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Synthesis of D-glyco-alkynone derivatives via carbonylative Sonogashira reaction†

Mariana P. Darbem, a C. Henrique A. Esteves, a Isadora M. de Oliveira, b Joel S. Reis, a Daniel C. Pimenta^c and Hélio A. Stefani b *a

A carbonylative Sonogashira coupling approach to the synthesis of glyco-alkynones is described. Eighteen examples were obtained in moderate do nearly quantitative yields under mild conditions employing Mo(CO)₆ as a safe carbon monoxide source. Functionalization of the alkynyl moiety via cycloaddition with organic azides provided six examples of glyco-triazoles.

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Introduction

Alkynones are attractive motifs in organic chemistry involved in the synthesis of medicinally valuable heteroaromatic compounds. These molecules are also important intermediates in the synthesis of natural products² and as part of biologically active molecules (Fig. 1).3

Consequently, a rich variety of methodologies targeting their synthesis has been reported, some of which involve the addition of borylated terminal alkynes to acyl chlorides,4 the addition of hypervalent alkynyl iodides to aldehydes via C-C bond cleavage, metal-catalyzed C-H bond activation of aldehydes5 or the oxidation of propargylic alcohols.6 While impressive, these methodologies present some drawbacks, such as excessive generation of chemical waste, instability of some of the substrates required and poor functional group tolerance. The

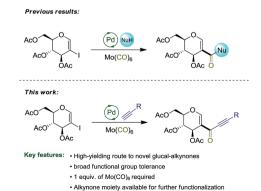
Anti-inflammatory Ginsenoyne E (cytotoxic activity) and Analgesic -hydroxy-2-nonyn-3-one (antifungal activity)

Fig. 1 Alkynones in biologically active compounds.

Pd-catalyzed carbonylative Sonogashira coupling, on the other hand, offers a route to alkynones that is mild, atom-economical and functional-group-tolerant.7 Attracted by these features, we decided to explore the construction of glyco-alkynones relying on this reaction as part of our ongoing research interest in the synthesis of functionalized glycals.8

In a previous report,8a we explored the synthesis of amidoglucals and glucal esters via the carbonylative coupling reaction of 2-iodo-p-glucal. Herein, we describe the synthesis of glycoalkynones via carbonylative Sonogashira coupling reaction, expanding the spectrum of reactions involving this important substrate (Scheme 1).

Taking advantage of the alkynyl group readily installed by this reaction, we also explored the synthesis of glyco-substituted triazoles via click chemistry. This approach has been of pivotal importance for carbohydrate chemistry as a tool to efficiently connect a sugar moiety to a molecule of interest via a triazole linker, improving the hydrophilicity, bioavailability and chemical profile of these fragments.9 Moreover, the biological activity demonstrated by several alkynone derivatives (e.g. triazoles) make new routes to these structures synthetically relevant (Fig. 2).10



Scheme 1 2-lodoglucal carbonylative coupling reactions.

[&]quot;Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, São Paulo, SP, Brazil. E-mail: hstefani@usp.br; Tel: +55 11 3091-3654

^bInstituto de Química, Universidade de São Paulo, São Paulo, SP, Brazil

^{&#}x27;Instituto Butantã, São Paulo, SP, Brazil

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Fig. 2 Examples of biologically active triazolic alkynones.

Scheme 2 Synthesis of 2-iodo-tri-O-acetyl-D-glucal

Results and discussion

We commenced our study by synthesizing 2-iodo-tri-*O*-acetyl-D-glucal (1) from tri-*O*-acetyl-D-glucal, *N*-iodosuccinimide (NIS) and AgNO₃ (Scheme 2).¹¹

With substrate 1 in hand, we next screened the reaction conditions for the carbonylation of 2-iodo-tri-O-acetyl-D-

glucal (1) with Mo(CO)₆ and 4-ethynyltoluene. Reactions were followed by TLC to ensure full conversion of the starting material 1 (Table 1).

We started by screening the effect of the catalyst on the reaction outcome. PdCl₂, Pd(PhCN)₂Cl₂, and Pd(Prol)₂ (Table 1, entries 1, 2 and 4, respectively) led to the formation of alkynone 3a in moderate yields. Catalysts containing ligands that are at the same time electron-rich and sterically demanding, such as xantphos and PEPPSI, ¹² delivered alkynone 3a in good to nearly quantitative yield (Table 1, entries 3, 5 and 6), with the combination PdCl₂/xantphos being the best. In order to seek other high-yielding set of conditions, the effect of the base was next examined. Organic and inorganic bases such as DIPEA, DBU, NaOAc and K₂CO₃ gave 3a in lower yields, with inorganic K₂CO₃ delivering the desired product in only 25%. Different solvents were also screened, however, only poor to modest yields of 3a were obtained.

With the optimized reaction conditions in hand, we set out to investigate the generality of this reaction (Scheme 3).

