



Cite this: *Nanoscale Adv.*, 2025, **7**, 934

## Extracellular vesicles: from intracellular trafficking molecules to fully fortified delivery vehicles for cancer therapeutics

Adham H. Mohamed, <sup>†a</sup> Tasneem Abaza, <sup>†bc</sup> Yomna A. Youssef, <sup>de</sup> Mona Rady, <sup>fg</sup> Sherif Ashraf Fahmy, <sup>h</sup> Rabab Kamel, <sup>i</sup> Nabila Hamdi, <sup>j</sup> Eleni Efthimiado, <sup>k</sup> Maria Braoudaki <sup>l</sup> and Rana A. Youness <sup>\*e</sup>

Extracellular vesicles (EVs) are emerging as viable tools in cancer treatment due to their ability to carry a wide range of theranostic activities. This review summarizes different forms of EVs such as exosomes, microvesicles, apoptotic bodies, and oncosomes. It also sheds the light onto isolation methodologies, characterization techniques and therapeutic applications of all discussed EVs. Evidence indicates that EVs are particularly effective in delivering chemotherapeutic medications, and immunomodulatory agents. However, the advancement of EV-based therapies into clinical practice is hindered by challenges including EVs heterogeneity, cargo loading efficiency, and *in vivo* stability. Overall, EVs have the potential to change cancer therapeutic paradigms. Continued research and development activities are critical for improving EV-based medications and increasing their therapeutic impact.

Received 10th May 2024  
Accepted 22nd December 2024

DOI: 10.1039/d4na00393d

[rsc.li/nanoscale-advances](http://rsc.li/nanoscale-advances)

## Introduction

Extracellular Vesicles (EVs) are lipid-bound structures released by all cell types, acting as essential mediators of intercellular communication across diverse organisms.<sup>1–4</sup> Initially thought to be mere cellular waste disposal mechanisms,<sup>5</sup> the

understanding of EVs has evolved dramatically over the past three decades. They are now recognized as crucial players in intracellular trafficking and the transportation of various biomolecules between cells.<sup>5–8</sup>

EVs represent a captivating class of nanosized, membrane-enclosed particles that act as cellular messengers.<sup>9</sup> These dynamic structures, primarily encompassing exosomes and microvesicles (MVs), serve as potent communication tools, shuttling a diverse cargo of molecules between cells.<sup>10</sup> Notably, specific cell types and conditions can further diversify EVs, leading to subtypes such as apoptotic bodies shed during programmed cell death and large oncosomes secreted by cancer cells.<sup>11,12</sup> This review will delve into the distinct biogenesis, release mechanisms, size, composition, and functional roles of these multifunctional EVs subtypes.<sup>1,13,14</sup>

EVs act as versatile communication hubs, transporting a meticulously selected cargo that includes DNA, various RNA species like messenger RNA (mRNA)<sup>15</sup> and non-coding RNAs (ncRNAs) such as microRNAs (miRNAs),<sup>16–18</sup> long non-coding RNAs (lncRNAs),<sup>19–22</sup> and circular RNAs (circRNAs).<sup>23–25</sup> Additionally, proteins, metabolites, lipids, and glycoconjugates derived from the parent cell can be packaged within EVs.<sup>26</sup> The protective lipid bilayer surrounding EVs shields their cargo from degradation by extracellular enzymes, allowing for long-distance travel through the bloodstream to reach distant target tissues.<sup>26</sup> The precise mechanisms governing cargo selection and packing remain an active area of research, as the cargo composition within EVs often differs significantly from the parent cell's internal environment.<sup>26</sup>

<sup>a</sup>Department of Chemistry, Faculty of Science, Cairo University, 12613, Giza, Egypt

<sup>b</sup>Biotechnology and Biomolecular Chemistry Program, Faculty of Science, Cairo University, 12613, Giza, Egypt

<sup>c</sup>Université Paris-Saclay, Université d'Evry Val D'Essonne, 91000 Évry-Courcouronnes, Île-de-France, France

<sup>d</sup>Department of Physiology, Faculty of Physical Therapy, German International University (GIU), 11835, Cairo, Egypt

<sup>e</sup>Molecular Biology and Biochemistry Department, Faculty of Biotechnology, German International University (GIU), 11835, Cairo, Egypt. E-mail: [rana.youness21@gmail.com](mailto:rana.youness21@gmail.com); [rana.youness@giu-uni.de](mailto:rana.youness@giu-uni.de)

<sup>f</sup>Microbiology, Immunology and Biotechnology Department, Faculty of Pharmacy and Biotechnology, German University in Cairo (GUC), 11835, Cairo, Egypt

<sup>g</sup>Faculty of Biotechnology, German International University, New Administrative Capital, 11835, Cairo, Egypt

<sup>h</sup>Department of Pharmaceutics and Biopharmaceutics, University of Marburg, Robert-Koch-Str. 4, 35037 Marburg, Germany

<sup>i</sup>Pharmaceutical Technology Department, National Research Centre, 12622, Cairo, Egypt

<sup>j</sup>Pharmacology and Toxicology Department, Faculty of Pharmacy and Biotechnology, German University in Cairo (GUC), 11835, Cairo, Egypt

<sup>k</sup>Inorganic Chemistry Laboratory, Chemistry Department, National and Kapodistrian University of Athens, Athens, Greece

<sup>l</sup>Department of Clinical, Pharmaceutical, and Biological Science, School of Life and Medical Sciences, University of Hertfordshire, Hatfield AL10 9AB, UK

<sup>†</sup> Equal contribution.



The impact of EVs on recipient cells is multifaceted and depends on several factors. These factors include the specific cell types and tissues interacting with the EVs, the cargo content and quantity, and the recipient cell's ability to process and utilize the cargo.<sup>26</sup> While the details of how EVs interact with recipient cells and how their cargo is delivered inside remain under investigation, various mechanisms are being actively explored.<sup>27</sup>

In the context of cancer, EVs emerge as intriguing players. They exhibit a selective packaging and delivery of cargo, fostering a dynamic communication network between tumors and the host organism.<sup>28</sup> This intricate interplay significantly influences all aspects of tumor biology, impacting tumor initiation, progression, and metastasis.<sup>29–32</sup> EVs, released by both cancerous and non-cancerous cells, can exert local and long-distance effects, influencing various cell types within the tumor microenvironment and potentially affecting all biological systems of the patient.<sup>33</sup>

The unique properties of EVs, including low toxicity, excellent biocompatibility, low immunogenicity, and inherent targeting capabilities, make them highly promising candidates for drug delivery systems in various diseases, including cancer.<sup>34</sup> Research efforts are directed towards engineering therapeutic EVs by modifying normal cells to optimize their cargo and targeting properties.<sup>24</sup> While initial studies explored cancer cell lines like HeLa cells for proof-of-concept, the oncological properties of cancer cell-derived EVs are now recognized.<sup>35</sup> Consequently, the focus has shifted towards utilizing intact normal cells as sources for therapeutic EVs production.

## Types of EVs and their composition

### Exosomes

Among the diverse EVs subtypes, exosomes stand out for their diminutive size, typically ranging 30–150 nm in diameter as shown in Table 1.<sup>58</sup> These nano-messengers are meticulously produced through the endosomal pathway. This intricate process involves inward budding of the limiting membrane of early endosomes, leading to the formation of multivesicular bodies (MVBs) (Fig. 1).<sup>59</sup> Once mature, MVBs can either fuse with the cell membrane and release their exosome cargo into the extracellular space, or they can fuse with lysosomes for degradation. Exosomes play a vital role in a multitude of biological processes.<sup>1,13,36,37</sup> They serve as crucial mediators of intercellular and intracellular communication, facilitating the exchange of proteins, lipids, and nucleic acids between cells.<sup>59</sup> Their involvement extends to various physiological functions such as immune response stimulation, myelin sheath formation, tissue repair, and neural survival.<sup>1,13,37</sup>

Biogenesis of exosomes is a complex process, exosomes are generated through intricate cellular mechanisms. While the endosomal sorting complex required for transport (ESCRT) machinery plays a pivotal role in their biogenesis, alternative pathways exist. The ESCRT pathway, comprising proteins like ALIX, TSG101, Chmp4, and SKD1, is instrumental in forming multivesicular bodies (MVBs) and subsequent exosome release.<sup>1,38,60</sup> However, ESCRT-independent mechanisms

involving proteins such as CD63, CD81, TSG101 (note: TSG101 participates in both pathways), ARF6, and heparanase also contribute to exosome production (Fig. 1).<sup>38</sup> These complex pathways underscore the diverse mechanisms governing exosome biogenesis and highlight its intricate relationship with cellular processes.

Exosome release is influenced by various factors, including intracellular calcium levels, hypoxia, growth factors, TNF- $\alpha$ , heparanase, and glutamate.<sup>61</sup> Once released, exosomes interact with recipient cells through multiple mechanisms, including clathrin-mediated endocytosis, caveolin-mediated endocytosis, macropinocytosis, and direct fusion with the plasma membrane.<sup>38,39</sup> These uptake processes determine the intracellular fate of exosomes and their cargo, ultimately influencing cellular responses.

Exosomes are heterogeneous nanovesicles characterized by a lipid bilayer encapsulating a diverse cargo of biomolecules.<sup>9,62</sup> This cargo encompasses proteins, lipids, and nucleic acids, including DNA, mRNA, various ncRNAs (miRNAs, lncRNAs, circRNAs), and metabolites.<sup>24,58,63–66</sup> The specific composition of exosomes is influenced by their cell of origin, highlighting their potential as biomarkers for distinct cellular states and pathologies.<sup>67,68</sup>

Lipid composition is a critical determinant of exosome structure and function. While sphingomyelin is enriched in some exosomes, particularly those derived from brain cells, other lipids like cardiolipins can also be abundant.<sup>6,40</sup> Notably, exosomal lipid profiles often differ from their parent cells, with elevated levels of glycolipids, free fatty acids, and phosphatidylserine, and decreased levels of phosphatidylcholine, diacylglycerol, phosphatidylinositol, and phosphatidylethanolamines.<sup>5,40</sup>

Protein content is another key aspect of exosome characterization. Exosomes carry a variety of proteins, including transmembrane and cytoplasmic proteins, with functions spanning cell adhesion, immune response, and intracellular signaling.<sup>69</sup> Specific protein markers like CD63, CD9, CD81, ALIX, TSG101, HSC70, and HSP90 $\beta$  are commonly used to identify and isolate exosomes (Table 1). However, the protein composition is highly diverse and reflects the origin and functional state of the parent cell.<sup>70</sup>

### Microvesicles

Microvesicles (MVs) are a heterogeneous population of EVs ranging in size 100–1000 nm.<sup>14</sup> Unlike exosomes, which originate from the endosomal pathway, MVs are directly shed from the plasma membrane *via* outward budding (Fig. 1).<sup>70</sup> This process is influenced by a complex interplay of factors, including phospholipid redistribution, cytoskeletal dynamics, and the involvement of specific proteins such as SNAREs, tethering factors, molecular motors, and components of the ESCRT machinery.<sup>47,71</sup>

MVs carry a diverse cargo, including proteins, lipids, and nucleic acids similar to exosomes.<sup>72</sup> Their lipid composition differs significantly from the parental cell membrane, with increased levels of phosphatidylserine and decreased levels of





Table 1 Classification of extracellular vesicles (EVs)

Extracellular vesicles (EVs)	Size	Composition	Biogenesis	Production and release stimuli	Markers	Functions	Ref.
Exosomes	30–150 nm	Nucleic acids: DNA, mRNA, miRs, other non-coding RNAs Lipids: ↓ phosphatidylcholine diacylglycerol, phosphatidylinositol, phosphatidylethanolamines ↑ Glycolipids, free fatty acids, phosphatidylserine Some are enriched with cardiolipins and sphingomyelin Proteins: CD81, Alix, TSG101, glycoproteins	Endosomal pathway (ESCRT-dependent or independent)	↑ Intracellular $\text{Ca}^{2+}$ , hypoxia Terminus (GIPC) depletion	Alix, CD63, CD9, CD81 TSG101, HSC70, and HSP90 $\beta$	Intercellular communications, immune response stimulation, tumor progression, myelin sheath formation, neural survival, tissue repair, others	1, 2, 6, 13 and 36–46
Microvesicles	100–1000 nm	Nucleic acids: DNA, mRNA, non-coding RNAs	Outward budding of plasma membrane initiated by translocases and ARF6	↑ Intracellular $\text{Ca}^{2+}$ , gamma-radiation	Integrins, selectins, CD40 ligand, ARF6, VCAMP3 and MMP	Intercellular communications, thrombosis, regulation of inflammatory response and angiogenesis, increasing apoptosis rate, autophagy promotion, cartilage regeneration Some act as anti-inflammatory modulators	1, 2, 6, 13, 38, 39, 41 and 47–50
Apoptotic bodies	50–5000 nm	Cellular organelles Nucleic acid: DNA, mRNA, non-coding RNAs Lipids: ↑ translocated phosphatidylserine Proteins: integrins, selectins, CD40 ligand, ARF6, VCAMP3 adhesion proteins	Outward budding of the plasma membrane of apoptotic cells <i>via</i> caspase-dependent and independent pathways	Heat, ROS, radiation, ↑ intracellular $\text{Ca}^{2+}$ , hypoxia	Thrombospondin, annexin V, C3b, phosphatidyl-serine	Debris clearance Immunomodulatory functions	1 and 51–53
Oncosomes	Small: 100–400 nm Large: 1–10 $\mu\text{m}$	Nucleic acid: DNAs, RNAs and non-coding RNAs Lipids: ↑ phosphatidylserine exposure and flipping ↑ Content of cholesterol Proteins: ARF6, MMP, Annexin A1, Annexin A2, oncogenic proteins complexes	Outward budding of the plasma membrane of tumor cells	Activation of EGFR & AKT pathways Silencing DIAPH3 by EKR	ARF6, CK18, GAPDH, MMP, Annexin A1, Annexin A2, oncogenic proteins complexes	Development, growth and metastasis of tumors	12 and 54–57

phosphatidylcholine, phosphatidylinositol, and phosphatidylethanolamine.<sup>73</sup> Sphingomyelin and ceramides are enriched in MV membranes, while cholesterol ester content varies significantly.<sup>40</sup>

The protein cargo of MVs is equally complex, encompassing proteins involved in various cellular functions, including signaling, adhesion, and structural maintenance.<sup>71</sup> Notably, MVs are enriched in specific marker proteins such as integrins, selectins, CD40 ligand, ARF6, and VCAMP3, facilitating their identification and characterization (Table 1).<sup>41,47,48</sup>

### Apoptotic bodies

Apoptotic bodies, the largest of the EVs subtypes, are generated during programmed cell death or apoptosis.<sup>74</sup> These membrane-bound structures, ranging from 50–5000 nm in diameter, encapsulate cellular debris, including organelles and nuclear fragments<sup>70</sup> (Fig. 1). Apoptosis, a tightly regulated process, is initiated by various stimuli such as heat, radiation, or oxidative stress and involves distinct pathways, including caspase-dependent and independent mechanisms.<sup>74,75</sup>

Beyond their role in cellular clearance, apoptotic bodies actively participate in several biological processes. They serve as immunomodulatory agents, influencing immune responses, and are recognized by phagocytes through “eat-me” signals, such as phosphatidylserine exposed on their outer membrane.<sup>47</sup>

The lipid composition of apoptotic bodies is distinct from that of living cells, with increased phosphatidylserine and oxidized lipids, facilitating their recognition and clearance.<sup>47</sup> Protein content is also characteristic, including heat shock proteins and other molecules involved in apoptotic processes.<sup>51</sup>

In contrast to exosomes and MVs, oxidative modifications to the apoptotic bodies' cell surface create binding sites for proteins like thrombospondin and complement component C3b, further facilitating phagocytosis as summarized in Table 1.<sup>47</sup> These markers, along with annexin V and T cell immunoglobulin mucin-4 (TIM4), are commonly used to identify apoptotic bodies.<sup>47,75</sup>

### Oncosomes

Oncosomes, a distinct subtype of EVs, were first described by Rak's group in 2008 as large vesicles shed from glioma cells.<sup>12</sup> These abnormally large EVs, ranging from 1–10  $\mu\text{m}$  in diameter, are characterized by their aberrant cargo, including oncogenic proteins.<sup>2</sup>

They are non-apoptotic blebs originating from the plasma membrane of invasive cancer cells. The loss of the cytoskeletal regulator diaphanous-related formin-3 (DIAPH3) promotes the creation and release of the oncosomes. Subsequently, this causes a shift in the cells' phenotype from mesenchymal to a more invasive, fast, and metastatic phenotype called “the amoeboid phenotype”. The formation of these oncosomes also requires various proteins such as GTPase RhoA or its effector ROCK.<sup>12,54</sup> Additionally, it has been reported that oncosomes can be released and formed by activating EGFR and AKT pathways (Fig. 1).<sup>2</sup>

In fact, only malignant cells produce a detectable quantity of lipo-oligosaccharide (LOs), which appears to correlate with tumor aggressiveness. Conversely, the production of LOs by benign cells is insignificant. Many tumor forms, such as those of the prostate, breast, bladder, lung cancer, and others, have the trait of tier shedding.<sup>76</sup> They play a big role in the development, growth, and metastasis of tumors as a result of oncogenic protein complexes and molecules being overexpressed and exported between tumor cells and stroma through the oncosomes.<sup>55</sup>

Similar to other EVs, oncosomes possess a lipid bilayer encapsulating a diverse cargo of proteins, lipids, and nucleic acids as summarized in Table 1.<sup>54,76</sup> However, they exhibit distinct compositional features, including elevated levels of phosphatidylserine and cholesterol compared to the parental cell membrane.<sup>12,54</sup> Moreover, oncosomes are enriched in specific proteins, such as ARF6, CK18, GAPDH, MMPs, annexins, and oncogenic protein complexes.<sup>41,54,56</sup> Notably, these large EVs like oncosomes carry substantial amounts of extracellular DNA, distinguishing them from smaller EVs subtypes such as exosomes.<sup>57</sup>

### Miscellaneous types

There are other types and classifications of EVs. For example, autophagic EVs, produced from the autophagy process are involved in the degradation of cellular components, contributing to cellular homeostasis.<sup>77</sup> Another type is stressomes (called damaged EVs) which emerge in response to cellular damage or stress, serving as messengers during challenging conditions.<sup>78</sup> Additionally, matrix vesicles, migrasomes, and others are recognized as distinct types of EVs.<sup>77</sup>

## EVs as drug delivery vehicles

EVs have emerged as promising platforms for drug delivery due to their intrinsic properties.<sup>79</sup> Possessing low toxicity, excellent biocompatibility, and inherent targeting capabilities, EVs offer significant advantages over traditional synthetic carriers.<sup>80</sup> Their ability to encapsulate a diverse array of therapeutic cargos, including nucleic acids, proteins, and small molecules, further enhances their potential as drug delivery vehicles.<sup>81</sup> To harness the full therapeutic potential of EVs, meticulous preparation is essential.<sup>80,82</sup> The development of EVs as drug delivery vehicles requires a systematic approach encompassing three key stages: EVs isolation, cargo loading and EVs engineering that will be discussed in the coming sections.

