


 Cite this: *RSC Adv.*, 2020, 10, 27103

Xanthan gum derivatives: review of synthesis, properties and diverse applications

 Jwala Patel,^a Biswajit Maji,^b N. S. Hari Narayana Moorthy^a and Sabyasachi Maiti ^{*a}

Natural polysaccharides are well known for their biocompatibility, non-toxicity and biodegradability. These properties are also inherent to xanthan gum (XG), a microbial polysaccharide. This biomaterial has been extensively investigated as matrices for tablets, nanoparticles, microparticles, hydrogels, buccal/transdermal patches, tissue engineering scaffolds with different degrees of success. However, the native XG has its own limitations with regards to its susceptibility to microbial contamination, unusable viscosity, poor thermal and mechanical stability, and inadequate water solubility. Chemical modification can circumvent these limitations and tailor the properties of virgin XG to fulfill the unmet needs of drug delivery, tissue engineering, oil drilling and other applications. This review illustrates the process of chemical modification and/crosslinking of XG via etherification, esterification, acetalation, amidation, and oxidation. This review further describes the tailor-made properties of novel XG derivatives and their potential application in diverse fields. The physicochemical modification and its impact on the properties of XG are also discussed. Overall, the recent developments on XG derivatives are very promising to progress further with polysaccharide research.

Received 16th May 2020

Accepted 13th July 2020

DOI: 10.1039/d0ra04366d

rsc.li/rsc-advances

1. Introduction

Polysaccharides are isolated from renewable sources and have nontoxic, biocompatible, biodegradable, and bioadhesive properties. These qualities account for their use in food, pharmaceutical, biomedical and cosmetic applications. Among them, xanthan gum (XG) has attracted considerable interest in the past two decades due to its safety clearance by the US FDA as food additive in 1969.¹ XG is a high molecular weight (2×10^6 to 20×10^6 Da) fermentation product of the Gram-negative bacterium *Xanthomonas campestris*.^{2,3} Chemically, it consists of β -1,4-D-glucopyranose glucan backbone with a pendant trisaccharide side chain, composed of mannose (β -1,4), glucuronic acid (β -1,2) and terminal mannose residues. The side chain is attached to alternate glucose residue in the backbone by α -1,3 linkages. In the side chain, the terminal mannose moiety is partially substituted with a pyruvate residue linked as an acetal to the 4- and 6-positions; the internal mannose sugar closest to the backbone is acetylated at C-6. The O-acetyl and pyruvate residues deprotonate at pH > 4.5, increase charge density along the xanthan chains, thus enabling XG for Ca²⁺ ion-mediated physical crosslinking.^{4,5} The presence of glucuronic acid in side chain gives birth to the polysaccharide

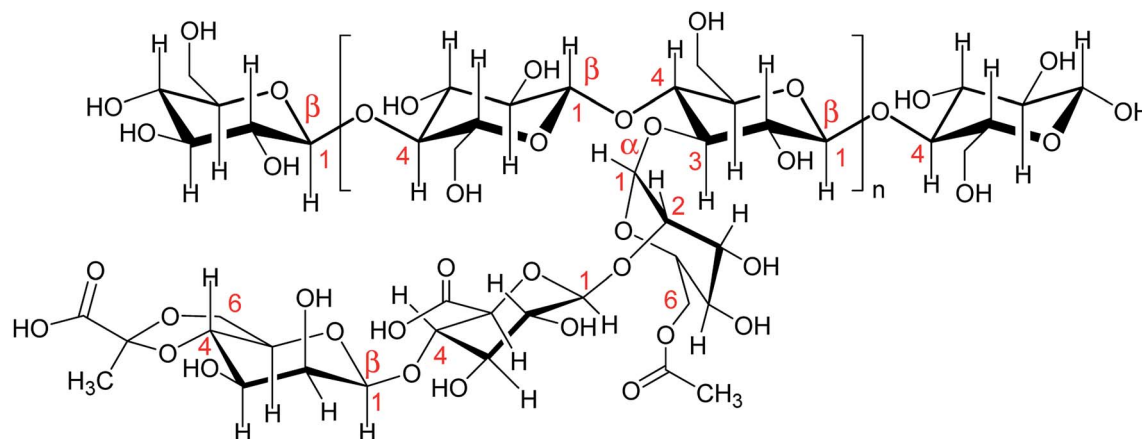
a polyanionic character. Chemical structure of XG is illustrated in Fig. 1a and b.

The hydroxy and carboxy polar groups of XG reconnoiter intramolecular and intermolecular hydrogen bonding interactions in aqueous solution.⁶ Due to high molecular weight and hydrogen bonding interactions, an aqueous solution of XG exhibits high intrinsic viscosity at low concentration,⁷ and behaves like a pseudoplastic fluid.⁸ High viscous rheology and pH- and salt-resistant properties of XG are satisfactory for use as thickening agent, suspending agent,^{9,10} and stabilizers for food, pharmaceuticals, cosmetics^{11–14} and drag reducer in oil drilling.¹⁵ XG assumes disorder conformation under low ionic strength or high temperature;¹⁶ however, the XG present themselves either in single or double helix conformations under reverse condition.¹⁷ As a consequence of 3-D association of XG chains, its aqueous solution confers weak gel-like properties.¹⁸ However, XG never forms true gels at any concentration,¹⁹ attributable to its weak, non-covalent intermolecular interactions. In comparison to gels, hydrogels are crosslinked 3-D networks of hydrophilic polymers which swell in water or biological fluids but do not dissolve. Hydrogels possess some other interesting characteristics such as biocompatibility, softness which put them forward as carriers for drug or proteins delivery.²⁰ Cross-linking of XG can be done either by physical or chemical methods.^{21,22} Physical cross-linking is a well-established method to fabricate XG-based hydrogels.²³ In this method, the concentrated polysaccharide is cross-linked through hydrogen bonding,²⁴ electrostatic ionic force,²⁵ π - π stacking,²⁶ hydrophobic interaction, or host-guest inclusion²⁷

