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Recent advances in redox-responsive nanoparticles for combined cancer therapy

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The combination of multiple therapeutic modalities has attracted increasing attention as it can achieve better therapeutic effects through different treatment mechanisms. However, traditional small molecule agents are non-specific to the tumor tissue, which leads to off-target toxic effects for healthy tissues. To solve this problem, a number of stimuli-responsive nanoscale drug-delivery systems have been developed. Among these stimuli, a high concentration of reactive oxygen species (ROS) and glutathione (GSH) are characteristic of the tumor microenvironment (TME), which can distinguish it from normal tissue. In this review, we summarize the redox-responsive nanoparticles (NPs) reported in the past three years classified by different functional groups, including GSH-responsive disulfide, ditelluride, and multivalent metal ions, ROS-responsive thioketal, arylboronic ester, aminoacrylate, and bilirubin as well as GSH/ROS dual-responsive diselenide and dicarbonyl thioethers. The prospects and challenges of redox-responsive NPs are also discussed.

1. Introduction

Despite rapid developments in medicinal and pharmaceutical chemistry, cancer treatment is still a major challenge.¹ Recently, as a new approach for curing cancer, phototherapy has demonstrated promising prospects due to its minimal invasiveness and high specificity.^{2–5} However, sole treatment approaches, including chemotherapy or phototherapy, show limitations because of their inherent deficiency and tumor

heterogeneity. For instance, chemotherapy can achieve systemic treatment, but always causes damage to normal tissues, making patients suffer from serious adverse reactions.^{6–9} In addition, the multidrug resistance (MDR) of tumor cells usually emerges after several courses of chemotherapy.¹⁰ Although phototherapy, including photothermal therapy (PTT) and photodynamic therapy (PDT), is efficient for irradiated tumor tissue, phototherapy is not capable of curing metastatic tumors.^{11–15} Therefore, the combination of multiple therapeutic modalities has attracted increasing attention as it can overcome the disadvantages of sole treatment to achieve enhanced therapeutic effects.^{16–20} For example, chemotherapy combined with phototherapy is one of the widely used strategies, and reduces

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the dosage of agents to minimize the side effects of each treatment modality, while chemoradiotherapy is a therapeutic paradigm of various cancers in clinic. In addition, immunotherapy modulates the autoimmune responses to suppress tumor growth and prevent tumor recurrence in combination with other modalities. However, most therapeutic reagents are non-specific to the tumor tissue, which can lead to serious toxic side effects on normal tissues. Nanoscale drug-delivery systems (DDSs) can passively target tumor sites through the enhanced permeability and retention (EPR) effect.^{21–27} NPs co-loaded with anticancer agents with a diameter of 10–200 nm (optimal size selection) tend to accumulate in the tumor tissue, which improves their availability and reduces systemic side effects.^{28,29} However, the premature leakage of cargo during blood circulation remains a major problem hindering the therapeutic efficacy.

To further enhance the precise release of anticancer agents, stimuli-responsive NPs have been extensively designed that can be activated by endogenous and exogenous stimuli.^{30–35} In general, endogenous stimuli refer to the tumor microenvironment (TME), including weak acid, hypoxia, disordered redox species, and overexpressed enzymes.³⁶ Numerous studies have focused on TME-responsive NPs for precise drug delivery to achieve excellent treatment effects.^{37,38} For example, NPs with pH-sensitive bonds or functional groups enabled precise drug release through bond cleavage or functional group isomerization.^{39–43} Hypoxic pathological environments lead to an overexpression of different biological reductases, including nitroreductase (NTR), azoreductase (AZR), and quinone

reductase.^{44,45} The targeting ability of nanomaterials can be significantly improved by incorporating hypoxia-sensitive groups, such as nitroimidazole (NI) and azobenzene (Azo) derivatives to achieve response to biological reductases.^{46,47} Moreover, some overexpressed enzymes, such as aminopeptidase N (APN), matrix metalloproteinases (MMPs), and carbonic anhydrase IX (CAIX), have also been used as the specific stimuli for cancer theranostics.^{48–51}

Among the aforementioned stimuli factors, redox species play an important role in the design of TME-responsive NPs.^{52–55} Reactive oxygen species (ROS) include non-radical species, including hydrogen peroxide (H₂O₂) and singlet oxygen (¹O₂), as well as radical species, such as hydroxyl radicals ([•]OH) and superoxide anion radicals (O₂^{•−}). Substantial evidence has shown that cancer cells produce higher concentration of ROS mainly due to mitochondrial dysfunction.^{56–59} In another aspect, glutathione (GSH) is a reduced biothiol, which is widespread in living organisms.^{60,61} It is worth noting that the concentration of GSH in tumor cells can reach 2–10 mM, which is 7–10 times higher than that in normal tissues.^{62,63} Therefore, both ROS and GSH are specific parameters in cancer cells. Moreover, ROS and GSH can react with specific functional groups (Table 1). Specially, thioketal can be oxidized by ROS to yield two thiol fragments and one acetone molecule, while arylboronic ester is oxidized by H₂O₂ and further hydrolyzes to release the phenol and boric acid, and the β-aminoacrylate linkage undergoes fast oxidative degradation, resulting in cleavage of the ester bond.^{64–66} Besides, bilirubin (BR) turns from a hydrophilic to hydrophobic group in the presence of

Table 1 Redox-responsive groups and their responsiveness behaviors

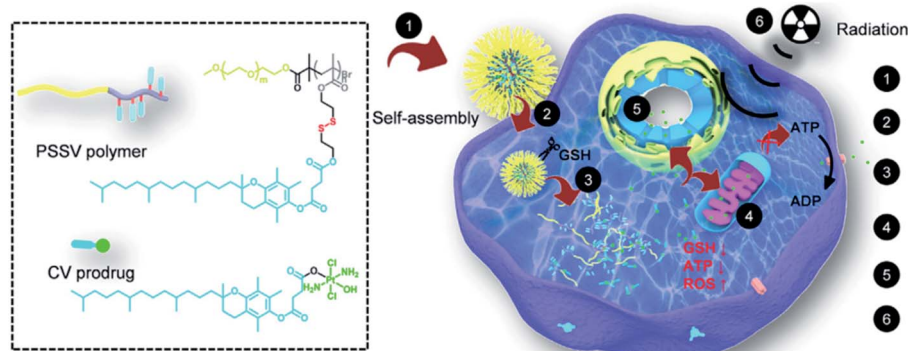
Responsive groups	Stimulus	Responsiveness behavior(s)	Therapeutic modalities	Ref.
Disulfide	GSH	$R^1-S-S-R^2 \xrightarrow{GSH} R^1-SH + HS-R^2$	Chemoradiotherapy PDT, PTT	83 87
Ditelluride	GSH	Similar to disulfide	Chemotherapy	88
Metal ions	GSH	$Fe^{3+} + GSH \longrightarrow Fe^{2+} + GSSG$	PDT, PTT, CDT	92
		$Cu^{2+} + GSH \longrightarrow Cu^+ + GSSG$	PDT, PTT, CDT	93
		$Mn^{4+} + GSH \longrightarrow Mn^{2+} + GSSG$	PTT, CDT	94
Thioketal	ROS	$R^1-S-C(CH_3)_2-S-R^2 \xrightarrow{[O]} R^1-SH + HS-R^2 + CH_3COCH_3$	Chemo-PDT	98
			Chemo-PDT	99
Arylboronic ester	H ₂ O ₂	$R^2O-B(OR^1)-C_6H_4-R^3 \xrightarrow{H_2O_2} R^2O-B(OR^1)-O-C_6H_4-R^3 \xrightarrow{H_2O} HO-C_6H_4-R^3$	PDT, PTT	102
			Chemoradiotherapy	105
Aminoacrylate	ROS	$R-O-C(=O)-CH=CH-N \xrightarrow{[O]} R-OH + O=CH-N$	Chemo-PDT	108
Bilirubin	ROS	Change from hydrophilic to hydrophobic	Chemo-photodynamic-immunotherapy	111
Diselenide	ROS/GSH	$R^1-Se-Se-R^2 \xrightarrow{[O]} R^1-Se(=O)-OH + HO-Se(=O)-R^2$	Chemo-immunotherapy	120
		$R^1-Se-Se-R^2 \xrightarrow{GSH} R^1-SeH + HSe-R^2$	Chemo-photodynamic-immunotherapy	121



Due to the uneven distribution of GSH between tumor cells and extracellular fluid, GSH-responsive moieties as linkers of nanostructured frameworks show promising application in DDSs.⁷⁶ Moreover, GSH, as a reducing agent, undergoes redox reactions with some metal-based nanomaterials. The reduced metal ions are further used in tumor diagnosis and therapy, which has been extensively studied.⁷⁷⁻⁷⁹

Disulfide bonds can be cleaved by biothiols, which are the most common moieties applied in designing GSH-responsive NPs.^{80–82} Commonly, disulfide bonds are introduced into the polymer chain to connect the prodrug molecules or as a carrier

However, polymer-based redox-responsive prodrugs or photophotosensitizers have demonstrated some drawbacks, including complex synthetic modifications and a low loading efficiency. In order to solve these problems, carrier-free nano-systems based on the self-assembly of amphiphilic small molecules have been developed.^{85,86} Pei and co-workers⁸⁷ synthesized amphiphilic small molecules consisting of a hydrophobic photosensitizer and a hydrophilic lactose, which was linked by GSH-responsive disulfide bonds (Fig. 2a). The amphiphilic small molecules enabled the self-assembly into NPs, targeting Hepg-2 cells through the lactose receptor on the