### Isolation of EVs

EVs are heterogeneous populations as described earlier that are characterized by their size, density, and surface composition. To isolate these nanosized particles, scientists employ a variety of techniques, each with its own advantages and limitations.<sup>83</sup> Conventional methods include: ultracentrifugation, membrane filtration, chromatography and microfluidics as an emerging promising approach, as microfluidics enables high-throughput EVs isolation with potential for automation.<sup>84</sup> The choice of



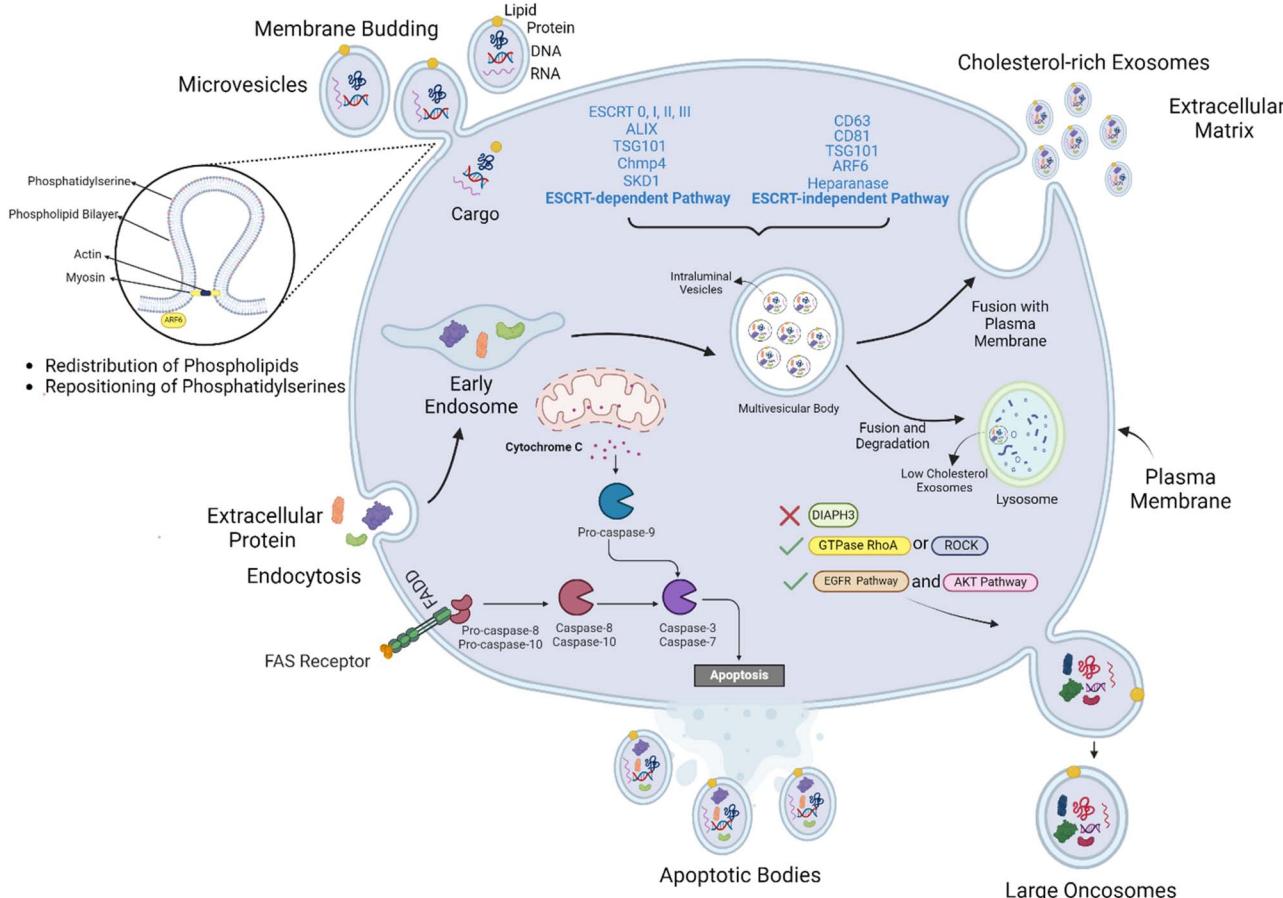


Fig. 1 Biogenesis and secretion of different types of extracellular vesicles.

isolation technique depends on factors such as the desired EVs purity, sample volume, available resources, and downstream applications. It is often necessary to combine multiple methods to achieve optimal EVs isolation and characterization.<sup>83</sup>

**Density-based EV isolation: ultracentrifugation techniques.** Ultracentrifugation remains the gold standard for EVs isolating based on their density and size.<sup>82</sup> This method involves sequential centrifugation steps at increasing speeds to fractionate biological samples as previously reviewed.<sup>85</sup> Differential ultracentrifugation is a commonly used approach where crude cell lysates or biological fluids are subjected to low-speed centrifugation ( $20\,000\times g$ ) to remove cellular debris, followed by higher-speed centrifugation to pellet EVs ( $100\,000\times g$ ) as shown in Fig. 2. However, this method is often associated with low EVs yield and purity, especially when working with viscous samples.<sup>82,86</sup>

On the other hand, it has been recommended that to enhance EVs purity and recovery, density gradient ultracentrifugation should be employed.<sup>85</sup> This technique involves layering a sample onto a preformed density gradient, allowing EVs to migrate and band at their corresponding buoyant density. This method provides better resolution and allows for the isolation of EVs subpopulations based on density differences as shown in Fig. 2. Despite its advantages, density gradient

ultracentrifugation is time-consuming and requires specialized equipment.<sup>82</sup>

#### Size-based methods

**Size-exclusion chromatography (SEC).** SEC separates EVs based on their size by utilizing a porous matrix column. Larger EVs elute first, while smaller particles are retained within the column.<sup>87</sup> This technique preserves EV biophysical properties, offering advantages over ultracentrifugation as shown in Table 2. According to a study performed by Carso *et al.*, EVs may be reasonably purified to 70–80% purity levels using a commercially available blind-elute-SEC, which is dependable and scalable.<sup>98</sup> Accordingly, challenges such as low recovery rates and potential contamination with larger particles may arise (Fig. 3).

**Polymer precipitation.** Polymer precipitation involves the addition of a polymer, such as polyethylene glycol (PEG), to induce EVs aggregation and subsequent precipitation (Fig. 3). PEG was employed to separate EVs from cell culture media quickly and inexpensively. A PEG/dextran aqueous two-phase system was also used, with a recovery efficiency of about 70%; four times higher than ultracentrifugation.<sup>90,91</sup> This method is rapid and cost-effective but often results in low purity due to co-precipitation of contaminants as summarized in Table 2.

**Filtration.** Filtration is a highly facile and efficient method for separating EVs based on their size.<sup>82</sup> Filtration leverages pore-sized membranes to retain EVs, offering a relatively fast and



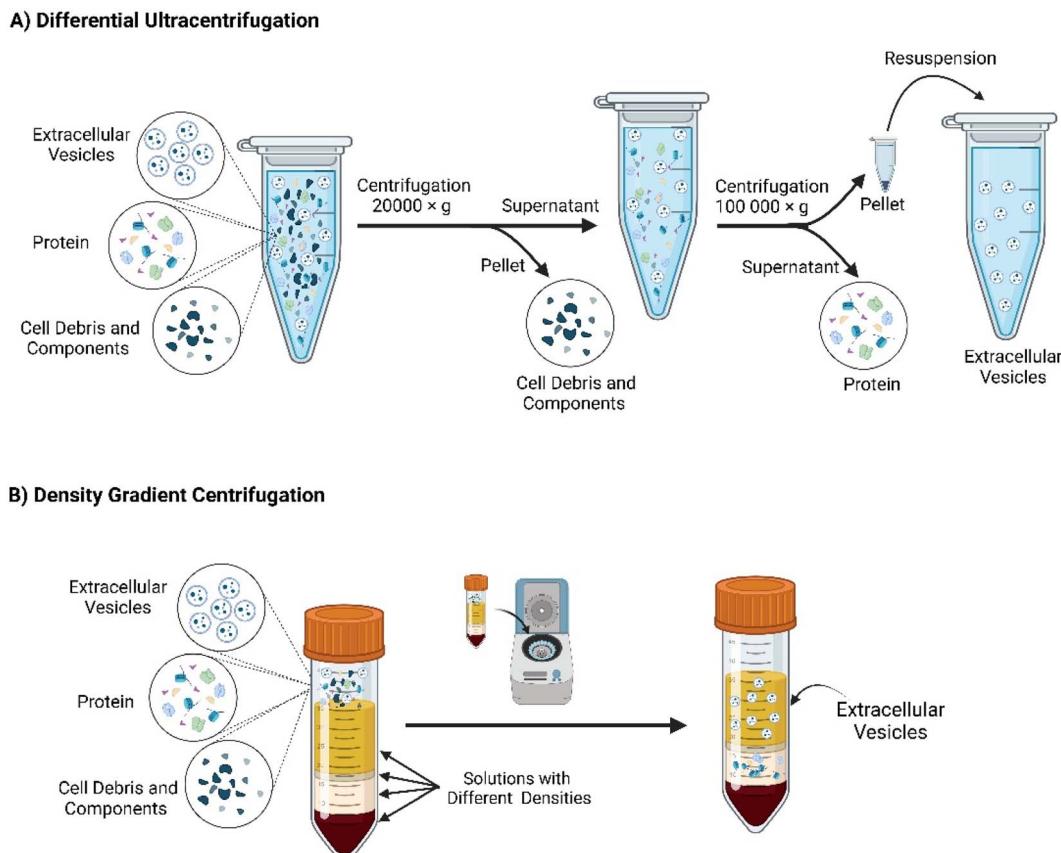


Fig. 2 Isolation of extracellular vesicles using density-based methods (A) differential ultracentrifugation in a two-step manner to eliminate cell debris and proteins, (B) density gradient centrifugation using pre-loaded centrifugal tubes.

scalable approach.<sup>99</sup> Sequential filtration using membranes with decreasing pore sizes can enhance EVs purity.<sup>92</sup> To separate tiny EVs from bigger EVs, a 100 nm track etch filter is employed in the third phase (Fig. 3),<sup>92</sup> and a filtration-based chip was also employed to increase the recovery rate.<sup>93</sup> The filtration-based techniques for EVs isolation are faster, more effective, automated, and exhibit superior recovery rates compared to ultracentrifugation. However, membrane clogging and potential EVs damage are still limitations of this technique as shown in Table 2.<sup>82</sup> Collectively, each isolation method exhibits distinct advantages and disadvantages as summarized in Table 2. The optimal choice of isolation method depends on the specific research question, desired EVs purity, and available resources. In many cases, a combination of techniques is employed to achieve optimal EVs isolation and characterization.

**Affinity-based and immunoaffinity approach.** Affinity-based and immunoaffinity capture methods leverage the specific recognition between target molecules and ligands to isolate EVs as previously reviewed in ref. 100. Antibodies, aptamers, and lipid-based probes are commonly employed to bind to specific surface antigens on EVs, enabling their capture and enrichment as briefly presented below.

**Enzyme-linked immunosorbent assay (ELISA).** ELISA is a commonly employed affinity-based method for EVs capture

and quantification.<sup>79</sup> This technique involves immobilizing antibodies specific to EVs surface antigens on a microplate. When an EVs sample is added, antigen–antibody interactions facilitate EVs capture onto the plate surface.<sup>94</sup> Subsequent steps involve washing to remove unbound components and the use of a labeled secondary antibody to detect and quantify the captured EVs (Fig. 4). While ELISA offers a relatively simple approach for EVs detection, it suffers from limitations. The technique is often restricted to small sample volumes and may compromise EV integrity due to the immobilization process, potentially affecting their biological activity.<sup>101</sup>

**Magneto-immunoprecipitation.** Magneto-immunocapture is an alternative affinity-based method for EVs isolation.<sup>97</sup> This approach involves the use of magnetic beads coated with antibodies specific to EVs surface antigens. Upon incubation with the sample, EVs bind to the antibody-coated beads, forming magnetic complexes.<sup>102</sup> These complexes can then be readily isolated using a magnetic field, allowing for efficient recovery of the target EVs (Fig. 4).

Compared to ELISA, magneto-immunocapture offers advantages in terms of sample volume and potential for higher capture efficiency due to the increased surface area of the magnetic beads as summarized in Table 2. However, similar to ELISA, this method relies on the specificity of the antibodies and may be susceptible to non-specific binding.<sup>103</sup>



Table 2 Isolation methods of extracellular vesicles (EVs)

Principle of separation	Isolation method	Assay principle	Advantages	Disadvantages	References
Density	Ultracentrifugation	Several centrifugation steps	Easy, very common	Costly, time-consuming, and less efficient with viscous body fluids	85
	Density gradient centrifugation	Pre-loaded centrifugal tubes, with different densities	Useful in isolating low-density EVs	Costly, time-consuming	88 and 89
Size	Size-exclusion chromatography	Chromatography columns packed with porous beads	Precise separation, maintain the structure of EVs	Time-consuming	87
	Polymer-based precipitation	Polymers decrease the solubility of EVs	Simple, easy, inexpensive, has mild effects on EVs	Co-precipitation of the polymer during separation	90 and 91
	Filtration	Ultrafiltration porous membranes	Eliminates small molecules, can sort out large and small EVs, is fast, more effective and has a higher recovery rate than centrifugation	EVs can clog the membrane, and releasing them jeopardizes their integrity	92 and 93
Immune-affinity	ELISA	Antibodies on microplates and EVs' antigens	Very specific, selective, can be used to identify the EVs	Not applicable for larger volumes, activity and function of EVs may be lost	94
	Magneto-immunoprecipitation	Antibodies on magnetic beads and EVs' antigens	Very specific, selective, has an increased surface area		
	Aptamer-based method	Oligonucleotide aptamers and EVs' proteins	Easier to use, aptamers cost less and have higher stability than antibodies	Less specific than antibodies	95
Others	Lipid-based probe method	Lipid probes and EVs' lipids	Can quickly isolate and retrieve highly pure EVs	Low specificity	82
	Electrophoretic isolation	Using electric field	Specific, quick	Inaccurate due to the presence of contaminants	96
	Microfluidics	Based on physical and biological features	Fast, efficient, small initial volumes	Low isolation capacity, needs high technical expertise	97

**Aptamer-based method.** Aptamers, synthetic oligonucleotides selected for their high affinity and specificity to target molecules, offer a promising alternative to antibody-based EVs isolation.<sup>104</sup> Unlike antibodies, aptamers possess several advantages, including ease of handling, low production costs, and superior stability as summarized in Table 2. By targeting specific protein markers on the EVs surface, aptamers enable selective capture and enrichment of EVs subpopulations (Fig. 4). However, the development of aptamers with high affinity and specificity for EVs antigens remains a challenge, limiting their widespread application.<sup>95</sup> Yet, despite this limitation, aptamer-based EVs isolation holds significant potential for advancing EVs research and clinical applications.

**Lipid-based probe method.** Exploiting the lipidic nature of EVs membranes, lipid-based probes offer a unique approach to EVs isolation.<sup>105</sup> These probes, designed to integrate into the EVs lipid bilayer, enable rapid and efficient EVs capture. Unlike antibody-based methods, which rely on specific surface

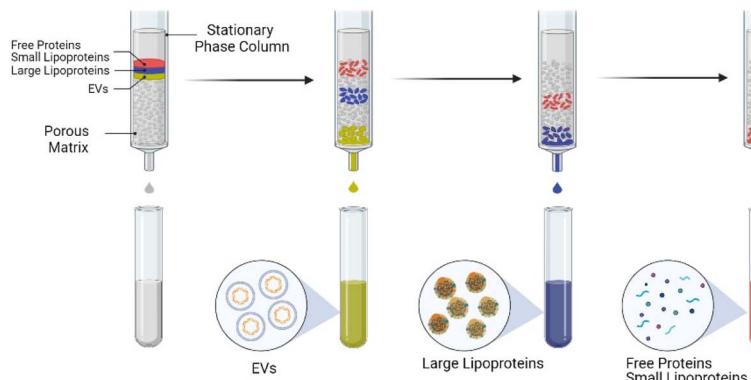
antigens, lipid probes offer a more universal approach, capable of capturing a broader range of EVs subpopulations (Fig. 4). This innovative technique provides several advantages, including high capture efficiency, speed, and the potential to preserve EVs integrity. However, the development of lipid probes with optimal selectivity and specificity remains an ongoing challenge as shown in Table 2. Collectively, it is important to note that no single isolation method is universally superior. The choice of technique often depends on the specific research question, desired EVs purity, and available resources.

#### Other methodologies

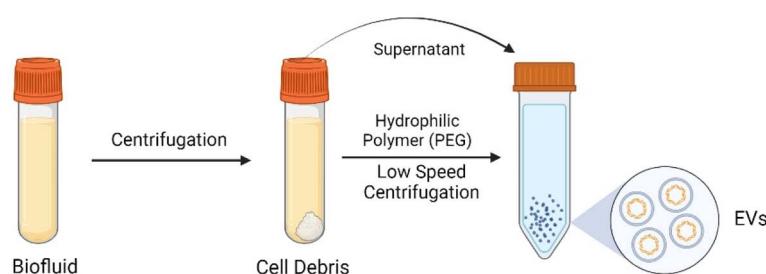
**Electrophoretic isolation.** Electrophoretic methods exploit the inherent charge of EVs to achieve their separation.<sup>106</sup> One approach involves the use of membrane filters, where an electric field is applied to expel proteins while retaining EVs.<sup>82,107</sup> However, this method suffers from limitations in terms of specificity and recovery as shown in Table 2.



### A) Size-Exclusion Chromatography (SEC)



### B) Polymer Precipitation



### C) Filtration Method

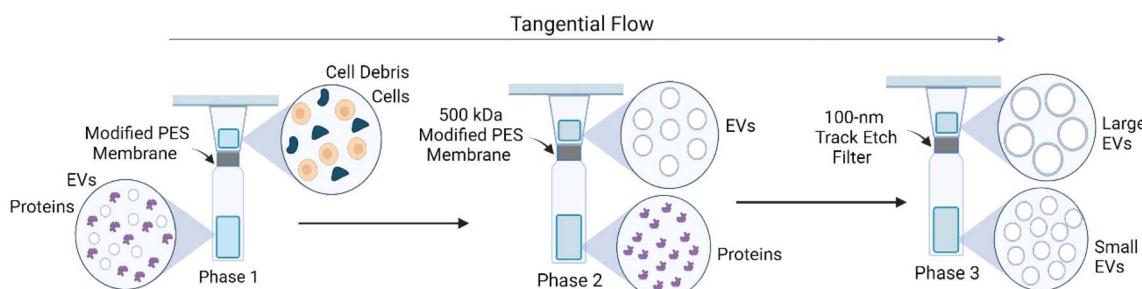


Fig. 3 Isolation of extracellular vesicles using size-based methods (A) size exclusion chromatography using columns packed with a porous matrix to retain smaller particles, (B) polymer precipitation where EVs are consequently precipitated at low-speed centrifugation, (C) filtration method employing several filtration membranes.

