor-1 alpha (HF-1 alpha), hepatocyte growth factor (HGF), B-cell lymphoma (Bcl), matrix metalloproteinases (MMP), RAF (rapidly accelerated fibrosarcoma), MEK/MAPK (mitogen activated protein kinase), ERK (extracellular signal-regulated kinase), PI3K (phosphatidylinositol-3 kinase), Akt (Akt kinase), mechanistic target of rapamycin (mTOR), nuclear factor- $\kappa$ B (Nf- $\kappa$ B), Rho GTPases (Rho), p21-activated kinases (PAK), early tumour shrinkage (ETS), FOS protooncogene (FOS), JUN protooncogene (JUN), mixed-lineage kinase (MLK), transforming growth factor- $\beta$ -activated kinase 1 (TAK1), apoptosis signal-regulating kinase 1 (ASK1), nuclear factor of activated T cells (NFAT), and ETS-like protein 1 (ELK1)].



**Table 1** Various isolated alkaloids, their biological sources and reported IC<sub>50</sub> against different types of cancer cell lines

| Isolated compound | Alkaloid (biological source)                     | Cancer cell line along with IC <sub>50</sub> (nM) value | References |
|-------------------|--------------------------------------------------|---------------------------------------------------------|------------|
| 1                 | <i>Berberis aetnensis</i> (Berberidaceae)        | MCF-7-230                                               | 18 and 19  |
| 2                 |                                                  | HepG2-170                                               |            |
| 3                 |                                                  | LNCaP-190                                               |            |
| 4                 |                                                  | PC-3165                                                 |            |
| 5                 |                                                  | MHCC97-L-400                                            |            |
| 6                 |                                                  | MDA-MB21-250                                            |            |
| 7                 |                                                  | HTB-94-200                                              |            |
| 8                 |                                                  | SMMC-7721-180                                           |            |
| 9                 | <i>Evodia rutaecarpa</i> (Rutaceae)              | MDA-MB-435-49                                           | 20         |
| 10                |                                                  | HCT116-90                                               |            |
| 11                |                                                  | U20S-26                                                 |            |
| 12                |                                                  | Panc-1-39                                               |            |
| 13                |                                                  | PC-3-65                                                 |            |
| 14                |                                                  | HL-60-60                                                |            |
| 15                |                                                  | Saos-2-95                                               |            |
| 16                | <i>Sophora flavescens</i> (Fabaceae)             | A549-200                                                | 21 and 22  |
| 17                |                                                  | HepG2-250                                               |            |
| 18                |                                                  | Panc-1-170                                              |            |
| 19                |                                                  | CCRFCEM-250                                             |            |
| 20                |                                                  | SGC7901-209                                             |            |
| 21                |                                                  | PC-3-170, DU145-175                                     |            |
| 22                | <i>Galanthus nivalis</i> (Amaryllidaceae)        | A-431-90                                                | 23 and 24  |
| 23                |                                                  | A549-255                                                |            |
| 24                |                                                  | BCA-1-200                                               |            |
| 25                |                                                  | B16F10-200                                              |            |
| 26                |                                                  | CEM-140                                                 |            |
| 27                |                                                  | HT29-230                                                |            |
| 28                |                                                  | HeLa-120                                                |            |
| 29                |                                                  | HepG2-156                                               |            |
| 30                |                                                  | Hs683-130                                               |            |
| 31                |                                                  | HL-60-145                                               |            |
| 32                |                                                  | B16F10-250                                              |            |
| 33                |                                                  | CEM-180                                                 |            |
| 34                | <i>Crinum bulbispermum</i> (Amaryllidaceae)      | BCA-1-220                                               | 25 and 26  |
| 35                |                                                  | A549-275                                                |            |
| 36                |                                                  | HT29-280                                                |            |
| 37                |                                                  | U373-280                                                |            |
| 38                | <i>Boophone disticha</i> (Amaryllidaceae)        | HL-60-120                                               | 27         |
|                   |                                                  | HeLa-150                                                |            |
|                   |                                                  | G-361-250                                               |            |
|                   |                                                  | MCF-7-200                                               |            |
| 39                | <i>Hymenocallis littoralis</i> (Amaryllidaceae)  | K562-280                                                | 28–30      |
|                   |                                                  | HL-60-150                                               |            |
|                   |                                                  | K562-180                                                |            |
|                   |                                                  | PC-3M-200                                               |            |
| 40                | <i>Amaryllis belladonna</i> L., (Amaryllidaceae) | A549-280                                                | 30         |
|                   |                                                  | OE21-220                                                |            |
|                   |                                                  | B16F10-290                                              |            |
|                   |                                                  | U373-360                                                |            |
| 41                | <i>Nerine bowdenii</i> (Amaryllidaceae)          | HL-60-200                                               | 31 and 32  |
|                   |                                                  | U937-290                                                |            |
|                   |                                                  | K562-360                                                |            |
|                   |                                                  | MOLT-4-270                                              |            |
| 42                | <i>Piper nigrum</i> L. (Piperaceae)              | LXFL 529L-240                                           | 31 and 32  |
|                   |                                                  | DU145-150                                               |            |
|                   |                                                  | HT-29-180, Caco-2-200, SW480-220                        |            |
|                   |                                                  | HRT-18-220                                              |            |
| 43                | <i>Sanguinaria canadensis</i> (Papaveraceae)     | A549-140                                                | 33         |
|                   |                                                  | DU145-210                                               |            |
|                   |                                                  | BEL-7402-280                                            |            |



Table 1 (Contd.)

| Isolated compound | Alkaloid (biological source)                    | Cancer cell line along with IC <sub>50</sub> (nM) value | References |
|-------------------|-------------------------------------------------|---------------------------------------------------------|------------|
| 44                | <i>Stephania tetrandra</i> (Menispermaceae)     | Hela-180                                                | 34 and 35  |
| 45                |                                                 | BGC-823-180                                             |            |
|                   |                                                 | HCT116-260                                              |            |
| 46                | <i>Piper arborescens</i> (Piperaceae)           | Hep G2-210                                              | 36 and 37  |
|                   |                                                 | A549-160                                                |            |
|                   |                                                 | KB-140                                                  |            |
|                   |                                                 | A549-180                                                |            |
| 47                | <i>Plumbago zeylanica</i> L. (Plumbaginaceae)   | P388-180                                                | 38 and 39  |
|                   |                                                 | HT29-260                                                |            |
|                   |                                                 | MG63-160                                                |            |
| 48                | <i>Nigella sativa</i> (Iridaceae)               | MCF7-230                                                | 40         |
| 49                |                                                 | PC3-300                                                 |            |
| 50                | <i>Cyrtanthus contractus</i> (Amaryllidaceae)   | LL/2-260                                                | 41         |
| 51                |                                                 | HeLa-280                                                |            |
|                   |                                                 | HeLa-200                                                |            |
| 52                | <i>Capsicum annum</i> (Solanaceae)              | MCF7-290                                                | 42         |
|                   |                                                 | A431-260                                                |            |
|                   |                                                 | HCT LoVo-250                                            |            |
|                   |                                                 | MCF7-200, MDA-MB231-240                                 |            |
| 53                | <i>Broussonetia papyrifera</i> (L.) (Moraceae)  | LNCaP-180                                               | 27         |
|                   |                                                 | HL-60-255                                               |            |
|                   |                                                 | PANC1-200                                               |            |
| 54                | <i>Narcissus jonquilla</i> (Amaryllidaceae)     | BEL-7402-185                                            | 43         |
|                   |                                                 | Hela-150                                                |            |
|                   |                                                 | PC3-290                                                 |            |
|                   |                                                 | LoVo-300                                                |            |
| 55                | <i>Hymenocallis littoralis</i> (Amaryllidaceae) | A549-350                                                | 44         |
|                   |                                                 | MCF-7-380                                               |            |
|                   |                                                 | PANC1-250                                               |            |
|                   |                                                 | MV4-11-110                                              |            |
|                   |                                                 | U87-160                                                 |            |
|                   |                                                 | MCF7-145                                                |            |
|                   |                                                 | OVCAR3-135                                              |            |
| 60                |                                                 | Hep G2-230                                              |            |
| 61                |                                                 | PANC1-280                                               |            |
| 62                |                                                 | U87-260                                                 |            |

Berberine (1) and its seven synthetic isomers having different substituents, such as [propyl benzene (1), 1-chloro-4-ethylbenzene (2), butane-1,1-diyl dibenzene (3), 4,4-diphenylbutyl (4), phenyl-pentyl (5), phenyl-hexyl (6), 5,5-diphenyl-amyl (7) and 6,6-diphenyl-hexyl (8)], were screened for prostate cancer, lung cancer, liver cancer, and chondrosarcoma and these were found to work in a variety of ways to prevent cancer. It inhibited cyclin D1 and E1 in lung cancer and CDK4 expression and modulating cyclin D1 in colorectal cancer and hepatoma cancer. Berberine upregulated the level of p53 and p21 in chondrosarcoma by regulating the PI3K/Akt and p38 signaling pathways.<sup>45</sup>

Evodiamine (9) and its six derivatives with varied substituents [4-chlorobenzoyl (10), 12-chloroevodiamine (11), 3-fluoro-evodiamine (12), 10-hydroxyevodiamine (13), 3-fluoro-10-hydroxyevodiamine (14) and 3-amino-10-hydroxyevodiamine (15)] were screened for the treatment of colon cancer, osteosarcoma, pancreatic carcinoma, prostate cancer, leukemia and breast cancer. By inhibiting the caspase inhibitor evodiamine

inhibits cervical cancer. Caspase inhibition causes alteration in Bax and Bcl-2 balance, which decreases apoptosis.<sup>46</sup> It suppresses the liver cancer by inducing apoptosis and inhibiting the PI3K/Akt pathway.<sup>47</sup>

Matrine (16) and its five derivatives with varying substituents, such as [11-(3-methylbenzyl)dodecahydro-1*H*,5*H*,10*H*-dipyrido[2,1-*f*:3',2',1'-*ij*][1,6]naphthyridin-10-one (17), 4-methyl-10-oxotetradecahydro-1*H*,5*H*-dipyrido[2,1-*f*:3',2'-*ij*][1,6]naphthyridin-4-ium iodide (18), (11(*Z*)-11-(6-bromonaphthalen-2-yl)(hydroxy)methylene)dodecahydro-1*H*,5*H*,10*H*-dipyrido[2,1-*f*:3',2',1'-*ij*][1,6]naphthyridin-10-one (19), 11-(hydroxy(naphthalen-2-yl)methyl)dodecahydro-1*H*,5*H*,10*H*-dipyrido[2,1-*f*:3',2',1'-*ij*][1,6]naphthyridin-10-one (20) and 11-(hydroxy(6-methoxynaphthalen-2-yl)methyl)dodecahydro-1*H*,5*H*,10*H*-dipyrido[2,1-*f*:3',2',1'-*ij*][1,6]naphthyridin-10-one (21)], showed anticancer potential against lung cancer, breast cancer, liver cancer, prostate cancer, leukemia and sarcoma. It causes caspase-mediated cell death in lung cancer by impeding the G<sub>1</sub>/G<sub>0</sub> phase of the cell cycle.<sup>48</sup> Matrine showed anticancer activity



Fig. 4 Structures of isolated alkaloids.

against pancreatic cancer by inducing ROS generation, and induced death.<sup>49</sup> Crinine-type (22) alkaloids and their fourteen derivatives [haemanthamine (23), haemanthidine (24), vitattine

(25), hydroxyvitattine (26), crinamine (27) 11-O-acetylcrinamine (28) 11-O-methylcrinamine (29), 6-hydroxycrinamine (30), hamayne (31), 3-O-acetylhamayne (32), 8-O-demethylmaritidine



(33), papyramine (34), dihydrocrinamine (35) and dihydrohamayne (36)] are also promising therapeutic candidates for the treatment of apoptosis-resistant tumors, particularly glioblastoma. The crinine-type alkaloid inhibits glioblastoma cell proliferation *via* cytostatic effects resulting from the rigidification of the actin cytoskeleton. Bulbispermine (37) showed anticancer activity against glioblastoma and leukemia by inhibiting apoptosis resistance.<sup>25</sup> Distichamine (38) in leukemia alter the cell cycle and induce death by activating the caspase 3 and 7.<sup>50</sup> Lycorine (39) and amarbellisine (40) reduce Mcl-1 at the translational level, which causes cell death in leukemia cells. Lycorine promotes the intrinsic apoptotic cascade in bladder cancer by decreasing the PI3K–Akt pathway and boosting the expression of the PTEN protein, which acts as a negative regulator of p-Akt.<sup>51</sup> ungeremine (41) showed cytotoxic effects against leukemia by inhibiting cell proliferation through caspase activation, matrix metalloproteinases (MMP) modification, and also increasing ROS production.<sup>52</sup> Piperine (42) inhibits cell proliferation by activation of apoptotic signalling pathways, modulation of ER stress and induction of detoxification of enzymes.<sup>53</sup> Sanguinarine (43) and its one derivative dihydrosanguinarine (44) showed their anticancer potential by suppressing the abnormally active signal transduction pathways, cell apoptosis, and cancer cell proliferation.<sup>54</sup> Tetrandrine (45) showed anti-cancer properties against lung, colon, bladder prostate, and many more, as shown in Table 1. Tetrandrine's anticancer properties may be linked to autophagy, cell cycle arrest, alleviate metastasis and suppression of tumor cell proliferation.<sup>55</sup> Piplartine (46) caused G2/M cell cycle arrest, followed by mitochondrial-dependent apoptosis, as shown by chromatin condensation and inter-nucleosomal DNA breakage.<sup>56</sup> Plumbagin (47) showed its anticancer potential through the NF- $\kappa$ , STAT3, and Akt regulatory signaling pathways. It was also a potent ROS inducer, a suppressor of cellular glutathione, and a novel proteasome inhibitor generating DNA double-strand breaks *via* oxidative DNA base damage.<sup>57</sup>

Thymoquinone (48) and its two derivatives [thymoquinone (49) and dithymoquinone (50)] have anti-cancer properties through a variety of mechanisms, including selective antioxidant activity, DNA structural interference, effects on carcinogenic signaling molecules/pathways, and immunomodulation.<sup>58</sup> Narciprimine's (51) effects on DNA topoisomerase have also been studied. The findings demonstrated that narciprimine was dose-dependently efficacious in DNA topoisomerase processes. The potential of this alkaloid to interfere with topoisomerase was somewhat associated with anticancer activity measured in HeLa, MCF-7, and A341 cells.<sup>41</sup> Multiple mechanisms were involved in capsaicin's (52) anti-cancer activity, including increased intracellular calcium, inhibition of p53, STAT3 and nuclear factor B.<sup>59</sup> Norchelerythrine (53) works as an anticancer agent by various methods, including apoptosis, inhibiting aromatase, disrupting tubulin aggregation, inhibiting topoisomerase, and inhibiting ER.<sup>60</sup> In the prostate and breast cancer cells, narciclasine (54) causes inactivation of mitochondrial membrane potential, cytochrome release and caspase activations.<sup>27</sup> Pancratistatin (55) and its seven synthetic analogues with varied substituents [JCTH-1 (56),

JCTH-2 (57), JCTH-3 (58), JCTH-4 (59), SVTH-5 (60) SVTH-6 (61) and SVTH-7 (62)] inhibit tumor xenograft growth by disrupting mitochondrial activity and by activating the intrinsic apoptotic pathway. SVTH-7 inhibits mitochondrial complex II and III, reducing pro-apoptotic effects on cancer cells and on mitochondria.<sup>44</sup>

### 3.1 *In vivo* anti-cancer studies of alkaloids

In the drug development process, preclinical data give complete information, including preliminary efficacy, toxicity, pharmacokinetics, and safety of potential lead compound. This information can be used to determine whether or not a compound should be pursued further for clinical trials. In this context, various *in vivo* studies reporting the anti-cancer evaluation of alkaloids have also been summarised as follows:

Berberine anticancer activity against colorectal cancer was tested in a xenograft model of BALB/c nude mice. Mice were injected with KM12C cell sublines, shCtrl, and shRXR. After the tumor had grown, the infected mice were given berberine (10 mg kg<sup>-1</sup>). Berberine reduced the length of the tumor which could be due to the induction of nuclear-catenin degradation, significantly reducing endogenous c-Cbl, Ki67, Cdc2, c-Myc, and CIP1. Berberine also shows its activity by inhibiting the  $\beta$ -catenin signaling pathway.<sup>61</sup> A xenograft model was used to test berberine's anticancer activity against endometrial cancer. Mice were injected with HEC-1-A. When the tumor had grown, mice were divided into three groups. Groups were given either 0.5% MC (vehicle control) or berberine (50 mg kg<sup>-1</sup>, p.o.qd or 100 mg kg<sup>-1</sup>, p.o.qd), orally. Berberine treatment significantly reduced the invasion of HEC-1-A cells at 50 mg kg<sup>-1</sup> and 100 mg kg<sup>-1</sup>,<sup>62</sup> and in lung cancer, it showed its potential at 200 mg kg<sup>-1</sup> and 25 mg kg<sup>-1</sup> in nude mice.<sup>63</sup> The anticancer activity of matrine against lung cancer was tested in a xenograft model of BALB/c nude mice by inserting the LA795 cell. The infected mice were given matrine (80 mg kg<sup>-1</sup>) and a vehicle. It reduced the length of the tumor by regulating transmembrane protein 16A.<sup>64</sup> Matrine anticancer activity in breast cancer was investigated by inserting the C57BL cell subcutaneously. Then 50 mg per kg matrine was injected once a day at an early stage of cancer. Mice were forfeited after 21 days. The tumor was collected and evaluated. The results indicated that matrine reduced breast cancer angiogenesis by inhibiting the Wnt/ $\beta$ -catenin signaling pathway.<sup>65</sup> Piperine anticancer activity against breast cancer was investigated in BALB/c mice. Then, 2  $\times$  10<sup>5</sup> EEMT6/P cells were injected subcutaneously. Following this, 25 mg per kg matrine was injected once a day at an early stage of cancer. Mice were forfeited after 14 days. The tumor was collected and evaluated.<sup>66</sup> Lycorine anticancer activity against prostate cancer was tested in a xenograft model of BALB/c nude mice by inserting the RM-1 cells. After the tumor had grown to about 20 mm<sup>3</sup> in diameter, the infected mice were given lycorine (10 mg kg<sup>-1</sup>) and a vehicle. It shows its anticancer potential by inhibiting the p65 and IKK- $\beta$  phosphorylation, downregulating the Ki-67 expression and increasing caspase 3 in tumor tissue.<sup>67</sup> Lycorine anticancer activity against liver cancer was tested in a xenograft model of Kunming mice. Then, 5  $\times$  10<sup>6</sup> of H22 cells were injected into



the axillary region of the right fore limb. The infected mice were given lycorine ( $10 \text{ mg kg}^{-1}$ ,  $20 \text{ mg kg}^{-1}$  and  $40 \text{ mg kg}^{-1}$ ) and a vehicle. Lycorine reduced the length of the tumor in a dose-dependent manner.<sup>68</sup> Evodiamine was tested for anticancer activity against tongue cancer in a xenograft model of BALB/c nude male mice. For 35 days, infected mice were given evodiamine ( $10 \text{ mg kg}^{-1}$ ) intraperitoneally. It reduced the tumor length by regulating the NF- $\kappa$ B pathway.<sup>69</sup> Evodiamine was tested for anticancer activity against lung cancer in a xenograft model of BALB/c nude female mice. For 22 days, the infected mice were given evodiamine ( $20 \text{ mg kg}^{-1}$ ) *via* gavage. Evodiamine reduced the tumor length by increasing CD8 + T cells and decreasing the MUC1-C/PD-L1 axis.<sup>70</sup> The anticancer activity of evodiamine against lymphoma was tested in a KM male mouse xenograft model. For 21 days, infected mice were given evodiamine ( $20 \text{ mg kg}^{-1}$ ) *via* gavage three times a day. Evodiamine shortened the tumor length by downregulating Ki-67 expression.<sup>71</sup> Evodiamine was tested for anticancer activity against colorectal carcinoma in a xenograft model of BALB/c nude female mice. For 22 days, infected mice were given evodiamine ( $10 \text{ mg kg}^{-1}$ , i.p.). Evodiamine shortened the tumor's length by suppressing hypoxia-inducible factor 1- $\alpha$ -mediated angiogenesis.<sup>72</sup> The anticancer activity of evodiamine against lung cancer was tested in a xenograft model SCID nude mice. For 14 days, the infected mice were given evodiamine ( $20 \text{ mg kg}^{-1}$ ) *via* gavage. Evodiamine showed its potential by inhibiting heat shock protein.<sup>73</sup>

Glycosides: the secondary metabolites, which produce at least one sugar fraction as well as one non-sugar fraction on hydrolysis, are termed glycosides. These include bufalin, antiaroside, papyriferoside, calotropin, ouabain, hyrcanoside, and many more glycosides. The antiproliferative activity of cardiac glycosides has attracted a lot of attention, because the sugar fraction increased solubility and their stereochemistry affected the binding affinity of the receptor protein.<sup>74</sup> Various isolated glycosides IC<sub>50</sub> values with their cytotoxicity on different cell lines are reported in Table 2, and the structures of the isolated glycosides are shown in Fig. 5.

Papyriferoside (63) and its two derivatives with different substitutions [(R<sub>1</sub> = O, R<sup>2</sup> =  $\alpha$ -L-araf-[1-6]- $\beta$ -D-Glcp) (63) (R<sub>1</sub> = O, R<sup>2</sup> =  $\beta$ -D-Api-[1-2]- $\beta$ -D-Glcp) (64) (R<sub>1</sub> = H<sub>2</sub>, R<sup>2</sup> =  $\beta$ -D-Glcp) (65)] show cytotoxic effects against lung cancer, colorectal cancer, and normal skin cancer by inducing apoptosis, resulting in cell cycle arrest, downregulation of IB phosphorylation and BCL-2, and over expression of cleaved caspase and BAX proteins.<sup>94</sup> Antiaroside (66) and its five derivatives with varied substituents [(R<sup>1</sup> = CHO, R<sup>2</sup> = OH, R<sup>3</sup> =  $\alpha$ -OH, R<sup>4</sup> =  $\beta$ -O- $\alpha$ -L-rhamnose) (67), (R<sup>1</sup> = CHO, R<sup>2</sup> = H, R<sup>3</sup> =  $\beta$ -OH, R<sup>4</sup> =  $\beta$ -O-6-deoxy- $\beta$ -D-glucose) (68), (R<sup>1</sup> = CHO, R<sup>2</sup> = H, R<sup>3</sup> =  $\beta$ -OH, R<sup>4</sup> =  $\beta$ -O-6-deoxy- $\beta$ -D-allose) (69), (R<sup>1</sup> = CHO, R<sup>2</sup> = H, R<sup>3</sup> =  $\beta$ -OH, R<sup>4</sup> =  $\beta$ -O- $\alpha$ -L-rhamnose) (70) and (R<sup>1</sup> = CH<sub>2</sub>OH, R<sup>2</sup> = H, R<sup>3</sup> =  $\beta$ -OH, R<sup>4</sup> =  $\beta$ -O-6-deoxy- $\beta$ -D-glucose) (71)] suppress lung cancer cell proliferation by inhibiting the cell migration and the epithelial-mesenchymal transition (EMT) processes.<sup>95</sup> Calotropin (72) and its derivative 12,16-dihydroxycalotropin (73) induce cell death through an apoptotic process that is caspase-dependent and ideally driven by an extrinsic pathway. These *A. subulata* cardenolide glycosides could be used as anticancer drugs. Acemophyllin A (74) and

Acemophyllin B (75) both promote apoptosis, damage DNA, and/or denature proteins, which trap free radicals and protect cellular macromolecules from oxidative mutilation.<sup>96</sup> Ouabain's (76) administration causes an increase in programmed cell death, intracellular ROS production, and breakage of DNA strands. Ouabain also inhibited STAT3-mediated transcription and downstream target proteins, as well as suppressing STAT3 levels and phosphorylation.<sup>79</sup> Hyrcanoside (77) and its two derivatives [deglucohyrcanoside (78) and cymarin (79)] showed anticancer potential against leukemia, lung adenocarcinoma, colorectal carcinoma, adenocarcinoma, breast carcinoma, and osteosarcoma by inducing cell cycle arrest in the G<sub>2</sub>/M phase.<sup>80</sup> Phyto-compounds digoxin (80), digitoxin (81), digitoxigenin (82), lanatoside (83), oleandrin (84) and neritaloside (85), reported in Table 2 are cardiac glycosides. The cardiac glycoside binding site has been investigated, in what manner the multi-functional groups of sodium pump is blocked. The first extracellular subunit channel is the most critical component of the binding site. The -1 subunit is overexpressed in several cancers including lung cancer, renal carcinoma, glioma, and melanoma.<sup>97</sup> Anti-proliferation, Na<sup>+</sup>/K<sup>+</sup>-ATPase activity targeting, and steroid receptor coactivator inhibitions were the key anticancer molecular mechanisms of bufalin (86).<sup>81</sup> 2-Methyl-1,3,6-trihydroxy-9,10-anthraquinone 3-O-(6'-O-acetyl)- $\alpha$ -rhamnosyl(1  $\rightarrow$  2)- $\beta$ -glucoside (87), 2-methyl-1,3,6-trihydroxy-9,10-anthraquinone (88), alizarin (89), purpurin (90) and lucidin- $\omega$ -methyl ether (91) can cause cell death in CNE cells by arresting CNE cells at the G1 stage.<sup>98</sup> Amygdalin (92) has also been demonstrated to prevent various cancer cells by reducing integrin expression and catenin levels, and inhibiting the Akt-mTOR pathway, which may contribute to cancer cell metastasis suppression.<sup>99</sup> Imperatorin (93) decreases the viability of HeLa cells and laryngeal carcinoma (Hep-2) cells by inducing apoptosis and elevating the activity of apoptosis mediator's caspase-3 and caspase-8 in both cell lines.<sup>84</sup> Esculetin (94) treats HN22 and HSC4 cells resulted in a substantial reduction of cancer cells, as well as the regulation of Sp1 regulatory protein.<sup>85</sup> Fraxini's (95) anti-proliferative effect in Hep3 cells was related to apoptosis and alterations in the mitochondrial structure.<sup>86</sup> Coumarin glycosides grandivittin (96), agasyllin (97) and aegelinol benzoate (98) have anticancer properties and showed their anticancer potential by lowering the mitochondrial depolarization potential, modulating the mitochondrial protein pathway, enhancing Bid, Bad, and Box protein expression, and lowering Bcl-xl and Mcl-1 expression.<sup>100</sup> Chartreusin's (99) anticancer actions are due to DNA binding and inhibition of topoisomerase II.<sup>101</sup> Paradoxoside (100) and its seven derivatives with disparate substituents [(R = GlcA R<sub>1</sub> = OH R<sub>2</sub> = Xyl-(1-4)-Rha-(1-2)-Ara) (101), (R = GlcA R<sub>1</sub> = OH R<sub>2</sub> = Rha-(1-3)-Xyl-(1-2)-Rha-(1-2)-Ara) (102), (R = GlcA R<sub>1</sub> = OH R<sub>2</sub> = Api-(1-3)-Xyl-(1-2)-Rha-(1-2)-Ara) (103), (R = GlcA R<sub>1</sub> = OH R<sub>2</sub> = Api-(1-3)-Xyl-(1-2)-Rha-(1-2)-Ara) (104), (R = MeGlcA R<sub>1</sub> = H R<sub>2</sub> = H) (105), (R = Glc-(1-3)-Glc R<sub>1</sub> = H R<sub>2</sub> = H) (106) and (R = GlcR<sub>1</sub> = H R<sub>2</sub> = H) (107)] showed anticancer activity on human leukemia, lung cancer, stomach cancer and breast cancer by regulating the microphthalmia-associated transcription factor (MITF), TRP-1 and TRP-2 expression.<sup>89</sup> Lycopene<sup>90</sup> (108) showed its potential



Table 2 Various isolated glycosides, their biological sources and reported IC<sub>50</sub> values against different types of cancer cell lines

| Isolated compound | Glycoside (biological source)                       | Cancer cell line along with IC <sub>50</sub> (nM) value | References |
|-------------------|-----------------------------------------------------|---------------------------------------------------------|------------|
| 63                | <i>Betula papyrifera</i> (Betulaceae)               | A-549-50                                                | 75         |
| 64                |                                                     | DLD-1-90                                                |            |
| 65                |                                                     | WS1-60                                                  |            |
| 66                | <i>Antiaris toxicaria</i> (Moraceae)                | KB-150                                                  | 76         |
| 67                |                                                     | 1A9-190                                                 |            |
| 68                |                                                     | CAKI-1-130                                              |            |
| 69                |                                                     | S-KMEL-2-200                                            |            |
| 70                | <i>Asclepias subulata</i> (Apocynaceae)             | KB-250                                                  | 77         |
| 71                |                                                     | S-KMEL-2-320                                            |            |
| 72                |                                                     | A549-180                                                |            |
| 73                |                                                     | LS 180, 147                                             |            |
| 74                | <i>Salix acmophylla</i> (Salicaceae)                | PC-3-90                                                 | 78         |
| 75                |                                                     | MCF7-184                                                |            |
| 76                | <i>Strophanthus gratus</i> (Apocynaceae)            | NCI-H460-210                                            | 79         |
|                   |                                                     | A549-12.66                                              |            |
|                   |                                                     | HCT116-10.44                                            |            |
|                   |                                                     | PANC1-42.36                                             |            |
| 77                | <i>Coronilla varia</i> (Fabaceae)                   | Hela-22.6                                               | 80         |
| 78                |                                                     | HCT116-144                                              |            |
| 79                |                                                     | MCF-7-165                                               |            |
| 80                | <i>Digitalis purpurea</i> (Plantaginaceae)          | U-2 OS-44                                               | 74         |
| 81                |                                                     | Hela-25.44                                              |            |
| 82                |                                                     | GSC-22                                                  |            |
|                   | <i>Digitalis purpurea</i> (Plantaginaceae)          | U2OS-18                                                 |            |
|                   |                                                     | SaOS2-15                                                |            |
|                   |                                                     | Huh7-22                                                 |            |
| 83                | <i>Digitalis purpurea</i> (Plantaginaceae)          | Mahlavu-19                                              |            |
| 84                |                                                     | DAOY-50                                                 |            |
| 85                |                                                     | U2OS-95                                                 |            |
|                   | <i>Bufo melanostictus</i> Schneider (Bufonidae)     | SaOS2-90                                                | 81         |
| 86                |                                                     | MDA-MB231-20                                            |            |
|                   |                                                     | Hela-16.6                                               |            |
|                   |                                                     | SW620 15.6                                              |            |
| 87                | <i>Rubia philippinensis</i> (Rubiaceae)             | A549-15.57                                              | 82         |
| 88                |                                                     | MCF7-240                                                |            |
| 89                |                                                     | SK-MEL5-175                                             |            |
| 90                | <i>Rubia philippinensis</i> (Rubiaceae)             | SK-MEL5-235                                             |            |
| 91                |                                                     | B16 F10-80                                              |            |
| 92                |                                                     | MCF-7-178                                               |            |
|                   | Amygdalin, <i>Amygdalus communis</i> (Rosaceae)     | TCCSUP-22.8                                             | 83         |
|                   |                                                     | HeLa-16.8                                               |            |
|                   |                                                     | SNU-C4-34.8                                             |            |
|                   |                                                     | HepG2-39.34                                             |            |
| 93                | <i>Angelica archangelica</i> (Apiaceae)             | SPC-A1-80                                               | 84         |
|                   |                                                     | SGC-7901-160                                            |            |
|                   |                                                     | HeLa-52.86                                              |            |
|                   |                                                     | K562-183                                                |            |
| 94                | <i>Artemisia capillaris</i> (Asteraceae)            | HN22-50.34                                              | 85         |
|                   |                                                     | HSC4-20.24                                              |            |
|                   |                                                     | Hep3B-19.34                                             |            |
| 95                | <i>Fraxinus rhynchophylla</i> (Oleaceae)            |                                                         | 86         |
| 96                | <i>Ferulago campestris</i> (Apiaceae)               | A549-29.34                                              | 87         |
| 97                | <i>Ferulago campestris</i> (Apiaceae)               | A549-180.4                                              |            |
| 98                | <i>Streptomyces chartreusis</i> (Streptomycetaceae) | A549-205.4                                              | 88         |
| 99                |                                                     | L1210-20                                                |            |
|                   |                                                     | P388-70                                                 |            |
|                   |                                                     | B16-90.34                                               |            |
| 100               | <i>Vitellaria paradoxa</i> (Sapotaceae)             | HL60-30                                                 | 89         |
| 101               |                                                     | A549-170                                                |            |
| 103               |                                                     | AZ521-78                                                |            |
| 104               |                                                     | SKBR-3120                                               |            |
| 105               |                                                     | AZ521-108                                               |            |
| 106               |                                                     | HL60-90                                                 |            |
| 107               |                                                     | A549-270                                                |            |



Table 2 (Contd.)

| Isolated compound | Glycoside (biological source)            | Cancer cell line along with IC <sub>50</sub> (nM) value | References |
|-------------------|------------------------------------------|---------------------------------------------------------|------------|
| 108               | <i>Solanum lycopersicum</i> (Solanaceae) | HT-29-70.89                                             | 90         |
| 109               | <i>Malus pumila</i> (Rosaceae)           | HeLa-70.12<br>AGS-40<br>A549-50<br>HepG2-13.16          | 91 and 92  |
| 110               | <i>Brassica napus</i> (Brassicaceae)     | PC3-100.9<br>HCT116-360<br>NCIH929-100.73               | 93         |

against colon cancer by significantly elevated cleaved caspase 3, BAX, cleaved PARP, and 8-oxo-dG levels in cancer cells. Quercetin<sup>91,92</sup> (**109**) diminished the viability of cervical cancer cells through the induction of G<sub>2</sub>/M phase cell cycle arrest and apoptosis, alongside the suppression of cell migration and invasion. In the context of gastric cancer, quercetin was observed to inhibit miR-143, while in HepG2 cells, p53 and miR-34a were found to be inhibited. Apigenin [**110**] showed its anticancer potential by inhibiting the STAT1/COX-2/iNOS signaling pathway.<sup>93</sup>

### 3.2 In vivo anti-cancer studies of glycosides

Recent pre-clinical studies of various glycosides from the natural sources with anti-cancer potential are summarised below:

