



Cite this: *Chem. Commun.*, 2022, 58, 7184

Received 2nd February 2022,
Accepted 30th May 2022

DOI: 10.1039/d2cc00671e

rsc.li/chemcomm

Hydride state accumulation in native [FeFe]-hydrogenase with the physiological reductant H₂ supports its catalytic relevance†

Moritz Senger,^a Tobias Kernmayr,^{‡b} Marco Lorenzi,^b Holly J. Redman^b and Gustav Berggren^b

Small molecules in solution may interfere with mechanistic investigations, as they can affect the stability of catalytic states and produce off-cycle states that can be mistaken for catalytically relevant species. Here we show that the hydride state (H_{hyd}), a proposed central intermediate in the catalytic cycle of [FeFe]-hydrogenase, can be formed in wild-type [FeFe]-hydrogenases treated with H₂ in absence of other, non-biological, reductants. Moreover, we reveal a new state with unclear role in catalysis induced by common low pH buffers.

Hydrogenases are redox enzymes catalysing molecular hydrogen (H₂) uptake and production. The high H₂ evolution frequency of prototypical [FeFe]-hydrogenases motivates basic research on the reaction mechanism and inspires catalyst design for a hydrogen economy.^{1–3} In these enzymes, hydrogen catalysis takes place at a hexa-iron cofactor, called the H-cluster. It is composed of a [4Fe4S]-cluster linked *via* a cysteine residue to a unique diiron site ([2Fe]) that binds two cyanide (CN[−]) and three carbonyl (CO) ligands. These ligands serve as intrinsic probes sensitive to infrared spectroscopy and report on reduction and protonation events during catalysis. The diiron site is bridged by an azadithiolate ligand (−SCH₂NHCH₂S−, ADT) that is proposed to shuttle protons *via* its amine group to the apical vacancy, the site of hydrogen catalysis, located at the Fe ion (Fe_d) distal to the [4Fe4S] cluster (Fig. 1). The most studied group of [FeFe]-hydrogenases (Group A) transfers protons to the active site *via* a conserved Proton Transfer Pathway (PTP) composed of amino acid residues and water molecules.^{4–6} Recently described [FeFe]-

hydrogenases in group D lack this PTP.^{7,8} Several redox and protonation states have been characterised and proposed as catalytic intermediates.^{1,2} The starting point of catalysis is the oxidised state H_{ox}. Under inert atmosphere conditions at low pH values and in presence of chemical reductant a blue-shifted variant of H_{ox} is formed, commonly denoted H_{ox}H.^{9,10} One electron reduction of H_{ox} results in population of two singly reduced states: H_{red}′ or H_{red}. H_{red}′ features a reduced [4Fe4S] cluster while the diiron site is still in an oxidised configuration. Formation of the diiron site reduced state H_{red} is coupled to a proton uptake event, although the site of protonation is under discussion.^{11,12} A second reduction step yields the super-reduced state (H_{sred}) or a hydride state (H_{hyd}). H_{sred} is characterised by a reduction of both the [4Fe4S] cluster and the diiron site.¹³ In contrast, H_{hyd} features a terminal hydride at Fe_d, yielding a

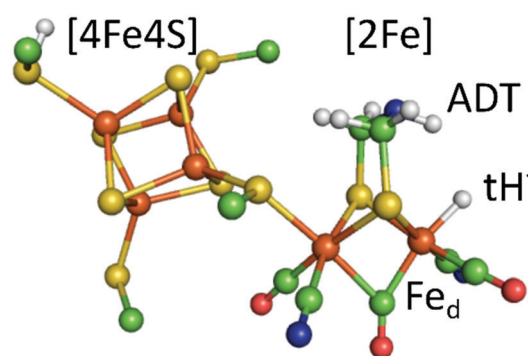


Fig. 1 The calculated hydride state structure of the H-cluster in [FeFe]-hydrogenases. The H-cluster consists of a protein bound [4Fe4S] cluster that is covalently linked *via* a cysteine thiolate ligand to the diiron site ([2Fe]), further ligated by three carbonyl (CO) and two cyanide (CN[−]) ligands. A terminal hydride (tH[−]) at the distal iron ion is proposed to combine with a proton delivered by the ADT ligand to yield molecular hydrogen in the last step of hydrogen catalysis. Colour code: orange-iron, yellow-sulphur, green-carbon, red-oxygen, blue-nitrogen, white-hydrogen. The structure is drawn after the PDB coordinates 4XDC optimized for the H_{hyd} state by DFT calculations. (ref. 33).