To address these challenges, tunable alternating current electrohydrodynamic (ac-EHD) technology has emerged. This method, also known as nanoshearing, utilizes a microfluidic system to generate high shear forces that disrupt EVs aggregates and enable their capture.<sup>82,96</sup> While promising, the presence of contaminants such as cell debris remains a hurdle for achieving high-purity EVs isolates (Table 2).

**Microfluidics.** Microfluidic platforms offer precise control over fluid flow and particle manipulation, making them attractive for EVs isolation.<sup>108</sup> These systems can incorporate various physical forces, including acoustic, electrophoretic, and

hydrodynamic forces, to separate EVs based on size, charge, and other properties as shown in Table 2. Microfluidic devices often demonstrate high efficiency and rapid processing times, but challenges related to sample loading, clogging, and integration with downstream analysis persist.<sup>109</sup>

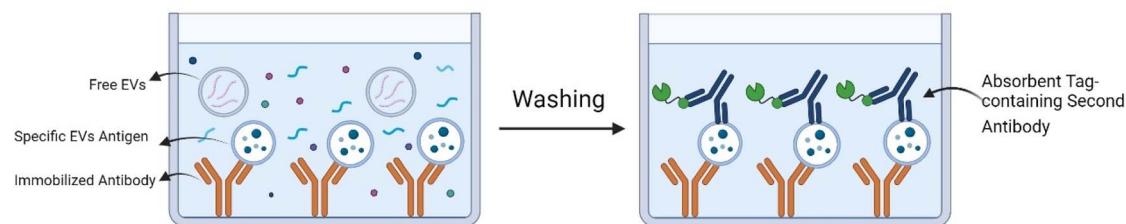
#### Cargo loading strategies

EVs serve as versatile carriers for a diverse array of therapeutic agents, including chemotherapeutics, nucleic acids (mRNA, miRNA, siRNA, snoRNA), and proteins.<sup>110</sup> Cargo loading strategies can be categorized into pre-isolation and post-isolation

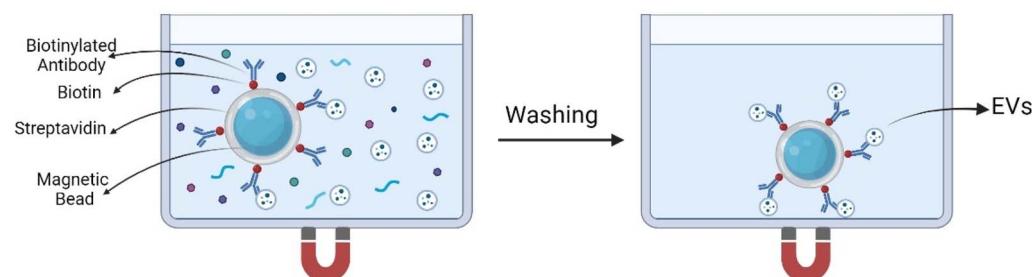




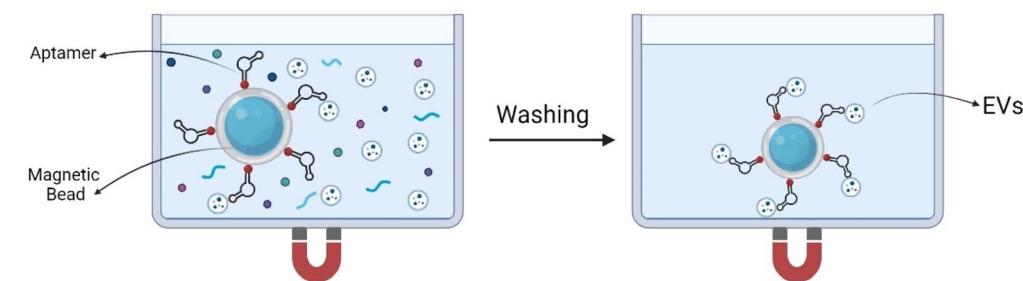
### A) Enzyme-Linked Immunosorbent Assay (ELISA)



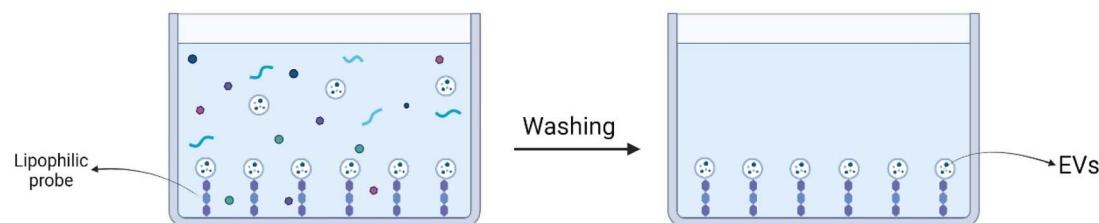
### B) Magneto-Immunoprecipitation



### C) Aptamer-Based Method



### D) Lipid-Based Probe Method



**Fig. 4** Affinity-based and immunoaffinity approach methods (A) enzyme-linked immunosorbent assay (ELISA) where antibodies are used to first capture antigen-expressing EVs then tag the immobilized EVs, (B) magneto-immunoprecipitation using biotinylated antibodies on streptavidin magnetic beads, (C) aptamer-based method to bind specific protein-expressing EVs, (D) lipid-based probe method exploiting EVs' lipid membrane.

approaches as shown in Table 3. Optimizing cargo loading strategies is crucial for developing effective EV-based therapeutics.<sup>118</sup> Cargo loading phase might be divided into two main approaches: pre-isolation loading and post-isolation loading. Pre-isolation loading is also sub-divided into several loading strategies such as endogenous loading during EVs biogenesis, cellular components, including drugs and biomolecules, can be naturally incorporated into the forming EVs.<sup>114</sup> On the other hand, transfection-based loading where donor cells are transfected with the desired cargo, allowing for its incorporation into newly formed EVs can also be another pre-loading strategy as previously reviewed in ref. 111 and summarized in Table 3.

While the post-isolation loading approach are sub-categorized into passive loading which includes hydrophobic drugs that can spontaneously integrate into the lipid bilayer of isolated EVs. However, this method is limited by cargo hydrophobicity and loading efficiency.<sup>112</sup> While the other category is known as active loading that enhance cargo loading, various techniques are employed, including electroporation, incubation, sonication, freeze-thaw cycles, and saponin-assisted incubation as shown in Table 3. These methods temporarily increase EVs membrane permeability, facilitating cargo entry.<sup>119</sup> Yet, the choice of loading method depends on the physico-chemical properties of the cargo, desired loading efficiency, and potential impact on EVs integrity.

### Modifications and engineering of EVs

Modification and engineering strategies for enhancing the distribution efficiency, targeting capability, and therapeutic efficacy of EVs involve various approaches. These include targeting ligands, stimuli-responsive elements, immune evasion properties as presented in Table 4, as well as hybridized membrane modifications and other modifications. These strategies have been shown to have promising applications not only in general EVs research but also in the specific context of cancer treatment.<sup>80,120</sup>

EVs can be modified through two primary methods: direct and indirect. The EVs themselves are modified in the direct method, while in the indirect method, modifications are made to the EV-producing cells.<sup>121</sup> It is important to note that no single optimum approach or standardized strategy exists for altering and modifying EVs. Each technique has its own benefits and applications as summarized in Table 4.<sup>121</sup>

## Characterization of the isolated EVs and their cargos

Following isolation, comprehensive characterization of EVs is essential to validate their identity, purity, and biological relevance.<sup>124</sup> A combination of techniques is often employed to assess various EVs properties, as discussed in the following section.

### Characterization of EVs

**Nanoparticle tracking analysis (NTA).** NTA is a widely adopted technique for characterizing EVs based on their brownian motion.<sup>125,126</sup> This method provides information on

Table 3 Cargo-loading strategies

Isolation strategy	Method	Nature of cargo	Advantages	Disadvantages	References
Pre-isolation method	Transfection of donor cells	Proteins, peptides, nucleic acids	High repeatability, stability, simple	Limited transfection efficiency, strong reliance on cell viability, potential toxicity, time-consuming	5 and 80
	Co-incubation	Drugs, nanoparticles	Very simple	Modest loading efficiency, applicable only for drugs that readily cross the plasma membrane, depends on parent cell type and drug concentration gradients	111
Post-isolation method	Electroporation	Nucleic acids, protein, peptides, drugs, nanomaterials	High loading efficiency	Require optimization, cause cargo aggregation	112
	Sonication	Drugs, proteins, nanoparticles	High loading efficiency	Cause aggregation of cargo and EVs, may damage EVs, not suitable for nucleic acids	113
	Incubation of EVs	Nucleic acids, protein, peptides, drugs, nanomaterials	Easy, practical	Low efficiency, applicable only for drugs that readily cross the plasma membrane	114
	Extrusion	Nucleic acids, protein, peptides, drugs, nanoparticles	High loading efficiency	Causes membrane integrity damage, makes EVs more cytotoxic	115
	Saponin as a surfactant	Proteins, peptides nanoparticles	High loading efficiency	Concentration must be regulated, requires further purification, and risk of hemolysis	116 and 117
	Freezing and thawing	Proteins, peptides	Simple, easy	Low efficiency of loading and low potency of the cargo, can cause vesicle degradation	116



Table 4 Engineering strategies of extracellular vesicles (EVs)

Engineering strategy	Type	Principle	Pros and cons	References
Direct method (EVs modification)	Lipid insertion	Lipids and lipid-tagged molecules are hydrophobically inserted by mixing and incubation	Quick, highly efficient, does not affect EV morphology or biological properties	116
	Chemical ligation (click chemistry)	Requires the presence of reactive groups in EV lipids/proteins that can interact with reactive fragment-tagged peptides	Reliable; change the characteristics of EVs, disrupt protein–protein interactions, costly	120
	Affinity binding	Molecules that have an affinity to EVs lipids/proteins are mixed and incubated with EVs	Does not affect the structure of the EV membrane; not as reliable as chemical or covalent bonding-based techniques	121
	Enzyme ligation	Enzymatic ligation between the targeted protein/peptide and the EV membrane protein	Produces permanent covalent modification without the need for chemical or genetic modification	122
	Hybridization	Combination of EVs with other lipid nanovesicles like liposomes	Improved colloidal stability, increased half-life, low immunogenicity; loss of biological functions of integral EVs	123
	Biomimetic EV production	Synthetic EV components that possess homing capabilities and cargo delivery effectiveness	Better pharmacokinetic and biocompatibility properties; require extra purification steps	
Indirect method (producing cell modification)	Genetic manipulation of the producing cells	Transgene production of proteins or chimeric proteins	Dual-targeting ability, low immunogenicity, low systemic toxicity; some peptides require protection against degradation	69
	Metabolic labeling	Cultivating donor cells in a medium containing saccharides or amino acids bearing reactive groups attaches them to EV membrane proteins	Reliable, effective; expensive	73 41 76

particle size distribution, concentration, and enables direct visualization of individual EVs in solution. NTA's ability to analyze particles within a size range of 10–1000 nm makes it particularly suitable for EVs characterization.<sup>125</sup>

Despite its advantages, NTA has certain limitations. The accuracy of size determination can be influenced by factors such as particle shape, refractive index, and the presence of contaminants.<sup>127</sup> Additionally, differentiating EVs from other nanoparticles in complex biological samples can be challenging, necessitating careful sample preparation and data analysis.<sup>127</sup> Accordingly, in an attempt to enhance the specificity of NTA, fluorescence-based approaches have been developed.<sup>128,129</sup> Labeling EVs with specific markers makes it possible to differentiate target EVs from non-EVs particles as shown in Table 5 and as recently reviewed in ref. 130. However, this method is limited by fluorophore bleaching and the availability of suitable antibodies or probes.<sup>129,131</sup> Overall, NTA is a valuable tool for EVs characterization but should be complemented with other techniques to comprehensively understand EVs properties.

**Dynamic light scattering (DLS).** DLS is another commonly used technique for EVs characterization.<sup>129,132</sup> By measuring the fluctuations in light scattered by particles in suspension, DLS can determine particle size distribution based on Brownian motion.<sup>129</sup> While DLS analysis is rapid and sensitive and can measure EVs (1 nm–6 µm) without needing pre-treatment, the results are only consistently accurate for monodisperse samples as shown in Table 5.<sup>133</sup> Moreover, DLS is susceptible to interference from larger particles and contaminants, which can affect the accuracy of size distribution measurements, especially in complex biological samples.<sup>133</sup> Therefore, the use of this approach for the investigation of varied heterogeneous EVs populations is limited as previously reviewed in<sup>134</sup>.

**Transmission electron microscopy (TEM).** TEM is the gold standard for visualizing individual EVs.<sup>135</sup> Its high resolution, capable of resolving particles smaller than 1 nm, allows for detailed morphological characterization of EVs size, shape, and internal structure.<sup>136</sup> When combined with immunogold labeling, TEM enables the localization of specific proteins or



Table 5 Comparison between nanoparticle tracking analysis (NTA) and dynamic light scattering (DLS)

Point of comparison	Nanoparticle tracking analysis (NTA)	Dynamic light scattering (DLS)
Principle	Characterizing EVs based on their brownian motion	
Type of detection	Size, concentration and distribution of EVs	Size of the EVs
Advantages	Ability to analyze particles within a size range of 10–1000 nm Can detect relatively low concentrations	Rapid Sensitive Can measure EVs (1 nm–6 $\mu$ m) without needing pre-treatment
Disadvantages	Limited size detection Differentiating EVs from other nanoparticles in complex biological samples can be challenging	Accurate size detection is limited to monodispersed samples Susceptible to interference from larger particles and contaminants

other biomolecules within EVs, providing insights into their composition and function.<sup>137</sup> However, TEM is associated with several limitations. The sample preparation process, which involves dehydration and embedding in resin, can introduce artifacts and distort EVs' true size and shape.<sup>138</sup> Additionally, the high vacuum environment required for imaging can lead to sample damage and the formation of cup-shaped structures, complicating accurate size measurements.<sup>139</sup> Despite these challenges, TEM remains an indispensable tool for obtaining high-resolution images of EVs, providing valuable information for their characterization and classification.<sup>70</sup>

**Atomic force microscopy (AFM).** Atomic force microscopy (AFM) offers high-resolution imaging of EVs at the nanoscale.<sup>129</sup> By scanning the sample surface with a sharp tip mounted on a cantilever, AFM generates detailed topographical images without the need for sample staining or fixation.<sup>140</sup> This technique enables precise measurement of EVs size and shape distribution, providing valuable insights into EVs heterogeneity.<sup>141</sup> Moreover, by functionalizing the AFM tip with specific antibodies or ligands, it is possible to differentiate EVs subpopulations based on surface markers.<sup>140,142</sup> AFM offers versatility through various imaging modes, including contact, tapping, non-contact, and peak force modes, allowing for tailored analysis of EVs properties.<sup>141,142</sup> However, factors such as tip condition, applied force, and environmental conditions can influence image quality and data accuracy.<sup>129,143</sup> To address these challenges, meticulous sample preparation and instrument calibration are essential for obtaining reliable AFM data. Yet, AFM remains a powerful tool for characterizing EVs morphology and physical properties.

**Resistive pulse sensing (RPS).** Resistive pulse sensing (RPS) uses the Coulter concept to assess the particles distribution and diameters in suspensions that roughly range between 50 and 100 000 nm.<sup>127,144</sup> The RPS used in the field of EVs characterization is often performed with qNano (Izon Science Ltd, Christchurch, New Zealand). Two fluid cells divided by a non-conductive membrane make up the qNano.<sup>125,145</sup> A single membrane pore is subjected to an electric current, and when particles flow through it, the signal experiences a transitory attenuation roughly proportionate to the volume of particles. The sample quantities used in the qNano can be as little as 10  $\mu$ L.<sup>125</sup> Nevertheless, there are numerous drawbacks to this method, such as: the need for various pore sizes; pores are

prone to blockage; minimal phenotypic information on the EV's origins is obtained; and it is impossible to distinguish EVs from pollutants of comparable sizes.<sup>125</sup>

**Flow cytometry (FCM).** FCM is another powerful technique for analyzing particle populations in suspension, including EVs.<sup>146</sup> By measuring light scatter and fluorescence signals from individual particles, FCM provides information about particle size, complexity, and surface markers.<sup>129</sup> Conventional FCM relies on forward scatter (FSC) to estimate particle size and side scatter (SSC) to assess particle complexity. However, the lower detection limit of conventional FCM is around 300 nm, limiting its ability to accurately characterize smaller EVs.<sup>129,147</sup> To overcome such limitation, labeling strategies, such as coupling EVs with larger fluorescent beads, have been employed to enhance detection sensitivity.<sup>148</sup> Additionally, nanoscale flow cytometry (nFCM) has emerged as a promising technique for analyzing particles as small as 100 nm.<sup>132,149</sup> By combining high-resolution optics with sensitive detectors, nFCM enables more accurate characterization of EVs size and surface markers, facilitating in-depth studies of EVs heterogeneity.<sup>129,150</sup> Despite these advancements, challenges remain in accurately differentiating EVs from other nanoparticles and achieving high-throughput analysis of large sample volumes. Further developments in flow cytometry instrumentation and sample preparation are necessary to fully realize the potential of this technique for EVs research.