Terminal alkynes bearing electron-neutral and electron-donating groups delivered the desired alkynones in good to excellent yields (3a-g). Electron-withdrawing groups such as the difluorinated moiety present in 2h and meta-chloro substituted 2i gave 3h and 3i in good yields, while meta-fluorinated 2j gave 3j in 67%. Incorporation of an heteroaromatic substituent was also tolerated, and alkynone 3k was obtained in 65% yield. Pleasingly, both cyclopropyl and TMS groups proved to be stable under the reaction

Table 1 Screening of reaction conditions

	Catalyst/ligand	Base			
Entry ^a		(3.0 equvi.)	Solvent	Reaction time (h)	Yield (%)
Effect of cata	lyst				
1	$PdCl_2$	$\mathrm{Et_{3}N}$	1,4-Dioxane	12	66
2	$Pd(PhCN)_2Cl_2$	Et_3N	1,4-Dioxane	12	58
3	Pd(PhCN) ₂ Cl ₂ /xantphos	$\mathrm{Et_{3}N}$	1,4-Dioxane	12	73
4	$Pd(Prol)_2$	Et_3N	1,4-Dioxane	12	63
5	PEPPSI-IPr	$\mathrm{Et_{3}N}$	1,4-Dioxane	12	75
6	PdCl ₂ /xantphos	$\mathrm{Et_{3}N}$	1,4-Dioxane	2	99
Effects of bas	se				
7	PdCl ₂ /xantphos	DIPEA	1,4-Dioxane	16	55
8	PdCl ₂ /xantphos	DBU	1,4-Dioxane	16	43
9	PdCl ₂ /xantphos	NaOAc	1,4-Dioxane	16	32
10	PdCl ₂ /xantphos	K_2CO_3	1,4-Dioxane	16	25
Effect of solv	ent				
11	PdCl ₂ /xantphos	Et_3N	Toluene	16	55
12	PdCl ₂ /xantphos	Et ₃ N	THF	16	43
13	PdCl ₂ /xantphos	Et_3N	DMF	16	32
14	PdCl ₂ /xantphos	$\mathrm{Et}_{3}\mathbf{N}$	MeCN	16	25

^a Reaction condition: 1 (0.2 mmol), catalyst (5 mol%), ligand (5 mol%), 4-ethynyltoluene (1.5 equvi.), base (3.0 equvi.), solvent (3 mL).

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Reaction conditions: 1 (0.2 mmol), terminal alkyne (1.5 equiv.), PdCl₂ (5.0 mol%), Xantphos (5.0 mol%), Et₃N (3.0 equiv.), Mo(CO)_e (1.0 equiv.), 1.4-dioxane, 2 h, 70 °C, ^a Reaction time; 4 h, ^b gram-scale reaction

Scheme 3 Sonogashira carbonylative coupling reaction of 2-iodo-p-glucal and terminal alkynes.

conditions, with products 31 and 3m being isolated in 72% and 80%, respectively, both leaving useful handles for further functionalization (see Scheme 4).13 Incorporation of terminal alkynes bearing alkyl moieties provided mixed results, with 2n and 20 delivering alkynones in moderate yields, while 2p and 2q, bearing a tertiary alcohol, provided 3p and 3q in good yields. 1,4-Diethynylbenzene 2r was subjected to the reaction conditions, giving the symmetrical alkynone 3r in 70%. Finally, the reaction with 2a was repeated on a gram scale, providing 3a in 80% isolated yield (Scheme 4).

In order to demonstrate the usefulness of this methodology, we decided to explore the formation of 1,2,3-triazoles via click chemistry. An in situ-generated terminal alkyne provided the desired triazoles 5a-f in the presence of organic azides, PMDTA and copper iodide (conditions found after a quick

Scheme 4 Gram-scale reaction.

screening).14 A variety of moieties were tolerated at the position 1 of the newly formed ring: a benzylic substituent (5a, 67%), heteroaromatic substituents (5b, 72% and 5c, 70%) and

Reaction conditions: 3m (0.25 mmol), Cul (1.0 equiv.), RN₃ (1.2 equiv.), PMDTA (1.2 equiv.), TBAF (1.2 equiv.), THF (3 mL), 0 °C for 2 h. PMDTA = N,N,N',N'',N''-pentamethyldiethylenetriamine.

Scheme 5 Synthesis of D-glyco-1,2,3-triazoles.

unactivated (5d, 65% and 5e, 58%) and activated aromatic rings (5f, 86%) (Scheme 5).

Conclusions

In conclusion, we have described a convenient palladium-catalyzed Sonogashira carbonylative coupling reaction for the synthesis of p-glyco-alkynones. This approach permitted the synthesis of 18 examples in moderate to nearly quantitative yields under mild conditions, employing Mo(CO)₆ as a safe carbon monoxide source. Further functionalization of a masked terminal alkynone allowed the synthesis of p-glyco-1,2,3-triazoles in moderate yields, demonstrating one of the potential applications of the alkynones described herein.