^a Department of Chemistry, Physical Chemistry, Uppsala University, 75120 Uppsala, Sweden. E-mail: moritz.senger@kemi.uu.se

^b Department of Chemistry, Molecular Biomimetics, Uppsala University, 75120 Uppsala, Sweden. E-mail: gustav.berggren@kemi.uu.se

† Electronic supplementary information (ESI) available: Materials and methods, FTIR spectra, Gas interaction kinetics. See DOI: <https://doi.org/10.1039/d2cc00671e>

‡ Current address: Department of Chemistry, Ludwig-Maximilians-Universität München, 81377 München, Germany.



formally oxidized diiron site (Fig. 1).^{14–16} As the last step in hydrogen turnover, the terminal hydride of H_{hyd} is proposed to combine with a proton delivered *via* the PTP to generate molecular hydrogen.^{1–3} A hydride state similar to H_{hyd} but with an additionally protonated ADT bridge has been proposed as the last intermediate in hydrogen evolution.^{17–20}

A terminal-hydride state of the H-cluster, considered to reflect the H_{hyd} state, can be stabilised by impairing the enzyme's catalytic function *via* amino acid variations in the PTP or cofactor alteration. This has enabled its characterization using a range of techniques, including NMR, NRVS, Mössbauer, EPR and FTIR spectroscopy.^{14–16,18,21,22} In native, fully functional, [FeFe]-hydrogenases the accumulation of a highly similar H_{hyd} state is commonly achieved by exposure to H_2 at low pH values in the presence of the non-physiological reductant sodium dithionite (NaDT, Table S1, ESI†) (with exceptions^{19,22–24}).^{16,17,19,22,25,26} Based on chronoamperometry experiments it was recently proposed that [FeFe]-hydrogenases are inhibited by NaDT at low pH values (or rather SO_2 , one of its oxidized by-products), *i.e.* conditions similar to H_{hyd} accumulation.²⁷ Considering its in-depth spectroscopic characterization, and proposed central importance in the catalytic cycle, the possibility that the to-date characterized H_{hyd} -state reflects an inhibited “artefact-state” would represent a significant setback in our mechanistic understanding of [FeFe]-hydrogenase.

Here, we selectively enrich the double reduced states, H_{hyd} and H_{sred} , by exposure to molecular hydrogen and follow the respective absorbance changes of cofactor ligand bands by Attenuated Total Reflection Fourier-transform Infrared (ATR-FTIR) Spectroscopy. Data is collected at different pH values and buffers. This study reveals that small carboxylic acids, often used in low pH buffers in enzyme electrochemistry experiments, are non-innocent in formation of a new H-cluster species similar to H_{ox} . At present it is not clear if this is an artefactual off-cycle state, or in fact a critical intermediate that previously have escaped detection. Moreover, we show the accumulation of H_{hyd} in native [FeFe]-hydrogenase, HydA1 from *Chlamydomonas reinhardtii*, at mildly basic and low pH values regardless of buffer choice and more importantly in the absence of NaDT. Our findings are well in accordance with the previously characterized H_{hyd} -state as a reaction intermediate rather than an experimental artefact.

A solution of the HydA1 [FeFe]-hydrogenase was applied to the crystal surface of the ATR-FTIR setup, following removal of any NaDT contaminations by chromatography and buffer exchange. The enzyme solution was subsequently dried and rehydrated under N_2 atmosphere to form an auto-oxidized protein film.^{28,29} The hydrogenase adopted the expected oxidized active-ready state H_{ox} , and was subsequently exposed to H_2 gas. The resulting difference spectra are displayed in Fig. 2. As a consequence of H_2 uptake, a mixture of the hydride state H_{hyd} (blue positive bands, 2081, 2066, 1979, 1960, 1860 cm^{-1}) and the super-reduced state H_{sred} (red positive bands, 2070, 2025, 1954, 1918, 1882 cm^{-1}) was populated while the oxidized state H_{ox} (grey negative bands, 2089, 2071, 1965, 1940, 1802 cm^{-1}) was depopulated. This holds true for different pH values, in a range of different buffers. Fig. 2 displays the ATR-FTIR difference

