

Dalton Transactions

An international journal of inorganic chemistry

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: J. Harland, A. B. LaLonde, D. Thomas, D. G. Castella, J. F. Kampf, M. Zeller, E. E. Alp, M. Hu, J. Zhao and N. Lehnert, *Dalton Trans.*, 2024, DOI: 10.1039/D4DT01632G.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.

The crystallographic data have been deposited in the Cambridge Crystallographic Data Centre, and the corresponding CCDC entry numbers are provided in the manuscript. UV-Vis, NMR and IR spectroscopic data are available from the corresponding author upon request.

Open Access Article. Published on 04 July 2024. Downloaded on 27/07/2024 7:26:47 PM.
This article is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported Licence.



Vibrational Properties of Heme-Nitrosoalkane Complexes in Comparison to their HNO Analogs, and Reactivity Studies towards Nitric Oxide and Lewis Acids

Jill B. Harland,[†] Ashley B. LaLonde,[†] Diamond J. Thomas,[†] Daniel G. Castella,[†] Jeff W. Kampf,[†] Matthias Zeller,[‡] E. Ercan Alp,[§] Michael Y. Hu,[§] Jiyong Zhao,[§] and Nicolai Lehnert^{*†}

[†] Department of Chemistry and Department of Biophysics, University of Michigan, Ann Arbor, MI 48109

[‡] Department of Chemistry, Purdue University, 560 Oval Drive, West Lafayette, Indiana 47907

[§] Advanced Photon Source, Argonne National Laboratory, Illinois 60439

^{*} Corresponding author: lehnert@umich.edu



Abstract

C-nitroso compounds (RNO, R = alkyl, aryl) are byproducts of drug metabolism, bind to heme proteins, and heme-RNO analogs are isoelectronic to ferrous nitroxyl (NO^-/HNO) complexes. Importantly, heme-HNO compounds are key intermediates in the reduction of NO to N_2O and nitrite to ammonium in the nitrogen cycle. Ferrous heme-RNO complexes act as stable analogs of these species, potentially allowing for the investigation of the vibrational and electronic properties of heme-HNO intermediates. In this paper, a series of six-coordinate ferrous heme-RNO complexes (where R=iPr, Ph) were prepared using the TPP^{2-} and 3,5-Me-BAFP $^{2-}$ coligands, and tetrahydrofuran, pyridine, and 1-methylimidazole as the axial ligands (bound trans to RNO). These complexes were characterized using different spectroscopic methods and X-ray crystallography. The complex $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$ was further utilized for nuclear resonance vibrational spectroscopy (NRVS), allowing for the detailed assignment of the Fe-N(R)O vibrations of a heme-RNO complex for the first time. The vibrational properties of these species were then correlated with those of their HNO analogs, using DFT calculations. Our studies support previous findings that RNO ligands in ferrous heme complexes do not elicit a significant *trans* effect. In addition, the complexes are air-stable, and do not show any reactivity towards NO. So although ferrous heme-RNO complexes are suitable structural and electronic models for their HNO analogs, they are unsuitable to model the reactivity of heme-HNO complexes. We further investigated the reaction of our heme-RNO complexes with different Lewis acids. Here, $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$ was found to be unreactive towards Lewis acids. In contrast, $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ is reactive towards all of the Lewis acids investigated here, but in most cases the iron center is simply oxidized, resulting in loss of the iPrNO ligand. In the case of the Lewis acid $\text{B}_2(\text{pin})_2$, the reduced product $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNH}_2)(\text{iPrNO})]$ was identified by X-ray crystallography.



Introduction

The oxidative metabolism of drugs (or xenobiotics) containing amine or hydroxylamine functional groups produces *C*-nitroso compounds (RNO, R = alkyl, aryl) as byproducts:

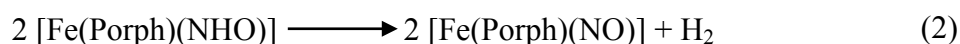


These harmful species can also be formed through reduction of nitroalkanes/arenes (RNO₂).¹⁻⁴ *C*-nitroso compounds bind to heme proteins, including myoglobin/hemoglobin (Mb/Hb), Cytochrome (Cyt.) P450s, and soluble guanylate cyclase (sGC), by coordination to the ferrous heme, thereby inhibiting function. This inhibition contributes to the general toxicity of nitro-containing compounds to cells, since they can be reduced to *C*-nitroso derivatives.⁵ Interestingly, thiols like glutathione in red blood cells react with nitrosoarenes to form sulfonamides, before they are metabolized in the liver, which actually constitutes a defense mechanism against these toxic molecules.⁶ In addition, RNOs react with cysteine residues in proteins.⁷⁻¹¹

Ferrous heme-RNO complexes are isoelectronic to ferrous nitroxyl (NO⁻/HNO) complexes, or *ls*-{FeN(H)O}⁸ species in the Enemark-Feltham notation (the superscript “8” represents the number of iron(d) plus NO(π*) electrons; *ls* = low-spin).¹² In the denitrification and DNRA pathways in the nitrogen cycle, heme-nitroxyl compounds are proposed as important intermediates in the reduction of NO to N₂O and nitrite to ammonium, respectively.¹ For example, it has been proposed based on quantum-chemical calculations that in the catalytic cycle of Cyt. P450_{nor} (fungal NO reductase), a ferrous heme-HNO intermediate is formed (termed “Intermediate I”) that subsequently reacts with NO directly under N-N bond formation, producing a ferric hyponitrite species in the central step of Cyt. P450_{nor} catalysis (see Scheme 1).¹³⁻¹⁵ Recently, experimental data have been obtained that further support these claims.¹⁶⁻¹⁹



Several heme-HNO adducts, in proteins and model complexes, have been reported in the literature. Farmer and coworkers prepared a heme-HNO complex in Mb, which was characterized in depth using different spectroscopic methods.^{20, 21} Computational work on Mb(II)-NHO shows that hydrogen bonds formed in the distal pocket between the coordinated HNO, His64 and a H₂O molecule likely aid in stabilizing the complex.²² Until recently, there were no heme model complexes available that could generate a stable ls-{FeNHO}⁸ species, allowing for further reactivity studies²³. This is due to disproportionation of the bound HNO ligand in simple [Fe(Porph)(NHO)] (Porph²⁻ = generic porphyrin dianion) complexes, as first reported by Ryan and coworkers.²⁴ This yields H₂ and the corresponding ls-{FeNO}⁷ complex:



Recently, Ryan's group was able to generate the complex [Fe(OEP)(NHO)] (OEP²⁻ = octaethylporphyrin dianion) upon reacting the corresponding nitroxyl complex [Fe(OEP)(NO)]⁻ with phenols (PhOH) as weak acids. Here, the [Fe(OEP)(NHO)] complex seems to form an equilibrium with the hydrogen-bonded complex [Fe(OEP)(NO---HOPh)]⁻.²⁵ Richter-Addo and coworkers were able to model the key hydride transfer step in the Cyt. P450_{nor} mechanism.²⁶ Direct hydride transfer from [NBu₄](BH₄) to the six-coordinate (6C) ls-{FeNO}⁶ model complex [Fe(OEP)(5-MI)(NO)]⁺ (5-MI = 5-methylimidazole) to form the corresponding 6C HNO complex, [Fe(OEP)(5-MI)(NHO)], was demonstrated.²⁶ Formation of this species was monitored by solution IR spectroscopy at -20 °C. This assignment was further confirmed by ¹H-NMR spectroscopy in conjunction with ¹⁵NO isotope labeling. Even at low temperature, this Fe^{II}-NHO complex disproportionates, forming the corresponding five-coordinate (5C) ls-{FeNO}⁷ complex [Fe(OEP)(NO)].²⁶ The total yield of [Fe(OEP)(5-MI)(NHO)] was only 11%, which precluded further characterization of this species. In a follow up study by the Richter-Addo group, it was



shown that the addition of NO to [Fe(OEP)(5-MI)(NHO)] leads to the generation of N₂O.¹⁹ In summary, the fast disproportionation of ferrous heme-HNO complexes in organic solvents has been a major obstacle for the preparation of these types of species.^{24, 27, 28} In contrast, the corresponding, deprotonated heme ls-{FeNO}⁸ complexes are stable.^{27, 29-31}

Recently, we prepared a long-lived 6C ls-{FeNHO}⁸ heme complex, [Fe(3,5-Me-BAFP)(MI)(NHO)], that is stabilized by steric bulk around the iron center, utilizing a bis-picket fence porphyrin co-ligand (3,5-Me-BAFP²⁻). In this case, the picket fence greatly slows down the disproportionation reaction of the bound HNO ligand. This complex has a half-life of 56 min at -30 °C, and was fully characterized by UV-Vis, ¹H- and ¹⁵N-NMR, IR, and nuclear resonance vibrational spectroscopy (NRVS). Upon reaction with NO, this complex produces quantitative amounts of N₂O.¹⁷ This result indicates that the initially formed HNO complex in the catalytic cycle of Cyt. P450nor (see Scheme 1) is catalytically competent and could react with the second equivalent of NO under N-N bond formation. Hence, Intermediate I could actually correspond to the heme-HNO complex.¹

We rationalized that we might obtain further insight into the vibrational and electronic properties of ferrous heme-HNO complexes and their reactivity by studying the much more stable ferrous heme-RNO analogs, as they have similar geometric and electronic structures.²² In this regard, note that a decent number of ferrous heme-RNO complexes with R = iPr, Ph, and Ph derivatives have been reported previously and characterized using different spectroscopic methods (IR, NMR, Raman, UV-Vis) and X-ray crystallography, starting with pioneering studies by Mansuy and coworkers in the 1970s, and 1980s.^{32, 33} Reviews are further available that provide a summary of the previous studies.³⁴⁻³⁸



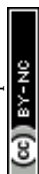
In this work, we have prepared a series of six-coordinate, ferrous heme-RNO complexes (where R = iPr, Ph) with the TPP²⁻ and 3,5-Me-BAFP²⁻ co-ligands, with the goal to (a) interrogate the vibrational properties of ferrous heme-RNO complexes, especially the low-energy Fe-N(R)O stretching and Fe-N-O bending modes, and (b) to study the reactivity of these complexes with NO (following the mechanism of Cyt. P450_{nor}) and Lewis acids. The latter investigation was inspired by previous work by the Richter-Addo group. In their study, they found that when [Fe(OEP)(NO)] was reacted with several Lewis acids (BF₃, B(C₆F₅)₃ and K⁺), the Lewis acid coordinated to the nitrosyl O atom, activating the NO ligand toward N-N bond formation to generate N₂O.³⁹ The 6C ferrous heme-RNO complexes prepared here were fully characterized using UV-Vis, IR, and ¹H-NMR spectroscopy and X-ray crystallography. Although crystal structures of ferrous heme-RNO complexes have been reported before (see Results & Discussion), X-Ray crystallographyThe weakness was that it was not entirely clear what was the aim of the work was used here to affirm the exact identity of our complexes, for example with respect to axial ligation (ligands bound trans to the RNO group). Typical axial ligands used here are tetrahydrofuran, pyridine, and 1-methylimidazole. Important results include the full vibrational characterization of the complex [Fe(TPP)(THF)(iPrNO)], using IR and nuclear resonance vibrational spectroscopy (NRVS), and the correlation of these results to the vibrational properties of the HNO analog, using DFT calculations. Interestingly, 6C ferrous heme-RNO complexes react with NO by simple displacement of the RNO ligand, but without undergoing N-N bond formation, in contrast to the HNO analogs. On the other hand, reactions of our heme-RNO complexes with different boron-based Lewis acids led merely to redox reactions, but no stable adduct formation was observed.

Experimental Procedures



All reactions were performed under inert conditions using Schlenk techniques. Preparation and handling of air sensitive materials was carried out under a dinitrogen atmosphere in an MBraun glovebox equipped with a circulating purifier (O_2 , H_2O < 0.1 ppm). All solvents (including deuterated solvents) and 1-methylimidazole (MI) were distilled from CaH_2 under dinitrogen, then degassed via five freeze-pump-thaw cycles. The purified solvents were stored over appropriately sized activated molecular sieves in the glovebox until used. Nitric oxide (Cryogenic Gases Inc., 99.5%) was purified by passage through an ascarite II column (NaOH on silica), followed by a cold trap at $-80^\circ C$ to remove higher order nitrogen oxide impurities. N-isopropylhydroxylamine hydrochloride was purchased from TCI America and used without further purification. N-phenylhydroxylamine, ^{15}N -aniline, and ^{15}N -sodium nitrite were purchased from Sigma Aldrich and used without any further purification. $[Fe(TPP)(Cl)]$,⁴⁰ $[Fe(TPP)(X)]$ (where $X = BF_4^-$, PF_6^- , and SbF_6^-),⁴¹ $[Fe(TPP)]$,²⁷ $[Fe(TPP)(PhNO)_2]$,⁴² $[Fe(3,5-Me-BAFP)(Cl)]$,²⁷ $[Fe(3,5-Me-BAFP)]$,²⁷ and $[Fe(3,5-Me-BAFP)(NO)]$ ²⁷ were synthesized as previously reported. Analogous ^{57}Fe complexes were synthesized in a similar manner using $^{57}FeCl_2$ dimethanol salt as the iron source.

Physical measurements. Infrared spectra were obtained from KBr disks on Perkin-Elmer BX or GX spectrometers at room temperature. Solution IR spectra were recorded on a Bruker Alpha-E FTIR spectrometer. Solution samples were measured in a thin-layer solution cell equipped with CaF_2 windows. Solid and gas-phase IR spectra were obtained on a Thermo-Nicolet IS-50 benchtop IR spectrometer. Gas samples were measured using a Pike HT gas cell (10 cm) with CaF_2 windows. Electronic absorption spectra were measured using an Analytical Jena Specord S600 instrument at room temperature. *In situ* UV-Vis measurements were conducted with a Hellma quartz immersion probe with a 10 mm path length. 1H and ^{13}C -NMR spectra were recorded on a Varian MR 400 MHz instrument or a Varian NMRS 500 MHz spectrometer at room temperature. The resonance



Raman (rRaman) experiments were performed using an INNOVA-301K* Krypton Ion gas laser system at an excitation wavelength of 407 nm with 20 mW power. Samples were transferred to a liquid nitrogen bath inside an EPR cold finger to cool the samples during the measurements. The scattered light from the samples was focused onto an Acton two-stage TriVista 555 monochromator and detected by a liquid N₂-cooled Princeton Instruments Spec-10:400B/LN CCD camera. The accumulation and exposure times for the samples were 5 acquisitions for 60 seconds. The spectral resolution was 0.3 cm⁻¹. Spectra were then plotted and processed using the OriginPro 9.0.0 (64-bit) software for baseline correction. Nuclear Resonance Vibrational spectroscopy (NRVS) measurements were carried out as previously described⁴³ at beamline 3-ID-XOR at the Advanced Photon Source (APS) at Argonne National Laboratory. This beamline provides about 2.5 x 10⁹ photons/sec in ~1 meV bandwidth (8 cm⁻¹) at 14.4125 keV in a 0.5 mm (vertical) x 0.5 mm (horizontal) spot. Samples were loaded into 4 x 7 x 1 mm copper cells. The final spectra represent averages of 4 scans. The program Phoenix was used to convert the NRVS raw data into the Vibrational Density of States (VDOS).^{44, 45}

Synthesis of Nitrosobenzene (PhNO): Nitrosobenzene was synthesized using a modified literature procedure.⁴⁶ A 4.6 mL portion of aniline (50.4 mmol) was stirred in a mixture of 15 mL methanol and 23 mL water in an ice bath. To this solution, 28 mL H₂O₂ (30% w/v) was slowly added and the reaction mixture was stirred for 10 minutes. Then, 0.76 g MoO₃ (5.28 mmol) was added followed by 5 mL KOH (1 M), which caused the solution to turn orange-brown in color. The solution was allowed to stir for 6 hours in an ice bath. When the reaction was complete the solid was filtered off and washed with water (15 mL) and cold methanol (15 mL) to give a light yellow solid. The light yellow solid was dissolved in 50 mL absolute ethanol and allowed to precipitate in the -33 °C freezer overnight. The following day the solution was vacuum filtered



through a frit to give a white solid. Yield: 4.23 g (82%). $^1\text{H-NMR}$ (CD_2Cl_2 , 400 MHz): δ = 7.621-7.906 (m, 5H). $^{13}\text{C-NMR}$: (CDCl_3 , 126 MHz) δ = 121.2 (s), 129.7 (s), 136.1 (s), 166.4 (s). IR $\nu(\text{N-O})$ = 1589 cm^{-1} . UV-Vis (CH_2Cl_2): 282, 304, 750 nm.

The corresponding ^{15}N -labeled nitrosobenzene compound was synthesized in the same way starting from ^{15}N -labeled aniline.

Synthesis of N-Isopropylhydroxylamine: 1.0 g N-isopropylhydroxylamine hydrochloride (9 mmol) and 0.694 g potassium methoxide (9.9 mmol) were dispersed in 30 mL dichloromethane and allowed to stir for several hours. The next day the white precipitate (KCl) was filtered off and the filtrate was dried via rotary evaporation to give a white solid. Yield: 0.686 g (100%) $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ : 3.379 – 3.412 (sep, 1H); 1.276, 1.292 (d, 6H); $^{13}\text{C-NMR}$ (CDCl_3 , 126 MHz): δ : 53.240 (s), 17.522 (s).

