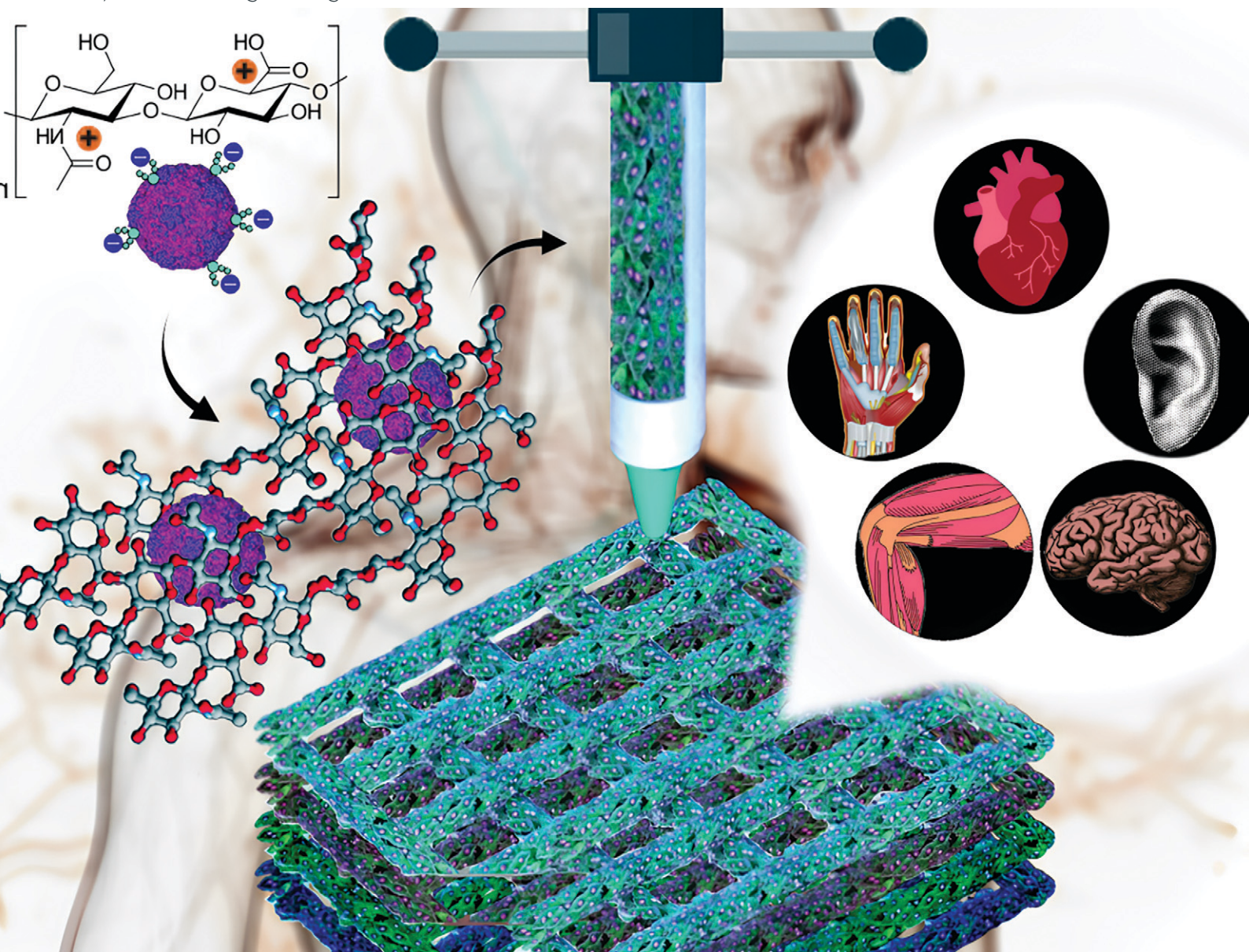


MSDE

Molecular Systems Design & Engineering

rsc.li/molecular-engineering



ISSN 2058-9689



Cite this: *Mol. Syst. Des. Eng.*, 2024, 9, 977

Recent advances in 3D bioprinting of polysaccharide-based bioinks for fabrication of bioengineered tissues

Kasula Nagaraja,^{†a} Pratik Dhokare,^{†a}
Amitava Bhattacharyya ^{abc} and Insup Noh ^{*ab}

Bioink in three-dimensional (3D) bioprinting of biomimetic tissue scaffolds has emerged as a key factor for the success of tissue engineering and regenerative medicine. The bioinks used for extrusion 3D bioprinting have hydrogel matrices with different kinds of polymeric biomaterials such as proteins, peptides, polysaccharides, hydrophilic synthetic polymers, and others. Natural polysaccharides such as alginate, chitosan, and hyaluronic acid have garnered significant attention as bioink materials due to their excellent biocompatibility, extracellular matrix mimetic properties, biodegradability, injectability, bioprintability and structural versatility among their many advantages, even though many research groups focus on the study of protein-based bioinks to utilize their high potential of cell adhesiveness. This review encompasses recent advancements of polysaccharide-based hydrogels and bioinks for bioengineered tissue regeneration and reconstruction, especially by focusing on fabrication of multilayered complex structures for biomimetic tissue engineering applications.

Received 1st January 2024,
Accepted 15th May 2024

DOI: 10.1039/d4me00001c

rsc.li/molecular-engineering

Design, System, Application

A bioprinting technology is a layer-by-layer 3D deposition of cell-laden hydrogel, *i.e.* a bioink, by extrusion-based printing technology to fabricate 3D constructs for its applications to regeneration of diseased tissues such as cartilage, bones, skin, skeletal muscle, and neural tissues. Even though this technology has attracted much attention in tissue engineering and regenerative medicine, the design and fabrication of tissue engineering scaffolds have yet to be solved by overcoming the difficulties of multilayered 3D scaffold construction in stable 3D structures. We here report the design of polysaccharide-based hydrogels, bioinks and 3D bioprinting scaffolds, targeting tissue engineering applications. For the 3D bioprinting and regeneration of diverse damaged tissues, the issues such as cell sources, biocompatibility, bioink and bioprinting properties, polymeric biomaterial selection, bioprintability performance, scaffold fabrication, and their applications to tissue regeneration have been described. Furthermore, the benefits, difficulties, and future directions of 3D bioprinting of polysaccharide-based bioinks for multilayered tissue engineering are discussed.

1. Introduction

Complex and multilayered tissues represent a fascinating frontier in the fields of biomaterials, 3D bioprinting, regenerative medicine, and biomedical engineering.^{1–4} These tissues are composed of various cell types and extracellular matrices (ECM) intricately arranged in three-dimensional molecular structures, mimicking the natural complexity of

human tissues and organs. 3D bioprinting allows the molecular, structural systems to be designed as a tailored scaffold construction of multilayered tissues, with the potential to revolutionize regenerative medicine. This cutting-edge technique leverages the precision of 3D bioprinting to create intricate, multilayered constructs that closely mimic the native architecture of human tissues and organs.^{5–7} By combining various natural biomaterials, including polysaccharides, with living cells and bioactive factors, 3D bioprinting allows the bioengineering of multilayered, functional and biocompatible tissue scaffolds. Within native tissues, diverse cell types like endothelial, epithelial, muscular, and adipose cells exist or co-exist in an intricate ECM in different tissues, intricately woven with interconnected networks.^{8,9} The goal of tissue engineering is to emulate this natural milieu, thereby fostering the creation and revitalization of fully functional tissues and organs

^a Department of Chemical and Biomolecular Engineering, Seoul National University of Science and Technology, Seoul 01811, Republic of Korea.

E-mail: insup@seoultech.ac.kr

^b Convergence Institute of Biomedical Engineering and Biomaterials, Seoul National University of Science and Technology, Seoul 01811, Republic of Korea

^c Medical Electronics Research Center, Seoul National University of Science and Technology, Seoul 01811, Republic of Korea

[†] Kasula Nagaraja and Pratik Dhokare are co-first authors with equal contribution and importance.

through designed biodegradable scaffolds. Recent technological advancements like bioprinting and advanced manufacturing have illuminated a promising trajectory for tissue engineering and reproductive medicine, profoundly shaping their future.^{10,11}

Polysaccharides serve a variety of purposes, such as structural properties, tissue regeneration,¹² antibacterial,¹³ antiviral,¹⁴ antitumor,^{15,16} and immune regulatory properties in tissue engineering.^{17,18} Hyaluronic acid and chitosan, derived from human connective tissue and arthropod exoskeletons respectively, along with alginic acid from brown algae, are key polysaccharides used in tissue engineering. These materials are processed into hydrogels and bioinks using various synthesis technologies. Hydrogels, known for their ability to retain significant amounts of water in a 3D polymeric network, differ from bioinks, which incorporate living cells within their polymer structure. Additionally, these materials can be enhanced with bioactive molecules like growth factors, nanoparticles, and drugs to tailor their properties for specific applications.¹⁹ The bioink is employed as a biomaterial for the fabrication of tissue engineering scaffolds. Thus, hydrogels are composed of highly hydrated 3D environments that resemble the ECM of tissues. Furthermore, because of their special architecture, which is porous and flexible, cells can migrate and communicate with each other, as well as receive nutrients, oxygen, and other water-soluble substances in a specific tissue.¹⁹ Different techniques can be used to design a hydrogel, based on the intended attributes and purpose in tissue engineering. Physically cross-linked hydrogels are created through molecular interactions.²⁰ The interactions can be influenced by changing the surrounding environment or applying mechanical forces. They are reversible and regenerated (associations break and continuously reform).²¹ For instance, ionic interactions with calcium ions (Ca^{2+}) can cross-link alginate to create a hydrophilic 3D network that resembles an egg-box model structure.²² Polymer chains joined by covalent bonds make up chemically cross-linked hydrogels.²³ There are two general methods for preparing these hydrogels, the first involves using a cross-linking agent to polymerize a hydrophilic monomer; the second method involves directly cross-linking water-soluble polymers.²⁴ Due to the strong covalent connections that form between polymeric chains, chemical crosslinking is typically more persistent than physical cross linking and has stronger mechanical qualities, making it appropriate for long-term use.²¹ Chemical hydrogels are formed by reacting polysaccharide functional groups ($-\text{COOH}$, $-\text{OH}$, $-\text{NH}_2$) with cross-linkers and other chemicals, ensuring that these groups network to create the gel. This process enhances the hydrogel's physical properties like viscosity, elasticity, and stability by forming a durable 3D structure with improved mechanical qualities.²⁵ These crosslinking induces formation of a hydrogel and makes layer-by-layer additive manufacturing of tissue engineering scaffolds by 3D bioprinting technology.

Proteins and their domain, *i.e.* peptides and oligopeptides, are essential for constructing hydrogel/bioink scaffolds in tissue engineering, thanks to their biocompatibility and ability to foster cell adhesion and then tissue regeneration. Proteins like collagen, fibronectin, gelatin, fibrin, and silk are crucial for their roles in enhancing cell interactions and scaffold functionality in tissue engineering.²⁶ Collagen, which is an abundant component in the ECM, supports cell adhesion and tissue regeneration, while its denatured form, gelatin, and its modified form (GelMA) have been employed as typical bioink hydrogels and allow for precise scaffold structures through photo cross-linking while maintaining biodegradability.²⁶ Fibrin is valued for its natural ability to form scaffolds that promote cell growth, especially in wound healing,²⁷ while silk fibroin's strength makes it suitable for structural repairs in bone and ligament contexts.²⁸ Incorporating peptides such as RGD, YIGSR, and IVAK have been employed as alternative biomaterials for proteins to prevent the issue of complex handling in synthesis by higher molecular weight. Peptides are the specific interaction domains of cell-adhesive proteins.²⁶ Even though this strategic application underscores their pivotal role in developing advanced, functional scaffolds for tissue engineering, they have limitations in their bioink applications, such as difficulty in gel synthesis compared to polysaccharide polymers.

Polysaccharides, with their diverse biophysical and chemical properties, are crucial in tissue engineering and 3D bioprinting. Classified as anionic, cationic, or neutral based on their natural charges, these polysaccharides such as alginate (ALG), chitosan, HA, gelatin (GEG), pectin, guar gum, and chondroitin sulfate are transformed into hydrogels through chemical and physical modifications. They are essential in various tissues within animal bodies, performing key biological, physiological, and structural functions influenced by their chemical composition, molecular weight, shape, and hydrophilicity.^{29,30} For example, sulfated marine polysaccharides like fucoidan and chondroitin sulfate have anti-inflammatory and antiviral properties, potentially blocking viruses from attaching to host cells. Chitosan, deacetylated from chitin, shows antibacterial effects by interfering with anions on bacterial cell walls, which can disrupt vital processes and kill the bacteria. Meanwhile, neutral dextran, due to its water solubility, resists protein adsorption.^{31,32}

The development of 3D bioprinting has been emphasized as a particular solution for creating unique 3D matrices (fabrication) similar to native tissues and organs, the regulated deposition of different biomaterials in layer-by-layer forms, and the shortcomings of the traditional approach. It is undoubtedly a groundbreaking method with several uses in tissue engineering, regenerative medicine, food industry, drug delivery and other bioengineering fields.^{33,34} 3D bioprinting is a biotechnology that uses biomacromolecules and biomedical engineering to create 3D structures that regulate cell functions, tissue regeneration

and disease treatment. It has applications in drug screening models, medical engineering, dentistry, surgery, and human-created structures like meniscus, vascular junctions, bone, tumor models, nerve and skin tissues. By harnessing the power of 3D bioprinting, researchers and medical professionals can advance healthcare and improve patient outcomes, making it a promising technology with significant implications in medical and biological fields.³⁵ However, there are no established standards or criteria for bioprinting or bioinks, which makes them challenging for researchers to determine which type of bioprinting or bioink they require for given tissue engineering applications.

Even while there are now a lot more commercially available bioprinting systems and bioinks than ever before, certain research organizations will probably still need to use unique setups and custom formulations of bioinks for specific tissues with different shapes and compositions.³⁶ Extrusion-based 3D bioprinting is one of the most frequently employed 3D bioprinting methods in tissue engineering applications, which deposits layers of tissue-specific cell-filled bioink on top of one another, indicating the importance of each layer of the bioprinted scaffolds. Natural polysaccharides have consistently attracted considerable attention in tissue engineering and extrusion-based bioprinting,^{37,38} and their hydrogels have gained broader applications in tissue engineering in recent decades. Notably, polysaccharides have diverse applications, for example, regeneration of encompassing bone and cartilage, muscle and cardiac tissue, neural tissue, skin, and other tissues. However, for the fabrication of large, multilayered complex structures through the techniques of polysaccharide-based hydrogel printing, the self-standing gel strength and post-printing shape fidelity are the issues of major concerns for the researchers working with such biomaterials in tissue engineering.^{39,40}

This review discusses the key properties of polysaccharides, their application in extrusion-based bioprinting technologies, and their role in tissue engineering. It delves into recent developments in polysaccharide-based hydrogels for creating multilayered scaffolds suitable for complex tissue structures. The review underscores the adaptability of these natural polymers in developing 3D bioprintable hydrogels and outlines their advantages and challenges in bioprinting. It also emphasizes the importance of bioink composition for effective 3D bioprinting and highlights extrusion-based bioprinting as a favorable method due to its versatility and reduced cell damage.

2. Characteristics of bioinks

Bioinks come in two categories: those that utilize scaffolds and those that do not. Scaffolds are fibrous or 3D structural biomaterials that permit not only transformation into tissues, but also biological liquids and gases to pass through. This is accomplished by making it simpler for cells to interact, deposit ECM and maintain viability with no or mild toxicity

and inflammation.⁴¹ The bioink should be printable and capable of maintaining cells without any cell damage, and also preserve the physio-mechanical and biological characteristics of the targeted tissue after bioprinting. The bioinks can be altered depending on the printing systems used to print on the target tissue for all the qualities to be achieved. Combining two or more nano-biomaterials is frequently necessary to improve the mechanical and biological properties, particularly for extrusion-based bioprinting.⁴² Multi-biomaterial bioinks are superior in bioprinting to those formed from a single biomaterial. Most single polymer-based bioinks lack the bioprintability, outstanding mechanical qualities, and functionality required to create tissues that mirror biological ones. The selection of polysaccharides is more constrained in 3D bioprinting because cells and post-printing fidelity are involved. It is important to consider mechanical qualities, biocompatibility, porosity, pH neutrality, hydrophilicity, and biodegradability while currently selecting the biomaterials for bioink.^{43,44} The majority of bioprinting techniques use additive manufacturing concepts by depositing tissue-specific cells and biomaterials collectively known as “bioinks” spatially and temporally in a regulated and sequential manner. Even though numerous bioinks have been created, no single bioprinter/bioink combination has proven effective in all tissue engineering applications to date, which is why the sector is still quite experimental. The following are some of the important characteristics that a bioink must have.