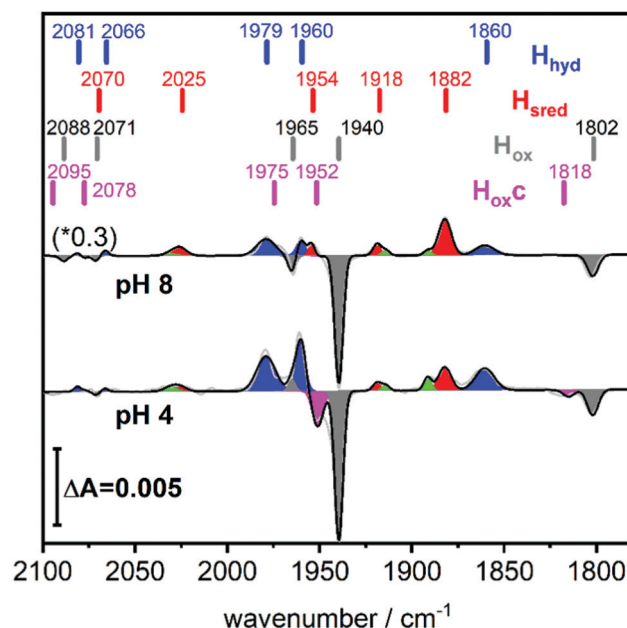


Fig. 2 Hydrogen uptake induced ATR-FTIR difference spectra of the CO/CN ligands of HydA1. Upon exposure to H_2 the oxidized state H_{ox} (grey negative bands) is depopulated while double reduced redox states H_{sred} (red positive bands) and H_{hyd} (blue positive bands) accumulate. For both pH values, pH 8 (top, 100 mM Tris buffer, scaled by 0.3) and pH 4 (bottom, 100 mM propionate buffer), the formation of H_{hyd} (blue) is observed in the absence of NaDT. Magenta negative bands at 1952 and 1815 cm^{-1} indicate the newly observed, unknown H-cluster species denoted H_{oxC} . Green bands belong to H_{red} . The band positions are indicated by bars.

spectra of HydA1 samples adjusted to pH 8 or pH 4 and subsequently exposed to H_2 (absolute spectra and pH adjustment in Fig. S1, ESI†). In each spectrum the signature of H_{hyd} (blue positive bands) is clearly detectable. A larger fraction of H_{sred} is formed at pH 8 (100 mM Tris buffer) while a larger fraction of H_{hyd} is populated at pH 4 (100 mM propionate buffer), in both cases a concomitant loss of H_{ox} is observed in contrast to earlier results where a depopulation of H_{ox} was observed at pH 4 due to the presence of NaDT.^{9,10,16} However, the preference for H_{hyd} accumulation at low pH is clearly independent of the presence of NaDT. As H_{hyd} and H_{sred} are generally considered to reflect two different tautomers, their relative stability as a function of pH value is arguably a consequence of the protonation state of the H-cluster surroundings. The reaction of auto-oxidized HydA1 films with H_2 gas proceeds within one-two seconds while auto-oxidation, observed when H_2 was replaced by N_2 , occurs over 10–20 seconds (compare Fig. S2, ESI†). Both kinetics report indirectly on the gas accessibility and hydration of the protein film in the ATR-FTIR setup. Dehydrated [FeFe]-hydrogenase samples were reported to be protected from exposure to gases.³⁰

Fig. 2 (bottom) shows that H_2 exposure causes H_{hyd} accumulation in the absence of NaDT preferentially at low pH values (compare Fig. 2 top). This trend had been reported by several groups before, however always involving NaDT.^{16,17,19,25} Apart from the recent proposed binding of NaDT (or one of its oxidized products) to the H-cluster, protonation has been suggested to stabilize the reduction at the [4Fe4S] cluster for



