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A review on phytochemical constituents and pharmacological potential of *Calotropis procera*

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Calotropis procera is locally known as Aak or Madar in Hindi, milk weed in English and belongs to the family Apocynaceae and subfamily Asclepiadoideae. Although a wasteland plant, it is of sacred use as its flowers are offered for worshipping Lord Shiva, a Hindu God. Tribes all over the world use the plant in treatment of various diseases like snake bite, body pain, asthma, epilepsy, cancer, sexual disorders, skin diseases and many more. This plant contains various phytoconstituents such as flavonoids, terpenoids, cardenolides, steroids oxypregnanes etc. Though literature searches reveal many reviews about ethnomedicinal uses, chemical composition and pharmacological activities, no recent papers are available that provide an overview of the therapeutic potential and toxicity of Calotropis procera. Hence, the insight of this review is to provide a systemic summary of phytochemistry, pharmacology, toxicology and therapeutic potential of Calotropis procera and to highlight the gaps in the knowledge so as to offer inspiration for future research.

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1. Introduction

Calotropis belongs to the Apocynaceae family, which is commonly known as milkweed or Aak. Plants of this genus are known as milkweeds due to the exudation of white and sticky latex from different plant parts. Genus Calotropis has two common species viz. Calotropis procera (Rakta arka) and Calotropis gigantea (Sweata arka), which are described as possessing vital pharmacological properties in Ayurvedic toxicology and therapeutics. Other species are C. sussuela and C. acia.

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Calotropis procera (Aiton) W. T. Aiton is an erect, soft wooded, evergreen perennial shrub and commonly known as 'Sodom apple' or 'Madar shrub'. In Bengali, it is known as 'Akanda' and in Hindi as 'Aak'. It manifests its wide utilization in Indian, Arabic and Sudanese traditional medicinal systems for healing global range of diseases.

The Dangas tribe in Gujarat, Singhum tribe in Bihar, tribes of Ghatigaon forest in Gwalior, tribes of Andhra Pradesh have been using this plant in the treatment of various disorders such as ear pain, cough, fever, abdominal pain, dysentery and elephantiasis.

Calotropis procera is more toxic than Calotropis gigantea and assumed to be even more poisonous than cobra venom. It is interesting that the cobra and other poisonous snakes cannot



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even bear its smell; hence snake charmers of Bengal use this plant for controlling or taming cobras.5

Earlier reviews6-16 have discussed on phytochemistry, ethnobotany and pharmacological potential of Calotropis procera. Review on Calotropis species17-20 comparing procera and gigantea have deliberated their therapeutic importance. The present review summarizes the phytochemistry, pharmacology, commercial aspects, traditional medicinal uses, toxicology and recent studies on Calotropis procera. The future scope of Calotropis procera has also been affirmed with a view to establish its multiple biological activities and mode of action.

Unique properties of Calotropis procera

2.1 Toxicity

C. procera finds its widespread distribution over many regions of the globe. What makes its phytochemistry interesting is the exudation of milky and toxic latex from all the plant parts. The



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latex is referred to as vegetable mercury as it shows mercury like effects on human body.21

Every part of this plant is toxic, but stem (latex) and roots are more poisonous than leaves. The leaves of this plant have three toxic glycosides calotropin, calotoxin and uscharin, whereas its latex contains calotropin, calotoxin and calactin, which are caustic and considered poisonous in nature. Besides this, the concentration of calactin, which is a toxic glycoside, gets increased as defense mechanism on encounter of grasshopper or insect attack and this is the rationale behind the plant not being consumed by cattles or other grazing animals.22 Other than this, osmotin, a laticifer protein purified from latex also provides protection to plant against phytopathogens.²³ Its milk is irritant, neurotoxic and has anticholinergic activity, which causes toxicity and fatal complications. Madar juice and latex has bitter taste and a burning pain which causes salivation, stomatitis, vomiting, diarrhoea, dilated pupils, titanic convulsion, collapse and death. The fatal period varies from half an hour to eight hours.24 If latex enters into the eye, it causes kerato-conjunctivitis, corneal edema and dimness of vision without any pain.25-27 Some cases showed permanent endothelial cell damage, which was evident after three weeks.^{5,28} C. procera was found toxic at the dose of 100 mg kg⁻¹ to chick embryo. Its toxicity caused hepatocellular degeneration in liver, brain congestion, dilation of central veins, sinusoids, underdeveloped lung and kidneys.29 Hence, bearing in mind the toxic effects of certain extracts and glycosides, further studies should be focused to explain toxicity and safe use of C. procera.

2.2 Ability to survive under extreme climatic conditions

Another interesting aspect of this plant is its ability to tolerate adverse environmental conditions like scarcity of water, arid environment or any kind of harsh climate. To understand this, Akhkha30 studied the effect of stress caused due to water scarcity and found that photosynthetic machinery remained uninfluenced, infact rate of photosynthesis gets raised at mild water



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Table 1 Ethnomedicinal applications of C. procera

Plant part	Disease	Preparation/administration	References
Root/root	Amoebic dysentery	Paste with/without opium taken orally	44-46
bark	Cholera	Powder orally taken or paste along with black pepper and ginger juice	44
	Dysentery	Powder orally taken	47
	Elephantiasis and hydrocele	Paste mixed with fermented rice water applied on the affected area	48-50
	Epilepsy	Grounded with goat milk and used as nasal drops	46
	Indigestion	Powder orally taken	47
	Jaundice	Taken with rice in grounded form	51
	Neuritis	Orally administered with cow butter	46
	Rheumatism Snake bite	Powder taken with milk and sugar Powder orally taken. Paste applied on wounds and internally	48 47 and 52
	Cuidor and insect hite	taken with ghee	40
	Spider and insect bite	Powdered and taken with vinegar	48
atox	Syphilis	Root bark powder taken orally	46
Latex	Boils Black scar on the face	Applied externally	46
	Ascites	Applied along with turmeric paste Applied externally	44 47
	Liver and spleen disorder	Taken after dilution	47 47
	-	Applied on the affected area	47
	Leprosy Migraine	Applied on the affected side vein of forehead	47
	Piles (haemorrhoids)	Applied externally	44
	Dog/jackal bite	Applied on wound	44 and 48
	Ring worm	Applied externally	44 and 48
	Scabies	Applied externally Applied externally	46
	Snake bite	Applied on wounds or taken orally (20–30 drops for adults and	46
	Shake bite	15–20 for infants)	
	Carl III a Language and Ladama	Five drops with 50 drops of distilled water injected hypodermally	46
	Syphilis, leprosy and odema	Applied externally with sesame oil	48 and 50
	Tooth ache	Applied on affected tooth	48 and 50
c	Vertigo	Applied on affected parts	53
Leaf	Cold, cough, asthma and bronchitis	Warmed along with ghee and bandaged on the chest of infants	44
	Calculus, liver and spleen disorder	Powder taken orally	48
	Ear ache or ear troubles	Juice along with fermented boiled rice water used as ear drops	50
	Eczema and skin eruptions Enlargement of abdominal viscera and	Applied externally along with turmeric and sesame oil Oral administration of powder	48, 50 and 5 48 and 51
	spleen		
	Gonorrhoea	Decoction used for washing and taken orally	51
	Inflammatory swellings	Covered on affected part after warming	51
	Joint pain	Powder taken	47
	Malaria and intermittent fever	Oral administration of fresh juice	46, 49 and 5
	Body pain	Paste applied after warming	51
	Paralysis and sciatica	Massaged after preparing decoction with sesame oil	47
	Snake bite	Oral administration of fresh juice	50
-1	Ulcers, wounds, sores	Powder orally administered or external application	47, 49 and 5
Flowers	Health tonic	Oral administration of powder	47
	Cough	Burnt to produce ash, then taken with honey	44
	Rat bite	Oral administration of powder	47 and 49
	Dog/jackal bite (rabies)	Seven tepals chewed with fine rice on seventh day of biting, continued for seven days decreasing one tepal everyday	44
	Feet pain	Decoction used for fomentation	46
	Epilepsy	Oral administration of paste with black pepper	46
	Asthma and bronchitis	Fruit taken with jaggery	3
	Liver and spleen disorder	Administered along with milk	46
₹ruit	Eye disorder	Decanted ash water applied on eye lids	44
	Anemia	Mixed with same quantity of red chilli, mineral salt and taken with milk.	46
Whole plant	Rheumatic pain and hyperacidity	Paste directly taken	44
Young wigs	Purgative	Juice taken	54

Table 2 Brief summary of the pharmacological properties

S. no.	Pharmacological activities	Parts/extracts/possible chemical constituents	References
1	Wound healing potential	Latex: aqueous extract	67
		Latex	68
		Bark: ethanolic extract	69
		Leaves: aqueous extract	70
		Bark: aqueous extract	71
	Anticoccidial activity	Dried leaves powder	72
	Toxicity activity	Leaves: aqueous extract	73 and 74
	J	Leaves and stem bark extracts	75
		Leaves and stem: ethanolic extract	29
		Leaves: ethanolic extract	79
	Biopesticidal/insecticidal activity	Leaves: extract	80 and 81
		Leaves: methanolic extract, latex protein fraction, flavonoids (quercetin-3- <i>O</i> -rutinoside)	35
;	Antimycoplasmal activity	Leaves: acetone extract	82
,	Hepatoprotective activity	Root bark: methanolic extract	83
	• •	Flowers: hydroethanolic extract	84
		Roots: chloroform extract	85
	Antimicrobial/antibacterial activity	Leaves: methanolic extract, flavonoids (quercetin-3- <i>O</i> -rutinoside)	86
	,	Leaves and latex: ethanol, aqueous, and chloroform extract	87
		Leaves and stem: aqueous, ethanolic, methanolic extract	88 and 89
		Endophytic fungi of <i>C. procera</i>	90
		Seeds: chloroform extract	91
		Root: pet. ether, methanolic extract	92
		Flowers: ethanolic extract	93
		Latex	94
		Leaves: methanolic extract	95
		Leaves, flower, root bark: ethanolic extract	96
		Leaves and latex: aqueous, ethanolic extract	97 and 98
		Leaves: aqueous, methanolic extract	99
		Latex: aqueous extract	78
	Central nervous system activity	Latex proteins	100
	Antioxidant activity	Leaves, flower, fruit, latex	100
	Antioxidant activity	Leaves: aqueous, methanolic extract, quercetin and its derivatives	76
		Leaves: aqueous and methanolic extract	102
		Leaves, flowers and fruits: methanolic extract	
		Bark: ethanolic extract	103
0	Antinopiaantivo activity		69
0	Antinociceptive activity	Latex protein	104
1	Antihelmintic activity	Flowers: crude powder, aqueous and methanolic extract	105
		Latex: fresh, dried aqueous extract	106 and
_	A + i i G	Perulatura	107
2	Antiinflammatory activity	Dry latex	108 and
		Completed all the Company of the Lorentz Labella of the Company	109
		Stem bark: chloroform and hydro-alcoholic extract	110
		Latex: hexane, dichloromethane, ethyl acetate, <i>n</i> -butanol and aqueous extract	77
		Latex: pet. ether, acetone, methanol extract	111
		Leaves: aqueous extract	112
_		Flowers: ethanolic extract	93
3	Antidiarroheal activity	Bark: Arkamula Tvarka (Ayurvedic preparation)	45
		Latex	113
4	Antifungal activity	Aqueous bark extract	114
		Leaves: aqueous, methanol, acetone and ethanol extract	115
		Root bark	116
	Antimycotic activity against dermatophytes	Latex	117
	Antimycofloral activity (fungi in wheat)	Fresh latex	118
5	Larvicidal activity	Crude latex and ethanolic extract of leaf	119
		Leaves: ethanolic extract	120
		Leaves: aqueous extract	121
		Flower, young bud, mature leaves and stems: ethanolic extract	122
		Flowers: aqueous extract	123
6	Tobacco mosaic virus (TMV) inhibitor	Latex	124
	activity		
	Antifertility activity		

Table 2 (Contd.)

10.	Pharmacological activities	Parts/extracts/possible chemical constituents	References
		Leaves: ethanolic extract	79
		Roots (calotropin)	59
	Abortifacient activity	Latex	126
	Antisperm activity	Root: chloroform extract	127
	Oestrogenic/antiovulatory activity	Roots: ethanolic and aqueous extract	128
18	Plasma clotting activity	Protein fraction isolated from fresh latex	129
19	Antiplasmodial activity	Different plant parts: ethyl acetate, ethanolic and acetone extract	130
		Leaves extract	131
20	Antipyretic activity	Dry latex: aqueous extract	132
		Flowers: ethanolic extract	93
21	Antiasthmatic activity	Flowers	133
22	Anticonvulsant activity	Root extracts	134
23	Cytotoxic activity	Root (2"-oxovoruscharin)	62
		Laticifer proteins (LP) recovered from latex	135
		Root: methanolic, aqueous, ethyl acetate, hexane extracts	136
		Plant: methanolic extract	137
		Stems: uzarigenin	138
		Root bark: calotroprocerol A	139
		Root: alcoholic, hydro-aqueous and aqueous	140
		Leaf: ethanolic extract	149
24	Analgesic activity	Flowers: Ethanolic extract	93
25.	Antihyperglycemic activity	Leaves: pet ether, methanol and aqueous extracts	141
26	Antiarthritis activity	Latex	142
	•	Protein sub fraction of latex	143
27	Antimolluscicidal activity	Latex: 95% aqueous ethanol (uscharin)	144
28	Antitermites activity	Latex	145
29	Antimigraine activity	Dried terminal leaves	146
30	Anti-ulcer activity	Root: chloroform extract	147
	,	Plant: 50% ethanolic extract	148
		Leaf: ethanolic extract	149
		Stem bark: chloroform and hydroalcoholic extract	110
31	Spasmolytic activity	Plant: aqueous extract	150
32	Allelopathic activity	Leaves: aqueous extract	151
33	Anti-keloidal activity	Latex	68
34	Anti-hyperbilirubinemic activity	Leaves: aqueous extract	70
35	Antiapoptotic activity	Latex	152

regime (50%) which can be considered as a compensatory mechanism. Further Ramadana $et~al.^{31}$ studied the influence of light and irrigation on cumulation of β -sitosterol in C.~procera. They hypothesized that β -sitosterol biosynthesis pathway supported the plant to bear drought and light intensity stress.