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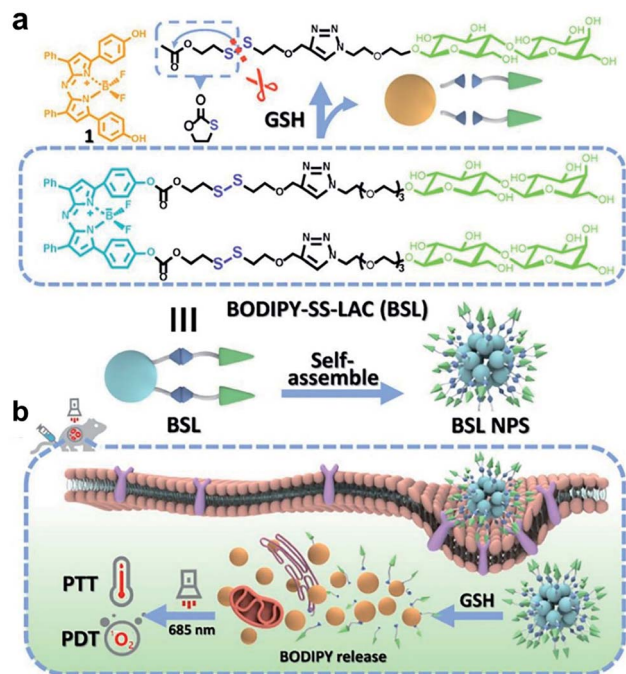


Fig. 2 (a) Chemical structure, self-assembly, and GSH responsiveness of BSL. (b) Schematic representation of BSL NPS for lactose-mediated endocytosis, intracellular BODIPY release triggered by GSH, and PDT with a NIR light of 685 nm wavelength.⁸⁷ Reproduced with permission from ref. 87, copyright © 2021, Elsevier Ltd.

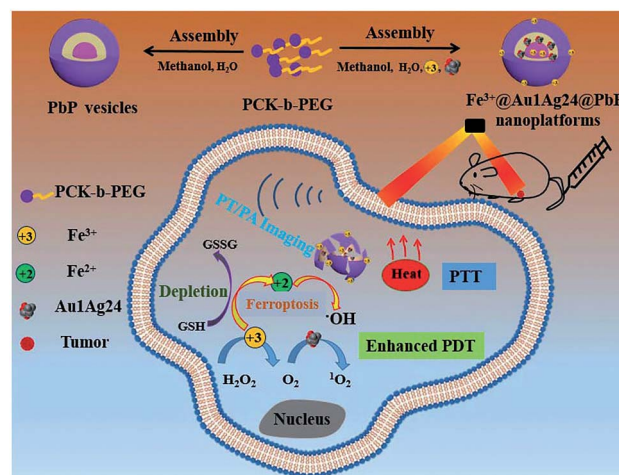


Fig. 4 Schematic illustration of the formation of the $\text{Fe}^{3+}@\text{Au}_1\text{Ag}_{24}@\text{PbP}$ nanoplatform and its imaging-guided synergistic anticancer therapy.⁹² Reproduced with permission from ref. 92. Copyright © 2021, The Royal Society of Chemistry.

cell membrane and the responsiveness to GSH. The NPs released the phototherapeutic agent BODIPY due to the cleavage of disulfide bonds by overexpressed GSH at the tumor site; thereby realizing PTT and PDT under the irradiation of a 685 nm laser (Fig. 2b). The carrier-free multifunctional NPSS exhibited good biocompatibility, enhanced enrichment in the tumor, and an excellent anticancer effect.

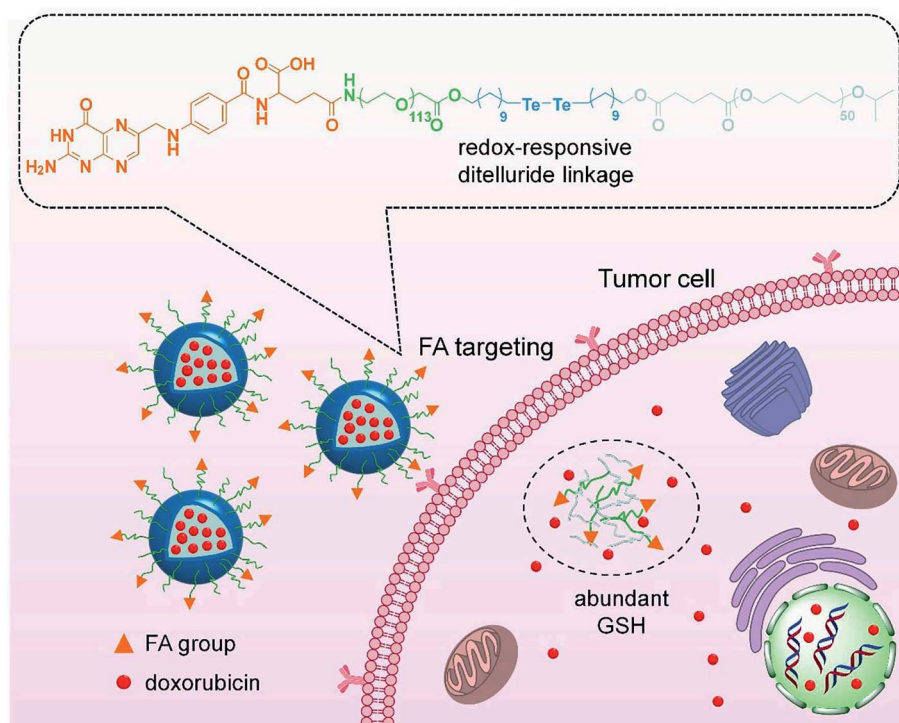


Fig. 3 Schematic of targeted drug delivery and GSH-responsive drug release.⁸⁸ Reproduced with permission from ref. 88, copyright © 2020, Frontiers Media SA.

2.2. Ditelluride bond-based GSH-responsive NPs

As a chalcogen, the chemical properties of tellurium are similar to sulfur. Compared with disulfide bonds, ditelluride bonds are more easily responsive to reducing substances, such as GSH, due to the lower bond energy. Although tellurium-containing polymers have been widely reported, nanoparticles based on ditellurium bonds are still in their infancy. In 2020, there was one case of ditellurium-based nanocarriers for sole chemotherapy. Sun and co-workers⁸⁸ prepared a ditelluride-containing PEGylated polycaprolactone modified with folic acid (FA) for active tumor targeting (Fig. 3). The polymer self-assembled to encapsulate DOX in aqueous solution, and the NPs exhibited GSH-responsive drug release due to the degradation of the

ditelluride bonds. FA could promote the uptake of nanoparticles by 4T1 breast cancer cells, promoting the accumulation of the NPs at the tumor site and further enhancing the growth inhibitory effect on 4T1 tumors. Combination therapy based on ditelluride-containing polymers is expected to be carried out in the future.

2.3. Metal-ion-based GSH-responsive NPs

Besides, multivalent metal ions exhibit GSH responsiveness *via* redox reactions. The reduced metal ions can undergo a Fenton/Fenton-like reaction and react with H₂O₂ to generate more cytotoxic ·OH for chemodynamic therapy (CDT).^{89–91} Common metal ions include Fe³⁺/Fe²⁺, Cu²⁺/Cu⁺, and Mn⁴⁺/Mn²⁺. Zhu