Lycopene anticancer activity against ovarian cancer was evaluated in egg-laying hens. First, 200 mg kg<sup>-1</sup> and 400 mg kg<sup>-1</sup> of lycopene were given to the hens daily for 12 months. At the end of 12 months, hens were sacrificed and ovarian tissues and blood were collected and evaluated. By reducing the expression of NF-κB and STAT3 and increasing the expression of heme oxygenase 1, lycopene shows its anticancer potential.<sup>102</sup> Lycopene consumption significantly reduced the metastatic load in an ovarian carcinoma-bearing rat model. Its consumption reduces the expression of CA125. The anti-proliferative and anti-metastatic effects were augmented by the down regulation of ITGB1, MMP9, ITGA5, FAK, ILK, and EMT markers, which reduced the MAPK activity and inhibited integrin 5 protein expression. Lycopene activity against tobacco-induced carcinogens was evaluated in male ferrets. For one month, six groups of ferrets were given 50 mg kg<sup>-1</sup> of NNK to induce lung and liver lesions. Following the induction of the lesions, each group was given dietary lycopene for 26 weeks at doses of 2.2 and 6.6 mg kg<sup>-1</sup> BW per day, respectively. Lycopene supplementation inhibited NNK-induced pulmonary α7 nAChR and hepatic CYP2E1, which were linked to lower mortality and occurrences of both pulmonary and hepatic lesions.<sup>103</sup> The anticancer potential of quercetin against colon cancer was tested in 4 week-old Balb/C mice. The control group received no treatment, whereas the treatment group received 10 mg kg<sup>-1</sup> of quercetin per day. The tumor volume was significantly reduced in the treatment group. According to the findings, quercetin has anticancer properties by inhibiting the expression of Notch-1,

Jagged 1, Hes-1, and Presenilin-1.<sup>104</sup> Then 2 × 10<sup>5</sup> MCF-7 cells were inserted into mice. Two groups were divided simultaneously; one group was the untreated group which receive only vehicle while the second group received quercetin (50 mg kg<sup>-1</sup> i.p.) twice a day for a month. Quercetin inhibits tumor by downregulation of VEGF, PKM2, beclin-1, and p-Akt/Akt.<sup>105</sup> Apigenin's anticancer activity against chondrosarcoma was investigated in athymic nude mouse xenografts. Then, 2 × 10<sup>5</sup> Sw1353 cells were inserted into mouse. The untreated group received no treatment, while the treatment group received 5 mg kg<sup>-1</sup> apigenin daily. In the treatment group, the tumor volume was significantly reduced. Apigenin has anti-cancer properties because it inhibits Ki67 expression. Apigenin-induced cell cycle arrest and apoptosis by regulating the expression of Bcl-2.<sup>106</sup> Additionally, apigenin (3 mg kg<sup>-1</sup>) inhibited NSCLC xenograft growth and metastasis by targeting the dipeptidyl peptidase IV (DPPIV) enzyme.<sup>107</sup> Digoxin anticancer activity against human lung cancer was investigated in BALB/c nude mouse xenograft model. Following this, 1 × 10<sup>7</sup> A549 cells were implanted in mouse. After the tumor had grown to about 100 mm<sup>3</sup> in diameter, the infected mice were daily given digoxin (1.0 mg kg<sup>-1</sup>). After 14 days, mice were forfeited, and the tumor volume was collected and evaluated. The results indicated that digoxin inhibits lung cancer by inhibiting both DNA DSB and SSB repairs.<sup>108</sup> Digitoxin anticancer activity against cervical cancer was investigated in a BALB/c nude mouse xenograft model. Then, 5 × 10<sup>6</sup> HeLa cells were implanted into mouse. After the tumor had grown to about 300 mm<sup>3</sup> in diameter, the infected mice were given digitoxin (1.0 to 2.0 mg kg<sup>-1</sup>) daily. After 19 days, mice were forfeited, and the tumor volume was collected and evaluated. Digitoxin shows its potential by arresting the cell.<sup>109</sup> Bufalin anticancer activity against human lung cancer was investigated in a BALB/c nude mouse xenograft model. Then, 8 × 10<sup>6</sup> A549 cells were implanted into mouse. After the tumor had grown to about 300 mm<sup>3</sup> in diameter, the infected mice were given bufalin (1 mg to 6 mg kg<sup>-1</sup>) daily. After 19 days, mice were forfeited, and the tumor volume was collected and evaluated. Bufalin shows its potential by activation of caspase-3 and the cleavage of PARP in A549 cells.<sup>110</sup> Bufalin anticancer activity against breast cancer was investigated in athymic nude mice. Then, 5 × 10<sup>6</sup> MB-231 cells were injected subcutaneously into both dorsal regions of mice and 10 μl of bufalin was injected once a day at an early stage of cancer. Mice were



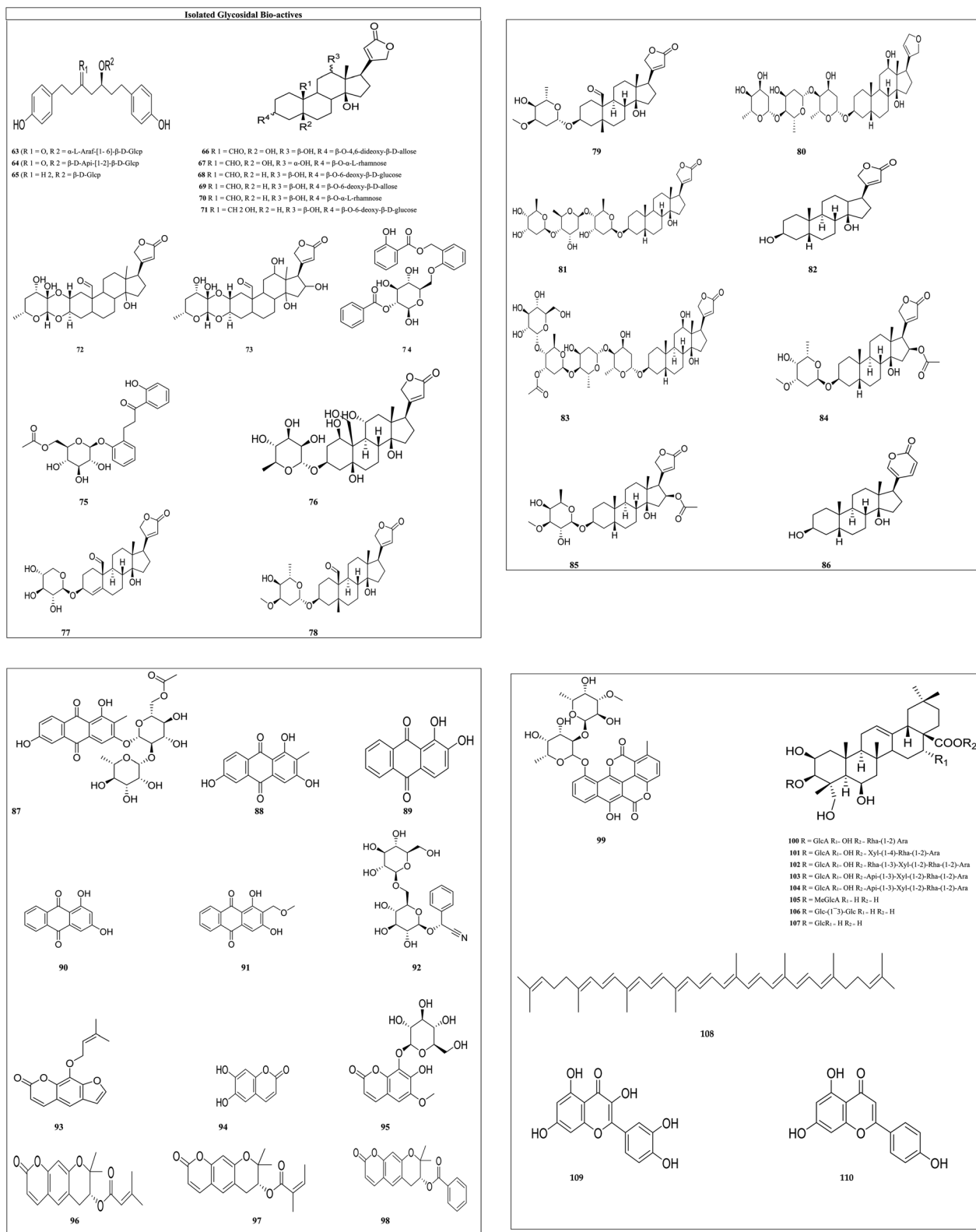


Fig. 5 Structures of isolated glycosides.

forfeited after 21 days. The tumor was collected and evaluated. The results indicated that bufalin reduced breast cancer angiogenesis by inhibiting the MAPK and NF- $\kappa$ B pathways.<sup>111</sup> Alizarin anticancer activity against pancreatic cancer was

investigated in the mouse xenograft model. Then  $5 \times 10^6$  MIA PaCa-2-luc cells were implanted into the mouse. After the tumor had grown to about 300 mm<sup>3</sup> in diameter, the infected mice were given alizarin (10 to 30 mg kg<sup>-1</sup>) daily. After 19 days, mice

were sacrificed, and the tumor volume was collected and evaluated. Digitoxin showed its potential by abrogating NF- $\kappa$ B activation.<sup>112</sup>

**Miscellaneous bio-actives.** Various plant bio-actives other than glycosides and alkaloids also possess the anticancer potential and showed significant activity against various types of cancers. These secondary compounds may contain terpenoids, terpenes, flavonoids, and lignans. Table 3 presents the various secondary metabolites exerting cytotoxicity on different cell lines, and the structures of miscellaneous bioactives are shown in Fig. 6.

In melanoma cell lines, 4-nerolidylcatechol (**111**) is reported as an inhibitor of cell invasiveness, owing to the G1 cell cycle arrest and inhibition of MMP-2 activity.<sup>113</sup> Melanoma has a high prevalence of B-RAF mutations. Cucurbitacin B (**112**) could be a possibility for inhibiting the signaling kinase pathway. Cucurbitacin B is a kinase inhibitor for B-RAF and MEK1.<sup>114</sup> Betulin's (**113**) anticancer action is based on the stimulation of apoptotic cell death. Betulin treatment caused cytomorphological changes that are typical of apoptotic cells, including cell rounding and the production of apoptotic bodies.<sup>115</sup> Camphor (**114**) white oil caused transcriptional alterations in immune-related genes identified by RNA-sequencing *in vivo*, leading to tumor regression mediated by cytotoxic T cells.<sup>116</sup> The cytotoxicity of myrcene (**115**) against leukemia cells was shown to be substantial. At 0.01 g ml<sup>-1</sup>, myrcene decreased *t*-butyl hydroperoxide-induced DNA damage in human B lymphoid NC-NC cells by 50%.<sup>146</sup> Linalool (**116**) inhibited mitochondrial complexes I and II, increased reactive oxygen species, and lowered ATP and GSH levels in HepG2 cells. Linalool also upregulated p53 and cyclin-dependent kinase inhibitors, which induced strong apoptosis in a variety of leukemia cells.<sup>147</sup> By decreasing the mRNA and protein expression of human telomerase reverse transcriptase, costunolide (**117**) inhibited proliferation in human B cell leukemia cells.<sup>148</sup> Acacetin (**118**) inhibited epidermal growth factor (EGF)-induced cell transformation and phosphorylation of p70S6K. Acacetin binds to the p110 subunit of PI3-K, interacting with Val828, Glu826 and Tyr813 residues.<sup>119</sup> Mitf, a transcription factor related to microphthalmia and found downstream of the Wnt/catenin pathway, has emerged as a key melanoma prognostic factor. Fisetin (**119**) (3,7,3',4'-tetrahydroxyflavone) treatment of melanoma cells resulted in decreased cell survival, G<sub>1</sub>-phase arrest, and inhibition of Wnt/catenin signaling. Fisetin-treated cells have higher intracellular levels of Axin and TrCP, as well as reduced glycogen synthase kinase 3 phosphorylation and catenin stabilization.<sup>120</sup> Silymarin (**120**) blocks cyclin-dependent kinase (CDK) activity and increases the levels of the CDK inhibitors p21CIP1 and p27KIP1 such that they are more tightly bound to CDKs, which suppresses EGFR signaling. Silymarin inhibits development at the G1 and G2 checkpoints.<sup>121</sup> The bioactive flavonoid, astragalin (**121**) heptaacetate (AHA) promotes apoptosis in HL-60 cells by releasing cytochrome c into the cytoplasm. Activation of Bax, caspase-3/-7, and p38MAPK, as well as intracellular ROS production and suppression of cell signaling pathways JNK/SAPK and ERK 1/2 also promote apoptosis in HL-60. TNF-induced NF-B

activation is significantly inhibited by astragalin in A549 and H1299 cells. Furthermore, astragalin-induced cell death is associated with a time- and dose-dependent increase in the Bax/Bcl-2 ratio, as well as increased cleavage of caspase-3/-9 and PARP.<sup>123</sup> Ginkgetin (**122**) decreased cell viability in breast cancer and blocked estrogen receptor (ER) expression at mRNA and protein levels. Ginkgetin therapy also reduced the expression of survivin, and cyclin D1, which are also ER targets.<sup>124</sup> Epigallocatechin gallate (**123**) inhibited cell proliferation by reducing the PCNA protein level and promoted apoptosis in melanoma by assessed cleavage of PARP, TUNEL assay. Treatment of melanoma cells with epigallocatechin gallate leads to a reduction in cyclin D1 and cdk2 protein levels, as well as stimulation of the cyclin kinase inhibitors (ckis) and p27KIP1.<sup>125</sup> Combretastatin (**124**) is the new molecule of vascular disrupting medicines that target tumor blood channels and prevent angiogenesis. Combretastatin affects DNA structure and function by interfering with nucleic acid production and transcription and inhibiting cell proliferation.<sup>149</sup> Resveratrol (stilbenoid) [**125**] was observed to halt the cell cycle at the G<sub>2</sub>/M phase, also elevating intracellular reactive oxygen species (ROS) and caspase 3 activity, and increasing the Bax/Bcl-2 protein ratio, all of which are indicative of apoptosis in hepatic cancer.<sup>126</sup> Elenoside (**126**) was screened for its anticancer potential on skin cancer cell lines but its mechanism of action is not known.<sup>150</sup> Sulfuraphane<sup>128</sup> (**127**) induced mitochondrion-mediated apoptosis in cancer cells through the activation of caspase-9, followed by the cleavage and subsequent activation of caspase-3 and caspase-7. 6-Shogaol<sup>129</sup> (**128**) has demonstrated the capability to inhibit the proliferation and migration of cervical cells through the suppression of the PI3K/Akt/mTOR signaling pathway. Allicin<sup>130</sup> (**129**) has been shown to exert its cytotoxic effects by targeting cancer cells during the S and G<sub>2</sub>/M phases of the cell cycle. Alpinumisoflavone<sup>131</sup> (**130**) modulates several signaling pathways, including PI3K/Akt, MAPK, and those regulating endoplasmic reticulum (ER) stress, ultimately leading to cell death and showcasing its therapeutic potential. Andrographolide<sup>132</sup> (**131**) acts against leukemia by inducing cell cycle arrest in the G<sub>0</sub>/G<sub>1</sub> phase, while also affecting the G<sub>2</sub>/M, G<sub>1</sub>, and S phases in hepatoma and colon cancer. The anti-tumor effects of baicalin<sup>133</sup> (**132**) in breast cancer may be attributed to a novel mechanism involving tumor-associated macrophages. Decursin<sup>134</sup> (**133**) reveals its potential by disrupting multiple signaling pathways; for instance, in gastric cancer, it alters the STAT3/c-Myc pathway and the MAPK/ERK1/2 pathways associated with colon and melanoma cancers. Additionally, decursin affects the PERK/ATF4 pathway, which plays a role in lung cancer. Decursinol<sup>135</sup> (**134**) exerts its cytotoxic properties through the regulation of the G<sub>0</sub>/G<sub>1</sub> phase in prostate cancer cells. The anticancer properties of dicumarol<sup>136</sup> (**135**) have been linked to the inhibition of NQO1. Genistein<sup>137</sup> (**136**) directly inhibits the PLK1 signaling pathway, demonstrating its anticancer efficacy. Gingerol<sup>138</sup> (**137**) has the capacity to induce the generation of reactive oxygen species (ROS) in chronic (K562) and acute myeloid leukemia (U937) tumor cell lines, resulting in the disruption of the G<sub>2</sub>/M cell cycle, a reduction in cell cycle protein expression (including cyclin B1, Cdk1, Cdc25B, and