**Enzyme-linked immunosorbent assay (ELISA).** ELISA is also a widely employed technique for quantifying and characterizing EVs.<sup>129,151</sup> By capturing EVs through specific antibody–antigen interactions on a microplate surface, ELISA detects and measures EV-associated proteins. This method offers several advantages, including high sensitivity, specificity, and the potential for high-throughput analysis. However, ELISA is susceptible to variations in assay conditions, leading to potential inconsistencies in results.<sup>151</sup>

**Zeta potential (ZP).** ZP is mainly concerned by the electrostatic repulsion among particles in colloidal solution.<sup>152</sup> ZP can be affected by the surface charge, which can be assessed from the electrophoretic mobility in the colloidal system.<sup>153</sup> The net surface charge of EVs, as shown by the ZP, is crucial in determining the stability of EVs.<sup>153,154</sup> Higher absolute ZP values suggest that the colloid, containing EVs, is more stable.<sup>155</sup> ZP can



be conjugated with other characterization techniques including DLS (like zetasizer), NTA (like zetaview or Z-NTA), RPS, or by on-chip microcapillary electrophoresis combined with microscopy technique for visualization and identification of EVs.<sup>152,156,157</sup>

### Characterization of EV content

**EVs protein content characterization.** Proteins found in EVs might give information about biological activities and how they affect cell communication. As a result, several studies characterized EVs from the perspective of their protein content.<sup>124</sup>

Two simple approaches are employed to quantify the total protein level, which are the Coomassie Brilliant Blue G-250 test and the bicinchoninic acid assay.<sup>48</sup> Both rely on measuring the absorbance of a colored complex formed between a protein and a reagent, using a calibration curve with known protein concentrations to quantify the protein.<sup>158</sup> While these tests are often employed, their applicability is restricted to assessing extremely pure EVs samples since protein impurities impair measurement accuracy.<sup>159</sup>

Western blot is another technique used for detecting, quantifying, and characterizing EVs protein, giving insight into EVs biology and discovering pathophysiological indicators of illnesses.<sup>160,161</sup> Nevertheless, EVs are diverse, thus it is challenging to provide a universal EV protein identifier or marker.<sup>162</sup> As a result, the International Society for EVs suggests characterizing various transmembrane and cytosolic proteins found in EVs,<sup>163</sup> following a set of guidelines (Minimal Information for Studies of EVs; MISEV2018) that should be followed for isolation, characterization, and functional studies of EVs. In this regards, there should be at least one cytosolic protein (such as TSG101 (tumour susceptibility gene 10), ALIX (ALG-2-interacting protein X), and syntenin) and one transmembrane protein (such as CD9, CD63, and CD81).<sup>48</sup> This approach has been used to detect and characterize proteins of EVs produced by tumors. For instance, Yoshioka *et al.* observed that regardless of the EV origin, all examined EVs recovered from four human prostate cell lines were positive for CD9 and CD81, with equal quantity. Conversely, the detection of other EV marker proteins, such as TSG101 and CD63, was uneven due to the heterogeneity of the cancer.<sup>164</sup>

Moreover, surface plasmon resonance (SPR) is another promising technique for detecting proteins in various EVs.<sup>165</sup> SPR allows for the extremely sensitive label-free detection of EVs by their immunological capture to a SPR-active surface, such as silver or gold nanoparticles.<sup>166</sup> Several of these methods have recently been used to quantify and characterize EVs produced from diseases, including cancer, using particular protein markers. For instance, it has been demonstrated that gold nanoparticles stabilized with DNA aptamers against particular surface proteins cause a noticeable color shift because EVs attach to these aptamers specifically.<sup>167</sup> This method enables a multiplexed analysis of EVs' protein composition using visual and spectrophotometric methods.<sup>167</sup>

The field of EVs has significantly benefited from mass spectrometry proteomics technologies, which have made it possible to create extensive protein profiles of EVs.<sup>124,168</sup>

Proteomics has been utilized in a number of studies to measure the presence of specific peptides and find changes in the EVs of biological samples from cancer patients compared to healthy people.<sup>169</sup> Notably, one study has shown that EVs generated from the serum of breast cancer patients have different protein profiles according to the cell line they derived from and so can be used to distinguish between the molecular subtypes of breast cancer, such as triple-negative or HER2 subtypes.<sup>170</sup>

Fluorescence-based techniques offer a versatile approach for detecting and quantifying EVs.<sup>171</sup> Fluorescently labeled antibodies targeting EV surface markers, such as CD9, CD63, and CD81, can be used to visualize and track EVs.<sup>148</sup> For instance, quenching and relabeling techniques have been employed to identify unique EVs marker clusters.<sup>172</sup> Integrated microfluidic-based approaches have also emerged as powerful tools for EVs isolation and quantification. ExoDEP chips, for example, utilize antibody-functionalized beads to capture EVs, followed by electrochemical detection.<sup>173</sup> Thermophoresis-assisted fluorescence detection is another promising method that enables the isolation and quantification of EVs based on size and specific markers.<sup>174,175</sup> The authors developed a novel approach called HOLMES-ExoPD-L1, which combines tumor-associated PD-L1 aptamers with thermophoresis to detect circulating PD-L1-positive EVs with high sensitivity and specificity, achieving an impressive limit of detection of 17.6 pg ml<sup>-1</sup>.<sup>174</sup>

**EVs RNA content characterization.** The presence of RNA (mRNAs, miRNAs, long noncoding RNAs and/or others) inside EVs is an essential tool in confirming the successful synthesis/isolation of EVs.<sup>176,177</sup> Their identification and characterization are crucial because they can provide information about the biological activities of EVs, their cell origin, and how they alter cell communication in both normal and pathological conditions.<sup>124</sup>

One of the widely used characterization assays for EVs' RNA profile is quantitative reverse transcriptase-PCR (qRT-PCR).<sup>178,179</sup> Several studies employed qRT-PCR to quantify and characterize the RNA content of EVs in malignancies, including pancreatic cancer,<sup>180</sup> breast cancer,<sup>181</sup> endometrial cancer,<sup>182</sup> and many other cancers.<sup>183</sup> However, it has a major limitation: the low RNA yield, particularly from clinical samples, which may restrict the identification and analysis.<sup>184</sup>

Digital droplet PCR (ddPCR) is a relatively recent technique that enables the absolute measurement of gene expression. This technology enables the susceptible measurement of RNA expression levels and DNA variations without standard curves.<sup>125</sup> Recently, ddPCR was used to analyze plasma-derived exosomal RNA from prostate cancer patients,<sup>185</sup> and EVs produced from blood and CSF of glioma patients.<sup>186</sup>

Furthermore, next-generation sequencing (NGS) has been used to characterize the RNA content of EVs from various sources, allowing for a more complete examination of the EV-RNA repertoire.<sup>187,188</sup> Many research have employed NGS to characterize EVs' RNA content in malignancies including bladder cancer,<sup>189</sup> ovarian cancer,<sup>190</sup> colorectal cancer,<sup>191</sup> pancreatic cancer,<sup>192</sup> and many malignancies. However, it



suffers from some limitations, including, library preparation concerns and adapter dimers that could hinder the analysis.<sup>187</sup>

**EVs lipid content characterization.** Lipids are an essential component of EVs as they contain markers from their original cell and act as a protective barrier for their load.<sup>193</sup> They are also involved in the transportation of biomolecules and membrane fusion events.<sup>193</sup> Mass spectrometry-based lipidomics approaches are among the most widely utilized methods for characterizing and quantifying the lipid composition of EVs. Moreover, these methodologies provide information regarding the lipid profile of EVs from different malignancies.<sup>194,195</sup>

For instance, using an MS-based lipomic approach, it was discovered that EVs derived from high metastatic breast cancer had a different lipid profile than EVs from low metastatic breast cancer.<sup>196</sup> It was also discovered that the EVs from high metastatic breast cancer had more unsaturated diacylglycerols (DGs) than those from low metastatic breast cancer, which means they have a greater capacity to promote angiogenesis.<sup>196</sup> Another study discovered that whereas sterol lipids, sphingolipids, and glycerophospholipids were more highly abundant in EVs from tumorigenic and metastatic prostate cancer cells, fatty acids, glycerolipids, and prenol lipids were more highly abundant in EVs from non-tumourigenic prostate cancer cells.<sup>197</sup>

**EVs metabolic content characterization.** Metabolites, such as steroid hormones, amino acids, or metabolic intermediates of lipid and nutrients, are a class of tiny molecules that result from different biological events.<sup>198</sup> Two primary analytical methods are used to characterize metabolites in EVs: nuclear magnetic resonance (NMR) spectroscopy and high-resolution mass spectrometry.<sup>199,200</sup> Although on the scarcity of information regarding EVs' metabolome, several studies highlighted the relevance of EVs as carriers of key metabolome fingerprints that may be utilized to define particular changes in cellular homeostasis that occur in both physiological and pathological conditions.<sup>201,202</sup> For instance, Čuperlović-Culf *et al.* (2020) studied the metabolome of sEVs generated from several glioblastoma cells using NMR spectroscopy.<sup>203</sup> When comparing EVs from different glioblastoma subtypes, there was a noticeable variation in their metabolic characteristics.<sup>203</sup> Another study that used MS-based methods found that the urine EVs from prostate cancer and benign prostatic hyperplasia had different metabolic profiles, with around 76 compounds different.<sup>199</sup> It was also detected that the steroid hormone, 3 beta-hydroxyandros-5-en-17-one-3-sulphate is higher in prostate cancer than in benign prostate hyperplasia so urine EVs can be used as a non-invasive biomarker for prostate cancer.<sup>199</sup> Furthermore, another study examined the metabolomic profile of EVs produced from pancreatic cancer cells (PANC-1) cultivated at various oxygen concentrations, as hypoxia contributes to the malignant activity of these cells.<sup>200</sup> This study also found that the metabolite composition of EVs differed depending on the cell of origin. A total of 140 hydrophilic metabolites were discovered in small EVs, and it was revealed that the metabolomic profile of small EVs altered during hypoxic stress, with an increase in the metabolites implicated in angiogenesis, growth, and metastasis of cancer<sup>200</sup>

## EVs: a novel frontier in cancer therapeutics

Cancer remains a formidable global health challenge, with conventional treatments often yielding suboptimal outcomes.<sup>20,64,65</sup> The emergence of EVs as versatile nanoparticulate carriers has sparked significant interest in their potential as therapeutic agents.<sup>70</sup> Unlike traditional drug delivery systems, EVs possess intrinsic advantages such as biocompatibility, low immunogenicity, and the ability to cross biological barriers.<sup>79</sup>

EVs can be harnessed for delivering a wide range of therapeutic payloads, including chemotherapeutics,<sup>68</sup> nucleic acids,<sup>81,204</sup> and immunomodulatory molecules.<sup>61</sup> Their capacity to target specific tissues and cells, coupled with their ability to evade immune clearance, positions EVs as promising candidates for overcoming challenges associated with conventional drug delivery systems.<sup>120,121</sup>

To fully realize the potential of EVs in cancer therapeutics, meticulous engineering and optimization are imperative. Researchers can develop highly effective EV-based therapeutics by carefully selecting cargo molecules, modifying EVs surface properties, and understanding the complex interactions within the tumor microenvironment. This review highlights the significant potential of EVs as a platform for delivering cancer treatments and emphasizes the need for continued research to overcome existing challenges and translate these promising findings into clinical applications.

### EVs as drug delivery vehicles for chemotherapeutic agents

Conventional chemotherapy regimens often exhibit limited efficacy due to systemic toxicity and drug resistance.<sup>63,66</sup> EVs have emerged as promising platforms for delivering chemotherapeutic agents to cancer cells, addressing these limitations.<sup>205</sup> By encapsulating chemotherapeutic drugs within their lipid bilayer, EVs can enhance drug delivery vehicles to tumor sites, prolong drug circulation time, and reduce off-target effects.<sup>120</sup>

Several studies have demonstrated the potential of EV-based chemotherapy.<sup>206</sup> For instance, LipHA-modified EVs loaded with doxorubicin (DOX) have shown enhanced efficacy in overcoming drug resistance in breast cancer cells by inhibiting P-glycoprotein.<sup>120,207</sup> Similarly, CC8-modified EV-like vesicles carrying Imperialine have demonstrated anti-tumor activity in non-small cell lung cancer (NSCLC).<sup>208</sup> Furthermore, a research study demonstrated that exosomes that are produced from human fibrosarcoma cells are able to carry DOX, and have the potential to target fibrosarcoma efficiently, increasing therapeutic retention and inhibiting cancer growth (Fig. 5).<sup>120,209</sup> It has also been reported that mimic or chimeric exosomes derived from red blood cells (RBCs) carrying DOX can enhance drug accumulation, decrease drug clearance, and prevent or reduce the growth of breast cancer cells.<sup>120,210</sup>

Moreover, EVs derived from tumor cells or healthy cells, such as red blood cells, have been employed to deliver chemotherapeutic agents like methotrexate, cisplatin, and paclitaxel,



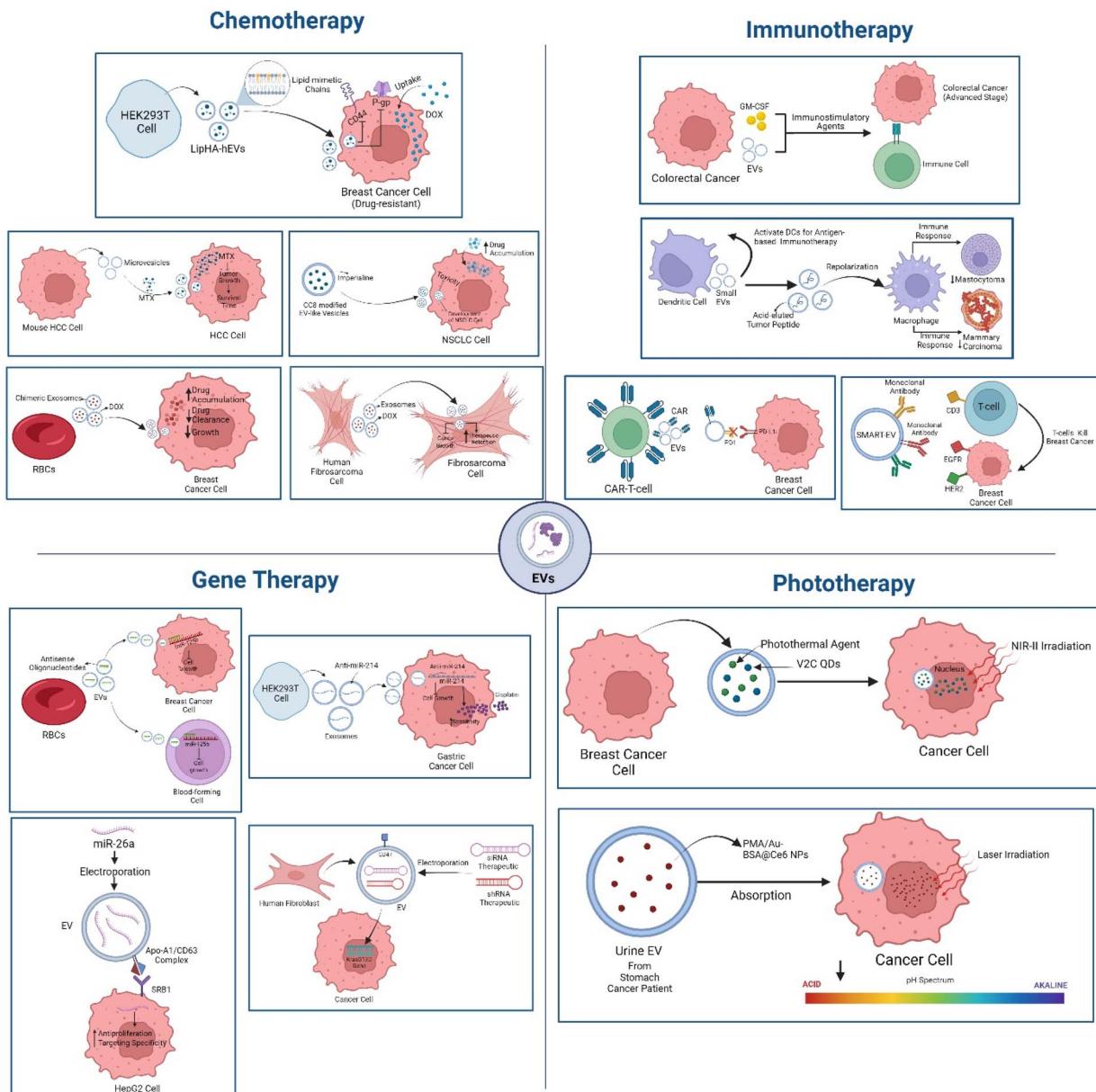


Fig. 5 Multifunctional role of extracellular vesicles in cancer therapeutics.

resulting in improved therapeutic outcomes<sup>211–213</sup> (Fig. 5 and Table 6). Collectively, these findings highlight the versatility of EVs as drug-delivery vehicles and their potential to improve cancer treatment by overcoming challenges associated with conventional chemotherapy.

#### EVs as drug delivery vehicles for gene therapy

EVs have emerged as promising platforms for delivering nucleic acid-based therapeutics, such as siRNA, shRNA, and miRNA, to target specific genes involved in cancer progression.<sup>214,215</sup> By encapsulating these therapeutic molecules within EVs, it is possible to overcome challenges associated with traditional gene delivery methods, such as limited cellular uptake and rapid degradation.<sup>216,217</sup>

Several studies have demonstrated the efficacy of EV-based gene therapy for cancer treatment. For example, EVs loaded with anti-miR-214 have shown anti-tumor effects in gastric cancer models.<sup>218</sup> A notable example is the use of RBC-derived EVs loaded with antisense oligonucleotides targeting miR-125b, which demonstrated anti-tumor effects in breast cancer and acute myeloid leukemia without inducing systemic toxicity<sup>219</sup> (Fig. 5). In parallel, EVs decorated with the Apo-A1/CD63 complex, a known target of scavenger receptor class B type 1, were electroporated directly with miR-26a, which is downregulated in liver cancer. When applied to HepG2 liver cancer cells, the miR-26a enriched EVs had a strong anti-proliferative impact and excellent targeting specificity (Fig. 5).<sup>220,221</sup>





Table 6 Overview of EV-based cancer therapeutics

Methods	Source	EVs type	Drug/Cargo	Treated tumor	Outcomes	Ref.
Monotherapy/targeted/combined chemotherapy	Mouse HCC cells RBCs	Microvesicles Mimics exosomes	MTX DOX	HCC Breast cancer	Inhibited tumor growth without side effects ↑ Drug accumulation, ↓ clearance of drug, inhibited cancer growth	120, 121
	Hek293t Cells	Lipid-modified EVs	DOX	MDR breast cancer	↑ Drug accumulation and drug sensitivity, prevented cancer growth	268
	Plasma	CC8 modified EV-like vesicles	Imperialine	NSCLC	↑ Drug accumulation, inhibited tumor growth with reduced systemic toxicity	269
	Mouse macrophages	Exosomes	PTX	Lung metastatic cancer	↑ Cytotoxicity, prevented lung metastasis	209
	Mouse macrophages	Exosomes mimetics	DM4	Lung metastatic breast cancer	Prevented lung cancer metastasis	210
	—	Folate-engineered microvesicles	Bcl-2 siRNA and paclitaxel	Breast cancer squamous cell carcinoma	Significantly ↑ synergistic antitumor efficacy of chemotherapy and gene therapy	235
	Hek293t cells	HER2-binding antibody, LAMP2, and GFP modified exosomes	FU and mir-21 inhibitor oligonucleotide	Colon cancer	Improved chemosensitivity and inhibited growth of colon cancer	236
	Human macrophages	RAG-modified exosomes	DOX and gold nanorods	Cervical cancer	Promote drug release and inhibit cervical cancer growth	237
	Colorectal cancer cells	EVs	Granulocyte-macrophage colony-stimulating factor	Colorectal cancer	Utilized as immunostimulatory agents to treat patients with advanced CRC	120
	Mature dendritic cells	EVs	Loaded with acid-eluted tumour peptide	Mastocytoma and mammary carcinoma	Stimulate DCs for tumor antigen-based cancer immunotherapy, prevention of tumor growth	121
Monotherapy/targeted/combined immuno-therapy	CAR-T-cell	CAR-T-cell-derived EVs with surface-expressed CAR	—	Breast cancer	Directly target tumor cells, safe, do not express programmed cell death protein 1 (PD1)	210
	Expi293F cells	Exosomes modified with CD3 and EGFR antibodies	CD3 and EGFR antibodies	Breast cancer	Exhibit strong anti-cancer immunity against EGFR-positive breast cancer cells	221
	Expi293F cells	Exosomes modified with CD3 and HER2 antibodies	CD3 and HER2 antibodies	Breast cancer	Redirected the T cells to kill HER2-positive breast cancer cells	230
	Tumor cells	Irradiated tumor cell-released microvesicles	—	Malignant pleural effusion	Repopolarized tumor-associated macrophages, resulted in immunogenic death	270
	Mouse melanoma cells	Exosomes modified with cpg DNA	SAV-LA fusion protein	Melanoma	Prevented tumor growth	231
	BM-MSCs (bone marrow mesenchymal stem cells)	Exosomes modified with oxaliplatin	Galectin-9 siRNAs and surface modified with oxaliplatin	Pancreatic ductal adenocarcinoma (PDAC)	Improved tumor targeting and enhanced drug accumulation in cancer site	233
	CT26 cells	Hybrid-modified exosomes fused with liposomes	ICG, immune adjuvant R837, overexpressed CD47	Colorectal cancer	Reduced tumors in tumor-bearing mice	232
	NK cells	Exosomes	Mirna, such as let 7a	Breast cancer and NB cells	Target tumors effectively and prevent their growth	238, 239, 240

Table 6 (Contd.)