Synthesis of ^{15}N -Isopropylhydroxylamine: The first step of this reaction was carried out according to a previous report.⁴⁷ Reaction of 2-iodopropane and ^{15}N -sodium nitrite delivers a crude product that contains ^{15}N -2-nitropropane which, according to the literature, is purified via vacuum distillation to remove impurities. We used the crude product for the following reaction based on a modified patent procedure, as described next.⁴⁸ ~0.5 g of crude ^{15}N -iPrNO₂ (~5.6 mmol), 96 μL acetic acid (16.8 mmol), and 0.550 g Zn powder (8.4 mmol) were added to 100 mL 95% ethanol and allowed to stir for 3 hours at 0 °C. Then, the solvent was evaporated under vacuum to give a grey powder. The grey solid was washed with dichloromethane (to dissolve the product) and the filtrate was dried by rotary evaporation. Crude Yield: ~0.357g (~89%). The $^1\text{H-NMR}$ spectrum showed that a large amount of acetic acid was present in the solid, so the product was further purified. For this purpose, ~0.375 g of crude ^{15}N -Isopropylhydroxylamine (~5 mmol) and 0.351 g KOMe (5 mmol) were stirred in 10 mL dichloromethane for 2 hours. Then, the reaction mixture



was filtered through a cotton plug and the solvent was evaporated via rotary evaporation. The resulting oil was redissolved in 2 mL dichloromethane and layered with 48 mL hexanes and placed in the -33 °C freezer. The next day the solution was filtered through a frit to give a white needle-like microcrystalline solid. The product was dried under vacuum for a few hours before further use. ¹H-NMR (CDCl₃, 400 MHz): δ: 3.150-3.118 (sep, 1H); 1.044, 1.067 (dd, 6H).

Synthesis of 2-nitrosopropane (iPrNO): 2-nitrosopropane was synthesized according to a modified literature procedure.⁴⁹ Under an inert atmosphere, 0.10 g (1.3 mmol) N-isopropylhydroxylamine was dissolved in 10 mL of dry methylene chloride, and 0.404 g Ag₂CO₃ (1.46 mmol) was added, causing a black precipitate to form. Then, the reaction was brought outside the glovebox and stirred for one hour. The solution was filtered through a celite pad in a frit and the resulting filtrate was dried to an oil via rotary evaporation. The crude material was chromatographed on silica gel with CH₂Cl₂ as the eluent (in a 9 inch Pasteur pipet). The solvent was removed via rotary evaporation and the product was further dried under vacuum to give a white solid (the solid is assumed to be in the dimer form based on previous literature reports).^{50, 51} Yield: 32 mg (15%). ¹H-NMR (CDCl₃, 400 MHz): δ: 5.315-5.347 (sep, 1H); 1.35, 1.36 (d, 6H).

Synthesis of [Fe(TPP)(THF)(PhNO)]: There are two literature methods^{32, 52} reported to synthesize ferrous porphyrin RNO complexes: (1) reaction of a ferric porphyrin chloride complex with excess RNHOH, and (2) reaction of a ferrous porphyrin complex with excess of the RNO ligand. In this work, we synthesized our ferrous heme-RNO complexes using modified versions of these methods as described below to obtain the ferrous PhNO/Ph¹⁵NO complexes. For the natural abundance isotopes (n.a.i.) complex, a 132 mg (0.175 mmol) portion of [Fe(TPP)(BF₄)] (also works with the PF₆⁻ or SbF₆⁻ counterions) and 95 mg N-Phenylhydroxylamine (0.880 mmol) were dissolved in 5 mL tetrahydrofuran. The solution was allowed to stir in the glovebox overnight

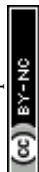


and the reaction progress was monitored by UV-Vis spectroscopy. The following day, the solution was layered with 40 mL hexanes and placed in the -33 °C glovebox freezer. The next day, the reaction mixture was vacuum filtered through a frit and washed with hexanes to give a dark purple powder. Yield: 120 mg (0.208 mmol, 89%). UV-Vis (CH₂Cl₂): 420, 507 nm. UV-Vis (THF): 420, 535 nm. ¹H-NMR (CD₂Cl₂, 400 MHz): δ = 8.58 (s); 8.014 (s); 7.72 (s); 6.339 – 6.509 (m); 6.014 (s). Anal. Calcd. for C₅₀H₃₃FeN₅O: C, 76.50; H, 4.87; N, 8.26. Found: C, 75.57; H, 4.59; N, 9.42. IR (KBr): ν(NO) = 1350, 1368 cm⁻¹.

Single crystals suitable for X-ray diffraction were grown by vapor diffusion of hexanes into a concentrated THF solution of [Fe(TPP)(THF)(PhNO)] in the -33 °C glovebox freezer.

Synthesis of [Fe(TPP)(THF)(Ph¹⁵NO)]: A 14 mg (0.021 mmol) portion of [Fe(TPP)] and 9 mg ¹⁵N-nitrosobenzene (0.083 mmol) were dissolved in 1 mL tetrahydrofuran and the resulting solution was allowed to stir in the glovebox. The reaction progress was monitored by UV-Vis spectroscopy. After one hour, the solution was layered with 15 mL hexanes and left in the -33 °C freezer. After 2 days, the solution was filtered through a frit to give a dark purple powder. Yield: 11 mg (0.0142 mmol, 68%). The UV-Vis data are identical to that of the n.a.i complex in THF: 420, 535 nm. ν(¹⁵NO) = 1328, 1348 cm⁻¹.

Synthesis of [Fe(TPP)(Pyr)(PhNO)]: First [Fe(TPP)(Pyr)₂] was synthesized by reacting [Fe(TPP)] (1 mmol) with excess pyridine (19 mmol) in methylene chloride. Under an inert atmosphere, 30 mg [Fe(TPP)(Pyr)₂] (1mmol) and 4 mg of PhNO were dissolved in 2 mL dry methylene chloride and stirred. The reaction mixture was then layered with 48 mL heptanes and placed in the -33 °C glovebox freezer. The following day, the reaction mixture was filtered through a frit and the resulting solid was washed with cold heptanes to give a dark purple solid.



Synthesis of [Fe(TPP)(MI)(PhNO)]: Under an inert atmosphere, 60 mg of [Fe(TPP)(MI)₂] (1 mmol) and 10 mg PhNO (1.2 mmol) was stirred overnight. The reaction mixture was then layered with 48 mL heptanes and placed in the -33 °C glovebox freezer. The following day, the reaction mixture was filtered through a frit and the resulting solid was washed with cold heptanes to give a quantitative yield. UV-Vis (CH₂Cl₂): 426, 540 nm.

Synthesis of [Fe(TPP)(THF)(iPrNO)]: Under an inert atmosphere, 100 mg [Fe(TPP)] (0.15 mmol) and 21 mg iPrNO (0.3 mmol) were dissolved in 2 mL dry tetrahydrofuran. The reaction was stirred for one hour, and then layered with 48 mL heptanes and placed in the -33 °C glovebox freezer. The following day, the reaction mixture was filtered through a frit and the resulting solid was washed with cold heptanes to give a dark purple solid. Yield: 81.7 mg (67%). UV-Vis (CH₂Cl₂): 415, 525 nm. UV-Vis (THF): 417, 535 nm. ¹H-NMR (CDCl₃, 400 MHz): δ = 8.731 (s); 8.108 (s); 7.704 (s); -0.61 (s); -2.065 (s). Anal. Calcd. for C₅₁H₄₃N₅O₂: C, 75.27; H, 5.33; N, 8.61. Found: C, 74.99; H, 5.11; N, 8.43.

Single crystals were grown by dissolving ~18 mg [Fe(TPP)(THF)(iPrNO)] in 0.2 mL THF and placing the solution in a 5 mm diameter glass tube. The solution was carefully layered with 2 mL hexanes, the tube was sealed with a septum, and left at room temperature. After 4 days crystals suitable for X-ray analysis were collected.

Synthesis of [Fe(TPP)(THF)(iPr¹⁵N)]: 70 mg [Fe(TPP)(SbF₆)] (0.077 mmol) was dissolved in 5 mL THF in the glovebox. To this solution, 29 mg (0.39 mmol) ¹⁵N-isopropylhydroxylamine was added and the reaction mixture was allowed to stir overnight. The reaction progress was monitored by UV-Vis spectroscopy. When the reaction was complete, the solution was layered with heptanes and placed in the glovebox freezer at -33 °C. The following day, the solution was filtered through a frit and the resulting solid was washed with cold heptanes to give a dark purple solid. Yield: 62



mg (100%). The UV-Vis and ^1H -NMR spectra are analogous to those of the n.a.i complex $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$.

The ^{57}Fe -labeled complexes, $[\text{}^{57}\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$ and $[\text{}^{57}\text{Fe}(\text{TPP})(\text{THF})(\text{iPr}^{15}\text{NO})]$, were synthesized using the same methods, except that the precursor complex $[\text{}^{57}\text{Fe}(\text{TPP})(\text{Cl})]$ was used.

Synthesis of $[\text{Fe}(\text{3,5-Me-BAFP})(\text{iPrNO})_2]$: Under an inert atmosphere, 30 mg $[\text{Fe}(\text{3,5-Me-BAFP})]$ (0.15 mmol) and 2.7 mg iPrNO (0.3 mmol) were dissolved in 2 mL dry tetrahydrofuran. The reaction was stirred for one hour, and then layered with 20 mL hexanes and placed in the -33°C glovebox freezer. The following day, the reaction mixture was filtered through a frit and the resulting solid was washed with cold hexanes to give a dark purple solid. Yield: 13.1 mg (41%). UV-Vis (THF): 425 nm. IR: N-O stretch 1459 cm^{-1} . EA calculated based on crystal structure with 20% THF and 80% iPrNO as the 6th ligand: $[\text{Fe}(\text{3,5-Me-BAFP})(\text{iPrNO})_2]$ Anal calcd for $\text{C}_{114.2}\text{H}_{106}\text{N}_5.8\text{O}_{10}\text{Fe} \cdot \text{CH}_2\text{Cl}_2 \cdot \text{C}_6\text{H}_{14}$: C, 74.67; H, 6.12; N, 3.36. Found: C, 74.78; H, 6.32; N, 4.17.

Single crystals of $[\text{Fe}(\text{3,5-Me-BAFP})(\text{iPrNO})_2]$ were grown by dissolving 2 mg of the complex in 0.2 mL of THF in a 5 mm glass tube layered with 0.8 mL of hexanes. The tube was sealed and left at room temperature in the glovebox for 5 days to grow crystals suitable for X-ray diffraction.

Reactions with NO Gas. For the reactions of our RNO complexes with NO, we first equilibrated a solution of dichloromethane with NO gas. Then, we titrated a solution of cobalt tetraphenylporphyrin $[\text{Co}(\text{TPP})]$ in THF with the $\text{CH}_2\text{Cl}_2/\text{NO}$ solution to form the corresponding $[\text{Co}(\text{TPP})(\text{NO})]$ complex, to determine the concentration of dissolved NO gas. From these data we approximated the NO concentration of the solution (typically $\sim 1\text{ mM}$), and then titrated $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$, $[\text{Fe}(\text{TPP})(\text{MI})(\text{PhNO})]$ and $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$ in THF with the calibrated $\text{CH}_2\text{Cl}_2/\text{NO}$ solution.



Lewis acid Reactions. Solutions of $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$, $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$, and $[\text{Fe}(3,5\text{-Me-BAFP})(\text{NO})]$ were prepared by dissolving the compounds in dry THF in the glovebox. The Lewis acids (2.5 equiv) were added to the solutions either by micro-syringe (for solutions), or were weighed out and then added (for solids). The reactions were monitored by UV-Vis spectroscopy. The solutions were then layered with 4X hexanes and left in the $-33\text{ }^\circ\text{C}$ glovebox freezer to precipitate overnight. The resulting solids were filtered off and collected the next day. In the case of the solutions where $\text{BF}_3\cdot\text{OEt}_2$ was added, the excess $\text{BF}_3\cdot\text{OEt}_2$ was difficult to remove and could be seen coating the solids, even after several washes with hexanes.

Single crystals of $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})(\text{iPrNH}_2)]$ were grown by adding 2 mg $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$, dissolved in 0.2 mL THF, to a 5 mm glass tube. 2.5 equiv of $\text{B}_2(\text{pin})_2$ were added to the solution. The solution was carefully layered with 0.8 mL of hexanes and suitable crystals for X-ray crystallography grew in 5 days.

DFT calculations. The structures of the PhNO and iPrNO complexes and their HNO analogs were fully optimized for their singlet ($S = 0$) ground states, using the BP86 functional^{53, 54} and the TZVP basis set.^{55, 56} In these calculations, the simple porphine (P^{2-}) ligand was used. Vibrational frequencies calculated for the structures show no imaginary frequencies. In all calculations, convergence was reached when the relative change in the density matrix between subsequent iterations was less than 1×10^{-8} . All of these calculations were performed using the program package Gaussian 09.⁵⁷

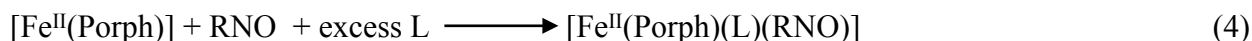
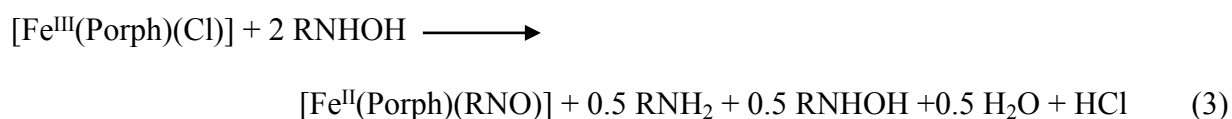
Results & Discussion

Preparation and Spectroscopic Characterization of $[\text{Fe}(\text{TPP})(\text{L})(\text{RNO})]$. For the synthesis of ferrous porphyrin RNO complexes, two different methods have been reported in the literature.^{32,}

⁵² The first method is based on the reaction of the ferric complexes, $[\text{Fe}(\text{Porph})(\text{Cl})]$ (where Porph^{2-}



= dianion of a generic porphyrin ligand), with excess N-(alkyl/aryl)hydroxylamines (RNHOH, R = alkyl, aryl), which leads to a disproportionation reaction to give the ferrous RNO complexes, as shown in equation (3). The second method of preparation is the direct reaction of the ferrous porphyrins, [Fe(Porph)], with nitrosoalkane ligands (RNO, R = alkyl, aryl) to give the corresponding ferrous RNO complexes. These reactions are usually carried out in the presence of excess neutral ligands (L), such as the N-donor ligands 1-methylimidazole (MI) or pyridine (Pyr), to form the corresponding six-coordinate (6C) complexes, [Fe(Porph)(L)(RNO)], as shown in equation (4). In this work, we used modified versions of both literature methods to synthesize our ferrous porphyrin RNO model complexes (where R = iPr, Ph; see Experimental Section).



We modified the reaction shown in equation (3) by starting from a ferric complex with a weakly coordinating anion, [Fe(TPP)(X)], where TPP²⁻ = tetraphenylporphyrin dianion, and X = BF₄⁻, PF₆⁻ or SbF₆⁻. For example, we reacted [Fe(TPP)(BF₄)] with excess N-Phenylhydroxylamine (PhNHOH) in tetrahydrofuran (THF) to yield the corresponding ferrous PhNO complex in ~90% yield. The UV-Vis spectrum of the reaction product (in CH₂Cl₂) exhibits the Soret band at 412 nm and the main Q band at 523 nm, indicative of a ferrous species as shown in Figure S4. Next, we further characterized the product by ¹H-NMR spectroscopy. The iron-PhNO complex shows a multiplet of peaks from 6.04 – 6.51 ppm that originate from the bound PhNO ligand, and the porphyrin ligand peaks range from 7.72 – 8.58 ppm; see Figure S4, bottom. These chemical shifts indicate that the product is pure, and they are characteristic of a diamagnetic, low-spin (ls)



ferrous porphyrin complex (the precursor is paramagnetic). It should be noted that these peaks were assigned based on the reported ^1H -NMR spectrum of $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$.⁴²

Despite the available, thorough structural and spectroscopic characterization of ferrous heme-RNO complexes,^{34, 35} the vibrational characterization of these complexes is incomplete. In particular, N-O stretching frequencies are often not confirmed by isotope labeling, and little is known about the Fe-N(R)O stretching and bending vibrations. In the case of PhNO complexes,^{32, 42, 52, 58, 59} N-O stretching frequencies have been reported in the 1340 – 1370 cm^{-1} range (see Table 1). To determine the N-O stretch of our mono-PhNO complex, we synthesized the corresponding Ph^{15}NO complex using the method according to equation (4). Surprisingly, the IR spectra in Figure 1, top show the presence of two isotope sensitive features at 1350 and 1368 cm^{-1} , where the latter band is significantly lower in intensity than the 1350 cm^{-1} feature. This observation prompted us to investigate whether the ferrous PhNO complex obtained in this way could be a mixture of different species. The reported N-O stretching frequency for $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$ is 1353 cm^{-1} ,⁴² but unfortunately, the IR spectrum of this complex is not shown in the previous report, so it is not clear how intense this IR band is. The IR spectrum of $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$ prepared by us, following the reported method of preparation,⁴² shows a very intense N-O stretching band at 1353 cm^{-1} that shifts to 1336 cm^{-1} with Ph^{15}NO ($\Delta = 17 \text{ cm}^{-1}$), as shown in Figure 1, bottom. The 1353 cm^{-1} band (and its relative intensity) in $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$ matches one of the bands in the IR spectrum of our “mono-PhNO” preparation (see Figure S5), which implies that our product contains a significant amount of the ferrous bis-PhNO complex. Additionally, the IR spectra in Figure S6 (top) show that it makes little difference for the relative amounts of the iron-PhNO products formed whether one equivalent or excess PhNO is used in the reaction with $[\text{Fe}(\text{TPP})]$. Richter-Addo and coworkers also reported the crystal structure of a mono-PhNO analog, $[\text{Fe}(\text{TPP})(\text{MI})(o\text{-tolNO})]$,



which has an N-O stretching frequency of 1366 cm^{-1} .⁵⁹ This value is very close to the other N-O band at 1368 cm^{-1} in our PhNO complex. Based on this result, we conclude that the bis- and mono-PhNO complexes have in fact different N-O stretching frequencies, and that the “mono-PhNO” product that we obtained is in fact a mixture of $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ and $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$.