Table 3 Different bio-actives, their biological sources and IC<sub>50</sub> values in different cancer cell lines

| Isolated compounds    | Biological source                             | Cancer cell line along with IC <sub>50</sub> (nM) value | References  |
|-----------------------|-----------------------------------------------|---------------------------------------------------------|-------------|
| 111                   | <i>Pothomorphe umbellata</i> (Piperaceae)     | SK-MEL2-95<br>SK-MEL103-100<br>SK-MEL147-90             | 113         |
| 112                   | <i>Cucumis sativus</i> (Cucurbitaceae)        | SK MEL28-45<br>A-375-30                                 | 114         |
| 113                   | <i>Betula pendula</i> (Betulaceae)            | SK-MEL 28-200<br>MSK-MEL2-198.4<br>G361-190             | 115         |
| 114                   | <i>Cinnamomum camphora</i> (Lauraceae)        | A549-18.7                                               | 116         |
| 115                   | <i>Cannabis sativa</i> (Cannabaceae)          | A549-120                                                | 117         |
| 116                   | <i>Cannabis sativa</i> (Cannabaceae)          | C6-22                                                   |             |
| 117                   | <i>Colchicum autumnale</i> (Colchicaceae)     | HA22T/VGH-90<br>OVCAR3-129<br>T24-180<br>MDA-MB-231-30  | 118         |
| 118                   | <i>Robinia pseudoacacia</i> (Fabaceae)        | LOVO-100                                                | 119         |
| 119                   | <i>Malus domestica</i> (Rosaceae)             | Mel 928-25                                              | 120         |
| 120                   | <i>Silybum marianum</i> (Asteraceae)          | DU145-24<br>MDAMB-468-46<br>MMP2-63.5<br>CD34-56.4      | 121 and 122 |
| 121                   | <i>Anaphalis neelgerriana</i> (Asteraceae)    | HaCaT-28<br>HL-60-40<br>A549-68<br>H1299-64             | 123         |
| 122                   | <i>Ginkgo biloba</i> (Ginkgoaceae)            | MCF7-48, T47D-52                                        | 124         |
| <b>Polyphenols</b>    |                                               |                                                         |             |
| 123                   | <i>Camellia sinensis</i> (Theaceae)           | A375 12.8<br>Hs294T-8.78                                | 125         |
| 124                   | <i>Combretum caffrum</i> (Combretaceae)       | P388-28                                                 | 36          |
| 125                   | <i>Cajanus cajan</i> (Fabaceae)               | HepG2-50.99<br>MCF-7-20.56<br>A549-60.18                | 126         |
| <b>Lignan</b>         |                                               |                                                         |             |
| 126                   | <i>Justicia hyssopifolia</i> L. (Acanthaceae) | MALME-3M-16<br>SK-MEL-5.32<br>UACC257-48                | 127         |
| <b>Isothiocyanate</b> |                                               |                                                         |             |
| 127                   | Sulforaphene                                  | MCF-7-41.1<br>HepG2-40.0<br>HT-29-42                    | 128         |
| 128                   | <i>Zingiber officinale</i> (Roscoe)           | HeLa-250.68<br>SiHa-370.52                              | 129         |
| 129                   | <i>Allium sativum</i> (Amaryllidaceae)        | HepG2-19.26<br>MCF7-28.51<br>A549-36<br>PC3-77.92       | 130         |
| 130                   | <i>Derris eriocarpa</i> (Leguminosae)         | KB-40.13<br>P-388-34.31<br>H2108-56.5                   | 131         |
| 131                   | <i>Andrographis paniculata</i> (Acanthaceae)  | HL 60-20.4<br>HepG2-40.2<br>Lovo-8.6                    | 132         |
| 132                   | <i>Scutellaria baicalensis</i> (Lamiaceae)    | MDA-MB-231-34.77<br>MCF7-41.78                          | 133         |
| 133                   | <i>Angelica gigas</i> (Apiaceae)              | SNU-216-50<br>HT29-293.064<br>A549-200<br>B16F10-80     | 134         |
| 134                   | <i>Angelica gigas</i> (Apiaceae)              | PC3-36.3                                                | 135         |

Table 3 (Contd.)

| Isolated compounds | Biological source                          | Cancer cell line along with IC <sub>50</sub> (nM) value | References |
|--------------------|--------------------------------------------|---------------------------------------------------------|------------|
| 135                | <i>Melilotus officinalis</i> (Fabaceae)    | MCF7-40                                                 | 136        |
| 136                | <i>Glycine max</i> (legumes)               | MCF7-15<br>HepG2-25<br>NCI-H1299-55                     | 137        |
| 137                | <i>Zingiber officinale</i> (Roscoe)        | S-180-19.18<br>HL-60-111.4                              | 138        |
| 138                | <i>Glycyrrhiza glabra</i> (Fabaceae)       | MDA-MB-231-84.22                                        | 139        |
| 139                | <i>Salvia involucrata</i> (Lamiaceae)      | MCF-7-25.44<br>HCC38-65.42                              | 140        |
| 140                | <i>Glycyrrhiza glabra</i> (Fabaceae)       | MCF-7-17.63<br>A549-11.55<br>DU-145-9.45                | 141        |
| 141                | <i>Azadirachta indica</i> (Meliaceae)      | EJ-30<br>MDA-MB-231-10.97<br>HT29-40<br>HCT116-75       | 142        |
| 142                | <i>Physalis pubescens</i> L. (Solanaceae)  | SKOV3-60.63                                             | 143        |
| 143                | <i>Polygonum cuspidatum</i> (Polygonaceae) | HeLa-30                                                 | 144        |
| 144                | Withaferin A                               | Panc1-10.24<br>BxPc-320.78<br>U87MG-10.4<br>GBM2-19     | 145        |

Cdc25C), and alterations in cellular oxidant status that promote mitochondrial ROS production. Glycyrrhizin<sup>139</sup> (138) offers protective and detoxifying effects by reducing the generation of reactive oxygen species, preserving glutathione (GSH), and differentially modulating apoptosis, as well as the Akt, ERK, and JNK pathways within the MAPK signaling cascade. Hispidulin<sup>140</sup> (139) has been shown to inhibit TGF- $\beta$ 1-induced Smad2/3 signaling and cell migration across breast cancer. Licochalcone A<sup>141</sup> (140) modulates the expression of various signaling pathways, including the EGFR/ERK, PI3K/Akt/mTOR, p38/JNK, MKK4/JNK, mitochondrial apoptosis pathway and the death receptor pathway. It inhibits the expression of proteins involved in the cell cycle and angiogenesis, and regulates both autophagy and apoptosis in cancer cells. Nimbolide<sup>142</sup> (141) blocks the attainment of cancer hallmarks such as sustained proliferation, evasion of apoptosis, invasion, angiogenesis, metastasis, and inflammation by influencing kinase-driven oncogenic signaling pathways and shows its potential. Furthermore, physapubescin B<sup>143</sup> (142) inhibited the transcriptional activity of STAT3, an oncogenic transcription factor implicated in numerous human malignancies, including ovarian cancer. Pterostilbene<sup>144</sup> (143) was linked to the induction of apoptosis in tumor cells, as well as the downregulation of the oncogene E6 and the upregulation of activated caspase-3 levels. Withaferin A<sup>145</sup> (144) was found to induce apoptosis and inhibit growth in pancreatic cancer cells through mitochondrial dysfunction and inactivation *via* the PI3K/Akt pathway.

### 3.3 *In vivo* anti-cancer studies of miscellaneous bio-actives

The pre-clinical data of various miscellaneous bio-actives including different terpenoids, flavones, polyphenols, *etc.*, are included as follows:

Epigallocatechin anticancer activity against lung cancer was evaluated in A/J female mice. Mice were injected with cisplatin for the induction of cancer. Epigallocatechin (1 mg ml<sup>-1</sup>, orally) was given to the mice. Male db/db mice were given tap water containing 40 ppm DEN for two weeks, followed by 34 weeks of drinking water containing 0.1% epigallocatechin gallate. The fortified drinking water containing epigallocatechin gallate significantly reduced the development of liver cell adenomas compared to the EGCG-untreated control group. In the livers of experimental mice, epigallocatechin gallate inhibited the phosphorylation of the ERK (extracellular signal-regulated kinase), Akt, Stat3, and JNK proteins. Chitosan-based nano-formulation of epigallocatechin gallate (10 mg ml<sup>-1</sup>) was also developed for the treatment of prostate cancer and the same was evaluated in a xenograft athymic nude mouse model. The formulation decreased the expression of Ki-67 and VEGF (markers of angiogenesis) in tissues of treated mice.<sup>151</sup> Emodin's anticancer activity against human lung epithelial cancer was investigated in BALB/c nude mice with 50 mg per kg emodin daily, which inhibits cell growth (A549) by inducing ER-dependent apoptosis. In hepatocellular cancer, emodin shows its potential by inhibiting the p-JNK expression and increasing ERK and p38 phosphorylation. Emodin's anticancer activity against breast cancer was investigated in C57BL/6 and BALB/c mice. Then, 2 × 10<sup>5</sup> EO771 or 4T1 cells were inserted into mice and 40 mg per kg emodin was injected once a day at an early stage of cancer. Mice were sacrificed at different time intervals and the tumor was collected and evaluated. The results indicated that emodin reduced breast cancer angiogenesis by inhibiting M2 polarization and macrophage infiltration and increasing T-cell activation.<sup>152</sup> Baicalein's anticancer activity against colon cancer was investigated in mouse xenograft. Then



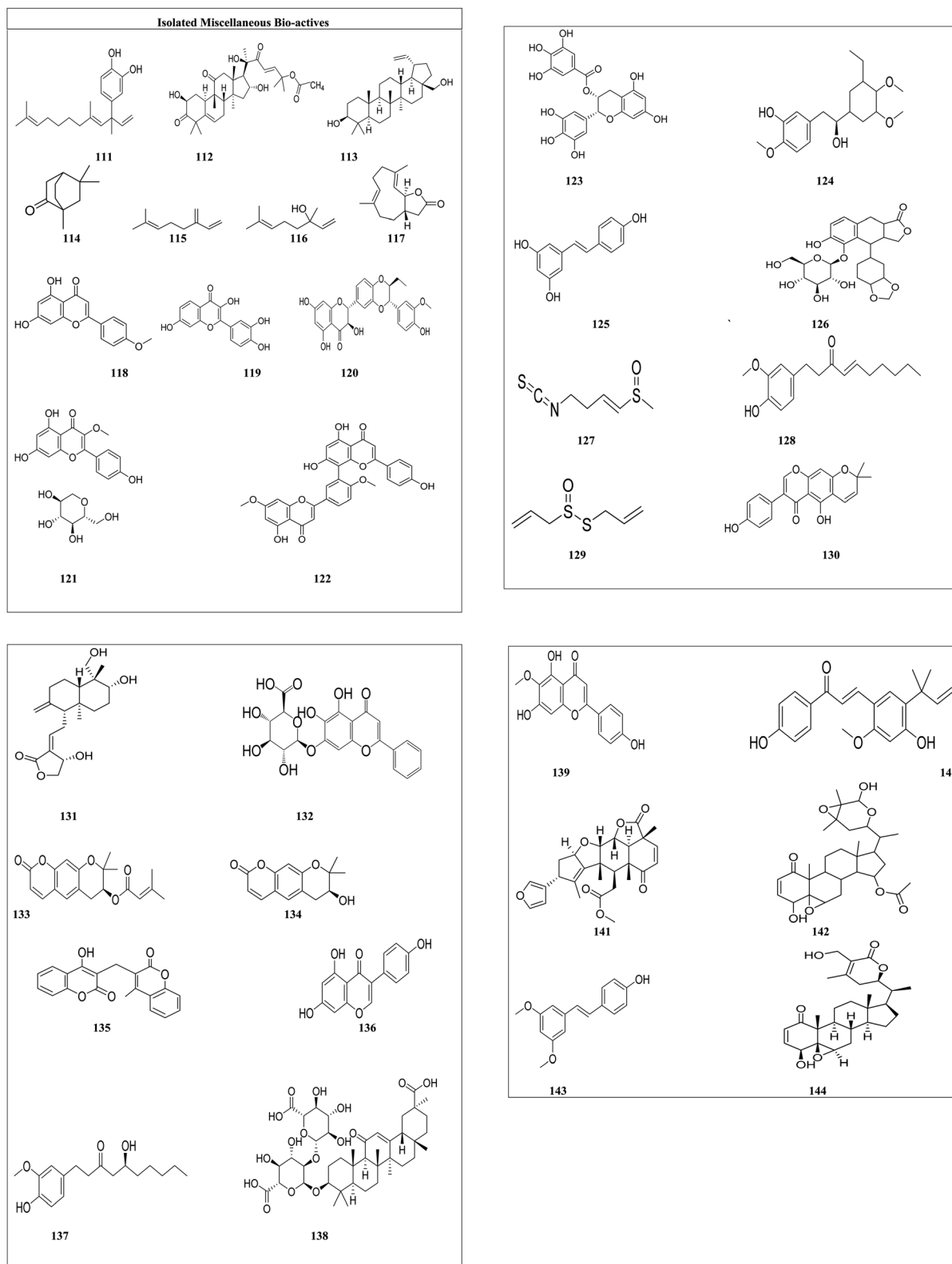


Fig. 6 Structures of isolated miscellaneous drugs.

50 mg kg<sup>-1</sup> of baicalein was given to the infected (HCT116 cell) mouse. Baicalein shows its activity by downregulating the mitogen-activated protein kinase (MAPK) and p38 signaling

pathways.<sup>153</sup> A nude mouse model was used to test withaferin A's anticancer activity against colorectal cancer. CRC cells were inserted into the mouse. The mouse was given 5 mg per kg

withaferin A orally after the onset of cancer. According to the findings, withaferin A has the potential to inhibit Akt over-expression and micro-vessel formation. Withaferin A's anti-cancer activity against hepatocellular carcinoma was investigated in athymic nude mouse xenografts. Then,  $5 \times 10^6$  HepG2 cells were injected subcutaneously into the mice. After 15 days of implantation, mice were divided into the untreated and treatment groups. The untreated group received no treatment, while the treatment group received 4 mg per kg withaferin A orally daily. After 5 weeks of treatment, mice were sacrificed and the tumor was collected and evaluated. Withaferin A showed its anticancer potential by inhibiting Ki67 expression while increasing the ERK, RSK, ELK1, and DR5 levels.<sup>154</sup> Some recent pre-clinical data related to the anti-cancer potential of plant bio-actives are also listed in Table 4.

### 3.4 Clinical trial data for plant-derived bioactives in cancer management

Despite the fact that an enormous number of anti-cancer molecules are currently being developed, clinical trials using phyto-chemicals to manage different cancers are still in the early stages.<sup>172</sup> The trials on anticancer moieties are focused on three important components: first, increasing cancer cells response to standard chemo- and radio-therapy; second, minimising the severe side effects of traditional cancer therapy; and third, identifying undesirable interactions with standard therapy. Preclinical studies of various phytoconstituents have revealed a high potential for treating various types of cancers. Due to a lack of research and knowledge regarding their mechanism of action, the specific site of action, and dose, they

failed to enter clinical trials. Currently, only seven phytoconstituents are under clinical trial as reported in Table 5.<sup>173–175</sup>

### 3.5 Structure–activity relationship (SAR) analysis of compounds with similar structures

Berberine and its derivatives have efficient cytotoxic potential against breast cancer, liver cancer and pancreatic cancer, and its structural analysis shows that its antitumor activity is mainly concentrated on C-9 and C-13, the derivatives being more potent than the parent compound. Propyl benzene and 4,4-diphenylbutyl on C-9 showed better anti-breast cancer cell toxicity, while 1-chloro-4-ethylbenzene and phenylpentyl on C-13 are strong electron-withdrawing groups and strengthen cytotoxicity potential against hepatic cancer cell toxicity compared to the parent compound.<sup>18</sup>

**Evodiamine and its derivatives.** Substitution at different positions of evodiamine such as fluorine at the C3 position, chlorine at both the C3 and C10 positions strengthen the cytotoxicity potential. The free hydroxyl group at the 10 position is also important for high antiproliferative activity. Substitutions at C3 and C10 have also been found to have synergistic effects. For example, 3-fluoro-10-hydroxyevodiamine and 3-amino-10-hydroxyevodiamine not only exhibit excellent antitumor activity but also have good water solubility. In addition, modifications of the D-shaped ring framework are also tolerated. Substitution of the C5 carbonyl group with a thio-carbonyl group or substitution of the N14 methyl group with an oxygen atom has a positive effect on antitumor activity.<sup>20</sup>

SAR analysis shows that analogs of pancratistatin depend largely on the hydroxyl group at C-7 and the functional group