Experimental section

General considerations

The compounds were all identified by usual analytical methods: ¹H NMR, ¹³C NMR, IR, and HR-MS (ESI). ¹H and ¹³C NMR spectra were measured in CDCl₃, in a Bruker DPX-300 instrument. ¹H chemical shifts were reported in ppm referenced relative to TMS internal standard (0.00 ppm) or the residual chloroform peak (7.26 ppm). Abbreviations to denote the multiplicity of a particular signal are: m (multiplet), s (singlet), d (doublet), t (triplet) and dd (doublet of doublets). ¹³C chemical shift were reported in ppm relative to the CDCl₃ triplet (77.16 ppm). IR spectra were measured on an Agilent Technologies Cary 630 and were reported in wavenumbers (cm⁻¹). High-resolution mass spectra (HRMS) were recorded on a Shimadzu LCMS-TOF, using ESI with 50% solution of acetonitrile/H2O and 0.1% formic acid as ionization method. Thin layer chromatography (TLC) was performed using silica gel UV₂₅₄ 0.20 mm thickness. For visualization, TLC plates were either placed under ultraviolet light, or stained with iodine or acidic vanillin solution. The solvents were purified by distillation or used without any purification in the case of HPLC-grade material. All other compounds were used as received.

General procedure for the synthesis of 3a-r

To a vial equipped with a magnetic stirrer bar and sealed with a rubber septum connected to a deflated balloon with a needle were added the tri-O-acetylated iodoglucal (0.2 mmol), 1,4-dioxane (3.0 mL), PdCl $_2$ (5 mol%), xantphos (5 mol%), Mo(CO) $_6$ (0.2 mmol, 1 equiv.), the alkyne (0.3 mmol, 1.5 equiv.) and Et $_3$ N (0.6 mmol, 3 equiv.). The reaction mixture was vigorously stirred at 70 °C for 2 to 4 h. The resulting mixture was washed with water and extracted with ethyl acetate. The organic layers were then combined and evaporated. The crude products were purified by flash chromatography using hexane and ethyl acetate as eluent (7 : 3).

General procedure for the synthesis of 5a-f

To a vial (20 mL) equipped with a magnetic stirrer bar under a nitrogen atmosphere containing CuI (0.25 mmol, 1 equiv.), THF (4 mL), an organic azide (0.3 mmol, 1.2 equiv.) and **3m** (0.25 mmol, 1 equiv.) was added PMDETA (0.3 mmol, 1.2 equiv.)

and the reaction mixture was stirred at 0 $^{\circ}$ C for 2 h. After this period, the reaction mixture was diluted with ethyl acetate and washed with aqueous NaCl. The organic phase was collected, dried over MgSO₄, filtered and the solvent was evaporated under reduced pressure. Purification was performed using flash chromatography (ethyl acetate/hexane, 4 : 6).

Analytical data of compounds 3a-r/5a-f

Product **3a** was obtained as a yellow oil (83 mg, 0.20 mmol, 99%). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.00$ (s, 1H), 7.40 (d, J = 7.9 Hz, 2H), 7.12 (d, J = 7.8 Hz, 2H), 5.73 (dd, J = 3.1, 1.6 Hz, 1H), 5.15 (t, J = 3.0 Hz, 1H), 4.70–4.51 (m, 1H), 4.40 (dd, J = 12.1, 7.8 Hz, 1H), 4.14 (dd, J = 12.1, 4.4 Hz, 1H), 2.31 (s, 3H), 2.15–1.89 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.5$, 170.2, 169.4, 169.1, 160.7, 141.3, 132.8, 129.4, 116.7, 114.9, 91.4, 84.8, 75.6, 65.6, 61.2, 60.9, 21.6, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2877, 2112, 1685, 1564, 1177, 1328, 1197, 1154, 1143, 991. HRMS (ESI-TOF) calc. $[C_{22}H_{22}O_8Na^+]$ 437.1212, found 437.1212.

Product 3b was obtained as a yellow oil (72 mg, 0.18 mmol, 92%). 1 H NMR (300 MHz, CDCl₃): δ = 8.08 (s, 1H), 7.65–7.55 (m, 2H), 7.49–7.34 (m, 3H), 5.82 (d, J = 1.8 Hz, 1H), 5.23 (t, J = 3.0 Hz, 1H), 4.74–4.60 (m, 1H), 4.48 (dd, J = 12.1, 7.8 Hz, 1H), 4.21 (dd, J = 12.1, 4.5 Hz, 1H), 2.18–2.01 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 174.4, 170.2, 169.4, 169.1, 160.9, 132.8, 130.6, 128.6, 119.8, 114.9, 90.8, 84.9, 75.6, 65.6, 61.2, 60.9, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 2959, 2864, 2127, 1682, 1566, 1324, 1266, 1175, 1151, 992. HRMS (ESI-TOF) calc. [$C_{21}H_{20}O_8Na^+$] 423.1056, found 423.1051.

Product 3c was obtained as a yellow oil (84 mg, 0.18 mmol, 88%). 1 H NMR (300 MHz, CDCl₃): δ = 8.10 (s, 1H), 7.74–7.53 (m, 6H), 7.49–7.36 (m, 3H), 5.93–5.79 (m, 1H), 5.24 (t, J = 3.1 Hz, 1H), 4.75–4.64 (m, 1H), 4.49 (dd, J = 12.0, 7.8 Hz, 1H), 4.22 (dd, J = 12.1, 4.5 Hz, 1H), 2.19–1.98 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 174.4, 170.2, 169.4, 169.1, 160.8, 143.5, 139.7, 133.3, 128.9, 128.1, 127.3, 127.1, 118.5, 114.9, 90.9, 85.7, 75.7, 65.6, 61.2, 60.9, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 2959, 2931, 2123, 1685, 1566, 1438, 1324, 1264, 1175, 1151, 991. HRMS (ESI-TOF) calc. [C_{27} H₂₄O₈Na $^{+}$] 499.1363, found 499.1361.