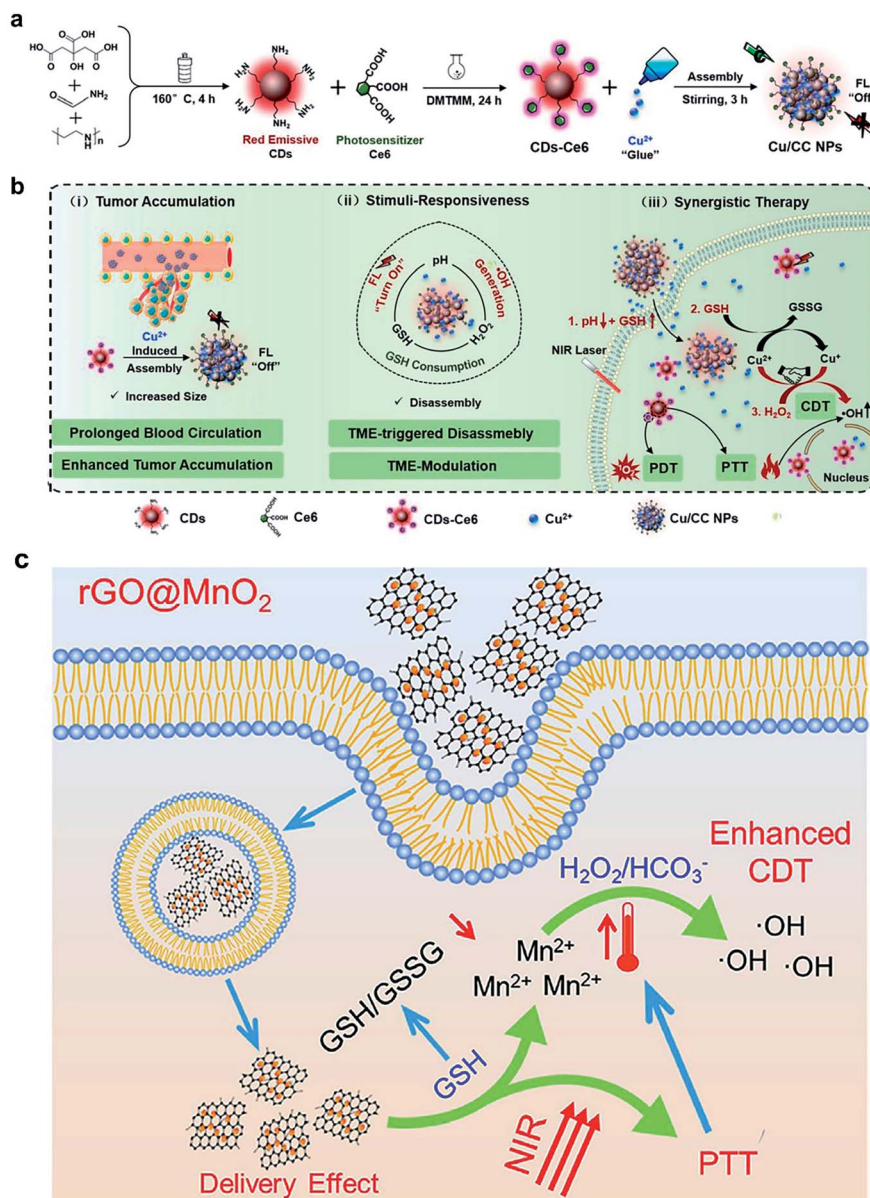


Fig. 5 Illustration of (a) the synthesis process of Cu/CC NPs and (b) their features for enhancing tumor accumulation, TME stimuli-response, and synergistic therapy.⁹³ (c) rGO@MnO₂-mediated PTT and photothermal/delivery effect enhanced CDT for synergistic cancer therapy.⁹⁴ Reproduced with permission from ref. 93 and 94. Copyright © 2020, Wiley-VCH. Copyright © 2021 Elsevier Ltd.

and co-workers⁹² prepared an amphiphilic block copolymer *via* grafting polyethylene glycol (PEG) onto pH-responsive polyketal (PCK), which readily self-assembled into a vesicle to load the oil-soluble metal nanoclusters Au₁Ag₂₄ as photosensitizers and water-soluble Fe³⁺. In this work, the composite NPs with pH and GSH dual-responsive properties exhibited synergetic PTT and PDT under near-infrared light irradiation. Meanwhile, the Fe³⁺ ions acted as oxidants to consume GSH in order to prevent ¹O₂ quenching. Besides, the formed Fe²⁺ ions underwent a Fenton reaction with endogenous H₂O₂ in tumors to generate [•]OH for ferroptosis-based tumor therapy (Fig. 4). The multi-responsive, multimodal nanotherapeutic platform was confirmed as an effective cancer therapy both *in vitro* and *in vivo*.

Several other multivalent metal-ion pairs containing Cu²⁺/Cu⁺ and Mn⁴⁺/Mn²⁺ also display similar functions to Fe ions. Lin and co-workers⁹³ developed a responsive diagnosis and treatment integrated nanoplatform *via* the assembly of Ce6-modified carbon dots (CDs) with Cu²⁺ ions (Fig. 5a). The nanosystem showed quenched fluorescence and photosensitization due to the aggregation of Ce6, and the fluorescence imaging and photosensitization were restored once activated by pH/GSH. In addition, the introduction of Cu²⁺ provided additional CDT through reaction with H₂O₂, and also enhanced intracellular oxidative stress by consuming the tumor's intracellular GSH. The TME-responsive NPs offer a new strategy for imaging-guided multiple therapeutic modalities, including PDT, PTT and chemodynamic therapy (Fig. 5b). Bianco and co-workers⁹⁴ synthesized a GSH-responsive composite by anchoring negatively charged BSA-modified MnO₂ NPs on the surface of polyethyleneimine-modified reduced graphene oxide nanosheets (rGO NSs) *via* electrostatic interactions. The MnO₂ NPs were reduced to Mn²⁺ ions by intracellular GSH, which converted H₂O₂ to [•]OH *via* a Fenton-like reaction for CDT and the rGO NSs further killed tumor cells due to PTT under NIR light irradiation (808 nm). Meanwhile, the high temperature induced by the photothermal conversion increased the rate of the Fenton-like reaction and enhanced the efficiency of CDT (Fig. 5c). Interestingly, the nanosystems exhibited a high lethality at low concentrations due to the presence of rGO, which facilitated the cellular uptake of MnO₂ NPs, thus further improving the biosafety.

3. ROS-responsive NPs for combined cancer therapy

A high level of ROS, especially H₂O₂, is a characteristic of TME.^{95,96} The elevated H₂O₂ concentration in tumor tissue possess a strong oxidative capacity, making it a suitable stimulus for responsive NPs. The development of ROS-responsive nanoplatforms has attracted considerable attention.⁶⁴ Hence, we review several representative nanoplatforms based on thio-ketal-, arylboronic ester-, and aminoacrylate moieties.

3.1. Thioketal-based ROS-responsive NPs

Thioketal is a sulfur-based ROS-responsive moiety that has been shown to react with most ROS, including H₂O₂, ¹O₂, [•]OH, and

O₂^{•-}.⁹⁷ Similar to disulfide bonds, thioketals are often grafted onto polymer chains to form polymer-based prodrugs or are used to link hydrophilic and hydrophobic prodrugs to generate amphiphilic small molecules. Sun and co-workers⁹⁸ synthesized an amphiphilic small molecule with the thioether- or thioketal-linked hypoxia-activated drugs PR104A and pyropheophorbide a (PPa). The amphiphilic small molecules self-assembled into nanoprodrugs that possessed a high dual drug-loading efficiency. Due to aggregation caused quenching (ACQ), 660 nm laser-excited PPa preferentially transferred electrons to the sulfur atoms of adjacent thioether or thioketal bonds *via* the photoinduced electron-transfer (PET) effect to form sulfur radical cations rather than generating ¹O₂, which led to the cleavage of C–S bonds and the release of PR104A and PPa. With the dissociation of the nanoprodrug, the ACQ effect of PPa was relieved and ROS were generated under light to further accelerate the cleavage of the thioether or thioketal bonds, releasing PR104A and PPa. Additionally, aggravated hypoxia by PDT activated the efficacy of the prodrug PR104A for combination chemo-photodynamic therapy (Fig. 6a). Qian and co-workers⁹⁹ incorporated thioketal within the PEG chain to connect camptothecin (CPT) and grafted the photosensitizer PPa into the polymer to construct a ROS-responsive nanoprodrug system (Fig. 6b). The self-assembly of the conjugate prevented drug leakage during systemic circulation, and the PPa fluorescence signal precisely tracked NPs at the tumor sites. PPa generated ROS under near-infrared laser irradiation, which cleaved thioketal to release CPT in a controllable and on-demand manner. The combined CPT-mediated chemotherapy and PPa-induced PDT suppressed the tumor growth.

3.2. Arylboronic ester-based ROS-responsive NPs

Arylboronic ester is also a common ROS-responsive group used in the design of stimuli-responsive nanomaterials. H₂O₂ nucleophilically attacks the boron center, which is further hydrolyzed to release phenol and boric acid.^{100,101} The nanoplatform-installed arylboronic ester moieties were degraded to release the cargo in the presence of H₂O₂. Cui and co-workers¹⁰² reported a light and ROS dual-responsive multistage nanocarrier consisting of phenylboronic acid-crosslinked polyethylene glycol-polycaprolactone, indocyanine green (ICG), and pyridine endoperoxide (PE) (Fig. 7a). Here, ICG acted as both a photosensitizer and a photothermal agent to generate ROS and heat. The excessive ROS in the tumor broke the phenylboronic acid to accelerate the release of ICG and PE, and the elevated temperature induced PE to continuously release oxygen for relieving deep tumor hypoxia to promote PDT. The nanocarriers generated multistage responses *in situ* to induce tumor double apoptosis, maximizing the therapeutic effect of PTT/PDT.