**Table 4** *In vivo* studies of some secondary metabolites from natural sources in different types of cancers

| Isolated compound | Plant                                        | Type of cancer             | Model/dose                                                                       | Reference |
|-------------------|----------------------------------------------|----------------------------|----------------------------------------------------------------------------------|-----------|
| 6-Shogaol         | <i>Zingiber officinale</i> (Roscoe)          | Non-small cell lung cancer | Nude mice model (10 mg kg <sup>-1</sup> )                                        | 155       |
| Allicin           | <i>Allium sativum</i> (Amaryllidaceae)       | Liver bile duct carcinoma  | BALB/c nude mice model (10 mg kg <sup>-1</sup> )                                 | 156       |
| Alpinum           | <i>Derris eriocarpa</i> (Leguminosae)        | Renal cell carcinoma       | In BALB/c nude mice xenograft (40 mg kg <sup>-1</sup> )                          | 157       |
| Andrographolide   | <i>Andrographis paniculata</i> (Acanthaceae) | Breast cancer              | Nude (BALB/c females, 6–8 weeks old) mice (25, 50, and 100 mg kg <sup>-1</sup> ) | 158       |
| Baicalin          | <i>Scutellaria baicalensis</i> (Lamiaceae)   | Colon cancer               | Nude mice (50 mg kg <sup>-1</sup> )                                              | 159       |
| Curcumin          | <i>Curcuma longa</i> (Zingiberaceae)         | Melanoma cancer            | Six-week-old female BALB/c nude mice (25 mg kg <sup>-1</sup> )                   | 160       |
| Decursin          | <i>Angelica gigas</i> (Apiaceae)             | Prostate cancer            | SCID-NSG mice xenograft (4.5 mg kg <sup>-1</sup> )                               | 161       |
| Dicumarol         | <i>Melilotus officinalis</i> (Fabaceae)      | Ovarian carcinoma          | BALB/c nude mouse xenograft model, DIC (30 mg kg <sup>-1</sup> )                 | 162       |
| Genistein         | <i>Glycine max</i> (legumes)                 | Leukemia                   | Male athymic BALB/c nu/nu mice 6–8 week (0.2 or 0.4 mg kg <sup>-1</sup> )        | 163       |
| Gingerol          | <i>Zingiber officinale</i> (Roscoe)          | Breast cancer              | Mice model (5 mg kg <sup>-1</sup> )                                              | 164       |
| Glycyrrhizin      | <i>Glycyrrhiza glabra</i> (Fabaceae)         | Non-small cell lung cancer | Athymic BALB/c nude mice xenograft (100 mg kg <sup>-1</sup> )                    | 165       |
| Hispidulin        | <i>Salvia involucrata</i> (Lamiaceae)        | Hepatocellular carcinoma   | Nude mice (20 mg kg <sup>-1</sup> )                                              | 166       |
| Stilbenoid        | <i>Polygonum cuspidatum</i> (Polygonaceae)   | Breast cancer              | Nude mouse mode (5 mg kg <sup>-1</sup> )                                         | 167       |
| Licochalcone A    | <i>Glycyrrhiza glabra</i> (Fabaceae)         | Glioma cell                | Athymic nude mice (10 mg kg <sup>-1</sup> )                                      | 168       |
| Nimbolide         | <i>Azadirachta indica</i> (Meliaceae)        | Pancreatic cancer          | Athymic nu/nu mouse model, (5 mg kg <sup>-1</sup> )                              | 169       |
| Physapubescin B   | <i>Physalis pubescens</i> L. (Solanaceae)    | Renal cell carcinoma       | Xenograft mouse model (30 mg kg <sup>-1</sup> )                                  | 170       |
| Pterostilbene     | <i>Polygonum cuspidatum</i> (Polygonaceae)   | Endometrial cancer         | Xenograft mouse model (30 mg kg <sup>-1</sup> )                                  | 171       |



substitution at C-1. Three new synthetic analogs, SVTH-5, SVTH-6, and SVTH-7, were examined, which possesses the complete anticancer pharmacophore of pancratistatin, including the hydroxyl group at C-7. As a result, SVTH-6 and SVTH-5 were more effective against cancer cells than related compounds JCTH-4 and JCTH 3, respectively, which lack this functional group. In addition, the functional group at C-1 significantly determines the effectiveness of the analogs. For example, JCTH-1 and JCTH-2 differ from JCTH-4 only in the functional group at C-1 and have hardly any anticancer activity. Similarly, SVTH-7 differs from SVTH-6 and SVTH-5 only in the C-1 group and is more effective against most cancer cell lines tested.<sup>43</sup>

**Antiaroside and its derivatives.** The structure–activity relationships of these compounds showed that the orientation of the C-3 and C-17 substituents plays an important role in the overall cytotoxicity profile. However, compounds with  $\alpha$ -orientation of the C-3 and C-17 substituents showed weaker anticancer activity. Similarly, compounds containing an  $\alpha$ -L-rhamnose residue at C-3 showed potent cytotoxic activity. The position of the glycosyl bond is also very important for cytotoxicity. For example, compounds with the sugar attached to C-19 showed lower cytotoxicity than compounds with this residue attached to C-3.<sup>76</sup>

Cymarín, hyrcanoside and deglucohyrcanoside contain a carbonyl group at C-19 and a  $\beta$ -hydroxyl group at C-5. While ouabain contains a  $\beta$ -hydroxyl group at C-1,  $\alpha$ -OH groups at C-10

and C-19, but lacks a  $\beta$ -OH group at C-5. The  $\beta$ -hydroxyl group at C-5 may contribute to general cytotoxicity. However, as reported in the literature, cytotoxicity is significantly affected by the carbonyl group at C-19; when the hydroxyl group at C-19 is replaced by a carbonyl group, the cytotoxicity of the resulting derivative increases.<sup>80</sup>

**Sophoridine and its derivatives.** From the SAR studies, it can be concluded that the substitution of a phenylmethylene group at the C-14 position of the parent sophoridine resulted in enhanced anticancer activity. The addition of a conjugated structure at the C-15 position of the carbonyl group *via* imine formation resulted in enhanced anticancer activity. The substitution of fatty acyl group at the C-12 position significantly improved the antitumor activity of *N*-substituted sophoridine derivatives. However, the side chain at the C-12 position is larger and not suitable for anticancer activity.<sup>176</sup>

## 4 Discussion

In this work, we reviewed a total of 144 isolated compounds which are having anticancer potential against different cell lines. Out of 144, there are 62 alkaloids, 47 glycosides, and 35 other isolated compounds, which include flavones, terpenoids, terpene, and polyphenols (Fig. 7). Out of 62 alkaloids, only one alkaloid berberine is currently under clinical trials in placebo-controlled phase 2/3 on colorectal cancer.

**Table 5** List of various bio-actives derived from plants under various stages of clinical trials for the management of different types of cancers

| Isolated compound                     | Biological source                          | Type of cancer                                            | Stage of trial                                                                                                                                                         | Identifier code |
|---------------------------------------|--------------------------------------------|-----------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------|
| Berberine hydrochloride (alkaloid)    | <i>Berberis</i> sp. (Berberidaceae)        | Colorectal cancer                                         | Placebo-controlled phase 2/3 trial berberine hydrochloride (1000 patients) (300 mg twice per day)                                                                      | NCT03281096     |
| Curcumin (polyphenol)                 | <i>Curcuma longa</i> (Zingiberaceae)       | Advanced and metastatic breast cancer                     | Placebo-controlled phase 2/3 trial curcumin (300 mg i.v. per day) along with paclitaxel (80 mg per m <sup>2</sup> BS; i.v.) once a week for 12 weeks                   | NCT03072992     |
| Epigallocatechin (flavonoids)         | <i>Camellia sinensis</i> (Theaceae)        | Colorectal cancer                                         | Early phase 1 trial Teavigo™ (highly purified and refined green tea extract providing 94% EGCG) (450 mg PO per day)                                                    | NCT02891538     |
| Lycopene (carotenoids)                | <i>Solanum lycopersicum</i> (Solanaceae)   | Metastatic colorectal cancer                              | Placebo-controlled phase 2 trial lycopene (20 mg PO per day) to reduce skin toxicity                                                                                   | NCT03167268     |
| Quercetin (carotenoids)               | <i>Glycyrrhiza glabra</i> (Leguminosae)    | Prostate cancer                                           | Phase 1 trial, placebo-controlled, two arm study of quercetin and green tea to enhance the bioavailability of green tea polyphenols in men scheduled for prostatectomy | NCT01912820     |
| Resveratrol (stilbenoid) (polyphenol) | <i>Polygonum cuspidatum</i> (Polygonaceae) | Low-grade GI neuroendocrine tumors                        | Placebo-controlled phase 1 trail, resveratrol (2.5 g p.o. twice per day) on Notch-1 signaling in low-grade gastrointestinal neuroendocrine tumors                      | NCT01476592     |
| Sulforaphane (isothiocyanate)         | <i>Brassica oleracea</i> (Brassicaceae)    | Former smokers with a high risk of developing lung cancer | Placebo-controlled phase 2 trial Avmacol (sulforaphane) tablets (120 $\mu$ M p.o. twice per day)                                                                       | NCT03232138     |





Fig. 7 Percentage of different categories of bioactive compounds with anticancer potential.

The  $IC_{50}$  value of berberine is 250 nM for breast cancer cell lines, 400 nM for colorectal cancer, and 200 nM for chondrosarcoma and lung cancer. Berberine's structural alteration for anticancer action had primarily focused on  $C_9$  and  $C_{13}$ . To increase the efficacy and bioavailability, some cycloberberine derivatives were also developed. By enhancing the moderate DNA-binding affinities of protoberberine alkaloids, five derivatives substantially inhibit human HepG2 and human colon cancer cell lines. With an  $IC_{50}$  of 200 nM, evodiamine has anticancer potential in human liver cancer cell lines (HepG2 and PLHC-1). The best derivative was 4-chlorobenzene, which had  $IC_{50}$  values of 8.6, 4.9, and 260 nM against A549 (lung cancer), MDA-MB435 (breast cancer), and HCT116 (colon cancer) cell lines with *N*-substitution series of evodiamine derivatives. Bulbispermine shows cytotoxic effects against glioblastoma (T98G and U373) and human leukemia (HL-60) with  $IC_{50}$  values of 90 nM, 380 nM, and 80 nM. Distichamine shows anticancer activity against HeLa, CEM, K562, MCF-7, and G-361 with  $IC_{50}$  values of 22–147 nM. Lycorine had  $IC_{50}$  ranging from 50 nM to 100 nM for different cell lines such as PC-3M, DU145, LNCaP, and 22RV1 and *in vivo* lycorine (5 mg per kg per day or 10 mg per kg per day) reduces prostate cancer.

We have also reviewed 47 glycosides that have anticancer potential against various cancer cell lines and digoxin was found to have the greatest potential to treat various cancers. Digoxin has undergone 27 clinical trials in which 11 trials are completed, 7 trials are under recruiting, 1 trial is not yet recruiting, 2 trials are active, 2 trials are terminated, 3 studies have unknown status, and 1 study is withdrawn. All trials involved digoxin alone or a combination of digoxin with other drugs such as enzalutamide, rosuvastatin, capecitabine, lapatinib, metformin, and simvastatin. These trials are conducted on various cancer cell lines such as prostate, head and neck, pancreatic lung, and breast cancer on neoplasm and solid tumours. Ouabain had  $IC_{50}$  values for H460 and PANC1 of 10.44 nM and 42.36 nM respectively. The  $IC_{50}$  value of bufalin is 20.0 nM for breast cancer, 16.6 nM for cervical cancer, 28.23 nM for gallbladder cancer and 15.57 nM for lung cancer. Imperatorin inhibits colon cancer with an  $IC_{50}$  value of 78 nM. In

combination with quercetin, imperatorin showed the synergistic effect by reducing the cell viability of HeLa cells to 52.86% and for Hep-2 cells to 39.34%. Esculetin inhibits the HCC cell with an  $IC_{50}$  value of 2.24 nM and reduces the tumor growth by 20.33, 40.37, and 55.42% in Hepa1-6 cell-containing mice. Cardiac glycosides such as digitoxin, digitoxigenin, lanatoside, oleandrin and neritaloside showed anticancer potential with significant  $IC_{50}$  values of 22 nM, 15 nM, 19 nM, 50 nM, and 90 nM on lung cancer (A549 and H1975), and of 59 nM, 43 nM, 45 nM, 1104 nM, and 165 nM on osteosarcoma (U2OS and SaOS-2) respectively.

Amongst miscellaneous isolated compounds epigallocatechin curcumin, lycopene, and resveratrol are under clinical trials. Totally 32 clinical trials were studied on epigallocatechin, of which 16 studies are completed, 6 trials are under recruiting, 1 study is enrolled by invitation, 1 study is active but this study is not recruiting, 6 studies are terminated and 2 trials are withdrawn. Camphor has one completed clinical trial related to the feasibility of recruiting pediatric patients receiving chemotherapy for cancer towards homeopathy. Fisetin has one clinical trial and this trial is not yet recruiting, which is related to the efficacy of the combination of dasatinib with quercetin and fisetin to reduce senescence and to improve frailty in adult survivors of childhood cancer. Silymarin has 6 clinical trials and all 6 are completed, and these trials were for colorectal, breast cancer, prostate cancer, upper GI cancer, colon, and leukemia. A total of 17 clinical trials have been done on combretastatin, in which 11 studies are completed, 4 studies are terminated, 1 trial has unknown status and 1 trial is withdrawn. These trials include head and neck cancer, sarcoma, neuroendocrine tumor, and solid tumors. Linalool shows anticancer potential against prostate cancer with  $IC_{50}$  values of 28.3 and 10.5 nM at 24 h and 48 h respectively.

Based on the aforementioned findings from the literature, we can state that each plant bioactive has a different method of action that targets different cancer cells. Alkaloids typically prevent cancer by blocking the replication of DNA and causing protein denaturation, and this leads to apoptosis. Moreover, alkaloids inhibit the caspase inhibitor and the  $G_2/M$  phase,





Fig. 8 Basic mechanism of action of some bioactive compounds in cancer management.

while glycosides decrease the proliferation of cells by altering the expression of IB phosphorylation, BCL-2, caspase 3, and BAX proteins. Cardiac glycosides block the transport of sodium ions across the membrane, and this causes an increase in the concentration of calcium ions in the plasma membrane, which is involved in the regulation of multiple signal pathways, including apoptosis. The basic mechanism of action of these secondary metabolites is also represented in Fig. 8. Among the seven compounds currently undergoing clinical trials, curcumin, resveratrol and berberine stand out as the most significant. Clinical investigations of curcumin are assessing its efficacy against various cancers, such as colorectal, pancreatic, and breast cancers. However, curcumin's low solubility in water restricts its absorption within the gastrointestinal tract, leading to a low concentration of the compound in the bloodstream and posing challenge in achieving therapeutic levels.<sup>177</sup> Researchers are actively seeking methods to improve its bioavailability, given that the natural form of curcumin is poorly absorbed by the body. Additionally, there is some evidence regarding the ideal dosage and administration frequency for curcumin, especially in the context of cancer treatment. This ambiguity complicates the design of clinical trials and hinders the ability to compare findings across different studies. In the case of resveratrol, clinical trials are investigating its potential in the prevention or treatment of cancers, including breast, prostate, and colon cancer. However, similar to curcumin, the low

bioavailability of resveratrol presents a challenge for therapeutic use, with resveratrol being rapidly metabolized and excreted from the body, resulting in low plasma concentrations. It is extensively metabolized in the liver and intestines, limiting the amount that enters the bloodstream and target tissues.<sup>178</sup> While resveratrol is generally considered safe at lower doses, higher doses may cause side effects, such as gastrointestinal upset and, in some cases, kidney damage. Clinical investigations of berberine are currently underway to test the effectiveness of berberine in various types of cancer, including lung and colon cancer, as well as to determine its potential synergistic effects with conventional chemotherapy. Berberine can inhibit enzymes involved in drug metabolism, such as cytochrome P450 enzymes, which may interact with other medications and reduce their effectiveness or increase the risk of side effects.<sup>179</sup>

## 5 Conclusion

Medicinal plants contain a significant number of secondary metabolites belonging to various categories such as alkaloids, glycosides, flavonoids, terpenes, and terpenoids. These compounds have shown promising anticancer properties against multiple types of cancers. Extensive literature reviews have indicated that each active compound from plants exhibits a distinct mechanism of action in the treatment of different cancers. Additionally, certain phytoconstituents such as vinca



alkaloids, taxane diterpenoids, camptothecin derivatives, and epipodophyllotoxin are currently utilized in cancer therapy. Meanwhile, berberine, curcumin, lycopene, quercetin, resveratrol, and sulforaphane are currently under clinical trials. In this work, the authors reviewed a large number of secondary metabolites, which play an important role in preventing and treating various types of cancers and are under different stages of clinical trials. To conclude, we can say that, plant-derived bioactives hold tremendous anticancer potential, which could lead to the establishment of novel therapeutic agents. However, persistent study is required to discover the uncovered moieties of new plants with anticancer potential, which may offer breakthrough for improving anticancer therapy.

## Data availability

No primary research results have been included and no new data were generated or analysed as part of this review.

## Conflicts of interest

All the authors declared no conflict for the submitted manuscript.

## Acknowledgements

The authors acknowledge the Vivekanand Library, Maharshi Dayanand University Rohtak, for providing necessary literature facilities for the present study.