Product 3d was obtained as a yellow oil (78 mg, 0.18 mmol, 90%). 1 H NMR (300 MHz, CDCl₃): $\delta = 8.06$ (s, 1H), 7.54 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 8.8 Hz, 2H), 5.87–5.78 (m, 1H), 5.23 (t, J = 3.0 Hz, 1H), 4.72–4.62 (m, 1H), 4.47 (dd, J = 12.1, 7.9 Hz, 1H), 4.21 (dd, J = 12.2, 4.5 Hz, 1H), 3.84 (s, 3H), 2.19–1.99 (m, 9H). 13 C NMR (75 MHz, CDCl₃): $\delta = 174.5$, 170.2, 169.4, 169.1, 161.6, 160.3, 134.8, 114.7, 114.4, 111.6, 91.9, 84.8, 75.5, 65.7, 61.3, 60.9, 55.4, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 2866, 2747, 2119, 1685, 1549, 1460, 1175, 1151, 1134, 991. HRMS (ESI-TOF) calc. $[C_{22}H_{22}O_9Na^{+1}]$ 453.1156, found 453.1159.

Product 3e was obtained as a yellow oil (88 mg, 0.19 mmol, 99%). ¹H NMR (300 MHz, CDCl₃): $\delta = 7.88$ (s, 1H), 7.29 (d, J = 8.5 Hz, 1H), 6.61–6.47 (m, 2H), 5.67–5.59 (m, 1H), 5.05 (t, J = 3.4 Hz, 1H), 4.54–4.41 (m, 1H), 4.33–4.23 (m, 1H), 4.12–3.95 (m, 1H), 3.62 (s, 3H), 2.29 (s, 3H), 1.96–1.80 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.5$, 170.2, 169.4, 169.1, 161.5, 160.4, 144.1, 135.2, 115.4, 115.0, 111.8, 111.6, 90.8, 88.4, 75.6, 65.6, 61.2, 60.9, 55.3, 21.0, 20.7, 20.6, 20.5. IR $(\nu, \text{cm}^{-1}) = 2821$, 2756, 2112, 1685,

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1566, 1549, 1324, 1259, 1179, 1151, 992. HRMS (ESI-TOF) calc. $\left[C_{23}H_{24}O_9Na^+\right]$ 467.1313, found 453.1311.

Product 3f was obtained as a yellow oil (91 mg, 0.19 mmol, 95%). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.05$ (s, 1H), 7.98 (s, 1H), 7.64 (m, 2H), 7.45 (d, J = 8.4 Hz, 1H), 7.11 (dd, J = 9.0, 2.4 Hz, 1H), 7.04 (d, J = 2.6 Hz, 1H), 5.85–5.70 (m, 1H), 5.28–5.06 (m, 1H), 4.64–4.56 (m, 1H), 4.52–4.35 (m, 1H), 4.24–4.11 (m, 1H), 3.85 (s, 3H), 2.19–1.96 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 174.5$, 170.2, 169.4, 169.2, 160.7, 159.3, 135.4, 133.9, 129.7, 128.9, 128.1, 127.2, 119.9, 114.9, 114.4, 105.9, 92.1, 85.0, 75.6, 65.7, 61.3, 61.0, 55.4, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2913, 2866, 2117, 1680, 1560, 1436, 1324, 1177, 1151, 991. HRMS (ESI-TOF) calc. [C₂₆H₂₄O₉Na⁺] 503.1313, found 503.1312.

Product 3g was obtained as a yellow oil (73 mg, 0.17 mmol, 85%). $^1{\rm H}$ NMR (300 MHz, CDCl₃): $\delta=9.21$ (s, 1H), 7.70 (s, 1H), 7.55–7.33 (m, 4H), 5.93 (s, 1H), 5.37 (s, 2H), 5.23 (t, J=3.4 Hz, 1H), 4.60–4.52 (m, 1H), 4.47 (dd, J=11.6, 7.7 Hz, 1H), 4.20 (dd, J=11.8, 4.1 Hz, 1H), 2.12–2.03 (m, 9H). $^{13}{\rm C}$ NMR (75 MHz, CDCl₃): $\delta=171.5$, 170.3, 169.6, 169.3, 154.2, 144.0, 131.9, 130.9, 128.3, 128.0, 120.4, 114.5, 96.0, 90.3, 74.3, 73.9, 66.2, 62.5, 61.0, 20.7, 20.7, 20.6. IR (ν , cm $^{-1}$) = 2861, 2080, 1680, 1574, 1527, 1324, 1177, 1145, 981, 732. HRMS (ESI-TOF) calc. [C $_{22}{\rm H}_{22}{\rm O}_{9}{\rm Na}^{+}$] 453.1156, found 453.1156.