Finally, we synthesized the Pyr- and MI-bound complexes, $[\text{Fe}(\text{TPP})(\text{Pyr})(\text{PhNO})]$ and $[\text{Fe}(\text{TPP})(\text{MI})(\text{PhNO})]$, first reported by Oldfield and coworkers,⁵⁸ which show the N-O stretch at 1342 cm^{-1} (1319 cm^{-1} with Ph^{15}NO ($\Delta = 23\text{ cm}^{-1}$), see Figure S7) and 1336 cm^{-1} (1313 cm^{-1} with Ph^{15}NO ($\Delta = 23\text{ cm}^{-1}$), see Figure S8), respectively. Interestingly, in the presence of the stronger ligands Pyr and MI (compared to THF), formation of the bis-PhNO complex is not observed.

To gain further insight into the vibrational properties of these PhNO complexes, we employed DFT calculations. We optimized a series of ferrous porphyrin-PhNO complexes as shown in Table 1 using BP86/TZVP. Comparison of the optimized structures to the available crystal structures shows good agreement, with deviations in bond lengths $<0.06\text{ \AA}$. Unfortunately, the experimental difference in the N-O stretching frequencies ($15\text{-}20\text{ cm}^{-1}$) between the mono- and bis-PhNO complexes is not reproduced by the DFT calculations. The calculated N-O stretching frequencies of $[\text{Fe}(\text{P})(\text{MI})(\text{PhNO})]$, $[\text{Fe}(\text{P})(\text{THF})(\text{PhNO})]$ and $[\text{Fe}(\text{P})(\text{PhNO})_2]$ (P^{2-} = porphine) are 1380 , 1386 and 1383 cm^{-1} , respectively (P^{2-} = porphine dianion). It should be noted that the 1383 cm^{-1} band of the $[\text{Fe}(\text{P})(\text{PhNO})_2]$ complex is the antisymmetric combination of the N-O stretching coordinates, while the symmetric mode is not IR active (predicted at 1403 cm^{-1}). Based on our combined experimental and theoretical results we conclude that the 1368 cm^{-1} band in the IR spectrum of our PhNO product belongs to the ferrous mono-PhNO complex (with a THF molecule bound to the iron center) and that the 1350 cm^{-1} band is the ferrous bis-PhNO complex, $[\text{Fe}(\text{TPP})(\text{PhNO})_2]$.



We envisioned that changing the R group of the RNO ligand to an isopropyl group would result in a better model for a ferrous heme-HNO complex, due to the strong conjugation of the nitroso group with the aromatic phenyl ring in PhNO. Using the method shown in equation (4) we reacted [Fe(TPP)] with a slight excess of iPrNO to form [Fe(TPP)(THF)(iPrNO)] in THF with a 67% yield. In the UV-Vis spectra in Figure 2, the Soret band of the precursor, [Fe(TPP)], shifts from 426 to 417 nm upon coordination of iPrNO but remains very symmetric, which is a feature characteristic of ferrous porphyrins. At the same time, the main Q band of [Fe(TPP)] shifts from 550 to 535 nm. The resulting UV-Vis spectrum of [Fe(TPP)(THF)(iPrNO)] is similar to that of [Fe(TPP)(THF)(PhNO)]. The ^1H -NMR spectrum of the isolated product is assigned based on a previous literature report.³² We observe the porphyrin resonances from 7.70 – 8.73 ppm, indicative of a diamagnetic ferrous heme complex (note that the ferrous precursor is paramagnetic), and two peaks shifted upfield to -0.61 and -2.07 ppm, which originate from the bound iPrNO ligand (due to the ring current of the porphyrin), as shown in Figure S9. X-ray crystallography (see below) further supports that [Fe(TPP)(THF)(iPrNO)] is obtained as a pure compound.

N-O stretching frequencies have previously been reported for 6C ferrous heme-iPrNO complexes with MI, Pyr, or methanol as axial ligands, which range from 1420 – 1440 cm^{-1} (see Table 2).^{32, 52, 59} We determined the N-O stretch of [Fe(TPP)(THF)(iPrNO)] to be 1440 cm^{-1} , which shifts to 1417 cm^{-1} with ^{15}N -labeling ($\Delta = 23 \text{ cm}^{-1}$), as shown in Figure 3, consistent with these reports. Based on the closer similarity of the iPrNO ligand with HNO, and the fact that [Fe(TPP)(THF)(iPrNO)] can be obtained as a pure compound, we chose this complex for further detailed NRVs studies (see below).

Crystallographic Studies of [Fe(TPP)(L)(RNO)]. Experimental details about the structural studies are provided in the Experimental Section. X-ray quality crystals of



[Fe(TPP)(THF)(PhNO)] were obtained by vapor diffusion of hexanes into a concentrated THF solution of the complex at -33 °C in the glovebox freezer. This resulted in a crystal with two different PhNO complexes in the unit cell, as shown in Figure 4 (top), but surprisingly, not the bis-PhNO complex as anticipated from the IR results (see above). One of the species in the unit cell is the target complex [Fe(TPP)(THF)(PhNO)], which has a THF molecule bound to the iron center. The crystal structure exhibits an Fe-N(Ph)O bond length of 1.813 Å and an Fe-N-O angle of 122°. These geometric parameters are very similar to those of [Fe(TPP)(MI)(PhNO)] that has an Fe-N(Ph)O bond length of 1.80 Å and an Fe-N-O bond angle of 125° (see Table 1).⁵⁸ The other heme complex in the unit cell is the phenylamine (PhNH₂) bound complex, [Fe(TPP)(PhNH₂)(PhNO)]. The phenylamine is formed during the disproportionation reaction of the ferric heme complex with excess PhNHOH, reflected in equation (3). This was a surprise, as we did not expect PhNH₂ to be bound in the presence of the coordinating solvent THF. This structure shows disorder of the bound PhNO and PhNH₂ ligands above and below the heme plane, resulting in corresponding Fe-N bond lengths of 1.963 and 1.994 Å from the structural fit. However, these bond lengths are significantly longer than the Fe-N(Ph)O and Fe-O(THF) bond lengths of [Fe(TPP)(THF)(PhNO)]. They are also much longer than all of the other reported Fe-N(Ph)O distances in ferrous heme-RNO complexes (1.80 - 1.90 Å), as shown in Table 1.^{32, 42, 52, 58, 59} On the other hand, the Fe-N(amine) bond length is shorter than that in bis(primary amine) ferrous porphyrins, where the Fe-NH₂R bond lengths range from 2.028 - 2.045 Å.⁶⁰ Based on these considerations, we feel that the Fe-N bond distances in [Fe(TPP)(PhNH₂)(PhNO)] are less well defined due to the disorder, and should be treated with caution.

X-ray quality crystals of [Fe(TPP)(THF)(iPrNO)] were obtained by layering a concentrated solution of the complex with hexanes at room temperature. The structure of



[Fe(TPP)(THF)(iPrNO)] has a THF molecule bound to the iron center as shown in Figure 4, as expected based on the synthetic conditions used. In the structure, the Fe-N(iPr)O bond length is 1.853 Å and the Fe-N-O bond angle of 121°. This is in good agreement with other 6C heme-iPrNO complexes that have Fe-N(iPr)O bond lengths between 1.80 – 1.89 Å and Fe-N-O bond angles from 116 – 124°, as shown in Table 2.^{32, 52} The Fe-N(iPr)O bond length of this complex is slightly longer than that of the MI adduct, [Fe(TPP)(MI)(iPrNO)] at 1.81 Å, but the Fe-N-O angle is quite similar (122°). These geometric parameters are also consistent with the crystal structure of the ls-{FeNO}⁸ complex [Fe(TFPPBr₈)(NO)]⁻ (where TFPPBr₈ = dianion of 2,3,7,8,12,13,17,18-octabromo-5,10,15,20-(tetra-(pentafluorophenyl)porphyrin), which exhibits an Fe-NO bond length of 1.814 Å and an Fe-N-O angle of 122°. ⁶¹ Also, all of these values compare well with the Mb(II)-NHO adduct where EXAFS fitting determined the Fe-NHO bond length to be 1.82 Å. ⁶² In contrast, ls-{FeNO}⁷ complexes, such as [Fe(TPP)(NO)] and [Fe(TPP)(MI)(NO)], exhibit much shorter Fe-NO bond lengths of 1.72⁶³ and 1.750 Å, ⁶⁴ respectively.

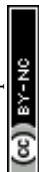
The heme is completely planar in all of our RNO crystal structures, in agreement with previous reports,⁵⁸ except in [Fe(TPP)(THF)(PhNO)] where the porphyrin shows some ruffling distortion as illustrated in Figure 5. This distortion is unusual for 6C ferrous porphyrins since the iron is located in the heme plane. A ruffling distortion is characterized by the rotation of *trans* pyrrole rings in the opposite direction around the Fe-N_{pyrrole} bonds of the porphyrin co-ligand. This out-of-plane distortion is measured by the root mean square deviation (RMSD) of the porphyrin atoms from the heme plane. The 25-atom core displacement of [Fe(TPP)(THF)(PhNO)] is 0.15 Å, whereas planar hemes are defined by a 25-atom core displacement of less than 0.10 Å. ⁶⁵ For example, the completely planar complex [Fe(TMP)(MI)₂](ClO₄) (TMP = dianion of tetramesitylporphyrin) has a 25-atom core displacement of only 0.02 Å, ⁶⁶ which is 7.5 times



smaller than that of [Fe(TPP)(THF)(PhNO)]. The RMSD for the 4-atom *meso* carbon displacement in [Fe(TPP)(THF)(PhNO)] is 0.15 Å.

Lastly, 6C ferrous heme-RNO complexes typically exhibit average Fe-N_{pyrrole} bond distances of ~2.0 Å (see Tables 1 and 2). This bond length is indicative of low-spin ferrous hemes, which exhibit average Fe-N_{pyrrole} bond distances between 1.981 – 2.008 Å.⁶⁷

Characterization of [Fe(3,5-Me-BAFP)(iPrNO)₂] and Crystallographic Studies. The iPrNO complex with our bulky bis-picket fence porphyrin, 3,5-Me-BAFP²⁻, was prepared under identical conditions as [Fe(TPP)(THF)(iPrNO)], where the reduced porphyrin was stirred with a slight excess of iPrNO in THF, and the product was precipitated with hexanes to yield a purple solid. The resulting complex was characterized by UV-Vis, IR, and rRaman spectroscopy, elemental analysis, and X-ray crystallography. It is interesting to note that unlike the TPP²⁻ version, this reaction forms the bis-iPrNO complex almost exclusively. For example, UV-Vis titration of [Fe(3,5-Me-BAFP)] with iPrNO in THF shows that at 1 equiv of iPrNO addition, the bis-iPrNO complex forms along with starting material, whereas the mono-iPrNO complex is not present in any significant amounts (Figure S18). The crystal structure of [Fe(3,5-Me-BAFP)(iPrNO)₂] is shown in Figure 6, where crystals were grown in a mixture of THF and hexanes. Two crystals were analyzed, but only one is discussed in the main text. It was found that in these crystals, 80-90% of the bis-iPrNO species is present, whereas 10-20% of the complex has THF as the 6th ligand. The crystal displays the iPrNO ligand disordered with a THF molecule. The occupancy ratio refined to 0.778(4) to 0.222(4). This indicates that Fe(3,5-Me-BAFP) has a stronger propensity towards the iPrNO ligand compared to Fe(TPP). We hypothesize that this could be due to the steric bulk in the bis-picket fence porphyrin, which increases the binding constant for the smaller iPrNO ligand compared to THF. Both porphyrins are completely planar.



The crystal structures exhibit average Fe-N_{pyrrole} bond distances of ~2.0 Å, again indicative for low-spin ferrous hemes.⁵¹ When comparing the geometric properties of [Fe(TPP)(THF)(iPrNO)] and [Fe(3,5-Me-BAFP)(iPrNO)₂] (Table 2), most of the distances and angles are relatively similar except that the Fe-N(iPr)O distance in the 3,5-Me-BAFP system (1.920 Å) is longer than in the TPP system (1.853 Å) and most of the porphyrin iPrNO complexes reported to date (see Table 2). [Fe(3,5-Me-BAFP)(iPrNO)₂] shows the Soret band at 425 nm and has an N-O stretching frequency of 1449 cm⁻¹, determined by IR spectroscopy (Figure S18).

Vibrational Analysis of [Fe(TPP)(THF)(iPrNO)]. Detailed vibrational assignments of ferrous heme-nitrosoalkane complexes that include metal-N(R)O stretching and bending modes are lacking. On the other hand, for the ferrous heme-HNO adduct in myoglobin (Mb), Farmer and coworkers reported the Fe-NHO stretch at 651 cm⁻¹, but the exact nature of this vibration was not further investigated (the assignment is simply based on the fact that this is the only low-energy mode observed by resonance Raman (rRaman) that is isotope sensitive).⁶² Recently, we showed that in the model complex [Fe(3,5-Me-BAFP)(MI)(NHO)], the Fe-NHO stretch and the Fe-N-O bend are located at 644 and 461 cm⁻¹, respectively, using nuclear resonance vibrational spectroscopy (NRVS) coupled to ¹⁵N-isotope labeling.⁶⁸ To investigate this further, we now measured the Fe-N(iPr)O stretching and bending frequencies of [Fe(TPP)(THF)(iPrNO)] using NRVS, and then analyzed these data using DFT calculations. The NRVS data of the ⁵⁷Fe-labeled iPrNO/iPr¹⁵NO complexes are shown in Figure 7, top. Two isotope sensitive bands are identified in the NRVS data. The first feature is observed 426 cm⁻¹, which shifts to 418 cm⁻¹ with iPr¹⁵NO ($\Delta = 8$ cm⁻¹). The second band is significantly lower in intensity and is found at 633 cm⁻¹. This feature shifts to 627 cm⁻¹ with iPr¹⁵NO ($\Delta = 6$ cm⁻¹). To determine the nature of these FeNO-based vibrations we employed DFT computations. The BP86/TZVP calculated geometric parameters of



the ferrous $iPrNO$ complexes are listed in Table 2. Overall, the calculated structure of $[Fe(P)(THF)(iPrNO)]$ is in good agreement with the crystal structure of $[Fe(TPP)(THF)(iPrNO)]$ determined here. The N-O stretching frequency is also within $\sim 20\text{ cm}^{-1}$ of the experimental value for this complex. With this, we predicted the NRVS data of the $[Fe(P)(iPrNO)]$ and $[Fe(P)(THF)(iPrNO)]$ complexes, as shown in Figure 7, bottom. Interestingly, the five-coordinate (5C) complex $[Fe(P)(iPrNO)]$ is predicted to show an $FeN(R)O$ -based feature at 608 cm^{-1} , which increases in energy (646 cm^{-1}) in the THF-bound complex. The calculated NRVS data of $[Fe(P)(THF)(iPrNO)]$, shown in Figure 7, bottom, exhibit isotope sensitive features at 646 and 447 cm^{-1} , which nicely reproduce the intensity pattern of the 633 and 426 cm^{-1} bands observed in our experimental data (Figure 7, top).