2.1 Cell source

When living cells are incorporated into a hydrogel, it becomes a measurable bioink. Since a necessary, key element of bioink is cells, the cell types and sources should be considered important factors. Since the chosen cell must continue to operate, reproduce successfully, imitate its physiological state, and regenerate the target tissues throughout the bioproduction process,⁴⁵ the right cell type, source, and density should be chosen carefully to create bioink and tissue engineering scaffolds. One of the most fundamental prerequisites for bioink's compatibility is that it must be robust and secure to support the loaded cells' survival and differentiation.⁴⁶

2.2 Hydrogel and its synthesis

A hydrogel, a 3D network of polymer, transforms into a bioink by adding cells into a hydrogel in 3D bioprinting and tissue engineering. It is synthesized by using diverse polymers such as HA, chitosan, gelatin, and chondroitin sulfate through the methods of chemical or physical crosslinking. The gelation process with cell incorporation should be easy to be handled, sterilizable and nontoxic because it can affect not only the cell viability but also the bioprinting processing and post-printing fidelity of the scaffolds.⁴⁷ Depending on how the composition and properties of the hydrogel and bioink are prepared, various

alternatives are available for the gelation procedure such as photo-crosslinking, ionic, hydrophobic, thermal, enzymatic, and stereo-complexing methods.

2.3 Biocompatibility

For the development of tissue engineering constructs by 3D bioprinting, cells in bioink should maintain their capacity to differentiate and proliferate *in vitro* without any damage. Biocompatibility and biodegradation were the primary concerns for the hydrogels and 3D printing.⁴⁸ Cutting edge researchers have expanded the concept of biocompatibility from the need for minimal cytotoxic effects in cell and biomaterial levels to a favorable biological relationship with the host, especially in tissue engineering and regenerative medicine area. Because they can simulate the natural cellular environment, the bioinks from natural biomaterials offer a nontoxic biomanufacturing approach, but they have to overcome the issues of poor mechanical qualities and immune response issues.⁴⁹

2.4 Bioprintability

Bioprintability refers to a biomaterial's ability to be printed in a controlled manner and shapes within a certain time frame. The idea behind 3D bioprinting is the layer-by-layer, controlled production of three-dimensional living objects using computer-aided design.⁵⁰ Several techniques, such as extrusion printing,⁵¹ inkjet printing,⁵² laser-assisted printing,⁵³ and stereolithography,⁵⁴ are used to bioprint cell-laden biomaterials, or bioinks. At present, 3D bioprinting technology has garnered significant interest and is being extensively researched for a variety of biomedical uses.⁵⁵ Rheology is essential to the extrusion-based 3D printing of hydrogels because it influences the behavior of the material during the printing process through its flow and deformation characteristics. Viscosity, shear thinning behavior, and viscoelasticity are examples of rheological qualities that are important in influencing the extrudability, shape retention, and general quality of printed structures.⁵⁶ Organ and tissue bioprinting relies on bioprinting of a gel with cells; however, the demanding bioprinting environment limits the bioink material options. Since the bioink's nature can substantially impact the bioprintability, the scaffold properties such as size accuracy and structural construction are all important considerations in 3D bioprinting and tissue engineering. Bioprinting technology varies based on its purposes; for example, micro-extrusion bioprinting requires adequate rheological properties and shear-thinning behaviors of bioinks, and preserves the preliminary shape fidelity of the post-bioprinting scaffolds.⁵⁷

2.5 Mechanical properties

Bioprinted hydrogels should have convincing mechanical qualities to withstand diverse biopressures throughout the bioprinting process and to sustain the scaffold structure after bioprinting for successful practical use as well as

differentiation during its biodegradation into specific tissue regeneration. At the same time, the bioink should guarantee the scaffold's mechanical qualities that can adapt to the target tissue's native environment. Due to their low mechanical properties, natural polysaccharide hydrogels do not provide good sacrificial nourishment during bioprinting and implant in the patient's body. Hence, synthetic polymers or nano-biomaterials are often used for higher mechanical qualities, keeping the bioink structurally and mechanically stable like genuine tissue, especially under mechanical stress like cartilage, cardiovascular tissues or bone.⁵⁸

3. Natural polysaccharides in bioinks

Polysaccharides and their derivatives are becoming more and more popular among the variety of biopolymers that can create hydrogels for 3D bioprinting applications, especially for the engineering and development of bioink formulations. Their primary characteristics—which include their vast diversity of chemical structures, ease of derivatization and functionalization, suitable mechanical and rheological qualities, and inherent biocompatibility and biodegradability—are what make them so appealing.⁵⁹ When it comes to bioprinting of the intricate and multilayered tissues, natural polysaccharides are essential building elements. Natural sources of polysaccharides as depicted in (Fig. 1) have the extraordinary capacity to produce hydrogels, which offer an environment that is favorable for cell encapsulation and growth as well as tissue regeneration. These polysaccharides function as the structural backbone of bioinks, enabling precise multilayer deposition during bioprinting procedures.²⁹ This integration gives the printed structures both mechanical stability and tissue engineering potential, by simulating the intricacy of original tissues in physical and biological aspects, presenting a possible path toward the creation of complex and useful biological constructs. The appropriate utilization of natural polysaccharides in bioprinting has enormous promise to transform regenerative medicine and advance the creation of realistic, multilayered tissues as researchers explore the frontier of tissue engineering.⁶⁰

3.1 Alginate or sodium salt of alginic acid (ALG)

One of the most common products made from brown algae is an alginate, ALG, with anionic, edible and hydrophilic properties. Due to its extremely low cost and biocompatibility, ALG is widely employed in biomedical applications.⁶¹ While ALG's low bioactivity also prevents the growth and proliferation of encapsulated cells, its stability is obtained by oxidation processing.^{62,63} Ca^{2+} and other divalent and trivalent cations are commonly used in the ionic gelation process to create ALG hydrogels in a stable state.⁶⁴ The drug release rate and the stability of the gels are modulated by control of the increase in ionic cross-linking.⁶⁵ ALG hydrogels' gel formation, mucoadhesive ability, biodegradability and nontoxicity, among other qualities, have

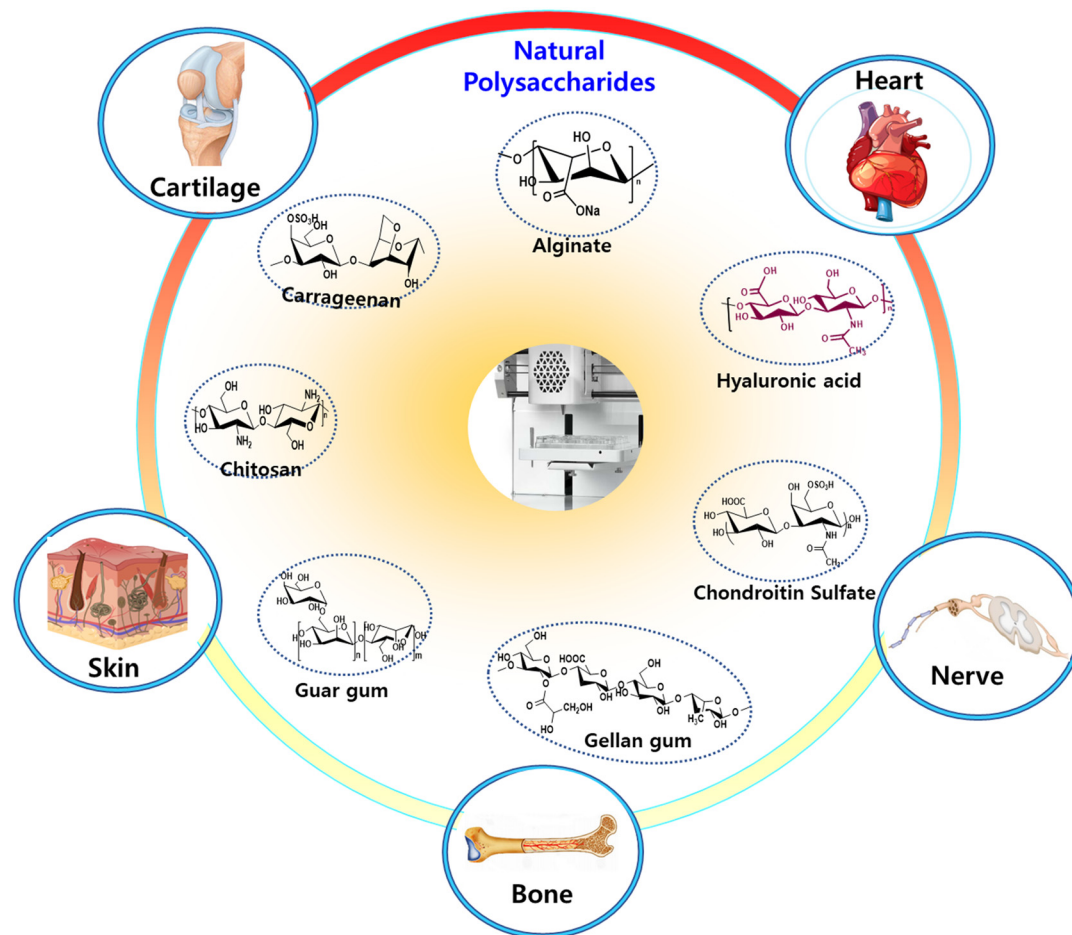


Fig. 1 Examples of natural polysaccharides used for 3D bioprinting tissue engineering.

encouraged the use of these biomaterials in multilayer 3D bioprinting, tissue engineering and drug delivery applications including wound healing, bioinks for 3D printing, *in vitro* models, or the delivery of anticancer medications.⁶⁶

In order to create hydrogels with potential uses in various multilayer 3D bioprinting and tissue engineering techniques, ALG has also been mixed with other polymers, in order to create skin tissue analogues using extrusion bioprinting; Somasekharan *et al.* reported a bioink based on alginate mixed with gelatin and diethylaminoethyl cellulose (DCEL). Because the gelatin backbone naturally contains RGD peptide sequences, the inclusion of gelatin boosted cell adherence, while the fibrous character of DCEL gave matrix stability and improved mechanical qualities. When compared to skin tissue, the bioprinted scaffolds had appropriate mechanical properties, such as elasticity, with an elongation of break of $91.7 \pm 9.36\%$ and a Young's modulus of 125 ± 22 kPa. For 21 days, the co-culturing of fibroblasts and keratinocytes was seen, along with the progression of cell differentiation within the scaffolds. Histological examination at this stage revealed the development of comparable dermal and epidermal components.⁶⁷

Dogan *et al.* went one step further in a recent study, creating bioprinted scaffolds using an alginate–collagen I

bioink loaded with mesodermal progenitor cells (hiMPCs) derived from human-induced pluripotent stem cells that were cultured with the addition of vascular endothelial growth factor (VEGF) to induce blood vessel formation. Following a 21 day incubation period, the bioprinted scaffolds showed signs of both small and big vessel creation. To evaluate their functionalities, the scaffolds were subsequently inserted into a chicken embryo's chorioallantoic membrane (CAM). The CAM model's appropriate blood perfusion was made possible by the printed vessels.⁶⁸

3.2 Chitosan

Chitosan is a linear polysaccharide, made up of randomly distributed-(1 → 4)-linked D-glucosamine (deacetylated unit molecules from chitin).⁶⁹ Chitosan's motivating properties are its low renewability, lack of toxicity, lower cost, biocompatibility, bio-adhesiveness, anti-bacterial activity, and biodegradability. Chitosan degrades more slowly than animal-derived polymers like collagen and fibrin^{70,71} and has several functional groups, including amine, amino, and hydroxyl groups, which can interact with both cationic and anionic molecules. This is particularly significant in the case of proteins. With the addition of various groups along the

polymer chain,⁷² the functionalization of the polysaccharide was possible, opening up a variety of applications for it, including hydrogel, bioink, 3D bioprinting, tissue engineering⁷³ and drug administration.⁷⁴

Chitosan hydrogels have been used for diverse purposes such as the repair of bone and cartilage, and other tissues. As examples, polyelectrolyte hydrogels can be produced in a simple way by combining chitosan with polysaccharides or proteins with opposed charges.⁷⁵ In recent years, further applications for chitosan hydrogels have included heart regeneration and wound healing.^{76,77} Tonda-Turo *et al.* created a bioink based on chitosan that combines photocrosslinking and thermally induced gelation in a dual crosslinking mechanism. To do this, β -glycerol phosphate salt and chitosan-methacrylate were combined to provide thermosensitive behavior, having an elastic modulus of about 6 kPa. After 48 hours, cell proliferation was observed from the extrusion bioprinting of these hydrogels loaded with NIH 3T3 cells, resulting in 3D constructs with good cell dispersion even after 24 hours.⁷⁸ The simultaneous inclusion of additional ingredients or nanostructures in the bioinks is another popular tactic used to boost the effectiveness of chitosan-based bioinks. For example, the development of a bioink based on chitosan and poly(γ -glutamic acid) γ PGA is described in the work of Pisani *et al.* The amine groups in chitosan and the carboxylic acid groups in γ PGA interacted ionically to produce the hydrogel. Human dermal fibroblasts were utilized by the authors to 3D bioprint grid-shaped constructs using this hydrogel. Cell viabilities of about 70% were seen 24 hours after the bioprinting procedure and maintained for 14 days in culture.⁷⁹

3.3 Hyaluronic acid (HA)

HA is available in all joints, neural, connective, and epithelial tissues in animals. The extracellular membrane of mammalian tissue (muscle), present in some organs including the central nervous system, contains HA with a repeating unit of *N*-acetyl-D-glucosamine and D-glucuronic acid in its chemical structure.⁸⁰ The capacity of HA to maintain moisture levels makes it a perfect substance for promoting wound healing. Depending its molecular weight and water solubility, HA has diverse properties of bioprinting, mechanical strength and stability.^{81,82}

By adding gelling agents, or chemical modifications, HA hydrogels can be created.⁸³ Combining HA with gelling agents, such as poloxamer,⁸⁴ ALG,^{2,85} collagen,⁸⁶ methylcellulose (MC),⁸⁷ or gelatin,⁶ is one of the frequently used techniques for producing physically cross-linked HA hydrogels. Cross-linking substances like divinyl sulfone or glutaraldehyde can be added to HA polymers to create crosslinked HA hydrogels.⁸⁸ However, the usage of cross-linking agents may be constrained since they might be hazardous or produce toxic byproducts to tissue engineering applications. For the creation of HA hydrogels, more

biocompatible chemical modification techniques including enzymatic cross-linking, click chemistry, or physical crosslinking have been investigated⁸⁹ such as oligonucleotides. For these new options to work, HA must be functionalized with additional chemical groups, which can be added to the carboxylate, hydroxyl, or amide groups.⁹⁰

Methacrylate groups can be chemically crosslinked with HA to generate HA hydrogels, a method that is frequently employed with various biopolymers. These platforms have been used to release PLs and GF. Further, even MSCs have been cultivated in these matrices to stimulate their differentiation for uses in periodontal regeneration,⁹¹ cartilage regeneration,⁹² bone regeneration, or skin regeneration.⁹³ A HA-based bioink made of HA, hydroxyethyl acrylate, and gelatin-methacryloyl was reported by Noh *et al.* They demonstrated that the HA-based bioink did not adversely affect the viability of embedded bone cells when used to manufacture multilayered construction. The hydrogel consisting of three components was biocompatible, and the gel printing procedure was good.⁸¹

3.4 Cellulose

Due to its rigid structure, cellulose, one of the most prevalent polymers in plant cell walls, contributes to its construction. It consists of a repeating unit of (1 $\rightarrow$ 4) connected-D-glucopyranosyl. As a renewable polysaccharide, cellulose offers intriguing properties due to its adaptability and sustainability.^{94,95} Natural cellulose possesses specific chain length, crystallinity, and intra-intermolecular hydrogen bonding, among other physicochemical properties. The great thickness of free OH groups in cellulose allows for hydrogen bonding, but they also contribute to the insoluble nature of cellulose.^{96,97}

Given that cellulose is not soluble in water, it makes sense to alter its structure to develop soluble derivatives with a high affinity for water, which can then be used to make hydrogels. Methylcellulose (MC), one of them, has been used in the creation of several hydrogels for their multilayered bioprinting.⁹⁸ For instance, Boonlai *et al.* combined CNCs, methylcellulose, and κ -carrageenan, and then used ionic crosslinking (with aqueous KCl) to make an appropriate hydrogel for 3D extrusion-based bioprinting of live constructions for applications such as bone tissue creation and regeneration. Better shear-thinning behavior resulted from adding 2 and 4 wt% of CNCs to the hydrogels because nanocellulose enhanced the bioink's rheological characteristics. Furthermore, while increasing the CNC content from 2 to 4 wt%, the compressive stress at 30% strain increased in the CNC-reinforced hydrogels from 20.03 $\pm$ 0.02 to 23.28 $\pm$ 0.01 kPa. The vitality of L929-loaded printed constructs was greater than 90% five days after bioprinting, demonstrating that the bioprinting procedure and the prepared biomaterials did not damage these cells.⁹⁹

3.5 Dextran

Dextran polymer is a naturally occurring, non-toxic, hydrophilic homopolysaccharide containing α -(1 $\rightarrow$ 6)-linked-D-glucopyranose. Dextran comprises solely hydroxyl(-OH) groups, which do not contribute to cellular adhesion,^{100,101} but its hydroxyl groups are simple to functionalize, making hydrogel preparation possible. DEX hydrogels can be created using physical or chemical cross-linking, including photo-crosslinking,¹⁰² Michael addition,¹⁰³ Schiff-base reaction,¹⁰⁴ and enzymatic cross-linking. Physically cross-linked hydrogels typically have lower mechanical strengths than chemically cross-linked ones, but because they may be made at softer circumstances, there is less risk of harm to the included biomacromolecules.