Product **3h** was obtained as a yellow oil (67 mg, 0.15 mmol, 77%). 1 H NMR (300 MHz, CDCl₃): δ = 7.93 (s, 1H), 7.45–7.30 (m, 1H), 6.80–6.68 (m, 2H), 5.63–5.56 (m, 1H), 5.05 (t, J = 2.8 Hz, 1H), 4.54–4.45 (m, 1H), 4.34–4.21 (m, 1H), 4.13–3.94 (m, 1H), 2.01–1.79 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 173.9, 170.2, 169.3, 169.1, 164.4, 164.3 (dd, J = 253.5 Hz, J = 7.5 Hz), 161.6, 135.7, 135.6, 115.0, 112.3, 112.2 (dd J = 22.2 Hz, 3.3 Hz), 105.2 (dd, J = 3.7 Hz), 104.7 (t, J = 24.7 Hz) 89.4, 75.7, 65.5, 61.0, 60.9, 20.7, 20.6. IR (ν , cm $^{-1}$) = 2976, 2136, 1685, 1560, 1456, 1326, 1175, 1151, 992, 937, 711. HRMS (ESI-TOF) calc. $[C_{21}H_{18}F_2O_8Na^+]$ 436.0862, found 436.0869.

Product 3i was obtained as a yellow oil (74 mg, 0.17 mmol, 85%). ¹H NMR (300 MHz, CDCl₃): δ = 7.57 (s, 1H), 7.51–7.39 (m, 1H), 7.35 (d, J = 7.8 Hz, 2H), 5.85–5.75 (m, 1H), 5.23 (t, J = 3.0 Hz, 1H), 4.70–4.68 (m, 1H), 4.56–4.42 (m, 1H), 4.23 (d, J = 4.5 Hz, 1H), 2.22–1.92 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.1, 170.2, 169.3, 169.1, 161.2, 134.5, 132.4, 130.9, 130.8, 129.9, 121.5, 114.9, 88.7, 85.5, 75.8, 65.5, 61.0, 60.9, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2975, 2130, 1682, 1562, 1426, 1365, 1266, 1173, 1151, 991. HRMS (ESI-TOF) calc. [C₂₁H₁₉ClO₈Na⁺] 457.0661, found 457.0660.

Product **3j** was obtained as a yellow oil (56 mg, 0.13 mmol, 67%). ¹H NMR (300 MHz, CDCl₃): $\delta=8.07$ (s, 1H), 7.43–7.37 (m, 2H), 7.32–7.23 (m, 1H), 7.24–7.07 (m, 1H), 5.88–5.78 (m, 1H), 5.23 (t, J=3.0 Hz, 1H), 4.82–4.64 (m, 1H), 4.49 (dd, J=12.1, 7.9 Hz, 1H), 4.21 (dd, J=12.2, 4.5 Hz, 1H), 2.18–2.01 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): $\delta=174.1$, 170.2, 169.3, 169.12, 162.53 (d, J=246.7 Hz), 161.1, 130.4 (d, J=8.5 Hz), 128.7 (d, J=3.2 Hz), 121.6 (d, J=9.3 Hz), 119.4 (d, J=23.3 Hz), 118.1 (d, J=21.2 Hz), 114.9, 88.9 (d, J=3.4 Hz), 85.2, 75.8, 65.5, 61.0, 60.9, 20.7, 20.6, 20.5. IR (ν , cm⁻¹) = 2970, 2132, 1685, 1564, 1326, 1268, 1177, 1151, 1113, 985, 849. HRMS (ESI-TOF) calc. [C₂₁H₁₉FO₈Na⁺] 441.0956, found 441.0956.

Product 3k was obtained as a yellow oil (52 mg, 0.13 mmol, 65%). 1 H NMR (300 MHz, CDCl₃): $\delta = 8.74$ (s, 1H), 8.60 (d, J = 4.0 Hz, 1H), 8.01 (s, 1H), 7.87–7.73 (m, 1H), 7.28 (dd, J = 7.9, 5.0 Hz, 1H), 5.74 (s, 1H), 5.16 (s, 1H), 4.68–4.58 (m, 1H), 4.42 (dd, J = 12.2, 7.9 Hz, 1H), 4.13 (dd, J = 12.1, 4.5 Hz, 1H), 2.09–1.90 (m, 9H). 13 C NMR (75 MHz, CDCl₃): $\delta = 173.9$, 170.2, 169.3, 169.1, 161.3, 153.0, 150.6, 139.6, 123.2, 117.2, 114.9, 87.5, 86.8, 75.8, 65.5, 61.0, 60.8, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 2859, 2119, 1680, 1566, 1326, 1181, 992. HRMS (ESI-TOF) calc. $[C_{20}H_{19}NO_8Na^{+}]$ 424.1003, found 444.1002.