Methods	Source	EVs type	Drug/Cargo	Treated tumor	Outcomes	Ref.
Monotherapy/targeted/combined gene-therapy	RBCs	EVs	miR-125b ASOs	Breast cancer and acute myelocytic leukemia	Inhibit tumor growth without observable cytotoxicity	120
HEK293T cells	Exosomes		Anti-mir-214	Cisplatin-resistant gastric cancer	Enhanced chemosensitivity and prevented tumor growth	221
HEK293T cells	SMA or EGFR aptamer, folate modified EV	Survivin siRNA		Prostate, breast, and colorectal cancer	Prevented tumor growth	218
Breast cancer	EVs	miR-134		Breast cancer	Reduced migration and invasion of cancer cells, and enhanced sensitivity to anti-Hsp90 drugs	220
HEK 293T cells	EVs decorated with the Apo-A1/CD63 EVs enriched in CD47	miR-26a		Liver cancer	It has a strong antiproliferative impact and great targeting specificity	271
Human fibroblast		siRNA or shRNA against KrasG12D		Pancreatic cancer cell	Suppressed tumor development and ↑ survival rate	241
Brain metastatic cancer	Apoptotic bodies (sabs)	Anti-TNF- $\alpha$ antisense oligonucleotide (ASO) paired with cationic konjac glucomannan (ckgm)	—		Extremely high brain-delivery efficiency due to the CD44v6 expressed on the apoptotic bodies helping them cross the BBB	
Mouse M1 macrophages	QDs modified exosomes	DOX and miR21-responded hairpin DNA	Breast cancer	Inhibited growth of the cancer		
Urine from gastric cancer patients	Exosomes	PMA/Au-BSA@@Ce6 nanovehicle	Gastric cancer	Enhanced penetration and retention, prevented growth	120	
Breast cancer cells	RGD-modified exosomes	TAT peptide-modified V2C QDs	Breast cancer	Penetrate the nucleus and perform low-temperature PTI with increased antitumor activity	226	
Mouse macrophages	NRP-1 targeted peptide-modified exosomes	3-Cureumun and SPIONs	Orthotopic glioma	Give decent results for imaging and therapy, penetrate the BBB, and prevent tumor growth	227	
Mouse HCC cells	Mps (microvesicles)	DOX and Bi2Se3 nanodots	HCC	Prevented and inhibited cancer growth	239	
Blood	Exosomes with chimeric peptides produced	Photosensitizers and nuclear translocation peptide	Breast cancer	Destroyed the membrane and the nucleus of breast cancer cells in mouse models	272	

It is also worth noting that EVs express CD47, a 'don't eat me' signal that protects them from immune clearance.<sup>222</sup> This immune evasion property, combined with their ability to deliver therapeutic cargos, makes EVs attractive for cancer therapy. Studies have shown that CD47-enriched EVs can efficiently deliver siRNA or shRNA targeting oncogenic KrasG12D, leading to tumor growth inhibition and improved survival in preclinical models.<sup>223</sup> This highlights the potential of EVs to overcome immune barriers and deliver therapeutic payloads directly to cancer cells. Collectively, these findings highlight the potential of EVs to deliver nucleic acid-based therapeutics with high specificity and efficacy, offering a promising approach for treating various types of cancer as shown in Table 6.

### EVs as promising platforms for phototherapy

EVs have shown promise as carriers for phototherapeutic agents.<sup>224</sup> By encapsulating photosensitizers or photothermal agents within their lipid bilayer, EVs can deliver these compounds to tumor sites, enhancing their therapeutic efficacy.<sup>225</sup> For instance, EVs loaded with vanadium carbide quantum dots have demonstrated effective low-temperature photothermal therapy,<sup>226</sup> while EVs carrying multifunctionalized nanoparticles have shown improved tumor accumulation and ROS generation upon laser irradiation<sup>225,226</sup> (Fig. 5). These approaches leverage the unique properties of EVs, such as biocompatibility and prolonged circulation time, to enhance the therapeutic potential of phototherapy as presented in Table 6.

In another study, high-purity urine EVs were collected from stomach cancer patients and electroporated with multifunctionalized PMA/Au-BSA@Ce6 NPs. Due to the decreased macrophage endocytosis and longer blood retention time, the modified nanovehicles are effectively absorbed into cancer cells. The designed nanovehicles are shattered and release enormous NPs within in response to laser irradiation and acidic conditions (Fig. 5). Consequently, a significant amount of singlet oxygen is released, thereby limiting tumor cell proliferation.<sup>227</sup>

### Novel EV-based immunotherapeutic strategies

Immunotherapy has shown to be a viable cancer treatment option, frequently exhibiting promising results when utilized in conjunction with conventional approaches such as radiation, chemotherapy, or surgery.<sup>16,228,229</sup> A growing number of studies have shown that EVs impact immunological processes, including immune response activation, antigen presentation modification, and tumor microenvironment modulation.<sup>221,230</sup>

Studies have demonstrated the potential of EVs derived from immune cells, such as dendritic cells (DCs), to stimulate anti-tumor immunity.<sup>221</sup> For instance, GM-CSF-loaded EVs derived from colorectal cancer (CRC) cells have shown efficacy in treating advanced CRC patients.<sup>171</sup>

In addition to utilizing immune cell-derived EVs, engineering EVs to express specific targeting and effector functions has been explored. CAR-T cell-derived EVs, expressing chimeric antigen receptors (CARs), have demonstrated the ability to

directly target and kill tumor cells while evading immune checkpoint inhibition (Fig. 5).<sup>120,231</sup> Similarly, synthetic antibody-targeted EVs (SMART-EVs) have been developed to redirect T cells towards cancer cells expressing specific antigens, such as EGFR and HER2.<sup>120,232</sup> Nonetheless, by genetically expressing two different types of antibodies on the EV membrane, monoclonal antibodies specific for T cell CD3 and cancer cell-associated EGFR, synthetic antibody-targeted EVs (SMART-EVs) were created and exhibited strong anti-cancer immunity against EGFR-positive breast cancer cells.<sup>120,233</sup>

These studies highlight the versatility of EVs for immunotherapy and their potential to overcome challenges associated with traditional immune-based therapies. Table 6 provides a comprehensive overview of different EV-based immunotherapy strategies and their corresponding outcomes.

### EVs in combination therapies: synergistic approaches

Combining EVs with other therapeutic modalities holds the potential to enhance treatment efficacy and overcome limitations associated with single-agent therapies.<sup>234</sup> This approach, often referred to as combination therapy, aims to achieve synergistic effects and reduce the risk of drug resistance. By combining chemotherapy, immunotherapy, gene therapy, and phototherapy, EVs has emerged as promising platforms for delivering multiple therapeutic payloads and thus producing synergistic effects, enhance treatment efficacy, and improve patient outcomes.

### EVs in chemotherapy-related combined therapies

EVs loaded with chemotherapeutic agents can be combined with other therapeutic cargos, such as siRNA or miRNA, to create multifunctional platforms.<sup>121</sup> For example, folate-engineered microvesicles loaded with Bcl-2 siRNA and paclitaxel demonstrated enhanced anti-tumor efficacy compared to single-agent therapies by targeting both apoptotic pathways and chemotherapy resistance.<sup>235</sup> Similarly, HER2-binding affibody-modified exosomes carrying 5-FU and miR-21 inhibitors have shown promising results in inhibiting colon cancer growth.<sup>120,236</sup>

Additionally, chemotherapy can be combined with phototherapy, such as RAG modified exosomes produced from human macrophages. These exosomes were loaded with DOX and gold nanorods as a source of hyperthermia. NIR irradiation was utilized to influence the permeability of the EVs membrane, promoting medication release, and limiting tumor recurrence in a controlled manner. This combination therapy inhibited cervical cancer growth<sup>120,237</sup> as shown in Table 6.

### EVs in immunotherapy-related combination therapy

EVs can be effectively combined with other therapeutic modalities to enhance anti-tumor responses and overcome treatment resistance.<sup>121</sup> For example, EVs loaded with galectin-9 siRNA and conjugated with oxaliplatin have demonstrated synergistic effects in treating pancreatic ductal adenocarcinoma (PDAC) by combining immunotherapy and chemotherapy.<sup>121,238</sup> This approach promotes tumor-suppressive macrophage polarization,



recruits cytotoxic T lymphocytes, and induces immunogenic cell death (ICD), leading to improved tumor control.

Similarly, combining phototherapy (PDT or PTT) with EV-based therapies can enhance anti-tumor effects by inducing ICD and creating an immunogenic tumor microenvironment.<sup>8</sup> Additionally, hybrid nanovesicles can be produced by mixing and fusing liposomes loaded with the photothermal agent ICG and immune adjuvant R837, along with exosomes with overexpressed CD47. This hybrid nanovesicle effectively reduced tumors in tumor-bearing mice by competitively connecting with SIRP alpha, prior to tumor cells, resulting in increased tumor cell phagocytosis by macrophages and to avoid immune clearance of the hybrid. They are also considered as an example of combining cancer immunotherapy with PTT.<sup>62,121,239</sup>

Furthermore, EVs can be used to deliver gene therapy cargos in combination with other therapeutic modalities as summarized in Table 6. Moreover, in an immunogenic multi-modality approach, self-assembly of biomimetic core–shell NPs featuring a dendrimer core loaded with therapeutic miRNA, such as let-7a, a hydrophilic shell of NK-cell-derived EVs, showed highly effective targeting and therapeutic miRNA delivery to both neuroblastoma cells and breast cancer cells. This approach resulted in dual tumor growth inhibition effects.<sup>120,240</sup> These findings highlight the potential of EVs to serve as versatile platforms for developing combination therapies with enhanced efficacy and reduced side effects.

### EVs in gene therapy-related and phototherapy-related combination therapies

As shown in Table 6, it has been repeatedly reported that immunomodulatory EVs or photo-sensitive EVs are usually combined with immunotherapeutic and chemotherapeutic agents.<sup>9,10</sup> It has been reported that the utilization of small apoptotic bodies (sABs) derived from brain metastatic cancer cells, loaded with anti-TNF-alpha antisense oligonucleotide and paired with cationic konjac glucomannan (cKGM), exhibits remarkable brain delivery efficiency. This is attributed to the presence of CD44v6 expressed on the apoptotic bodies, facilitating their crossing of the blood–brain barrier.<sup>241</sup> Another study, which employed phototherapy combined with gene therapy approach, showed that exosomes with chimeric peptide produced from blood loaded with photosensitizers and nuclear translocation peptide destroyed the membrane and the nucleus of breast cancer cells in a mouse model.<sup>242</sup> Collectively, these strategies elaborated in Table 6 demonstrate the versatility of EVs as platforms for developing complex therapeutic approaches. However, further research is needed to optimize EVs production, cargo loading, and delivery for clinical translation.

## Challenges and obstacles with EVs-based cancer treatment

Despite the promising potential of EVs as therapeutic carriers, several challenges hinder their clinical translation.<sup>243</sup> Ensuring consistent EVs quality and composition for reliable therapeutic

outcomes has been considered one of the formidable barriers for the clinical translation of EVs-based onco-therapeutics.

### EVs heterogeneity

A significant hurdle in the development of EV-based therapeutics is the inherent heterogeneity of EVs populations.<sup>244,245</sup> EVs derived from the same cell type can exhibit variations in size, composition, and biological functions due to factors such as cellular origin, activation state, and extracellular environment.<sup>243,244</sup>

For instance, miRNAs, a common cargo of EVs as highlighted, exhibit differential distribution among EVs subpopulations.<sup>246</sup> This heterogeneity is attributed to both passive and active loading mechanisms, with some miRNAs being enriched in specific EVs subtypes.<sup>247,248</sup> According to earlier research, miRNAs in EVs are loaded by two mechanisms into the EVs: (a) highly expressed cellular miRNAs that enter the EVs passively by an osmotic-like effect; or (b) selectively released miRNAs that actively pack into EVs according to the particular RNA molecule sequence.<sup>243,249,250</sup> For instance, Pigati *et al.* discovered that, dependent on the amount of cytoplasmic miRNA, around 66% of the released miRNAs are passively secreted by EVs, whereas 30% of exosomal miRNAs do not match the cellular profile, indicating that they are released selectively.<sup>251</sup>

Similarly, protein composition can vary significantly among EVs subpopulations,<sup>252–254</sup> as demonstrated by the identification of distinct protein profiles in low-density and high-density EVs derived from the same cell line (B16F10 melanoma cells). Accordingly, the authors have classified LD (low density)-Exo and HD (high density)-Exo.<sup>254</sup> They found that both EVs contain the same proteins such as TSG101 and Alix. In addition to distinct protein species, LD-Exo has two unique proteins: cyclin Y and actinin alpha 4. In contrast, HD-Exo tightly encloses ephrin type-A receptor 2. Additionally, they found that there was some variation in the relative abundance of the some of common proteins. It has been partly explained that such protein-heterogeneity of EVs is related to the fact that EVs relies on ESCRT-dependent or -independent sorting machinery in sorting and packing some molecules including proteins.<sup>4,243</sup> Communally, the heterogeneity of EVs poses challenges for isolating specific EVs subpopulations with desired properties and for understanding their precise biological functions. To address this issue, advanced characterization techniques and novel isolation methods are required.

### Cargo loading efficiency

Optimizing the loading of multiple therapeutic agents into EVs without compromising their biophysical properties is a significant challenge.<sup>255</sup> Therapeutic payloads of interest can be loaded into EVs in a variety of methods as explained earlier. However, EVs loading efficiency is comparatively lower than liposome loading efficiency for instance.<sup>255,256</sup> There may not be enough room for exogenous medications to be loaded into EVs because EVs themselves retain some of the contents of their



parent cells during creation.<sup>255,257</sup> As such, loading of exogenous medicines into EVs is a significant obstacle.<sup>255,258</sup>

Since multiple techniques were employed to load oncotherapeutic agents into the same EVs as previously described, the techniques could be precisely compared.<sup>255</sup> A comprehensive literature review was performed to identify and compare different loading strategies. By categorizing these methods into high, medium, and low loading efficiency, researchers can select the most appropriate approach based on the specific cargo and desired outcome. However, literature has been highly controversial at that point. Haney *et al.* discovered, for instance, that the loading amount of catalase into EVs was raised in the following order: incubation, freeze/thaw cycle, sonication, extrusion.<sup>259</sup> According to Kim *et al.* there was an increase in the amount of PTX (paclitaxel) loaded into exosomes in the following order: incubation, electroporation, sonication.<sup>255,260</sup> Another study, when compared to incubation, electroporation, and extrusion, it was revealed that the amount of loading pharmaceuticals of saponin or hypotonic dialysis was up to 11 times higher.<sup>115,255</sup> However, the loading efficiency of EVs depends not only on loading techniques but also on the chemical composition of the EVs, the drug content, and lipophilicity.<sup>255</sup> Therefore, further work is required to create novel loading strategies in future studies and to optimize existing loading technologies.<sup>255</sup>

### Stability and storage

Preserving the integrity and bioactivity of EVs during storage is crucial for their therapeutic application.<sup>261</sup> While low temperatures, typically  $-80^{\circ}\text{C}$ , are commonly employed, factors such as storage buffer, cryoprotectants, and lyophilization can influence EVs stability.<sup>243,262</sup>

Although EVs were typically resuspended in phosphate buffer saline.<sup>46</sup> Yet, it has been reported that Trehalose has great advantage as a cryoprotectant, enhancing EVs stability during freezing and thawing cycles.<sup>263,264</sup> Lyophilization, another potential storage method, can reduce the need for ultra-low temperature storage, although its impact on EVs integrity requires further investigation.<sup>265</sup> Charoenviriyakul *et al.* reported that after lyophilization, the sample was held at room temperature and trehalose was added as a cryoprotectant to shield the exosomes from osmotic destruction. The outcomes demonstrate that lyophilization has minimal impact on the physical and biological properties of exosomes.<sup>265</sup> Optimizing EVs storage conditions is essential for maintaining their therapeutic potential and ensuring consistency in downstream applications.

### Lack of standardized isolation and purification method for EVs

A significant hurdle in advancing EV-based therapeutics is the lack of a standardized isolation method.<sup>255</sup> The heterogeneity of EVs populations, coupled with the diverse range of isolation techniques available, has hindered the development of reproducible and reliable EVs preparations.

To address the challenges associated with EVs isolation, various methods have been developed, each with its own strengths and limitations. These methods can be categorized based on their recovery and specificity: (1) high recovery, low specificity: ultracentrifugation, filtration, and precipitation techniques are commonly used due to their simplicity and relatively high yield, but they often result in low purity EVs preparations.<sup>266,267</sup> (2) Intermediate specificity and recovery: size-exclusion chromatography (SEC) offers improved purity compared to ultracentrifugation, but with lower recovery rates. (3) High specificity, low recovery: immunoaffinity capture and microfluidic-based techniques provide high purity but often suffer from low yield and limited throughput.<sup>266,267</sup> The ideal EVs isolation method would combine high recovery, specificity, efficiency, and reproducibility. However, no single method currently meets all these criteria.

To overcome these challenges, the development of standardized protocols, advanced characterization techniques, and innovative isolation methods is essential.

## Conclusion

In conclusion, EVs have emerged as versatile platforms with substantial potential to revolutionize cancer therapy. This review provides a thorough overview of EVs, emphasizing their promise as innovative drug delivery vehicles for cancer therapeutics. The authors highlight different types of EVs, isolation techniques, and different characterization methods. Moreover, the authors spot the light onto the journey and the transformation process of EVs from intracellular trafficking molecules to fully fortified drug delivery vehicles and especially focused on cancer therapeutics. The promising ability of EVs to carry diverse cargos has also been highlighted, including chemotherapeutic agents, nucleic acids, immunomodulatory molecules, and photosensitizers thus offering unique advantages over conventional therapeutic modalities.