Arylboronic ester is a component of a proalkylating agent that can be activated by H₂O₂ and can irreversibly deplete GSH to increase oxidative stress in tumor cells.^{103,104} Yan and co-workers¹⁰⁵ synthesized silica-coated bismuth NPs with premixed H₂O₂-responsive arylboronic ester-containing prodrugs and lauric acid on the surfaces. Lauric acid was melted under near-infrared light irradiation to release the prodrug, which was



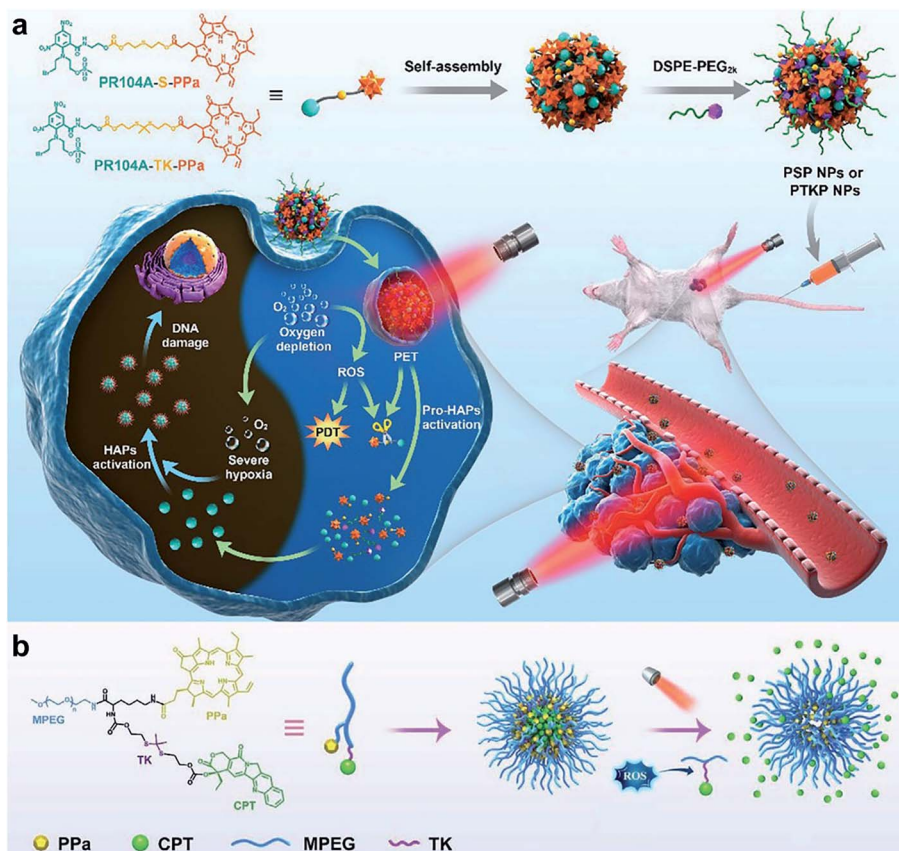


Fig. 6 (a) Schematic illustration of the preparation of self-assembled heterodimeric prodrug-NPs, the light-triggered dual-modality activation of the prodrug-NPs, and the PDT-induced activation of HAPs.⁹⁸ (b) Schematic illustration of the MPEG-(TK-CPT)-PPa NPs for photo-triggered ROS-activatable CPT prodrug release and local tumor therapy.⁹⁹ Reproduced with permission from ref. 98 and 99. Copyright © 2020, The Royal Society of Chemistry. Copyright © 2020, Wiley-VCH.

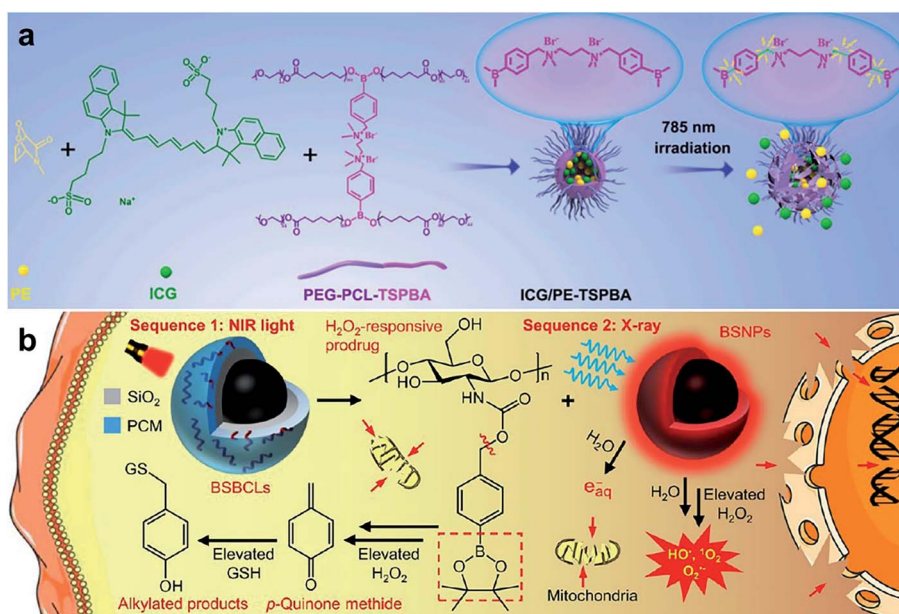


Fig. 7 (a) Fabrication and responsive breakage of the nanocarrier ICG/PE-TSPBA.¹⁰² (b) Schematic illustration of the BBSLs NPs for NIR- and X-ray-triggered ROS-activatable prodrug release.¹⁰⁵ Reproduced with permission from ref. 102 and 105. Copyright © 2021, Elsevier Ltd. Copyright © 2021, American Chemical Society.



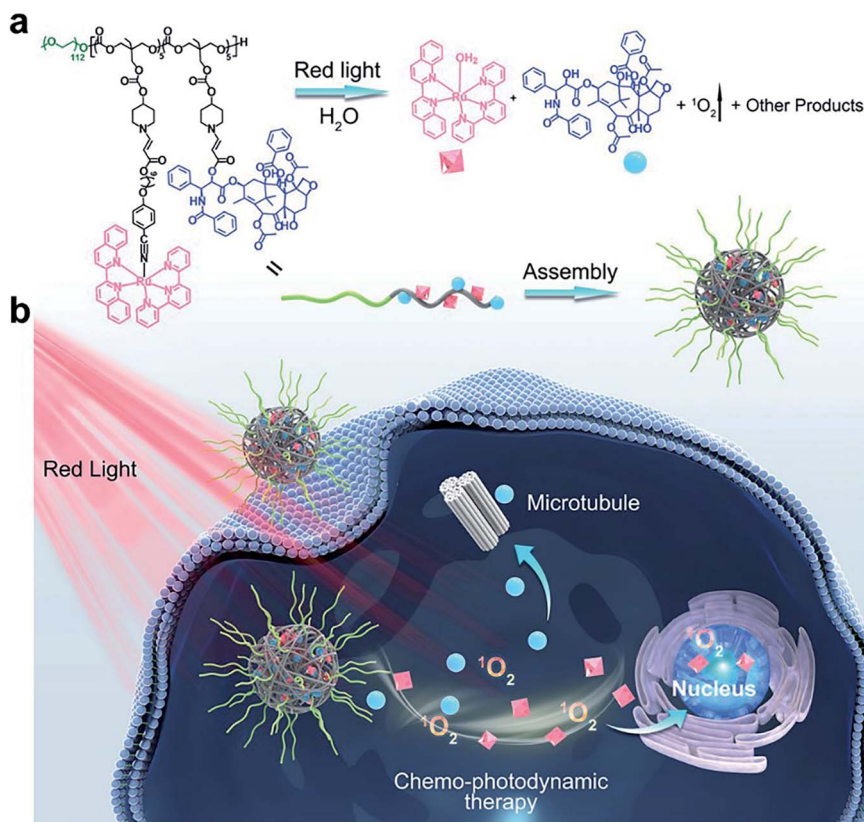


Fig. 8 (a) Chemical structure of the amphiphilic copolymer poly(Ru/PTX). Red-light irradiation induced cleavage of the Ru complex, the generation of ¹O₂, and release of the anticancer drug PTX. (b) Schematic illustration of the self-assembly, cell internalization, and chemo-photodynamic therapy using poly(Ru/PTX).¹⁰⁸ Reproduced with permission from ref. 108, copyright © 2021, Wiley-VCH.

converted into the electrophilic *p*-quinone methyl ether in the presence of H₂O₂, irreversibly depleting GSH and enhancing tumor oxidative stress. The generated heat promoted blood flow

into the tumor tissue and relieved the hypoxic microenvironment. Bismuth NPs, as radiosensitizers, generated a large amount of ROS under X-ray irradiation, synergistically inducing

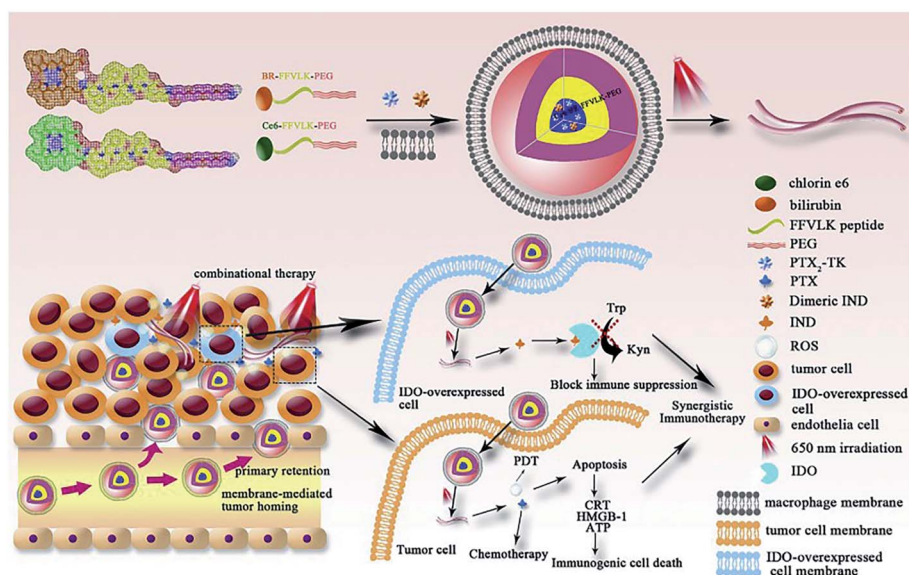


Fig. 9 Illustration of the construction of I-P@NPs@M and its function in the tumor region.¹¹¹ Reproduced with permission from ref. 111, copyright © 2020, Elsevier. Ltd.

cell death through DNA fragmentation and mitochondria-mediated apoptosis (Fig. 7b). This strategy effectively inhibited tumor growth *in vivo* with high tumor specificity and reduced side effects.