In comparison, for RNO adducts of ferrous heme proteins (sGC, Hb, and Mb, where $R = \text{Me}$, Pentyl, and Octyl), $Fe-N(R)O$ stretching modes have been assigned to rRaman features in the $540 - 560\text{ cm}^{-1}$ range in the literature.⁶⁹⁻⁷⁴ However, these assignments are merely based on the similarity of these vibrational energies to those of the supposed $Fe-NO$ stretches of $6C\text{ ls-}\{FeNO\}^7$ porphyrin complexes ($545 - 565\text{ cm}^{-1}$), but no further analysis of the vibrational data was provided. Note that it was later determined, using NRVS, that the vibrational feature around $545-565\text{ cm}^{-1}$ in $6C\text{ ls-}\{FeNO\}^7$ porphyrin complexes (for example, at 551 cm^{-1} in $Mb(II)-NO$ and 563 cm^{-1} in $[Fe(TPP)(MI)(NO)]$) has primary $Fe-N-O$ bending character.^{43, 75, 76}

To make further comparisons between the analogous heme-HNO and -RNO complexes, we also optimized the structures of $[Fe(P)(L)(NHO)]$, where $L = THF$ and MI , using MI is a model for histidine (see Table 2). The calculated structure of $[Fe(P)(MI)(NHO)]$ agrees well with the EXAFS data for the $Mb(II)-HNO$ species. However, the N-O stretching frequency for the HNO complex is overestimated by $\sim 75\text{ cm}^{-1}$. It is possible that the hydrogen bonding interactions



between the HNO ligand and the distal histidine in the active site of Mb need to be included in order to accurately predict the N-O stretch. In this regard, Zhang and coworkers have shown that such hydrogen bonding interactions are important for reproducing experimental data of the Mb(II)-NHO complex with computational models.⁷⁷ On the other hand, the experimental Fe-NHO stretching vibrations in Mb(II)-NHO (651 cm⁻¹) and [Fe(3,5-Me-BAFP)(MI)(NHO)] (644 cm⁻¹) are very close in energy to the DFT calculated feature at 653 cm⁻¹. The calculated NRVS data of the HNO adducts are shown in Figure S10. The calculated vibrational frequencies of the iron(II)-NHO complex, with the Fe-NHO stretch and the Fe-N-O bend predicted at 653 cm⁻¹ and 447 cm⁻¹, respectively, are therefore in very good agreement with the experimental results for [Fe(3,5-Me-BAFP)(MI)(NHO)], where the Fe-NHO stretch was assigned to the mode at higher frequency (~650 cm⁻¹; also confirmed by Farmer's work on Mb(II)-NHO as mentioned above⁶²), whereas the Fe-N-O bend was assigned to the peak at 461 cm⁻¹.

Note that in the bent geometry of the FeNHO unit (with an Fe-N-O angle of about 130°), significant mixing between the Fe-NO stretching and Fe-N-O bending internal coordinates can be expected, as we have previously investigated in detail for ls-{FeNO}⁷ complexes that contain bent FeNO units.^{43, 75, 78} Vibrational analysis of our DFT results for [Fe(P)(NHO)]-type complexes shows indeed that the vibrational features at ~650 and ~460 cm⁻¹ contain significant Fe-NO stretching, and Fe-N-O and Fe-N-H bending character, with the higher frequency mode best assigned to the Fe-NO stretch, whereas the Fe-N-O bend resembles the lower energy mode. On the other hand, based on our DFT model [Fe(P)(THF)(iPrNO)], we conclude that the low-intensity feature at 633 cm⁻¹ in [Fe(TPP)(THF)(iPrNO)] is predominantly an isopropyl bending mode with some Fe-N-O bending character, whereas the intense feature at 426 cm⁻¹ is the Fe-N-O bending mode, with some admixture of Fe-NO stretching character (calculated at ~450 cm⁻¹). The main



component of the Fe-NO stretch is calculated as a weaker feature at 422 cm^{-1} , but no corresponding peak could be unequivocally identified in the experimental NRVS data of $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$. The distinct differences in vibrational properties between the analogous Fe-N(H)O and Fe-N(iPr)O complexes are due to (a) a weaker Fe-N(R)O bond in the iPrNO complex (calculated Fe-N(R)O force constants are $3.1\text{ mdyne/\AA}$ for the HNO compared to $2.7\text{ mdyne/\AA}$ for the iPrNO complex), and (b), strong coupling between the low-energy modes of the FeNO unit and bending modes of the isopropyl substituent in the iPrNO complex. These two effects combined result in the observed differences between vibrational features in the iPrNO and HNO model complexes.

Reactivity with NO. To the best of our knowledge, the reactivity of ferrous heme-RNO complexes with NO gas has not been previously studied. However, as pointed out in the Introduction, the reaction of the Fe(II)-NHO adduct in Cyt. P450nor with NO is one possible route for N-N bond formation and N_2O generation in this enzyme.¹⁷ Hence, we decided to investigate whether our RNO complexes would react with NO, and what products would result from this reaction. The UV-Vis spectra in Figure 8 show the titration of $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ in THF with an NO-saturated CH_2Cl_2 solution (typically $\sim 1\text{ mM}$) at room temperature. The reaction is complete with ~ 1 equiv. of NO, and the UV-Vis data of the reaction product exhibit features at 412, 476, and 540 nm that are identical to those of the $\text{ls-}\{\text{FeNO}\}^7$ complex, $[\text{Fe}(\text{TPP})(\text{NO})]$. We hypothesize that this $\text{ls-}\{\text{FeNO}\}^7$ complex is generated by formation of the intermediate, $[\text{Fe}(\text{TPP})(\text{NO})(\text{PhNO})]$, which, due to the strong thermodynamic σ -*trans* effect of NO (also referred to as a *trans* “interaction”, since it is a ground state effect), causes the PhNO ligand to dissociate, as shown in Scheme 3.⁷⁸⁻⁸⁰ It is important to point out that the proposed intermediate, $[\text{Fe}(\text{TPP})(\text{PhNO})(\text{NO})]$, would likely not be observed at room temperature, due to the large binding



constant of NO to a ferrous heme ($K_{eq} = 10^{10-12} \text{ M}^{-1}$).⁸¹⁻⁸³ With this result, we hypothesized that binding of an N-donor ligand, MI, to the iron center, leading to the corresponding 6C complex, $[\text{Fe}(\text{TPP})(\text{MI})(\text{PhNO})]$, could force the NO to react with the PhNO ligand, rather than NO simply binding to the iron center in trans position to PhNO, and then, due to the σ -*trans* effect of NO, dissociating the RNO ligand. The complex $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ in dichloromethane binds ~ 1 equivalent of MI to form the 6C complex, $[\text{Fe}(\text{TPP})(\text{MI})(\text{PhNO})]$, as shown in Figure S11. The UV-Vis spectra show that the Soret band shifts from 412 to 426 nm and the Q band at 523 shifts to 540 nm upon MI binding. Interestingly, this implies that RNO ligands in heme complexes have essentially no *trans* effect, in contrast to NO. This result compares well with a previous DFT study from our laboratory that predicts that heme-HNO complexes have a significantly weaker σ -*trans* effect compared to NO, more comparable to that of CO.⁸⁴

We then reacted $[\text{Fe}(\text{TPP})(\text{MI})(\text{PhNO})]$ with low equivalents of NO gas in dichloromethane at room temperature. The UV-Vis spectra of the products formed, shown in Figure S12, are again identical to that of $[\text{Fe}(\text{TPP})(\text{NO})]$. We envisioned that this could arise from (1) a weak Fe-RNO bond, or (2) the NO ligand having a higher binding constant than MI. Hypothesis (1) was tested by adding excess MI to a solution of $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ in dichloromethane. In Figure S13, the UV-Vis spectra show that the addition of 1 equiv. or 50 equiv. of MI results in no change in the UV-Vis spectrum. We expected the formation of a ferrous bis-imidazole complex if the Fe-RNO bond is weak, but this is not the case (the ferrous bis-imidazole adduct has UV-Vis bands at 430 and 536 nm). In summary, ferrous RNO complexes have strong Fe-N(R)O bonds like ferrous heme-nitrosyls ($\text{ls-}\{\text{FeNO}\}^7$) and ferrous heme-CO complexes. Based on these findings, we can rule out hypothesis (1) and we believe that the NO ligand first



displaces the imidazole (MI) ligand, and then again forces out the RNO ligand via the σ -*trans* effect (see Scheme 3).

Finally, we investigated the reactivity of [Fe(TPP)(THF)(iPrNO)] with an NO-saturated CH₂Cl₂ solution at room temperature. Unfortunately, we observed the same general findings as with the ferrous PhNO complexes. In summary, even in the presence of MI we observe [Fe(TPP)(NO)] as the product of both reactions (see Figures S14 – S15). Overall, this shows that heme-RNO complexes with the TPP²⁻ porphyrin are generally unreactive toward NO and do not lead to N-N bond formation, in contrast to heme-HNO complexes.¹⁷

Reactivity with Lewis Acids. iPrNO is a better structural and isoelectronic mimic of HNO than PhNO, due to the strong conjugation of the nitroso group with the aromatic phenyl ring in PhNO. Thus, the following studies focused on iPrNO. The reactivity of [Fe(TPP)(THF)(iPrNO)] with Lewis acids was first explored. BF₃•OEt₂ (boron trifluoride dietherate) and bis(pinacolato)diboron (B₂(pin)₂) were chosen as the Lewis acids. BF₃•OEt₂ is a much stronger Lewis acid compared to B₂(pin)₂. BF₃•OEt₂ was also used successfully by the Richter-Addo group to obtain Lewis acid adducts of [Fe(OEP)(NO)].³⁹ Another important factor in choosing Lewis acids is size. While Fe(TPP) is a completely open scaffold and can interact with any size Lewis acid, the sterically bulky Fe(3,5-Me-BAFP) complex has a limited pocket size. Both of the Lewis acids, BF₃•OEt₂ and B₂(pin)₂, are small enough to fit into the pocket of 3,5-Me-BAFP²⁻ (results described below). [Fe(TPP)(THF)(iPrNO)] was stirred in THF with both Lewis acids and the reaction was monitored by UV-Vis spectroscopy over the course of 2 hrs. No reaction was observed. No shift in the Soret band in UV-Vis spectroscopy was observed and there was no shift in the N-O stretching band by IR spectroscopy. Fe(3,5-Me-BAFP) was investigated next, to see if



adding a non-polar, sterically protected pocket would help promoting the binding of the Lewis acid to the corresponding ferrous iPrNO complex.

$[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ was reacted with 2.5 equiv of $\text{BF}_3 \cdot \text{OEt}_2$, $\text{B}(\text{OEt})_3$ (triethoxy borate), $\text{B}(\text{C}_6\text{F}_5)(\text{OH})_2$ (pentafluorophenylboronic acid), $\text{B}(\text{Mes})_2\text{Br}$ (dimesitylboron bromide), and $\text{B}_2(\text{pin})_2$ in THF and the reaction mixtures were monitored by UV-Vis spectroscopy. The reaction of $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ with $\text{BF}_3 \cdot \text{OEt}_2$ caused a shift in the Soret band to 407 nm, whereas the reaction with $\text{B}_2(\text{pin})_2$ shifted the Soret band to 416 nm (Figure S19). Both reactions were monitored over the course of 2 hrs by UV-Vis spectroscopy, but no further changes occurred after the initial 2 min. $\text{B}(\text{C}_6\text{F}_5)(\text{OH})_2$ and $\text{B}(\text{Mes})_2\text{Br}$ reacted much slower, and over the course of 90 min, the Soret band shifted to 403 nm (Figure S19), which is similar to the product obtained with $\text{BF}_3 \cdot \text{OEt}_2$. $\text{B}_2(\text{pin})_2$ therefore forms a unique product, compartmented to the other Lewis acids, with the Soret band at 416 nm. $\text{B}(\text{OEt})_3$, which is a very weak Lewis acid, did not fully convert to a new product (Figure S19).

The products of these reactions were then further characterized. In the case of $\text{BF}_3 \cdot \text{OEt}_2$, the UV-Vis spectrum of the product shows the Soret and Q band at 403 and 527 nm, which matches with the absorption spectrum of $[\text{Fe}^{\text{III}}(3,5\text{-Me-BAFP})(\text{THF})_2]$. This indicates that the Lewis acids ($\text{BF}_3 \cdot \text{OEt}_2$, $\text{B}(\text{C}_6\text{F}_5)(\text{OH})_2$, and $\text{B}(\text{Mes})_2\text{Br}$) oxidize the iron center and this causes the iPrNO ligand to dissociate. This is in line with previous observations that oxidation of ferrous heme-RNO adducts leads to RNO release.^{32, 72} In order to test this hypothesis, $\text{B}(\text{Mes})_2\text{Br}$ was reacted with $[\text{Fe}^{\text{II}}(3,5\text{-Me-BAFP})]$ and the progress of the reaction was monitored by UV-Vis spectroscopy. Upon addition of the Lewis acid, the Soret band of $[\text{Fe}^{\text{II}}(3,5\text{-Me-BAFP})]$ shifted to 403 nm, verifying the oxidizing ability of these Lewis acids.



In order to identify the product of the reaction of $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ with $\text{B}_2(\text{pin})_2$, we crystallized the resulting complex as shown in Figure 9. The structure was solved by isomorphous replacement using the $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ crystal structure. The disorder model was adjusted (two methylene chloride moieties, no THF molecule, partial oxygen loss). The isopropyl nitrosyl ligand was found to be disordered with an isopropyl amine ligand, i.e. the oxygen atom at the nitrogen was partially replaced by two hydrogen atoms. No disorder was refined for the N atom or the isopropyl group. The occupancy ratio refined to 0.522(6) to 0.478(6) for $\text{iPrNO}:\text{iPrNH}_2$. A solvate occupied area was refined as major hexanes and minor methylene chloride (two moieties). The crystal structure in Figure 9 depicts the $\sim 50\%$ disorder of $\text{iPrNO}:\text{iPrNH}_2$ as the mixed complex, where iPrNO is coordinated on one side of the heme and iPrNH_2 is bound in trans position, on the other side of the heme. Just based on the crystallographic data, however, it is also possible that the 50% disorder is represented as half bis- iPrNO and half bis- iPrNH_2 complex.

The reaction product was also characterized by rRaman spectroscopy (Figure S20), which shows the oxidation state marker band at 1369 cm^{-1} and the spin state marker band at 1568 cm^{-1} .⁸⁵⁻
⁸⁷ There are some slight shifts compared to the rRaman spectrum of the starting complex $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$, indicating that oxidation state and spin state remain the same upon addition of the Lewis acid, and that the complex is in fact the mixed-ligand compound, $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNH}_2)(\text{iPrNO})]$, as shown in Figure 9. Further evidence for this conclusion comes from UV-Vis analysis, showing that upon addition of the Lewis acid, the Soret band shifts into the region of the mono- iPrNO species. This indicates that the Lewis acid not only attacks the iPrNO group, but is also able to abstract an oxygen atom from the iPrNO ligand, followed by reduction of the remaining ligand fragment to the amine (iPrNH_2). The required electrons likely originate



from the Lewis acid $B_2(\text{pin})_2$, which explains why this reaction is not observed with any of the other Lewis acids studied here.

Summary and Conclusions

In this work, we prepared and fully characterized several ferrous heme-RNO complexes, where R is an isopropyl or phenyl group, using the TPP^{2-} and $3,5\text{-Me-BAFP}^{2-}$ coligands. Although a significant number of complexes of this type have been reported in the literature, as reviewed in refs.^{34,35}, their vibrational characterization is incomplete, and reactivity studies of these complexes with NO and Lewis acids have not been reported. These complexes are isoelectronic to $\text{ls-}\{\text{FeNHO}\}^8$ complexes; however, they do not suffer from the same stability problems as these reactive species. In ferrous heme-RNO complexes, iron is in the low-spin (ls) state, resulting in complexes that are diamagnetic with a closed-shell ground state. Previously, Mansuy and coworkers noted that five-coordinate (5C) $[\text{Fe}(\text{Porph})(\text{RNO})]$ complexes bind nitrogenous bases, L, with high affinity to form the corresponding six-coordinate (6C) complexes $[\text{Fe}(\text{Porph})(\text{L})(\text{RNO})]$.³² Our work shows that $[\text{Fe}(\text{TPP})(\text{THF})(\text{RNO})]$ complexes (R = iPr, Ph) generate the corresponding 6C complexes, $[\text{Fe}(\text{TPP})(\text{MI})(\text{RNO})]$, in the presence of ~1 equiv. of 1-methylimidazole (MI). Therefore, RNO ligands in ferrous heme complexes do not elicit a significant thermodynamic σ -*trans* effect (interaction), unlike the NO ligand in ferrous heme-nitrosyl complexes.^{64, 75, 78, 88, 89} This is further supported by crystal structures of $[\text{Fe}(\text{TPP})(\text{MI})(\text{RNO})]$ (R = iPr, PhNO), $[\text{Fe}(\text{TPP})(\text{MI})(\text{NO})]$, and $[\text{Fe}(\text{TPP})(\text{MI})(\text{CO})]$, which exhibit Fe- N_{MI} bond lengths of 2.05/2.03 Å, 2.173 Å, and 2.071 Å, respectively.^{58, 59, 64, 90} By comparison, in $[\text{Fe}(\text{TPP})(\text{MI})_2]$, where no *trans* interaction exists, the Fe- N_{MI} bond length is 1.997 Å.⁹¹ Hence, ferrous heme-RNO complexes are most similar to ferrous heme-CO complexes that



bind to ferrous hemes predominately via strong π -backbonds, which do not result in a strong *trans* effect. This provides strong support for previous, computational work on ferrous heme-HNO complexes.^{22, 84, 92, 93}

Using ^{15}N isotope labeling, we further assigned the N-O stretching modes in our PhNO and iPrNO complexes. Our data show that the 6C [Fe(TPP)(L)(PhNO)] complexes with L = THF, pyridine (Pyr) and MI have N-O stretching frequencies of 1368, 1342 and 1336 cm^{-1} , respectively. In the presence of a weak axial ligand like THF, the complexes further have a propensity to form the bis-PhNO analog, which shows the (antisymmetric) N-O stretch as a very intense feature at $\sim 1350 \text{ cm}^{-1}$ in the IR spectrum.⁴² In the case of iPrNO, N-O stretches are located at 1440, 1429 and 1423 cm^{-1} for the 6C [Fe(TPP)(L)(iPrNO)] complexes with L = THF, Pyr and MI ligands, respectively. Interestingly, the N-O stretch of [Fe(TPP)(MI)(iPrNO)] at 1423 cm^{-1} is close to the N-O stretch of ferrous heme-HNO complexes, observed in the 1380 – 1400 cm^{-1} range.^{17, 18, 32, 62}

In order to further compare the vibrational properties of heme-iPrNO and -HNO complexes, we then obtained NRVS data for the complex [$^{57}\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})$]. In this case, two ^{15}N -isotope sensitive features are observed at 426 and 633 cm^{-1} , which are assigned to the Fe-N-O bend and a primarily isopropyl bending mode with some Fe-N-O bending character, respectively. NRVS data on the heme-HNO model complex [Fe(3,5-Me-BAFP)(MI)(NHO)] show similar features, in this case at 461 and 644 cm^{-1} , but interestingly, the vibrational assignments are quite different, with the Fe-N(H)O stretch at higher (644 cm^{-1}) and the Fe-N-O bend at lower (461 cm^{-1}) frequency.¹⁷ DFT calculations further confirm these assignments. These results indicate that the Fe-N(R)O bond is weaker than the Fe-NHO bond, causing the majority of the observed differences.