## References

- 1 D. T. Debela, S. G. Muzazu, K. D. Heraro, M. T. Ndalama, B. W. Mesele, D. C. Haile, S. K. Kitui and T. Manyazewal, New approaches and procedures for cancer treatment: current perspectives, *SAGE Open Med.*, 2021, **9**, 20503121211034370.
- 2 R. L. Siegel, A. N. Giaquinto and A. Jemal, Cancer statistics, *Ca-Cancer J. Clin.*, 2024, **74**, 12–49.
- 3 F. Artosi, G. Costanza, M. Di Prete, V. Garofalo, F. Lozzi, E. Dika, T. Cosio, L. Diluvio, R. G. Shumak, S. Lambiase, C. Di Raimondo, S. Campa, P. Piscitelli, A. Miani, L. Bianchi and E. Campione, Epidemiological and clinical analysis of exposure-related factors in non-melanoma skin cancer: a retrospective cohort study, *Environ. Res.*, 2024, **247**, 118117.
- 4 H. Sung, J. Ferlay, R. L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal and F. Bray, Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries, *Ca-Cancer J. Clin.*, 2021, **71**, 209–249.
- 5 M. Kapalczyńska, T. Kolenda, W. Przybyła, M. Zajączkowska, A. Teresiak, V. Filas, M. Ibbs, R. Bliźniak, Ł. Łuczewski and K. Lamperska, 2D and 3D cell cultures—a comparison of different types of cancer cell cultures, *Arch. Med. Sci.*, 2018, **14**, 910–919.
- 6 U. Leiter, U. Keim and C. Garbe, Epidemiology of Skin Cancer: Update 2019, in: *Sunlight, Vitamin D and Skin Cancer*, *Adv. Exp. Med. Bio.*, ed. J. Reichrath, 2020, p. 1268.
- 7 I. G. Lempesis, V. E. Georgakopoulou, P. Papalexis, G. P. Chrousos and D. A. Spandidos, Role of stress in the pathogenesis of cancer, *Int. J. Oncol.*, 2023, **63**, 1–4.
- 8 O. V. Bukhtoyarov and D. M. Samarin, Pathogenesis of Cancer: Cancer Reparative Trap, *J. Cancer Ther.*, 2015, **6**, 399–412.
- 9 Z. Wang and X. Wu, Abnormal function of telomere protein TRF2 induces cell mutation and the effects of environmental tumor-promoting factors, *Oncol. Rep.*, 2021, **46**, 1–20.
- 10 S. P. Leong and M. H. Witte, Cancer metastasis through the lymphatic versus blood vessels, *Clin. Exp. Metastasis*, 2024, **41**, 387–402.
- 11 E. N. Kontomanolis, A. Koutras, A. Syllaios, D. Schizas, S. Kalagasidou, A. Pagkalos, D. Alatzidou, P. Kantari, T. Ntounis and Z. Fasoulakis, Basic principles of molecular biology of cancer cell-Molecular cancer indicators, *J. BUON*, 2021, **26**, 1723–1734.
- 12 G. Gaobotse, S. Venkataraman, P. D. Brown, K. Masisi, T. E. Kwape, D. O. Nkwe, G. Rantong and A. Makhzoum, The use of African medicinal plants in cancer management, *Front. Pharmacol.*, 2023, **14**, 1122388.
- 13 P. Bajpai, S. Usmani, R. Kumar and O. Prakash, Recent advances in anticancer approach of traditional medicinal plants: a novel strategy for cancer chemotherapy, *Intell. Pharm.*, 2024, **2**, 291–304.
- 14 H. Y. Yip and A. Papa, Signaling pathways in cancer: therapeutic targets, combinatorial treatments, and new developments, *Cells*, 2021, **10**, 659.
- 15 J. Yi, F. Peng, J. Zhao and X. Gong, METTL3/IGF2BP2 axis affects the progression of colorectal cancer by regulating m6A modification of STAG3, *Sci. Rep.*, 2023, **13**, 17292.
- 16 M. Ghanbari-Movahed, T. Kaceli, A. Mondal, M. H. Farzaei and A. Bishayee, Recent Advances in Improved Anticancer Efficacies of Camptothecin Nano-Formulations: A Systematic Review, *Biomedicines*, 2021, **9**, 480.
- 17 E. Matrineo, G. Casamassima, S. Castiglione, E. Cellupica, S. Pantalone, F. Papagni, M. Rui, A. M. Siciliano and S. Collina, Vinca alkaloids and analogues as anti-cancer agents: looking back, peering ahead, *Bioorg. Med. Chem. Lett.*, 2018, **28**(17), 2816–2826.
- 18 C. Zhang, J. Sheng, G. Li, L. Zhao, Y. Wang, W. Yang, X. Yao, L. Sun, Z. Zhang and R. Cui, Effects of Berberine and Its Derivatives on Cancer: A Systems Pharmacology Review, *Front. Pharmacol.*, 2020, **10**, 1461.
- 19 C. Wen, L. Wu, L. Fu, X. Zhang and H. Zhou, Berberine enhances the anti-tumor activity of tamoxifen in drug-sensitive MCF-7 and drug-resistant MCF-7/TAM cells, *Mol. Med. Rep.*, 2016, **14**, 2250–2256.
- 20 C. Sheng, Z. Miao and W. Zhang, Topoisomerase I Inhibitors Derived from Natural Products: Structure–Activity Relationships and Antitumor Potency, Atta-ur-Rahman editors, *Stud. Nat. Prod. Chem.*, 2016, **47**, 1–28.



- 21 H. Zhang, L. Chen, X. Sun, Q. Yang, L. Wan and C. Guo, Matrine: A Promising Natural Product With Various Pharmacological Activities, *Front. Pharmacol.*, 2020, **11**, 588.
- 22 L. Jiang, L. Wu, F. Yang, N. Almosnid, X. Liu, J. Jiang, E. Altman, L. Wang and Y. Gao, Synthesis, biological evaluation and mechanism studies of matrine derivatives as anticancer agents, *Oncol. Lett.*, 2017, **14**, 3057–3064.
- 23 J. J. Nair, J. Bastida, F. Viladomat and J. van Staden, Cytotoxic agents of the crinine series of amaryllidaceae alkaloids, *Nat. Prod. Commun.*, 2012, **7**, 1677–1688.
- 24 B. Abebe, S. Tadesse, A. Hymete and D. Bisrat, Antiproliferative Effects of Alkaloids from the Bulbs of *Crinum abyscinicum* Hochst. ExA. Rich, *Evid.-Based Complementary Altern. Med.*, 2020, **2020**, 2529730.
- 25 G. Luchetti, R. Johnston, V. Mathieu, F. Lefranc, K. Hayden, A. Andolfi, D. Lamoral-Theys, M. R. Reisenauer, C. Champion, S. C. Pelly, W. A. Van Otterlo, I. V. Magedov, R. Kiss, A. Evidente, S. Rogelj and A. Kornienko, Bulbispermine: a crinine-type Amaryllidaceae alkaloid exhibiting cytostatic activity toward apoptosis-resistant glioma cells, *ChemMedChem*, 2012, **7**, 815–822.
- 26 M. M. He, C. R. Qu, O. D. Gao, X. M. Hu and X. C. Hong, Biological and pharmacological activities of amaryllidaceae alkaloids, *RSC Adv.*, 2015, **5**, 16562–16574.
- 27 K. Habartová, L. Cahliková, M. Řezáčová and R. Havelek, The Biological Activity of Alkaloids from the Amaryllidaceae: From Cholinesterases Inhibition to Anticancer Activity, *Nat. Prod. Commun.*, 2016, **11**, 1587–1594.
- 28 M. Roy, L. Liang, X. Xiao, P. Feng, M. Ye and J. Liu, Lycorine: a prospective natural lead for anticancer drug discovery, *Biomed. Pharmacother.*, 2018, **107**, 615–624.
- 29 G. Van Goietsenoven, A. Andolfi, B. Lallemand, A. Cimmino, D. Lamoral-Theys, T. Gras, A. Abou-Donia, J. Dubois, F. Lefranc, V. Mathieu, A. Kornienko, R. Kiss and A. Evidente, Amaryllidaceae alkaloids belonging to different structural subgroups display activity against apoptosis-resistant cancer cells, *J. Nat. Prod.*, 2010, **73**, 1223–1227.
- 30 J. J. Nair and J. van Staden, Cytotoxicity studies of lycorine alkaloids of the Amaryllidaceae, *Nat. Prod. Commun.*, 2014, **9**(8), 1193–1210.
- 31 A. Manayi, S. M. Nabavi, W. N. Setzer and S. Jafari, Piperine as a Potential Anti-cancer Agent: A Review on Preclinical Studies, *Curr. Med. Chem.*, 2018, **25**, 4918–4928.
- 32 E. Turrini, P. Sestili and C. Fimognari, Overview of the anticancer potential of the “King of Spices” *Piper nigrum* and its main constituent piperine, *Toxins*, 2020, **12**, 747.
- 33 M. Sun, W. Lou, J. Y. Chun, D. S. Cho, N. Nadiminty, C. P. Evans, J. Chen, J. Yue, Q. Zhou and A. C. Gao, Sanguinarine suppresses prostate tumor growth and inhibits survivin expression, *Genes Cancer*, 2010, **1**, 283–292.
- 34 S. Q. Pang, G. Q. Wang, J. S. Lin, Y. Diao and R. A. Xu, Cytotoxic activity of the alkaloids from *Broussonetiapapyrifera* fruits, *Pharm. Biol.*, 2014, **52**, 1315–1319.
- 35 J. Lu, J. Bao, X. Chen, M. C. Huang and Y. Wang, Alkaloids Isolated from Natural Herbs as the Anticancer Agents, *Evid.-Based Complementary Altern. Med.*, 2012, 1–12.
- 36 K. Piska, A. Gunia-Krzyżak, P. Koczurkiewicz, K. Wójcik-Pszczola and E. Pękala, Piperlongumine (piplartine) as a lead compound for anticancer agents - synthesis and properties of analogues: a mini-review, *Eur. J. Med. Chem.*, 2018, **156**, 13–20.
- 37 K. I. Mohd, A. Barman and N. Qais, Anti-Cancer constituents from plants: a brief review, *Dhaka Univ. J. Pharm. Sci.*, 2020, **19**, 83–96.
- 38 C. H. Yan, F. Li and Y. C. Ma, Plumbagin shows anticancer activity in human osteosarcoma (MG-63) cells via the inhibition of S-phase checkpoints and down-regulation of c-myc, *Int. J. Clin. Exp. Med.*, 2015, **8**, 14432–14439.
- 39 X. Q. Zhang, C. Y. Yang, X. F. Rao and J. P. Xiong, Plumbagin shows anti-cancer activity in human breast cancer cells by the upregulation of p53 and p21 and suppression of G1 cell cycle regulators, *Eur. J. Gynaecol. Oncol.*, 2016, **37**, 30–35.
- 40 M. A. Khan, H. C. Chen, M. Tania and D. Z. Zhang, Anticancer activities of *Nigella sativa* (black cumin), *Afr. J. Tradit., Complementary Altern. Med.*, 2011, **8**, 226–232.
- 41 J. J. Nair, J. van Staden and J. Bastida, Apoptosis-Inducing Effects of Amaryllidaceae Alkaloids, *Curr. Med. Chem.*, 2016, **23**, 161–185.
- 42 A. M. Chapa-Oliver and L. Mejía-Teniente, Capsaicin: From Plants to a Cancer-Suppressing Agent, *Molecules*, 2016, **21**, 931.
- 43 D. Ma, C. Pignatelli, D. Tarade, T. Gilbert, M. Noel, F. Mansour, S. Adams, A. Dowhayko, K. Stokes, S. Vshyvenko, T. Hudlicky, J. McNulty and S. Pandey, Cancer cell mitochondria targeting by pancratistatin analogs is dependent on functional complex ii and iii, *Sci. Rep.*, 2017, **7**, 42957.
- 44 C. Griffin, C. Hamm, J. McNulty and S. Pandey, Pancratistatin induces apoptosis in clinical leukemia samples with minimal effect on non-cancerous peripheral blood mononuclear cells, *Cancer Cell Int.*, 2010, **10**, 6.
- 45 Y. Wang, Y. Liu, X. Du, H. Ma and J. Yao, The Anti-Cancer Mechanisms of Berberine: A Review, *Cancer Manage. Res.*, 2020, **12**, 695–702.
- 46 T. Koltai, Re-Purposing Evodiamine as an Anti-Cancer Drug: Effects on Migration and Apoptosis, *Open Access J. Oncol. Med.*, 2018, **1**(3), 37–51.
- 47 J. Jia, X. Kang, Y. Liu and J. Zhang, Inhibition of human liver cancer cell growth by evodiamine involves apoptosis and deactivation of PI3K/AKT pathway, *Appl. Biol. Chem.*, 2020, **63**, 67.
- 48 Q. An, C. Han, Y. Zhou, F. Li, D. Li, X. Zhang, Z. Yu, Z. Duan and Q. Kan, Matrine induces cell cycle arrest and apoptosis with recovery of the expression of miR-126 in the A549 non-small cell lung cancer cell line, *Mol. Med. Rep.*, 2016, **14**, 4042–4048.



- 49 H. Huang, Q. Wang, T. Du, C. Lin, Y. Lai, D. Zhu, W. Wu, X. Ma, S. Bai, Z. Li, L. Liu and Q. Li, Matrine inhibits the progression of prostate cancer by promoting expression of GADD45B, *Prostate*, 2018, **78**, 327–335.
- 50 J. J. Nair, L. Rárová, M. Strnad, J. Bastida and J. van Staden, Apoptosis-inducing effects of distichamine and narciprimine, rare alkaloids of the plant family Amaryllidaceae, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 6195–6199.
- 51 G. Xin, M. Yu, Y. Hu, S. Gao, Z. Qi, Y. Sun, W. Yu and J. He, Yubin, Effect of lycorine on the structure and function of hepatoma cell membrane in vitro and in vivo, *Biotechnol. Biotechnol. Equip.*, 2020, **34**, 104–114.
- 52 A. T. Mbaveng, G. Bitchagno, V. Kuete, P. Tane and T. Efferth, Cytotoxicity of ungeremine towards multi-factorial drug resistant cancer cells and induction of apoptosis, ferroptosis, necroptosis and autophagy, *Phytomedicine*, 2019, **60**, 152832.
- 53 R. A. Rather and M. Bhagat, Cancer chemoprevention and piperine: molecular mechanisms and therapeutic opportunities, *Front. Cell Dev. Biol.*, 2018, **6**, 10.
- 54 I. W. Achkar, F. Mraiche, R. M. Mohammad and S. Uddin, Anticancer potential of sanguinarine for various human malignancies, *Future Med. Chem.*, 2017, **9**, 933–950.
- 55 F. Luan, X. He and N. Zeng, Tetrandrine: a review of its anticancer potentials, clinical settings, pharmacokinetics and drug delivery systems, *J. Pharm. Pharmacol.*, 2020, **72**, 1491–1512.
- 56 D. P. Bezerra, C. Pessoa, M. O. de Moraes, N. Saker-Neto, E. R. Silveira and L. V. Costa-Lotufo, Overview of the therapeutic potential of piplartine (piperlongumine), *Eur. J. Pharm. Sci.*, 2013, **48**, 453–463.
- 57 Z. Yin, J. Zhang, L. Chen, Q. Guo, B. Yang, W. Zhang and W. Kang, Anticancer Effects and Mechanisms of Action of Plumbagin: Review of Research Advances, *BioMed Res. Int.*, 2020, **2020**, 6940953.
- 58 M. A. Khan, M. Tania, S. Fu and J. Fu, Thymoquinone, as an anticancer molecule: from basic research to clinical investigation, *Oncotarget*, 2017, **8**, 51907–51919.
- 59 J. R. Friedman, N. A. Nolan, K. C. Brown, S. L. Miles, A. T. Akers, K. W. Colclough, J. M. Seidler, J. M. Rimoldi, M. A. Valentovic and P. Dasgupta, Anticancer Activity of Natural and Synthetic Capsaicin Analogs, *J. Pharmacol. Exp. Ther.*, 2018, **364**, 462–473.
- 60 G. C. Arya, K. Kaur and V. Jaitak, Isoxazole derivatives as anticancer agent: a review on synthetic strategies, mechanism of action and SAR studies, *Eur. J. Med. Chem.*, 2021, **221**, 113511.
- 61 H. Ruan, Y. Y. Zhan, J. Hou, B. Xu, B. Chen, Y. Tian, D. Wu, Y. Zhao, Y. Zhang, X. Chen, P. Mi, L. Zhang, S. Zhang, X. Wang, H. Cao, W. Zhang, H. Wang, H. Li, Y. Su, X. K. Hu and T. Zhang, Berberine binds RXR  $\alpha$  to suppress beta-catenin signaling in colon cancer cells, *Oncogene*, 2017, **36**, 6906–6918.
- 62 Y. Wang and S. L. Zhang, Berberine suppresses growth and metastasis of endometrial cancer cells via miR-101/COX-2, *Biomed. Pharmacother.*, 2018, **103**, 1287–1293.
- 63 R. G. Xiong, S. Y. Huang, S. X. Wu, D. D. Zhou, Z. J. Yang, A. Saimaiti, C. N. Zhao, A. Shang, Y. J. Zhang, R. Y. Gan and H. B. Li, Anticancer Effects and Mechanisms of Berberine from Medicinal Herbs: An Update Review, *Molecules*, 2022, **27**, 4523.
- 64 S. Guo, Y. Chen, C. Pang, X. Wang, S. Shi, H. Zhang, H. An and Y. Zhan, Matrine is a novel inhibitor of the TMEM16A chloride channel with antilung adenocarcinoma effects, *J. Cell. Physiol.*, 2019, **234**(6), 8698–8708.
- 65 X. Xiao, M. Ao, F. Xu, X. Li, J. Hu, Y. Wang, D. Li, X. Zhu, C. Xin and W. Shi, Effect of matrine against breast cancer by downregulating the vascular endothelial growth factor via the Wnt/ $\beta$ -catenin pathway, *Oncol. Lett.*, 2018, **15**(2), 1691–1697.
- 66 W. H. Talib, Regressions of Breast Carcinoma Syngraft Following Treatment with Piperine in Combination with Thymoquinone, *Sci. Pharm.*, 2017, **85**, 27.
- 67 J. Liu, S. Sun, C. Zhou, Z. Sun, Q. Wang and C. Sun, In vitro and in vivo anticancer activity of Lycorine in prostate cancer by inhibiting NF- $\kappa$ B signaling pathway, *J. Cancer*, 2022, **13**, 3151–3159.
- 68 G. Xin, M. Yu, Y. Hu, S. Gao, Z. Qi, Y. Sun, W. Yu, J. He and Y. Ji, Effect of lycorine on the structure and function of hepatoma cell membrane in vitro and in vivo, *Biotechnol. Biotechnol. Equip.*, 2020, **34**, 104–114.
- 69 Q. Guo, Y. Liu, J. Zhao, J. Wang, Y. Li, Y. Pang, J. Chen and J. Wang, Evodiamine inactivates NF- $\kappa$ B and potentiates the antitumor effects of gemcitabine on tongue cancer both in vitro and in vivo, *OncoTargets Ther.*, 2018, **12**, 257–267.
- 70 Z. B. Jiang, J. M. Huang, Y. J. Xie, Y. Z. Zhang, C. Chang, H. L. Lai, W. Wang, X. J. Yao, X. X. Fan, Q. B. Wu, C. Xie, M. F. Wang and E. L. Leung, Evodiamine suppresses non-small cell lung cancer by elevating CD8<sup>+</sup> T cells and downregulating the MUC1-C/PD-L1 axis, *J. Exp. Clin. Cancer Res.*, 2020, **39**, 249.
- 71 Y. H. Deng, Y. C. Dai and B. Tang, Effect of Evodiamine on the Ki-67 Expression in Tumor Tissue of Lymphoma Transplanted Mice Model, *Clin. Basic Bridging Res.*, 2020, **36**, 655–657.
- 72 D. Zeng, P. Zhou, R. Jiang, X. P. Li, S. Y. Huang, D. Y. Li, G. L. Li, L. S. Li, S. Zhao, L. Hu, J. H. Ran, D. L. Chen, Y. P. Wang and J. Li, Evodiamine inhibits vasculogenic mimicry in HCT116 cells by suppressing hypoxia-inducible factor 1- $\alpha$ -mediated angiogenesis, *Anticancer Drugs*, 2021, **32**, 314–322.
- 73 S. Y. Hyun, H. T. Le, H. Y. Min, H. Pei, Y. Lim, I. Song, Y. T. K. Nguyen, S. Hong, B. W. Han and H. Y. Lee, Evodiamine inhibits both stem cell and non-stem-cell populations in human cancer cells by targeting heat shock protein 70, *Theranostics*, 2021, **11**, 2932–2952.
- 74 N. F. Z. Schneider, C. Cerella, C. M. O. Simões and M. Diederich, Anticancer and Immunogenic Properties of Cardiac Glycosides, *Molecules*, 2017, **22**, 1932.
- 75 V. L. Muilenburg, P. L. Phelan, P. Bonello and D. A. Herms, Inter- and intra-specific variation in stem phloem phenolics of paper birch (*Betula papyrifera*) and European white birch (*Betula pendula*), *J. Chem. Ecol.*, 2011, **37**, 1193–1202.