3.3. Aminoacrylate-based ROS-responsive NPs

Aminoacrylate is another ROS-responsive group, which is sensitive to $^1\text{O}_2$. The β -aminoacrylate linkage undergoes fast oxidative degradation, resulting in the cleavage of ester bonds.^{106,107} Following the mechanism, our group¹⁰⁸ synthesized a red-light-responsive metallopolymer with a biodegradable backbone structure containing a Ru complex as the photosensitizer and a chemo-drug paclitaxel (PTX) linked *via* $^1\text{O}_2$ activatable aminoacrylate linker (Fig. 8a). Under the irradiation of 650 nm red light, the cyano coordination bond between the polymer and the Ru ligand was cut, which then released the Ru complex to generate $^1\text{O}_2$ for PDT. Subsequently, the generated $^1\text{O}_2$ triggered cleavage of the β -aminoacrylate linker, resulting in the release of PTX for chemotherapy (Fig. 8b). The sequential dual-drug-release strategy minimized the premature drug release *in vivo* and enabled combined chemotherapy and PDT for enhanced anticancer efficiency.

3.4. Bilirubin-based ROS-responsive NPs

Bilirubin (BR) is an endogenous small molecule that can be converted from hydrophilic to hydrophobic with ROS. In some reports, BR as a ROS-responsive group was used to design nanocarriers for controlled anticancer agent delivery.^{109,110} Gao and co-workers¹¹¹ used BR or Ce6 as the hydrophobic end, PEG as the hydrophilic tail, and the polypeptide chain for hydrogen bond formation to synthesize two amphiphilic triblock molecules, which co-assembled into micelles to encapsulate the prodrug PTX and pro-immune reagent indoximod (IND). The NPs were further coated on the macrophage membrane to facilitate tumor targeting. ROS were generated by Ce6 upon 650 nm laser irradiation, triggering the transformation of the micelles into nanofibers and facilitating the release of anticancer agents. Chemophotodynamic therapy significantly inhibited tumor proliferation and triggered immunogenic cell death (ICD) effects, which were combined with IND to activate the immune response (Fig. 9).

4. ROS and GSH dual-responsive NPs for combined cancer therapy

Due to the heterogeneity of tumors, different concentrations of ROS and GSH are found in various tumors, and even different GSH/ROS levels are found in different regions of the same tumor.¹¹² Thus, GSH or ROS single-responsive DDSs may be limited in selectivity and response efficiency.^{113,114} Compared with single-responsive NPs, redox dual-triggered NPs exhibit a faster response rate and higher drug-release efficiency. Therefore, DDSs with dual responsiveness are more promising in diverse pathological conditions.^{115,116} Some chemical moieties can react with ROS and GSH simultaneously, such as diselenide and α -dicarbonyl thioether, which are utilized for the design of responsive NPs.

4.1. Diselenide-based ROS and GSH dual-responsive NPs

As known, diselenide linkers can not only be cleaved and oxidized to seleninic acid by ROS, but also reduced to selenol in the presence of a reducing agent.^{117,118} So far, diselenide bonds have been widely used in inorganic and polymeric nanomaterial frameworks for the controlled release of cargoes. Shao and co-workers¹¹⁹ designed diselenide-bond-bridged mesoporous silica nanoparticles (MSNs) that possessed redox dual-responsive properties for cancer therapeutics (Fig. 10a). The cytotoxic RNase A was encapsulated into the pores of diselenide bond-bridged MSNs and released *via* matrix degradation in the presence of ROS or GSH. The NPs were endowed with homologous targeting features after being coated on the cancer cell membrane. The diselenide bond-bridged MSNs exhibited high anticancer properties *in vitro* and *in vivo*. The research group¹²⁰ found that diselenide bond-bridged MSNs also possessed X-ray responsiveness. The MSNs showed material degradation to control the release of DOX with low-dose X-ray irradiation, which resulted in an ICD effect for chemo-immunotherapy to inhibit tumor growth and metastasis (Fig. 10b). Further, our group¹²¹ utilized the diselenide bond-bridged MSNs to co-encapsulate the chemo-drug DOX and photosensitizer methylene blue (MB) for red-light-responsive cascade-amplifying chemo-photodynamic-immunotherapy. Under 660 nm red-light irradiation, MB generated ROS, which resulted in a collapse of the organosilica matrix and release of the dual drug for combined chemotherapy and PDT. Interestingly, the cascaded chemophotodynamic therapy enhanced ICD and the antitumor immune responses, which suppressed tumor growth, metastasis, and recurrence combined with anti-PD-1 (Fig. 10c).

5. Conclusion and perspective

Redox-responsive NPs have exhibited great potential in DDSs and targeted tumor combined therapy due to their easy preparation, specificity, easy modifiability, and low toxicity. In the past three years, researchers have made great efforts to develop GSH-responsive NPs based on disulfide bonds, ditelluride bonds, and metal ions, ROS-responsive NPs based on thioketal, arylboronic ester, aminoacrylate, and BR, as well as ROS/GSH dual-responsive NPs containing diselenide and dicarbonyl thioether groups. These reviewed examples demonstrated excellent anticancer effects both *in vitro* and *in vivo*. Causing oxidative stress can result in tumor cell apoptosis through disruption of the redox balance in cancer cells; thus, the modulation of ROS production combined with ROS-activated nanomedicine could trigger cascades of the therapeutic effects on tumor tissues. Therefore, many ROS-responsive nanocarriers have been developed and made significant progress in cancer therapy. In addition, given that GSH plays a role in the quenching of ROS, a strategy to amplify oxidative stress by promoting the generation of ROS and the consumption of GSH was proposed to inhibit tumor growth. These designs have attracted great interest in controlled drug release to enhance therapeutic efficacy.



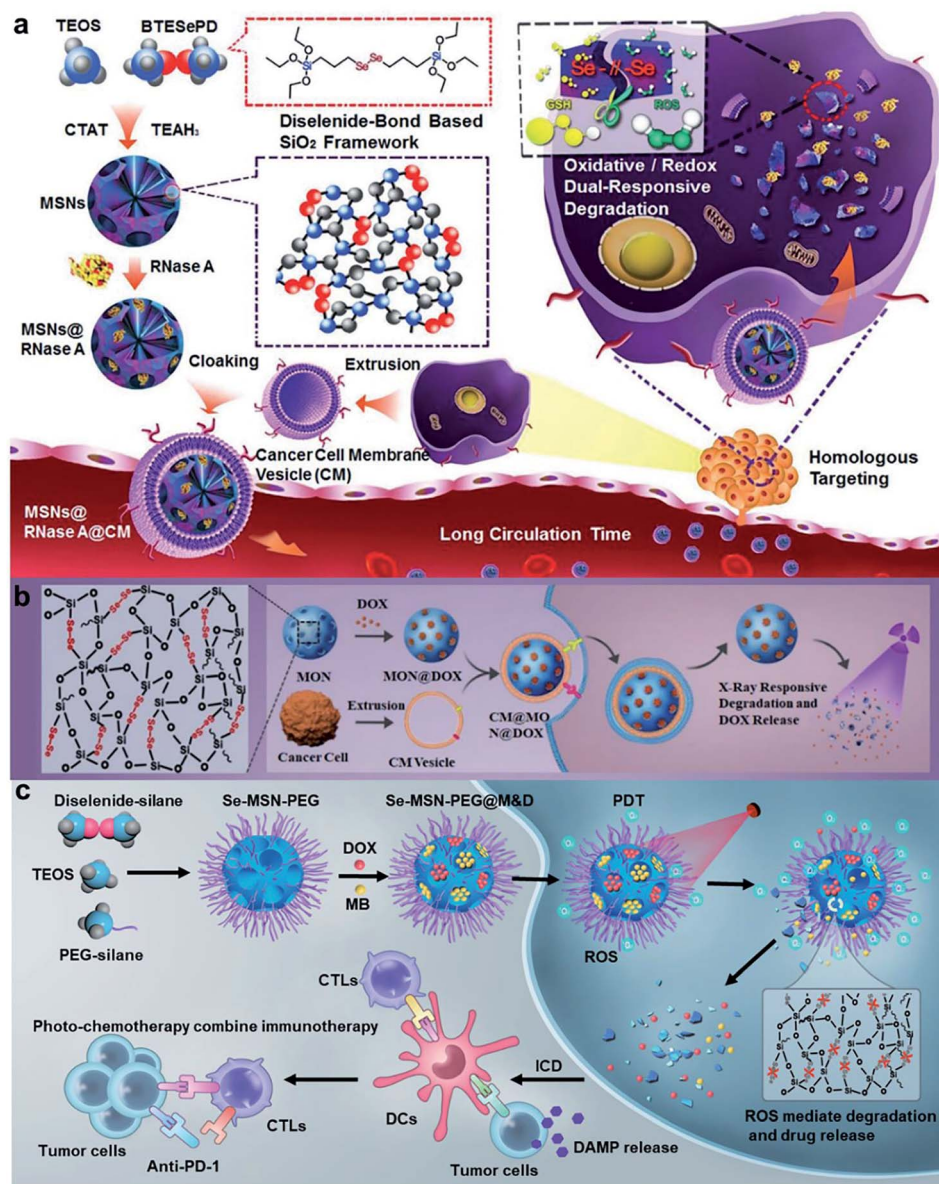


Fig. 10 (a) Schematic illustration of the synthesis procedure of biodegradable diselenide-bridged MSNs and their application for dual-responsive, cancer cell-membrane-mimetic protein delivery.¹¹⁹ (b) Schematic of the synthesis of diselenide-bond-bridged MONs for low-dose X-ray radiation-controllable drug release.¹²⁰ (c) Schematic of the synthetic procedure of Se-MSN-PEG with cascading drug release and amplifying ICD manners and their application for efficient and safe cancer chemo-photo-immunotherapy.¹²¹ Reproduced with permission from ref. 119, 120 and 121. Copyright © 2018, 2020, Wiley-VCH. Copyright © 2022, Elsevier Ltd.