Actually, utilizing a promising core/shell extrusion-based 3D bioprinting technology, only one study to date has described the bioprinting of a vascularized construct for wound care employing oxidized dextran.^{69,105} This was accomplished by using a hydrogel consisting of periodate-oxidized dextran and peptide-functionalized succinylated chitosan (C) as the shell and GelMA as the core. In this experiment, two cell types were used: BMSC in the shell and HUVECs in the core. With HUVEC-specific markers visible 21 days after bioprinting, the 3D bioprinted peptide-CD/GelMA constructs offered a suitable milieu for cell growth and development, indicating the presence of endothelial cells within the tube-like structures. Although this work is the only one on the usage of dextran-based bioinks to date, it provides a way to employ this polysaccharide to create vascularized structures, which addresses one of the primary obstacles in 3D bioprinting live tissues.¹⁰⁶

3.6 Gellan gum

An anionic polysaccharide with a linear chemical structure is GEG. As a result of the fermentation process, it is secreted by the bacteria *P. elodea*. L-rhamnose, two D-glucose subunit chains, and repeated chain units of D-glucuronic acid are all present in gellan gum. It has effective mechanical and gelling capabilities and exhibits cytocompatibility, biocompatibility, and biodegradation.^{107,108} Gellan gum is used in tissue engineering to bio print skeletal cartilage, tissue, and structures that resemble the human brain.¹⁰⁹ In a different direction, gellan gum and pure starch were mixed to create 3D-printed scaffolds with varied printing gaps for Schwann cell seeding. The printed constructs were shown to be stable, to have sufficient swelling ratios, and not to be harmful to the L929 fibroblast cell line. Because these methods work well for 3D bioprinting, the usage of gellan gum is expected to be increased in the formulation of cell-laden bioinks in the future.¹¹⁰

3.7 Agarose

A repeating chain like arrangement of D-galactose and 3,6-anhydro-L-galactose units makes up the agarose polymer. Along with agarpectin, agarose is a naturally occurring

linear polysaccharide composed of D-galactose and 3,6-anhydro-L-galactopyranose units.¹¹¹

Because of its thermo-responsive reversible gelling, agarose polysaccharide, generated from red seaweed, is frequently used in the tissue engineering sector.^{112,113} Instead of using bioink components, alternative uses for agarose hydrogels in bioprinting procedures have also been investigated.¹¹⁴ Agarose hydrogels are easily molded and used to cast sub-millimetric geometries at low concentrations (less than 1% (w/v)).¹¹⁴ Utilizing this characteristic, Aydin *et al.* created a bioink using alginate and agarose that was created using a microwave-assisted technique. The agarose acted as a self-eroding sacrificial component to cast tubular structures within the alginate that was packed with cells, creating living printed constructs with a vascularized network. In particular, this sacrificial bioink made it possible to bioprint a 2 cm tubular structure in 2 minutes, preserving its shape and enabling very high cell survival (up to 95%) in a 3 day culture of MSCs generated from human adipose tissue (AT MSCs).¹¹⁵

3.8 Konjac gum

A water-soluble hydrophilic polymer derived from konjac tubers is called konjac glucomannan (KJG). China, Japan, and Southeast Asia have traditionally used KJG as a food source and traditional medicine. The α -(1 $\rightarrow$ 4)-pyranoside bond between D-glucose and D-mannose polymerizes the main structural chain of KJG, with a decreased number of acetyl groups at C-6 locations in the side chain unit.¹¹⁶ KJG is a desirable biopolymer because of its notable properties, which include biodegradability, biocompatibility, the ability to form films, and the capacity to gel.¹¹⁷ Alves *et al.* presented a KJG-xanthan gum hydrogel for wound dressing application. The dressing was then assessed by physicochemical and biological experiments.¹¹⁶ The use of a KJG hydrogel in the treatment of acute wounds was investigated by Yang *et al.* after its synthesis by adjusting the KJG concentration during the preparation process. The hydrogel was demonstrated to have antibacterial activity, biocompatibility, and water-holding capacity.¹¹⁸

3.9 Guar gum(GG)

GG is a plant-based polysaccharide derived from *Cyamopsis tetragonolobus* and has a high molecular weight. It is made up of α -(1 $\rightarrow$ 6), connected galactose side chain units and β -(1 $\rightarrow$ 4)-D-mannan linear chains.¹⁰⁸ An affordable hydrophilic polyhydroxy polysaccharide is guar gum. Due to its strong biocompatibility, biodegradability, and outstanding rheological qualities, it is frequently utilized in multilayered and complex tissue engineering application.^{119,120} Indurkar *et al.* presented a novel, cost-effective bioink made of gelatin and varying guar gum concentrations. This novel biopolymer combination was created to provide bloom strengths, G' and G'' , that range widely, and then $\tan \delta$ was computed using G' and G'' , and then the bloom test was conducted. Following the study of filament creation, these physical parameters

were associated with the printability of the bioink. The guar gum–gelatin bioinks with bloom strength between 480 and 750 and $\tan \delta$ between 0.15 and 0.2 were reported to be printable because they could form filaments during extrusion.¹²⁰

3.10 Xanthan gum (XTG)

XTG is an anionic polysaccharide generated from *Xanthomonas* bacteria that produces a viscous solution (high molecular weight) and has a low shearing rate. The side chains of XTG are composed of a trisaccharide made up of D-mannose beta (β)-(1 $\rightarrow$ 4), D-glucuronic acid beta(β)-(1 $\rightarrow$ 2), and D-mannose, which are linked to substitute residues of glucose groups by alpha(α)-(1 $\rightarrow$ 3)-linkage in the backbone. XTG is a branched polysaccharide.¹²¹ The remarkable qualities of XTG, which include exceptional physicochemical properties, stability over a wide pH range and temperature range, high viscosity at low concentration, high degree of pseudoplastic behavior, biodegradability, and non-toxicity, are suitable for multilayered and complex tissue engineering application.¹²²

Since xanthan gum has outstanding rheological properties, it is often used to enhance the mechanical and rheological properties of bioinks.^{123,124} To be more precise, Lim *et al.* used xanthan gum's ability to thin shear and mixed it with CMC as well as with alginate, GelMA, and hMSCs to create a suitable bioink for extrusion-based bioprinting.¹²⁵

4. Polysaccharide-based hydrogels in complex/multilayered tissue engineering applications

Polysaccharide-based hydrogels are increasingly used in tissue engineering for regenerating various tissue types, including soft, hard, connective, and functional tissues. These hydrogels are formulated into bioinks that encapsulate cells and bioactive molecules, offering properties such as biocompatibility, appropriate rheology, and elasticity, crucial for bioprinting of complex or multilayered scaffolds like those needed for cartilage, bone, skin and nerves. However, single-polymer hydrogels often fall short of meeting the complex demands of multilayered tissues. To address this, combinations of multiple bioactive polymers, nanobiomaterials, and crosslinking techniques are used to enhance the mechanical, biological, and post-printing properties of the constructs, ensuring effective regeneration of multilayered complex tissues. Further control of the interactions between layers is a crucial requirement to achieve multi-layer post-printing stability and integration.

Table 1 summarizes polysaccharide-based hydrogels used for 3D bioprinting of multilayered complex structures. To successfully regenerate specific functional tissues, cellular level bioengineering is essential during and after 3D bioprinting. The details of each tissue reconstruction using

polysaccharide-based bioinks are discussed in the subsections below.

4.1 Cartilage

Collagen fibers, matrix, and chondrocytes make up cartilage tissue.¹⁴¹ Cartilage is devoid of lymphatic and blood vessels. From the perichondrium's blood arteries, nutrients enter the intercellular matrix and feed the osteocytes.¹⁴² This histological trait significantly restricts the injured cartilage's ability to regenerate itself. Thus, physicians and researchers have been baffled by the damage healing challenge.¹⁴³ Combining cartilage scaffolds with 3D bioprinting technology offers a potent strategy to address and revitalize cartilage imperfections. These scaffolds are central in fostering essential cellular processes such as adhesion, differentiation, proliferation, and migration of the cells in the 3D bioprinted scaffolds. Moreover, they actively promote the regeneration of new tissue after implantation.^{144,145}

3D printing of bone and cartilage has utilized ALG, a biocompatible and organic polymer, for bioink synthesis. Although progress has been made in 3D bioprinting of ALG for orthopedic applications, issues such as poor bioink component distributions and mechanical properties, lack of long-term stability, the absence of functional moieties for cell adhesion and proliferation, *etc.* remained as problems to be solved.¹⁴⁶ Hence, it is mixed with other proteins, polypeptides, and bioactive molecules to strengthen its structure and improve its biological activities. The uniform biomixing of multiple components present in today's advanced bioink is crucial for the controlled and homogeneous tissue regeneration. In a recent study, Bhattacharyya *et al.* demonstrated a screw extruder head with an ionic gel (ALG), alpha-tricalcium phosphate (α -TCP) micro/nanoparticles, and osteoblast cells. 10 layers were 3D bioprinted using the bioink samples with a screw extruder which showed better bioprintability, with better mechanical and biological properties than the conventional mixing ones. The live cell distribution in the printed constructs was significantly superior to conventional mixing even with continuous feeding and extrusion-based bioink printing. The extrusion head ensured the control of uniform micro/nanomaterials and cell distribution throughout the directly mixed printable bioink with minimal cell damage as shown in Fig. 2(a and b).¹⁴⁷

Govindharaj *et al.* developed a bioink hydrogel combining dECM with ALG and *Phallusia nigra* supports. When 3D bioprinted using extrusion with human mesenchymal stem cells (hMSCs), the hydrogel exhibited high cell viability, substantially increasing hMSC proliferation and promoting chondrogenesis for cartilage repair applications.¹⁴⁹ Chae *et al.* developed a 3D meniscus graft using mesenchymal stem cells and ECM. The implants showed good biocompatibility and strong mechanical properties in a rabbit meniscectomy model. They also stimulated the development

Table 1 Polysaccharide-based extrusion 3D bioprinting of multilayered complex structures for tissue engineering applications

S. No	Polysaccharide	Preparation of bioink	Cell type	TE application	Features/limitations	Ref.
1	HAMA-CSMA-gel	UV-crosslink	SMSCs	Cartilage	1. Easy extrusion and stable 15-layer scaffolds w/ fidelity 2. Significant elastic and compressive strengths; robust after cross-linking 3. <i>In vitro</i> proliferation and differentiation of SMSCs 4. <i>In vivo</i> cartilage regeneration and functional recovery	126
2	HA-ALG	Ionic Ca^{+2}	ADSCs	Cartilage	1. Double cross-linking w/ biotinylated HA and streptavidin for printability and structural integrity 2. Strengthened w/ Ca^{2+} for mechanical stability & durability 3. High precision in printing both square & round shape 4. Chondrogenic differentiation for cartilage tissue engineering	127
3	GEG/lignin	Magnesium (Mg^{+2})	hMSCs	Cartilage	1. Elation properties & lignin's shear-thinning behavior 2. High stability mimicking the cartilage environment 3. Chondrogenic differentiation	128
4	GEG/gel	Ionic Ca^{+2}	MSCs	Cartilage	1. Rheological properties, shear recovery, and shape fidelity 2. The 30-layer scaffold w/ mechanical strength 3. Initial cell viability in scaffolds is over 80%	129
5	ALG/gel	Ionic Ca^{+2}	MSCs	Bone	1. Stiffness controlled by alginate concentrations 2. Matrix mineralization and cellular organization 3. Osteogenic development effectively at high cell densities 4. Softer scaffolds excel in matrix deposition and cellular activity	130
6	GEG	UV-crosslink	MC3T3-E1	Bone	1. Good shear-thinning properties 2. UV cross-linking post-printing significantly w/ mechanical stability 3. High shape accuracy and structural integrity in complex structures	131
7	ALG	UV-crosslink	HASCs	Bone	1. Osteogenic potential of bioink by bone-derived ECM 2. Modified dECM enhances the mechanical properties of the bioink 3. Printing of complex, cell-laden structures w/ precise multi-layered, micro-sized struts 4. Osteogenic development w/ high <i>in situ</i> cell viability (>90%)	132
8	GEG MA/GelMA	Ionic Ca^{+2} and UV-crosslink	BMSCs	Bone	1. Structural stability of scaffolds w/ double cross-linking 2. Post-printing shape integrity 3. Diverse printing across a broad temperature range 4. Angiogenic and osteogenic properties	133
9	XTG/ALG	Ionic Ca^{+2} (electrostatics interaction)	CCD-986	Skin	1. Viscoelastic properties, printability & structural stability w/ catechol-functionalization 2. Compressive and tensile strengths of the gel 3. Catechol functionalization w/ the biocompatibility	134
10	ALG/gelatin	Ionic Ca^{+2}	PBMCs	Skin	1. Gel stability and offers strong antibacterial properties w/ gallium 2. Bioink's reproducibility and scalability 3. High-resolution & free printing of refined structures	135
11	Pectin methacrylate	Ionic Ca^{+2}	Human neonatal dermal fibroblast	Skin	1. Tunable mechanical properties w/ UV cross-linking & ionic gelation 2. Shear-thinning behavior in extrusion printing 3. Mechanical strength for dermal tissue support	136
12	ALG/DCEL/gel	Ionic Ca^{+2}	hFBs	Skin	1. Enhanced mechanical integrity and stacking 2. Printing various shapes durable for manual handling 3. Supports growth and function of skin cells	67
13	ALG/gel	Ionic Ca^{+2}	Schwann cells	Nerve	1. Post-printing stability 2. High viability rate of 91.87% in the scaffold 3. Enhanced release of neurotrophic factors (NGF, BDNF, GDNF <i>etc.</i>) 4. Positive immunofluorescence after 4 weeks	137
14	HA acid/ALG	Ionic Ca^{+2}	Schwann cells	Nerve	1. Multilayer scaffolds w/ varied geometries 2. Balance stabilization speed & flowability for effective bioprinting	138

Table 1 (continued)

S. No	Polysaccharide	Preparation of bioink	Cell type	TE application	Features/limitations	Ref.
15	ALG/BNC/GelMA	Ionic Ca^{+2}	Schwann cells	Nerve	3. High Schwann cell viability and functional activity 1. BNC addition improves mechanical & rheological properties 2. Post-printing shape fidelity & line thickness 3. Oriented growth and high viability of Schwann cells	139
16	HA acid/ALG/gel	Ionic Ca^{+2}	Schwann cells	Nerve	1. Curable <i>via</i> iron(III) chelation or photocuring, adjusting mechanical properties & degradation rates 2. Intricate tissue architecture printing w/ post-printing stability 3. High cell viability in Schwann cells, neuronal cells, and glioma cell-laden constructs	140
17	HA/gelatin	Genipin	MC3T3		1. Gelatin concentration affects hydrogel's viscosity and shear-thinning, improving extrusion 2. Printed complex, multi-layer (50 and 100 layers) structures 3. Robust cell growth & high cell viability	6
18	ALG/HA	Ionic Ca^{+2}	MC3T3		1. Good printability for complex structures w/ stable viscosity 2. 100 layers printing w/ post-printing integrity under stress by Ca^{2+} 3. Cell survival and controlled release of medicinal substances	2

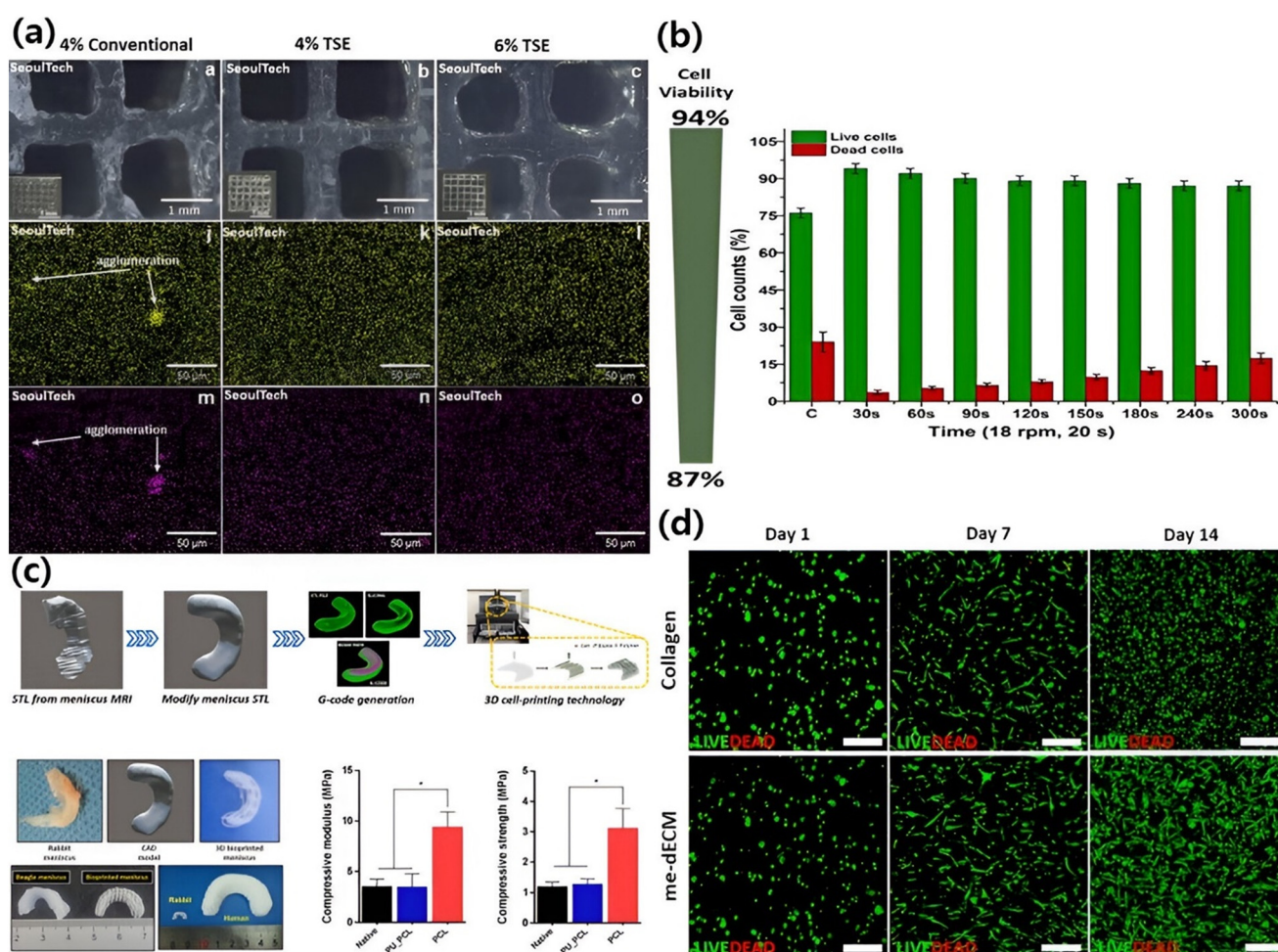


Fig. 2 (a) 3D biprinted (top view) micro/nanocomposite ALG gels showing improved dispersion of α -TCP particles in the bioprinting pen process; (b) cell viability of ALG gels (permission from Elsevier¹⁴⁷); (c) bioink from the meniscus was used to 3D print living, functional meniscus grafts; (d) live/dead staining (permission from Elsevier¹⁴⁸).