Another important difference between ferrous heme-HNO and -RNO complexes is their overall stability and reactivity. Whereas heme-RNO complexes are completely air stable,³² the HNO complexes are sensitive to O₂. We therefore wondered how the reactivity of these complexes with NO compares. Previous studies have shown that heme-HNO complexes react with NO under N-N bond formation giving a quantitative N₂O yield, and are therefore possible candidates for the key Intermediate I in Cyt. P450_{nor} catalysis.^{17, 26} In contrast, in this study we found that ferrous heme-RNO complexes are generally unreactive toward NO, resulting in the formation of ls-{FeNO}⁷ complexes rather than N-N bond formation. Based on these findings, we conclude that the presence of the R group greatly stabilizes the RNO ligand and corresponding ferrous heme complexes compared to HNO, and hence, whereas ferrous heme-RNO complexes are good structural and electronic models for their HNO analogs,²² they do not model the reactivity of the heme-HNO complexes at all. For future studies, a ferrous RNO complex with an axial anionic thiolate ligand could be synthesized, to examine this point further.

In addition, the Lewis acid reactivity of heme-RNO complexes was investigated, to determine if these compounds would form Lewis acid adducts and could be activated in this way for reaction with NO. This work was in part inspired by a previous report in the literature, where the formation of Lewis acid adducts of ls-{FeNO}⁷ heme complexes was reported.³⁹ We found that [Fe(TPP)(L)(RNO)] complexes are unreactive towards all Lewis acids tested here: BF₃•OEt₂, B(OEt)₃, B(C₆F₅)(OH)₂, B(Mes)₂Br, and B₂(pin)₂ in THF. We then changed the porphyrin to a bulkier scaffold, the bis-picked fence porphyrin 3,5-Me-BAFP²⁻. Surprisingly, the complex [Fe(3,5-Me-BAFP)(iPrNO)₂] is reactive towards Lewis acids, in contrast to the TPP²⁻ complexes. With BF₃•OEt₂, B(OEt)₃, B(C₆F₅)(OH)₂, B(Mes)₂Br, simple oxidation of the iron center to the ferric oxidation state was observed, likely followed by loss of the iPrNO ligand. In contrast,



reaction of $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ with $\text{B}_2(\text{pin})_2$ leads to O-atom abstraction from one of the iPrNO ligands, followed by reduction, to yield the corresponding amine, iPrNH_2 . From these reactions, the mixed ligand complex $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNH}_2)(\text{iPrNO})]$ was isolated and characterized using different spectroscopic methods and X-ray crystallography. These results provide the foundation for further studies to be developed to ultimately capture a Lewis-acid appended heme-RNO species.

Acknowledgements

The authors acknowledge Dr. Benedict J. Elvers for his support with X-Ray crystallography. This work was supported by a grant from the National Science Foundation (CHE-1464696 to NL). This research used resources of the Advanced Photon Source, a U.S. Department of Energy (DOE) Office of Science User Facility at Argonne National Laboratory and is based on research supported by the U.S. DOE Office of Science-Basic Energy Sciences, under Contract No. DE-AC02-06CH11357.

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website. The SI includes UV-Vis, NMR, and FT-IR spectra for $[\text{Fe}(\text{TPP})(\text{L})(\text{PhNO})]$ where $\text{L} = \text{THF}$, MI , and pyridine, and $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$; UV-Vis, IR, and rRaman spectra for $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$ and for products from the reaction with Lewis acids; crystallographic information for $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]/[\text{Fe}(\text{TPP})(\text{PhNH}_2)(\text{PhNO})]$, $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$, $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNO})_2]$, $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNH}_2)(\text{iPrNO})]$, and $[\text{Fe}(3,5\text{-Me-BAFP})(2\text{-MeTHF})(\text{PhNO})]$ (CCDC entries: #2340714, #2341278, #2339812, #2339811, and #2357458).



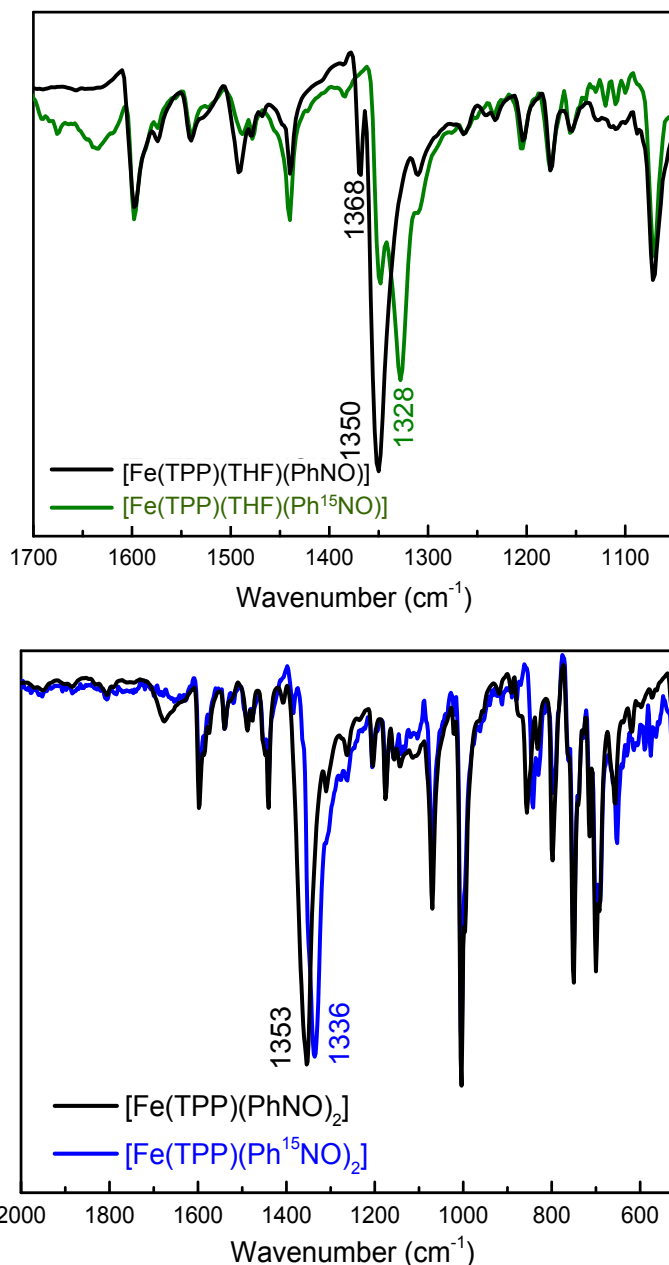
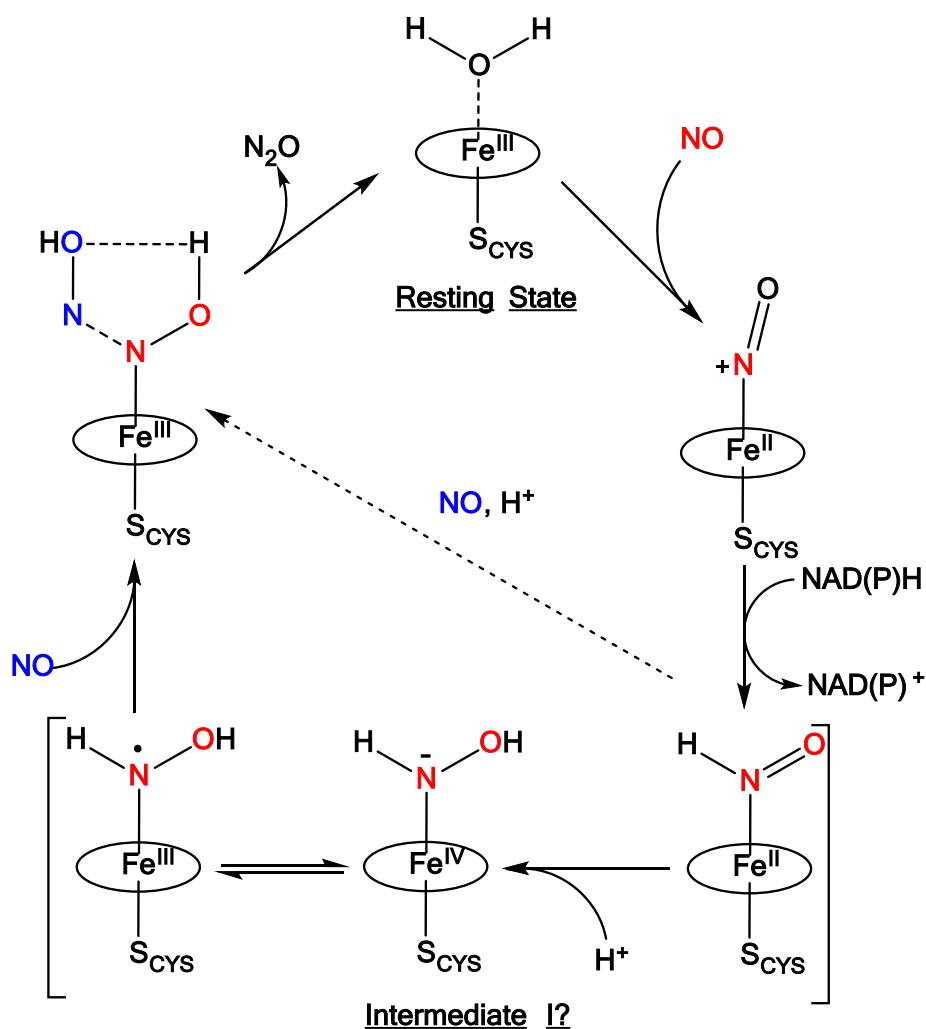


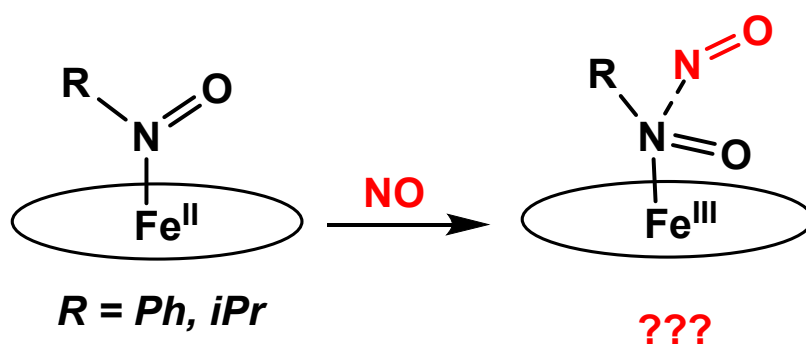
Figure 1. Top: Overlay of the IR spectra of [Fe(TPP)(THF)(PhNO)] (black), and of the ¹⁵N-labeled complex, [Fe(TPP)(THF)(Ph¹⁵NO)] (green), measured in KBr pellets (expanded view of the N-O stretching frequency region.) Bottom: Overlay of the IR spectra of [Fe(TPP)(PhNO)₂] (black) and the ¹⁵N-labeled complex, [Fe(TPP)(Ph¹⁵NO)₂] (blue), measured in KBr pellets. See text for further discussion.





Scheme 1. Proposed mechanism for N₂O formation in Cyt. P450nor. The oval represents a generic porphyrin ligand. Reproduced with permission from ref. ¹. Copyright 2021 American Chemical Society.





Scheme 2. Proposed reactions of iron(II)-RNO complexes with NO to form a ferric hyponitrite-type species. The oval represents a generic porphyrin ligand.



Table 1. Comparison of geometric and vibrational parameters^a and N-O stretching frequencies (cm⁻¹) of experimental and BP86/TZVP-calculated structures of ferrous porphyrin PhNO complexes (P = porphine²⁻).

Complex	$\Delta\text{Fe-N}_p^b$	$\Delta\text{Fe-N(PhNO)}$	$\Delta\text{Fe-X (X = O, N)}$	$\Delta\text{N-O}$	$\angle\text{Fe-N-O}$	$\nu(\text{N-O})$	Ref.
<i>Crystal Structures</i>							
[Fe(TPP)(THF)(PhNO)] ^{c,d}	1.998	1.813	2.141	1.253	122	1368	t.w.
[Fe(3,5-Me-BAFP)(2-MeTHF)(PhNO)]	2.002	1.906	2.088	1.259	125	n.d.	t.w.
[Fe(TPP)(PhNH ₂)(PhNO)] ^{c,d}	2.002	1.994, 1.963	1.994, 1.963	1.172, 1.194	119, 120	1350 ^f	t.w.
[Fe(TPP)(PhNO) ₂] ^c	1.998	1.874, 1.899	-	1.237, 1.227	124, 123	1353	42
[Fe(TPP)(Pyr)(PhNO)]	1.990	1.819	2.106	1.249	124	1342	53
[Fe(TPP)(MI)(PhNO)] ^{c,d}	1.986	1.8	2.029	1.254	125	1336	58
	2.004	1.802, 1.807, 1.812	2.03	1.267, 1.263, 1.26	123, 124, 124	n.d.	
[Fe(TPP)(MI)(<i>o</i> -tolNO)]	2.007	1.841	2.065	1.257	123	1366	59
[Fe(TPP)(Pyr)(NODMA)] ^e	1.981	1.859	2.095	1.252	120	n.d.	58
[Fe(OEP)(MI)(PhNO)] ^c	2.002	1.809, 1.802	2.092, 2.094	1.269, 1.258	123, 124	n.d.	58
<i>Calculated Structures</i>							
[Fe(P)(PhNO)]	-	1.779	-	1.245	123	1383	t.w.
[Fe(P)(THF)(PhNO)]	-	1.795	2.220	1.245	124	1386	t.w.
[Fe(P)(MI)(PhNO)]	-	1.830	2.084	1.246	124	1380	t.w.
[Fe(P)(PhNO) ₂]	-	1.936	1.936	1.239	121	1382	t.w.
[Fe(P)(PhNO)(PhNHOH)]	-	1.814	2.223	1.245, 1.456	110, 123	1386	t.w.