Despite the rapid development of redox-responsive nano-platforms, the research is still in the preclinical stage with many obstacles for clinical use. First, although redox-responsive NPs have exhibited better antitumor ability than common DDSs due to their specificity to redox conditions, their targeting ability mainly depends on the EPR effect, which requires further improvement. Studies have shown that combining tumor-specific ligands or cancer-cell-derived membrane with DDSs can largely enhance the tumor-targeting ability.^{122,123,124,125,126} Thus, redox-responsive NPs with enhanced tumor-targeting ability are expected to be investigated and developed in the near future. Second, the biosafety of NPs is a matter of great concern. The biocompatibility of most existing NPs was only

investigated during the treatment. The fate of the NPs *in vivo* after the therapeutic agents are released needs to be studied, including their metabolic pathways and long-term side effects. Only redox-responsive NPs with excellent biosafety can be confidently used in future clinical research. Third, the tumor models used in the current research were constructed in mice, but the microenvironment of human tumors is different. In order to achieve clinical therapeutic use, it is necessary to better understand the human tumor microenvironment and to design related redox-responsive NPs. With the enhancement of tumor specificity, biosafety, and sensitivity, redox-responsive NPs will be promising for more desirable therapeutic modalities, enabling their future clinical applications.

Conflicts of interest

There are no conflicts to declare.

References

- 1 R. L. Siegel, K. D. Miller and A. Jemal, *Ca-Cancer J. Clin.*, 2020, **70**, 7–30.
- 2 P. Agostinis, K. Berg, K. A. Cengel, T. H. Foster, A. W. Girotti, S. O. Gollnick, S. M. Hahn, M. R. Hamblin, A. Juzeniene, D. Kessel, M. Korbelik, J. Moan, P. Mroz, D. Nowis, J. Piette, B. C. Wilson and J. Golab, *Ca-Cancer J. Clin.*, 2011, **61**, 250–281.
- 3 H. Shi and P. J. Sadler, *Br. J. Cancer*, 2020, **123**, 871–873.
- 4 Q. Zheng, X. Liu, Y. Zheng, K. W. K. Yeung, Z. Cui, Y. Liang, Z. Li, S. Zhu, X. Wang and S. Wu, *Chem. Soc. Rev.*, 2021, **50**, 5086–5125.
- 5 G. Lan, K. Ni and W. Lin, *Coord. Chem. Rev.*, 2019, **379**, 65–81.
- 6 V. T. DeVita Jr and E. Chu, *Cancer Res.*, 2008, **68**, 8643–8653.
- 7 S. B. Park, D. Goldstein, A. V. Krishnan, C. S. Lin, M. L. Friedlander, J. Cassidy, M. Koltzenburg and M. C. Kiernan, *Ca-Cancer J. Clin.*, 2013, **63**, 419–437.
- 8 G. S. Karagiannis, J. S. Condeelis and M. H. Oktay, *Clin. Exp. Metastasis*, 2018, **35**, 269–284.
- 9 M. Yu, D. Su, Y. Yang, L. Qin, C. Hu, R. Liu, Y. Zhou, C. Yang, X. Yang, G. Wang and H. Gao, *ACS Appl. Mater. Interfaces*, 2019, **11**, 176–186.
- 10 K. Nurgali, R. T. Jagoe and R. Abalo, *Front. Pharmacol.*, 2018, **9**, 245.
- 11 X. Li, J. F. Lovell, J. Yoon and X. Chen, *Nat. Rev. Clin. Oncol.*, 2020, **17**, 657–674.
- 12 D. Xi, M. Xiao, J. Cao, L. Zhao, N. Xu, S. Long, J. Fan, K. Shao, W. Sun, X. Yan and X. Peng, *Adv. Mater.*, 2020, **32**, 1907855.
- 13 D. Xi, N. Xu, X. Xia, C. Shi, X. Li, D. Wang, S. Long, J. Fan, W. Sun and X. Peng, *Adv. Mater.*, 2022, **34**, 2106797.
- 14 Y. Qin, F. Tong, W. Zhang, Y. Zhou, S. He, R. Xie, T. Lei, Y. Wang, S. Peng, Z. Li, J. Leong, H. Gao and L. Lu, *Adv. Funct. Mater.*, 2021, **31**, 2104645.
- 15 X.-Q. Zhou, M. Xiao, V. Ramu, J. Hilgendorf, X. Li, P. Papadopolou, M. A. Siegler, A. Kros, W. Sun and S. Bonnet, *J. Am. Chem. Soc.*, 2020, **142**, 10383–10399.
- 16 S.-Y. Qin, Y.-J. Cheng, Q. Lei, A.-Q. Zhang and X.-Z. Zhang, *Biomaterials*, 2018, **171**, 178–197.
- 17 W. Sang, Z. Zhang, Y. Dai and X. Chen, *Chem. Soc. Rev.*, 2019, **48**, 3771–3810.
- 18 Y. Wang, J. Liu, X. Ma and X.-J. Liang, *Nano Res.*, 2018, **11**, 2932–2950.
- 19 H. Lei, X. Wang, S. Bai, F. Gong, N. Yang, Y. Gong, L. Hou, M. Cao, Z. Liu and L. Cheng, *ACS Appl. Mater. Interfaces*, 2020, **12**, 52370–52382.
- 20 J. Liu, M. Wu, Y. Pan, Y. Duan, Z. Dong, Y. Chao, Z. Liu and B. Liu, *Adv. Funct. Mater.*, 2020, **30**, 1908865.
- 21 J. Park, Y. Choi, H. Chang, W. Um, J. H. Ryu and I. C. Kwon, *Theranostics*, 2019, **9**, 8073–8090.
- 22 S. S. Lucky, K. C. Soo and Y. Zhang, *Chem. Rev.*, 2015, **115**, 1990–2042.
- 23 A. S. Thakor and S. S. Gambhir, *Ca-Cancer J. Clin.*, 2013, **63**, 395–418.
- 24 S. Guo, K. Li, B. Hu, C. Li, M. Zhang, A. Hussain, X. Wang, Q. Cheng, F. Yang, K. Ge, J. Zhang, J. Chang, X. J. Liang, Y. Weng and Y. Huang, *Exploration*, 2021, **1**, 35–49.
- 25 K. Y. W. Teo, C. Sevensan, Y. A. Cheow, S. Zhang, D. T. Leong and W. S. Toh, *Small Science*, 2022, **2**, 2100116.
- 26 B. Zhang, D. Zheng, S. Yiming, K. Oyama, M. Ito, M. Ikari, T. Kigawa, T. Mikawa and T. Miyake, *Small Science*, 2021, **1**, 2100069.
- 27 Y. Wang, Y. Zhang, J. Wang and X. J. Liang, *Adv. Drug Delivery Rev.*, 2019, **143**, 161–176.
- 28 Y. Yamada, S. Atrialdi, M. Hibino, D. Sasaki, J. Abe and H. Harashima, *Adv. Drug Delivery Rev.*, 2020, **154–155**, 187–209.
- 29 L. Duan, L. Yang, J. Jin, F. Yang, D. Liu, K. Hu, Q. Wang, Y. Yue and N. Gu, *Theranostics*, 2020, **10**, 462–483.
- 30 W. Cai, J. Wang, C. Chu, W. Chen, C. Wu and G. Liu, *Adv. Sci.*, 2019, **6**, 1801526.
- 31 Z. R. Lu, V. E. A. Laney, R. Hall and N. Ayat, *Adv. Healthcare Mater.*, 2021, **10**, 2001294.
- 32 C. Y. Ang, S. Y. Tan, C. Teh, J. M. Lee, M. F. Wong, Q. Qu, L. Q. Poh, M. Li, Y. Zhang, V. Korzh and Y. Zhao, *Small*, 2017, **13**, 1602379.
- 33 J. Li, X. Huang, X. Zhao, L. J. Chen and X. P. Yan, *Angew. Chem., Int. Ed.*, 2021, **60**, 2398–2405.
- 34 M. He, F. Chen, D. Shao, P. Weis, Z. Wei and W. Sun, *Biomaterials*, 2021, **275**, 120915.
- 35 F. Chen, F. Zhang, D. Shao, W. Zhang, L. Zheng, W. Wang, W. Yang, Z. Wang, J. Chen, W.-f. Dong, F. Xiao and Y. Wu, *Appl. Mater. Today*, 2020, **19**, 100558.
- 36 N. M. Anderson and M. C. Simon, *Curr. Biol.*, 2020, **30**, R921–R925.
- 37 S. Yang and H. Gao, *Pharmacol. Res.*, 2017, **126**, 97–108.
- 38 Y. Zhou, X. Chen, J. Cao and H. Gao, *J. Mater. Chem. B*, 2020, **8**, 6765–6781.
- 39 C. Gong, X. Zhang, M. Shi, F. Li, S. Wang, Y. Wang, Y. Wang, W. Wei and G. Ma, *Adv. Sci.*, 2021, **8**, 2002787.
- 40 E. Perez-Herrero and A. Fernandez-Medarde, *Acta Pharm. Sin. B*, 2021, **11**, 2243–2264.
- 41 S. Chen, D. Li, X. Du, X. He, M. Huang, Y. Wang, X. Yang and J. Wang, *Nano Today*, 2020, **35**, 100924.
- 42 J. Zhao, Y. Y. Peng, D. Diaz-Dussan, J. White, W. Duan, L. Kong, R. Narain and X. Hao, *Mol. Pharm.*, 2022, **19**, 1766–1777.
- 43 J. Mu, H. Zhong, H. Zou, T. Liu, N. Yu, X. Zhang, Z. Xu, Z. Chen and S. Guo, *J. Controlled Release*, 2020, **326**, 265–275.
- 44 W. R. Wilson and M. P. Hay, *Nat. Rev. Cancer*, 2011, **11**, 393–410.
- 45 A. Patel and S. Sant, *Biotechnol. Adv.*, 2016, **34**, 803–812.
- 46 D. Cui, J. Huang, X. Zhen, J. Li, Y. Jiang and K. Pu, *Angew. Chem., Int. Ed.*, 2019, **58**, 5920–5924.
- 47 S. Zhou, X. Hu, R. Xia, S. Liu, Q. Pei, G. Chen, Z. Xie and X. Jing, *Angew. Chem., Int. Ed.*, 2020, **59**, 23198–23205.