2.3 Commercial prospective

2.3.1 As biofuel. *C. procera* is rich in hydrocarbons and contains biologically degradable materials similar to that found in other agricultural crops. Traore³² conducted fermentation experiments and found that it is a good substrate for biogas synthesis. Barbosa *et al.*³³ found that oil composition of its seeds varies from 19.7 to 24.0% which proves its future potential as biodiesel, specially in those areas where people rely mainly on wood as source of energy production.

2.3.2 As biopesticide. Laticifer proteins (LP) from *Calotropis procera* were assayed for insecticidal activity against different crop pests to assess the biological role of latex. Diets

containing 4% latex led to decreased weight gain ($ED_{50} = 3.07\%$) and affected survival ($LD_{50} = 4.61\%$) of third instars of *Ceratitis capitata*.³⁴ The crude flavonoid fraction (Cf), the latex protein fraction (LP) and the leaf methanolic extract showed significant insecticidal activity.³⁵ These studies suggest that it can be developed as natural biopesticidal agent.

2.4 Industrial prospective

2.4.1 Cheese making agent. In West Africa, crude aqueous extract of *C. procera* is used as milk clotting enzyme in traditional method of cheese production.³⁶ It displayed an optimum activity at a temperature of 75 °C, which is essential for cheese production.³⁷ Calotropain enzyme found in the plant is more efficient than papain, ficin and bromelin, moreover it can lead to milk coagulation, digestion of meat, casein and gelatin.^{38,39} These studies supported its traditional use as cheese making agent.

2.4.2 As surfactant. *C. procera* milk latex was used as a surfactant for facile synthesis of Eu³⁺ activated La(OH)₃ and

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Table 3 Summary of cytotoxic studies of C. procera

C. procera: plant part/chemical constituent	Cancer cell lines/model	Method of analysis/assay	Mechanism of action/investigation Observation	Observation	References
Uscharin and its derivatives 2"-Oxovoruscharin and its derivatives	Lung cancer (A549) Two glioblastoma (Hs683, U373) and two colon cancer (HCT-15 and LoVo)	MTT colorimetric assay, intraperitoneal (ip) injection- related toxicity	Na ⁺ /K ⁺ -ATPase inhibition activity	Cardenolides derived from 2"- oxovoruscharin exhibited significant <i>in vitro</i> antitumor activity and high <i>in vivo</i> tolerance	62
Laticifer proteins (LP) recovered from latex	HL60 (promoyelocytic leukemia), HCT-8 (colon), MDA-MB- 435(breast), SF-295(brain)	3-(4,5-Dimethyl-2-thiazolyl)-2,5- diphenyl-2 <i>H</i> -tetrazolium bromide MTT	LP is a target for DNA topoisomerase I triggering apoptosis in cancer cell lines	IC ₅₀ values for LP ranged from 0.42 135 to 1.36 μg mL ⁻¹ to SF-295, MDA-MB-435 respectively	2 135
Root: methanolic, aqueous, ethyl acetate, hexane extracts (1, 5, 10, 25 $\mu g \ m L^{-1}$)		Tetrazolium bromide (MTT), colorimetty	Treatment initiated apoptotic mechanism by blocking the cell cycle at S-phase and thus preventing cells from entering proliferative (G2/M) phase	Ethyl acetate extract showed strongest cytotoxic effect	136
Plant: methanolic extract (0, 5, 10, 20 and 40 $\mu g \; m L^{-1})$	Plant: methanolic extract (0, 5, 10, Human skin melanoma cells (SK-20 and 40 $\mu g \; m L^{-1})$ MEL-2)	Annexin-V FITC flow cytometry method, MTS assay	t te G2/M cell hase	At 40 µg mL ⁻¹ late apoptotic cell percentage was increased up to 80%. <i>C. procera</i> exerted cytotoxic potential	137
5-Hydroxy-3,7-dimethoxyflavone-4- O-6-glucopyranoside; uzarigenin; β- anhydroepidigitoxigenin; 2β,19- epoxy-3β,14β-dihydroxy-19- methoxy-5α-card-20(22)-enolide; β- anhydroepidigitoxigenin-3β-O- glucopyranoside	5-Hydroxy-3,7-dimethoxyflavone-4- HT 29, HepG2 (human cancer cell O-β-glucopyranoside; uzarigenin; β- lines), NIH-3T3 (mouse fibroblast anhydroepidigitoxigenin; 2β,19- cell line) epoxy-3β,14β-dihydroxy-19- methoxy-5 α -card-20(22)-enolide; β- anhydroepidigitoxigenin-3β- O - glucopyranoside	CellTiter-Blue® cell viability assay		Uzarigenin showed moderate cytotoxicity	138
Calotroprocerol A; calotroproceryl acetate A; calotroprocerone A, B; pseudo-taraxasterol acetate; taraxasterol; calotropursenyl acetate B; stigmasterol; (E)-octadec. 7-enoic acid	A549 non-small cell lung cancer 3-(4,5-Dimet (NSCLC), the U373 glioblastoma diphenyltetr (GBM) and the PC-3 prostate cancer (MTT) assay cell lines	3-(4,5-Dimethylthiazol-2,yl)- 2,5- diphenyltetrazolium bromide (MTT) assay	Growth inhibition action	Calotroprocerol A exhibited in vitro 139 growth inhibitory activity in all the three cancer cell lines with effects comparable to those of cisplatin and carboplatin	o 139
Calotroposide H; calotroposide I; calotroposide J; calotroposide K; Calotroposide L; calotroposide M; calotroposide N	A549 non-small cell lung cancer (NSCLC), U373 glioblastoma (GBM), and PC-3 prostate cancer cell lines	MTT colorimetric assay	Calotroposide K and M exhibited subnanomolar growth inhibition activity with IC ₅₀ ranging from 0.5 to 0.7 μM against U373 glioblastoma (GBM) and PC-3 prostate cancer cell lines	C. procera exhibited cytotoxic potential	153
Calotroposide S	PC-3 prostate cancer, A549 non-small cell lung cancer (NSCLC), and U373 glioblastoma (GBM) cell lines	MTT colorimetric assay	Calotroposide S showed potent anti <i>C. procera</i> exerted anti-proliferative 154 proliferative activity	C. procera exerted anti-proliferativ activity	e 154
Latex: hexane, chloroform, ethyl acetate and aqueous extract. Calactin; 15β-hydroxy calactin;	A549 (lung) and hela (cervix) cancer MTT colorimetric assay cell lines using cisplatin as a positive control	MTT colorimetric assay	Growth inhibition action	Highest cytotoxic activity was displayed by chloroform extract. Amongst isolated compounds,	65

Table 3 (Contd.)

La₂O₃ nanophosphors through green mediated hydrothermal route. The latex reflected good capping potency for controlling the morphology and phase of the nanophosphor.⁴⁰ Hence its latex can be a good source of natural surfactant.

- **2.4.3 As corrosion inhibitor.** Extract of *C. procera* was studied for its corrosion inhibition action by weight loss, electrochemical, SEM and UV methods, significant corrosion inhibitive effect in sulphuric acid medium on mild steel was observed. ⁴¹ Hence, it can be used as green corrosion inhibitor.
- **2.4.4 As dehairing agent of leather.** Latex peptidases of *C. procera* when assayed against skin representative substrates, revealed complete dehairing process, while no changes in leather structure were observed. Thus, it can be an appropriate environment friendly dehairing agent as compared to toxic sodium sulphite treatment for tanneries.⁴²

3. Ethnomedicinal uses

An insight into Ayurveda, Unani and folk uses of different parts of *C. procera* and *C. gigantea* to cure various ailments was compiled by Misra *et al.*⁴³ Ethnomedicinal uses of plant parts of *C. procera* in curing various diseases have been summarized in Table 1.

4. Major milestone of *Calotropis* phytochemistry

Phytochemistry of *Calotropis procera* has always attracted the attention of researchers because despite its toxicity, it employs wide applications in traditional medicinal system till date. Dating back to 1936, Hesse *et al.*⁵⁵ identified calotropin as the first compound from this plant. Further Hesse and his coworkers^{56,57} isolated heart poisons or cardiac glycosides namely calotropin, calotoxin, calactin, uscharin, voruscharin and uscharidin.⁵⁸ Root powder of this plant is used in tribes to induce abortion in women and as an uterotonic since ancient period. Later it was found that it was due to the compound calotropin. Gupta *et al.*⁵⁹ administered calotropin to gerbils and rabbits and observed reduction in spermatids count by 65% and 94% respectively.

In 1955, Rajagopalan *et al.*⁶⁰ identified chemical constituents of seed *viz.* coroglaucigenin, corotoxigenin and frugoside (cardenolides). Later Bruschweiler *et al.*⁶¹ identified three additional cardenolides *viz.* uzarigenin, syriogenin and procerosid. A novel cardenolide, 2ⁿ-oxovoruscharin was isolated from the root bark by Quaquebeke *et al.*⁶² and modified into its semi-synthetic derivative, *i.e.*, UNBS1450. Akhtar and Malik⁶³ isolated a new cardenolide named proceragenin from the hexane-insoluble fraction of *C. procera*.

A fascinating feature of the plant is its potential to curb Alzheimer's disease (AD), the most predominant root cause of dementia, a neurodegenerative disease. Its dried latex showed attenuation of β -amyloid deposition in mouse brain and cerebral protective activities. Hence, it is imperative to evaluate the mechanism of metabolites, so that it can lead to promising direction to search new scaffolds for AD treatment. In 2015.

Table 4 Summary of in vivo studies of wound healing potential of C. procera

Model	C. procera extract/dose/ duration	Negative control	Investigation	Result	References
Guinea pigs	20 mL of 1.0% sterile solution of the latex twice daily for 7 days	Excision wounds	Wounds exhibited marked dryness, no visual sign of inflammation	Significant prohealing property	67
Male albino- Wistar rats	Ethanolic extract of bark (50 mg per wound)	Incision and excision wounds	Extract demonstrated wound healing effect by accelerating wound closure and epithelialization	Excellent dermal wound healing potential	69
Wistar rats	Aqueous extract of <i>C.</i> procera (25 mg and 50 mg kg ⁻¹)	Incision and excision wounds	Significant ($P < 0.05$) increase in breaking strength and percentage wound contractions with decreased epithelization period was observed	Significant wound healing property	70

Mohamed et al. isolated three non-glycosidic cardenolides namely calactoprocin, procegenin A and procegenin B from the latex.65

A patent claimed that polar extract of C. procera showed antiulcerative colitis activity in dose-dependent manner in a subject mammal and was found to be more effective than the standard drug Prednisolone.66

Pharmacology

Over the last many years, researchers have carried out numerable pharmacological activities, which are summarized in Table 2.

The details enumerated in the Table 2 is indicative of the fact that the different plant parts demonstrate large number of

Table 5 Summary of in vivo anti-inflammatory potential of C. procera

Model	C. procera extract/dose/ duration	Negative control	Investigation	Result	References
Male albino rats and albino guinea pigs	50 mg, 200 mg 500 mg and 1 g kg ⁻¹ dry latex	Carrageenan-induced oedema test, cotton pellet granuloma and vascular permeability etc.	Dry latex suppressed fluid exudation, due to its influence on vascular permeability and also delayed the onset and intensity of UV induced erythema	Significant anti- inflammatory potential	108
Male albino rats	Dry latex	Carrageenin and formalin-induced pedal oedema test	At dose 5 mg per rat, showed 71% inhibition in the case of the carrageenin-induced oedema ($P < 0.005$) and 32% inhibition for the formalin-induced oedema ($P < 0.05$). At higher dose (50 mg per rat), 96% and 98%, for carrageenin- and formalin-induced oedema groups respectively	Potent anti- inflam-matory activity	109
Albino rats of either sex	Stem bark: chloroform and hydro-alcoholic extract	Carrageenan-induced paw oedema	Significant reduction in the inflammation at 100, 200 and 400 mg kg ⁻¹ displayed by chloroform extract	Significant anti- inflammatory potential	110
Male Wistar rats	Dry latex: petroleum ether, acetone, methanol and aqueous extracts (50 mg per rat)	Carrageenan induced paw oedema	Maximum anti-inflammatory effect (59% and 53% inhibition) by the aqueous and acetone extracts respectively compared to (63%) inhibition exhibited by phenylbutazone	Latex of <i>C.</i> procera exerted anti- inflammatory property	111
Male Wistar rats	Crude latex: hexane, dichloromethane, ethyl acetate, <i>n</i> -butanol and aqueous fractions (1.0, 5.0 or 10.0 mg kg ⁻¹ and 0.2 mL)	Carrageenan-induced peritonitis	Dichloromethane, ethyl acetate, and aqueous fractions inhibited carrageenan-induced neutrophil migration in rats at the ratios 67%, 56%, and 72%, respectively	Latex of <i>C.</i> procera possess anti- inflammatory property	77