^a All values are given in Å and °; ^b Average value; ^c Data from the disordered structural components; ^d Two different molecules in the unit cell;^e NODMA = 4-nitroso-*N,N*-dimethylaniline; ^f Likely from an impurity of the bis-PhNO complex

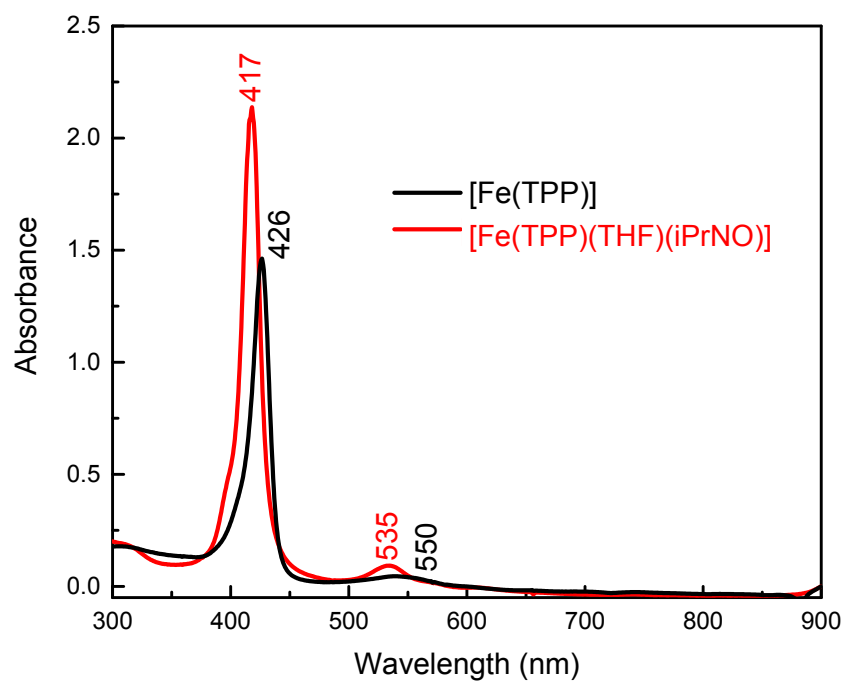


Figure 2. UV-Vis spectra of the precursor, [Fe(TPP)] (black), and of the isolated reaction product, [Fe(TPP)(THF)(iPrNO)] (red), in THF at room temperature.



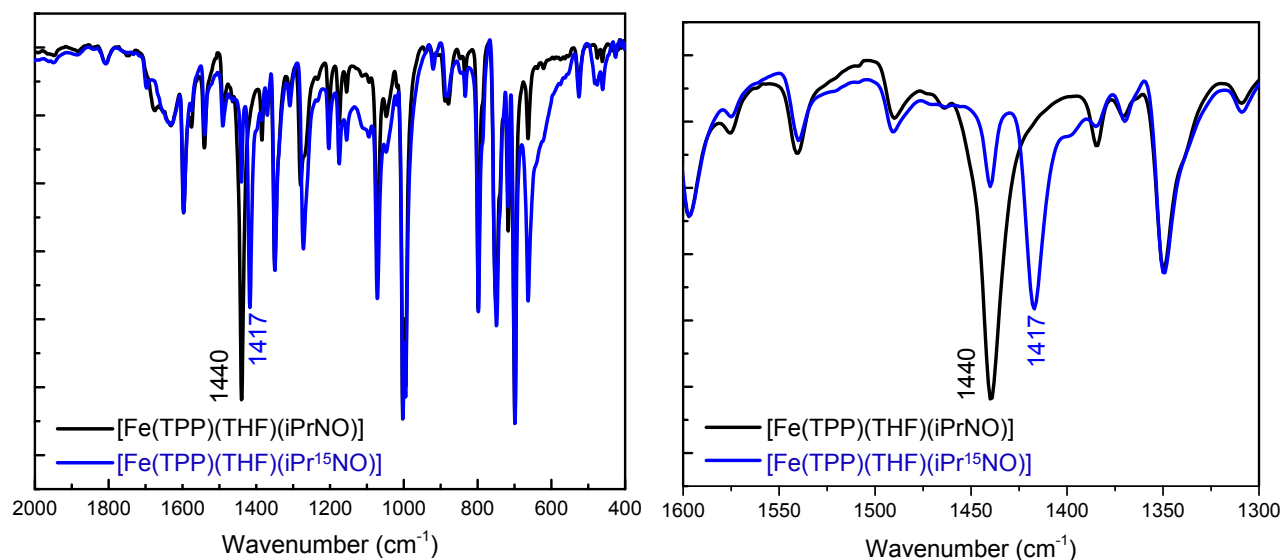


Figure 3. Left: Overlay of the IR spectra of [Fe(TPP)(THF)(iPrNO)] (black) and of the ¹⁵N-labeled complex, [Fe(TPP)(THF)(iPr¹⁵N)] (blue). The data were obtained in KBr pellets. Right: Expanded region of the N-O stretching frequency. The natural abundance isotopes (n.a.i.) complex [Fe(TPP)(THF)(iPrNO)] has one isotope sensitive feature at 1440 cm⁻¹ that shifts to 1417 cm⁻¹ upon ¹⁵N-labeling.



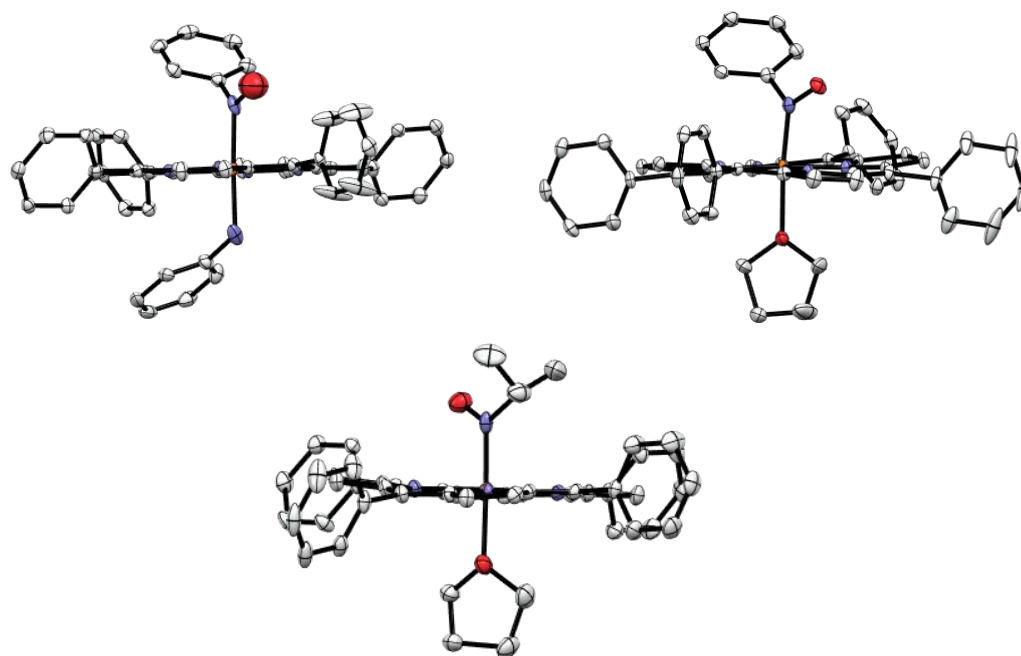


Figure 4. Top: Crystal structures of the two unique ferrous PhNO complexes in the unit cell of the co-crystallized compounds $[\text{Fe}(\text{TPP})(\text{PhNH}_2)(\text{PhNO})]$ (left) and $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ (right). Bottom: Crystal structure of $[\text{Fe}(\text{TPP})(\text{THF})(\text{iPrNO})]$. In the structures, the hydrogen atoms, disorder, and solvent molecules are omitted for clarity. Thermal ellipsoids are shown at 50% probability.



Table 2. Comparison of geometric and vibrational parameters^a and N-O stretching frequencies (cm⁻¹) of experimental and BP86/TZVP-calculated structures of selected ferrous iPrNO/NHO heme complexes (P = porphine²⁻).

Complex	$\Delta\text{Fe-N}_p^b$	$\Delta\text{Fe-N(NO)}$	$\Delta\text{Fe-X (X = O, N)}$	$\Delta\text{N-O}$	$\angle\text{Fe-N-O}$	$\nu(\text{N-O})$	Ref.
<i>Crystal Structures</i>							
[Fe(TPP)(THF)(iPrNO)] ^c	1.998	1.853	2.090	1.212	121	1440	t.w.
[Fe(3,5-Me-BAFP)(iPrNO) ₂] ^c	2.000	1.920	-	1.239	122	1449	t.w.
[Fe(3,5-Me-BAFP)(iPrNH ₂)(iPrNO)] ^c	2.000	1.920	1.959	1.239	122	1449	t.w.
[Fe(TPP)(MI)(iPrNO)]	1.996	1.81	2.050	1.24	122	1433	³²
[Fe(TPP)(iPrNH ₂)(iPrNO)] ^c	1.993	1.86, 1.87	2.105, 2.094	1.26	124	1432	⁵²
[Fe(TTP)(MI)(iPrNO)] ^c	1.997	1.81, 1.86	2.05, 2.03	1.25,	123,	1428	³²
				1.28	121		
[Fe(OEP)(MeOH)(iPrNO)]	2.010	1.89	2.15	1.26	116	1433	³²
[Fe(OEP)(MI)(iPrNO)] ^c	2.014	1.8	2.03, 2.11	1.26	123	1423	³²
[Fe(OEP)(Pyr)(iPrNO)] ^c	2.009	1.84	2.12	1.23	124	1429	³²
<i>Calculated Structures</i>							
[Fe(P)(iPrNO)]	-	1.770	-	1.235	124	1464/(1468, 1261) ^d	t.w.
[Fe(P)(THF)(iPrNO)]	-	1.795	2.245	1.236	124	1461/(1466, 1265) ^d	t.w.
[Fe(P)(MI)(iPrNO)]	-	1.831	2.089	1.237	124	1457/(1464, 1268) ^d	t.w.
[Fe(P)(NHO)]	-	1.741	-	1.235	132	1466	t.w.
[Fe(P)(THF)(NHO)]	-	1.762	2.219	1.235	132	1466	t.w.
[Fe(P)(MI)(NHO)]	-	1.788	2.08254	1.236	132	1459	t.w.

^a Values are given in Å and °; ^b Average value; ^c Data from the disordered structural components; ^d Other vibrations that are coupled to the N-O stretching mode.



Figure 5. View looking down the *meso* carbon atoms of the heme, taken from the crystal structure of [Fe(TPP)(THF)(PhNO)]. The graph clearly shows the out-of-plane distortion of the heme plane. All of the hydrogen atoms and phenyl groups of the porphyrin ligand are omitted for clarity.

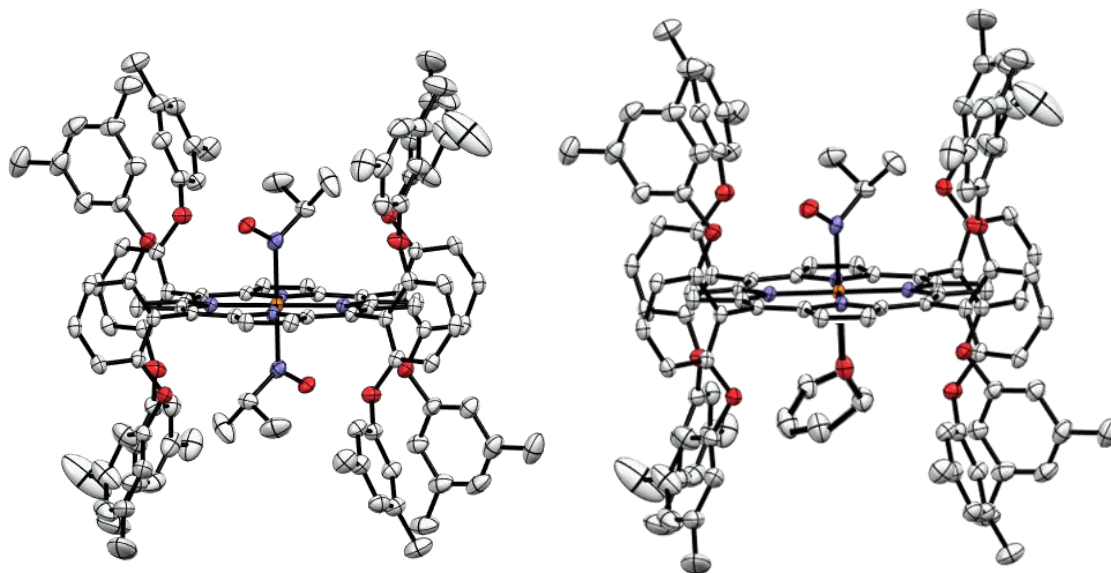


Figure 6. Crystal structures of [Fe(3,5-Me-BAFP)] reacted with iPrNO. Left: [Fe(3,5-Me-BAFP)(iPrNO)₂]. Right: [Fe(3,5-Me-BAFP)(THF)(iPrNO)]. In the crystal, 10-20% of the 6th ligand is THF as shown on the right and 80-90% is the bis-iPrNO complex, shown on the left.



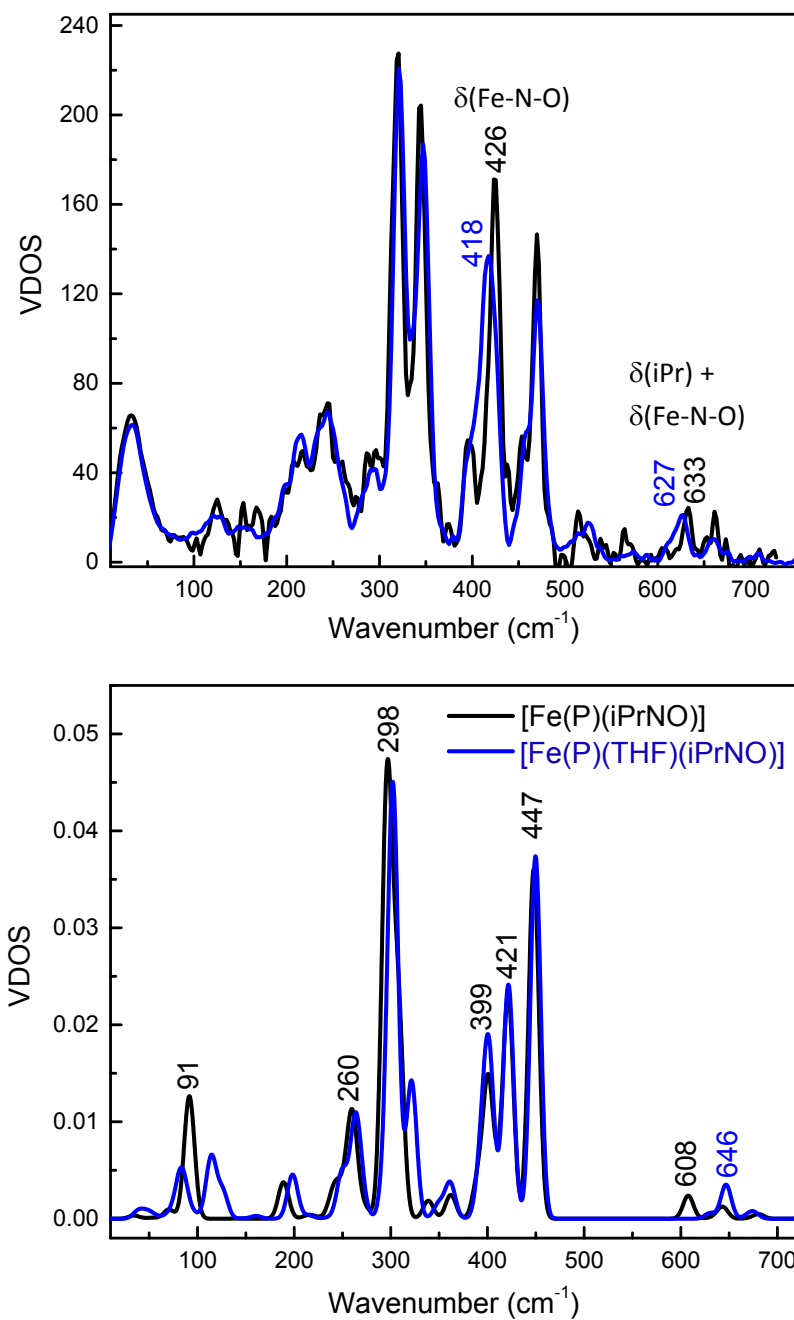


Figure 7. Top: NRVS-derived vibrational density of states (VDOS) for [⁵⁷Fe(TPP)(THF)(iPrNO)] (black) and the ¹⁵N-labeled iPrNO complex (blue). Bottom: BP86/TZVP calculated NRVS VDOS for [Fe(P)(iPrNO)] (black; P²⁻ = porphine dianion) and [Fe(P)(THF)(iPrNO)] (blue).