- 48 Q. Mou, Y. Ma, F. Ding, X. Gao, D. Yan, X. Zhu and C. Zhang, *J. Am. Chem. Soc.*, 2019, **141**, 6955–6966.
- 49 L. Luo, Z. Yin, Y. Qi, S. Liu, Y. Yi, X. Tian, Y. Wu, D. Zhong, Z. Gu, H. Zhang and K. Luo, *Appl. Mater. Today*, 2021, **23**, 100996.
- 50 Y. Liu, Y. Lu, X. Zhu, C. Li, M. Yan, J. Pan and G. Ma, *Biomaterials*, 2020, **242**, 119933.
- 51 J. Choi, M. K. Shim, S. Yang, H. S. Hwang, H. Cho, J. Kim, W. S. Yun, Y. Moon, J. Kim, H. Y. Yoon and K. Kim, *ACS Nano*, 2021, **15**, 12086–12098.
- 52 D. Li, R. Zhang, G. Liu, Y. Kang and J. Wu, *Adv. Healthcare Mater.*, 2020, **9**, 2000605.
- 53 P. H. Hsu and A. Almutairi, *J. Mater. Chem. B*, 2021, **9**, 2179–2188.
- 54 R. Li, F. Peng, J. Cai, D. Yang and P. Zhang, *Asian J. Pharm. Sci.*, 2020, **15**, 311–325.
- 55 T. Lei, Z. Yang, X. Xia, Y. Chen, X. Yang, R. Xie, F. Tong, X. Wang and H. Gao, *Acta Pharm. Sin. B*, 2021, **11**, 4032–4044.
- 56 S. H. Lee, M. K. Gupta, J. B. Bang, H. Bae and H. J. Sung, *Adv. Healthcare Mater.*, 2013, **2**, 908–915.
- 57 C. R. Reczek and N. S. Chandel, *Annu. Rev. Cancer Biol.*, 2017, **1**, 79–98.
- 58 J. Wang, L. Liu, L. Yin and L. Chen, *J. Biomed. Mater. Res., Part A*, 2018, **106**, 2955–2962.
- 59 G. Liu, Z. Bao and J. Wu, *Chin. Chem. Lett.*, 2020, **31**, 1817–1821.
- 60 J. M. Estrela, A. Ortega and E. Obrador, *Crit. Rev. Clin. Lab. Sci.*, 2006, **43**, 143–181.
- 61 J. A. Cook, D. Gius, D. A. Wink, M. C. Krishna, A. Russo and J. B. Mitchell, *Semin. Radiat. Oncol.*, 2004, **14**, 259–266.
- 62 X. Zhong, X. Wang, L. Cheng, Y. Tang, G. Zhan, F. Gong, R. Zhang, J. Hu, Z. Liu and X. Yang, *Adv. Funct. Mater.*, 2020, **30**, 1907954.
- 63 C. R. Corso and A. Acco, *Crit. Rev. Oncol. Hematol.*, 2018, **128**, 43–57.
- 64 G. Saravanakumar, J. Kim and W. J. Kim, *Adv. Sci.*, 2017, **4**, 1600124.
- 65 M. S. Shim and Y. Xia, *Angew. Chem., Int. Ed.*, 2013, **52**, 6926–6929.
- 66 M. Bio, G. Nkepeng and Y. You, *Chem. Commun.*, 2012, **48**, 6517–6519.
- 67 C. Lin, F. Tong, R. Liu, R. Xie, T. Lei, Y. Chen, Z. Yang, H. Gao and X. Yu, *Acta Pharm. Sin. B*, 2020, **10**, 2348–2361.
- 68 M. H. Lee, Z. Yang, C. W. Lim, Y. H. Lee, S. Dongbang, C. Kang and J. S. Kim, *Chem. Rev.*, 2013, **113**, 5071–5109.
- 69 J. Li, L. Jiao, W. Xu, H. Yan, G. Chen, Y. Wu, L. Hu and W. Gu, *Sens. Actuators, B*, 2021, **329**, 129247.
- 70 H. Pelicano, D. Carney and P. Huang, *Drug Resist. Updates*, 2004, **7**, 97–110.
- 71 F. Liu, T. Bing, D. Shangguan, M. Zhao and N. Shao, *Anal. Chem.*, 2016, **88**, 10631–10638.
- 72 C. Shi, M. Li, Z. Zhang, Q. Yao, K. Shao, F. Xu, N. Xu, H. Li, J. Fan, W. Sun, J. Du, S. Long, J. Wang and X. Peng, *Biomaterials*, 2020, **233**, 119755.
- 73 Z. Deng, Y. Qian, Y. Yu, G. Liu, J. Hu, G. Zhang and S. Liu, *J. Am. Chem. Soc.*, 2016, **138**, 10452–10466.
- 74 Y. H. Kim, X. T. Cao, S.-S. Hong, Y. S. Gal and K. T. Lim, *Mol. Cryst. Liq. Cryst.*, 2016, **635**, 107–113.
- 75 M. Wen, J. Ouyang, C. Wei, H. Li, W. Chen and Y. N. Liu, *Angew. Chem., Int. Ed.*, 2019, **58**, 17425–17432.
- 76 X. Guo, Y. Cheng, X. Zhao, Y. Luo, J. Chen and W. E. Yuan, *J. Nanobiotechnol.*, 2018, **16**, 74.
- 77 F. Gong, M. Chen, N. Yang, Z. Dong, L. Tian, Y. Hao, M. Zhuo, Z. Liu, Q. Chen and L. Cheng, *Adv. Funct. Mater.*, 2020, **30**, 2002753.
- 78 L. S. Lin, J. Song, L. Song, K. Ke, Y. Liu, Z. Zhou, Z. Shen, J. Li, Z. Yang, W. Tang, G. Niu, H. H. Yang and X. Chen, *Angew. Chem., Int. Ed.*, 2018, **57**, 4902–4906.
- 79 L. H. Fu, Y. Wan, C. Qi, J. He, C. Li, C. Yang, H. Xu, J. Lin and P. Huang, *Adv. Mater.*, 2021, **33**, 2006892.
- 80 L. Lu, X. Zhao, T. Fu, K. Li, Y. He, Z. Luo, L. Dai, R. Zeng and K. Cai, *Biomaterials*, 2020, **230**, 119666.
- 81 S. Li, P. E. Saw, C. Lin, Y. Nie, W. Tao, O. C. Farokhzad, L. Zhang and X. Xu, *Biomaterials*, 2020, **234**, 119760.
- 82 X. Yi, J. J. Hu, J. Dai, X. Lou, Z. Zhao, F. Xia and B. Z. Tang, *ACS Nano*, 2021, **15**, 3026–3037.
- 83 K. Luo, W. Guo, Y. Yu, S. Xu, M. Zhou, K. Xiang, K. Niu, X. Zhu, G. Zhu, Z. An, Q. Yu and Z. Gan, *J. Controlled Release*, 2020, **326**, 25–37.
- 84 L. Luo, Y. Qi, H. Zhong, S. Jiang, H. Zhang, H. Cai, Y. Wu, Z. Gu, Q. Gong and K. Luo, *Acta Pharm. Sin. B*, 2022, **12**, 424–436.
- 85 X. Xie, C. Zhan, J. Wang, F. Zeng and S. Wu, *Small*, 2020, **16**, 2003451.
- 86 Y. Yang, B. Sun, S. Zuo, X. Li, S. Zhou, L. Li, C. Luo, H. Liu, M. Cheng, Y. Wang, S. Wang, Z. He and J. Sun, *Sci. Adv.*, 2020, **6**, eabc1725.
- 87 W. Feng, Y. Lv, Z. Chen, F. Wang, Y. Wang, Y. Pei, W. Jin, C. Shi, Y. Wang, Y. Qu, W. Ji, L. Pu, X.-W. Liu and Z. Pei, *Chem. Eng. J.*, 2021, **417**, 129178.
- 88 Z. Pang, J. Zhou and C. Sun, *Front. Chem.*, 2020, **8**, 156.
- 89 H. Deng, Z. Yang, X. Pang, C. Zhao, J. Tian, Z. Wang and X. Chen, *Nano Today*, 2022, **42**, 101337.
- 90 X. Lin, R. Zhu, Z. Hong, X. Zhang, S. Chen, J. Song and H. Yang, *Adv. Funct. Mater.*, 2021, **31**, 2101278.
- 91 Y. Yan, Y. Hou, H. Zhang, W. Gao, R. Han, J. Yu, L. Xu and K. Tang, *Colloids Surf., B*, 2021, **208**, 112103.
- 92 H. Cheng, X. Wang, X. Liu, X. Wang, H. Wen, Y. Cheng, A. Xie, Y. Shen, R. Tang and M. Zhu, *Nanoscale*, 2021, **13**, 10816–10828.
- 93 S. Sun, Q. Chen, Z. Tang, C. Liu, Z. Li, A. Wu and H. Lin, *Angew. Chem., Int. Ed.*, 2020, **59**, 21041–21048.
- 94 B. Ma, Y. Nishina and A. Bianco, *Carbon*, 2021, **178**, 783–791.
- 95 V. J. Thannickal and B. L. Fanburg, *Am. J. Physiol.: Lung Cell. Mol. Physiol.*, 2000, **279**, L1005–L1028.
- 96 S. Parvez, M. J. C. Long, J. R. Poganik and Y. Aye, *Chem. Rev.*, 2018, **118**, 8798–8888.
- 97 X. Xu, P. E. Saw, W. Tao, Y. Li, X. Ji, S. Bhasin, Y. Liu, D. Ayyash, J. Rasmussen, M. Huo, J. Shi and O. C. Farokhzad, *Adv. Mater.*, 2017, **29**, 1700141.