Table 6 Summary of larvicidal potential of C. procera

Vector species	C. procera extract/dose/ duration	Observation	Result	References
Culex quinquefasciatus 3 rd instar larvae	Crude latex and ethanolic extract of leaves	100% larval mortality at 300 ppm concentration of latex and at 1000 ppm concentration of ethanolic leaf extract. LC $_{50}$ values of the latex and ethanolic leaves extract were 57.3 and 388.7 ppm respectively	Crude latex exerted stronger larvicidal potential than ethanolic extract	119
Musca domestica 3 rd instar larvae	Ethanolic extract of leaves (500 mg L^{-1})	100% mortality at 500 ppm. LC ₅₀ value of the extract 282.5 ppm	Leaves exerted insecticidal potential	120
Anopheles arabiensis and Culex quinquefasciatus 2 nd , 3 rd , 4 th instar larvae	Aqueous extract of leaves (1000, 500, 200 ppm)	LC ₅₀ value 273.53, 366.44, 454.99 ppm for 2 nd , 3 rd and 4 th instar larvae	Leaves showed oviposition deterrent, larvicidal and adult emergence activity	121
Anopheles stephansi 3 rd instar larvae	Ethanolic extracts of different parts <i>viz.</i> flower, young bud, mature leaves and stems (100 to 5000 ppm)	Mature leaves extract exhibited 100% mortality at 2000 ppm after 48 hours of incubation	Mature leaves showed high larvicidal activity against tested larvae	122
Culex species 4 th instar	Aqueous extract of flowers (1%, 2.5% and 5%)/24 h	At 1% concentration, the mortality rate was 0%, 60% and 100% and at 2.5% concentration, mortality rate was 20%, 80% and 100% at the end of 1, 3 and 4 days of exposure, and at 5% concentration, 100% mortality was recorded at the end of third day	Flowers exhibited remarkable larvicidal properties against the pupae and late 4 th instar larvae of <i>Culex</i> sp.	123

Table 7 Summary of in vivo and in vitro studies of anthelmintic potential of C. procera

Model	C. procera extract/dose	Compared with drug	Observation	Result	References
In vivo: sheep infected with mixed species of nematodes in vitro: Haemonchus contortus	Crude powder (CP), crude aqueous (CAE) and crude methanolic extracts (CME)	Levamisole	88.4%, 77.8% and 20.9% reduction in egg count percent for CAE, CP and CME respectively	Aqueous extract of <i>C. procera</i> has good anthelmintic potential	105
Earthworms	Aqueous extract of dry latex $(5, 10, 50 \text{ and } 100 \text{ mg mL}^{-1})$ and fresh latex $(1.45, 7.25, 29, 72.5 \text{ and } 145 \text{ mg mL}^{-1})$	Piperazine	At 5 to 10 mg mL $^{-1}$ concentration paralysis at 90 min, at 100 mg mL $^{-1}$ death within 60 min. Fresh latex also showed dose-dependent paralysis	Latex showed wormicidal activity, hence can be used as an anthelmintic agent	106

pharmacological activities. Moreover, maximum number of activities were conducted at extract level, therefore horizons for further research is still bright, wherein the active principle constituents responsible for the activities may be identified. Here some of the very vital biological activites are being discussed in detail.

5.1 Cytotoxic potential

Various phytoconstituents and plant extracts were examined for their *in vitro* anticancer potential on various cancer cell lines, and showed significant cytotoxic activities as summarized in Table 3.

Over past decade, cytotoxic activities of various extracts and chemical constituents of *C. procera* have been carried out. Majority of studies were conducted on various cancer cell line models *in vitro*, except the one conducted using UNBS1450. UNBS1450, a semi-synthesized cardenolide was compared to reference anticancer agents and classic cardenolides in prostate cancer cell line *in vitro* and *in vivo* following s.c. (subcutaneous) and orthotopic prostate cancer cell grafting into mice; it was

Table 8 Summary of in vitro studies of antioxidant potential of C. procera

C. procera part	Extract/dose/duration	Investigation	Result	References
Leaves, fruits, flowers and latex	Methanolic solution of dried extract	DPPH radical scavenging assay	Leaves exhibited maximum DPPH radical scavenging activity with $IC_{50} = 0.18$ mg mL ⁻¹ , whereas latex showed minimum activity with $IC_{50} = 0.42$ mg mL ⁻¹	101
Leaves	Aqueous and methanolic extract $(1, 5, 10, 50, 100 \text{ and } 500 \mu\text{g mL}^{-1})$	DPPH radical scavenging assay	IC ₅₀ of the methanol extract was 110.25 μ g mL ⁻¹ , the aqueous extract showed mild antioxidant activity	102
Leaves	2–100 mg mL ⁻¹ for quercetin in methanol and 20–100 mg mL ⁻¹ for AME and quercetin derivatives with different methoxy substitution	DPPH radical scavenging assay	Varying degrees of antioxidant activity was exerted by quercetin derivatives, but quercetin was found to be most active	76
Leaves, flowers and fruits	Methanolic extracts of the samples of different concentrations (100–1000 ppm)	DPPH radical scavenging assay	IC_{50} values in leaves, fruits and flowers were 16.08, 16.06 and 10.31 μg mL ⁻¹ respectively, showing strong antioxidant activity of <i>C.</i> procera	103

Table 9 Summary of in vitro schizontocidal activity of C. procera

Model	C. procera extract/dose	Investigation	Result	References
Chloroquine sensitive strain, MRC 20 and a chloroquine resistant strain, MRC 76 of Plasmodium falciparum	Ethyl acetate, acetone, methanol fractions of flower, bud, root: (62–125 mg mL ⁻¹)	Percentage inhibition varied from 7.51 to 61.38% between the various fractions against MRC 20 and for MRC 76, percentage inhibition varied from 3.437 to 41.08% between the various fractions	At the lower dose range, the root extracts of <i>C. procera</i> found to be the most effective for both <i>P. falciparum</i> MRC 20 and MRC 76. Hence, <i>C. procera</i> exerted antiplasmodial potential	130

Table 10 Summary of in vivo hepatoprotective potential of C. procera

Model	C. procera extract/dose	Negative control	Investigation	Result	References
Albino rats of either sex	Methanol extract (MCP) of root and its sub fractions <i>viz</i> . hexane (HCP), ethyl acetate (ECP) and chloroform (CCP) (200 mg kg ⁻¹)	Carbon tetra chloride	MCP and its sub fractions HCP, ECP displayed hepatoprotective effect by reducing the elevated serum levels of, serum glutamic pyruvic transaminase, alkaline phosphatase and serum glutamic oxaloacetic transaminase, it increased high density lipoprotein. CCP does not show effective results	C. procera exerted hepatoprotective potential	83
Wistar rats of either sex	Hydro-ethanolic extract of <i>C. procera</i> flowers $(200 \text{ mg kg}^{-1} \text{ and}$ $400 \text{ mg kg}^{-1})$	Paracetamol- induced hepatitis	Improvement in the hepatic architecture was observed	C. procera flowers have hepatoprotective effect	84

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found to be more effective than tested reference compounds, such as mitoxantrone, taxol, oxaliplatin, irinotecan and temozolomide and less toxic than cardenolides. ^{155,156} Mechanism of UNBS1450 was studied and proven to be a potent sodium pump inhibitor as it inhibits NF-kB transactivation and triggers apoptosis by recruitment of pro-apoptotic Bak and Bax protein thereby leading to cell death. ^{157,158} Carrying out further *in vivo* studies will play a crucial role in ascertaining the safer use of UNBS1450. Therefore, further studies are necessary to obtain the clinically important lead molecules for the development of potent anticancer drugs.

5.2 Wound healing potential

C. procera has folk medicinal reputation as a wound healing agent. *In vivo* studies proved its wound healing potential as summarized in Table 4.

These data strongly support its ethnomedicinal use in wound healing potential and skin problems. *In vivo* screening showed considerable results in dose-dependent manner when compared to positive controls. A future perspective of studying the side effects and toxicity of the extracts at the dose level can also be unravelled.

5.3 Anti-inflammatory potential

Anti-inflammatory potential of extracts from *C. procera* have been summarized in Table 5.

On the basis of studies mentioned in Table 5, it can be concluded that the anti-inflammatory effect of dry latex needs to be further characterized as well as the nature of active principle leads responsible for anti-inflammatory activity remains to be identified.

5.4 Larvicidal/insecticidal potential

Aqueous and ethanolic extracts of leaves and other parts of *C. procera* showed significant larvicidal activities against various vector species as summarized in Table 6.

Above studies indicated that aqueous and ethanolic extracts of leaves of *C. procera* possessed phenomenal oviposition deterrent and larvicidal effect, thus it can be developed as environment friendly alternative for the synthetic insecticides for mosquito control.

5.5 Anthelmintic potential

C. procera is used as an anthelmintic by ruminant farmers as proved by activities summarized in Table 7.

5.6 Antioxidant potential

Leaves of *C. procera* displayed highest antiradical activity as evident from activities summarized in Table 8.

Above activities proved that quercetin, aqueous and methanolic extracts of leaves of *C. procera* possessed remarkable antiradical activity. Evaluation of the *in vivo* antioxidant potential would be indispensable, so that it can be used as natural antioxidant ingredients in food and drug industries.

5.7 Antiplasmodial potential

Traditional practitioners use *C. procera* as antimalarial agent. Activity summarized in Table 9.

Over past decades, reduction in efficiency of chloroquine has been observed, thus resistivity to antimalarial drugs can be a threat to control malaria. The hunt for analogues with reduced toxicity and improved antimalarial activity still prevails. The possibilities of finding active compounds and correlating with specific dose effective antimalarial activity, from those parts of the plant, which are used separately or together could be further pursued.

5.8 Hepatoprotective activity

In vivo experimental study proves that *C. procera* has hepatoprotective potential as summarized in Table 10.

5.9 Miscellaneous activities

Antiapoptotic activity of latex of *C. procera* was carried out by Sayed *et al.* (2016) on catfishes exposed to (100 μ g L⁻¹) 4-non-ylphenol as chemical pollutant. Significant (P < 0.05) decrease in apoptotic cells, enzymes (superoxidase dismutase, acetylcholinesterase cortisol *etc.*) and ions validified antiapoptotic activity of the crude latex against the toxicity of 4-non-ylphenol.¹⁵² Hence, crude latex exerted antiapoptotic activities against the toxicity of 4-non-ylphenol.

Anti-hyperbilirubinemic activity of leaves was evaluated using phenylhydrazine and paracetamol induced Wistar rats. Significant (P < 0.05) decrease in concentrations of serum total bilirubin in hyperbilirubinemic rats proved bilirubin lowering activity of aqueous extracts of *C. procera.*⁷⁰

Recent studies indicated that *C. procera* has significantly broader range of beneficial effects as it contains bioactive phytochemicals with therapeutic potential. By far only cytotoxic studies on cancer cell lines have been well established in clinical trials, whereas other activities have been evidenced by basic studies. Most of the studies are limited to *in vitro* studies which lack exploration of molecular mechanism of action. Therefore, mechanism based *in vitro* and *in vivo* studies should be carried out, which can lead to understanding of underlying mechanism related to traditional uses.

6. Phytochemistry

C. procera contains cardenolides, flavonoids, sterols, oxypregnanes triterpenoids, glycosides and other constituents as elaborated in Table 11.⁷ Flavonoid and its glycosides (Fig. 1) are the major compounds isolated from the leaves of *C. procera*. Steroids (Fig. 2) and cardenolides (Fig. 3) are the major secondary metabolites found in the latex. Cardenolides have also been reported from other plant genera of the family Apocynaceae or Asclepiadaceae like *Strophanthus*, *Cerbera*, *Apocynum*, *Nerium*, and *Thevetia*. ¹⁵⁹ Traditionally they are employed in curing of congestive heart failure. ¹⁶⁰ Cardenolides are C23 steroids with steroid nucleus having a glycoside moiety at C-3 and a lactone moiety at C-17. ⁶ Cardiac glycosides can be novel antineoplastic agents as cancer cells are more prone to these compounds. ¹⁵⁹ Terpenoids (ursane, olenane type and pentacyclic triterpenes *etc.*) (Fig. 4) have been