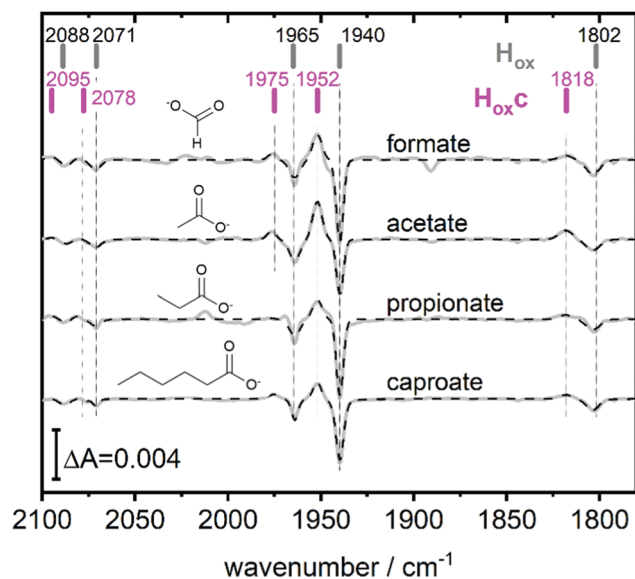


Fig. 3 The full spectroscopic signature of $H_{ox}C$. Difference spectra highlighting the full band pattern of $H_{ox}C$ (positive bands) for transitions from pH 8 (10 mM Tris, negative bands) to pH 4 with, from top to bottom, 100 mM formate, 100 mM acetate, 100 mM propionate and 100 mM caproate buffer. Data presented in grey and fit in dashed black. The band positions of H_{ox} and $H_{ox}C$ are indicated by bars. Acetate and propionate spectrum scaled by 2 and 0.5 respectively.

H_{red} ^{9,10,31} and a similar stabilisation effect was reported for H_{hyd} from both purely computational and spectroscopic approaches.^{32,33} Our findings verify that NaDT is not required to stabilize H_{hyd} . The observed pH dependent shift between H_{sred} and H_{hyd} instead supports a model of a protonatable site, either near the reduced [4Fe4S] cluster as previously proposed,^{3,31,34} or *e.g.* in the PTP. Residual traces of NaDT have been shown to lead to population of $H_{ox}H$ (2089, 2079, 1970, 1945, 1810 cm^{-1}),²⁷ which is absent in our spectra. The negative bands in the pH 4 difference spectrum (Fig. 2 bottom) at 1952 and 1815 cm^{-1} belong to an unknown band pattern significantly different from $H_{ox}H$ (full spectroscopic signature Fig. 3), and we denote this new state as $H_{ox}C$.

This blue shifted H_{ox} population was observable with pH 4 buffers composed of monocarboxylic acids, *i.e.* formic acid ($pK_{a1} = 3.8$), acetic acid ($pK_a = 4.8$), propionic acid ($pK_{a1} = 4.9$) and caproic acid ($pK_{a1} = 4.9$) (Fig. 3). Conversely, di- and tricarboxylic acids such as succinic acid ($pK_{a1} = 4.2$), oxalic acid ($pK_{a2} = 4.14$) or citric acid ($pK_{a1} = 3.1$) did not induce the new species (Fig. S3, ESI†). Thus, stabilizations of $H_{ox}C$ appear to be determined by the number of the carboxylic acid groups, rather than molecular weight of the acid.

Compared to H_{ox} the overall band pattern of $H_{ox}C$ is retained and in analogy to H_{red} (red shifted) and $H_{ox}H$ (blue shifted) we propose that the H-cluster geometry and electronic structure is highly similar. Moreover, the requirement for low pH in forming the state implies that the buffer molecule interacts in its protonated, uncharged, form (Fig. S4, ESI†). A complete characterisation of this new H-cluster state is beyond the scope of the current study, but we note that it could be of high relevance