EVs exhibit heterogeneity in terms of size, composition, and biological function, necessitating the development of standardized isolation and characterization methods. While challenges remain in optimizing EVs production, cargo loading, and delivery, the potential benefits of EV-based therapies warrant continued research and development.

By addressing the limitations associated with EVs heterogeneity and developing innovative strategies for EVs engineering and combination therapies, the field can progress towards the clinical translation of this promising technology. Ultimately, the successful development of EV-based therapeutics holds the potential to improve patient outcomes and transform cancer treatment.

## Data availability

This manuscript does not involve any experimental work.



## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

The authors acknowledge the financial support and sponsorship received from the Alexander von Humboldt Foundation, Germany.

## References

- 1 L. M. Doyle and M. Z. Wang, Overview of extracellular vesicles, their origin, composition, purpose, and methods for exosome isolation and analysis, *Cells*, 2019, **8**(7), 727.
- 2 R. Jaiswal and L. M. Sedger, Intercellular vesicular transfer by exosomes, microparticles and oncosomes-implications for cancer biology and treatments, *Front. Oncol.*, 2019, **9**, 125.
- 3 B. L. Deatherage and B. T. Cookson, Membrane vesicle release in bacteria, eukaryotes, and archaea: a conserved yet underappreciated aspect of microbial life, *Infect. Immun.*, 2012, **80**(6), 1948–1957.
- 4 G. Van Niel, G. d'Angelo and G. Raposo, Shedding light on the cell biology of extracellular vesicles, *Nat. Rev. Mol. Cell Biol.*, 2018, **19**(4), 213–228.
- 5 Z. Wang, *et al.*, Extracellular vesicles as an emerging drug delivery system for cancer treatment: Current strategies and recent advances, *Biomed. Pharmacother.*, 2022, **153**, 113480.
- 6 M. P. Zaborowski, *et al.*, Extracellular vesicles: composition, biological relevance, and methods of study, *Bioscience*, 2015, **65**(8), 783–797.
- 7 R. A. Youness, *et al.*, Oral Delivery of Psoralidin by Mucoadhesive Surface-Modified Bilosomes Showed Boosted Apoptotic and Necrotic Effects against Breast and Lung Cancer Cells, *Polymers*, 2023, **15**(6), 1464.
- 8 A. Ramzy, *et al.*, Drugless nanoparticles tune-up an array of intertwined pathways contributing to immune checkpoint signaling and metabolic reprogramming in triple-negative breast cancer, *Biomed. Mater.*, 2022, **18**(1), DOI: [10.1088/1748-605X/aca85d](https://doi.org/10.1088/1748-605X/aca85d).
- 9 R. A. Youness, *et al.*, A Snapshot of Photoresponsive Liposomes in Cancer Chemotherapy and Immunotherapy: Opportunities and Challenges, *ACS Omega*, 2023, **8**(47), 44424–44436.
- 10 N. K. Sedky, *et al.*, Box-Behnken design of thermo-responsive nano-liposomes loaded with a platinum(iv) anticancer complex: evaluation of cytotoxicity and apoptotic pathways in triple negative breast cancer cells, *Nanoscale Adv.*, 2023, **5**(19), 5399–5413.
- 11 D. K. Jeppesen, *et al.*, Reassessment of exosome composition, *Cell*, 2019, **177**(2), 428–445.
- 12 V. R. Minciucchi, M. R. Freeman, and D. Di Vizio, Extracellular vesicles in cancer: exosomes, microvesicles and the emerging role of large oncosomes, in *Seminars in Cell & Developmental Biology*, Elsevier, 2015.
- 13 F. T. Borges, L. Reis and N. Schor, Extracellular vesicles: structure, function, and potential clinical uses in renal diseases, *Braz. J. Med. Biol. Res.*, 2013, **46**, 824–830.
- 14 T. Huyan, *et al.*, Extracellular vesicles-advanced nanocarriers in cancer therapy: Progress and achievements, *Int. J. Nanomed.*, 2020, 6485–6502.
- 15 Y. A. ZeinElAbdeen, A. AbdAlSeed and R. A. Youness, Decoding Insulin-Like Growth Factor Signaling Pathway From a Non-coding RNAs Perspective: A Step Towards Precision Oncology in Breast Cancer, *J. Mammary Gland Biol. Neoplasia*, 2022, **27**(1), 79–99.
- 16 R. A. Youness, *et al.*, Contradicting interplay between insulin-like growth factor-1 and miR-486-5p in primary NK cells and hepatoma cell lines with a contemporary inhibitory impact on HCC tumor progression, *Growth Factors*, 2016, **34**(3–4), 128–140.
- 17 Y. M. Shaalan, *et al.*, Destabilizing the interplay between miR-1275 and IGF2BPs by Tamarix articulata and quercetin in hepatocellular carcinoma, *Nat. Prod. Res.*, 2018, **32**(18), 2217–2220.
- 18 N. A. Selem, *et al.*, Let-7a/cMyc/CCAT1/miR-17-5p Circuit Re-sensitizes Atezolizumab Resistance in Triple Negative Breast Cancer through Modulating PD-L1, *Pathol. Res. Pract.*, 2023, **248**, 154579.
- 19 R. M. Abdallah, *et al.*, Hindering the synchronization between miR-486-5p and H19 lncRNA by hesperetin halts breast cancer aggressiveness through tuning ICAM-1, *Anti Cancer Agents Med. Chem.*, 2022, **22**(3), 586–595.
- 20 R. A. Youness, *et al.*, A novel role of sONE/NOS3/NO signaling cascade in mediating hydrogen sulphide bilateral effects on triple negative breast cancer progression, *Nitric Oxide*, 2018, **80**, 12–23.
- 21 N. A. Selem, R. A. Youness and M. Z. Gad, What is beyond lncRNAs in breast cancer: A special focus on colon cancer-associated Transcript-1 (CCAT-1), *Non-coding RNA Res.*, 2021, **6**(4), 174–186.
- 22 R. Y. Mekky, *et al.*, MALAT-1: Immunomodulatory lncRNA hampering the innate and the adaptive immune arms in triple negative breast cancer, *Transl. Oncol.*, 2023, **31**, 101653.
- 23 A. Dawoud, *et al.*, Circular RNAs: New layer of complexity evading breast cancer heterogeneity, *Non-coding RNA Res.*, 2023, **8**(1), 60–74.
- 24 S. A. Fahmy, *et al.*, Molecular Engines, Therapeutic Targets, and Challenges in Pediatric Brain Tumors: A Special Emphasis on Hydrogen Sulfide and RNA-Based Nano-Delivery, *Cancers*, 2022, **14**(21), 5244.
- 25 T. Abaza, *et al.*, Emerging Role of Circular RNAs in Hepatocellular Carcinoma Immunotherapy, *Int. J. Mol. Sci.*, 2023, **24**(22), 16484.
- 26 S. E. Wang, Extracellular vesicles in cancer therapy, in *Seminars in Cancer Biology*, Elsevier, 2022.
- 27 N. M. Elelman, *et al.*, Expression of GPR68, an Acid-Sensing Orphan G Protein-Coupled Receptor, *Breast Cancer Front. Oncol.*, 2022, **12**, 847543.
- 28 N. M. Elelman, *et al.*, Editorial: Understanding the crosstalk between immune cells and the tumor microenvironment in



cancer and its implications for immunotherapy, *Front. Med.*, 2023, **10**, 1202581.

29 A. Becker, *et al.*, Extracellular vesicles in cancer: cell-to-cell mediators of metastasis, *Cancer cell*, 2016, **30**(6), 836–848.

30 A. R. Chin and S. E. Wang, Cancer tills the premetastatic field: mechanistic basis and clinical implications, *Clin. Cancer Res.*, 2016, **22**(15), 3725–3733.

31 S. E. Wang, Extracellular vesicles and metastasis, *Cold Spring Harb. Perspect. Med.*, 2020, **10**(7), a037275.

32 Y. Zhang and X.-F. Wang, A niche role for cancer exosomes in metastasis, *Nat. Cell Biol.*, 2015, **17**(6), 709–711.

33 N. I. Rizk, *et al.*, Revealing the role of serum exosomal novel long non-coding RNA NAMPT-AS as a promising diagnostic/prognostic biomarker in colorectal cancer patients, *Life Sci.*, 2024, **352**, 122850.

34 Y. K. Abd El Fattah, *et al.*, CCDC144NL-AS1/hsa-miR-143-3p/HMGA2 interaction: In-silico and clinically implicated in CRC progression, correlated to tumor stage and size in case-controlled study; step toward ncRNA precision, *Int. J. Biol. Macromol.*, 2023, **253**, 126739.

35 T. A. Shtam, *et al.*, Exosomes are natural carriers of exogenous siRNA to human cells in vitro, *Cell Commun. Signal.*, 2013, **11**(1), 1–10.

36 M. Bakhti, C. Winter and M. Simons, Inhibition of myelin membrane sheath formation by oligodendrocyte-derived exosome-like vesicles, *J. Biol. Chem.*, 2011, **286**(1), 787–796.

37 S. Wang, *et al.*, Synapsin I is an oligomannose-carrying glycoprotein, acts as an oligomannose-binding lectin, and promotes neurite outgrowth and neuronal survival when released via glia-derived exosomes, *J. Neurosci.*, 2011, **31**(20), 7275–7290.

38 E. R. Abels and X. O. Breakefield, Introduction to extracellular vesicles: biogenesis, RNA cargo selection, content, release, and uptake, *Cell. Mol. Neurobiol.*, 2016, **36**, 301–312.

39 L. A. Mulcahy, R. C. Pink and D. R. F. Carter, Routes and mechanisms of extracellular vesicle uptake, *J. Extracell. Vesicles*, 2014, **3**(1), 24641.

40 R. A. Haraszti, *et al.*, High-resolution proteomic and lipidomic analysis of exosomes and microvesicles from different cell sources, *J. Extracell. Vesicles*, 2016, **5**(1), 32570.

41 C. N. Suire and M. D. Hade, Extracellular vesicles in type 1 diabetes: A versatile tool, *Bioengineering*, 2022, **9**(3), 105.

42 G. Raposo and W. Stoorvogel, Extracellular vesicles: exosomes, microvesicles, and friends, *J. Cell Biol.*, 2013, **200**(4), 373–383.

43 M. P. Bebelman, *et al.*, Biogenesis and function of extracellular vesicles in cancer, *Pharmacol. Therapeut.*, 2018, **188**, 1–11.

44 W. Möbius, *et al.*, Immunoelectron microscopic localization of cholesterol using biotinylated and non-cytolytic perfringolysin O, *J. Histochem. Cytochem.*, 2002, **50**(1), 43–55.

45 B. J. Tauro, *et al.*, Comparison of ultracentrifugation, density gradient separation, and immunoaffinity capture methods for isolating human colon cancer cell line LIM1863-derived exosomes, *Methods*, 2012, **56**(2), 293–304.

46 K. W. Witwer, *et al.*, Standardization of sample collection, isolation and analysis methods in extracellular vesicle research, *J. Extracell. Vesicles*, 2013, **2**(1), 20360.

47 J. C. Akers, *et al.*, Biogenesis of extracellular vesicles (EV): exosomes, microvesicles, retrovirus-like vesicles, and apoptotic bodies, *J. Neuro Oncol.*, 2013, **113**, 1–11.

48 C. Théry, *et al.*, Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines, *J. Extracell. Vesicles*, 2018, **7**(1), 1535750.

49 H. Cai, K. Reinisch and S. Ferro-Novick, Coats, tethers, Rabs, and SNAREs work together to mediate the intracellular destination of a transport vesicle, *Dev. Cell*, 2007, **12**(5), 671–682.

50 C. Tricarico, J. Clancy and C. D'Souza-Schorey, Biology and biogenesis of shed microvesicles, *Small GTPases*, 2017, **8**(4), 220–232.

51 X. Xu, Y. Lai and Z.-C. Hua, Apoptosis and apoptotic body: disease message and therapeutic target potentials, *Biosci. Rep.*, 2019, **39**(1), BSR20180992.

52 M. Battistelli and E. Falcieri, Apoptotic bodies: particular extracellular vesicles involved in intercellular communication, *Advances in Medical Biochemistry, Genomics, Physiology, and Pathology*, 2021, pp. 473–486.

53 S. Elmore, Apoptosis: a review of programmed cell death, *Toxicol. Pathol.*, 2007, **35**(4), 495–516.

54 M. C. Cufaro, *et al.*, Extracellular vesicles and their potential use in monitoring cancer progression and therapy: the contribution of proteomics, *J. Oncol.*, 2019, 2019.

55 V. R. Minciachchi, *et al.*, Large oncosomes contain distinct protein cargo and represent a separate functional class of tumor-derived extracellular vesicles, *Oncotarget*, 2015, **6**(13), 11327.

56 S. R. Li, *et al.*, Tissue-derived extracellular vesicles in cancers and non-cancer diseases: Present and future, *J. Extracell. Vesicles*, 2021, **10**(14), e12175.

57 T. Vagner, *et al.*, Large extracellular vesicles carry most of the tumour DNA circulating in prostate cancer patient plasma, *J. Extracell. Vesicles*, 2018, **7**(1), 1505403.

58 S. A. Fahmy, *et al.*, Emerging tendencies for the nano-delivery of gambogic acid: a promising approach in oncotherapy, *RSC Adv.*, 2024, **14**(7), 4666–4691.

59 M. Dimik, *et al.*, The exosome: a review of current therapeutic roles and capabilities in human reproduction, *Drug Delivery Transl. Res.*, 2023, **13**(2), 473–502.

60 T. Wollert and J. H. Hurley, Molecular mechanism of multivesicular body biogenesis by ESCRT complexes, *Nature*, 2010, **464**(7290), 864–869.

61 N. M. Elelman, *et al.*, Pharmacogenomic and epigenomic approaches to untangle the enigma of IL-10 blockade in oncology, *Expert Rev. Mol. Med.*, 2024, **26**, e1.

62 A. H. Soliman, *et al.*, Phytochemical-derived tumor-associated macrophage remodeling strategy using Phoenix dactylifera L. boosted photodynamic therapy in melanoma via H19/iNOS/PD-L1 axis, *Photodiagnosis Photodyn. Ther.*, 2023, **44**, 103792.



63 H. Nafea, *et al.*, Dual targeting of H(2)S synthesizing enzymes; cystathionine  $\beta$ -synthase and cystathionine  $\gamma$ -lyase by miR-939-5p effectively curbs triple negative breast cancer, *Helyon*, 2023, **9**(10), e21063.

64 A. Dawoud, *et al.*, Pan-inhibition of the three H(2)S synthesizing enzymes restrains tumor progression and immunosuppression in breast cancer, *Cancer Cell Int.*, 2024, **24**(1), 136.

65 R. A. Youness, *et al.*, Targeting hydrogen sulphide signaling in breast cancer, *J. Adv. Res.*, 2021, **27**, 177–190.

66 M. K. A. El-Aziz, *et al.*, Decoding hepatocarcinogenesis from a noncoding RNAs perspective, *J. Cell. Physiol.*, 2023, **238**(9), 1982–2009.

67 F. H. El Kilany, *et al.*, miR-744/eNOS/NO axis: A novel target to halt triple negative breast cancer progression, *Breast Dis.*, 2021, **40**(3), 161–169.

68 S. A. Fahmy, *et al.*, Hijacking 5-Fluorouracil Chemoresistance in Triple Negative Breast Cancer via microRNAs-Loaded Chitosan Nanoparticles, *Int. J. Mol. Sci.*, 2024, **25**(4), 2070.

69 R. Kalluri and V. S. LeBleu, The biology, function, and biomedical applications of exosomes, *Science*, 2020, **367**(6478), eaau6977.

70 M. A. Kumar, *et al.*, Extracellular vesicles as tools and targets in therapy for diseases, *Signal Transduct. Targeted Ther.*, 2024, **9**(1), 27.

71 R. A. Youness, *et al.*, Heat Shock Proteins: Central Players in Oncological and Immuno-Oncological Tracks, *Adv. Exp. Med. Biol.*, 2023, **1409**, 193–203.

72 Z. Hassan, *et al.*, How Viruses Hijack and Modify the Secretory Transport Pathway, *Cells*, 2021, **10**(10), 2535.

73 E. Calzada, O. Onguka and S. M. Claypool, Phosphatidylethanolamine Metabolism in Health and Disease, *Int. Rev. Cell Mol. Biol.*, 2016, **321**, 29–88.

74 K. S. Negara, *et al.*, The role of caspase-dependent and caspase-independent pathways of apoptosis in the premature rupture of the membranes: A case-control study, *Int. J. Reprod. Biomed.*, 2020, **18**(6), 439.

75 E. Petrovčíková, K. Vičíková and V. Leksa, Extracellular vesicles–biogenesis, composition, function, uptake and therapeutic applications, *Biologia*, 2018, **73**(4), 437–448.

76 C. Ciardiello, *et al.*, Focus on extracellular vesicles: new frontiers of cell-to-cell communication in cancer, *Int. J. Mol. Sci.*, 2016, **17**(2), 175.

77 M. Sheta, *et al.*, Extracellular vesicles: New classification and tumor immunosuppression, *Biology*, 2023, **12**(1), 110.

78 T. Eguchi, *et al.*, Cell stress induced stressome release including damaged membrane vesicles and extracellular HSP90 by prostate cancer cells, *Cells*, 2020, **9**(3), 755.

79 R. A. Assal, *et al.*, A Novel Epigenetic Strategy to Concurrently Block Immune Checkpoints PD-1/PD-L1 and CD155/TIGIT in Hepatocellular Carcinoma, *Transl. Oncol.*, 2024, **45**, 101961.

80 S. Walker, *et al.*, Extracellular vesicle-based drug delivery systems for cancer treatment, *Theranostics*, 2019, **9**(26), 8001.

81 H. H. Rashwan, *et al.*, Harnessing the supremacy of MEG3 LncRNA to defeat gastrointestinal malignancies, *Pathol. Res. Pract.*, 2024, **256**, 155223.

82 S. Tiwari, *et al.*, Preparation and characterization of extracellular vesicles, *Am. J. Reprod. Immunol.*, 2021, **85**(2), e13367.

83 K. Brennan, *et al.*, A comparison of methods for the isolation and separation of extracellular vesicles from protein and lipid particles in human serum, *Sci. Rep.*, 2020, **10**(1), 1039.

84 A. Akbar, *et al.*, Methodologies to Isolate and Purify Clinical Grade Extracellular Vesicles for Medical Applications, *Cells*, 2022, **11**(2), 186.

85 M. Y. Konoshenko, *et al.*, Isolation of Extracellular Vesicles: General Methodologies and Latest Trends, *Biomed. Res. Int.*, 2018, **2018**, 8545347.