Table 11 Compounds isolated from Calotropis procera

S. No.	Compound name (molecular formula)	Extract/fraction	Eluent	Plant part & references
Elav	onoids			
	5-Hydroxy-3,7-dimethoxyflavone-4'- <i>O</i> -β-glucopyranoside (C ₂₃ H ₂₄ O ₁₁)	Ethanolic extract	Benzene-chloroform	Stem ¹³⁸
2	Isorhamnetin 3- <i>O</i> -β-D-rutinoside (C ₂₈ H ₃₂ O ₁₆)	85% methanolic extract	10-40% methanol	Leaves ^{76,164}
	Isorhamnetin 3- <i>O</i> - β -D-robinoside ($C_{28}H_{32}O_{16}$)	85% methanolic extract	10-40% methanol	Leaves ^{76,164}
	Isoquercitrin ($C_{21}H_{20}O_{12}$)	85% methanolic extract	70% methanol	Leaves ⁷⁶
	Quercetagetin-6-methyl ether 3- O - β -D- 4 C $_1$ -galacturonopyranoside ($C_{22}H_{20}O_{14}$)	85% methanolic extract	40-60% methanol	Leaves ⁷⁶
6	Quercetin $(C_{15}H_{10}O_7)$	85% methanolic extract	80% methanol	Leaves ⁷⁶
	Isorhamnetin (C ₁₆ H ₁₂ O ₇)	85% methanolic extract	80% methanol	Leaves ⁷⁶
	Azaleatin $(C_{16}H_{12}O_7)$	85% methanolic extract	80% methanol	Leaves ⁷⁶
	3,3'-Dimethoxy quercetin $(C_{17}H_{14}O_7)$	85% methanolic extract	50-60% ethyl acetate	Leaves ⁷⁶
	$3,6,3',4'$ -Tetramethoxy quercetin ($C_{18}H_{16}O_7$)	85% methanolic extract	50-60% ethyl acetate	Leaves ⁷⁶
	$3,6,7,3',4'$ -Pentamethoxy quercetin ($C_{19}H_{18}O_7$)	85% methanolic extract	60-100% ethyl acetate	Leaves ⁷⁶
	Kaempferol-3- O -rutinoside ($C_{27}H_{30}O_{15}$)	Methanolic extract	Ethyl acetate: water: formic acid: glacial acetic acid (100: 26: 11: 11, v/v)	Leaves ⁸⁶
13	Quercetin-3- O -rutinoside ($C_{27}H_{30}O_{16}$)	Methanolic extract	Ethyl acetate: water: formic acid: glacial acetic acid (100: 26: 11: 11, v/v)	Leaves ⁸⁶
14	Luteolin $(C_{15}H_{10}O_6)$	Ethanol-water extract (60 : 40)/butanol fraction	<i>n</i> -Hexane–acetone (70 : 30)	Stem bark ¹⁶⁵
15	Epicatechin (C ₁₅ H ₁₄ O ₆)	Ethanol-water extract (60 : 40)/butanol fraction	<i>n</i> -Hexane–acetone (60 : 40)	Stem bark ¹⁶⁵
	Kaempferol 3-O- α -1-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside ($C_{27}H_{30}O_{15}$)	Ethanolic extract	Water-methanol $(1:1)$	Fruits ¹⁴⁹
Ster	oids			
	Stigmasterol (C ₂₉ H ₄₈ O)	Methanolic extract/ hexane fraction	Hexane-ethyl acetate	Flowers, 166 root bark, 139 latex 165
18	$β$ -Sitosterol ($C_{29}H_{50}O$)	Ethanolic extract/ chloroform fraction	Hexane-ethyl acetate	bark, 139 latex 165 Flowers, 166 latex, 167 aerial part 168
19	Daucosterol or $\beta\text{-sitosterol}$ glucoside $(C_{35}H_{60}O_6)$	Ethanolic extract/ chloroform fraction	10% aq. methanol and hexane	Latex, aerial part, 168 roots 169
20	Benzoyllineolone ($C_{28}H_{36}O_6$)	Ether extract/chloroform fraction	Benzene-chloroform	Root bark ¹⁷⁰
21	Benzoylisolineolone ($C_{28}H_{36}O_6$)	Ether extract/chloroform fraction	Benzene-chloroform	Root bark ¹⁷⁰
22	Lineolone $(C_{21}H_{32}O_5)$	Ether extract	_	Root bark ¹⁷⁰
	Isolineolone (C ₂₁ H ₃₂ O ₅)	Ether extract	_	Root bark 170
	Cyclosadol (C ₃₁ H ₅₂ O)	Methanolic extract	_	Flowers ¹⁶⁶
	β-Sitost-4-en-3-one $(C_{29}H_{48}O)$	Methanolic extract	<i>n</i> -Hexane–ethyl acetate (95 : 5)	Flowers ¹⁶⁶
Ster	oids : cardenolides			
26	Calactin $(C_{29}H_{40}O_9)$	Ethanolic extract/ chloroform fraction	10% aq. methanol and hexane	Roots, ⁶² latex, ⁶⁵ aerial part ¹⁶⁸
	15 β -Hydroxycalactin ($C_{29}H_{40}O_{10}$)	Ethanolic extract/ chloroform fraction	_	Latex ⁶⁵
	Calactoprocin or 14 β ,15 β -dihydroxy-19-oxo-2 α ,3 β -[(2 S ,3 S :4 R ,6 R)-tetrahydro-3-hydroxy-4-methoxy-6-methyl-2 H -pyran-2,3-diyl] bis(oxy)-5 α -card-20(22)-enolide (3' β -methoxy-15 β -hydroxy calactin) (C ₃₀ H ₄₂ O ₁₀)	Ethanolic extract/ chloroform fraction	_	Latex ⁶⁵
	Afroside $(C_{29}H_{42}O_9)$	Ethanolic extract/ chloroform fraction	_	Latex ⁶⁵
30	Calotoxin $(C_{29}H_{40}O_{10})$	Ethanolic extract/ chloroform fraction	_	Aerial part, ¹⁶⁸ latex ⁶⁵
31	Calotropin $(C_{29}H_{40}O_9)$	Ethanolic extract/ chloroform fraction	_	Root bark, ⁶² latex and aerial part ¹⁶⁸
32	12 β -Hydroxycoroglaucigenin ($C_{23}H_{34}O_6$)	Ethanolic extract/ chloroform fraction	_	Latex ⁶⁵

Table 11 (Contd.)

S. No.	Compound name (molecular formula)	Extract/fraction	Eluent	Plant part & references
33	Procegenin A or 3α ,12β,14β-trihydroxy-19-hydroxymethyl- 5α -card-20(22)-enolide or 3 - epi ,12β-hydroxycoroglaucigenin ($C_{23}H_{34}O_6$)	Ethanolic extract/ chloroform fraction	_	Latex ⁶⁵
34	Procegenin B or 3α,12β,14β-trihydroxy-19-oxo-5α-card-20 (22)-	Ethanolic extract/	_	Latex ⁶⁵
35	enolide or 12 β -hydroxy carpogenin ($C_{23}H_{32}O_6$) Afrogenin ($C_{23}H_{34}O_6$)	chloroform fraction Ethanolic extract/	_	Latex ⁶⁵
36	Desglucouzarin $(C_{29}H_{44}O_9)$	chloroform fraction Ethanolic extract/ chloroform : ethyl acetate fraction	Chloroform–methanol $(9:1)$	Stem ¹⁷¹
37	Frugoside ($C_{29}H_{44}O_9$)	Ethanolic extract/ chloroform : ethyl acetate fraction	Chloroform-methanol $(9:1)$	Seeds, ⁶⁰ stem, ¹⁷¹ root bark ¹⁷²
38	Uzarigenin ($C_{23}H_{34}O_4$)	Ethanolic extract/ chloroform : ethyl acetate fraction	Chloroform–methanol (9.5 : 0.5)	Latex ⁶¹ Stem ^{168,171,173}
39	Uzarigenone ($C_{23}H_{32}O_4$)	Ethanolic extract/ benzene	Chloroform-methanol (9.5 : 0.5)	Stem ¹⁷¹
40	$\beta \hbox{-} Anhy droepidigitox igenin-3} \beta \hbox{-} {\it O} \hbox{-} glucopyranos ide } \left(C_{29} H_{42} O_8 \right)$	Ethanolic extract/ benzene : chloroform	Chloroform-methanol (9 : 1)	Stem ¹³⁸
41	β-Anhydroepidigitoxigenin or 3β-hydroxy-5α-carda-14(15),20(22)-dienolide ($C_{23}H_{32}O_3$)		Chloroform-methanol (9:2)	Stem ¹³⁸
	Calotropagenin $(C_{23}H_{32}O_6)$	Chloroform extract	Hexane-diethyl ether (9:11)	Aerial part ¹⁷⁴
	Ischarin (C ₃₁ H ₄₁ NO ₈ S)	Ethanolic extract Ethanolic extract/10%	Chloroform methanol (08 - 2)	Aerial part ¹⁶⁸ Aerial part ¹⁶⁸
44	Ischaridin $(C_{29}H_{42}O_8)$	aq. methanol and hexane fraction	Chloroform-methanol (98 : 2)	Aeriai part
45	$2''$ -Oxovoruscharin ($C_{31}H_{41}NO_9S$)	Methanolic extract	Dichloromethane-methanol (98 : 2)	Root bark ⁶²
46	Proceraside A $(C_{31}H_{44}O_{10})$	Methanolic extract/ethyl acetate fraction		Root bark ¹⁷²
47	Syriogenin (C ₂₃ H ₃₄ O ₅)	Methanolic extract	Water-methanol	Latex ⁶¹
48	Proceroside $(C_{29}H_{40}O_{10})$	Methanolic extract	Water-methanol	Latex ⁶¹
49	Uscharidin (C ₂₉ H ₃₈ O ₉)	Ethanolic extract	_	Aerial part ⁵⁶
	Voruscharin (C ₃₁ H ₄₃ NO ₈ S)	Methanolic extract	Acetone–methanol (8:2)	Roots ⁶²
	Coroglaucigenin (C ₂₃ H ₃₄ O ₅)	Chloroform extract	_	Seeds ⁶⁰
	Corotoxigenin (C ₂₃ H ₃₂ O ₅)	Ether extract	_	Seeds ⁶⁰
53	3-[β-(4- <i>O</i> -β- _D -Glucopyranosyl-β- _D -6-desoxyallopyranosyl)oxy]	70% ethanolic extract/	Chloroform–methanol (9 : 1.5)	Stem ¹⁷³
- 4	uzarigenin (C ₃₅ H ₅₄ O ₁₃)	benzene : chloroform		C+173
54	Uzarin or 3-[β-(2- <i>O</i> -β- _D -glucopyranosyl-β- _D -glucopyranosyl)oxy] uzarigenin (C ₃₅ H ₅₄ O ₁₄)	70% ethanolic extract/ benzene : chloroform	Chloroform-methanol (9:2)	Stem ¹⁷³
55	15β-Hydroxyuscharin (C ₃₁ H ₄₁ NO ₉ S)	Ethanolic extract	Chloroform	Latex ⁶⁵
	Uscharin ($C_{31}H_{41}NO_8S$)	Methanolic extract	Chloroform-methanol (70 : 30)	Aerial part, ¹⁶⁸ latex ^{65,168}
57	Proceragenin or 7 β ,14 β -dihydroxy-5 α -card-20(22)-enolide ($C_{23}H_{34}O_4$)	Methanolic extract/ chloroform fraction	Hexane-chloroform (1:9)	Aerial part ⁶³
58	2 β ,19-Epoxy-3 β ,14 β -dihydroxy-19-methoxy-5- α -card-20(22)-enolide (C ₂₄ H ₃₄ O ₆)	Ethanolic extract/ benzene : chloroform fraction	Chloroform-methanol (9 : 2)	Stem ¹³⁸
59	Procesterol or (24S)-24-ethyl-stigmast-4-en-6 α -ol-3-one (C ₂₉ H ₄₈ O ₂)	Ethanolic extract/ chloroform fraction	Hexane–chloroform (3 : 2)	Fresh and undried flowers ¹⁷⁶
	penes/terpenoids α -Amyrin ($C_{30}H_{50}O$)	Methanolic extract/ hexane : ethyl acetate gradients	Dichloromethane-methanol (1:1)	Flowers ¹⁷⁶
61	β -Amyrin ($C_{30}H_{50}O$)	Methanolic extract/ hexane : ethyl acetate gradients	Dichloromethane-methanol (1:1)	Flowers ¹⁷⁶
		gradients		

Table 11 (Contd.)