of neo-fibro cartilage *in vivo* due to a meniscus-specific environment. This approach, combining cells, biomaterials, and bioactive proteins, holds promise for promoting regenerative healing of complex cartilage tissues (as depicted in Fig. 2(c) and (d)).¹⁴⁸ Sadeghianmaryan *et al.* enhanced 3D bioprinting using a chitosan-based bioink with optimized crosslinking methods. The biocompatibility of the chitosan-printed bioink was validated through ATDC5 cell testing. This bioink holds promise for applications in cartilage regeneration and joint repair due to the articular cartilage's role as a shock absorber and lubricant in the joints.¹⁵⁰

To increase the strength of the chitosan hydrogel, He *et al.* first treated the chitosan with calcium and ethylene diamine tetraacetic acid (EDTA), and then 3D printed a scaffold. The bioink has been altered to exhibit enhanced mechanical characteristics and to encourage the growth of chondrocytes and the expression of genes related to cartilage.¹⁵¹ One derivative of chitosan with outstanding injectability and biocompatibility is hydroxybutyl-chitosan (HBC). Liu *et al.* synthesized the HBC-nanofiber (NF) hydrogel by combining HBC hydrogel with poly(lactic acid-co-glycolic acid) copolymer nanofiber. The use of nanofibers considerably improved the HBC-NF hydrogel's capacity to encourage cartilage development. The HBC-NF hydrogel was then injected into a 3D printed PCL skeleton that had internal microchannels to produce a mechanical concentration scaffold resembling a real cartilage.¹⁵² Antich *et al.* printed 3D scaffolds for articular cartilage by combining HA and alginate. The novel biological ink that encourages the regeneration of articular cartilage and has appropriate mechanical qualities is made of HA and alginate.¹⁵³ Angelis *et al.* printed 3D CH/HAA scaffolds using chitosan (CH) and hydroxyapatite (HAA) as raw materials. During a protracted culturing period, the printed scaffold's adequate porosity offers ideal conditions for chondrocyte colonization and synthesis. A unique idea for articular cartilage tissue engineering is provided by the scaffold cultured in platelet lysate (PL), which can better retain the phenotype of chondrocytes and enhance gene expression of the most relevant ECM components, utilizing the fact that HAA encourages cartilage regeneration.¹⁵⁴

Olate-Moya *et al.* mixed alginate, gelatin, and chondroitin sulfate (CS). To enhance the shape fidelity and resolution of the 3D printing scaffold, graphene oxide was further added to the composite hydrogel as a nanofiller material. A $30 \times 30 \times 1 \text{ mm}^3$ (length $\times$ width $\times$ height) scaffold was designed. An interior pattern resembling a mesh was established, with a thread spacing of 1.5 mm. Human adipose tissue mesenchymal stem cells were used to demonstrate that the novel nanocomposite hydrogel had a higher rate of cell proliferation than pure sodium alginate. Additionally, the scaffold made with the innovative hydrogel nanocomposite shows superior cartilage induction.¹⁵⁵

4.2 Bone

Bone is a sophisticated composition comprising hierarchical tissue, mineralized collagen fibers, and a network of blood vessels. Remarkably, bones can autonomously mend minor cracks and fractures due to their strong regenerative capacity. However, when a bone injury exceeds the size of two centimeters, its natural healing mechanisms become insufficient. Restoring both form and function necessitates tailored grafts with tissue regeneration capacity. Conventional hydrogels lack precise control for internal structure and growth factor distribution.

Innovative 3D bioprinting methods have been proposed to get over these constraints.^{156,157} In a recent advancement, Curti *et al.* created a scaffold-based 3D bioprint bioink from fish gelatin (FG), cuttlebone (CB), and ALG. It was accomplished by bonding the bioink after it had been produced. Conical nozzles with an inner diameter of 0.25 mm (G25) were employed; eight layers with a deposition direction ranging from 0 to 90°, and a layer thickness equal to 75% of the nozzle diameter. The composite FG-CB-ALG bio-ink hydrogel encapsulated to pre-osteoclast cells MC3T3-E1 and most bone ECMs were replicated using marine materials. The *in vitro* biocompatibility assay revealed that the composites could support MC3T3-E1 murine pre-osteoblast growth and proliferation and that the hydrogel may boost the bone regeneration potential of the 3D bioprint scaffold.¹⁵⁸ Seunghyun *et al.* introduced PDANPs for their notable affinity to bind with calcium ions, ALG, and temporally oxidized cellulose nanofibrils, all vital components within the bioink designed for bone tissue construction. Once integrated into each bioink, osteoblasts were extruded using 3D printing technology, and hydrogels were printed as a grid structure of 6 layers, followed by a curing period lasting up to 7 days. Notably, most cells in each sample remained viable. The printed cells' viability rate (% of living cells) surpassed 80% for all three models on both days 1 and 7.¹⁵⁹

For the objective of bone tissue engineering, a unique MeGC-based cell-laden 3D bioink was created. For creating biocompatible and structurally stable cell structures, the bioink solutions were tuned. MeGC bioink was created by photocuring the precursor solution for 70 seconds. The produced five-layer 3D bioprinting constructions showed increased cell proliferation, distinct bone differentiation, and good shape fidelity.¹⁶⁰

Wu *et al.* introduced a dual-network hydrogel approach, by combining the shear-thinning and recovery properties of GEG with the rapid photo crosslinking capabilities of poly(ethylene glycol)-diacrylate (PEGDA). The bioprinted multilayered scaffold provided an open and interconnected network (Fig. 3(a to l)), facilitating efficient oxygen and nutrient exchange. Within these hydrogels, several human-scale structures were printed with good stability and fidelity. Furthermore, researchers incorporated MC3T3-E1 cells and murine BMSCs. Throughout the 21 day cell culture, both cell

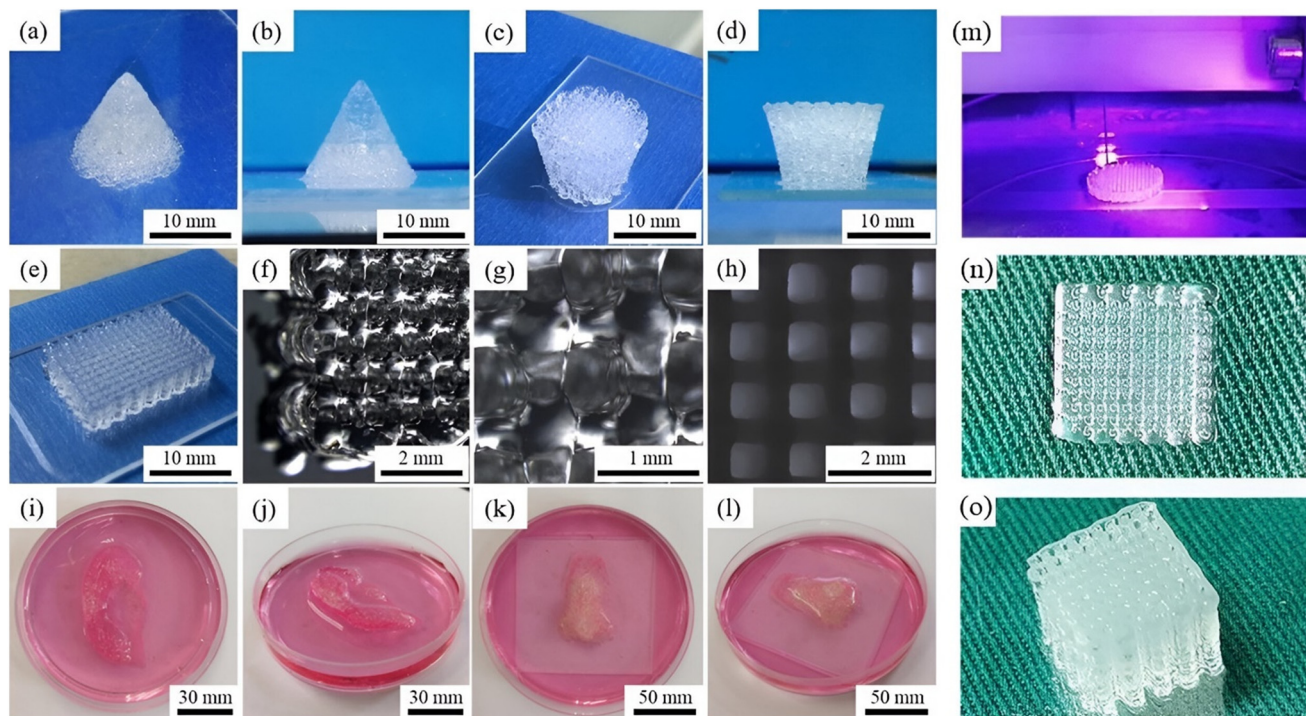


Fig. 3 Images of several 3D printed scaffolds by Ink G1.5P10. Aerial view (a) and left view (b) images of a sharp cone structure. Aerial view (c) and left view (d) images of a reverse square prism structure. (e) The aerial view image of a cuboid structure. Its partial enlarged images obtained by a stereoscope shown as (f)–(h) (after being treated with liquid nitrogen). Top view (i) and aerial view (j) images of a 3D printed gel human ear. Top view (k) and aerial view (l) images of a 3D printed gel human nose. (permission from Elsevier).¹³¹ (m)–(o) Printing process of the scaffold, top view and side view of the scaffold with 60 layers (permission from Elsevier¹³³).

types demonstrated both stability and viability, with cell survival exceeding 87%.¹³¹ The deferoxamine-loaded GelMA/GGMA-based hybrid bio-ink was improved by Zhihui *et al.* using ionic and light crosslinking techniques to create a 3D-printed scaffold of 60 layers as shown in (Fig. 3(m)–(o)). The hydrogels made of GGMA/GelMA had the best printing and tensile qualities. The regulated release of pharmaceuticals from GGMA/GelMA hydrogels could be used to simulate *in vivo* bone reserve, HUVEC tube improvement, and cell migration. A rat cranial defect model study demonstrated that angiogenesis and bone repair might occur *in vivo* by activating the hypoxia-inducible factor 1 (one)-alpha signaling pathway. To repair the bond defect, hybrid hydrogel 3D bioprinting can assist with osteogenesis and angiogenesis.¹³³

4.3 Skin

The body's largest organ, the skin, serves as a protective barrier against the external environment. Bioprinted skin must possess key attributes to facilitate effective nutrient/waste exchange and promote robust skin regeneration. These include biocompatibility, mechanical properties aligned with natural skin tissue, and a meticulously interconnected network of pores.^{161,162} Lately, there has been a growing trend in developing 3D multicellular HA-based constructs.⁶ In addition to HA, researchers have explored various other biocompatible materials like collagen, fibrin, decellularized

dermis and epidermis, ALG,¹⁴⁷ chitosan,^{7,147} gelatin, and cellulose,⁹⁵ aiming to facilitate skin wound healing. Liu *et al.* introduced an innovative approach by combining ALG/gelatin to produce a bioink with enhanced printability of multilayered scaffold structures. Through an in-depth rheological analysis, they identified the optimal blend comprising 2% ALG and 15% gelatin. Introducing 2% CaCl₂ triggered a remarkable ionic gelation process driven by the ALG component, which substantially stabilizes the fabricated structures. This hybrid bioink encapsulated hAECs and was employed for bioprinting a skin-like, bi-layered membrane construct. The outcomes of the *in vitro* study demonstrated a notable enhancement in the epithelial cell phenotype of the hAECs.¹⁶³ In a study by Zhao *et al.*, the approach involved the incorporation of poly(*p*-phenylene vinyl acetate) (PPV) and HA into ALG/gelatin ink blends, resulting in skin patches imbued with distinctive biological properties. The ALG/HA/Gel-A5G81/PPV skin patches proved instrumental in effectively healing the wound and fostering the development of new skin tissue. Importantly, these patches exhibited *in vivo* biodegradability, signifying their potential as a promising avenue for advanced wound healing approaches.¹⁶⁴ Due to the uniform distribution of chitosan and kaolinite nanoclay utilizing the screw-based pen-type extruder, Bhattacharyya *et al.* in this study demonstrated the structural stability of the multicomponent ALG-based hydrogel. Extrudable multilayered big and sophisticated

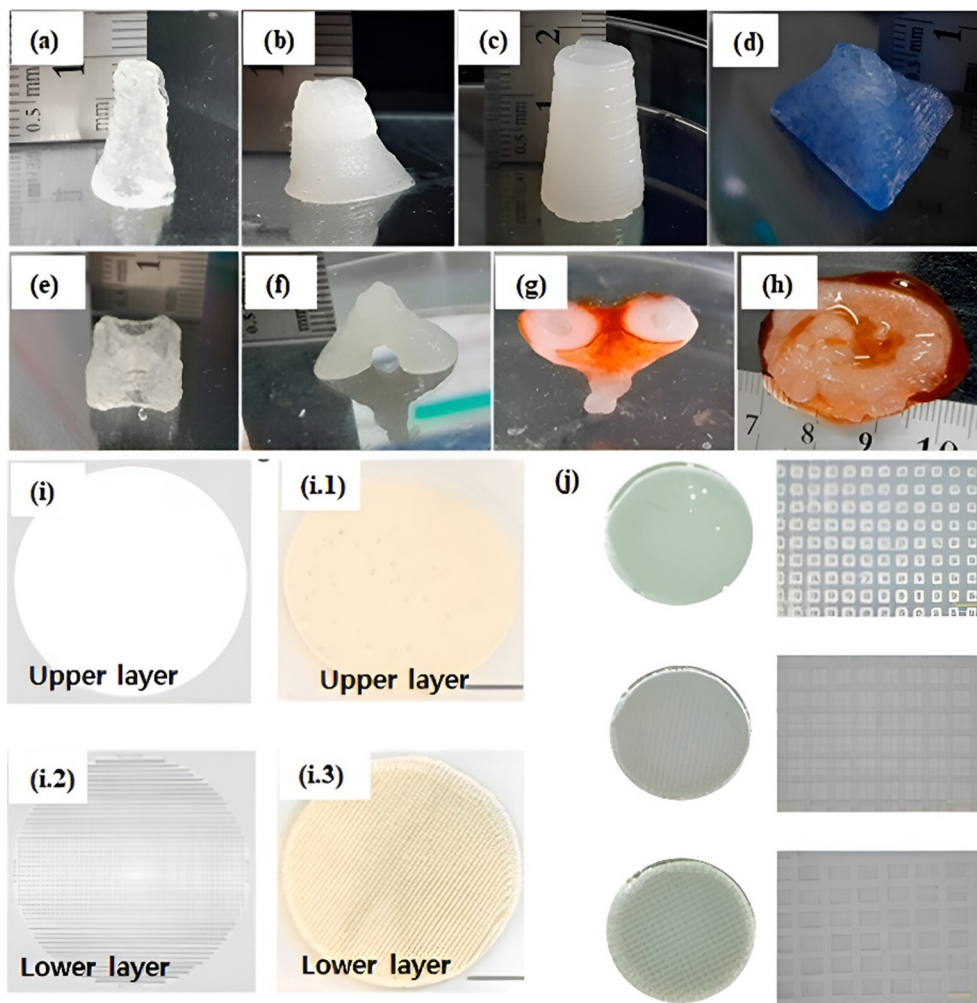


Fig. 4 (a)–(c) 3D printed hollow cylinder with varying nanocomposite concentration, (d) pyramid, (e) star, (f) top view of the bifurcated tube, (g) bottom view of the bifurcated tube, (h) and human ear (permission from Elsevier⁷). (i) & (i.1) The upper dense layer is dense mimicking the epidermis, (i.2) and (i.3) the lower porous layer resembling the dermis. (j) The printed products by DLP-based 3D printing showed complex structures of CAD images of 3D biprinted products.¹⁶⁵

tissue engineering constructs can be created using this multiple crosslinking method of the self-standing hydrogel with good post-printing shape fidelity. For large scale 3D bioprinting, the “Biowork™” pen with a roller head attached to a 3D bioprinter is utilized. Bioinks made from osteoblast and fibroblast cells in varying concentrations of kaolin-dispersed alginate/chitosan hydrogels were used, and various multilayered 3D biprinted constructs were demonstrated as given in (Fig. 4(a)–(h)). Additionally, it increases the cellular activity of two distinct cell types (osteoblast and fibroblast), which are targeted towards bone and skin tissue engineering.⁷ The double crosslinked hydrogel with HA as its foundation showed excellent swelling characteristics, biocompatibility, and extremely controlled breakdown. To treat diabetic foot ulcers, this material is being evaluated as a novel gel wound dressing. Zhou *et al.* printed functional live skin using a newly developed biomimetic GelMA/HA-NB/LAP hydrogel using digital light processing based 3D printing technology (Fig. 4 (i), (i.1)–(i.3) and (j)).¹⁶⁵