- 76 H. Khan, M. Saeedi, S. M. Nabavi, M. S. Mubarak and A. Bishayee, Glycosides from medicinal plants as potential anticancer agents: emerging trends towards future drugs, *Curr. Med. Chem.*, 2019, **26**, 2389–2406.
- 77 L. A. Rascón-Valenzuela, C. Velázquez, A. Garibay-Escobar, W. Vilegas, L. A. Medina-Juárez, N. Gámez-Meza and R. E. Robles-Zepeda, Apoptotic activities of cardenolide glycosides from *Asclepias subulata*, *J. Ethnopharmacol.*, 2016, **193**, 303–311.
- 78 Z. A. Shah, A. Hameed, A. Ahmed, S. U. Simjee, A. Jabeen, A. Ullah and F. Shaheen, Cytotoxic and anti-inflammatory salicin glycosides from leaves of *Salix acmophylla*, *Phytochem. Lett.*, 2016, **17**, 107–113.
- 79 J. Du, L. Jiang, F. Chen, H. Hu and M. Zhou, Cardiac glycoside ouabain exerts anticancer activity via downregulation of STAT3, *Front. Oncol.*, 2021, **11**, 684316.
- 80 S. Rimpelová, T. Zimmermann, P. B. Drašar, B. Dolenský, J. Bejček, E. Kmoníčková, P. Cihlářová, S. Gurská, L. Kuklíková, M. Hajdůch, T. Ruml, L. Opletal, P. Džubák and M. Jurásek, Steroid glycosides hyrcanoside and deglucohyrcanoside: on isolation, structural identification, and anticancer activity, *Foods*, 2021, **10**, 136.
- 81 C. S. Cheng, J. Wang, J. Chen, K. T. Kuo, J. Tang, H. Gao, L. Chen, Z. Chen and Z. Meng, New therapeutic aspects of steroidal cardiac glycosides: the anticancer properties of Huachansu and its main active constituent Bufalin, *Cancer Cell Int.*, 2019, **19**, 92.
- 82 V. K. Bajpai, M. B. Alam, K. T. Quan, H. J. Choi, H. An, M. K. Ju, S. H. Lee, Y. S. Huh, Y. K. Han and M. Na, Cytotoxic properties of the anthraquinone derivatives isolated from the roots of *Rubia philippinensis*, *BMC Complementary Altern. Med.*, 2018, **18**, 200.
- 83 E. Jaszczak-Wilke, Ż. Polkowska, M. Koprowski, K. Owsianik, A. E. Mitchell and P. Bałczewski, Amygdalin: toxicity, anticancer activity and analytical procedures for its determination in plant seeds, *Molecules*, 2021, **26**, 2253.
- 84 E. Koziół and K. Skalicka-Woźniak, Imperatorin-pharmacological meaning and analytical clues: profound investigation, *Phytochem. Rev.*, 2016, **15**, 627–649.
- 85 J. H. Cho, J. C. Shin, J. J. Cho, Y. H. Choi, J. H. Shim and J. I. Chae, Esculetin (6,7-dihydroxycoumarin): a potential cancer chemopreventive agent through suppression of Sp1 in oral squamous cancer cells, *Int. J. Oncol.*, 2015, **46**, 265–271.
- 86 P. Yang, Y. Jiang, Y. Pan, X. Ding, P. Rhea, J. Ding, D. H. Hawke, D. Felsner, G. Narla, Z. Lu and R. T. Lee, Mistletoe extract Fraxini inhibits the proliferation of liver cancer by down-regulating c-Myc expression, *Sci. Rep.*, 2019, **9**, 6428.
- 87 S. Rosselli, A. M. Maggio, N. Faraone, V. Spadaro, S. L. Morris-Natschke, K. F. Bastow, K. H. Lee and M. Bruno, The cytotoxic properties of natural coumarins isolated from roots of *Ferulago campestris* (Apiaceae) and of synthetic ester derivatives of aegelinol, *Nat. Prod. Commun.*, 2009, **4**, 1701–1706.
- 88 J. Portugal, Chartreusin, elsamycin A and related anticancer antibiotics, *Curr. Med. Chem.: Anti-Cancer Agents*, 2003, **3**, 411–420.
- 89 J. Zhang, M. Kurita, T. Shinozaki, M. Ukiya, K. Yasukawa, N. Shimizu, H. Tokuda, E. T. Masters, M. Akihisa and T. Akihisa, Triterpene glycosides and other polar constituents of shea (*Vitellaria paradoxa*) kernels and their bioactivities, *Phytochemicals*, 2014, **108**, 157–170.
- 90 D. Ataseven, A. Öztürk, M. Özkara and Z. Joha, Anticancer activity of lycopene in HT-29 colon cancer cell line, *Med. Oncol.*, 2023, **24**(40), 127.
- 91 X. Chen, P. Xu, H. Zhang, X. Su, L. Guo, X. Zhou, J. Wang, P. Huang, Q. Zhang and R. Sun, EGFR and ERK activation resists flavonoid quercetin-induced anticancer activities in human cervical cancer cells in vitro, *Oncol. Lett.*, 2021, **22**, 1–3.
- 92 K. Andrijauskaite, J. Morris and M. J. Wargovich, *Natural Anticancer Agents*, Academic Press, 2019, pp. 49–73.
- 93 A. N. Adham, S. Abdelfatah, A. M. Naqishbandi, N. Mahmoud and T. Efferth, Cytotoxicity of apigenin toward multiple myeloma cell lines and suppression of iNOS and COX-2 expression in STAT1-transfected HEK293 cells, *Phytomedicine*, 2021, **80**, 153371.
- 94 J. E. Lee, N. T. T. Thuy, J. Lee, N. Cho and H. M. Yoo, Platyphylloside isolated from *betula platyphylla* is antiproliferative and induces apoptosis in colon cancer and leukemic cells, *Molecules*, 2019, **24**, 2960.
- 95 W. P. Zheng, F. Y. Huang, S. Z. Dai, J. Y. Wang, Y. Y. Lin, Y. Sun, G. H. Tan and Y. H. Huang, Toxicarioside O Inhibits Cell Proliferation and Epithelial-Mesenchymal Transition by Downregulation of Trop2 in Lung Cancer Cells, *Front. Oncol.*, 2021, **10**, 609275.
- 96 N. Tawfeek, M. F. Mahmoud, D. I. Hamdan, M. Sobeh, N. Farrag, M. Wink and A. M. El-Shazly, Phytochemistry, Pharmacology and Medicinal Uses of Plants of the Genus *Salix*: An Updated Review, *Front. Pharmacol.*, 2021, **12**, 593856.
- 97 H. A. Elbaz, T. A. Stueckle, W. Tse, Y. Rojanasakul and C. Z. Dinu, Digitoxin and its analogs as novel cancer therapeutics, *Exp. Hematol. Oncol.*, 2012, **1**, 4.
- 98 G. Y. Yao, W. Dai, M. Ye, R. Huang, Y. M. Pan, Z. Liao and H. S. Wang, Synthesis and antitumor properties of novel alizarin analogs, *Med. Chem. Res.*, 2014, **23**, 5031–5042.
- 99 P. Liczbiński and B. Bukowska, Molecular mechanism of amygdalin action in vitro: review of the latest research, *Immunopharmacol. Immunotoxicol.*, 2018, **40**, 212–218.
- 100 R. K. Singh, T. S. Lange, K. K. Kim and L. Brard, A coumarin derivative (RKS262) inhibits cell-cycle progression, causes pro-apoptotic signaling and cytotoxicity in ovarian cancer cells, *Invest. New Drugs*, 2011, **29**, 63–72.
- 101 J. Rohr and C. Hertweck, Type II PKS, in *Comprehensive Natural Products II*, Elsevier, 2010, pp. 227–303.
- 102 K. Sahin, E. Yenice, M. Tuzcu, C. Orhan, C. Mizrak, I. H. Ozercan, N. Sahin, B. Yilmaz, B. Bilir, B. Ozpolat and O. Kucuk, Lycopene Protects Against Spontaneous Ovarian Cancer Formation in Laying Hens, *J. Cancer Prev.*, 2018, **23**, 25–36.
- 103 K. Aizawa, C. Liu, S. Tang, S. Veeramachaneni, K. Q. Hu, D. E. Smith and X. D. Wang, Tobacco carcinogen induces both lung cancer and non-alcoholic steatohepatitis and



- hepatocellular carcinomas in ferrets which can be attenuated by lycopene supplementation, *Int. J. Cancer*, 2016, **139**, 1171–1181.
- 104 Y. Li, Z. Wang, J. Jin, S. X. Zhu, G. Q. He, S. H. Li, J. Wang and Y. Cai, Quercetin pretreatment enhances the radiosensitivity of colon cancer cells by targeting Notch-1 pathway, *Biochem. Biophys. Res. Commun.*, 2020, **523**, 947–953.
  - 105 L. Jia, S. Huang, X. Yin, Y. Zan, Y. Guo and L. Han, Quercetin suppresses the mobility of breast cancer by suppressing glycolysis through Akt-mTOR pathway mediated autophagy induction, *Life Sci.*, 2018, **208**, 123–130.
  - 106 X. B. Yan, T. Xie, S. D. Wang, Z. Wang, H. Y. Li and Z. M. Ye, Apigenin inhibits proliferation of human chondrosarcoma cells via cell cycle arrest and mitochondrial apoptosis induced by ROS generation-an in vitro and in vivo study, *Int. J. Clin. Exp. Med.*, 2018, **11**, 1615–1631.
  - 107 J. H. Chang, C. W. Cheng, Y. C. Yang, W. S. Chen, W. Y. Hung, J. M. Chow, P. S. Chen, M. Hsiao, W. J. Lee and M. H. Chien, Downregulating CD26/DPPIV by apigenin modulates the interplay between Akt and Snail/Slug signaling to restrain metastasis of lung cancer with multiple EGFR statuses, *J. Exp. Clin. Cancer Res.*, 2018, **37**, 199.
  - 108 Y. Wang, Q. Ma, S. Zhang, H. Liu, B. Zhao, B. Du, W. Wang, P. Lin, Z. Zhang, Y. Zhong and D. Kong, Digoxin enhances the anticancer effect on non-small cell lung cancer while reducing the cardiotoxicity of adriamycin, *Front. Pharmacol.*, 2020, **11**, 186.
  - 109 H. Gan, M. Qi, C. Chan, P. Leung, G. Ye, Y. Lei, A. Liu, F. Xue, D. Liu, W. Ye, D. Zhang, L. Deng and J. Chen, Digoxin inhibits HeLa cell growth through the induction of G2/M cell cycle arrest and apoptosis in vitro and in vivo, *Int. J. Oncol.*, 2020, **57**, 562–573.
  - 110 M. Liu, L. X. Feng, P. Sun, W. Liu, W. Y. Wu, B. H. Jiang, M. Yang, L. H. Hu, D. A. Guo and X. Liu, A novel bufalin derivative exhibited stronger apoptosis-inducing effect than bufalin in A549 lung cancer cells and lower acute toxicity in mice, *PLoS One*, 2016, **11**, e0159789.
  - 111 H. T. Chen, D. Sun, Y. C. Peng, P. H. Kao and Y. L. Wu, Novel augmentation by bufalin of protein kinase C-induced cyclooxygenase-2 and IL-8 production in human breast cancer cells, *Innate Immun.*, 2017, **23**, 54–66.
  - 112 Z. Xu, Y. Hou, C. Zou, H. Liang, J. Mu, X. Jiao, Y. Zhu, L. Su, M. Liu, X. Chen, C. Qian, X. Zhu, W. Gong, Q. Dong and F. Zhang, Alizarin, a nature compound, inhibits the growth of pancreatic cancer cells by abrogating NF- $\kappa$ B activation, *Int. J. Biol. Sci.*, 2022, **18**, 2759–2774.
  - 113 C. A. Brohem, T. C. Sawada, R. R. Massaro, R. L. Almeida, D. P. Rivelli, C. D. Ropke, V. V. da Silva, T. M. de Lima, R. Curi, S. B. Barros and S. S. Maria-Engler, Apoptosis induction by 4-nerolidylcatechol in melanoma cell lines, *Toxicol. In Vitro*, 2008, **23**, 111–119.
  - 114 M. S. Ahmed and F. T. Halaweish, Cucurbitacins: potential candidates targeting mitogen-activated protein kinase pathway for treatment of melanoma, *J. Enzyme Inhib. Med. Chem.*, 2014, **29**, 162–167.
  - 115 S. K. Król, M. Kielbas, A. Rivero-Müller and A. Stepulak, Comprehensive review on betulin as a potent anticancer agent, *BioMed Res. Int.*, 2015, **2015**, 584189.
  - 116 M. Schröder, M. Petrova, Z. Vlahova, G. M. Dobrikov, I. Slavchev, E. Pasheva and I. Ugrinova, In vitro anticancer activity of two ferrocene-containing camphor sulfonamides as promising agents against lung cancer cells, *Biomedicines*, 2022, **10**, 1353.
  - 117 I. Begum, R. Sharma and H. K. Sharma, A review on plants having anti-cancer activity, *Curr. Trends Pharm. Res.*, 2017, **4**, 39–62.
  - 118 X. Lin, Z. Peng and C. Su, Potential anti-cancer activities and mechanisms of costunolide and dehydrocostuslactone, *Int. J. Mol. Sci.*, 2015, **16**, 10888–10906.
  - 119 S. Singh, P. Gupta, A. Meena and S. Luqman, Acacetin, a flavone with diverse therapeutic potential in cancer, inflammation, infections and other metabolic disorders, *Food Chem. Toxicol.*, 2020, **145**, 111708.
  - 120 D. N. Syed, F. Afaq, N. Maddodi, J. J. Johnson, S. Sarfaraz, A. Ahmad, V. Setaluri and H. Mukhtar, Inhibition of human melanoma cell growth by the dietary flavonoid fisetin is associated with disruption of Wnt/ $\beta$ -catenin signaling and decreased Mitf levels, *J. Invest. Dermatol.*, 2011, **131**, 1291–1299.
  - 121 K. Ramasamy and R. Agarwal, Multitargeted therapy of cancer by silymarin, *Cancer Lett.*, 2008, **269**, 352–362.
  - 122 D. Delmas, J. Xiao, A. Vejux and V. Aires, Silymarin and cancer: a dual strategy in both in chemoprevention and chemosensitivity, *Molecules*, 2020, **25**, 2009.
  - 123 A. Riaz, A. Rasul, G. Hussain, M. K. Zahoor, F. Jabeen, Z. Subhani, T. Younis, M. Ali, I. Sarfraz and Z. Selamoglu, Astragalin: A Bioactive Phytochemical with Potential Therapeutic Activities, *Adv. Pharmacol. Sci.*, 2018, **2018**, 9794625.
  - 124 Y. Park, S. H. Woo, S. K. Seo, H. Kim, W. C. Noh, J. K. Lee, B. M. Kwon, K. N. Min, T. B. Choe and I. C. Park, Ginkgetin induces cell death in breast cancer cells via downregulation of the estrogen receptor, *Oncol. Lett.*, 2017, **14**, 5027–5033.
  - 125 M. Nihal, N. Ahmad, H. Mukhtar and G. S. Wood, Anti-proliferative and proapoptotic effects of (-)-epigallocatechin-3-gallate on human melanoma: possible implications for the chemoprevention of melanoma, *Int. J. Cancer*, 2005, **114**, 513–521.
  - 126 N. Zhang, X. Shen, X. Jiang, J. Cai, X. Shen, Y. Hu and S. X. Qiu, Two new cytotoxic stilbenoid dimers isolated from *Cajanus cajan*, *J. Nat. Med.*, 2018, **201**(72), 304–309.
  - 127 E. Navarro, S. J. Alonso, P. J. Alonso, J. Trujillo, E. Jorge and C. Perez, Pharmacological effects of elenoside, an aryl-naphthalene lignan, *Biol. Pharm. Bull.*, 2001, **24**, 254–258.
  - 128 S. B. Kntayya, M. D. Ibrahim, N. M. Ain, R. Iori, C. Ioannides and A. F. A. Razis, Induction of apoptosis and cytotoxicity by isothiocyanate sulforaphene in human hepatocarcinoma HepG2 cells, *Nutrients*, 2018, **10**, 718.