86 C. Théry, *et al.*, Isolation and characterization of exosomes from cell culture supernatants and biological fluids, *Curr. Protoc. Cell Biol.*, 2006, **30**(1), 1–29.

87 P. M. Vanderboom, *et al.*, A size-exclusion-based approach for purifying extracellular vesicles from human plasma, *Cell Rep. Methods*, 2021, **1**(3), 100055.

88 L. Mincheva-Nilsson, *et al.*, Isolation and characterization of exosomes from cultures of tissue explants and cell lines, *Curr. Protoc. Im.*, 2016, **115**(1), 1–21.

89 F. Momen-Heravi, *et al.*, Current methods for the isolation of extracellular vesicles, *Biol. Chem.*, 2013, **394**(10), 1253–1262.

90 M. A. Rider, S. N. Hurwitz and D. G. Meckes Jr, ExtraPEG: a polyethylene glycol-based method for enrichment of extracellular vesicles, *Sci. Rep.*, 2016, **6**(1), 23978.

91 H. Shin, *et al.*, High-yield isolation of extracellular vesicles using aqueous two-phase system, *Sci. Rep.*, 2015, **5**(1), 13103.

92 M. L. Heinemann, *et al.*, Benchtop isolation and characterization of functional exosomes by sequential filtration, *J. Chromatogr. A*, 2014, **1371**, 125–135.

93 F. Liu, *et al.*, The exosome total isolation chip, *ACS Nano*, 2017, **11**(11), 10712–10723.

94 M. Logozzi, *et al.*, Immunocapture-based ELISA to characterize and quantify exosomes in both cell culture supernatants and body fluids, *Methods Enzymol.*, 2020, **645**, 155–180.

95 D. Sun, *et al.*, Recent progress in aptamer-based microfluidics for the detection of circulating tumor cells and extracellular vesicles, *J. Pharm. Anal.*, 2023, **13**(4), 340–354.

96 R. Vaidyanathan, *et al.*, Detecting exosomes specifically: a multiplexed device based on alternating current electrohydrodynamic induced nanoshearing, *Anal. Chem.*, 2014, **86**(22), 11125–11132.

97 J. Zhang, *et al.*, Exosome and exosomal microRNA: trafficking, sorting, and function, *Genomics, Proteomics Bioinf.*, 2015, **13**(1), 17–24.

98 G. Corso, *et al.*, Reproducible and scalable purification of extracellular vesicles using combined bind-elute and size exclusion chromatography, *Sci. Rep.*, 2017, **7**(1), 11561.



99 V. S. Chernyshev, *et al.*, Asymmetric depth-filtration: A versatile and scalable method for high-yield isolation of extracellular vesicles with low contamination, *J. Extracell. Vesicles*, 2022, **11**(8), e12256.

100 S. Cheng, *et al.*, Advances in microfluidic extracellular vesicle analysis for cancer diagnostics, *Lab Chip*, 2021, **21**(17), 3219–3243.

101 J. Suthar, *et al.*, Recent developments in biosensing methods for extracellular vesicle protein characterization, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2023, **15**(1), e1839.

102 L. M. Doyle and M. Z. Wang, Overview of Extracellular Vesicles, Their Origin, Composition, Purpose, and Methods for Exosome Isolation and Analysis, *Cells*, 2019, **8**(7), 727.

103 V. C. Kok and C. C. Yu, Cancer-Derived Exosomes: Their Role in Cancer Biology and Biomarker Development, *Int. J. Nanomed.*, 2020, **15**, 8019–8036.

104 M. Domsicova, *et al.*, New Insights into Aptamers: An Alternative to Antibodies in the Detection of Molecular Biomarkers, *Int. J. Mol. Sci.*, 2024, **25**(13), 6833.

105 A. D. Flynn and H. Yin, Lipid-Targeting Peptide Probes for Extracellular Vesicles, *J. Cell. Physiol.*, 2016, **231**(11), 2327–2332.

106 D. Buschmann, V. Mussack and J. B. Byrd, Separation, characterization, and standardization of extracellular vesicles for drug delivery applications, *Adv. Drug Deliv. Rev.*, 2021, **174**, 348–368.

107 S. Cho, *et al.*, Isolation of extracellular vesicle from blood plasma using electrophoretic migration through porous membrane, *Sens. Actuators, B*, 2016, **233**, 289–297.

108 F. G. Ortega-Sánchez, *et al.*, Microfluidic systems in extracellular vesicles single analysis. A systematic review, *Trac. Trends Anal. Chem.*, 2023, **159**, 116920.

109 T. A. Duncombe, A. M. Tentori and A. E. Herr, Microfluidics: reframing biological enquiry, *Nat. Rev. Mol. Cell Biol.*, 2015, **16**(9), 554–567.

110 Y. Zeinelabdeen, *et al.*, MIAT LncRNA: A multifunctional key player in non-oncological pathological conditions, *Non-coding RNA Res.*, 2024, **9**(2), 447–462.

111 B. S. Joshi, D. Ortiz and I. S. Zuhorn, Converting extracellular vesicles into nanomedicine: loading and unloading of cargo, *Mat. Today Nano*, 2021, **16**, 100148.

112 A. Al-Jipouri, *et al.*, Liposomes or extracellular vesicles: a comprehensive comparison of both lipid bilayer vesicles for pulmonary drug delivery, *Polymers*, 2023, **15**(2), 318.

113 A. Jeyaram, *et al.*, Enhanced Loading of Functional miRNA Cargo via pH Gradient Modification of Extracellular Vesicles, *Mol. Ther.*, 2020, **28**(3), 975–985.

114 Y. Han, *et al.*, Overview and Update on Methods for Cargo Loading into Extracellular Vesicles, *Processes*, 2021, **9**(2), 356.

115 G. Fuhrmann, *et al.*, Active loading into extracellular vesicles significantly improves the cellular uptake and photodynamic effect of porphyrins, *J. Contr. Release*, 2015, **205**, 35–44.

116 Q. Liu, *et al.*, Targeted therapy using engineered extracellular vesicles: principles and strategies for membrane modification, *J. Nanobiotechnol.*, 2023, **21**(1), 334.

117 I. Podolak, A. Galanty and D. Sobolewska, Saponins as cytotoxic agents: a review, *Phytochemistry Rev.*, 2010, **9**, 425–474.

118 F. Villa, R. Quarto and R. Tasso, Extracellular Vesicles as Natural, Safe and Efficient Drug Delivery Systems, *Pharmaceutics*, 2019, **11**(11), 557.

119 C. Chen, *et al.*, Active cargo loading into extracellular vesicles: Highlights the heterogeneous encapsulation behaviour, *J. Extracell. Vesicles*, 2021, **10**(13), e12163.

120 X. Zhang, *et al.*, Engineered extracellular vesicles for cancer therapy, *Adv. Mater.*, 2021, **33**(14), 2005709.

121 J. Chen, *et al.*, Engineered extracellular vesicles: potentials in cancer combination therapy, *J. Nanobiotechnol.*, 2022, **20**(1), 132.

122 L. Ramasubramanian, P. Kumar and A. Wang, Engineering extracellular vesicles as nanotherapeutics for regenerative medicine, *Biomolecules*, 2019, **10**(1), 48.

123 G. Rosso and V. Cauda, Biomimicking Extracellular Vesicles with Fully Artificial Ones: A Rational Design of EV-BIOMIMETICS toward Effective Theranostic Tools in Nanomedicine, *ACS Biomater. Sci. Eng.*, 2022, 5924–5932.

124 K. P. De Sousa, *et al.*, Isolation and characterization of extracellular vesicles and future directions in diagnosis and therapy, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2023, **15**(1), e1835.

125 M. I. Ramirez, *et al.*, Technical challenges of working with extracellular vesicles, *Nanoscale*, 2018, **10**(3), 881–906.

126 C. Gardiner, *et al.*, Techniques used for the isolation and characterization of extracellular vesicles: results of a worldwide survey, *J. Extracell. Vesicles*, 2016, **5**(1), 32945.

127 E. Van der Pol, *et al.*, Particle size distribution of exosomes and microvesicles determined by transmission electron microscopy, flow cytometry, nanoparticle tracking analysis, and resistive pulse sensing, *J. Thromb. Haemostasis*, 2014, **12**(7), 1182–1192.

128 D. Fortunato, *et al.*, Opportunities and pitfalls of fluorescent labeling methodologies for extracellular vesicle profiling on high-resolution single-particle platforms, *Int. J. Mol. Sci.*, 2021, **22**(19), 10510.

129 C. Bağcı, *et al.*, Overview of extracellular vesicle characterization techniques and introduction to combined reflectance and fluorescence confocal microscopy to distinguish extracellular vesicle subpopulations, *Neurophotonics*, 2022, **9**(2), 021903.

130 A. Shehzad, *et al.*, Extracellular vesicles in cancer diagnostics and therapeutics, *Pharmacol. Ther.*, 2021, **223**, 107806.

131 K. E. Thane, A. M. Davis and A. M. Hoffman, Improved methods for fluorescent labeling and detection of single extracellular vesicles using nanoparticle tracking analysis, *Sci. Rep.*, 2019, **9**(1), 12295.



132 R. Szatanek, *et al.*, The methods of choice for extracellular vesicles (EVs) characterization, *Int. J. Mol. Sci.*, 2017, **18**(6), 1153.

133 C. M. Hoo, *et al.*, A comparison of atomic force microscopy (AFM) and dynamic light scattering (DLS) methods to characterize nanoparticle size distributions, *J. Nanoparticle Res.*, 2008, **10**, 89–96.

134 W. C. S. Cho, Extracellular Vesicles: Biology and Potentials in Cancer Therapeutics, *Int. J. Mol. Sci.*, 2021, **22**(17), 9586.

135 K. P. De Sousa, *et al.*, Isolation and characterization of extracellular vesicles and future directions in diagnosis and therapy, *WIREs Nanomed. Nanobiotechnol.*, 2023, **15**(1), e1835.

136 A. Kotrbová, *et al.*, TEM ExosomeAnalyzer: a computer-assisted software tool for quantitative evaluation of extracellular vesicles in transmission electron microscopy images, *J. Extracell. Vesicles*, 2019, **8**(1), 1560808.

137 Z. Zhao, *et al.*, Isolation and analysis methods of extracellular vesicles (EVs), *Extracell. Vesicles Circ. Nucleic Acids*, 2021, **2**, 80–103.

138 D. Bachurski, *et al.*, Extracellular vesicle measurements with nanoparticle tracking analysis—An accuracy and repeatability comparison between NanoSight NS300 and ZetaView, *J. Extracell. Vesicles*, 2019, **8**(1), 1596016.

139 M. Malenica, *et al.*, Perspectives of Microscopy Methods for Morphology Characterisation of Extracellular Vesicles from Human Biofluids, *Biomedicines*, 2021, **9**(6), 603.

140 Y. Yuana, *et al.*, Atomic force microscopy: a novel approach to the detection of nanosized blood microparticles, *J. Thromb. Haemostasis*, 2010, **8**(2), 315–323.

141 P. Parisse, *et al.*, Atomic force microscopy analysis of extracellular vesicles, *Eur. Biophys. J.*, 2017, **46**, 813–820.

142 M. i. Li, *et al.*, Multiparametric atomic force microscopy imaging of single native exosomes, *Acta Biochim. Biophys. Sin.*, 2021, **53**(3), 385–388.

143 K. Singh, *et al.*, Separation of distinct exosome subpopulations: isolation and characterization approaches and their associated challenges, *Analyst*, 2021, **146**(12), 3731–3749.

144 T. Ito, *et al.*, Comparison of nanoparticle size and electrophoretic mobility measurements using a carbon-nanotube-based coulter counter, dynamic light scattering, transmission electron microscopy, and phase analysis light scattering, *Langmuir*, 2004, **20**(16), 6940–6945.

145 S. L. Maas, M. L. Broekman and J. de Vrij, Tunable resistive pulse sensing for the characterization of extracellular vesicles. Exosomes and microvesicles, *Methods Protoc.*, 2017, 21–33.

146 A. Morales-Kastresana and J. C. Jones, Flow Cytometric Analysis of Extracellular Vesicles, *Methods Mol. Biol.*, 2017, **1545**, 215–225.

147 E. Van Der Pol, *et al.*, Optical and non-optical methods for detection and characterization of microparticles and exosomes, *J. Thromb. Haemostasis*, 2010, **8**(12), 2596–2607.

148 K. P. De Sousa, *et al.*, Isolation and characterization of extracellular vesicles and future directions in diagnosis and therapy, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2023, **15**(1), e1835.

149 J. M. Carnino, H. Lee and Y. Jin, Isolation and characterization of extracellular vesicles from Bronchoalveolar lavage fluid: a review and comparison of different methods, *Respir.*, 2019, **20**, 1–11.

150 N. Salmond, *et al.*, Nanoscale flow cytometry for immunophenotyping and quantitating extracellular vesicles in blood plasma, *Nanoscale*, 2021, **13**(3), 2012–2025.

151 E. Serrano-Pertierra, *et al.*, Extracellular vesicles: current analytical techniques for detection and quantification, *Biomolecules*, 2020, **10**(6), 824.

152 D. L. Rupert, *et al.*, Methods for the physical characterization and quantification of extracellular vesicles in biological samples, *Biochim. Biophys. Acta Gen. Subj.*, 2017, **1861**(1), 3164–3179.

153 G. Midekessa, *et al.*, Zeta potential of extracellular vesicles: toward understanding the attributes that determine colloidal stability, *ACS Omega*, 2020, **5**(27), 16701–16710.

154 S. Y. Liu, *et al.*, Diagnostic Role of Extracellular Vesicles in Cancer: A Comprehensive Systematic Review and Meta-Analysis, *Front. Cell Dev. Biol.*, 2021, **9**, 705791.

155 E. M. Veziroglu and G. I. Mias, Characterizing extracellular vesicles and their diverse RNA contents, *Front. Genet.*, 2020, **11**, 700.

156 E. Weatherall and G. R. Willmott, Applications of tunable resistive pulse sensing, *Analyst*, 2015, **140**(10), 3318–3334.

157 T. Akagi, *et al.*, On-chip immunoelectrophoresis of extracellular vesicles released from human breast cancer cells, *One*, 2015, **10**(4), e0123603.

158 K. P. De Sousa, *et al.*, Proteomic identification of the contents of small extracellular vesicles from *in vivo* Plasmodium yoelii infection, *Int. J. Parasitol.*, 2022, **52**(1), 35–45.

159 Y. Yuana, A. Sturk and R. Nieuwland, Extracellular vesicles in physiological and pathological conditions, *Blood Rev.*, 2013, **27**(1), 31–39.

160 J. Conde-Vancells, *et al.*, Characterization and comprehensive proteome profiling of exosomes secreted by hepatocytes, *J. Proteome Res.*, 2008, **7**(12), 5157–5166.

161 H. Shao, *et al.*, New technologies for analysis of extracellular vesicles, *Chem. Rev.*, 2018, **118**(4), 1917–1950.

162 R. R. Mizenko, *et al.*, Tetraspanins are unevenly distributed across single extracellular vesicles and bias sensitivity to multiplexed cancer biomarkers, *J. Nanobiotechnol.*, 2021, **19**(1), 250.

163 J. Lötvall, A. F. Hill, F. Hochberg, E. I. Buzás, D. Di Vizio, C. Gardiner, Y. S. Gho, I. V. Kurochkin, S. Mathivanan, P. Quesenberry, S. Sahoo, H. Tahara, M. H. Wauben, K. W. Witwer and C. Théry, Minimal experimental requirements for definition of extracellular vesicles and their functions: a position statement from the International Society for Extracellular Vesicles, *J. Extracell. Vesicles*, 2014, **3**, 26913.



164 Y. Yoshioka, *et al.*, Comparative marker analysis of extracellular vesicles in different human cancer types, *J. Extracell. Vesicles*, 2013, **2**(1), 20424.

165 J. Suthar, *et al.*, Recent developments in biosensing methods for extracellular vesicle protein characterization, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2023, **15**(1), e1839.

166 T. A. Hartjes, *et al.*, Extracellular vesicle quantification and characterization: common methods and emerging approaches, *Bioengineering*, 2019, **6**(1), 7.

167 Y. Jiang, *et al.*, Aptamer/AuNP biosensor for colorimetric profiling of exosomal proteins, *Angew. Chem. Int. Ed.*, 2017, **56**(39), 11916–11920.

168 B. Domon and R. Aebersold, Options and considerations when selecting a quantitative proteomics strategy, *Nat. Biotechnol.*, 2010, **28**(7), 710–721.

169 A. Y. T. Wu, K. Ueda and C. P. K. Lai, Proteomic analysis of extracellular vesicles for cancer diagnostics, *Proteomics*, 2019, **19**(1–2), 1800162.

170 S. Rontogianni, *et al.*, Proteomic profiling of extracellular vesicles allows for human breast cancer subtyping, *Commun. Biol.*, 2019, **2**(1), 325.

171 J. Zhou, *et al.*, A sensitive detection assay based on signal amplification technology for Alzheimer's disease's early biomarker in exosome, *Anal. Chim. Acta*, 2018, **1022**, 124–130.

172 K. Lee, *et al.*, Multiplexed profiling of single extracellular vesicles, *ACS Nano*, 2018, **12**(1), 494–503.

173 W. Zhao, *et al.*, Microsphere mediated exosome isolation and ultra-sensitive detection on a dielectrophoresis integrated microfluidic device, *Analyst*, 2021, **146**(19), 5962–5972.

174 M. Huang, *et al.*, Homogeneous, low-volume, efficient, and sensitive quantitation of circulating exosomal PD-L1 for cancer diagnosis and immunotherapy response prediction, *Angew. Chem.*, 2020, **132**(12), 4830–4835.

175 Y. Li, *et al.*, Molecular identification of tumor-derived extracellular vesicles using thermophoresis-mediated DNA computation, *J. Am. Chem. Soc.*, 2021, **143**(3), 1290–1295.

176 S. A. Hinger, *et al.*, Diverse long RNAs are differentially sorted into extracellular vesicles secreted by colorectal cancer cells, *Cell Rep.*, 2018, **25**(3), 715–725.

177 K. O'Brien, *et al.*, RNA delivery by extracellular vesicles in mammalian cells and its applications, *Nat. Rev. Mol. Cell Biol.*, 2020, **21**(10), 585–606.

178 V. N. Aushev, *et al.*, Comparisons of microRNA patterns in plasma before and after tumor removal reveal new biomarkers of lung squamous cell carcinoma, *PloS One*, 2013, **8**(10), e78649.

179 L. Manterola, *et al.*, A small noncoding RNA signature found in exosomes of GBM patient serum as a diagnostic tool, *Neuro Oncol.*, 2014, **16**(4), 520–527.

180 D. Qin, *et al.*, Detection of pancreatic ductal adenocarcinoma by a qPCR-based normalizer-free circulating extracellular vesicles RNA signature, *J. Cancer*, 2021, **12**(5), 1445.