4.4 Nerve

The nervous system is pivotal in transmitting impulses across different body regions, thereby governing their coordinated movement. In addressing central and peripheral nerve injuries, the restoration of nerve function through regeneration is of paramount importance. However, the intricacy lies in the fact that nerve regeneration is particularly challenging for the central nervous system compared to the peripheral nervous system, as the former exhibits limited inherent self-repair capabilities.¹⁶⁶ Efforts directed at brain tissue regeneration through bioprinting entail the creation of 3D models that encompass elevated cell densities and a diverse array of cell types. This orchestrated complexity serves as a vital substrate for mimicking the intricate architecture of brain tissue. Moreover, the success of these models hinges upon the precise administration of external cues, which govern cell behavior and sustain the intricate web of cellular interactions.¹⁶⁷ By harnessing bioprinting, a dynamic

platform is established for fostering the renewal and preservation of the nervous system, addressing the challenges posed by nerve injuries and contributing to the advancement of regenerative therapies.¹⁶⁸ Peripheral nerve damage presents a clinically intricate and challenging scenario, often leading to compromised sensory and motor capabilities due to inherent regenerative potential limitations. Despite various efforts, their efficacy is frequently confined. In the pursuit of addressing peripheral nerve regeneration, nerve-guided catheters have emerged as captivating alternatives. Among

the therapeutic options available, autologous nerve transplantation, while popular, is burdened with shortcomings such as donor scarcity and potential neurological issues in the donor area.^{169,170} Presently, the predominant choice for malleable nerve cells in neural tissue engineering revolves around Schwann cells. These versatile cells possess the capacity to express an array of nerve growth factors (NGF), encompassing NGF, BDNF, GDNF, and PDGF, thereby instigating the formation of nerve synapses. Particularly adept at promoting the regeneration of the

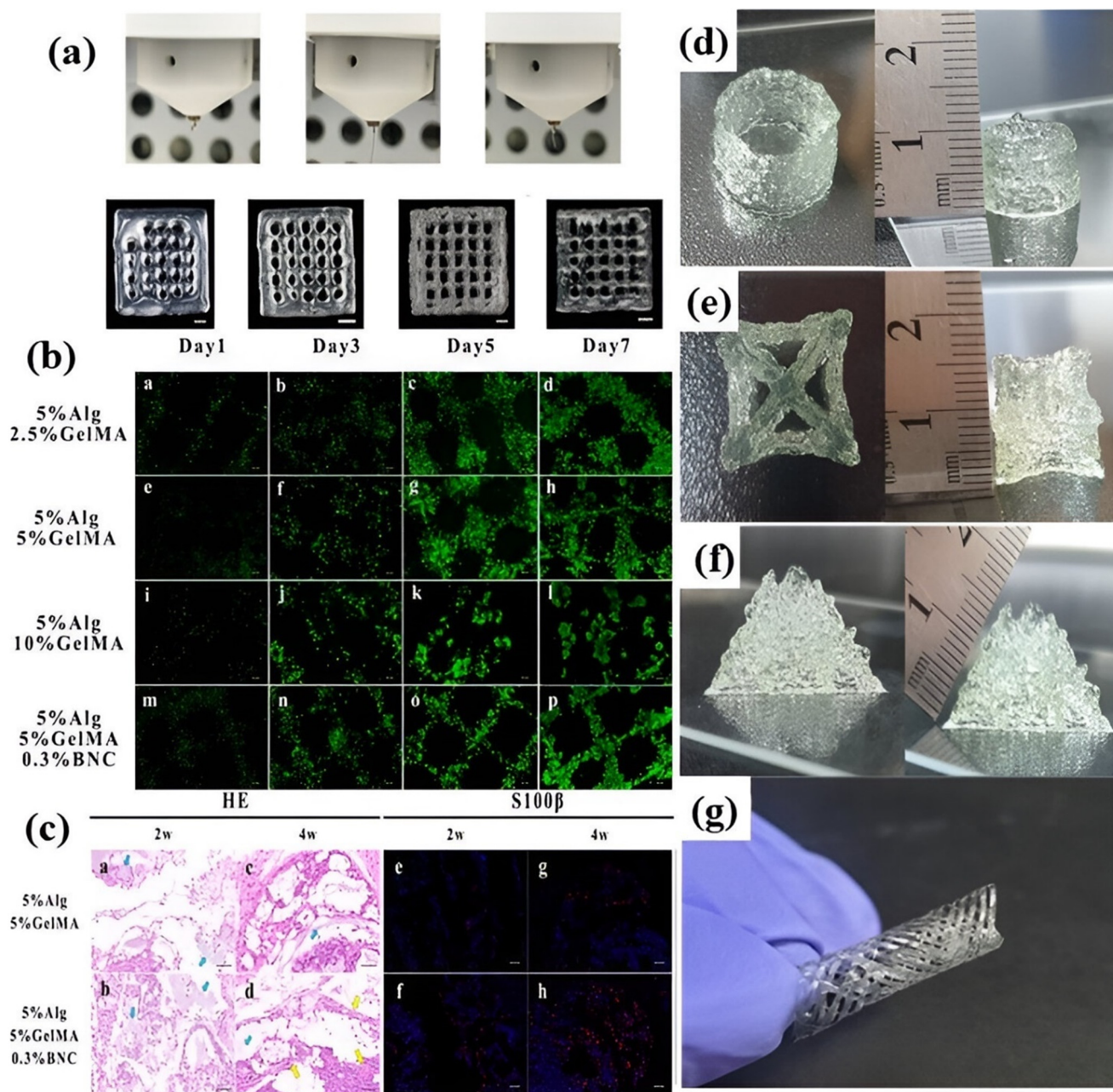


Fig. 5 Application of nerve tissue regeneration: (a) different concentrations of ALG BNC, and GelMA, excessive fluidity of extruded liquid, extruded lines suitable for printing. (b) and (c) Cell viability staining is performed on the hydrogel (permission Elsevier).¹³⁹ 3D bioprinted structures: (d) hollow tube structure, (e) star shape, (f) pyramid shape & (g) four-axis printed tubular shape with a bio responsive gelatin-hyaluronic acid hydrogel (permission MDPI⁶).

peripheral nervous system, Schwann cells find extensive application. Consequently, integrating ALG-based hydrogels housing Schwann cells can effectively facilitate the expression of multiple nerve growth factors, contributing to a conducive environment for nerve development.¹³⁷ As an illustration, the hydrogel framework, crafted from a blend of low-viscosity ALG, HA, and fibrinogen, notably fosters heightened activity and proliferation of Schwann cells, accompanied by the expression of essential biological proteins. Moreover, this scaffold exerts a regulatory influence on the maturation of the dorsal root ganglion, concurrently promoting the alignment of Schwann cells.¹³⁸ The study explored the printability aspects of ALG, GelMA and a hybrid bioink composed of bacterial nano-cellulose (BNC), accompanied by an in-depth analysis of the resulting material's physicochemical attributes reported by Wu *et al.* Subsequently, RSC96 cells were integrated into the bioink and used for bioprinting. The incorporation of ALG and BNC in the composition reveals that post photo-crosslinking and chemical crosslinking, the BNC and GelMA composite exhibited the most delicate lines compared to other groups. This suggests that the addition of BNCs enables the preservation of the scaffold's original shape, highlighting its potential to maintain inherent structural integrity as shown in (Fig. 5a). Furthermore, the biocompatibility of the engineered construct was confirmed through experiments involving unclothed mice. To facilitate nerve regeneration, the combination of BNC with ALG/GelMA was employed in the hydrogel construct. Notably, *in vivo* investigations revealed compelling outcomes, wherein directed growth, proliferation, and gene expression of rat Schwann cells were promoted as shown in (Fig. 5b and c).¹³⁹ Haring *et al.* employed a combination of natural and synthetic polymers to successfully bioprint 3D micro-physiological brain tissue, facilitating research on disease models and regenerative potential. The bioprinting process involved using a specially formulated bioink of HA, ALG, and gelatin, which could be crosslinked using CaCl_2 . This investigation was conducted in comparison to the stiffness of the original brain tissue. Intriguingly, it was discovered that the hydrogel attained a stiffness level comparable to that of native human tissue at higher concentrations of gelatin.¹⁴⁰ Similarly, Khatun *et al.* demonstrated genipin crosslinked with a gelatin–HA bioresponsive hydrogel for high shape fidelity and resolution. A variety of forms, including hollow tubes and star shapes that were each made up of about 50 layers and could reach a height of 1 cm (Fig. 5d and e), were successfully printed. These intricate printed forms have a porous surface morphology that was visible after lyophilization. The gelatin–HA gel-based 3D printed structures, in particular the 1.5 cm tall pyramid-shaped structure (Fig. 5f) with about 100 layers, demonstrated exceptional stability and shape integrity. An intriguing finding was that the printed structures stuck to the substrates very well without the need for further adhesive, indicating how highly sticky the hydrogel is to the surface.⁶

4.5 Muscle

Muscle tissue provides for around 45% of total body weight and accounts for a considerable percentage of the body's composition, with a significant portion interwoven with bones. Peripheral nerves intricately innervate muscles, particularly those within motor organs. In nerve injury, particularly to peripheral nerves, the innervated muscles undergo a process of atrophy, leading to a decline in size and function.¹⁷¹ Hence, the restoration of muscle tissue involves a combination of strategies encompassing nerve and vascular regeneration techniques. Currently, the principal avenues for promoting muscle regeneration involve administering growth factors and cell transplantation. Notably, ALG is a widely favored biomaterial for 3D printing muscle constructs, particularly in ALG-based bioink with cells.¹⁷² Lee *et al.* demonstrated a bioink composed of collagen, GelMA, and dECM, restoring muscle tissue using 3D bioprinting with a novel photo-crosslinking technique. This crosslinking method was used to create multilayered 3D cell-laden structures, mechanically stable and biologically safe, cell-laden struts. When administered to a volumetric muscle defect injury in mice, the bioprinted hASC-containing constructs showed noticeably improved muscle regeneration over those made with a traditional bioprinting method.¹⁷³ The C2C12 myogenic cells, a widely used cell type in tissue engineering for muscle regeneration, were studied in those 3 commercial biomaterials. The results showed robust cell proliferation and successful differentiation across all 3 muscular scaffolds. The cells also demonstrated significant myotubular organization along a linear structure and the ability to coalesce and fuse, generating structurally aligned multinucleated myotubes.¹⁷⁴

4.6 Other tissues

Tuning the polysaccharide-based hydrogels through a simple modification or crosslinking method enabled researchers to use them in tissue engineering and bioprinting tissues such as retina, liver, heart, *etc.*¹⁷⁵ The kidney's substantial complexity and intricate structure pose a formidable challenge for complete bioprinting in a single endeavor. The formidable nature of this task arises from both the intricacy of the organ's design and its considerable size. The most practical approach involves employing 3D bioprinting to create microscopic components, often called “mini tissue building blocks.” Subsequently, these intricate building blocks can be assembled to construct larger, functional kidney structures.¹⁷⁵ To circumvent the intricacies of organ size and cellular diversity, the utilization of organoids presents a viable avenue for bioprinting endeavors. However, achieving functional viability for the kidney necessitates addressing critical concerns related to the organ's ductal system and vasculature, which present their own complex challenges.¹⁷⁶ Cardiovascular disease (CD) is the foremost global cause of mortality, accounting for 31% of all deaths. This prevalent health issue primarily arises from plaque

accumulation, a condition known as atherosclerosis, which constricts arteries and other vital blood vessels. This broader category of cardiovascular ailments encompasses conditions like myocardial infarction, congestive heart failure, stroke, and valvular heart disease.¹⁷⁷ A composite bioink (ALG/GelMA) is intricately deposited using the coaxial extrusion bioprinting technology, encasing endothelial cells, and creating micro fibrous scaffolds with intricate micro-channels. This novel approach significantly promotes the process of scaffold reendothelialization, encouraging the growth and proliferation of HUVECs. After this, the deendothelialized scaffold serves as a foundation for introducing cardiomyocytes, culminating in developing a deendothelialized myocardial scaffold. This scaffold configuration closely mimics the structural attributes of native myocardial tissue. Remarkably, the scaffold orchestrates synchronized and spontaneous contractions and maintains a consistent rhythm, yielding an impressive heartbeat rate ranging from 55 to 75 beats per minute.¹⁷⁸ LA *et al.* created axially symmetric aortic valve scaffolds with a 12 mm internal diameter using PEGDA and ALG, similar to double nozzle extrusion printing. The hybrid scaffolds had an elastic modulus of 5.3–74.6 kPa and a nearly 100% survival rate for cultured PAVIC over 21 days.¹⁷⁹ The bioink derived from ALG can be used to create 3D scaffolds that enhance the mechanical and biological properties of the hydrogels. These scaffolds, when adapted to the liver's honeycomb architecture, can significantly improve hepatocyte proliferation, replication, and overall liver function preservation.¹⁸⁰ Rania *et al.* used a core-shell bioprinting technique to enhance the complexity of hepatocytes' microenvironment. They used distinct core-shell structures housing different cell types and incorporated Matrigel into ALG/MA to mimic the honeycomb-like arrangement in liver tissue. This led to significant increase in hepatocyte activity and proliferation within the scaffold. The introduction of plasma or fibrin into the core of ALG/MA facilitated the organization of fibroblasts into a network, creating a more intricate and realistic microenvironment.¹⁸¹ A 3D retina-equivalent construct was 3D printed using GM and triethylamine to methacrylate-HA at different degrees. Cell-binding motifs were added to HA-GM polymers, followed by photoirradiation using a LAP photo initiator. The hydrogels were further enhanced with RGD to improve adhesion properties. The construct, with a base layer of 125 μm and a top layer of 250 μm , showed over 70% vitality and matured into retinal photoreceptors, ganglion cells, and bipolar cells within 14 days, resembling real retina layers.^{182,183}

5. Future directions

3D bioprinting has made significant progress, but the challenge lies in creating tissue engineering structures with desired bioink properties. Natural polysaccharides continue to be the preferred option for creating bioinks for 3D

bioprinting tissue engineering. Because of their greater bioactivity, lower immune response and biocompatibility, polysaccharides are frequently preferred over proteins and synthetic polymers like PCL, PEG, and Pluronic. These naturally occurring biomaterials are excellent at simulating the ECM and offer vital signals that support cell adhesion, proliferation, differentiation, and tissue regeneration, all of which are necessary for not only tissue regeneration but also proper integration with host tissues and operation of bioengineered tissues. Although they have limitations in terms of processing and cell interactions in 3D bioprinting tissue engineering, a number of interesting methods and chemical alterations have already shown how to overcome these restrictions. 3D bioprinting has made substantial progress using polysaccharide-based bioinks due to their biocompatibility and effective mimicry of the ECM in different tissues. These hydrogels can work as a major bioink component for engineering complex, multilayered structures and are favored for their non-toxicity, natural degradability with minimal side effects, and diverse crosslinking properties. Current research is enhancing these bioinks' properties with modifications of hydrogels to their chemical and physical structures similar to ECM's proteoglycans including cell-adhesive oligopeptides, aiming to closely replicate native tissue environments and improve the functionality of the engineered tissues. The field is moving towards integrating bioactive molecules such as growth factors and nanoparticles, as well as incorporating real-time monitoring technologies to refine the bioprinting process. This integration is expected to drive significant advances in personalized regenerative medicine, leading to more effective tissue and organ replacements. Especially in complex tissue regeneration, fabrication of multilayered tissues such as skin, blood vessel and osteocartilage emerges as an essential tool. Furthermore, development of bioinks and hydrogels is critical in tissue engineering and regenerative medicine with post-printing fidelity, *i.e.* higher interaction between the layers of the tissue engineering constructs. The convergence of these technologies holds promise for accelerating the development of personalized, functional tissue replacements, ushering in a new era of 3D bioprinting of multilayered complex tissue scaffolds, regenerative medicine and tissue engineering.