S. No.	Compound name (molecular formula)	Extract/fraction	Eluent	Plant part & references
	Procerursenyl acetate or urs-18α- <i>H</i> -12,20(30)-diene-3β-yl acetate $(C_{22}H_{50}O_2)$			
64	Calotropenyl acetate or urs-19(29)-3 β -yl acetate ($C_{32}H_{52}O_2$)	Chloroform extract	Benzene-hexane (60 : 40)	Flower, ¹⁷⁵ latex and aerial part ¹⁶⁸
65	Calotropoleanyl ester or olean-13(18)-en-3β-yl acetate (C ₃₂ H ₅₂ O ₂)	Ethanolic extract	Pet. ether	Root bark ¹⁷⁸
66	Calotroprocerol A or ursa-5,12,20(30)-trien-18 α H-3 β -ol ($C_{30}H_{46}O$)	Methanolic extract	n-Hexane-ethyl acetate	Root bark ¹³⁹
67	Calotroproceryl acetate A or ursa-5,12,20(30)-trien-18 αH -3 β -yl acetate (C32 H_{48} O2)	Methanolic extract	<i>n</i> -Hexane–ethyl acetate	Root bark ¹³⁹
	Calotroprocerone A or ursa-5,12,20(30)-trien-18 αH -3-one ($\mathrm{C}_{30}\mathrm{H}_{44}\mathrm{O}$)	Methanolic extract	n-Hexane–ethyl acetate	Root bark ¹³⁹
	Calotroproceryl acetate B or ursa-5,12,20-trien-18α H -3 β -yl acetate ($C_{32}H_{48}O_2$)		n-Hexane–ethyl acetate	Root bark ¹³⁹
	Calotropursenyl acetate B or urs-12,19(29)-diene-3 β -yl acetate ($C_{32}H_{50}O_2$)	Methanolic extract	n-Hexane–ethyl acetate	Root bark ^{139,186}
	Pseudo-taraxasterol acetate (C ₃₂ H ₅₂ O ₂)	Methanolic extract	n-Hexane–ethyl acetate	Root bark ¹³⁹
72	Taraxasterol $(C_{30}H_{50}O)$	Methanolic extract	n-Hexane–ethyl acetate	Root bark ¹³⁹
73	Proceroleanenol A or olean-13(18)-en- 9α -ol ($C_{30}H_{50}O$)	Ethanolic extract	Benzene chloroform	Root bark ¹⁷⁸
	Proceroleanenol B or olean-5,13(18)-dien-3α-ol (C ₃₀ H ₄₈ O)	Ethanolic extract Ethyl acetate extract	Benzene-chloroform (1:1)	Root bark ¹⁷⁸ Flowers ¹⁶⁶
75 76	Cycloart-23-ene-3 β ,25-diol ($C_{30}H_{50}O_2$) Lupeol ($C_{30}H_{50}O$)	Ethanolic extract	Hexane-ethyl acetate (2 : 1)	Latex ¹⁷⁹
		Ethanolic extract		Latex Latex
77 78	3-epi-Moretenol ($C_{30}H_{50}O$) Multiflorenol ($C_{30}H_{50}O$)	Pet. ether fraction	Chloroform–ethyl acetate (3 : 2)	Flowers, 166 latex 167
79	Urs-19(29)-en-3-β-ol (C ₃₀ H ₅₀ O)	Acetone fraction	Pet. ether–acetone (8 : 2)	Latex ¹⁶⁷
80	Calotropenyl acetate or urs-19(29)-en-3-yl acetate (C ₃₂ H ₅₂ O ₂)	Pet. ether fraction	Chloroform-ethyl acetate (3:5)	Latex ¹⁶⁷
81	3β ,27-Dihydroxy-urs-18-en-13,28-olide ($C_{30}H_{46}O_4$)	Ethyl acetate fraction	Benzene-ethyl acetate (8 : 2)	Latex ¹⁶⁷
82	Calotropfriedelenyl acetate or friedelin-1-ene-3 β -yl acetate ($C_{32}H_{52}O_2$)	Ethanolic extract	_	Root bark ¹⁸⁰
83	Calotropterpenyl ester or 6,10,14-trimethylpentadec-6-enyl- $2'$, $4'$, $8'$, $12'$, $16'$ -pentamethyl nonadecane ester ($C_{42}H_{82}O_2$)	Ethanolic extract	_	Root bark ¹⁸⁰
	Phytyl iso-octyl ether or 3,7,11,15-tetramethyl hexadecanyl-6'-methyl hept-5'-enyl ether ($C_{28}H_{56}O$)	Methanolic extract	Pet. ether-chloroform (1:3)	Roots ¹⁸¹
85	Dihydrophytoyl tetraglucoside or 3,7,11,15 tetramethylhexadecanoyl- β -D-glucopyranosyl- $(2 \rightarrow 1)$ - β -D-glucopyranosyl $(2 \rightarrow 1)$ - β -D-glucofuranoside $(C_{44}H_{80}O_{22})$	Methanolic extract	Chloroform-methanol (3:2)	Roots ¹⁸¹
86	Procerasesterterpenoyl triglucoside or 2,6,10,14,18-pentamethylnonadecanoyl- β -D-glucopyranosyl- $(2 \rightarrow 1)$ - β -D-glucopyranosyl- $(2 \rightarrow 1)$ - β -D-glucopyranoside ($C_{42}H_{78}O_{17}$)	Methanolic extract	Chloroform-methanol (3 : 1)	Roots ¹⁸¹
87	Oleanolic acid $(C_{30}H_{48}O_3)$	Chloroform extract/ butanol fraction	Benzene-ethyl acetate (10 : 1-1 : 10)	Stem bark ¹⁶⁵
	Lupeol-3- O -acetate ($C_{32}H_{52}O_2$) Proceraursenolide or 18- αH -urs-12-en-3,25-olide ($C_{30}H_{46}O_2$)	Ethanolic extract Ethanolic extract	Chloroform-methanol $(9.3:0.7)$ Pet. ether-chloroform $(1:3)$	Leaves ¹⁴⁹ Roots ¹⁸³
Ova	pregnane oligoglycosides			
•	Calotroposide H or 12-O-benzoylisolineolon-3-O- β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-oleandropyranosyl- $(1 \rightarrow 4)$ - β -D-	Methanolic extract/ <i>n</i> -butanol fraction	Chloroform-methanol (85 : 15)	Root bark ¹⁵³
91	cymaropyranosyl ($C_{63}H_{96}O_{21}$) Calotroposide I or 12- O -benzoylisolineolon-3- O - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-oleandropyranosyl- $(1 \rightarrow 4)$ - $(1 \rightarrow 4)$	Methanolic extract/ <i>n</i> -butanol fraction	Chloroform-methanol (85 : 15)	Root bark ¹⁵³
92	oleandropyranosyl ($C_{63}H_{96}O_{21}$) Calotroposide J or 12-O-benzoylisolineolon-3-O- β -D-cymaropyranosyl-(1 \rightarrow 4)- β -D-cymaropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranosyl-(1 \rightarrow 4)- β -D-cymaropyranosyl-(1 \rightarrow 4)-(6-O-acetyl)- β -D- glucopyranoside	Methanolic extract/ <i>n</i> -butanol fraction	Chloroform-methanol (85 : 15)	Root bark ¹⁵³
93	$(C_{71}H_{108}O_{27})$ Calotroposide K or 12- <i>O</i> -benzoylisolineolon-3- <i>O</i> -β-D-cymaropyranosyl- $(1 \rightarrow 4)$ -β-D-cymaropyranosyl- $(1 \rightarrow 4)$ -β-D-	Methanolic extract/ <i>n</i> -butanol fraction	Chloroform-methanol (85 : 15)	Root bark ¹⁵³

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Table 11 (Contd.)

o. Compound name (molecular formula)	Extract/fraction	Eluent	Plant part & references
oleandropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranosyl-(1 \rightarrow 4)- β -D)-		
cymaropyranosyl-(1 \rightarrow 4)- β -D- glucopyranoside ($C_{69}H_{106}O_{26}$)			153
Calotroposide L or 12-O-benzoylisolineolon-3-O-β-D-	Methanolic extract/n- butanol fraction	Chloroform–methanol (85 : 15)	Root bark ¹⁵³
cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-glucopyranosyl- $(1 \rightarrow 4)$ - β -D-glucopyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-	butanoi fraction		
oleandropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranoside (C ₆₈ H ₁₀₄ O	O_{28}		
Calotroposide M or 12- <i>O</i> -benzoylisolineolon-3- <i>O</i> -β-D-	Methanolic extract/n-	Chloroform-methanol (85 : 15)	Root bark ¹⁵³
cymaropyranosyl-(1 \rightarrow 4)- β -D-cymaropyranosyl-(1 \rightarrow 4)- β -D-	butanol fraction		
oleandropyranosyl- $(1 \rightarrow 4)$ - β -D-oleandropyranosyl- $(1 \rightarrow 4)$ - β -D			
oleandropyranosyl- $(1 \rightarrow 4)$ -β-D-oleandropyranoside- $(1 \rightarrow 4)$ - (6 acetyl) -β-D-glucopyranoside $(C_{78}H_{120}O_{30})$	I - O-		
5 Calotroposide N or 12- <i>O</i> -benzoylisolineolon-3- <i>O</i> -β-D-	Methanolic extract/n-	Chloroform-methanol (85:15)	Root bark ¹⁵³
cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-	butanol fraction	,	
oleandropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranosyl-(1 \rightarrow 4)- β -D)-		
oleandropyranosyl- $(1 \rightarrow 4)$ - β -D-glucopyranoside- $(1 \rightarrow 4)$ - β -D-			
gluopyranoside ($C_{75}H_{116}O_{31}$) Calotroposide S or 12-benzoylisolineolon-3- O -β-D-	Methanolic extract/n-	Chloroform-methanol (85 : 15)	Root bark ¹⁵⁴
cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-	butanol fraction	Cinorororiii inechanor (65 : 15)	Root bark
oleandropyranosyl- $(1 \rightarrow 4)$ - β -D-oleandropyranosyl- $(1 \rightarrow 4)$ - β -D)-		
cymaropyranosyl- $(1 \rightarrow 4)$ - β -D-oleandropyranosyl- $(1 \rightarrow 4)$ - β -D-			
oleandro-pyranosyl-(1 \rightarrow 4)- β -D-oleandropyranoside ($C_{84}H_{132}C_{132}$	O ₃₀)		
iphatic and phenolic glycoside			
B Methyl resorcinyl triglycoside or <i>O</i> -methyl resorcinyl-β-D-	Methanolic extract	Chloroform-methanol $(3:2)$	Roots ¹⁶⁹
glucuronopyranosyl (2 \rightarrow 1)- β -D-glucopyranosyl-(2 \rightarrow 1)- β -D-			
glucopyranoside (C ₂₅ H ₃₆ O ₁₈) (phenolic glycoside)	Mathanalia aytraat	Chloroform mothenol (4.1)	Roots ¹⁶⁹
Butanediol diglucuronoside or (<i>n</i> -butan-1,4-diol-1,4-β-D-diglucuronopyranoside) (C ₁₆ H ₂₆ O ₁₄) (aliphatic glycoside)	Methanolic extract	Chloroform–methanol (4 : 1)	ROOIS
(0) (E)-3-(4-Methoxyphenyl-2- O - β -D- 4 C ₁ -glucopyranoside)-methyl	85% methanolic extract	40-60% aqueous methanol	Leaves ⁷⁶
propenoate (C ₁₇ H ₂₂ O ₉)		•	
01 Methyl 4- <i>O</i> -β-D-glucopyranosyl ferulate (C ₁₇ H ₂₂ O ₉)	Ethanolic extract	Water–methanol $(1:1)$	Flowers ¹⁴⁹
gnan glycoside			
2 7'-Methoxy-3'-O-demethyl-tanegool-9-O-β-D-glucopyranoside	Ethanolic extract	Water-methanol (6:4)	Flowers 149
$(C_{26}H_{34}O_{12})$			- 140
33 Pinoresinol-4- <i>O</i> -glucoside (C ₂₆ H ₃₂ O ₁₁)	Ethanolic extract	Water methanol (1:1)	Flowers ¹⁴⁹ Fruits ¹⁴⁹
4 Syringaresinol-4- O -glucoside ($C_{28}H_{36}O_{13}$)	Ethanolic extract	Water–methanol $(1:1)$	FIUIUS
erpene glycoside			
05 Labdan-18-ol-β-ɒ-galactofuranoside (C ₂₆ H ₄₈ O ₆)	Methanolic extract	Chloroform-methanol (9:1)	Roots ¹⁸²
6 Proceralabdanoside/labdan-3β-ol-11,15-olide-18,20-dioic acid- 3β-p-galactofuranoside (C ₂₆ H ₄₀ O ₁₂)	Methanolic extract	Chloroform-methanol (9:1)	Roots ¹⁸²
$5p$ -b-galactolularioside ($O_{26}\Pi_{40}O_{12}$)			
affeic acid derivatives			
7 Methyl caffeate $(C_{10}H_{10}O_4)$		30-50% aqueous methanol	Leaves ⁷⁶
08 Caffeic acid (C ₉ H ₈ O ₄)		30–50% aqueous methanol	Leaves ⁷⁶ Flowers ¹⁴⁹
99 Rosmarinic acid ($C_{18}H_{16}O_8$) 10 Methyl rosmarinate ($C_{19}H_{18}O_8$)	Ethanolic extract Ethanolic extract	Chloroform–methanol (8.5 : 1.5) Chloroform–methanol (8.5 : 1.5)	Flowers Flowers Flowers
(0191-1008)	_maione enduce		_10015
thers			- 186
1 2-Propenyl-2Z-hydroxyethyl carbonate	— Methanolic extract	Bot other ableratorm (1 · 2)	Leaves ¹⁸⁶ Roots ¹⁷⁷
2 Glyceryl mono-oleolyl-2-phosphate (C ₂₁ H ₄₁ O ₇ P) 3 Methyl behenate (C ₂₃ H ₄₆ O ₂)	Methanolic extract	Pet. ether-chloroform (1 : 3) Chloroform-methanol (99 : 1)	Roots Roots 177
4 N-Dotriacont-6-ene ($C_{32}H_{64}$)	Methanolic extract	Pet. ether-chloroform (3 : 1)	Roots ¹⁷⁷
5 Methyl myrisate $(C_{15}H_{30}O_2)$	Methanolic extract	Chloroform	Roots ¹⁷⁷
6 Glyceryl-1,2-dicapriate-3-phosphate (C ₂₃ H ₄₅ O ₈ P)	Methanolic extract	Chloroform–methanol (97 : 3)	Roots ¹⁷⁷
17 (E)-Octadec-7-enoic acid ($C_{18}H_{34}O_2$)	Methanolic extract/n- hexane fraction	<i>n</i> -Hexane–ethyl acetate	Root bark ¹³⁹
18 Proceranol or <i>n</i> -triacontan-10β-ol (C ₃₀ H ₆₂ O)	Methanolic extract	Chloroform-methanol (99:1)	Roots ¹⁷⁷
19 Methyl ferulate	Methanolic extract	Chloroform–methanol (8.5 : 1.5)	Flowers ¹⁴⁹
20 1,2-Dihexadecanoyl-3-phosphatyl glycerol (C ₃₅ H ₆₉ O ₈ P)	Methanolic extract	Chloroform-methanol (99 : 1)	Roots ¹⁸¹
21	Methanolic extract	Pet. ether-chloroform (1:3)	Roots ¹⁸³

Table 11 (Contd.)