Product 3l was obtained as a yellow oil (53 mg, 0.14 mmol, 72%). 1 H NMR (300 MHz, CDCl $_3$): $\delta=7.90$ (s, 1H), 5.72 (s, 1H), 5.18 (s, 1H), 4.64–4.54 (m, 1H), 4.44 (dd, J=12.1, 7.8 Hz, 1H), 4.19 (d, J=4.5 Hz, 1H), 2.19–1.91 (m, 9H), 1.45–1.42 (m, 1H), 1.07–0.86 (m, 4H). 13 C NMR (75 MHz, CDCl $_3$): $\delta=174.8$, 174.8, 170.5, 169.7, 169.5, 160.8, 115.1, 98.5, 75.8, 66.0, 61.5, 61.3, 21.0, 21.0, 20.9, 9.8, 9.8. IR (ν , cm $^{-1}$) = 2915, 2138, 1682, 1566, 1365, 1175, 1149, 991, 864. HRMS (ESI-TOF) calc. [$C_{18}H_{20}O_8Na^{+1}$] 387.1050, found 387.1051.

Product 3m was obtained as a yellow oil (64 mg, 0.16 mmol, 80%). ¹H NMR (300 MHz, CDCl₃): δ = 7.82 (s, 1H), 5.60–5.48 (m, 1H), 5.02 (t, J = 3.1 Hz, 1H), 4.51–4.39 (m, 1H), 4.26 (dd, J = 12.2, 7.9 Hz, 1H), 4.01 (dd, J = 12.2, 4.5 Hz, 1H), 1.99–1.81 (m, 9H), 0.08 (s, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 174.8, 170.9, 170.03, 169.8, 162.1, 115.6, 99.7, 98.6, 76.4, 66.3, 61.7, 61.7, 21.4, 21.3, 21.3, 0.0. IR (ν , cm⁻¹) = 2864, 2028, 1914, 1685, 1566, 1324, 1261, 1175, 1151, 987, 817. HRMS (ESI-TOF) calc. [C₁₈H₂₄O₈-SiNa⁺] 419.1133, found 419.1135.

Product 3n was obtained as a yellow oil (53 mg, 0.14 mmol, 70%). 1 H NMR (300 MHz, CDCl $_3$): δ = 7.95 (s, 1H), 5.73 (dd, J = 3.1, 1.7 Hz, 1H), 5.19 (t, J = 3.1 Hz, 1H), 4.69–4.58 (m, 1H), 4.45 (dd, J = 12.1, 7.8 Hz, 1H), 4.17 (dd, J = 12.1, 4.5 Hz, 1H), 2.39 (t, J = 7.0 Hz, 2H), 2.18–1.96 (m, 9H), 1.69–1.36 (m, 4H), 0.94 (t, J = 7.3 Hz, 3H). 13 C NMR (75 MHz, CDCl $_3$): δ = 174.7, 170.2, 169.3, 169.1, 160.7, 114.9, 94.1, 77.0, 75.5, 65.6, 61.1, 60.9, 29.7, 22.0, 20.7, 20.6, 20.6, 18.6, 13.4. IR (ν , cm $^{-1}$) = 2838, 2862, 2147, 1685, 1566, 1365, 1324, 1175, 1149, 991, 864. HRMS (ESI-TOF) calc. [C_{19} H $_{24}$ O $_{8}$ Na $^{+}$] 403.1363, found 403.1361.

Product 3o was obtained as a yellow oil (45 mg, 0.12 mmol, 62%). 1 H NMR (300 MHz, CDCl $_3$): $\delta = 7.76$ (s, 1H), 5.53 (dd, J = 3.1, 1.7 Hz, 1H), 4.99 (t, J = 3.1 Hz, 1H), 4.49–4.37 (m, 1H), 4.25 (dd, J = 12.1, 7.8 Hz, 1H), 3.97 (dd, J = 12.1, 4.5 Hz, 1H), 2.17 (t, J = 7.1 Hz, 2H), 1.94–1.81 (m, 9H), 1.44 (h, J = 7.2 Hz, 2H), 0.84 (t, J = 7.4 Hz, 3H). 13 C NMR (75 MHz, CDCl $_3$): $\delta = 174.7$, 170.2, 169.3, 169.1, 160.7, 114.9, 93.9, 77.8, 75.5, 65.6, 61.1, 60.9, 21.2, 20.8, 20.7, 20.6, 20.6, 13.5. IR (ν , cm $^{-1}$) = 2916, 2879, 1680, 1560, 1141989, 836, 724. HRMS (ESI-TOF) calc. [$C_{18}H_{22}O_8Na^+$] 389.1207, found 403.1361.

Product **3p** was obtained as a yellow oil (67 mg, 0.15 mmol, 78%). 1 H NMR (300 MHz, CDCl₃): δ = 7.61 (s, 1H), 7.32–7.11 (m, 5H), 5.65–5.52 (m, 1H), 5.16–5.04 (m, 1H), 4.57–4.49 (m, 1H), 4.36 (dd, J = 11.8, 7.8 Hz, 1H), 4.07 (dd, J = 12.0, 4.5 Hz, 1H), 2.84 (t, J = 7.3 Hz, 2H), 2.63 (t, 2H), 2.05–1.94 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 174.5, 170.2, 169.3, 169.1, 161.1, 139.5, 128.5, 128.3, 126.7, 114.9, 92.7, 78.3, 75.4, 65.6, 60.9, 60.8, 33.8, 21.0, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 2926, 2840, 2149, 1685,

1566, 1324, 1261, 1175, 1151, 1017, 991, 678. HRMS (ESI-TOF) calc. $[C_{23}H_{24}O_8Na^{\dagger}]$ 451.1363, found 451.1361.