6. Abbreviations

DECL	Diethylaminoethyl cellulose
ALG	Alginate (sodium salt)
CHN	Chitosan
HA	Hyaluronic acid
CEL	Cellulose
DEX	Dextran
ECM	Extracellular matrix
GEG	Gellan gum
AGR	Agarose
KJG	Konjac gum

GG	Guar gum
XTG	Xanthan gum
CS	Chondroitin sulfate
PRP	Platelet-rich plasma
BMSCs	Bone marrow mesenchymal stromal cells
dECM	Decellularized extracellular matrix
MFC	Meniscal fibrocartilage cells
TGF	Transforming growth factor beta-1
MSCs	Mesenchymal stem cells
MECM	Meniscal extracellular matrix
SMSc	Synovial mesenchymal stem cells
hMSCs	Human mesenchymal stem cells
ATDC5	Mouse teratocarcinoma cell
MC3T3-E1	Clonal murine cell line of immature osteoblasts
MA	Methacrylate
HASCs	Human adipose-derived stem cells
BMP-2	Bone morphogenetic protein-2
GGMA	Gellan gum methacrylate
HUVEC	Human umbilical vein endothelial cells
hAECs	Human amniotic epithelial cells
NGF	Nerve growth factor
BDNF	Brain-derived neurotrophic factor
GDNF	Glial cell-derived neurotrophic factor
PDGF	Platelet-derived growth factor
BNC	Bacterial nanocellulose
RSc	Rat Schwann Cell 96
CaCl ₂	Calcium chloride
CD	Cardiovascular disease
PAVIC	Porcine aortic valve interstitial cells
BTE	Bone tissue engineering
CCD-986	Human skin fibroblast cells
PBMCs	Human peripheral blood mononuclear cells
hFBs	Human fibroblasts
MeGC	Methacrylated glycol chitosan
FG	Fish gelatin
CB	Cuttle bone
CMP	Cuttlefish melanin nanoparticles
PLs	Platelet lysates
GFs	Growth factor
CNC	Carboxylated-cellulose nanocrystal
PDANP	Polydopamine nanoparticles

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was financially supported by Seoul National University of Science and Technology.

References

- 1 D. Lee, J.-Y. Park, Y.-W. Seo, X. Jin, J. Hong and A. Bhattacharyya, Photo-crosslinked gelatin methacryloyl strengthened with nanoparticles for regeneration of rabbit calvarial defects, *J. Periodontal Implant Sci.*, 2023, **53**, 321–335.
- 2 G. Janarthanan, J. H. Kim, I. Kim, C. Lee, E.-J. Chung and I. Noh, Manufacturing of self-standing multi-layered 3D-bioprinted alginate-hyaluronate constructs by controlling the cross-linking mechanisms for tissue engineering applications, *Biofabrication*, 2022, **14**(3), 035013.
- 3 A. Bhattacharyya, V. K. Priya, J.-h. Kim, M. R. Khatun, R. Nagarajan and I. Noh, Nanodiamond enhanced mechanical and biological properties of extrudable gelatin hydrogel cross-linked with tannic acid and ferrous sulphate, *Biomater. Res.*, 2022, **26**(1), 37.
- 4 A. Bhattacharyya, M. R. Khatun, S. Narmatha, R. Nagarajan and I. Noh, Modulation of 3D Bioprintability in Polysaccharide Bioink by Bioglass Nanoparticles and Multiple Metal Ions for Tissue Engineering, *Tissue Eng. Regen. Med.*, 2023, 1–15.
- 5 H. N. Tran, I. G. Kim, J. H. Kim, A. Bhattacharyya, E. J. Chung and I. Noh, Incorporation of Cell-Adhesive Proteins in 3D-Printed Lipoic Acid-Maleic Acid-Poly(Propylene Glycol)-Based Tough Gel Ink for Cell-Supportive Microenvironment, *Macromol. Biosci.*, 2023, **23**(11), 2300316.
- 6 M. R. Khatun, A. Bhattacharyya, M. Gunbayar, M. Jung and I. Noh, Study on Bioresponsive Gelatin-Hyaluronic Acid-Genipin Hydrogel for High Cell-Density 3D Bioprinting, *Gels*, 2023, **9**, 601.
- 7 A. Bhattacharyya, H.-w. Ham, J. Sonh, M. Gunbayar, R. Jeffy and R. Nagarajan, *et al.*, 3D bioprinting of complex tissue scaffolds with in situ homogeneously mixed alginate-chitosan-kaolin bioink using advanced portable biopen, *Carbohydr. Polym.*, 2023, 121046.
- 8 M. P. Sekar, H. Budharaju, A. Zennifer, S. Sethuraman, N. Vermeulen, D. Sundaramurthi and D. M. Kalaskar, Current standards and ethical landscape of engineered tissues—3D bioprinting perspective, *J. Tissue Eng.*, 2021, **12**, 20417314211027677.
- 9 H. Budharaju, A. Subramanian and S. Sethuraman, Recent advancements in cardiovascular bioprinting and bioprinted cardiac constructs, *Biomater. Sci.*, 2021, **9**(6), 1974–1994.
- 10 P. M. Sivakumar, A. A. Yetisgin, S. B. Sahin, E. Demir and S. Cetinel, Bone tissue engineering: Anionic polysaccharides as promising scaffolds, *Carbohydr. Polym.*, 2022, **283**, 119142.
- 11 A. Zaszczynska, M. Moczulska-Heljak, A. Gradys and P. Sajkiewicz, Advances in 3D printing for tissue engineering, *Materials*, 2021, **14**(12), 3149.
- 12 P. Wu, X. Xi, R. Li and G. Sun, Engineering polysaccharides for tissue repair and regeneration, *Macromol. Biosci.*, 2021, **21**(9), 2100141.
- 13 Y. Zhou, X. Chen, T. Chen and X. Chen, A review of the antibacterial activity and mechanisms of plant polysaccharides, *Trends Food Sci. Technol.*, 2022, **123**, 264–280.
- 14 H. Claus-Desbonnet, E. Nikly, V. Nalbantova, D. Karcheva-Bahchevanska, S. Ivanova and G. Pierre, *et al.*, Polysaccharides and their derivatives as potential antiviral molecules, *Viruses*, 2022, **14**(2), 426.

- 15 R. Guo, M. Chen, Y. Ding, P. Yang, M. Wang and H. Zhang, *et al.*, Polysaccharides as potential anti-tumor biomacromolecules—a review, *Front. Nutr.*, 2022, **9**, 838179.
- 16 H. Jin, M. Li, F. Tian, F. Yu and W. Zhao, An overview of antitumour activity of polysaccharides, *Molecules*, 2022, **27**(22), 8083.
- 17 E. J. Murphy, G. W. Fehrenbach, I. Z. Abidin, C. Buckley, T. Montgomery and R. Pogue, *et al.*, Polysaccharides—Naturally Occurring Immune Modulators, *Polymer*, 2023, **15**(10), 2373.
- 18 Y. Hu, Y. He, Z. Niu, T. Shen, J. Zhang and X. Wang, *et al.*, A review of the immunomodulatory activities of polysaccharides isolated from Panax species, *J. Ginseng Res.*, 2022, **46**(1), 23–32.
- 19 M. Mahinroosta, Z. J. Farsangi, A. Allahverdi and Z. Shakoobi, Hydrogels as intelligent materials: A brief review of synthesis, properties and applications, *Mater. Today Chem.*, 2018, **8**, 42–55.
- 20 J. N. Israelachvili, *Intermolecular and surface forces*, Academic press, 2011.
- 21 Y. Liu, J. Wang, H. Chen and D. Cheng, Environmentally friendly hydrogel: A review of classification, preparation and application in agriculture, *Sci. Total Environ.*, 2022, **846**, 157303.
- 22 D. Kühbeck, J. Mayr, M. Häring, M. Hofmann, F. Quignard and D. D. Díaz, Evaluation of the nitroaldol reaction in the presence of metal ion-crosslinked alginates, *New J. Chem.*, 2015, **39**(3), 2306–2315.
- 23 M. F. Akhtar, M. Hanif and N. M. Ranjha, Methods of synthesis of hydrogels... A review, *Saudi Pharm. J.*, 2016, **24**(5), 554–559.
- 24 E. Caló and V. V. Khutoryanskiy, Biomedical applications of hydrogels: A review of patents and commercial products, *Eur. Polym. J.*, 2015, **65**, 252–267.
- 25 N. Reddy, R. Reddy and Q. Jiang, Crosslinking biopolymers for biomedical applications, *Trends Biotechnol.*, 2015, **33**(6), 362–369.
- 26 A. Sheikhi, J. de Rutte, R. Haghniaz, O. Akouissi, A. Sohrabi, D. Di Carlo and A. Khademhosseini, Microfluidic-enabled bottom-up hydrogels from annealable naturally-derived protein microbeads, *Biomaterials*, 2019, **192**, 560–568.
- 27 T. A. Ahmed, E. V. Dare and M. Hincke, Fibrin: a versatile scaffold for tissue engineering applications, *Tissue Eng., Part B*, 2008, **14**(2), 199–215.
- 28 M. A. de Moraes, E. Paternotte, D. Mantovani and M. M. Beppu, Mechanical and biological performances of new scaffolds made of collagen hydrogels and fibroin microfibers for vascular tissue engineering, *Macromol. Biosci.*, 2012, **12**(9), 1253–1264.
- 29 A. Gandini, T. M. Lacerda, A. J. Carvalho and E. Trovatti, Progress of polymers from renewable resources: furans, vegetable oils, and polysaccharides, *Chem. Rev.*, 2016, **116**(3), 1637–1669.
- 30 D. Impellizzeri, R. Siracusa, M. Cordaro, A. F. Peritore, E. Gugliandolo and R. D'amico, *et al.*, Protective effect of a new hyaluronic acid-carnosine conjugate on the modulation of the inflammatory response in mice subjected to collagen-induced arthritis, *Biomed. Pharmacother.*, 2020, **125**, 110023.
- 31 H. Q. Song, Y. Fan, Y. Hu, G. Cheng and F. J. Xu, Polysaccharide-peptide conjugates: a versatile material platform for biomedical applications, *Adv. Funct. Mater.*, 2021, **31**(6), 2005978.
- 32 P. Sahariah and M. Másson, Antimicrobial chitosan and chitosan derivatives: A review of the structure-activity relationship, *Biomacromolecules*, 2017, **18**(11), 3846–3868.
- 33 L. Lin, Y. Fang, Y. Liao, G. Chen, C. Gao and P. Zhu, 3D printing and digital processing techniques in dentistry: a review of literature, *Adv. Eng. Mater.*, 2019, **21**(6), 1801013.
- 34 L. Pugliese, S. Marconi, E. Negrello, V. Mauri, A. Peri and V. Gallo, *et al.*, The clinical use of 3D printing in surgery, *Updates Surg.*, 2018, **70**, 381–388.
- 35 S. Song, X. Liu, J. Huang and Z. Zhang, Neural stem cell-laden 3D bioprinting of polyphenol-doped electroconductive hydrogel scaffolds for enhanced neuronal differentiation, *Biomater. Adv.*, 2022, **133**, 112639.
- 36 S. Freeman, S. Calabro, R. Williams, S. Jin and K. Ye, Bioink formulation and machine learning-empowered bioprinting optimization, *Front. bioeng. biotechnol.*, 2022, **10**, 913579.
- 37 A. I. Cernescu and M. Ioniță, The current state of the art in gellan-based printing inks in tissue engineering, *Carbohydr. Polym.*, 2023, 120676.
- 38 M. Jin, J. Shi, W. Zhu, H. Yao and D.-A. Wang, Polysaccharide-based biomaterials in tissue engineering: a review, *Tissue Eng., Part B*, 2021, **27**(6), 604–626.
- 39 M. S. Algahtani, A. A. Mohammed and J. Ahmad, Extrusion-based 3D printing for pharmaceuticals: contemporary research and applications, *Curr. Pharm. Des.*, 2018, **24**(42), 4991–5008.
- 40 J. K. Placone and A. J. Engler, Recent advances in extrusion-based 3D printing for biomedical applications, *Adv. Healthcare Mater.*, 2018, **7**(8), 1701161.
- 41 X. Cui, J. Li, Y. Hartanto, M. Durham, J. Tang and H. Zhang, *et al.*, Advances in extrusion 3D bioprinting: a focus on multicomponent hydrogel-based bioinks, *Adv. Healthcare Mater.*, 2020, **9**(15), 1901648.
- 42 N. Khoshnood and A. Zamanian, A comprehensive review on scaffold-free bioinks for bioprinting, *Bioprinting*, 2020, **19**, e00088.
- 43 N. Ashammakhi, S. Ahadian, C. Xu, H. Montazerian, H. Ko and R. Nasiri, *et al.*, Bioinks and bioprinting technologies to make heterogeneous and biomimetic tissue constructs, *Mater. Today Bio*, 2019, **1**, 100008.
- 44 A. Dhawan, P. M. Kennedy, E. B. Rizk and I. T. Ozbolat, Three-dimensional bioprinting for bone and cartilage restoration in orthopaedic surgery, *J. Am. Acad. Orthop. Surg.*, 2019, **27**(5), e215–e226.
- 45 G. Gao, H. Kim, B. S. Kim, J. S. Kong, J. Y. Lee and B. W. Park, *et al.*, Tissue-engineering of vascular grafts containing endothelium and smooth-muscle using triple-coaxial cell printing, *Appl. Phys. Rev.*, 2019, **6**, 041402.
- 46 R. El Khoury, N. Nagiah, J. A. Mudloff, V. Thakur, M. Chattopadhyay and B. Joddar, 3D bioprinted spheroidal

- droplets for engineering the heterocellular coupling between cardiomyocytes and cardiac fibroblasts, *Cyborg Bionic Syst.*, 2021, **2021**, 9864212.
- 47 M. Kesti, M. Müller, J. Becher, M. Schnabelrauch, M. D'Este, D. Eglin and M. Zenobi-Wong, A versatile bioink for three-dimensional printing of cellular scaffolds based on thermally and photo-triggered tandem gelation, *Acta Biomater.*, 2015, **11**, 162–172.
 - 48 C. B. Rodell, J. E. Mealy and J. A. Burdick, Supramolecular guest–host interactions for the preparation of biomedical materials, *Bioconjugate Chem.*, 2015, **26**(12), 2279–2289.
 - 49 R. Khoeini, H. Nosrati, A. Akbarzadeh, A. Eftekhari, T. Kavetsky and R. Khalilov, *et al.*, Natural and synthetic bioinks for 3D bioprinting, *Adv. NanoBiomed Res.*, 2021, **1**(8), 2000097.
 - 50 M. Burke, B. M. Carter and A. W. Perriman, Bioprinting: uncovering the utility layer-by-layer, *J. 3D Print. Med.*, 2017, **1**(3), 165–179.
 - 51 D. X. Chen and D. X. Chen, *Extrusion bioprinting of scaffolds*, Springer, 2019.
 - 52 P. Kumar, S. Ebbens and X. Zhao, Inkjet printing of mammalian cells—Theory and applications, *Bioprinting*, 2021, **23**, e00157.
 - 53 A. Zennifer, A. Subramanian and S. Sethuraman, Design considerations of bioinks for laser bioprinting technique towards tissue regenerative applications, *Bioprinting*, 2022, **27**, e00205.
 - 54 A. Zennifer, S. Manivannan, S. Sethuraman, S. G. Kumbar and D. Sundaramurthi, 3D bioprinting and photocrosslinking: emerging strategies & future perspectives, *Biomater. Adv.*, 2022, **134**, 112576.
 - 55 I. T. Ozbolat, W. Peng and V. Ozbolat, Application areas of 3D bioprinting, *Drug Discovery Today*, 2016, **21**(8), 1257–1271.
 - 56 H. Herrada-Manchón, M. A. Fernández and E. Aguilar, Essential guide to hydrogel rheology in extrusion 3D printing: how to measure it and why it matters?, *Gels*, 2023, **9**(7), 517.
 - 57 D. M. Kirchmayer and I. R. Gorkin, An overview of the suitability of hydrogel-forming polymers for extrusion-based 3D-printing, *J. Mater. Chem. B*, 2015, **3**(20), 4105–4117.
 - 58 L. Han, K. Liu, M. Wang, K. Wang, L. Fang and H. Chen, *et al.*, Mussel-inspired adhesive and conductive hydrogel with long-lasting moisture and extreme temperature tolerance, *Adv. Funct. Mater.*, 2018, **28**(3), 1704195.
 - 59 D. Diekjürgen and D. W. Grainger, Polysaccharide matrices used in 3D in vitro cell culture systems, *Biomaterials*, 2017, **141**, 96–115.
 - 60 M. R. Khatun, A. Bhattacharyya, S. Taheri, H. Hw, H. Kim, S. H. Chang and I. Noh, High Molecular Weight Fucoidan Loading Into and Release from Hyaluronate-Based Prefabricated Hydrogel and its Nanogel Particles Controlled by Variable Pitch and Differential Extensional Shear Technology of Advanced Twin Screw-Based System, *Adv. Mater. Technol.*, 2023, **8**(5), 2201478.
 - 61 C. Benwood, J. Chrenek, R. L. Kirsch, N. Z. Masri, H. Richards, K. Teetzen and S. M. Willerth, Natural biomaterials and their use as bioinks for printing tissues, *Bioengineering*, 2021, **8**(2), 27.
 - 62 Q. Zhou, H. Kang, M. Bielec, X. Wu, Q. Cheng, W. Wei and H. Dai, Influence of different divalent ions cross-linking sodium alginate-polyacrylamide hydrogels on antibacterial properties and wound healing, *Carbohydr. Polym.*, 2018, **197**, 292–304.
 - 63 S. Khalil and W. Sun, Bioprinting endothelial cells with alginate for 3D tissue constructs, *J. Biomech. Eng.*, 2009, **131**(11), 111002.
 - 64 G. Janarthanan, S. Lee and I. Noh, 3D printing of bioinspired alginate-albumin based instant gel ink with electroconductivity and its expansion to direct four-axis printing of hollow porous tubular constructs without supporting materials, *Adv. Funct. Mater.*, 2021, **31**(45), 2104441.
 - 65 I. Braccini and S. Pérez, Molecular basis of Ca²⁺-induced gelation in alginates and pectins: the egg-box model revisited, *Biomacromolecules*, 2001, **2**(4), 1089–1096.
 - 66 K. Y. Lee and S. H. Yuk, Polymeric protein delivery systems, *Prog. Polym. Sci.*, 2007, **32**(7), 669–697.
 - 67 L. T. Somasekharan, R. Raju, S. Kumar, R. Geevarghese, R. P. Nair, N. Kasoju and A. Bhatt, Biofabrication of skin tissue constructs using alginate, gelatin and diethylaminoethyl cellulose bioink, *Int. J. Biol. Macromol.*, 2021, **189**, 398–409.
 - 68 L. Dogan, R. Scheuring, N. Wagner, Y. Ueda, S. Schmidt and P. Wörsdörfer, *et al.*, Human iPSC-derived mesodermal progenitor cells preserve their vasculogenesis potential after extrusion and form hierarchically organized blood vessels, *Biofabrication*, 2021, **13**(4), 045028.
 - 69 M. C. Teixeira, N. S. Lameirinhas, J. P. Carvalho, A. J. Silvestre, C. Vilela and C. S. Freire, A guide to polysaccharide-based hydrogel bioinks for 3D bioprinting applications, *Int. J. Mol. Sci.*, 2022, **23**(12), 6564.
 - 70 M. Rajabi, M. McConnell, J. Cabral and M. A. Ali, Chitosan hydrogels in 3D printing for biomedical applications, *Carbohydr. Polym.*, 2021, **260**, 117768.
 - 71 Q. Wu, D. Therriault and M.-C. Heuzey, Processing and properties of chitosan inks for 3D printing of hydrogel microstructures, *ACS Biomater. Sci. Eng.*, 2018, **4**(7), 2643–2652.
 - 72 N. M. Alves and J. F. Mano, Chitosan derivatives obtained by chemical modifications for biomedical and environmental applications, *Int. J. Biol. Macromol.*, 2008, **43**(5), 401–414.
 - 73 M. Sahranavard, A. Zamanian, F. Ghorbani and M. H. Shahrezaee, A critical review on three dimensional-printed chitosan hydrogels for development of tissue engineering, *Bioprinting*, 2020, **17**, e00063.
 - 74 S. Peers, A. Montembault and C. Ladavière, Chitosan hydrogels for sustained drug delivery, *J. Controlled Release*, 2020, **326**, 150–163.
 - 75 M. B. Oliveira, H. X. Bastos and J. F. Mano, Sequentially moldable and bondable four-dimensional hydrogels