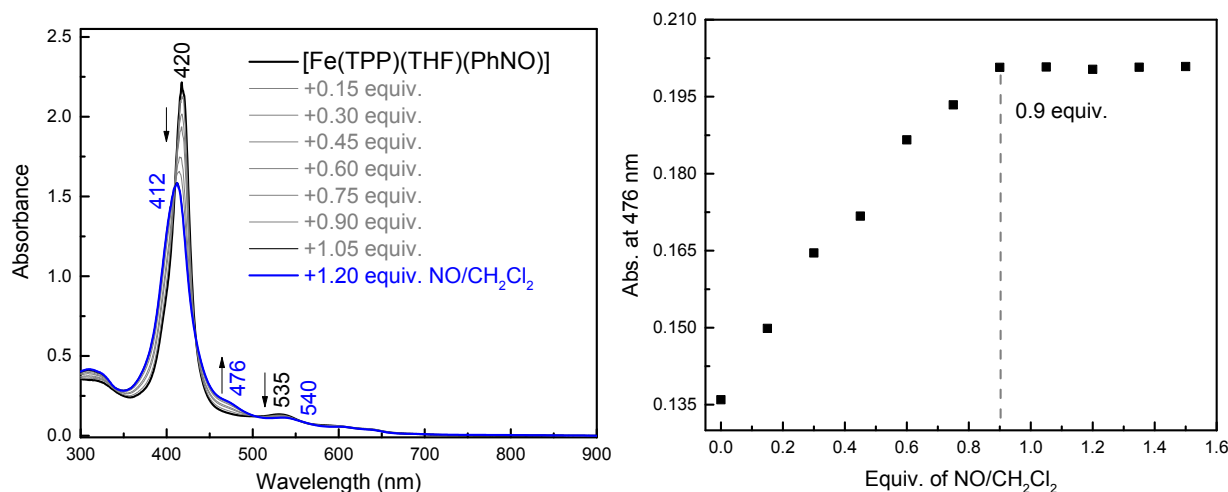
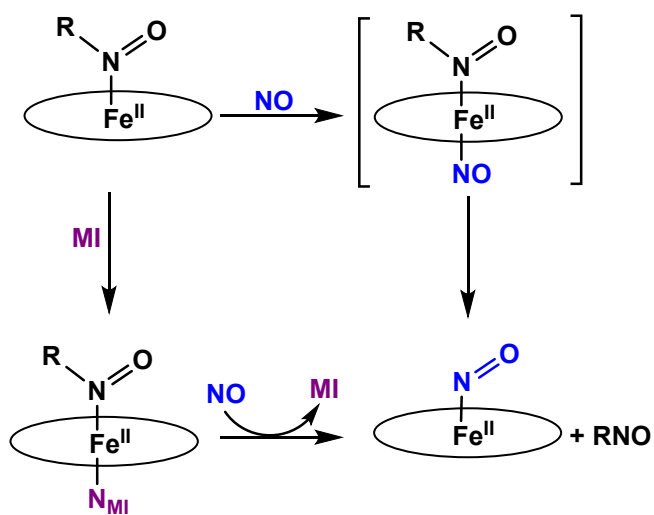


Figure 8. UV-Vis spectra of the titration of $\sim 14 \mu\text{M}$ $[\text{Fe}(\text{TPP})(\text{THF})(\text{PhNO})]$ (black) in THF with a NO-saturated solution of dichloromethane, forming the ferrous NO complex $[\text{Fe}(\text{TPP})(\text{NO})]$ (blue). Right: Changes in the absorption at 476 nm upon addition of the NO solution, showing that the reaction is complete after the addition of ~ 1 equivalent of the NO-saturated dichloromethane solution.





Scheme 3. Summary of the reactivity observed for five- and six-coordinate ferrous heme-RNO (R = IPr, PhNO) complexes with the TPP²⁻ coligand with NO.



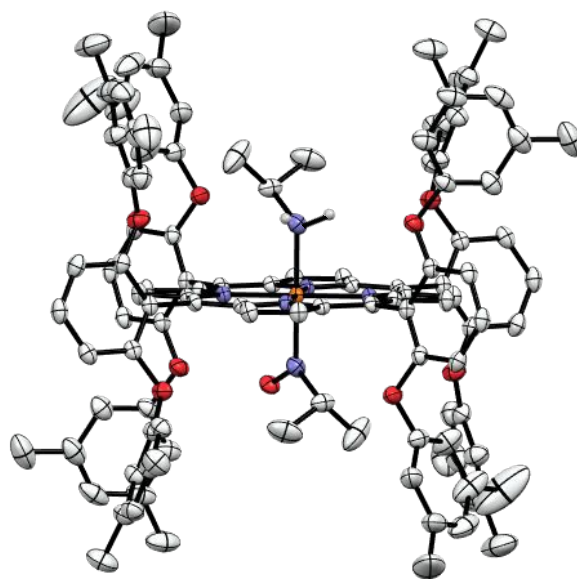


Figure 9. Crystal structure of $[\text{Fe}(3,5\text{-Me-BAFP})(\text{iPrNH}_2)(\text{iPrNO})]$. Hydrogen atoms, except for the amine group of iPrNH_2 , are omitted for clarity. Thermal ellipsoids are shown at 50% probability.



References

- (1) Lehnert, N.; Kim, E.; Dong, H. T.; Harland, J. B.; Hunt, A. P.; Manickas, E. C.; Oakley, K. M.; Pham, J.; Reed, G. C.; Alfaro Sosa, V. The biologically relevant coordination chemistry of iron and nitric oxide: electronic structure and reactivity. *Chem. Rev.* **2021**, *121* (24), 14682-14905.
- (2) Mansuy, D.; Rouer, E.; Bacot, C.; Gans, P.; Chottard, J. C.; Leroux, J. P. Interaction of aliphatic N-hydroxylamines with microsomal cytochrome P450: Nature of the different derived complexes and inhibitory effects on monooxygenases activities. *Biochem. Pharmacol.* **1978**, *27* (8), 1229-1237. DOI: [http://dx.doi.org/10.1016/0006-2952\(78\)90456-2](http://dx.doi.org/10.1016/0006-2952(78)90456-2).
- (3) Bensoussan, C.; Delaforge, M.; Mansuy, D. Particular ability of cytochromes P450 3A to form inhibitory P450-iron-metabolite complexes upon metabolic oxidation of aminodrugs. *Biochem. Pharmacol.* **1995**, *49* (5), 591-602. DOI: [http://dx.doi.org/10.1016/0006-2952\(94\)00477-4](http://dx.doi.org/10.1016/0006-2952(94)00477-4).
- (4) Delaforge, M.; Jaouen, M.; Mansuy, D. Dual effects of macrolide antibiotics on rat liver cytochrome P-450: Induction and formation of metabolite-complexes: A structure-activity relationship. *Biochem. Pharmacol.* **1983**, *32* (15), 2309-2318. DOI: [http://dx.doi.org/10.1016/0006-2952\(83\)90178-8](http://dx.doi.org/10.1016/0006-2952(83)90178-8).
- (5) O'Brien, P. J.; Wong, W. C.; Silva, J.; Khan, S. Toxicity of nitrobenzene compounds towards isolated hepatocytes: dependence on reduction potential. *Xenobiotica* **1990**, *20* (9), 945-955. DOI: 10.3109/00498259009046910.
- (6) Belisario, M. A.; Pecce, R.; Garofalo, A.; Sannolo, N.; Malorni, A. Erythrocyte enzymes catalyze 1-nitropyrene and 3-nitrofluoranthene nitroreduction. *Toxicology* **1996**, *108* (1), 101-108. DOI: [http://dx.doi.org/10.1016/0300-483X\(95\)03293-O](http://dx.doi.org/10.1016/0300-483X(95)03293-O).
- (7) Ringe, D.; Turesky, R. J.; Skipper, P. L.; Tannenbaum, S. R. Structure of the single stable hemoglobin adduct formed by 4-aminobiphenyl in vivo. *Chem. Res. Toxicol.* **1988**, *1* (1), 22-24. DOI: 10.1021/tx00001a003.
- (8) Eyer, P. Reactions of nitrosobenzene with reduced glutathione. *Chem.-Biol. Interact.* **1979**, *24* (2), 227-239. DOI: [http://dx.doi.org/10.1016/0009-2797\(79\)90011-5](http://dx.doi.org/10.1016/0009-2797(79)90011-5).
- (9) Dölle, B.; Töpner, W.; Neumann, H. G. Reaction of aryl nitroso compounds with mercaptans. *Xenobiotica* **1980**, *10* (7-8), 527-536. DOI: 10.3109/00498258009033787.
- (10) Klehr, H.; Eyer, P.; Schäfer, W. On the mechanism of reactions of nitrosoarenes with thiols. Formation of a common intermediate "semimercaptal". *Biol Chem Hoppe Seyler* **1985**, *366* (8), 755-760. PubMed.
- (11) Eyer, P.; Lierheimer, E. Biotransformation of nitrosobenzene in the red cell and the role of glutathione. *Xenobiotica* **1980**, *10* (7-8), 517-526. DOI: 10.3109/00498258009033786.
- (12) Enemark, J. H.; Feltham, R. D. Principles of structure, bonding, and reactivity for metal nitrosyl complexes. *Coord. Chem. Rev.* **1974**, *13*, 339-406.
- (13) Riplinger, C.; Neese, F. The reaction mechanism of cytochrome P450 NO reductase: A detailed quantum mechanics/molecular mechanics study. *ChemPhysChem* **2011**, *12*, 3192-3203.
- (14) McQuarters, A. B.; Wirgau, N. E.; Lehnert, N. Model complexes of key intermediates in fungal cytochrome P450 nitric oxide reductase (P450nor). *Curr. Opin. Chem. Biol.* **2014**, *19*, 82-89. DOI: <http://dx.doi.org/10.1016/j.cbpa.2014.01.017>.
- (15) Lehnert, N.; Praneeth, V. K. K.; Paulat, F. Electronic structure of iron (II)-porphyrin nitroxyl complexes: Molecular mechanism of fungal nitric oxide reductase (P450nor). *J. Comput. Chem.* **2006**, *27* (12), 1338-1351.



- (16) Nomura, T.; Kimura, T.; Kanematsu, Y.; Yamada, D.; Yamashita, K.; Hirata, K.; Ueno, G.; Murakami, H.; Hisano, T.; Yamagiwa, R. Short-lived intermediate in N₂O generation by P450 NO reductase captured by time-resolved IR spectroscopy and XFEL crystallography. *Proc. Natl. Acad. Sci.* **2021**, *118* (21), e2101481118.
- (17) Manickas, E. C.; LaLonde, A. B.; Hu, M. Y.; Alp, E. E.; Lehnert, N. Stabilization of a Heme-HNO Model Complex Using a Bulky Bis-Picket Fence Porphyrin and Reactivity Studies with NO. *J. Am. Chem. Soc.* **2023**, *145* (42), 23014-23026.
- (18) Abucayon, E. G.; Khade, R. L.; Powell, D. R.; Zhang, Y.; Richter-Addo, G. B. Hydride Attack on a Coordinated Ferric Nitrosyl: Experimental and DFT Evidence for the Formation of a Heme Model-HNO Derivative. *Journal of the American Chemical Society* **2016**, *138* (1), 104-107. DOI: 10.1021/jacs.5b12008.
- (19) Abucayon, E. G.; Khade, R. L.; Powell, D. R.; Shaw, M. J.; Zhang, Y.; Richter-Addo, G. B. Over or under: hydride attack at the metal versus the coordinated nitrosyl ligand in ferric nitrosyl porphyrins. *Dalton Transactions* **2016**, *45* (45), 18259-18266, 10.1039/C6DT03860C. DOI: 10.1039/C6DT03860C.
- (20) Farmer, P. J.; Sulc, F. Coordination chemistry of the HNO ligand with hemes and synthetic coordination complexes. *J. Inorg. Biochem.* **2005**, *99* (1), 166-184.
- (21) Kumar, M. R.; Fukuto, J. M.; Miranda, K. M.; Farmer, P. J. Reactions of HNO with heme proteins: New routes to HNO-Heme complexes and insight into physiological effects. *Inorg. Chem.* **2010**, *49* (14), 6283-6292.
- (22) Ling, Y.; Mills, C.; Weber, R.; Yang, L.; Zhang, Y. NMR, IR/Raman, and Structural Properties in HNO and RNO (R = Alkyl and Aryl) Metalloporphyrins with Implication for the HNO-Myoglobin Complex. *J. Am. Chem. Soc.* **2010**, *132* (5), 1583-1591. DOI: 10.1021/ja907342s.
- (23) Mazzeo, A.; Pellegrino, J.; Doctorovich, F. Water-Soluble Nitroxyl Porphyrin Complexes Fe^{II}TPPSHNO and Fe^{II}TPPSNO⁻ Obtained from Isolated Fe^{II}TPPSNO[•]. *J. Am. Chem. Soc.* **2019**, *141*, 18521-18530.
- (24) Choi, I.-K.; Liu, Y.; Feng, D.; Paeng, K.-J.; Ryan, M. D. Electrochemical and spectroscopic studies of iron porphyrin nitrosyls and their reduction products. *Inorg. Chem.* **1991**, *30*, 1832-1839.
- (25) Rahman, M. H.; Ryan, M. D. Redox and spectroscopic properties of iron porphyrin nitroxyl in the presence of weak acids. *Inorg. Chem.* **2017**, *56* (6), 3302-3309.
- (26) Abucayon, E. G.; Khade, R. L.; Powell, D. R.; Zhang, Y.; Richter-Addo, G. B. Hydride Attack on a Coordinated Ferric Nitrosyl: Experimental and DFT Evidence for the Formation of a Heme Model-HNO Derivative. *J. Am. Chem. Soc.* **2016**, *138* (1), 104-107. DOI: 10.1021/jacs.5b12008.
- (27) Goodrich, L. E.; Roy, S.; Alp, E. E.; Zhao, J.; Hu, M. Y.; Lehnert, N. Electronic Structure and Biologically Relevant Reactivity of Low-Spin {FeNO}⁸ Porphyrin Model Complexes: New Insight from a Bis-Picket Fence Porphyrin. *Inorg. Chem.* **2013**, *52* (13), 7766-7780. DOI: 10.1021/ic400977h.
- (28) Rahman, M. H.; Liu, Y.; Ryan, M. D. Proton transfer versus hydrogen bonding in a reduced iron porphyrin nitrosyl complex. *Inorg. Chem.* **2019**, *58* (20), 13788-13795.
- (29) Pellegrino, J.; Bari, S. E.; Bikiel, D. E.; Doctorovich, F. Successful Stabilization of the Elusive Species {FeNO}⁸ in a Heme Model. *Journal of the American Chemical Society* **2010**, *132* (3), 989-995. DOI: 10.1021/ja905062w.
- (30) Kundakarla, N.; Lindeman, S.; Rahman, M. H.; Ryan, M. D. X-ray Structure and Properties of the Ferrous Octaethylporphyrin Nitroxyl Complex. *Inorganic Chemistry* **2016**, *55* (5), 2070-2075. DOI: 10.1021/acs.inorgchem.5b02384.



- (31) Hu, B.; Li, J. One Electron Makes Differences: From Heme {FeNO}₇ to {FeNO}₈. *Angewandte Chemie International Edition* **2015**, *54* (36), 10579-10582. DOI: <https://doi.org/10.1002/anie.201505166>.
- (32) Mansuy, D.; Battioni, P.; Chottard, J. C.; Riche, C.; Chiaroni, A. Nitrosoalkane complexes of iron-porphyrins: analogy between the bonding properties of nitrosoalkanes and dioxygen. *J. Am. Chem. Soc.* **1983**, *105* (3), 455-463. DOI: 10.1021/ja00341a027.
- (33) Mansuy, D.; Beaune, P.; Chottard, J. C.; Bartoli, J. F.; Gans, P. The nature of the “455 nm absorbing complex” formed during the cytochrome P450 dependent oxidative metabolism of amphetamine. *Biochemical Pharmacology* **1976**, *25* (5), 609-612. DOI: [https://doi.org/10.1016/0006-2952\(76\)90398-1](https://doi.org/10.1016/0006-2952(76)90398-1).
- (34) Lee, J.; Chen, L.; West, A. H.; Richter-Addo, G. B. Interactions of Organic Nitroso Compounds with Metals. *Chemical Reviews* **2002**, *102* (4), 1019-1066. DOI: 10.1021/cr0000731.
- (35) Xu, N.; Richter-Addo, G. B. Interactions of Nitrosoalkanes/arenes, Nitrosamines, Nitrosothiols, and Alkyl Nitrites with Metals. In *Progress in Inorganic Chemistry: Volume 59*, John Wiley & Sons, Inc., 2014; pp 381-446.
- (36) Abucayon, E. G.; Powell, D. R.; Richter-Addo, G. B. Carbon–Nitrogen and Nitrogen–Nitrogen Bond Formation from Nucleophilic Attack at Coordinated Nitrosyls in Fe and Ru Heme Models. *Journal of the American Chemical Society* **2017**, *139* (28), 9495-9498. DOI: 10.1021/jacs.7b05209.
- (37) Emhoff, K. A.; Balaraman, L.; Salem, A. M. H.; Mudarmah, K. I.; Boyd, W. C. Coordination chemistry of organic nitric oxide derivatives. *Coordination Chemistry Reviews* **2019**, *396*, 124-140. DOI: <https://doi.org/10.1016/j.ccr.2019.06.011>.
- (38) Herrera, V. E.; Charles, T. P.; Scott, T. G.; Prather, K. Y.; Nguyen, N. T.; Sohl, C. D.; Thomas, L. M.; Richter-Addo, G. B. Insights into Nitrosoalkane Binding to Myoglobin Provided by Crystallography of Wild-Type and Distal Pocket Mutant Derivatives. *Biochemistry* **2023**, *62* (8), 1406-1419. DOI: 10.1021/acs.biochem.2c00725.
- (39) Abucayon, E. G.; Khade, R. L.; Powell, D. R.; Zhang, Y.; Richter-Addo, G. B. Lewis acid activation of the ferrous heme–NO fragment toward the N–N coupling reaction with NO to generate N₂O. *J. Am. Chem. Soc.* **2018**, *140* (12), 4204-4207.
- (40) Adler, A. D.; Longo, F. R.; Kampas, F.; Kim, J. On the preparation of metalloporphyrins. *J. Inorg. Nucl. Chem.* **1970**, *32* (7), 2443-2445. DOI: [http://dx.doi.org/10.1016/0022-1902\(70\)80535-8](http://dx.doi.org/10.1016/0022-1902(70)80535-8).
- (41) McQuarters, A. B.; Goodrich, L. E.; Goodrich, C. M.; Lehnert, N. Disproportionation of O-Benzylhydroxylamine Catalyzed by a Ferric Bis-Picket Fence Porphyrin Complex. *Z. Anorg. Allg. Chem.* **2013**, *639* (8-9), 1520-1526. DOI: 10.1002/zaac.201300125.
- (42) Wang, L.-S.; Chen, L.; Khan, M. A.; Richter-Addo, G. B. The first structural studies of nitrosoarene binding to iron-(II) and -(III) porphyrins. *Chem. Comm.* **1996**, (3), 323-324, 10.1039/CC9960000323. DOI: 10.1039/CC9960000323.
- (43) Paulat, F.; Berto, T. C.; DeBeer George, S.; Goodrich, L.; Praneeth, V. K. K.; Sulok, C. D.; Lehnert, N. Vibrational Assignments of Six-Coordinate Ferrous Heme Nitrosyls: New Insight from Nuclear Resonance Vibrational Spectroscopy. *Inorg. Chem.* **2008**, *47* (24), 11449-11451. DOI: 10.1021/ic801626w.
- (44) Sage, J. T.; Paxson, C.; Wyllie, G. R. A.; Sturhahn, W.; Durbin, S. M.; Champion, P. M.; Alp, E. E.; Scheidt, W. R. Nuclear resonance vibrational spectroscopy of a protein active-site mimic. *J. Condens. Matter Phys.* **2001**, *13* (34), 7707.