Product 3**q** was obtained as a yellow oil (73 mg, 0.17 mmol, 86%). 1 H NMR (300 MHz, CDCl₃): $\delta = 7.91$ (s, 1H), 5.68 (d, J = 2.3 Hz, 1H), 5.12 (t, J = 3.0 Hz, 1H), 4.62–4.52 (m, 1H), 4.37 (dd, J = 12.2, 7.8 Hz, 1H), 4.12 (dd, J = 12.1, 4.4 Hz, 1H), 2.68 (s, 1H), 2.09–1.98 (m, 9H), 1.93–1.84 (m, 2H), 1.72–1.43 (m, 8H). 13 C NMR (75 MHz, CDCl₃): $\delta = 174.3$, 170.2, 169.5, 169.1, 161.0, 114.7, 95.5, 79.9, 75.6, 65.5, 61.1, 60.9, 39.1, 39.1, 24.9, 22.9, 20.7, 20.6, 20.6. IR (ν , cm $^{-1}$) = 3363, 2840, 2766, 2136, 1691, 1568, 1326, 1182, 1156, 996. HRMS (ESI-TOF) calc. [$C_{21}H_{26}O_9Na^+$] 445.1469, found 445.1467.

Product 3r was obtained as a yellow oil (101 mg, 0.14 mmol, 70%). 1 H NMR (300 MHz, CDCl₃): $\delta = 8.07$ (s, 2H), 7.61 (d, J = 2.6 Hz, 4H), 5.95–5.73 (m, 2H), 5.29–5.10 (m, 2H), 4.77–4.58 (m, 2H), 4.58–4.46 (m, 2H), 4.23 (dd, J = 9.7, 5.6 Hz, 2H), 2.25–1.97 (m, 18H). 13 C NMR (75 MHz, CDCl₃): $\delta = 174.0$, 170.2, 169.3, 169.1, 161.2, 132.8, 122.0, 114.9, 89.0, 87.0, 75.8, 65.5, 61.0, 60.8, 20.7, 20.6, 20.6. IR (ν , cm⁻¹) = 2870, 2129, 1685, 1564, 1324, 1264, 1177, 1151, 989. HRMS (ESI-TOF) calc. [$C_{36}H_{34}O_{16}Na^{+}$] 745.1739, found 745.1735.

Product 5a was obtained as a yellow oil (61 mg, 0.13 mmol, 67%). 1 H NMR (300 MHz, CDCl₃): $\delta = 9.15$ (s, 1H), 8.02 (s, 1H), 7.46–7.34 (m, 5H), 5.96–5.85 (m, 1H), 5.55 (d, J = 1.4 Hz, 1H), 5.30 (s, 2H), 4.64 (d, J = 5.3 Hz, 1H), 4.54–4.44 (m, 1H), 4.23 (dd, J = 12.2, 4.7 Hz, 1H), 2.19–2.05 (m, 9H). 13 C NMR (75 MHz, CDCl₃): $\delta = 180.3$, 169.3, 168.4, 168.2, 161.0, 132.6, 128.3, 128.1, 127.3, 126.5, 126.5, 111.5, 73.6, 64.9, 60.5, 60.1, 53.4, 19.7, 19.6, 19.6. IR (ν , cm $^{-1}$) = 3261, 2866, 2836, 1680, 1560, 1475, 1324, 1179, 1149, 989, 706. HRMS (ESI-TOF) calc. [C₂₂H₂₃N₃O₈Na $^+$] 480.1377, found 480.1375.

Product **5b** was obtained as a yellow oil (69 mg, 0.14 mmol, 72%). ¹H NMR (300 MHz, CDCl₃): δ = 9.15 (s, 1H), 8.67 (s, 1H), 8.45 (s, 1H), 7.87 (s, 1H), 7.55–7.42 (m, 2H), 7.30 (d, J = 2.8 Hz, 1H), 6.68–6.52 (m, 1H), 5.97–5.84 (m, 1H), 5.23 (t, J = 2.9 Hz, 1H), 4.65–4.56 (m, 1H), 4.46 (m, 1H), 4.21 (m, 1H), 2.06–1.97 (m, 9H). ¹³C NMR (75 MHz, CDCl₃): δ = 181.5, 170.3, 169.5, 169.3, 162.0, 148.1, 135.90, 128.1, 126.7, 126.2, 126.2, 115.5, 113.5, 112.6, 112.1, 103.5, 74.7, 66.0, 61.6, 61.2, 20.8, 20.7, 20.6. IR (ν , cm⁻¹) = 2834, 2862, 2779, 1682, 1560, 1475, 1460, 1324, 1162, 1011, 989, 700. HRMS (ESI-TOF) calc. [C₂₃H₂₂N₄O₈Na[†]] 505.1330, found 505.1329.