in particular as *e.g.* acetic acid (acetate) is commonly employed in protein film electrochemistry studies of [FeFe]-hydrogenase.^{27,35–45} Formaldehyde has previously been observed to act as an inhibitor of the H-cluster under reducing conditions.^{46,47} In the latter case, binding of formaldehyde on the [2Fe] subsite *via* a Fe–C bond was proposed from a combination of ENDOR spectroscopy and DFT calculations,⁴⁸ and further supported by studies of model complexes.⁴⁹ Whether a similar binding model can be applied also to carboxylic acids remains to be verified. The extent of the new state ($H_{ox}C$) formed in electrochemistry setups, its role in catalysis and its implication on catalytic currents, especially at low pH, remain unclear. In contrast to the H_{ox} state, the same H_{hyd} state forms regardless of the nature of the buffer, rendering a stabilizing role of the buffer or other additives unlikely (Fig. 2 and Fig. S1, ESI†). Our data shows clearly that H_{hyd} can be accumulated in the absence of NaDT and without introducing loss-of-function mutations. In closing, this underscores the flat energy landscape of the H-cluster during catalysis, and it is tempting to conclude that the previously characterized H_{hyd} state indeed represents one of the final intermediates in hydrogen evolution catalysis of [FeFe]-hydrogenases.

Investigation by M. S. and T. K., M. L. (enzyme purification), H. J. R. (cofactor synthesis), writing (original draft) by M. S., writing (review & editing) M. S., T. K., M. L., H. J. R. and G. B.

The authors would like to thank Leif Hammarström for critically reviewing the manuscript. Open access funding was provided by Uppsala University. The European Research Council (ERC, to GB, Contract No. 714102) as well as the European Union's Horizon 2020 research and innovation program (Marie Skłodowska Curie Grant No. 897555 to M.S.) are gratefully acknowledged for funding.

Conflicts of interest

There are no conflicts to declare.

References

- J. T. Kleinhaus, F. Wittkamp, S. Yadav, D. Siegmund and U. P. Apfel, *Chem. Soc. Rev.*, 2021, **50**, 1668–1784.
- H. Land, M. Senger, G. Berggren and S. T. Stripp, *ACS Catal.*, 2020, **10**, 7069–7086.
- F. Wittkamp, M. Senger, S. T. Stripp and U. P. Apfel, *Chem. Commun.*, 2018, **54**, 5934–5942.
- M. Winkler, J. Esselborn and T. Happe, *Biochim. Biophys. Acta*, 2013, **1827**, 974–985.
- J. Duan, M. Senger, J. Esselborn, V. Engelbrecht, F. Wittkamp, U. P. Apfel, E. Hofmann, S. T. Stripp, T. Happe and M. Winkler, *Nat. Commun.*, 2018, **9**, 4726.
- M. Senger, V. Eichmann, K. Laun, J. Duan, F. Wittkamp, G. Knor, U. P. Apfel, T. Happe, M. Winkler, J. Heberle and S. T. Stripp, *J. Am. Chem. Soc.*, 2019, **141**, 17394–17403.
- H. Land, A. Sekretareva, P. Huang, H. J. Redman, B. Nemeth, N. Polidori, L. S. Meszaros, M. Senger, S. T. Stripp and G. Berggren, *Chem. Sci.*, 2020, **11**, 12789–12801.
- H. Land, P. Ceccaldi, L. S. Meszaros, M. Lorenzi, H. J. Redman, M. Senger, S. T. Stripp and G. Berggren, *Chem. Sci.*, 2019, **10**, 9941–9948.
- M. Senger, K. Laun, F. Wittkamp, J. Duan, M. Haumann, T. Happe, M. Winkler, U. P. Apfel and S. T. Stripp, *Angew. Chem., Int. Ed.*, 2017, **56**, 16503–16506.