- compatible with cell encapsulation, *Biomacromolecules*, 2018, **19**(7), 2742–2749.
- 76 Y.-H. Lee, Y.-L. Hong and T.-L. Wu, Novel silver and nanoparticle-encapsulated growth factor co-loaded chitosan composite hydrogel with sustained antimicrobidity and promoted biological properties for diabetic wound healing, *Mater. Sci. Eng., C*, 2021, **118**, 111385.
 - 77 T. Lim, Q. Tang, Z. Zhu, X. Wei and C. Zhang, Sustained release of human platelet lysate growth factors by thermosensitive hydroxybutyl chitosan hydrogel promotes skin wound healing in rats, *J. Biomed. Mater. Res., Part A*, 2020, **108**(10), 2111–2122.
 - 78 C. Tonda-Turo, I. Carmagnola, A. Chiappone, Z. Feng, G. Ciardelli, M. Hakkarainen and M. Sangermano, Photocurable chitosan as bioink for cellularized therapies towards personalized scaffold architecture, *Bioprinting*, 2020, **18**, e00082.
 - 79 S. Pisani, R. Dorati, F. Scocozza, C. Mariotti, E. Chiesa and G. Bruni, *et al.*, Preliminary investigation on a new natural based poly(γ -glutamic acid)/Chitosan bioink, *J. Biomed. Mater. Res., Part B*, 2020, **108**(7), 2718–2732.
 - 80 J. A. Burdick, C. Chung, X. Jia, M. A. Randolph and R. Langer, Controlled degradation and mechanical behavior of photopolymerized hyaluronic acid networks, *Biomacromolecules*, 2005, **6**(1), 386–391.
 - 81 I. Noh, N. Kim, H. N. Tran, J. Lee and C. Lee, 3D printable hyaluronic acid-based hydrogel for its potential application as a bioink in tissue engineering, *Biomater. Res.*, 2019, **23**(1), 1–9.
 - 82 J. K. Carrow, P. Kerativitayanan, M. K. Jaiswal, G. Lokhande and A. K. Gaharwar, *Polymers for bioprinting Essentials of 3D biofabrication and translation*, Elsevier, 2015, pp. 229–248.
 - 83 S. Trombino, C. Servidio, F. Curcio and R. Cassano, Strategies for hyaluronic acid-based hydrogel design in drug delivery, *Pharmaceutics*, 2019, **11**(8), 407.
 - 84 H.-Y. Hsieh, W.-Y. Lin, A. L. Lee, Y.-C. Li, Y.-J. Chen, K.-C. Chen and T.-H. Young, Hyaluronic Acid on the Urokinase Sustained Release with a Hydrogel System Composed of Poloxamer 407, *bioRxiv*, 2020, preprint, DOI: [10.1371/journal.pone.0227784](https://doi.org/10.1371/journal.pone.0227784).
 - 85 S. Ansari, I. M. Diniz, C. Chen, T. Aghaloo, B. M. Wu, S. Shi and A. Moshaverinia, Alginate/hyaluronic acid hydrogel delivery system characteristics regulate the differentiation of periodontal ligament stem cells toward chondrogenic lineage, *J. Mater. Sci.: Mater. Med.*, 2017, **28**, 1–12.
 - 86 R. Tsaryk, A. Gloria, T. Russo, L. Anspach, R. De Santis and S. Ghanaati, *et al.* Collagen-low molecular weight hyaluronic acid semi-interpenetrating network loaded with gelatin microspheres for cell and growth factor delivery for nucleus pulposus regeneration, *Acta Biomater.*, 2015, **20**, 10–21.
 - 87 Z. He, H. Zang, L. Zhu, K. Huang, T. Yi, S. Zhang and S. Cheng, An anti-inflammatory peptide and brain-derived neurotrophic factor-modified hyaluronan-methylcellulose hydrogel promotes nerve regeneration in rats with spinal cord injury, *Int. J. Nanomed.*, 2019, 721–732.
 - 88 L. Li, G. Jiang, W. Yu, D. Liu, H. Chen and Y. Liu, *et al.*, A composite hydrogel system containing glucose-responsive nanocarriers for oral delivery of insulin, *Mater. Sci. Eng., C*, 2016, **69**, 37–45.
 - 89 N. K. Agrawal, P. Allen, Y. H. Song, R. A. Wachs, Y. Du, A. D. Ellington and C. E. Schmidt, Oligonucleotide-functionalized hydrogels for sustained release of small molecule (aptamer) therapeutics, *Acta Biomater.*, 2020, **102**, 315–325.
 - 90 G. Janarthanan, H. S. Shin, I.-G. Kim, P. Ji, E.-J. Chung, C. Lee and I. Noh, Self-crosslinking hyaluronic acid-carboxymethylcellulose hydrogel enhances multilayered 3D-printed construct shape integrity and mechanical stability for soft tissue engineering, *Biofabrication*, 2020, **12**(4), 045026.
 - 91 P. S. Babo, R. L. Pires, L. Santos, A. Franco, F. Rodrigues and I. Leonor, *et al.*, Platelet lysate-loaded photocrosslinkable hyaluronic acid hydrogels for periodontal endogenous regenerative technology, *ACS Biomater. Sci. Eng.*, 2017, **3**(7), 1359–1369.
 - 92 Y. Deng, A. X. Sun, K. J. Overholt, Z. Y. Gary, M. R. Fritch and P. G. Alexander, *et al.*, Enhancing chondrogenesis and mechanical strength retention in physiologically relevant hydrogels with incorporation of hyaluronic acid and direct loading of TGF- β , *Acta Biomater.*, 2019, **83**, 167–176.
 - 93 S. Thönes, S. Rother, T. Wippold, J. Blaszkiewicz, K. Balamurugan and S. Moeller, *et al.*, Hyaluronan/collagen hydrogels containing sulfated hyaluronan improve wound healing by sustained release of heparin-binding EGF-like growth factor, *Acta Biomater.*, 2019, **86**, 135–147.
 - 94 H. Mao, L. Yang, H. Zhu, L. Wu, P. Ji, J. Yang and Z. Gu, Recent advances and challenges in materials for 3D bioprinting, *Prog. Nat. Sci.: Mater. Int.*, 2020, **30**(5), 618–634.
 - 95 M. M. Pillai, H. N. Tran, G. Sathishkumar, K. Manimekalai, J. Yoon and D. Lim, *et al.*, Symbiotic culture of nanocellulose pellicle: A potential matrix for 3D bioprinting, *Mater. Sci. Eng., C*, 2021, **119**, 111552.
 - 96 J. Y. Shin, Y. H. Yeo, J. E. Jeong, S. A. Park and W. H. Park, Dual-crosslinked methylcellulose hydrogels for 3D bioprinting applications, *Carbohydr. Polym.*, 2020, **238**, 116192.
 - 97 N. Reddy and Y. Yang, Citric acid cross-linking of starch films, *Food Chem.*, 2010, **118**(3), 702–711.
 - 98 M. I. Rial-Hermida, A. Rey-Rico, B. Blanco-Fernandez, N. Carballo-Pedrares, E. M. Byrne and J. F. Mano, Recent progress on polysaccharide-based hydrogels for controlled delivery of therapeutic biomolecules, *ACS Biomater. Sci. Eng.*, 2021, **7**(9), 4102–4127.
 - 99 W. Boonlai, V. Tantishaiyakul and N. Hirun, Characterization of κ -carrageenan/methylcellulose/cellulose nanocrystal hydrogels for 3D bioprinting, *Polym. Int.*, 2022, **71**(2), 181–191.
 - 100 S. G. Lévesque, R. M. Lim and M. S. Shoichet, Macroporous interconnected dextran scaffolds of controlled porosity for tissue-engineering applications, *Biomaterials*, 2005, **26**(35), 7436–7446.

- 101 L. Pescosolido, W. Schuurman, J. Malda, P. Matricardi, F. Alhaique and T. Coviello, *et al.*, Hyaluronic acid and dextran-based semi-IPN hydrogels as biomaterials for bioprinting, *Biomacromolecules*, 2011, **12**(5), 1831–1838.
- 102 A. Brunsen, U. Ritz, A. Mateescu, I. Hoefer, P. Frank and B. Menges, *et al.*, Photocrosslinkable dextran hydrogel films as substrates for osteoblast and endothelial cell growth, *J. Mater. Chem.*, 2012, **22**(37), 19590–19604.
- 103 G. Peng, J. Wang, F. Yang, S. Zhang, J. Hou and W. Xing, *et al.*, In situ formation of biodegradable dextran-based hydrogel via Michael addition, *J. Appl. Polym. Sci.*, 2013, **127**(1), 577–584.
- 104 Z. Wei and S. Gerecht, A self-healing hydrogel as an injectable instructive carrier for cellular morphogenesis, *Biomaterials*, 2018, **185**, 86–96.
- 105 P. R. Turner, E. Murray, C. J. McAdam, M. A. McConnell and J. D. Cabral, Peptide chitosan/dextran core/shell vascularized 3D constructs for wound healing, *ACS Appl. Mater. Interfaces*, 2020, **12**(29), 32328–32339.
- 106 P. Datta, B. Ayan and I. T. Ozbolat, Bioprinting for vascular and vascularized tissue biofabrication, *Acta Biomater.*, 2017, **51**, 1–20.
- 107 R. Levato, J. Visser, J. A. Planell, E. Engel, J. Malda and M. A. Mateos-Timoneda, Biofabrication of tissue constructs by 3D bioprinting of cell-laden microcarriers, *Biofabrication*, 2014, **6**(3), 035020.
- 108 R. Mohammadinejad, A. Kumar, M. Ranjbar-Mohammadi, M. Ashrafzadeh, S. S. Han, G. Khang and Z. Roveimiab, Recent advances in natural gum-based biomaterials for tissue engineering and regenerative medicine: A review, *Polymer*, 2020, **12**(1), 176.
- 109 Y. Niu, T. Yang, R. Ke and C. Wang, Preparation and characterization of pH-responsive sodium alginate/humic acid/konjac hydrogel for L-ascorbic acid controlled release, *Mater. Express*, 2019, **9**(6), 563–569.
- 110 L. Zhang, T. Zheng, L. Wu, Q. Han, S. Chen and Y. Kong, *et al.*, Fabrication and characterization of 3D-printed gellan gum/starch composite scaffold for Schwann cells growth, *Nanotechnol. Rev.*, 2021, **10**(1), 50–61.
- 111 P. Zarrintaj, S. Manouchehri, Z. Ahmadi, M. R. Saeb, A. M. Urbanska, D. L. Kaplan and M. Mozafari, Agarose-based biomaterials for tissue engineering, *Carbohydr. Polym.*, 2018, **187**, 66–84.
- 112 L. E. Bertassoni, J. C. Cardoso, V. Manoharan, A. L. Cristino, N. S. Bhise and W. A. Araujo, *et al.*, Direct-write bioprinting of cell-laden methacrylated gelatin hydrogels, *Biofabrication*, 2014, **6**(2), 024105.
- 113 S. H. Moon, H. N. Choi and Y. J. Yang, Natural/synthetic polymer materials for bioink development, *Biotechnol. Bioprocess Eng.*, 2022, **27**(4), 482–493.
- 114 B. Pokusaev, A. Vyazmin, N. Zakharov, S. Karlov, D. Nekrasov, V. Reznik and D. Khramtsov, Thermokinetics and rheology of agarose gel applied to bioprinting technology, *Therm. Sci.*, 2020, **24**(1 Part A), 347–353.
- 115 L. Aydin, S. Kucuk and H. Kenar, A universal self-eroding sacrificial bioink that enables bioprinting at room temperature, *Polym. Adv. Technol.*, 2020, **31**(7), 1634–1647.
- 116 A. Alves, S. P. Miguel, A. R. Araujo, M. J. de Jesús Valle, A. Sánchez Navarro and I. J. Correia, *et al.* Xanthan gum–Konjac glucomannan blend hydrogel for wound healing, *Polymer*, 2020, **12**(1), 99.
- 117 D. Yang, Y. Yuan, L. Wang, X. Wang, R. Mu and J. Pang, *et al.*, A review on konjac glucomannan gels: Microstructure and application, *Int. J. Mol. Sci.*, 2017, **18**(11), 2250.
- 118 L. Yang, Q. Zhao, Z. Guo, Y. Liu, W. Gao, Y. Pu and B. He, Konjac glucomannan hydrogel dressing and its combination with Chinese medicine for the wound treatment, *New J. Chem.*, 2022, **46**(48), 23077–23087.
- 119 X. Pan, Q. Wang, D. Ning, L. Dai, K. Liu and Y. Ni, *et al.*, Ultraflexible self-healing guar gum-glycerol hydrogel with injectable, antifreeze, and strain-sensitive properties, *ACS Biomater. Sci. Eng.*, 2018, **4**(9), 3397–3404.
- 120 A. Indurkar, P. Bangde, M. Gore, P. Reddy, R. Jain and P. Dandekar, Optimization of guar gum-gelatin bioink for 3D printing of mammalian cells, *Bioprinting*, 2020, **20**, e00101.
- 121 E. Bakirci, B. Toprakhisar, M. C. Zeybek, G. Ince and B. Koç, Cell sheet based bioink for 3D bioprinting applications, *Biofabrication*, 2017, **9**(2), 024105.
- 122 H. Baniyadi, E. Kimiaei, R. T. Polez, R. Ajdary, O. J. Rojas, M. Österberg and J. Seppälä, High-resolution 3D printing of xanthan gum/nanocellulose bio-inks, *Int. J. Biol. Macromol.*, 2022, **209**, 2020–2031.
- 123 B. Piola, M. Sabbatini, S. Gino, M. Invernizzi and F. Renò, 3D bioprinting of gelatin–xanthan gum composite hydrogels for growth of human skin cells, *Int. J. Mol. Sci.*, 2022, **23**(1), 539.
- 124 S. Muthusamy, S. Kannan, M. Lee, V. Sanjairaj, W. F. Lu and J. Y. Fuh, *et al.*, 3D bioprinting and microscale organization of vascularized tissue constructs using collagen-based bioink, *Biotechnol. Bioeng.*, 2021, **118**(8), 3150–3163.
- 125 W. Lim, S. Y. Shin, J. M. Cha and H. Bae, Optimization of polysaccharide hydrocolloid for the development of bioink with high printability/biocompatibility for Coextrusion 3D bioprinting, *Polymer*, 2021, **13**(11), 1773.
- 126 S. Sang, X. Mao, Y. Cao, Z. Liu, Z. Shen and M. Li, *et al.*, 3D Bioprinting using synovium-derived MSC-laden photo-cross-linked ECM bioink for cartilage regeneration, *ACS Appl. Mater. Interfaces*, 2023, **15**(7), 8895–8913.
- 127 S. Nedunchezian, P. Banerjee, C.-Y. Lee, S.-S. Lee, C.-W. Lin and C.-W. Wu, *et al.*, Generating adipose stem cell-laden hyaluronic acid-based scaffolds using 3D bioprinting via the double crosslinked strategy for chondrogenesis, *Mater. Sci. Eng., C*, 2021, **124**, 112072.
- 128 M. A. Bonifacio, S. Cometa, A. Cochis, A. Scalzone, P. Gentile and A. C. Scalia, *et al.*, A bioprintable gellan gum/lignin hydrogel: a smart and sustainable route for cartilage regeneration, *Int. J. Biol. Macromol.*, 2022, **216**, 336–346.
- 129 A. R. Akkineni, B. S. Elci, A. Lode and M. Gelinsky, Addition of high acyl gellan gum to low acyl gellan gum enables the blends 3d bioprintable, *Gels*, 2022, **8**(4), 199.