Product 5c was obtained as a yellow oil (70 mg, 0.14 mmol, 70%). 1 H NMR (300 MHz, CDCl₃): δ = 9.13 (s, 1H), 9.07 (s, 1H), 8.58 (s, 1H), 8.37 (t, 1H), 8.24 (d, J = 8.0 Hz, 1H), 7.82 (d, J = 8.9 Hz, 1H), 5.93–5.89 (m, 1H), 5.24–5.22 (m, 1H), 4.64–4.57 (m, 1H), 4.47 (dd, J = 12.0, 7.8 Hz, 1H), 4.20 (dd, J = 12.0, 4.5 Hz, 1H), 2.08–1.94 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 181.1, 170.3, 169.4, 169.2, 162.2, 162.1, 156.2, 148.6, 135.2, 126.0, 124.9, 120.0, 119.3, 115.1, 112.7, 74.8, 65.9, 61.5, 61.1, 20.8, 20.7, 20.6. IR (ν , cm $^{-1}$) = 2985, 2864, 2037, 1687, 1562, 1186, 1154, 994, 838, 855. HRMS (ESI-TOF) calc. [$C_{22}H_{20}N_4O_8SNa^+$] 523.0894, found 523.0890.

Product 5**d** was obtained as a yellow oil (89 mg, 0.13 mmol, 65%). 1 H NMR (300 MHz, CDCl₃): $\delta = 9.09$ (s, 1H), 8.48 (s, 1H), 7.95 (d, J = 8.4 Hz, 1H), 7.49 (s, 1H), 7.40–7.21 (m, 11H), 6.28 (d, J = 3.5 Hz, 1H), 5.98–5.85 (m, 1H), 5.29 (s, 2H), 5.18 (s, 2H),

4.68–4.54 (m, 1H), 4.50–4.41 (m, 1H), 4.18 (dd, J=12.1, 4.5 Hz, 1H), 2.10–1.92 (m, 9H). 13 C NMR (75 MHz, CDCl₃): $\delta=201.9$, 180.2, 170.4, 169.8, 169.1, 164.9, 163.5, 159.3, 139.6, 135.6, 135.5, 133.7, 128.7, 128.5, 128.2, 128.2, 128.2, 127.2, 125.6, 121.5, 113.7, 112.7, 111.6, 105.9, 71.8, 71.1, 67.1, 65.3, 61.1, 60.5, 20.8, 20.6, 20.5. IR (ν , cm⁻¹) = 2967, 2931, 1687, 1559, 1195, 1169, 1046, 998. HRMS (ESI-TOF) calc. [C₃₆H₃₃N₃O₁₁Na⁺] 706.2007, found 706.2004.

Product **5e** was obtained as a yellow oil (57 mg, 0.12 mmol, 58%). 1 H NMR (300 MHz, CDCl₃) δ = 9.07 (s, 1H), 8.69–8.59 (m, 2H), 8.30 (d, J = 8.0 Hz, 1H), 8.11 (d, J = 8.5 Hz, 1H), 7.74 (t, J = 8.1 Hz, 1H), 6.28 (d, J = 3.5 Hz, 1H), 5.92–5.83 (m, 1H), 4.66–4.57 (m, 1H), 4.52–4.41 (m, 1H), 4.19 (dd, J = 12.1, 4.5 Hz, 1H), 2.09–1.98 (m, 9H). 13 C NMR (75 MHz, CDCl₃): δ = 180.1, 170.4, 169.8, 169.1, 163.6, 137.0, 131.2, 126.0, 125.7, 123.9, 115.8, 115.8, 113.7, 112.7, 74.9, 71.8, 61.5, 61.1, 20.7, 20.7, 20.6. IR (ν , cm $^{-1}$) = 2868, 1687, 1562, 1486, 1309, 1188, 1171, 994, 717. HRMS (ESITOF) calc. [$C_{21}H_{20}N_4O_{10}Na^+$] 511.1072, found 511.1071.

Product **5f** was obtained as a yellow oil (81 mg, 0.17 mmol, 86%). 1 H NMR (300 MHz, CDCl₃): $\delta = 9.11$ (s, 1H), 8.41 (s, 1H), 7.58 (d, J = 9.0 Hz, 2H), 6.98 (d, J = 8.9 Hz, 2H), 5.89 (t, 2H), 5.21 (t, J = 3.1 Hz, 1H), 4.64–4.55 (m, 1H), 4.44 (dd, J = 12.1, 7.7 Hz, 1H), 4.19 (dd, J = 12.0, 4.8 Hz, 3H), 2.08–1.94 (m, 9H). 13 C NMR (75 MHz, CDCl₃): $\delta = 181.3$, 170.3, 169.4, 169.2, 162.0, 160.3, 129.6, 127.8, 125.7, 122.3, 115.0, 112.6, 74.7, 65.9, 61.6, 61.1, 55.6, 20.7, 20.7, 20.6. IR (ν , cm $^{-1}$) = 2902, 2875, 1687, 1564, 1471, 1326, 1262, 1184, 1153, 998, 838. HRMS (ESI-TOF) calc. [$C_{22}H_{23}N_3O_9Na^+$] 496.1327, found 496.1329.

Conflicts of interest

There are no conflicts to declare.

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