S. No. Compound name (molecular formula)	Extract/fraction	Eluent	Plant part & references
n-Tetradecanyl palmitoleate/n-tetradecanyl n-hexadec-9-enoate			
$(C_{30}H_{58}O_2)$			
122 Tricapryl glyceride (C ₃₃ H ₆₂ O ₆)	Methanolic extract	Pet. ether	Roots ¹⁸³
23 Oleodipalmityl glyceride (C ₅₃ H ₁₀₀ O ₆)	Methanolic extract	Pet. ether-chloroform (9:1)	Roots ¹⁸³
24 Tribehenyl glyceride (C ₆₉ H ₁₃₄ O ₆)	Methanolic extract	Pet. ether-chloroform (1:1)	Roots ¹⁸³
25 Capryl glucoside/ <i>n</i> -decanoyl-β-p-glucopyranoside (C ₁₆ H ₃₁ O ₇)	Methanolic extract	Chloroform-methanol (49:1)	Roots ¹⁸²
26 Palmityl glucoside/ n -hexacosanoyl- β-D-glucopyranoside $(C_{22}H_{43}O_6)$	Methanolic extract	Chloroform-methanol (19:1)	Roots ¹⁸²
.27 Stearyl glucoside/ <i>n</i> -octadecanoyl-β-p-glucopyranoside (C ₂₄ H ₄₇ O ₇)	Methanolic extract	Chloroform-methanol (93:7)	Roots ¹⁸²
.28 <i>n</i> -Heptanoate/heptylate ($C_8H_{16}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
29 <i>n</i> -Octanoate/caprylate (C ₉ H ₁₈ O ₂)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
30 <i>n</i> -Nonanoate $(C_{10}H_{20}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
31 <i>n</i> -Tridecanoate/tridecylat (C ₁₄ H ₂₈ O ₂)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
32 <i>n</i> -Pentadecanoate/pantadecylate (C ₁₆ H ₃₂ O ₂)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
33 <i>n</i> -Hexadecanoate/palmitate (C ₁₆ H ₃₄ O ₂)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
34 <i>n</i> -Heptadecanoate/margorate ($C_{18}H_{36}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part
35 Methyl nonanotetracnoate $(C_{10}H_{12}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
36 <i>n</i> -Decenoic acid ($C_{10}H_{18}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part
37 9-Decenoate $(C_{11}H_{20}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
38 Undecadienoate $(C_{12}H_{20}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
39 9-Dodecenoate $(C_{13}H_{24}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part
40 Tridecatrienoate $(C_{14}H_{22}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part
41 2,4,5-Tetradecatrienoate ($C_{14}H_{22}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part
42 Hiragonate $(C_{17}H_{28}O_2)$	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
43 Heptadecadienoate ($C_{18}H_{22}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
43 Heptadecenoate ($C_{18}H_{22}O_2$) 44 Heptadecenoate ($C_{18}H_{38}O_2$)	Ethanolic extract	Hexane-chloroform	Aerial part
44 Reptatecenoate (C ₁₈ H ₃₈ O ₂) 45 9-Eicosenoate/gadoleate (C ₂₁ H ₄₀ O ₂)	Ethanolic extract	Hexane-chloroform	Aerial part ¹⁶
0 (== == == == == == == == == == == == ==			Aerial part ¹⁸
46 Gallic acid (C ₇ H ₆ O ₅)	Ethanolic extract	HPLC analysis	Aeriai part
47 Ferulic acid $(C_{10}H_{10}O_4)$	Ethanolic extract	HPLC analysis	Aerial part ¹⁸
48 <i>p</i> -Coumaric acid (C ₉ H ₈ O ₃)	Ethanolic extract	HPLC analysis	Aerial part ¹⁸
49 Vanillic acid (C ₈ H ₈ O ₄)	Ethanolic extract	HPLC analysis	Aerial part ¹⁸
50 Rutin $(C_{27}H_{30}O_{16})$	Ethanolic extract	HPLC analysis	Aerial part ¹⁸
51 4-Hydroxy-4-methylpentan-2-one $(C_6H_{12}O_2)$	Acetone extract	GC-MS analysis	Latex ¹⁶¹
$52 2,3,4$ -Trimethylhexane (C_9H_{20})	Acetone extract	GC-MS analysis	Latex ¹⁶¹
53 Decane $(C_{10}H_{22})$	Acetone extract	GC-MS analysis	Latex ¹⁶¹
54 <i>n</i> -Pentadecane (C ₁₅ H ₃₂)	Acetone extract	GC-MS analysis	Latex ¹⁶¹
55 2,6-Dimethyl tetra-1,5-decaene (C ₁₆ H ₂₈)	Acetone extract	GC-MS analysis	Latex ¹⁶¹
56 n -Eicosane ($C_{20}H_{42}$)	Acetone extract	GC-MS analysis	Latex ¹⁶¹
57 3,7,11-Trimethyl-2,6,10,12-pentadecatrien-1-ol ($C_{18}H_{30}O$)	Acetone extract	GC-MS analysis	Latex ¹⁶¹
58 2,6,10,15,19,23-Hexamethyl-2,6,10,14,18,22-tetracosahexaene $(C_{30}H_{50})$	Acetone extract	GC-MS analysis	Latex ¹⁶¹
59 1,3,5-Tri-isopropylbenzene (C ₁₅ H ₂₄)	Acetone extract	GC-MS analysis	Latex ¹⁶¹
60 6,10,14-Trimethyl-pentadecanone-2 ($C_{18}H_{36}O$)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵
61 9-Octadecenoic acid (Z)-(C ₁₈ H ₃₄ O)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵
62 (6Z,9Z)-Pentadecadien-1-ol ($C_{15}H_{28}O$)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵
63 Farnesol isomer (C ₁₅ H ₂₆ O)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵
64 Tetratetracontane (C ₄₄ H ₉₀)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵
.65 Ergost-5-en-3-ol (C ₂₈ H ₄₈ O)	Hexane extract	GC-MS analysis	Leaves ¹⁸⁵

isolated from flowers, root bark and latex. Oxypregnane glycosides (Fig. 5) have recently been reported from root bark of this plant. ^{153,154} They have steroidal skeleton containing a 2-deoxy sugar moiety. These oxypregnanes have benzoyl moiety at C-12 and a straight 5–7 units sugar chain connected to C-3 of the aglycone. ⁶ Some glycosides (Fig. 6), lignan glycosides (Fig. 7), terpene glycosides (Fig. 8) and caffeic acid derivatives (Fig. 9) have also been isolated from this plant.

A number of hydrocarbons, saturated and unsaturated fatty acids were also identified from *C. procera* extract by GC-MS. ^{161,162} Similarly fatty acid ester, phthalate derivatives, and pentacyclic triterpenes were identified from chloroform extract of roots of *Calotropis procera*. ¹⁶³

Apart from the compounds mentioned in Table 11, terpenoids named α -calotropeol and β -calotropeol have been isolated from ethanolic extract of latex. ¹⁷⁹ A cardenolide named 19-

Fig. 1 Chemical structures of flavonoids.

13.

Fig. 2 Chemical structures of steroids.

Chemical structures of cardenolides

53. R= Glu(1 → 4)6-desoxyallosyl-O

R= Glu(1 →2)Glu-O

dihydrocalotropagenin and flavonoid named 3'-O-methyl-quercetin-3-O-rutinoside have also been reported from ethanolic extract of aerial parts.168

55. R=OH

Conclusion, discussion and future perspectives

In the present review, the research progress in phytochemistry and pharmacology of C. procera have been summarized. There

have been acquirements in the research; still some gaps came across our studies which are as follows:

58.

- (1) Folks and tribes have been using C. procera since ancient times; still investigations can be carried out on inception time of traditional uses of C. procera.
- (2) Secondary metabolites of plant vary according to several factors like region, environment, quality of soil, age of plant etc. Moreover, latex and root bark seem to be exhaustively investigated for phytoconstituents, not much research on flowers, pods and seeds for phyoconstituentsis have been conducted.

59

Fig. 4 Chemical structures of terpenoids.

Further exploring these parts can lead to discovery of new phytoconsituents of interest.

88

(3) The plant can be employed commercially as scientific studies have proved its use as cheese making agent, dehairing of leather, natural surfactant, biopesticide and corrosion inhibitor.

(4) Numerous activities on validation of its cytotoxic and anti-inflammatory potential have been conducted. A few have been carried out on its antimigraine, antiplasmodial and anti-convulsant effects. Carrying out further scientific studies in these fields can provide medical science with effective and promising new drugs.

89.

Fig. 5 Chemical structures of oxypregnanes.

(5) Most of the cytotoxic activities conducted are in vitro except the one conducted on UNBS1450; a semi-synthesized cardenolide. Further studies should be carried out to examine its in vivo potential.

Ole

Ole

Ole

Cym

Ole

Ole

(6) Right route and right dose can convert a dreadful toxicant into an outstanding drug whereas even a drug in lack of proper dosage and route can become a fatal poison. Folk practitioners have been employing C. procera as antifertility and uterotonic

Cym

Fig. 6 Chemical structures of glycosides.

Fig. 7 Chemical structures of lignan glycosides

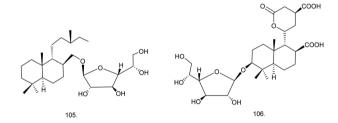


Fig. 8 Chemical structures of terpene glycosides.

Fig. 9 Chemical structures of caffeic acid derivatives.

agent. Further studies using positive controls, study of toxicity and side effects can lead to discovery of effective and natural contraceptive drugs.

- (7) Active principles behind many of the activities are unknown, except the one known for cytotoxic, antibacterial, antifertility, antimolluscicidal and insecticidal activity. More research can be carried out to know the active principles so that potent drugs can be made.
- (8) Replicable and environment benign sources of energy are the need of hour, *Calotropis procera* being rich source of various hydrocarbons, thus can prove to be a promising biofuel agent.

Overall, the pharmacology, toxicology, traditional uses, use of secondary metabolites, clinical trials and quality control has been reviewed in this paper. However, there seems to be a good correspondence between pharmacological activities and traditional uses. Further research in this field is essential to determine the active principles and the underlying mechanisms.

Author contributions

Barkha Darra Wadhwani: literature collection, evaluation and draft manuscript preparation. Deepak Mali and Pooja Vyas: literature collection: pharmacological activity and analyses of chemicals constituents of *C. procera*. Rashmy Nair: reviewing and editing. Poonam Khandelwal: concept development; idea generation; manuscript preparation; reviewing and editing.

Conflicts of interest

The authors confirm that this article content has no conflict of interest.

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References

Review

- 1 M. C. Joshi, M. B. Patel and P. J. Mehta, *Bull. Med.-ethno-bot. Res.*, 1980, 1, 8–24.
- 2 K. Chandra and U. N. Pandey, Some folk medicines of Singhbhum (Bihar), *Sachitra Ayurveda*, 1984, 37, 253–357.
- 3 L. S. Bhatnagar, V. K. Singh and G. Pandey, *J. Res. Indian Med.*, 1973, **8**(2), 67–100.
- 4 J. Venkateswarulu, P. V. Bhairavamurthy and N. Rao, *The Flora of Visakhapatnam*, Andhra Pradesh Academy of Sciences, Hyderabad, 1972, p. 128.
- H. S. Al-Mezaine, A. A. Al-Rajhi, A. Al-Assiri and M. D. Wagoner, *Am. J. Ophthalmol.*, 2005, 139, 199–202.
- 6 E. W. C. Chan, N. I. Sweidan, S. K. Wong and H. T. Chan, Rec. Nat. Prod., 2017, 11(4), 334-344.
- 7 P. M. Ranjit, G. E. Rao, M. Krishnapriya, V. Nagalakshmi, P. Silpa and M. Anjali, FS J. Pharm. Res., 2012, 1, 18–25.
- 8 R. Sharma, G. Thakur, B. S. Sanodiya, A. Savita, M. Pandey, A. Sharma and P. S. Bisen, *IOSR J. Pharm. Biol. Sci.*, 2012, 4(3), 42–57.
- P. A. Karale and M. A. Karale, *Asian J. Pharm. Clin. Res.*, 2017, 10, 27–34.
- 10 G. Parihar and N. Balekar, *Thai J. Pharm. Sci.*, 2016, **40**, 115–131.
- 11 R. K. Upadhyay, Int. J. Green Pharm., 2014, 8(3), 135-146.
- 12 R. P. Mali, P. S. Rao and R. S. Jadhav, *J. Drug. Deliv. Ther.*, 2019, **9**, 947–951.
- 13 H. S. Alzahrani, M. Mohamemd, S. Kulvinder and M. R. Rizgallah, *J. Appl. Environ. Biol. Sci.*, 2017, 7(10), 232–240.
- 14 A. K. Khairnar, S. R. Bhamare and H. P. Bhamare, *Adv. Res. Pharm. Biol.*, 2012, **2**, 142–156.
- 15 A. Ranade and R. Acharya, *Glob. J. Res. Med. Plants Indig. Med.*, 2014, 3(12), 475–488.
- 16 Z. Yaniv and H. Koltai, Isr. J. Plant Sci., 2018, 65, 55-61.
- 17 S. M. Bairagi, P. Ghule and R. Gilhotra, *Ars Pharm.*, 2018, **59**(1), 37–44.
- 18 N. Ranjan, S. K. Singh and C. Kumari, *Int. J. Curr. Microbiol. App. Sci.*, 2017, **6**(4), 1640–1648.
- 19 Poonam and G. Punia, Global J. Res. Med. Plants & Indigen. Med., 2013, 2(5), 392-400.
- 20 (a) S. Quazi, K. Mathur and S. Arora, *Indian J. Drugs*, 2013, 1(2), 63–69; (b) A. Bera, S. Maiti and N. Banerjee, *Int. J. Pharm. Sci. Res.*, 2020, 11(11), 5425–5433; (c) I. Pavani and S. Udayavani, *World J. Pharm. Res.*, 2020, 9(14), 1381–1392; (d) A. Kaur, D. R. Batish, S. Kaur and B. S. Chauhan, *Front. Plant Sci.*, 2021, 12, 690806, DOI: 10.3389/fpls.2021.690806.
- 21 P. Chandrawat and R. A. Sharma, *Res. J. Recent Sci.*, 2016, 5(1), 61–70.