- 98 D. Zhao, W. Tao, S. Li, L. Li, Y. Sun, G. Li, G. Wang, Y. Wang, B. Lin, C. Luo, Y. Wang, M. Cheng, Z. He and J. Sun, *Nanoscale Horiz.*, 2020, **5**, 886–894.
- 99 B. Chu, Y. Qu, X. He, Y. Hao, C. Yang, Y. Yang, D. Hu, F. Wang and Z. Qian, *Adv. Funct. Mater.*, 2020, **30**, 2005918.
- 100 C.-C. Song, R. Ji, F.-S. Du and Z.-C. Li, *Macromolecules*, 2013, **46**, 8416–8425.
- 101 M. Zhang, C. C. Song, S. Su, F. S. Du and Z. C. Li, *ACS Appl. Mater. Interfaces*, 2018, **10**, 7798–7810.
- 102 H. Luo, C. Huang, J. Chen, H. Yu, Z. Cai, H. Xu, C. Li, L. Deng, G. Chen and W. Cui, *Biomaterials*, 2021, **279**, 121194.
- 103 C. Lux, S. Joshi-Barr, T. Nguyen, E. Mahmoud, E. Schopf, N. Fomina and A. Almutairi, *J. Am. Chem. Soc.*, 2012, **134**, 15758–15764.
- 104 A. Gennari, C. Gujral, E. Hohn, E. Lallana, F. Cellesi and N. Tirelli, *Bioconjugate Chem.*, 2017, **28**, 1391–1402.
- 105 H. Xiang, Y. Wu, X. Zhu, M. She, Q. An, R. Zhou, P. Xu, F. Zhao, L. Yan and Y. Zhao, *J. Am. Chem. Soc.*, 2021, **143**, 11449–11461.
- 106 M. Bio, P. Rajaputra, G. Nkepan, S. G. Awuah, A. M. Hossion and Y. You, *J. Med. Chem.*, 2013, **56**, 3936–3942.
- 107 P. Thapa, M. Li, M. Bio, P. Rajaputra, G. Nkepan, Y. Sun, S. Woo and Y. You, *J. Med. Chem.*, 2016, **59**, 3204–3214.
- 108 M. He, G. He, P. Wang, S. Jiang, Z. Jiao, D. Xi, P. Miao, X. Leng, Z. Wei, Y. Li, Y. Yang, R. Wang, J. Du, J. Fan, W. Sun and X. Peng, *Adv. Sci.*, 2021, **8**, 2103334.
- 109 X. Yang, C. Hu, F. Tong, R. Liu, Y. Zhou, L. Qin, L. Ouyang and H. Gao, *Adv. Funct. Mater.*, 2019, **29**, 1901896.
- 110 R. Liu, M. Yu, X. Yang, C. S. Umeshappa, C. Hu, W. Yu, L. Qin, Y. Huang and H. Gao, *Adv. Funct. Mater.*, 2019, **29**, 1808462.
- 111 R. Liu, Y. An, W. Jia, Y. Wang, Y. Wu, Y. Zhen, J. Cao and H. Gao, *J. Controlled Release*, 2020, **321**, 589–601.
- 112 L. Han, X. Y. Zhang, Y. L. Wang, X. Li, X. H. Yang, M. Huang, K. Hu, L. H. Li and Y. Wei, *J. Controlled Release*, 2017, **259**, 40–52.
- 113 M. Ye, Y. Han, J. Tang, Y. Piao, X. Liu, Z. Zhou, J. Gao, J. Rao and Y. Shen, *Adv. Mater.*, 2017, **29**, 1702342.
- 114 Y. Kang, X. Ju, L. S. Ding, S. Zhang and B. J. Li, *ACS Appl. Mater. Interfaces*, 2017, **9**, 4475–4484.
- 115 Y. Tian, M. Lei, L. Yan and F. An, *Polym. Chem.*, 2020, **11**, 2360–2369.
- 116 B. Z. Hailemeskel, K. D. Addisu, A. Prasannan, S. L. Mekuria, C.-Y. Kao and H.-C. Tsai, *Appl. Surf. Sci.*, 2018, **449**, 15–22.
- 117 N. Ma, Y. Li, H. Xu, Z. Wang and X. Zhang, *J. Am. Chem. Soc.*, 2010, **132**, 442–443.
- 118 L. Wang, W. Cao, Y. Yi and H. Xu, *Langmuir*, 2014, **30**, 5628–5636.
- 119 D. Shao, M. Li, Z. Wang, X. Zheng, Y. H. Lao, Z. Chang, F. Zhang, M. Lu, J. Yue, H. Hu, H. Yan, L. Chen, W. F. Dong and K. W. Leong, *Adv. Mater.*, 2018, **30**, 1801198.
- 120 D. Shao, F. Zhang, F. Chen, X. Zheng, H. Hu, C. Yang, Z. Tu, Z. Wang, Z. Chang, J. Lu, T. Li, Y. Zhang, L. Chen, K. W. Leong and W. F. Dong, *Adv. Mater.*, 2020, **32**, 2004385.
- 121 Y. Yang, F. Chen, N. Xu, Q. Yao, R. Wang, X. Xie, F. Zhang, Y. He, D. Shao, W. F. Dong, J. Fan, W. Sun and X. Peng, *Biomaterials*, 2022, **281**, 121368.
- 122 S. Pandit, D. Dutta and S. Nie, *Nat. Mater.*, 2020, **19**, 478–480.
- 123 S. Sindhwani, A. M. Syed, J. Ngai, B. R. Kingston, L. Maiorino, J. Rothschild, P. MacMillan, Y. Zhang, N. U. Rajesh, T. Hoang, J. L. Y. Wu, S. Wilhelm, A. Zilman, S. Gadde, A. Sulaiman, B. Ouyang, Z. Lin, L. Wang, M. Egeblad and W. C. W. Chan, *Nat. Mater.*, 2020, **19**, 566–575.
- 124 P. Wei and J. S. Moodera, *Nat. Mater.*, 2020, **19**, 481–482.
- 125 J. He, C. Li, L. Ding, Y. Huang, X. Yin, J. Zhang, J. Zhang, C. Yao, M. Liang, R. P. Pirraco, J. Chen, Q. Lu, R. Baldrige, Y. Zhang, M. Wu, R. L. Reis and Y. Wang, *Adv. Mater.*, 2019, **31**, 1902409.
- 126 Y. Ding, Y. Wang and Q. Hu, *Exploration*, 2022, **2**, 20210106.