- (45) Sturhahn, W. CONUSS and PHOENIX: Evaluation of nuclear resonant scattering data. *Hyperfine Interact.* **2000**, *125* (1), 149-172. DOI: 10.1023/A:1012681503686.
- (46) Defoin, A. Simple Preparation of Nitroso Benzenes and Nitro Benzenes by Oxidation of Anilines with H₂O₂ Catalysed with Molybdenum Salts. *Synthesis* **2004**, *2004* (05), 706-710. DOI: 10.1055/s-2004-815964.
- (47) Leinisch, F.; Jiang, J.; Deterding, L. J.; Mason, R. P. Simplified Synthesis of Isotopically Labeled 5,5-Dimethyl-pyrroline N-Oxide. *Molecules* **2011**, *16* (10). DOI: 10.3390/molecules16108428.
- (48) Sherwin, M. B.; Pichaichanarong, P. Hydrogenation of nitroalkanes to hydroxylamines. Google Patents: 1994.
- (49) Maassen, J. A.; de Boer, T. J. Silver carbonate, a convenient reagent for preparing C-nitroso compounds from hydroxylamines. *Recl.Trav. Chim. Pays-Bas* **1971**, *90* (4), 373-376. DOI: 10.1002/recl.19710900405.
- (50) Gowenlock, B. G.; Luttkie, W. Structure and properties of C-nitroso-compounds. *Quarterly Reviews, Chem. Soc.* **1958**, *12* (4), 321-340, 10.1039/QR9581200321. DOI: 10.1039/QR9581200321.
- (51) Emmons, W. D. The Synthesis of Nitrosoalkane Dimers. *J. Am. Chem. Soc.* **1957**, *79* (24), 6522-6524. DOI: 10.1021/ja01581a043.
- (52) Sohl, C. D.; Lee, J.; Alguindigue, S. S.; Khan, M. A.; Richter-Addo, G. B. Synthesis and solid-state molecular structures of nitrosoalkane complexes of iron porphyrins containing methanol, pyridine, and 1-methylimidazole ligands. *J. Inorg. Biochem.* **2004**, *98* (7), 1238-1246. DOI: <http://dx.doi.org/10.1016/j.jinorgbio.2004.04.008>.
- (53) Perdew, J. P. Density-functional approximation for the correlation energy of the inhomogeneous electron gas. *Phys. Rev. A* **1986**, *33* (12), 8822-8824.
- (54) Becke, A. D. Density-functional exchange-energy approximation with correct asymptotic behavior. *Phys. Rev. A* **1988**, *38* (6), 3098-3100.
- (55) Schäfer, A.; Horn, H.; Ahlrichs, R. Fully optimized contracted Gaussian basis sets for atoms Li to Kr. *J. Chem. Phys.* **1992**, *97* (4), 2571-2577. DOI: <http://dx.doi.org/10.1063/1.463096>.
- (56) Schäfer, A.; Huber, C.; Ahlrichs, R. Fully optimized contracted Gaussian basis sets of triple zeta valence quality for atoms Li to Kr. *J. Chem. Phys.* **1994**, *100* (8), 5829-5835. DOI: <http://dx.doi.org/10.1063/1.467146>.
- (57) *Gaussian 09*; Gaussian, Inc.: Wallingford, CT, USA, 2009. (accessed).
- (58) Godbout, N.; Sanders, L. K.; Salzmann, R.; Havlin, R. H.; Wojdelski, M.; Oldfield, E. Solid-State NMR, Mössbauer, Crystallographic, and Density Functional Theory Investigation of Fe–O₂ and Fe–O₂ Analogue Metalloporphyrins and Metalloproteins. *J. Am. Chem. Soc.* **1999**, *121* (16), 3829-3844. DOI: 10.1021/ja9832820.
- (59) Abucayon, E. G.; Awasabisah, D.; Powell, D. R.; Richter-Addo, G. B. (1-Methyl-1H-imidazole-κN₃)(1-methyl-2-nitrosobenzene-[kappa]N)(5,10,15,20-tetraphenylporphyrinato-κ4N)iron(II) dichloromethane monosolvate. *Acta Cryst. E* **2014**, *70* (2), m51-m52. DOI: doi:10.1107/S160053681400083X.
- (60) Munro, O. Q.; Madlala, P. S.; Warby, R. A. F.; Seda, T. B.; Hearne, G. Structural, Conformational, and Spectroscopic Studies of Primary Amine Complexes of Iron(II) Porphyrins. *Inorg. Chem.* **1999**, *38* (21), 4724-4736. DOI: 10.1021/ic990178q.
- (61) Hu, B.; Li, J. One Electron Makes Differences: From Heme {FeNO}⁷ to {FeNO}⁸. *Angew. Chem. Int. Ed. Engl.* **2015**, *54* (36), 10579-10582. DOI: 10.1002/anie.201505166.



- (62) Immoos, C. E.; Sulc, F.; Farmer, P. J.; Czarnecki, K.; Bocian, D. F.; Levina, A.; Aitken, J. B.; Armstrong, R. S.; Lay, P. A. Bonding in HNO-Myoglobin as Characterized by X-ray Absorption and Resonance Raman Spectroscopies. *J. Am. Chem. Soc.* **2005**, *127* (3), 814-815. DOI: 10.1021/ja0433727.
- (63) Scheidt, W. R.; Frisse, M. E. Nitrosylmetalloporphyrins. II. Synthesis and molecular stereochemistry of nitrosyl-.alpha.,.beta.,.gamma.,.delta.-tetraphenylporphyrinatoiron(II). *J. Am. Chem. Soc.* **1975**, *97* (1), 17-21. DOI: 10.1021/ja00834a005.
- (64) Wyllie, G. R. A.; Schulz, C. E.; Scheidt, W. R. Five- to Six-Coordination in (Nitrosyl)iron(II) Porphyrinates: Effects of Binding the Sixth Ligand. *Inorg. Chem.* **2003**, *42* (18), 5722-5734. DOI: 10.1021/ic034473t.
- (65) Walker, F. A. Nitric oxide interaction with insect nitrophorins and thoughts on the electron configuration of the {FeNO}⁶ complex. *J. Inorg. Biochem.* **2005**, *99* (1), 216-236. DOI: <http://dx.doi.org/10.1016/j.jinorgbio.2004.10.009>.
- (66) Safo, M. K.; Walker, F. A.; Raitsimring, A. M.; Walters, W. P.; Dolata, D. P.; Debrunner, P. G.; Scheidt, W. R. Axial Ligand Orientation in Iron(III) Porphyrinates: Effect of Axial π -Acceptors. Characterization of the Low-Spin Complex [Fe(TPP)(4-CNPy)₂](ClO₄). *J. Am. Chem. Soc.* **1994**, *116* (17), 7760-7770. DOI: 10.1021/ja00096a037.
- (67) Scheidt, W. R.; Reed, C. A. Spin-state/stereochemical relationships in iron porphyrins: implications for the hemoproteins. *Chem. Rev.* **1981**, *81* (6), 543-555. DOI: 10.1021/cr00046a002.
- (68) McQuarters, A. B.; Blaes, E. J.; Kampf, J. W.; Alp, E. E.; Zhao, J.; Hu, M.; Krebs, C.; Lehnert, N. Synthetic model complex of the key intermediate in cytochrome P450 nitric oxide reductase. *Inorg. Chem.* **2019**, *58* (2), 1398-1413.
- (69) Derbyshire, E. R.; Tran, R.; Mathies, R. A.; Marletta, M. A. Characterization of Nitrosoalkane Binding and Activation of Soluble Guanylate Cyclase. *Biochemistry* **2005**, *44* (49), 16257-16265. DOI: 10.1021/bi0515671.
- (70) Denninger, J. W.; Marletta, M. A. Guanylate cyclase and the $\cdot$ NO/cGMP signaling pathway. *Biochim. Biophys. Acta - Bioenerg.* **1999**, *1411* (2-3), 334-350. DOI: [http://dx.doi.org/10.1016/S0005-2728\(99\)00024-9](http://dx.doi.org/10.1016/S0005-2728(99)00024-9).
- (71) Karow, D. S.; Pan, D.; Davis, J. H.; Behrends, S.; Mathies, R. A.; Marletta, M. A. Characterization of Functional Heme Domains from Soluble Guanylate Cyclase. *Biochemistry* **2005**, *44* (49), 16266-16274. DOI: 10.1021/bi051601b.
- (72) Mansuy, D.; Chottard, J. C.; Chottard, G. Nitrosoalkanes as Fe(II) Ligands in the Hemoglobin and Myoglobin Complexes Formed from Nitroalkanes in Reducing Conditions. *Eur. J. Biochem.* **1977**, *76* (2), 617-623. DOI: 10.1111/j.1432-1033.1977.tb11632.x.
- (73) Stone, J. R.; Marletta, M. A. The Ferrous Heme of Soluble Guanylate Cyclase: Formation of Hexacoordinate Complexes with Carbon Monoxide and Nitrosomethane. *Biochemistry* **1995**, *34* (50), 16397-16403. DOI: 10.1021/bi00050a021.
- (74) Romberg, R. W.; Kassner, R. J. Nitric oxide and carbon monoxide equilibria of horse myoglobin and (N-methylimidazole)protoheme. Evidence for steric interaction with the distal residues. *Biochemistry* **1979**, *18* (24), 5387-5392. DOI: 10.1021/bi00591a020.
- (75) Lehnert, N.; Sage, J. T.; Silvernail, N.; Scheidt, W. R.; Alp, E. E.; Sturhahn, W.; Zhao, J. Oriented Single-Crystal Nuclear Resonance Vibrational Spectroscopy of [Fe(TPP)(MI)(NO)]: Quantitative Assessment of the trans Effect of NO. *Inorg. Chem.* **2010**, *49* (15), 7197-7215. DOI: 10.1021/ic1010677.



- (76) Zeng, W.; Silvernail, N. J.; Wharton, D. C.; Georgiev, G. Y.; Leu, B. M.; Scheidt, W. R.; Zhao, J.; Sturhahn, W.; Alp, E. E.; Sage, J. T. Direct probe of iron vibrations elucidates NO activation of heme proteins. *J. Am. Chem. Soc.* **2005**, *127* (32), 11200-11201.
- (77) Yang, L.; Ling, Y.; Zhang, Y. HNO Binding in a Heme Protein: Structures, Spectroscopic Properties, and Stabilities. *J. Am. Chem. Soc.* **2011**, *133* (35), 13814-13817. DOI: 10.1021/ja204072j.
- (78) Praneeth, V. K. K.; Näther, C.; Peters, G.; Lehnert, N. Spectroscopic Properties and Electronic Structure of Five- and Six-Coordinate Iron(II) Porphyrin NO Complexes: Effect of the Axial N-Donor Ligand. *Inorganic Chemistry* **2006**, *45* (7), 2795-2811. DOI: 10.1021/ic050865j.
- (79) Berto, T. C.; Praneeth, V. K. K.; Goodrich, L. E.; Lehnert, N. Iron-porphyrin NO complexes with covalently attached N-donor ligands: Formation of a stable six-coordinate species in solution. *J. Am. Chem. Soc.* **2009**, *131* (47), 17116-17126.
- (80) Lehnert, N.; Scheidt, W. R.; Wolf, M. W. Structure and Bonding in Heme-Nitrosyl Complexes and Implications for Biology. In *Nitrosyl Complexes in Inorganic Chemistry, Biochemistry and Medicine II*, Mingos, D. M. P. Ed.; Springer Berlin Heidelberg, 2014; pp 155-223.
- (81) Lehnert, N.; Berto, T. C.; Galinato, M. G. I.; Goodrich, L. E. The Role of Heme-Nitrosyls in the Biosynthesis, Transport, Sensing, and Detoxification of Nitric Oxide (NO) in Biological Systems: Enzymes and Model Complexes. In *The Handbook of Porphyrin Science*, Kadish, K. M., Smith, K. M., Guilard, R., Eds.; World Scientific: New Jersey, 2011; Vol. 14, pp 1-247.
- (82) Cooper, C. E. Nitric oxide and iron proteins. *Biochim. Biophys. Acta* **1999**, *1411*, 290-309.
- (83) Hoshino, M.; Ozawa, K.; Seki, H.; Ford, P. C. Photochemistry of nitric oxide adducts of water-soluble iron(III) porphyrin and ferrihemoproteins studied by nanosecond laser photolysis. *J. Am. Chem. Soc.* **1993**, *115*, 9568-9575.
- (84) Goodrich, L. E.; Lehnert, N. The *trans* effect of nitroxyl (HNO) in ferrous heme systems: Implications for soluble guanylate cyclase activation by HNO. *J. Inorg. Biochem.* **2013**, *118*, 179-186.
- (85) Spiro, T. G.; Strekas, T. C. Resonance Raman spectra of heme proteins. Effects of oxidation and spin state. *Journal of the American Chemical Society* **1974**, *96* (2), 338-345. DOI: 10.1021/ja00809a004.
- (86) Spiro, T. G. The Resonance Raman Spectroscopy of Metalloporphyrins and Heme Proteins. In *Iron Porphyrins (Part Two)*, Lever, A. B. P., Gray, H. B. Eds.; Addison-Wesley Publishing Company, 1983; pp 89-159.
- (87) Paulat, F.; Praneeth, V. K. K.; Näther, C.; Lehnert, N. Quantum Chemistry-Based Analysis of the Vibrational Spectra of Five-Coordinate Metalloporphyrins [M(TPP)Cl]. *Inorg. Chem.* **2006**, *45*, 2835-2856.
- (88) Traylor, T. G.; Sharma, V. S. Why NO? *Biochemistry* **1992**, *31*, 2847-2849.
- (89) Goodrich, L. E.; Paulat, F.; Praneeth, V. K. K.; Lehnert, N. Electronic Structure of Heme-Nitrosyls and Its Significance for Nitric Oxide Reactivity, Sensing, Transport, and Toxicity in Biological Systems. *Inorg. Chem.* **2010**, *49* (14), 6293-6316. DOI: 10.1021/ic902304a.
- (90) Silvernail, N. J.; Roth, A.; Schulz, C. E.; Noll, B. C.; Scheidt, W. R. Heme Carbonyls: Environmental Effects on ν_{C-O} and $Fe-C/C-O$ Bond Length Correlations. *Journal of the American Chemical Society* **2005**, *127* (41), 14422-14433. DOI: 10.1021/ja053148x.
- (91) Li, J.; Nair, S. M.; Noll, B. C.; Schulz, C. E.; Scheidt, W. R. Relative Axial Ligand Orientation in Bis(imidazole)iron(II) Porphyrinates: Are "Picket Fence" Derivatives Different? *Inorg. Chem.* **2008**, *47* (9), 3841-3850. DOI: 10.1021/ic702498c.



(92) Linder, D. P.; Rodgers, K. R. Structural, Electronic, and Vibrational Characterization of Fe–HNO Porphyrinates by Density Functional Theory. *Inorganic Chemistry* **2005**, *44* (23), 8259-8264. DOI: 10.1021/ic0504745.

(93) Yang, L.; Fang, W.; Zhang, Y. Metal centre effects on HNO binding in porphyrins and the electronic origin: metal's electronic configuration, position in the periodic table, and oxidation state. *Chemical Communications* **2012**, *48* (32), 3842-3844, 10.1039/C2CC31016C. DOI: 10.1039/C2CC31016C.