- 22 A. K. Meena, A. Yadav and M. M. Rao, *Asian J. Tradit. Med.*, 2011, **6**(2), 45–53.
- 23 C. D. T. de Freitas, J. L. Lopes, L. M. Beltramini, R. S. B. de Oliveira, J. T. A. Oliveira and M. V. Ramos, *Biochim. Biophys. Acta*, 2011, **1808**, 2501–2507.
- 24 P. J. Modi, *Medical Jurisprudence and Toxicology, 2006*, first reprint Dr Mathiharan, K., Dr Patnaik, A.K. Lexis Nexis, New Delhi, 23rd edn, 2007, pp. 234–238.
- 25 B. Biedner and L. R. A. Witztum, *Isr. J. Med. Sci.*, 1977, **13**, 914–916.
- 26 W. Laukanjanaratand and M. Tovanich, *Thai. J. Ophthalmol.*, 1997, 1, 87–90.
- 27 T. Devasari, Indian J. Pharmacol., 1965, 27, 272–275.
- 28 S. K. Basak, A. Bhaumik, A. Mohanta and P. Singhal, *Indian J. Ophthalmol.*, 2009, 57(3), 232–234.
- 29 H. Tavakkoli, A. Derakhshanfar, J. Moayedi, A. P. Fard, S. Behrouz, M. A. Piltan and M. N. Soltani-Rad, *Comp. Clin. Pathol.*, 2019, 28, 195–202.
- 30 A. Akhkha, Biosci. Biotechnol. Res. Asia, 2009, 6(2), 653-658.
- 31 M. A. Ramadana, A. A. Azeiz, S. Baabada, S. Hassanein, N. O. Gadalla, S. Hassan, M. Algandaby, S. Bakr, T. Khan, H. H. Abouseadaa, H. M. Ali, A. Al-Ghamdi, G. Osman, S. Edris, H. Eissa and A. Bahieldin, Steroids, 2019, 141, 1–8.
- 32 A. S. Traore, Bioresour. Technol., 1992, 41, 105-109.
- 33 M. O. Barbosa, J. S. de Almeida-Cortez, S. I. da Silva and A. F. M. de Oliveira, *J. Am. Oil Chem. Soc.*, 2014, **91**, 1433–1441.
- 34 M. V. Ramos, C. D. T. Freitas and F. Staniscuaski, *Plant Science*, 2007, 173, 349–357.
- 35 G. E. Nenaah, Ind. Crops Prod., 2013, 45, 327-334.
- 36 O. C. Aworh and S. Nakai, J. Food Sci., 1986, 51, 1569-1570.
- 37 D. Raheem, N. Suri and P. E. Saris, *Int. J. Food Sci. Technol.*, 2007, 42, 220–223.
- 38 C. K. Atal and P. D. Sethi, Planta Med., 1962, 10(1), 77-90.
- 39 D. A. R. Agossou Yao, Y. Sprycha, S. Porembski and R. Horn, *Genet. Resour. Crop. Evol.*, 2015, **62**, 863–878.
- 40 M. Chandrashekar, H. Nagabhushana, S. C. Sharma, Y. S. Vidya, K. S. Anantharaju, D. Prasad, S. C. Prashantha, D. Kavyashree and P. S. Maiya, *Mater. Res. Express*, 2015, 2(4), 045402, DOI: 10.1088/2053-1591/2/4/045402.
- 41 P. B. Raja and M. G. Sethuraman, *Pigm. Resin Technol.*, 2009, **38**(1), 33–37.
- 42 L. Lopez, C. Viana, M. Errasti, M. L. Garro, J. E. Martegani, G. A. Mazilli, C. D. T. Freitas, I. M. S. Araujo, R. O. da silva and M. V. Ramos, *Bioprocess Biosyst. Eng.*, 2017, 40, 1391–1398.
- 43 M. K. Misra, M. K. Mohanty and P. K. Das, *Anc. Sci. Life*, 1993, **13**, 40–56.
- 44 L. Misra, Sahaja Chikichcha (in Oriya), ed. K. Devi Puri, 1959.
- 45 P. K. Jain, R. Verma, N. Kumar and A. Kumar, *Jour. Res. Ay. Sid.*, 1985, **6**, 88–91.
- 46 M. Garg, Sudhanidhi (Hindi edition) and D. Karyalaya, *Bijoygarh*, Uttar Pradesh, 1986, vol. 5, pp. 165–202.
- 47 K. R. Kirtikar and B. D. Basu, *Indian Medicinal Plants*, ed. B. Singh and M. Singh, Dehra Dun, 1933, vol. 3, pp. 1606–1611.

48 B. Tripathy, *Dravyaguna Kalpadruma (Oriya edition)*, ed. D. Tripathy, Nayagarh, 1953, pp. 22–28.

RSC Advances

- 49 Anon., *The wealth of India (Raw Materials)*, Council of Scientific and Industrial Research, New Delhi, 1959, vol. 2, pp. 20–23.
- 50 R. R. Pathak, *Therapeutic guide of Ayurvedic medicines*, Baidyanath Ayurveda Bhawan, Patna, 1970.
- 51 J. F. Dastur, *Medicinal Plants of India and Pakistan*, D. B. Taraporevalla Sons & Co., Bombay, 1970, pp. 43–44.
- 52 S. K. Jain, D. K. Banerjee and D. C. Pal, Medicinal Plants among certain Adivasis in India, *Bull. Bot. Surv. India*, 1973, 15, 85–91.
- 53 P. V. Sharma, *Dravyaguna Vigyana*, Choukamba Bharati Academy, Varanasi, India, 5th hindi edn, 1985.
- 54 P. K. Hajra and A. K. Baishya, *Ethnobotanical notes on the Miris (Mishings) of Assam Plains*, ed. S. K. Jain, Glimpses of Indian Ethnobotany, Oxford & IBH Publishing Co., New Delhi, 1981, pp. 161–169.
- 55 G. Hesse and F. Reicheneder, *Justus Liebigs Ann. Chem.*, 1936, 526, 252-276.
- 56 V. G. Hesse, F. Reicheneder and H. Eysenbach, *Justus Liebigs Ann. Chem.*, 1939, 537, 67–86.
- 57 G. Hesse and G. Ludwig, *Justus Liebigs Ann. Chem.*, 1960, 632, 158–171.
- 58 D. H. G. Crout, C. H. Hassall and T. L. Jones, *J. Chem. Soc.*, 1964, 2187–2194.
- 59 R. S. Gupta, N. Sharma and V. P. Dixit, Anc. Sci. life, 1990, 9(4), 224–230.
- 60 S. Rajagopalan, Ch. Tamm and T. Reichstein, Helv. Chim. Acta., Fasciculus, 1955, 38(7), 1809–1824.
- 61 F. Bruschweiler, W. Stocklin, K. Atockel and T. Reichstein, *Helv. Chem. Acta.*, 1969, 52, 2086–2106.
- 62 V. E. Quaquebeke, G. Simon, A. Andre, J. Dewelle, M. E. Yazidi, F. Bruyneel, J. Tuti, O. Nacoulma, P. Guissou, C. Decaestecker, J. C. Braekman, R. Kiss and F. Darro, J. Med. Chem., 2005, 48, 849–856.
- 63 N. Akhtar and A. Malik, Phytochemistry, 1992, 31(8), 2821– 2824.
- 64 H. Joshi, V. Havannavar, C. Gavimat, H. Pooja and P. Praveena, *J. Alzheimer's Assoc.*, 2008, 4(4), T502.
- 65 N. H. Mohamed, M. Liu, W. M. Abdel-Mageed, L. H. Alwahibi, H. Dai, M. A. Ismail, G. Badr, R. J. Quinn, X. Liu, L. Zhang and A. A. M. Shoreit, *Bioorg. Med. Chem. Lett.*, 2015, 25, 4615–4620.
- 66 A. S. Awaad, G. M. Zain, M. Reham, H. F. Alkanhal and V. D. Seshadri, *Calotropis procera* extracts as antiulcerative colitis agents, *US Pat.*, 9533019B1, 2017.
- 67 A. M. Rasik, R. Raghubir, A. Gupta, A. Shukla, M. P. Dubey, S. Srivastava, H. K. Jain and D. K. Kulshrestha, *J. Ethnopharmacol.*, 1999, 68, 261–266.
- 68 A. O. Aderounmua, A. E. Omonisib, J. A. Akingbasotec, M. Makanjuolad, R. A. Bejide, L. O. Orafidiya and K. A. Adelusolae, *Afr. J. Tradit. Complement. Altern. Med.*, 2013, 10(3), 574–579.
- 69 D. A. Tsala, N. Nga, M. B. N. Thiery, M. T. Bienvenueand and D. Theophile, *J. Intercult. Ethnopharmacol.*, 2015, 4(1), 64–69.

- 70 R. A. Patil and A. B. Makwana, *Indian J. Pharmacol.*, 2015, 47(4), 398–402.
- 71 R. P. Samy and V. T. K. Chow, *Evid. Based Complement. Alternat. Med.*, 2012, 294528, DOI: 10.1155/2012/294528, PMID: 22973400, .
- 72 A. S. Seddek, A. A. El-Ghoneimy, M. W. Dina, S. El-hamd and E. G. Mahmoud, *Egypt. J. Chem. Environ. Health*, 2015, 1(1), 768–784.
- 73 J. D. Mbako, Z. Adamu, J. K. Afutu, A. Aliyu, S. David, M. B. Umar and C. Nduaka, *Afr. J. Biotechnol.*, 2009, 8(19), 5071–5075.
- 74 G. B. Pouokam, H. Ahmed, C. Dawurung, A. Atiku, S. David and O. Philipe, *J. Toxicol. Environ. Health Sci.*, 2011, 3(5), 119–126
- 75 A. M. Dieye, M. A. Tidjani, A. Diouf, E. Bassene and B. Faye, *Dakar Med.*, 1993, **38**(1), 69–72.
- 76 M. A. Mohamed, M. M. Hamed, W. S. Ahmed and A. M. Abdou, *Z. Naturforsch., C: J. Biosci.*, 2011, **66**, 547–554.
- 77 T. L. Juca, M. V. Ramos, F. B. M. Batista Moreno, M. P. V. de Matos, J. D. B. Marinho-Filho, R. A. Moreira and A. C. de Oliveira Monteiro-Moreiro, *Sci. World J.*, 2013, 615454, DOI: 10.1155/2013/615454.
- 78 E. A. A. Sadaqa and K. S. Ali, *Int. J. Pharm. and Pharm. Res.*, 2019, **16**(4), 400–407.
- 79 E. S. A. Toson, S. A. Habib, E. A. Saad and N. H. Harraz, *Int. J. Biochem.*, 2014, 195, 328–338.
- 80 A. B. Abbasi, R. Bibi, A. A. Khan, M. S. Iqbal, J. Sherani and A. M. Khan, J. Biofertil. Biopestici., 2012, 3, 126.
- 81 P. S. Jahan, A. Mannan, A. R. Khan and P. Karmakar, *Bangladesh J. Zool.*, 1991, **19**(2), 261–262.
- 82 I. A. Muraina, A. O. Adaudi, M. Mamman, H. M. Kazeem, J. Picard, L. J. McGaw and J. N. Elof, *Pharm. Biol.*, 2010, 48(10), 1103–1107.
- 83 R. Chavda, K. R. Vadalia and R. Gokani, *Int. J. Pharmacol.*, 2010, **6**(6), 937–943.
- 84 S. R. Setty, A. A. Quereshi and A. H. M. Viswanath Swamy, *Fitoterapia*, 2007, 78, 451–454.
- 85 A. Basu, T. Sen, R. N. Ray and A. K. Nag-Chaudhuri, *Fitoterapia*, 1992, **63**(6), 507–514.
- 86 G. Nenaah, World J. Microbiol. Biotechnol., 2013, **29**, 1255–1262.
- 87 S. O. Kareem, I. Akpan and O. P. Ojo, *Afr. J. Biomed. Res.*, 2008, **11**, 105–110.
- 88 H. O. Oladimeji, R. Nia and E. E. Essien, *Afr. J. Biomed. Res.*, 2006, **9**, 205–211.
- 89 S. C. Jain, R. Sharma, R. Jain and R. A. Sharma, *Fitoterpia*, 1996, **67**(3), 275–277.
- 90 T. L. Nascimento, Y. Oki, D. M. M. Lima, J. S. Almeida-Cortez, G. W. Fernandes and C. M. Souza-Motta, *Fungal Ecol.*, 2015, 14, 79–86.
- 91 V. H. Bhaskar, Asian J. Chem., 2000, 21(7), 5788-5790.
- 92 B. Desta, J. Ethnopharmacol., 1993, 39(2), 129-139.
- 93 N. Mascolo, R. Sharma, S. C. Jain and F. Capasso, *J. Ethnopharmacol.*, 1988, **22**(2), 211–221.
- 94 O. P. Shukla and C. R. Krishnamurti, *J. Sci. Ind. Res.*, 1961, **20**(8), 225–226.