- 10 M. Senger, S. Mebs, J. Duan, O. Shulenina, K. Laun, L. Kertess, F. Wittkamp, U. P. Apfel, T. Happe, M. Winkler, M. Haumann and S. T. Stripp, *Phys. Chem. Chem. Phys.*, 2018, **20**, 3128–3140.
- 11 S. Mebs, M. Senger, J. Duan, F. Wittkamp, U. P. Apfel, T. Happe, M. Winkler, S. T. Stripp and M. Haumann, *J. Am. Chem. Soc.*, 2017, **139**, 12157–12160.
- 12 C. Sommer, A. Adamska-Venkatesh, K. Pawlak, J. A. Birrell, O. Rudiger, E. J. Reijerse and W. Lubitz, *J. Am. Chem. Soc.*, 2017, **139**, 1440–1443.
- 13 A. Silakov, C. Kamp, E. Reijerse, T. Happe and W. Lubitz, *Biochemistry-Us*, 2009, **48**, 7780–7786.
- 14 D. W. Mulder, Y. Guo, M. W. Ratzloff and P. W. King, *J. Am. Chem. Soc.*, 2017, **139**, 83–86.
- 15 E. J. Reijerse, C. C. Pham, V. Pelmenchikov, R. Gilbert-Wilson, A. Adamska-Venkatesh, J. F. Siebel, L. B. Gee, Y. Yoda, K. Tamasaku, W. Lubitz, T. B. Rauchfuss and S. P. Cramer, *J. Am. Chem. Soc.*, 2017, **139**, 4306–4309.
- 16 M. Winkler, M. Senger, J. Duan, J. Esselborn, F. Wittkamp, E. Hofmann, U. P. Apfel, S. T. Stripp and T. Happe, *Nat. Commun.*, 2017, **8**, 16115.
- 17 D. W. Mulder, M. W. Ratzloff, E. M. Shepard, A. S. Byer, S. M. Noone, J. W. Peters, J. B. Broderick and P. W. King, *J. Am. Chem. Soc.*, 2013, **135**, 6921–6929.
- 18 D. W. Mulder, M. W. Ratzloff, M. Bruschi, C. Greco, E. Koonce, J. W. Peters and P. W. King, *J. Am. Chem. Soc.*, 2014, **136**, 15394–15402.
- 19 L. S. Meszaros, P. Ceccaldi, M. Lorenzi, H. J. Redman, E. Pfizner, J. Heberle, M. Senger, S. T. Stripp and G. Berggren, *Chem. Sci.*, 2020, **11**, 4608–4617.
- 20 A. Adamska, A. Silakov, C. Lambert, O. Rudiger, T. Happe, E. Reijerse and W. Lubitz, *Angew. Chem., Int. Ed.*, 2012, **51**, 11458–11462.
- 21 P. Knorzer, A. Silakov, C. E. Foster, F. A. Armstrong, W. Lubitz and T. Happe, *J. Biol. Chem.*, 2012, **287**, 1489–1499.
- 22 M. W. Ratzloff, J. H. Artz, D. W. Mulder, R. T. Collins, T. E. Furtak and P. W. King, *J. Am. Chem. Soc.*, 2018, **140**, 7623–7628.
- 23 S. Morra, J. Duan, M. Winkler, P. A. Ash, T. Happe and K. A. Vincent, *Dalton Trans.*, 2021, **50**, 12655–12663.
- 24 P. S. Corrigan, J. L. Tirsch and A. Silakov, *J. Am. Chem. Soc.*, 2020, **142**, 12409–12419.
- 25 C. Lorent, S. Katz, J. Duan, C. J. Kulka, G. Caserta, C. Teutloff, S. Yadav, U. P. Apfel, M. Winkler, T. Happe, M. Horch and I. Zebger, *J. Am. Chem. Soc.*, 2020, **142**, 5493–5497.
- 26 V. Pelmenchikov, J. A. Birrell, C. C. Pham, N. Mishra, H. Wang, C. Sommer, E. Reijerse, C. P. Richers, K. Tamasaku, Y. Yoda, T. B. Rauchfuss, W. Lubitz and S. P. Cramer, *J. Am. Chem. Soc.*, 2017, **139**, 16894–16902.
- 27 M. A. Martini, O. Rudiger, N. Breuer, B. Noring, S. DeBeer, P. Rodriguez-Macia and J. A. Birrell, *J. Am. Chem. Soc.*, 2021, **143**, 18159–18171.
- 28 M. Senger, S. Mebs, J. Duan, F. Wittkamp, U. P. Apfel, J. Heberle, M. Haumann and S. T. Stripp, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, 8454–8459.