- 130 J. Zhang, E. Wehrle, P. Adamek, G. R. Paul, X.-H. Qin, M. Rubert and R. Müller, Optimization of mechanical stiffness and cell density of 3D bioprinted cell-laden scaffolds improves extracellular matrix mineralization and cellular organization for bone tissue engineering, *Acta Biomater.*, 2020, **114**, 307–322.
- 131 D. Wu, Y. Yu, J. Tan, L. Huang, B. Luo, L. Lu and C. Zhou, 3D bioprinting of gellan gum and poly(ethylene glycol) diacrylate based hydrogels to produce human-scale constructs with high-fidelity, *Mater. Des.*, 2018, **160**, 486–495.
- 132 J. Lee, J. Hong, W. Kim and G. H. Kim, Bone-derived dECM/alginate bioink for fabricating a 3D cell-laden mesh structure for bone tissue engineering, *Carbohydr. Polym.*, 2020, **250**, 116914.
- 133 Z. Li, S. Li, J. Yang, Y. Ha, Q. Zhang, X. Zhou and C. He, 3D bioprinted gelatin/gellan gum-based scaffold with double-crosslinking network for vascularized bone regeneration, *Carbohydr. Polym.*, 2022, **290**, 119469.
- 134 M. Puertas-Bartolomé, M. K. Włodarczyk-Biegun, A. Del Campo, B. Vázquez-Lasa and J. San Román, Development of bioactive catechol functionalized nanoparticles applicable for 3D bioprinting, *Mater. Sci. Eng., C*, 2021, **131**, 112515.
- 135 H. Rastin, M. Ramezanpour, K. Hassan, A. Mazinani, T. T. Tung, S. Vreugde and D. Losic, 3D bioprinting of a cell-laden antibacterial polysaccharide hydrogel composite, *Carbohydr. Polym.*, 2021, **264**, 117989.
- 136 R. F. Pereira, A. Sousa, C. C. Barrias, P. J. Bártolo and P. L. Granja, A single-component hydrogel bioink for bioprinting of bioengineered 3D constructs for dermal tissue engineering, *Mater. Horiz.*, 2018, **5**(6), 1100–1111.
- 137 Z. Wu, Q. Li, S. Xie, X. Shan and Z. Cai, In vitro and in vivo biocompatibility evaluation of a 3D bioprinted gelatin-sodium alginate/rat Schwann-cell scaffold, *Mater. Sci. Eng., C*, 2020, **109**, 110530.
- 138 L. Ning, H. Sun, T. Lelong, R. Guilloteau, N. Zhu, D. J. Schreyer and X. Chen, 3D bioprinting of scaffolds with living Schwann cells for potential nerve tissue engineering applications, *Biofabrication*, 2018, **10**(3), 035014.
- 139 Z. Wu, S. Xie, Y. Kang, X. Shan, Q. Li and Z. Cai, Biocompatibility evaluation of a 3D-bioprinted alginate-GelMA-bacteria nanocellulose(BNC) scaffold laden with oriented-growth RSC96 cells, *Mater. Sci. Eng., C*, 2021, **129**, 112393.
- 140 A. P. Haring, E. G. Thompson, Y. Tong, S. Laheri, E. Cesewski, H. Sontheimer and B. N. Johnson, Process-and bio-inspired hydrogels for 3D bioprinting of soft free-standing neural and glial tissues, *Biofabrication*, 2019, **11**(2), 025009.
- 141 N. Li, R. Guo and Z. J. Zhang, Bioink formulations for bone tissue regeneration, *Front. bioeng. biotechnol.*, 2021, **9**, 630488.
- 142 S. Taheri, H. S. Ghazali, Z. S. Ghazali, A. Bhattacharyya and I. Noh, Progress in biomechanical stimuli on the cell-encapsulated hydrogels for cartilage tissue regeneration, *Biomater. Res.*, 2023, **27**(1), 1–17.
- 143 C. Qi, J. Liu, Y. Jin, L. Xu, G. Wang, Z. Wang and L. Wang, Photo-crosslinkable, injectable sericin hydrogel as 3D biomimetic extracellular matrix for minimally invasive repairing cartilage, *Biomaterials*, 2018, **163**, 89–104.
- 144 N. Liu, X. Zhang, Q. Guo, T. Wu and Y. Wang, 3D bioprinted scaffolds for tissue repair and regeneration, *Front. Mater.*, 2022, **9**, 925321.
- 145 M. Li, D. Sun, J. Zhang, Y. Wang, Q. Wei and Y. Wang, Application and development of 3D bioprinting in cartilage tissue engineering, *Biomater. Sci.*, 2022, **10**(19), 5430–5458.
- 146 S. Murab, A. Gupta, M. K. Włodarczyk-Biegun, A. Kumar, P. van Rijn and P. Whitlock, *et al.* Alginate based hydrogel inks for 3D bioprinting of engineered orthopedic tissues, *Carbohydr. Polym.*, 2022, 119964.
- 147 A. Bhattacharyya, G. Janarthanan, H. N. Tran, H. J. Ham, J. Yoon and I. Noh, Bioink homogeneity control during 3D bioprinting of multicomponent micro/nanocomposite hydrogel for even tissue regeneration using novel twin screw extrusion system, *Chem. Eng. J.*, 2021, **415**, 128971.
- 148 S. Chae, S.-S. Lee, Y.-J. Choi, G. Gao, J. H. Wang and D.-W. Cho, 3D cell-printing of biocompatible and functional meniscus constructs using meniscus-derived bioink, *Biomaterials*, 2021, **267**, 120466.
- 149 M. Govindharaj, N. Al Hashimi, S. S. Soman, S. Kanwar and S. Vijayavenkataraman, 3D Bioprinting of human Mesenchymal Stem Cells in a novel tunic decellularized ECM bioink for Cartilage Tissue Engineering, *Materialia*, 2022, **23**, 101457.
- 150 A. Sadeghianmaryan, S. Naghieh, H. A. Sardroud, Z. Yazdanpanah, Y. A. Soltani, J. Sernaglia and X. Chen, Extrusion-based printing of chitosan scaffolds and their in vitro characterization for cartilage tissue engineering, *Int. J. Biol. Macromol.*, 2020, **164**, 3179–3192.
- 151 Y. He, S. Derakhshanfar, W. Zhong, B. Li, F. Lu, M. Xing and X. Li, Characterization and application of carboxymethyl chitosan-based bioink in cartilage tissue engineering, *J. Nanomater.*, 2020, **2020**, 1–11.
- 152 X. Liu, S. Song, J. Huang, H. Fu, X. Ning, Y. He and Z. Zhang, HBC-nanofiber hydrogel scaffolds with 3D printed internal microchannels for enhanced cartilage differentiation, *J. Mater. Chem. B*, 2020, **8**(28), 6115–6127.
- 153 C. Antich, J. de Vicente, G. Jiménez, C. Chocarro, E. Carrillo and E. Montañez, *et al.* Bio-inspired hydrogel composed of hyaluronic acid and alginate as a potential bioink for 3D bioprinting of articular cartilage engineering constructs, *Acta Biomater.*, 2020, **106**, 114–123.
- 154 E. De Angelis, R. Saleri, P. Martelli, L. Elviri, A. Bianchera and C. Bergonzi, *et al.* Cultured horse articular chondrocytes in 3d-printed chitosan scaffold with hyaluronic acid and platelet lysate, *Front. Vet. Sci.*, 2021, **8**, 671776.
- 155 F. Olate-Moya, L. Arens, M. Wilhelm, M. A. Mateos-Timoneda, E. Engel and H. Palza, Chondroinductive alginate-based hydrogels having graphene oxide for 3D printed scaffold fabrication, *ACS Appl. Mater. Interfaces*, 2020, **12**(4), 4343–4357.

- 156 K. Elkhoury, M. Morsink, L. Sanchez-Gonzalez, C. Kahn, A. Tamayol and E. Arab-Tehrany, Biofabrication of natural hydrogels for cardiac, neural, and bone Tissue engineering Applications, *Bioact. Mater.*, 2021, **6**(11), 3904–3923.
- 157 M. Ojansivu, A. Rashad, A. Ahlinder, J. Massera, A. Mishra and K. Syverud, *et al.*, Wood-based nanocellulose and bioactive glass modified gelatin–alginate bioinks for 3D bioprinting of bone cells, *Biofabrication*, 2019, **11**(3), 035010.
- 158 F. Curti, A. Serafim, E. Olaret, S. Dinescu, I. Samoila and B. S. Vasile, *et al.*, Development of Biocomposite Alginate–Cuttlebone–Gelatin 3D Printing Inks Designed for Scaffolds with Bone Regeneration Potential, *Mar. Drugs*, 2022, **20**(11), 670.
- 159 S. Im, G. Choe, J. M. Seok, S. J. Yeo, J. H. Lee and W. D. Kim, *et al.*, An osteogenic bioink composed of alginate, cellulose nanofibrils, and polydopamine nanoparticles for 3D bioprinting and bone tissue engineering, *Int. J. Biol. Macromol.*, 2022, **205**, 520–529.
- 160 H. K. Chang, D. H. Yang, M. Y. Ha, H. J. Kim, C. H. Kim and S. H. Kim, *et al.*, 3D printing of cell-laden visible light curable glycol chitosan bioink for bone tissue engineering, *Carbohydr. Polym.*, 2022, **287**, 119328.
- 161 C. Gao, C. Lu, Z. Jian, T. Zhang, Z. Chen and Q. Zhu, *et al.*, 3D bioprinting for fabricating artificial skin tissue, *Colloids Surf., B*, 2021, **208**, 112041.
- 162 R. Augustine, Skin bioprinting: a novel approach for creating artificial skin from synthetic and natural building blocks, *Prog. Biomater.*, 2018, **7**(2), 77–92.
- 163 P. Liu, H. Shen, Y. Zhi, J. Si, J. Shi, L. Guo and S. G. Shen, 3D bioprinting and in vitro study of bilayered membranous construct with human cells-laden alginate/gelatin composite hydrogels, *Colloids Surf., B*, 2019, **181**, 1026–1034.
- 164 H. Zhao, J. Xu, H. Yuan, E. Zhang, N. Dai and Z. Gao, *et al.*, 3D printing of artificial skin patches with bioactive and optically active polymer materials for anti-infection and augmenting wound repair, *Mater. Horiz.*, 2022, **9**(1), 342–349.
- 165 F. Zhou, Y. Hong, R. Liang, X. Zhang, Y. Liao and D. Jiang, *et al.*, Rapid printing of bio-inspired 3D tissue constructs for skin regeneration, *Biomaterials*, 2020, **258**, 120287.
- 166 R. Poongodi, Y.-L. Chen, T.-H. Yang, Y.-H. Huang, K. D. Yang, H.-C. Lin and J.-K. Cheng, Bio-scaffolds as cell or exosome carriers for nerve injury repair, *Int. J. Mol. Sci.*, 2021, **22**(24), 13347.
- 167 P. Zhuang, A. X. Sun, J. An, C. K. Chua and S. Y. Chew, 3D neural tissue models: From spheroids to bioprinting, *Biomaterials*, 2018, **154**, 113–133.
- 168 M. Habibizadeh, S. Nadri, A. Fattahi, K. Rostamizadeh, P. Mohammadi and S. Andalib, *et al.*, Surface modification of neurotrophin-3 loaded PCL/chitosan nanofiber/net by alginate hydrogel microlayer for enhanced biocompatibility in neural tissue engineering, *J. Biomed. Mater. Res., Part A*, 2021, **109**(11), 2237–2254.
- 169 G. Zhou, Y. Chen, F. Dai and X. Yu, Chitosan-based nerve guidance conduit with microchannels and nanofibers promotes schwann cells migration and neurite growth, *Colloids Surf., B*, 2023, **221**, 112929.
- 170 S. Houshyar, A. Bhattacharyya and R. Shanks, Peripheral nerve conduit: materials and structures, *ACS Chem. Neurosci.*, 2019, **10**(8), 3349–3365.
- 171 Q. Lian, L. Zhou, X. Li, W. Mao and D. Li, Perfusive and osmotic capabilities of 3D printed hollow tube for fabricating large-scaled muscle scaffold, *Rapid Prototyp. J.*, 2020, **26**(1), 1–10.
- 172 S. Ostrovidov, S. Salehi, M. Costantini, K. Suthiwanich, M. Ebrahimi and R. B. Sadeghian, *et al.*, 3D bioprinting in skeletal muscle tissue engineering, *Small*, 2019, **15**(24), 1805530.
- 173 J. Lee, H. Lee, E.-J. Jin, D. Ryu and G. H. Kim, 3D bioprinting using a new photo-crosslinking method for muscle tissue restoration, *npj Regener. Med.*, 2023, **8**(1), 18.
- 174 F. L. Ronzoni, F. Aliberti, F. Scocozza, L. Benedetti, F. Auricchio and M. Sampaulesi, *et al.*, Myoblast 3D bioprinting to burst in vitro skeletal muscle differentiation, *J. Tissue Eng. Regener. Med.*, 2022, **16**(5), 484–495.
- 175 A. Bhattacharyya, G. Janarthanan and I. Noh, Nano-biomaterials for designing functional bioinks towards complex tissue and organ regeneration in 3D bioprinting, *Addit. Manuf.*, 2021, **37**, 101639.
- 176 H.-W. Kang, S. J. Lee, I. K. Ko, C. Kengla, J. J. Yoo and A. Atala, A 3D bioprinting system to produce human-scale tissue constructs with structural integrity, *Nat. Biotechnol.*, 2016, **34**(3), 312–319.
- 177 G. Cattelan, A. Guerrero Gerbolés, R. Foresti, P. P. Pramstaller, A. Rossini, M. Miragoli and M. C. Caffarra, Alginate formulations: current developments in the race for hydrogel-based cardiac regeneration, *Front. bioeng. biotechnol.*, 2020, **8**, 414.
- 178 Y. S. Zhang, A. Arneri, S. Bersini, S.-R. Shin, K. Zhu and Z. Goli-Malekabadi, *et al.*, Bioprinting 3D microfibrous scaffolds for engineering endothelialized myocardium and heart-on-a-chip, *Biomaterials*, 2016, **110**, 45–59.
- 179 L. Hockaday, K. Kang, N. Colangelo, P. Cheung, B. Duan and E. Malone, *et al.*, Rapid 3D printing of anatomically accurate and mechanically heterogeneous aortic valve hydrogel scaffolds, *Biofabrication*, 2012, **4**(3), 035005.
- 180 Y. Wu, A. Wenger, H. Golzar and X. Tang, 3D bioprinting of bicellular liver lobule-mimetic structures via microextrusion of cellulose nanocrystal-incorporated shear-thinning bioink, *Sci. Rep.*, 2020, **10**(1), 20648.
- 181 R. Taymour, D. Kilian, T. Ahlfeld, M. Gelinsky and A. Lode, 3D bioprinting of hepatocytes: Core-shell structured co-cultures with fibroblasts for enhanced functionality, *Sci. Rep.*, 2021, **11**(1), 5130.
- 182 P. Wang, X. Li, W. Zhu, Z. Zhong, A. Moran and W. Wang, *et al.*, 3D bioprinting of hydrogels for retina cell culturing, *Bioprinting*, 2018, **12**, e00029.
- 183 M. P. Sekar, S. Suresh, A. Zennifer, S. Sethuraman and D. Sundaramurthi, Hyaluronic Acid as Bioink and Hydrogel Scaffolds for Tissue Engineering Applications, *ACS Biomater. Sci. Eng.*, 2023, **9**(6), 3134–3159.