- 129 X. D. Pei, Z. L. He, H. L. Yao, J. S. Xiao, L. Li, J. Z. Gu, P. Z. Shi, J. H. Wang and L. H. Jiang, 6-Shogaol from ginger shows anti-tumor effect in cervical carcinoma via PI3K/Akt/mTOR pathway, *Eur. J. Nutr.*, 2021, **1**, 1–3.
- 130 M. Ossama, R. M. Hathout, D. A. Attia and N. D. Mortada, Enhanced allicin cytotoxicity on HEPG-2 cells using glycyrrhetic acid surface-decorated gelatin nanoparticles, *ACS Omega*, 2019, **4**, 11293–11300.
- 131 S. B. Ateba, M. A. Mvondo, S. Djiogue, S. Zingué, L. Krenn and D. Njamen, A pharmacological overview of alpinumisoflavone, a natural prenylated isoflavonoid, *Front. Pharmacol.*, 2019, **10**, 952.
- 132 R. Raghavan, S. Cheriyaundath and J. Madassery, Exploring the mechanisms of cytotoxic and anti-inflammatory property of andrographolide and its derivatives, *Pharmacogn. Rev.*, 2018, **12**, 56–65.
- 133 X. Zhao, J. Qu, X. Liu, J. Wang, X. Ma, X. Zhao, Q. Yang, W. Yan, Z. Zhao, Y. Hui and H. Bai, Baicalein suppress EMT of breast cancer by mediating tumor-associated macrophages polarization, *Am. J. Cancer Res.*, 2018, **8**, 1528–1540.
- 134 Y. Chu, Q. Yuan, H. Jiang, L. Wu, Y. Xie, X. Zhang and L. Li, A comprehensive review of the anticancer effects of decursin, *Front. Pharmacol.*, 2024, **15**, 1303412.
- 135 Z. A. Rahman, F. Hidayatullah, P. K. Pratama, D. P. Andhika and L. Hakim, Effects of decursinol angelate on viability and apoptosis in PC-3 prostate cancer cells: in vitro study, *Narra J*, 2024, **4**, e948.
- 136 D. Aras, O. Cinar, Z. Cakar, S. Ozkavukcu and A. Can, Can dicoumarol be used as a gonad-safe anticancer agent: an in vitro and in vivo experimental study, *Mol. Hum. Reprod.*, 2016, **22**, 57–67.
- 137 H. S. Chae, R. Xu, J. Y. Won, Y. W. Chin and H. Yim, Molecular targets of genistein and its related flavonoids to exert anticancer effects, *Int. J. Mol. Sci.*, 2019, **20**, 2420.
- 138 S. Nafees, M. Zafaryab, S. H. Mehdi, B. Zia, M. A. Rizvi and M. A. Khan, Anti-cancer effect of gingerol in cancer prevention and treatment, *Anti-Cancer Agents Med. Chem.*, 2021, **21**, 428–432.
- 139 Y. Cai, B. Zhao, Q. Liang, Y. Zhang, J. Cai and G. Li, The selective effect of glycyrrhizin and glycyrrhetic acid on topoisomerase II $\alpha$  and apoptosis in combination with etoposide on triple negative breast cancer MDA-MB-231 cells, *Eur. J. Pharmacol.*, 2017, **809**, 87–97.
- 140 H. A. Kim and J. Lee, Hispidulin modulates epithelial-mesenchymal transition in breast cancer cells, *Oncol. Lett.*, 2021, **21**, 155.
- 141 N. Deng, M. Qiao, Y. Li, F. Liang, J. Li and Y. Liu, Anticancer effects of licochalcones: a review of the mechanisms, *Front. Pharmacol.*, 2023, **14**, 1074506.
- 142 S. Nagini, R. Nivetha, M. Palrasu and R. Mishra, Nimbolide, a neem limonoid, is a promising candidate for the anticancer drug arsenal, *J. Med. Chem.*, 2021, **64**, 3560–3577.
- 143 X. Zhao, L. Huang, W. Xu, X. Chen, Y. Shen, W. Zeng and X. Chen, Physapubescin B inhibits tumorigenesis and circumvents taxol resistance of ovarian cancer cells through STAT3 signaling, *Oncotarget*, 2017, **8**, 70130.
- 144 K. Chatterjee, S. Mukherjee, J. Vanmanen, P. Banerjee and J. E. Fata, Dietary polyphenols, resveratrol and pterostilbene exhibit antitumor activity on an HPV E6-positive cervical cancer model: an in vitro and in vivo analysis, *Front. Oncol.*, 2019, **9**, 352.
- 145 V. S. Sivasankarapillai, R. M. K. Nair, A. Rahdar, S. Bungau, D. C. Zaha, L. Aleya and D. M. Tit, Overview of the anticancer activity of withaferin A, an active constituent of the Indian ginseng *Withania somnifera*, *Environ. Sci. Pollut. Res.*, 2020, **27**, 26025–26035.
- 146 A. M. Tomko, E. G. Whynot, L. D. Ellis and D. J. Dupré, Anti-Cancer Potential of Cannabinoids, Terpenes, and Flavonoids Present in Cannabis, *Cancers*, 2020, **12**, 1985.
- 147 M. V. Sobral, A. L. Xavier, T. C. Lima and D. P. de Sousa, Antitumor activity of monoterpenes found in essential oils, *Sci. World J.*, 2014, **2014**, 953451.
- 148 D. Y. Kim and B. Y. Choi, Costunolide-A Bioactive Sesquiterpene Lactone with Diverse Therapeutic Potential, *Int. J. Mol. Sci.*, 2019, **20**, 2926.
- 149 G. Ş. Karatoprak, E. K. Akkol, Y. Genç, H. Bardakci, Ç. Yücel and E. Sobarzo-Sánchez, Combretastatins: An Overview of Structure, Probable Mechanisms of Action and Potential Applications, *Molecules*, 2020, **25**, 2560.
- 150 S. Ijaz, N. Akhtar, M. S. Khan, A. Hameed, M. Irfan, M. A. Arshad, S. Ali and M. Asrar, Plant derived anticancer agents: a green approach towards skin cancers, *Biomed. Pharmacother.*, 2018, **103**, 1643–1651.
- 151 K. J. Min and T. K. Kwon, Anticancer effects and molecular mechanisms of epigallocatechin-3-gallate, *Integr. Med. Res.*, 2014, **3**, 16–24.
- 152 A. S. Choudhari, P. C. Mandave, M. Deshpande, P. Ranjekar and O. Prakash, Phytochemicals in Cancer Treatment: From Preclinical Studies to Clinical Practice, *Front. Pharmacol.*, 2020, **10**, 1614.
- 153 J. Dou, Z. Wang, L. Ma, B. Peng, K. Mao, C. Li, M. Su, C. Zhou and G. Peng, Baicalein and baicalin inhibit colon cancer using two distinct fashions of apoptosis and senescence, *Oncotarget*, 2018, **9**, 20089–20102.
- 154 P. Kuppusamy, A. Nagalingam, N. Muniraj, N. K. Saxena and D. Sharma, Concomitant activation of ETS-like transcription factor-1 and death receptor-5 via extracellular signal-regulated kinase in withaferin A-mediated inhibition of hepatocarcinogenesis in mice, *Sci. Rep.*, 2017, **7**, 17943.
- 155 M. O. Kim, M. H. Lee, N. Oi, S. H. Kim, K. B. Bae, Z. Huang, D. J. Kim, K. Reddy, S. Y. Lee, S. J. Park, J. Y. Kim, H. Xie, J. K. Kundu, Z. Y. Ryoo, A. M. Bode, Y. J. Surh and Z. Dong, [6]-Shogaol inhibits growth and induces apoptosis of non-small cell lung cancer cells by directly regulating Akt1/2, *Carcinogenesis*, 2014, **35**, 683–691.
- 156 H. Chen, B. Zhu, L. Zhao, Y. Liu, F. Zhao, J. Feng, Y. Jin, J. Sun, R. Geng and Y. Wei, Allicin inhibits proliferation and invasion in vitro and in vivo via shp-1-mediated stat3 signaling in cholangiocarcinoma, *Cell. Physiol. Biochem.*, 2018, **47**, 641–653.



- 157 T. Wang, Y. Jiang, L. Chu, T. Wu and J. You, Alpinumisoflavone suppresses tumour growth and metastasis of clear-cell renal cell carcinoma, *Am. J. Cancer Res.*, 2017, **7**, 999–1015.
- 158 J. Li, C. Zhang, H. Jiang and J. Cheng, Andrographolide inhibits hypoxia-inducible factor-1 through phosphatidylinositol 3-kinase/AKT pathway and suppresses breast cancer growth, *OncoTargets Ther.*, 2015, **8**, 427–435.
- 159 Y. Tao, S. Zhan, Y. Wang, G. Zhou, H. Liang, X. Chen and H. Shen, Baicalin, the major component of traditional Chinese medicine *Scutellaria baicalensis* induces colon cancer cell apoptosis through inhibition of oncomiRNAs, *Sci. Rep.*, 2018, **8**, 14477.
- 160 G. Zhao, X. Han, S. Zheng, Z. Li, Y. Sha, J. Ni, Z. Sun, S. Qiao and Z. Song, Curcumin induces autophagy, inhibits proliferation and invasion by downregulating AKT/mTOR signaling pathway in human melanoma cells, *Oncol. Rep.*, 2016, **35**, 1065–1074.
- 161 W. Wu, S. N. Tang, Y. Zhang, M. Puppala, T. K. Cooper, C. Xing, C. Jiang and J. Lü, Prostate cancer xenograft inhibitory activity and pharmacokinetics of decursinol, a metabolite of angelica gigas pyranocoumarins, in mouse models, *Am. J. Chin. Med.*, 2017, **45**, 1773–1792.
- 162 W. Zhang, J. Su, H. Xu, S. Yu, Y. Liu, Y. Zhang, L. Sun, Y. Yue and X. Zhou, Dicumarol inhibits PDK1 and targets multiple malignant behaviors of ovarian cancer cells, *PLoS One*, 2017, **12**, e0179672.
- 163 Y. C. Hsiao, S. F. Peng, K. C. Lai, C. L. Liao, Y. P. Huang, C. C. Lin, M. L. Lin, K. C. Liu, C. C. Tsai, Y. S. Ma and J. G. Chung, Genistein induces apoptosis in vitro and has antitumor activity against human leukemia HL-60 cancer cell xenograft growth in vivo, *Environ. Toxicol.*, 2019, **34**, 443–456.
- 164 A. C. B. M. Matrine, A. M. Fuzer, A. B. Becceneri, J. A. da Silva, R. Tomasin, D. Denoyer, S. H. Kim, K. A. McIntyre, H. B. Pearson, B. Yeo, A. Nagpal, X. Ling, H. S. Selistre-de-Araújo, P. C. Vieira, M. R. Cominetti and N. Pouliot, [10]-Gingerol induces apoptosis and inhibits metastatic dissemination of triple negative breast cancer in vivo, *Oncotarget*, 2017, **8**, 72260–72271.
- 165 X. Wu, W. Wang, Y. Chen, X. Liu, J. Wang, X. Qin, D. Yuan, T. Yu, G. Chen, Y. Mi, J. Mou, J. Cui, A. Hu, Y. E and D. Pei, Glycyrrhizin Suppresses the Growth of Human NSCLC Cell Line HCC827 by Downregulating HMGB1 Level, *BioMed Res. Int.*, 2018, **2018**, 6916797.
- 166 M. Han, H. Gao, J. Xie, Y. P. Yuan, Q. Yuan, M. Q. Gao, K. L. Liu, X. H. Chen, Y. T. Han and Z. W. Han, Hispidulin induces ER stress-mediated apoptosis in human hepatocellular carcinoma cells in vitro and in vivo by activating AMPK signaling pathway, *Acta Pharmacol. Sin.*, 2019, **40**, 666–676.
- 167 A. S. Choudhari, P. C. Mandave, M. Deshpande, P. Ranjekar and O. Prakash, Phytochemicals in Cancer Treatment: From Preclinical Studies to Clinical Practice, *Front. Pharmacol.*, 2020, **10**, 1614.
- 168 W. J. Lu, G. J. Wu, R. J. Chen, C. C. Chang, L. M. Lien, C. C. Chiu, M. F. Tseng, L. T. Huang and K. H. Lin, Licochalcone A attenuates glioma cell growth in vitro and in vivo through cell cycle arrest, *Food Funct.*, 2018, **9**, 4500–4507.
- 169 R. Subramani, E. Gonzalez, A. Arumugam, S. Nandy, V. Gonzalez, J. Medel, F. Camacho, A. Ortega, S. Bonkougou, M. Narayan, A. K. Dwivedi and R. Lakshmanaswamy, Nimbolide inhibits pancreatic cancer growth and metastasis through ROS-mediated apoptosis and inhibition of epithelial-to-mesenchymal transition, *Sci. Rep.*, 2018, **6**, 19819.
- 170 L. Chen, G. Xia, F. Qiu, C. Wu, A. P. Denmon and X. Zi, Physapubescin selectively induces apoptosis in VHL-null renal cell carcinoma cells through down-regulation of HIF-2 $\alpha$  and inhibits tumor growth, *Sci. Rep.*, 2016, **6**, 32582.
- 171 W. Wen, G. Lowe, C. M. Roberts, J. Finlay, E. S. Han, C. A. Glackin and T. H. Dellinger, Pterostilbene, a natural phenolic compound, synergizes the antineoplastic effects of megestrol acetate in endometrial cancer, *Sci. Rep.*, 2017, **7**, 12754.
- 172 A. Bishayee and G. Sethi, Bioactive natural products in cancer prevention and therapy: progress and promise, in *Seminars in Cancer Biology*, Academic Press, 2016, pp. 1–3.
- 173 F. Majolo, L. K. Delwing, D. J. Marmitt, I. C. Bustamante-Filho and M. I. Goettert, Medicinal plants and bioactive natural compounds for cancer treatment: important advances for drug discovery, *Phytochem. Lett.*, 2019, **1**, 196–207.
- 174 S. Kumar, D. Kumar, A. Bhat and A. Kumar, Phytochemicals in Clinical Studies: Current Perspective, in *Functional Food and Human Health*, ed. V. Rani and U. Yadav, Springer Nature Singapore Pte Ltd, 2018, pp. 471–511.
- 175 J. Fleminger and B. Goldacre, Prevalence of clinical trial status discrepancies: a cross-sectional study of 10,492 trials registered on both clinicaltrials.gov and the european union clinical trials register, *PLoS One*, 2018, **13**, e0193088.
- 176 H. ur Rashid, S. Rasool, Y. Ali, K. Khan and M. A. U. Martines, Martines Anti-cancer potential of sophoridine and its derivatives: recent progress and future perspectives, *Bioorg. Chem.*, 2020, **99**, 103863.
- 177 S. Sabet, A. Rashidinejad, L. D. Melton and D. J. McGillivray, Recent advances to improve curcumin oral bioavailability, *Trends Food Sci. Technol.*, 2021, **110**, 253–266.
- 178 L. X. Zhang, C. X. Li, M. U. Kakar, M. S. Khan, P. F. Wu, R. M. Amir, D. F. Dai, M. Naveed, Q. Y. Li, M. Saeed and J. Q. Shen, Resveratrol (RV): a pharmacological review and call for further research, *Biomed. Pharmacother.*, 2021, **43**, 112164.
- 179 P. Bathaei, M. Imenshahidi and H. Hosseinzadeh, Effects of *Berberis vulgaris*, and its active constituent berberine on cytochrome P450: a review. Naunyn Schmiedebergs, *Arch. Pharmacol.*, 2024, **14**, 1–24.