- 29 S. T. Stripp, *ACS Catal.*, 2021, **11**, 7845–7862.
- 30 J. Noth, R. Kositzki, K. Klein, M. Winkler, M. Haumann and T. Happe, *Sci. Rep.*, 2015, **5**, 13978.
- 31 K. Laun, I. Baranova, J. Duan, L. Kertess, F. Wittkamp, U. P. Apfel, T. Happe, M. Senger and S. T. Stripp, *Dalton Trans.*, 2021, **50**, 3641–3650.
- 32 P. E. M. Siegbahn and R. Z. Liao, *J. Phys. Chem. A*, 2020, **124**, 10540–10549.
- 33 S. Mebs, R. Kositzki, J. Duan, L. Kertess, M. Senger, F. Wittkamp, U. P. Apfel, T. Happe, S. T. Stripp, M. Winkler and M. Haumann, *Biochim. Biophys. Acta, Bioenerg.*, 2018, **1859**, 28–41.
- 34 M. Haumann and S. T. Stripp, *Acc. Chem. Res.*, 2018, **51**, 1755–1763.
- 35 P. Rodriguez-Macia, N. Breuer, S. DeBeer and J. A. Birrell, *ACS Catal.*, 2020, **10**, 13084–13095.
- 36 P. Rodriguez-Macia, L. Kertess, J. Burnik, J. A. Birrell, E. Hofmann, W. Lubitz, T. Happe and O. Rudiger, *J. Am. Chem. Soc.*, 2019, **141**, 472–481.
- 37 O. Lampret, A. Adamska-Venkatesh, H. Konegger, F. Wittkamp, U. P. Apfel, E. J. Reijerse, W. Lubitz, O. Rudiger, T. Happe and M. Winkler, *J. Am. Chem. Soc.*, 2017, **139**, 18222–18230.
- 38 L. Kertess, F. Wittkamp, C. Sommer, J. Esselborn, O. Rudiger, E. J. Reijerse, E. Hofmann, W. Lubitz, M. Winkler, T. Happe and U. P. Apfel, *Dalton Trans.*, 2017, **46**, 16947–16958.
- 39 L. Kertess, A. Adamska-Venkatesh, P. Rodriguez-Macia, O. Rudiger, W. Lubitz and T. Happe, *Chem. Sci.*, 2017, **8**, 8127–8137.
- 40 O. Lampret, J. Duan, E. Hofmann, M. Winkler, F. A. Armstrong and T. Happe, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 20520–20529.
- 41 V. Hajj, C. Baffert, K. Sybirna, I. Meynial-Salles, P. Soucaille, H. Bottin, V. Fourmond and C. Leger, *Energy Environ. Sci.*, 2014, **7**, 715–719.
- 42 M. Del Barrio, M. Sensi, L. Fradale, M. Bruschi, C. Greco, L. de Gioia, L. Bertini, V. Fourmond and C. Leger, *J. Am. Chem. Soc.*, 2018, **140**, 5485–5492.
- 43 M. Winkler, J. Duan, A. Rutz, C. Felbek, L. Scholtysek, O. Lampret, J. Jaenecke, U. P. Apfel, G. Gilardi, F. Valetti, V. Fourmond, E. Hofmann, C. Leger and T. Happe, *Nat. Commun.*, 2021, **12**, 756.
- 44 S. Hardt, S. Stapf, D. T. Filmon, J. A. Birrell, O. Rudiger, V. Fourmond, C. Leger and N. Plumere, *Nat. Catal.*, 2021, **4**, 251–258.
- 45 A. Fasano, H. Land, V. Fourmond, G. Berggren and C. Leger, *J. Am. Chem. Soc.*, 2021, **143**, 20320–20325.
- 46 A. F. Wait, C. Brandmayr, S. T. Stripp, C. Cavazza, J. C. Fontecilla-Camps, T. Happe and F. A. Armstrong, *J. Am. Chem. Soc.*, 2011, **133**, 1282–1285.
- 47 C. E. Foster, T. Kramer, A. F. Wait, A. Parkin, D. P. Jennings, T. Happe, J. E. McGrady and F. A. Armstrong, *J. Am. Chem. Soc.*, 2012, **134**, 7553–7557.
- 48 A. Bachmeier, J. Esselborn, S. V. Hexter, T. Kramer, K. Klein, T. Happe, J. E. McGrady, W. K. Myers and F. A. Armstrong, *J. Am. Chem. Soc.*, 2015, **137**, 5381–5389.
- 49 F. Zhang, T. J. Woods, L. Zhu and T. B. Rauchfuss, *Chem. Sci.*, 2021, **12**, 15673–15681.