Review

95 M. S. Kumar and U. K. Chanhan, *Geobios*, 1992, **19**, 135–137.

- 96 N. Nawazisht, I. Malik and M. I. D. Chugtai, *Pak. J. Sci.*, 1979, 31, 127–129.
- 97 A. H. Kawo, A. Mustapha, B. A. Abdullahi, L. D. Rogo, Z. A. Gaiyaand and A. S. Kumurya, *Bayero. J. Pure Appl. Sci.*, 2009, 2(1), 34–40.
- 98 P. O. Akindele, O. A. Fatunla, K. A. Ibrahim and C. O. Afolayan, *J. Complement. Altern. Med. Res.*, 2017, 2(1), 1–14.
- 99 V. Talsaniya, T. Patel, N. Saiyad, S. Desai, D. Patel and D. Meshram, *Int. J. Pharm. Sci. Rev. Res.*, 2014, 25(2), 241– 244.
- 100 R. Lima, N. Lima, E. Chaves, L. Leal, M. Patrocinio, R. Lobato, M. Ramos, F. C. F. Sousa, K. Carvalho and S. Vasconcelos, J. Complement. Integr. Med., 2010, 7, 1–9.
- 101 S. Gholamshahi, A. V. Mohammad, S. Fatemeh and A. Salehi, *Int. J. Biosci.*, 2014, 4(7), 159–164.
- 102 M. N. Yesmin, S. N. Uddin, S. Mubassara and M. A. Akond, *American-Eurasian J. Agric. & Environ. Sci.*, 2008, 4(5), 550–553.
- 103 S. Loonker, W. A. Qadri and J. Singh, *Int. J. Cur. Res. Rev.*, 2015, 7, 55–59.
- 104 P. M. Soares, S. R. Lima, S. G. Matos, M. M. Andrade, M. C. A. Patrocinio, C. D. T. de Freitas., M. V. Ramos, D. N. Criddle, B. A. Cardi, K. M. Carvalho, A. M. S. Assreuy and S. M. M. Vasconcelos, J. Ethnopharmocol., 2005, 99, 125–129.
- 105 Z. Iqbal, M. Lateef, A. Jabbar, G. Muhammad and M. N. Khan, *J. Ethnopharmacol.*, 2005, **102**, 256–261.
- 106 Y. M. Shivkar and V. L. Kumar, *Pharm. Biol.*, 2003, **41**(4), 263–265.
- 107 A. A. Al-Qarawi, O. M. Mahmoud, M. A. Sobaih, E. M. Haroum and S. E. I. Adam, *Vet. Res. Commun.*, 2001, 25, 61–70.
- 108 H. Sangraula, S. Dewan and V. L. Kumar, *Inflammopharmacology*, 2002, **9**(3), 257–264.
- 109 V. L. Kumar and N. Basu, *J. Ethnopharmacol.*, 1994, **44**, 123–125.
- 110 N. S. Tour and G. S. Talele, Rev. Bras. Farmacogn., 2011, 21(6), 1118–1126.
- 111 P. K. Majumdar and V. L. Kumar, *Phytother. Res.*, 1997, 11(2), 166–167.
- 112 C. R. Jangde, C. G. Raut and V. V. Bisan, *Livestock Advisor*, 1994, **19**(3), 29–31.
- 113 S. Kumar, S. Dewan, H. Sangraula and V. L. Kumar, *J. Ethnopharmacol.*, 2001, **76**(1), 115–118.
- 114 O. J. Olaitan, S. U. R. Wasagu, A. A. Adepoju-Bello, K. U. Nwaeze and A. Olufunsho, *Nig. Q. J. Hosp. Med.*, 2013, 23(4), 338–341.
- 115 D. Srivastav and P. Singh, World J. Pharm. Res., 2015, 4(3), 1123–1135.
- 116 M. Larhsini, M. Bonsaid, H. Lazrek, M. Jana and H. Amarouch, *Fitoterapia*, 1997, **68**(4), 371–373.
- 117 R. M. Aliyu, M. B. Abubakar, Y. U. Dabai, N. Lawal, M. B. Bello and A. Y. Fardami, *J. Intercult. Ethnopharmacol.*, 2015, 4(4), 314–317.

- 118 N. Pathak and R. K. Zaidi, Ann. Biol. Res., 2013, 4(4), 1-6.
- 119 A. M. Mashlawi, M. K. H. Ali and E. S. Tarek, *Int. J. Mosq. Res.*, 2017, 4(1), 1–6.
- 120 N. Begum, B. Sharma and R. S. Pandey, *J. Biofertil. Biopestici.*, 2010, 1, 101.
- 121 A. M. Elimam, K. H. Elimalik and F. S. Ali, *J. Biol. Sci.*, 2009, **16**, 95–100.
- 122 H. Doshi, H. Satodiya, M. C. Thakur, F. Parabia and A. Khan, *Int. J. Plant Res.*, 2011, 1(1), 29–33.
- 123 N. M. Azmathullah, M. A. Sheriff and A. K. S. Mohideen, *Int. J. Pharm. Biol. Arch.*, 2011, **26**, 1718–1721.
- 124 S. M. P. Khurana and S. Singh, *Phytopathol. Z.*, 1972, 73, 341–346.
- 125 J. V. Kamath and A. C. Rana, *Fitoterapia*, 2002, 73(2), 111–115.
- 126 S. M. A. El-Badwi and A. O. Bakhiet, *Sci. Res. Essays*, 2010, 5(17), 2404–2408.
- 127 M. A. Qureshi, N. M. Qureshi, R. Arshad and R. Begum, *Pak. J. Zool.*, 1991, 23(2), 161–165.
- 128 C. Circosta, R. Sanogo and F. Occhiuto, *IL Farmaco*, 2001, **56**, 373–378.
- 129 M. V. Ramos, C. A. Viana and A. F. Silva, Naunyn Schmiedebergs Arch. Pharmacol., 2012, 385(5), 455-463.
- 130 P. Sharma and J. D. Sharma, *J. Ethnopharmacol.*, 1999, **68**, 83–95.
- 131 S. Y. Mudi and A. Bukar, Biochemistry, 2011, 23, 29-34.
- 132 S. Dewan, S. Kumar and V. L. kumar, *Ind. J. Pharmacol.*, 2000, **32**, 252–253.
- 133 U. P. Upadhyay, J. Sci. Res. Plant. Med., 1979, 1(1), 52-55.
- 134 S. S. Jalalpure, Pharm. Biol., 2009, 47(2), 162-167.
- 135 J. S. Oliveira, D. P. Bezerra, C. D. T. Freitas, J. D. B. Marinho-Filho, M. O. de Moraes, C. Pessoa, L. C. V. Costa-Lotufo and M. V. Ramos, *Toxicol. In. Vitro.*, 2007, 21, 1563–1573.
- 136 R. Mathur, S. K. Gupta, S. R. Mathur and T. Velpandian, *Indian J. Exp. Biol.*, 2009, 47(5), 343–348.
- 137 A. L. Joshi, P. H. Roham, R. Mhaske, M. Jadhava, K. Krishnadasa, A. Kharatb, B. Hardikarc and R. K. Kiran, *Nat. Prod. Res.*, 2015, **29**, 2261–2264.
- 138 K. H. Shaker, N. Morsy, H. Zinecker, J. F. Imhoff and B. Schneider, *Phytochem. Lett.*, 2010, 3, 212–216.
- 139 S. R. M. Ibrahim, G. A. Mohamed, L. A. Shaala, L. M. Y. Banuls, G. V. Goietsenoven, R. Kiss and D. T. A. Youssef, *Phytochem. Lett.*, 2012, 5(3), 490–495.
- 140 M. Bhagat, J. S. Arora and A. K. Saxena, *Int. J. Green. Pharm.*, 2010, 4, 286–288.
- 141 V. H. Bhaskar and S. A. Sumant, *Global J. Pharmacol.*, 2009, 3, 95–98.
- 142 V. L. Kumar and S. Roy, *Phytother. Res.*, 2009, 23, 1–5.
- 143 P. Chaudhary, M. V. Ramos, Md S. Vasconcelos and V. L. Kumar, *Pharmacogn. Mag.*, 2016, **12**, 147–151.
- 144 H. T. Hussein, A. Kamel, M. Abou-Zeid, A. K. H. El-Sebae and M. A. Saleh, Uscharin, *J. Chem. Ecol.*, 1994, **20**(1), 135–140.
- 145 G. Giridhar, S. Santosh and P. Vesudevan, *Pesticides*, 1988, 22, 31–33.
- 146 G. Prasad, J. Nat. Med. Assoc., 1985, 27, 7-10.

- 147 A. Basu, T. Sen, S. Pal, F. Capasso and A. Nagchaudhri, Phytother. Res., 1997, 11, 163-165.
- 148 S. K. Bhatnagar and S. K. Verma, J. Econ. Taxon. Bot., 1986, 8, 489-490.
- 149 A. M. Al-Taweel, S. Perveen, G. A. Fawzy, A. U. Rehman, A. Khan, R. Mehmood and L. M. Fadda, Evid. Based Complement. Alternat. Med., 2017, 2017, 1-10.
- 150 E. O. Iwalewa, A. O. Elujoba and A. Olanrewaju, Fitoterapia, 2005, 76(2), 250-253.
- 151 S. B. S. Aliyu-Umar and Y. Mustapha, Unique. Res. J. Agric. Sci., 2014, 2(4), 37-41.
- 152 A. D. Sayed, N. H. Mohammed, M. A. Ismail, W. M. Abdel-Mageedand and A. A. Shoreit, Ecotoxicol. Environ. Saf., 2016, 128, 189-194.
- 153 S. R. M. Ibrahim, G. A. Mohamed, L. A. Shaala, L. M. Y. Banuls, R. Kiss and D. T. A. Youssef, Steroids, 2015, 96, 63-72.
- 154 S. R. M. Ibrahim, G. A. Mohamed, L. A. Shaala and D. T. A. Youssef, Rec. Nat. Prod., 2016, 10, 761-765.
- 155 T. Mijatovic, F. Lefranc, V. E. Quaquebeke, F. V. Vynckt, F. Darro and R. Kiss, Drug Dev. Res., 2007, 68, 164-173.
- 156 T. Mijatovic, D. V. Neve, P. Gailly, V. Mathieu, B. Haibe-Kains, G. Bontempi, J. Lapeira, C. Decaestecker, V. Facchini and R. Kiss, Mol. Cancer Ther., 2008, 7, 1285-1296.
- 157 T. Juncker, M. Schumacher, M. Dicato and M. Diederich, Biochem. Pharmacol., 2009, 78, 1-10.
- 158 T. Juncker, C. Cerella, M. H. Teiten, F. Morceau, M. Schumacher, J. Ghelfi, F. O. Gaascht, M. Schnekenburger, E. Henry, M. Dicato M. Diederich, Biochem. Pharmacol., 2011, 81, 13-23.
- 159 S. Wen, Y. Chen, Y. Lu, Y. Wang, L. Ding and M. Jiang, Fitoterapia, 2016, 112, 74-84.
- 160 I. Prassas and E. P. Diamandis, Nat. Rev. Drug. Discov., 2008, 7, 926-935.
- 161 H. V. Doshi, F. M. Parabia, F. K. Sheth, I. L. Kothari, M. H. Parabia and A. Ray, Int. J. Plant. Res., 2012, 2(2), 28-30.
- 162 S. K. Khanzada, W. Shaikh, T. G. Kazi, S. Sofia, A. Kabir, K. Usmanghani and A. A. Kandhro, Pak. J. Bot., 2008, 40(5), 1913-1921.
- 163 A. A. Ibrahim and E. H. Tuhami, Sci. J. Anal. Chem., 2019, 4(2), 20-24.
- 164 R. S. Gallegos-Olea, M. O. R. Borges, A. C. R. Borges, M. F. Freire, L. M. S. Silveira, W. Vilegas,

- C. M. Rodrigues, A. V. Oliveira and J. L. Costa, Rev. Bras. Pl. Med., Botucatu., 2008, 10(1), 29-33.
- 165 N. S. Tour and G. S. Talele, Chem. Nat. Compd., 2012, 48(4), 708-709.
- 166 A. Q. Khan and A. Malik, Fitoterapia, 1990, 61(1), 89.
- 167 S. J. Chundattu, V. K. Agrawal and N. Ganesh, Arab. J. Chem., 2016, 9, S230-S234.
- 168 N. I. Sweidan and M. H. Abu Zarga, J. Asian Nat. Prod. Res., 2015, 17, 900-907.
- 169 A. Mittal and M. Ali, Int. J. Pharmtech. Res., 2012, 4(1), 213-217.
- 170 R. F. Chandler, R. G. Coombe and T. R. Watson, Aust. J. Chem., 1968, 21(6), 1625–1631.
- 171 M. H. A. Elgamal, A. G. Hanna, N. A. M. Morsy, H. Duddeck, A. Simon, T. Gati and G. Toth, J. Mol. Struct., 1999, 477, 201-208.
- 172 S. R. M. Ibrahim, G. A. Mohamed, L. A. Shaala, L. Moreno, Y. Banuls, R. Kiss and D. T. A. Youssef, Nat. Prod. Res., 2014, 28, 1322-1327.
- 173 A. G. Hanna, M. H. A. Elgamal, N. A. M. Morsy, H. Duddeck, J. Kovacs and G. Toth, Magn. Reson. Chem., 1999, 37, 754-
- 174 B. Singh and R. P. Rastogi, *Phytochemistry*, 1972, **11**(2), 757– 762.
- 175 A. Q. Khan, Z. Ahmed, S. N. Kazmi and A. Malik, J. Nat. Prod., 1988, 51, 925-928.
- 176 A. Q. Khan and A. Malik, Phytochemistry, 1989, 28, 2859-2861.
- 177 P. Alam and M. Ali, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2009, 48, 443-446.
- 178 S. H. Ansari and M. Ali, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2000, 39, 287-290.
- 179 R. Pant and K. Chaturvedi, Curr. Sci., 1989, 58, 740–724.
- 180 S. H. Ansari and M. Ali, Pharmazie, 2001, 56(2), 175-177.
- 181 A. Mittal and M. Ali, J. Saudi. Chem. Soc., 2015, 19, 59-63.
- 182 A. Mittal and M. Ali, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2013, 52, 641-645.
- 183 A. Mittal and M. Ali, Int. Res. J. Pharm., 2011, 2(9), 52-54.
- 184 M. A. Khasawneh, H. M. Elwy, N. M. Fawzi, A. A. Hamza, A. R. Chevidenkandy and A. H. Hassan, Res. J. Phytochem., 2011, 5(2), 80-88.
- 185 B. Dwivedi, A. Singh, S. Mishra, R. Singh, P. Pant, L. K. Thakur and M. M. Padhi, World J. Pharm. Res., 2014, 3, 708-715.
- 186 R. S. Gallegos Olea, A. V. Oliveira, L. M. Silveira and E. R. Silveira, Fitoterapia, 2002, 73, 263-265.