181 P. M. Ozawa, *et al.*, Identification of miRNAs enriched in extracellular vesicles derived from serum samples of breast cancer patients, *Biomolecules*, 2020, **10**(1), 150.

182 H. Xu, *et al.*, Circular RNA expression in extracellular vesicles isolated from serum of patients with endometrial cancer, *Epigenomics*, 2018, **10**(2), 187–197.

183 L. Dong, *et al.*, Circulating long RNAs in serum extracellular vesicles: their characterization and potential application as biomarkers for diagnosis of colorectal cancer, *Cancer Epidemiol. Biomarkers Prev.*, 2016, **25**(7), 1158–1166.

184 L. Moldovan, *et al.*, Analyzing the circulating microRNAs in exosomes/extracellular vesicles from serum or plasma by qRT-PCR. Circulating MicroRNAs, *Methods Protoc.*, 2013, 129–145.

185 M. Del Re, *et al.*, Reply to Ugo De Giorgi, Vincenza Conteduca, and Emanuela Scarpi's Letter to the Editor re: Marzia Del Re, Elisa Biasco, Stefania Crucitta, *et al.* The Detection of Androgen Receptor Splice Variant 7 in Plasma-derived Exosomal RNA Strongly Predicts Resistance to Hormonal Therapy in Metastatic Prostate Cancer Patients. *Eur Urol* 2017; **71**: 680–7, *Eur. Urol.*, 2017, **73**(1), e11–e12.

186 W. W. Chen, *et al.*, BEAMing and droplet digital PCR analysis of mutant IDH1 mRNA in glioma patient serum and cerebrospinal fluid extracellular vesicles, *Mol. Ther. Nucleic Acids*, 2013, **2**, e109.

187 M. G. Amorim, *et al.*, A total transcriptome profiling method for plasma-derived extracellular vesicles: applications for liquid biopsies, *Rep.*, 2017, **7**(1), 1–11.

188 L. Cheng, *et al.*, Characterization and deep sequencing analysis of exosomal and non-exosomal miRNA in human urine, *Kidney Int.*, 2014, **86**(2), 433–444.

189 A. A. Sabo, *et al.*, Small non-coding RNA profiling in plasma extracellular vesicles of bladder cancer patients by next-generation sequencing: expression levels of miR-126-3p and piR-5936 increase with higher histologic grades, *Cancers*, 2020, **12**(6), 1507.

190 J. D. Kuhlmann, *et al.*, Extracellular vesicle-associated miRNAs in ovarian cancer—design of an integrated NGS-based workflow for the identification of blood-based biomarkers for platinum-resistance, *Clin. Chem. Lab. Med.*, 2019, **57**(7), 1053–1062.

191 T. Bjørnetrø, *et al.*, Next-generation sequencing reveals mitogenome diversity in plasma extracellular vesicles from colorectal cancer patients, *BMC Cancer*, 2023, **23**(1), 650.

192 J. W. Roy, *et al.*, Small RNA sequencing analysis of peptide-affinity isolated plasma extracellular vesicles distinguishes pancreatic cancer patients from non-affected individuals, *Sci. Rep.*, 2023, **13**(1), 9251.

193 S. Kreimer, *et al.*, Mass-spectrometry-based molecular characterization of extracellular vesicles: lipidomics and proteomics, *J. Proteome Res.*, 2015, **14**(6), 2367–2384.

194 A. Zebrowska, *et al.*, Metabolome of exosomes: Focus on vesicles released by cancer cells and present in human body fluids, *Int. J. Mol. Sci.*, 2019, **20**(14), 3461.



195 D. Schwudke, *et al.*, Shotgun lipidomics on high resolution mass spectrometers, *Cold Spring Harbor Perspect. Biol.*, 2011, **3**(9), a004614.

196 N. Nishida-Aoki, *et al.*, Lipidomic analysis of cells and extracellular vesicles from high-and low-metastatic triple-negative breast cancer, *Metabolites*, 2020, **10**(2), 67.

197 J. S. Brzozowski, *et al.*, Lipidomic profiling of extracellular vesicles derived from prostate and prostate cancer cell lines, *Lipids Health Dis.*, 2018, **17**, 1–12.

198 T. Altadill, *et al.*, Enabling metabolomics based biomarker discovery studies using molecular phenotyping of exosome-like vesicles, *PLoS One*, 2016, **11**(3), e0151339.

199 M. Clos-Garcia, *et al.*, Metabolic alterations in urine extracellular vesicles are associated to prostate cancer pathogenesis and progression, *J. Extracell. Vesicles*, 2018, **7**(1), 1470442.

200 R. Hayasaka, *et al.*, Metabolomic analysis of small extracellular vesicles derived from pancreatic cancer cells cultured under normoxia and hypoxia, *Metabolites*, 2021, **11**(4), 215.

201 M. Harmati, *et al.*, The role of the metabolite cargo of extracellular vesicles in tumor progression, *Cancer Metastasis Rev.*, 2021, 1–19.

202 M. Palviainen, *et al.*, Cancer alters the metabolic fingerprint of extracellular vesicles, *Cancers*, 2020, **12**(11), 3292.

203 M. Čuperlović-Cuflik, *et al.*, Analysis and simulation of glioblastoma cell lines-derived extracellular vesicles metabolome, *Metabolites*, 2020, **10**(3), 88.

204 R. A. Youness, *et al.*, A comprehensive insight and *in Silico* analysis of CircRNAs in hepatocellular carcinoma: a step toward ncRNA-based precision medicine, *Cells*, 2024, **13**(15), 1245.

205 Y. Ma, *et al.*, Extracellular vesicles: an emerging nanoplatform for cancer therapy, *Front. Oncol.*, 2021, **10**, 606906.

206 D. Sousa, R. T. Lima and M. H. Vasconcelos, Intercellular transfer of cancer drug resistance traits by extracellular vesicles, *Trends Mol. Med.*, 2015, **21**(10), 595–608.

207 J. Liu, *et al.*, Functional extracellular vesicles engineered with lipid-grafted hyaluronic acid effectively reverse cancer drug resistance, *Biomaterials*, 2019, **223**, 119475.

208 Q. Lin, *et al.*, Exosome-like nanoplatform modified with targeting ligand improves anti-cancer and anti-inflammation effects of imperialine, *J. Contr. Release*, 2019, **311–312**, 104–116.

209 L. Qiao, *et al.*, Tumor cell-derived exosomes home to their cells of origin and can be used as Trojan horses to deliver cancer drugs, *Theranostics*, 2020, **10**(8), 3474.

210 K.-L. Zhang, *et al.*, Artificial chimeric exosomes for anti-phagocytosis and targeted cancer therapy, *Chem. Sci.*, 2019, **10**(5), 1555–1561.

211 Y. Tian, *et al.*, A doxorubicin delivery platform using engineered natural membrane vesicle exosomes for targeted tumor therapy, *Biomaterials*, 2014, **35**(7), 2383–2390.

212 M. J. Haney, *et al.*, Macrophage-derived extracellular vesicles as drug delivery systems for triple negative breast cancer (TNBC) therapy, *J. Neuroimmune Pharmacol.*, 2020, **15**, 487–500.

213 R. Kanchanapally, *et al.*, Drug-loaded exosomal preparations from different cell types exhibit distinctive loading capability, yield, and antitumor efficacies: a comparative analysis, *Int. J. Nanomed.*, 2019, 531–541.

214 G. Rashid, *et al.*, miRNA expression in PCOS: Unveiling a paradigm shift toward biomarker discovery, *Arch. Gynecol. Obstet.*, 2024, **309**(5), 1707–1723.

215 S. M. El-Daly, R. M. Talaat, M. Braoudaki, R. A. Youness and W. C. Cho, Editorial: Recent breakthroughs in the decoding of circulating nucleic acids and their applications to human diseases, *Front. Mol. Biosci.*, 2023, **10**, 1203495.

216 M. Muskan, *et al.*, Therapeutic potential of RNA-enriched extracellular vesicles: the next generation in RNA delivery via biogenic nanoparticles, *Mol. Ther.*, 2024, 2939–2949.

217 H. I. Kim, *et al.*, Recent advances in extracellular vesicles for therapeutic cargo delivery, *Exp. Mol. Med.*, 2024, **56**(4), 836–849.

218 X. Wang, *et al.*, Exosomes serve as nanoparticles to deliver anti-miR-214 to reverse chemoresistance to cisplatin in gastric cancer, *Mol. Ther.*, 2018, **26**(3), 774–783.

219 H. Huang, *et al.*, MicroRNA profiling of exosomes derived from red blood cell units: implications in transfusion-related immunomodulation, *BioMed Res. Int.*, 2019, **2019**(1), 2045915.

220 G. Liang, *et al.*, Engineered exosome-mediated delivery of functionally active miR-26a and its enhanced suppression effect in HepG2 cells, *Int. J. Nanomed.*, 2018, 585–599.

221 M. Pirisinu, *et al.*, Extracellular vesicles as natural therapeutic agents and innate drug delivery systems for cancer treatment: Recent advances, current obstacles, and challenges for clinical translation, in *Seminars in Cancer Biology*, Elsevier, 2022.

222 Z. Belhadj, *et al.*, A combined "eat me/don't eat me" strategy based on extracellular vesicles for anticancer nanomedicine, *J. Extracell. Vesicles*, 2020, **9**(1), 1806444.

223 T. L. Whiteside, Therapeutic targeting of oncogenic KRAS in pancreatic cancer by engineered exosomes, *Transl. Cancer Res.*, 2017, **6**(Suppl 9), S1406–S1408.

224 S. Nag, *et al.*, Nanomaterials-assisted photothermal therapy for breast cancer: State-of-the-art advances and future perspectives, *Photodiagnosis Photodyn. Ther.*, 2024, **45**, 103959.

225 M. Chehelgerdi, *et al.*, Progressing nanotechnology to improve targeted cancer treatment: overcoming hurdles in its clinical implementation, *Mol. Cancer*, 2023, **22**(1), 169.

226 Y. Cao, *et al.*, Engineered exosome-mediated near-infrared-II region V2C quantum dot delivery for nucleus-target low-temperature photothermal therapy, *ACS Nano*, 2019, **13**(2), 1499–1510.

227 S. Pan, *et al.*, Passion fruit-like exosome-PMA/Au-BSA@ Ce6 nanovehicles for real-time fluorescence imaging and enhanced targeted photodynamic therapy with deep penetration and superior retention behavior in tumor, *Biomaterials*, 2020, **230**, 119606.



228 M. Abdel-Latif and R. A. Youness, Why natural killer cells in triple negative breast cancer?, *World J. Clin. Oncol.*, 2020, **11**(7), 464.

229 M. Abdel-Latif, *et al.*, MALAT-1/p53/miR-155/miR-146a ceRNA circuit tuned by methoxylated quercitin glycoside alters immunogenic and oncogenic profiles of breast cancer, *Mol. Cell. Biochem.*, 2022, **477**(4), 1281–1293.

230 N. N. Parayath, S. Padmakumar and M. M. Amiji, Extracellular vesicle-mediated nucleic acid transfer and reprogramming in the tumor microenvironment, *Cancer Lett.*, 2020, **482**, 33–43.

231 W. Fu, *et al.*, CAR exosomes derived from effector CAR-T cells have potent antitumour effects and low toxicity, *Nat. Commun.*, 2019, **10**(1), 4355.

232 X. Shi, *et al.*, Genetically engineered cell-derived nanoparticles for targeted breast cancer immunotherapy, *Mol. Ther.*, 2020, **28**(2), 536–547.

233 Q. Cheng, *et al.*, Reprogramming exosomes as nanoscale controllers of cellular immunity, *J. Am. Chem. Society*, 2018, **140**(48), 16413–16417.

234 M. Wu, *et al.*, Extracellular vesicles: Emerging anti-cancer drugs and advanced functionalization platforms for cancer therapy, *Drug Deliv.*, 2022, **29**(1), 2513–2538.

235 L. Zhu, *et al.*, Folate-engineered microvesicles for enhanced target and synergistic therapy toward breast cancer, *ACS Appl. Mater. Interfaces*, 2017, **9**(6), 5100–5108.

236 G. Liang, *et al.*, Engineered exosomes for targeted co-delivery of miR-21 inhibitor and chemotherapeutics to reverse drug resistance in colon cancer, *J. Nanobiotechnol.*, 2020, **18**(1), 1–15.

237 J. Wang, *et al.*, Designer exosomes for active targeted chemo-photothermal synergistic tumor therapy, *Adv. Funct. Mater.*, 2018, **28**(18), 1707360.

238 W. Zhou, *et al.*, Pancreatic cancer-targeting exosomes for enhancing immunotherapy and reprogramming tumor microenvironment, *Biomaterials*, 2021, **268**, 120546.

239 L. Cheng, *et al.*, Gene-engineered exosomes-thermosensitive liposomes hybrid nanovesicles by the blockade of CD47 signal for combined photothermal therapy and cancer immunotherapy, *Biomaterials*, 2021, **275**, 120964.

240 G. Wang, *et al.*, Cocktail strategy based on NK cell-derived exosomes and their biomimetic nanoparticles for dual tumor therapy, *Cancers*, 2019, **11**(10), 1560.

241 Y. Wang, *et al.*, Delivering antisense oligonucleotides across the blood-brain barrier by tumor cell-derived small apoptotic bodies, *Adv. Sci.*, 2021, **8**(13), 2004929.

242 H. Cheng, *et al.*, Chimeric peptide engineered exosomes for dual-stage light guided plasma membrane and nucleus targeted photodynamic therapy, *Biomaterials*, 2019, **211**, 14–24.

243 S. Li, *et al.*, Engineering extracellular vesicles for cancer therapy: recent advances and challenges in clinical translation, *Biomater. Sci.*, 2020, **8**(24), 6978–6991.

244 R. Kalluri, The biology and function of exosomes in cancer, *J. Clin. Investig.*, 2016, **126**(4), 1208–1215.

245 J. Palma, *et al.*, MicroRNAs are exported from malignant cells in customized particles, *Nucleic Acids Res.*, 2012, **40**(18), 9125–9138.

246 S. Y. Ko, *et al.*, The glycoprotein CD147 defines miRNA-enriched extracellular vesicles that derive from cancer cells, *J. Extracell. Vesicles*, 2023, **12**(4), 12318.

247 L. T. Vu, *et al.*, microRNA exchange via extracellular vesicles in cancer, *Cell Prolif.*, 2020, **53**(11), e12877.

248 H. Sork, *et al.*, Heterogeneity and interplay of the extracellular vesicle small RNA transcriptome and proteome, *Sci. Rep.*, 2018, **8**(1), 10813.

249 B. N. Hannafon, *et al.*, Plasma exosome microRNAs are indicative of breast cancer, *Breast Cancer Res.*, 2016, **18**, 1–14.

250 A. Turchinovich, L. Weiz and B. Burwinkel, Extracellular miRNAs: the mystery of their origin and function, *Trends Biochem. Sci.*, 2012, **37**(11), 460–465.

251 L. Pigati, *et al.*, Selective Release of MicroRNA Species from Normal and Malignant Mammary Epithelial Cells, *PLoS One*, 2010, **5**(10), e13515.

252 R. Crescitelli, *et al.*, Subpopulations of extracellular vesicles from human metastatic melanoma tissue identified by quantitative proteomics after optimized isolation, *J. Extracell. Vesicles*, 2020, **9**(1), 1722433.

253 T. Vagner, *et al.*, Protein composition reflects extracellular vesicle heterogeneity, *Proteomics*, 2019, **19**(8), 1800167.

254 E. Willms, *et al.*, Cells release subpopulations of exosomes with distinct molecular and biological properties, *Sci. Rep.*, 2016, **6**(1), 22519.

255 W. Meng, *et al.*, Prospects and challenges of extracellular vesicle-based drug delivery system: considering cell source, *Drug Deliv.*, 2020, **27**(1), 585–598.

256 X. Luan, *et al.*, Engineering exosomes as refined biological nanoplatforms for drug delivery, *Acta Pharmacol. Sin.*, 2017, **38**(6), 754–763.

257 P. Vader, *et al.*, Extracellular vesicles for drug delivery, *Adv. Drug Deliv. Rev.*, 2016, **106**, 148–156.

258 R. van der Meel, *et al.*, Extracellular vesicles as drug delivery systems: lessons from the liposome field, *J. Contr. Release*, 2014, **195**, 72–85.

259 M. J. Haney, *et al.*, Exosomes as drug delivery vehicles for Parkinson's disease therapy, *J. Contr. Release*, 2015, **207**, 18–30.

260 M. S. Kim, *et al.*, Development of exosome-encapsulated paclitaxel to overcome MDR in cancer cells, *Nanomed. Nanotechnol. Biol. Med.*, 2016, **12**(3), 655–664.

261 A. Jeyaram and S. M. Jay, Preservation and storage stability of extracellular vesicles for therapeutic applications, *AAPS J.*, 2018, **20**, 1–7.

262 J. H. Crowe, *et al.*, Stabilization of dry phospholipid bilayers and proteins by sugars, *Biochem. J.*, 1987, **242**(1), 1.

263 K. A. Murray and M. I. Gibson, Chemical approaches to cryopreservation, *Nat. Rev. Chem.*, 2022, **6**(8), 579–593.

264 S. Bosch, *et al.*, Trehalose prevents aggregation of exosomes and cryodamage, *Sci. Rep.*, 2016, **6**(1), 36162.

265 C. Charoenviriyakul, *et al.*, Erratum to 'Preservation of exosomes at room temperature using lyophilization', *Int. J.*



*Pharm.*, 2018, **553**, 1–7. International Journal of Pharmaceutics, 2019. **559**: pp. 427–428.

266 R. Stranska, *et al.*, Comparison of membrane affinity-based method with size-exclusion chromatography for isolation of exosome-like vesicles from human plasma, *J. Transl. Med.*, 2018, **16**, 1–9.

267 G. Vergauwen, *et al.*, Confounding factors of ultrafiltration and protein analysis in extracellular vesicle research, *Sci. Rep.*, 2017, **7**(1), 1–12.

268 Q. Lin, *et al.*, Exosome-like nanoplatform modified with targeting ligand improves anti-cancer and anti-inflammation effects of imperialine, *J. Contr. Release*, 2019, **311**, 104–116.

269 K. Tang, *et al.*, Delivery of chemotherapeutic drugs in tumour cell-derived microparticles, *Nat. Commun.*, 2012, **3**(1), 1282.

270 C. Admyre, *et al.*, Direct exosome stimulation of peripheral human T cells detected by ELISPOT, *Eur. J. Immunol.*, 2006, **36**(7), 1772–1781.

271 E. Koh, *et al.*, Exosome-SIRP $\alpha$ , a CD47 blockade increases cancer cell phagocytosis, *Biomaterials*, 2017, **121**, 121–129.

272 D. Wang, *et al.*, Engineered cell-derived microparticles Bi2Se3/DOX@ MPs for imaging guided synergistic photothermal/low-dose chemotherapy of cancer, *Adv. Sci.*, 2020, **7**(3), 1901293.