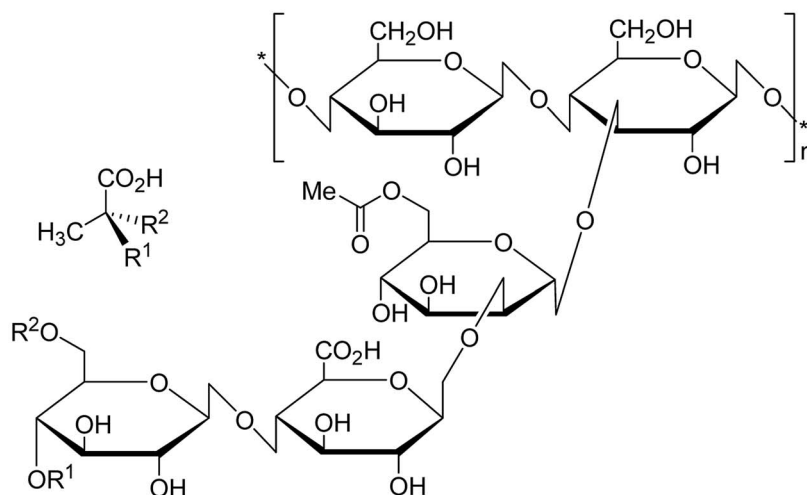
^aDepartment of Pharmacy, Indira Gandhi National Tribal University, Amarkantak, Madhya Pradesh-484887, India. E-mail: sabya245@rediffmail.com; Tel: +91 9474119931

^bDepartment of Chemistry, Indira Gandhi National Tribal University, Amarkantak, Madhya Pradesh-484887, India

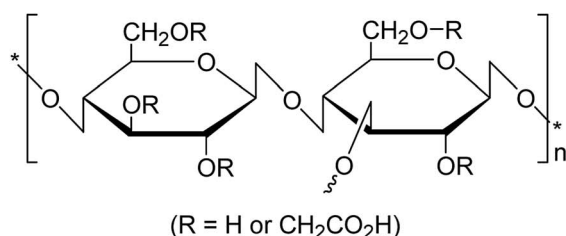
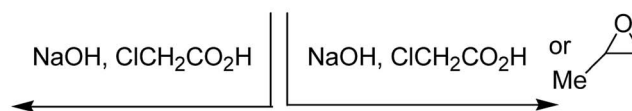




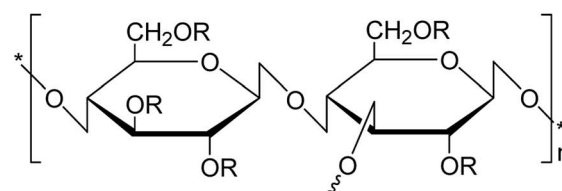
(a) Xanthan Gum (Chair)



(b) Xanthan Gum (Haworth Projection)

(R = H or CH₂CO₂H)

(c) O-Carboxymethyl XG

(R = H or CH₂CO₂H) or

(d) Carboxymethyl hydroxypropyl XG

Fig. 1 Chemical structure of (a) XG (chair conformation); (b) XG (Haworth projection); (c) carboxymethyl XG; (d) carboxymethyl hydroxypropyl XG.

to obtain hydrogels. However, the physically-formed hydrogels suffer from poor mechanical strength, perhaps due to intermolecular secondary interactions, generally weaker than

covalent bonds.²⁸ Further, the lack of thermal stability, pH or salt stability on aging limits their applications. On heating or dilution, XG physical gels are easily solubilized in aqueous



solution²⁹ and thus, they are not suitable as controlled drug release carrier. In contrast, the coordination bonding between XG polymer chains and trivalent metal ions leads to stable gels *via* dimeric or polymeric ionic bridges.^{30,31}

XG has also been put forward as a component of hydrogel preparations with other polymers such as alginate, locust bean gum, chitosan in the form of microparticles^{32,33} and nanoparticles.^{34,35} One fascinating example is the combination of XG with locust bean gum. In equal proportion, they can form a firm, thermoreversible gel in aqueous medium through synergistic interaction.³⁶ This observation captivates this polymer combination for use as control drug release carriers.³⁷

It is noteworthy that XG can act as a stabilizer for the synthesis of gold nanoparticles. The presence of mannose sugars in XG further assist in the delivery of chemotherapeutic agents/siRNA/shRNA to mannose receptor overexpressing cancer cells.³⁵ Nonetheless, XG-stabilized silver nanoparticles have been studied for their antibacterial and catalytic applications.³⁸ As a coating material, XG appears promising for the production of iron nanoparticles of 20–80 nm diameters.³⁹ Recently, liposomes coated with chitosan–XG have been proposed as potential carriers for direct drug administration to lungs due to their improved liposomal stability and mucoadhesion properties.^{40,41}

Native XG has been extensively studied as matrix forming materials in the design of tablets either alone^{42–47} or in combination with other natural polymers such as guar gum,⁴⁸ alginate,^{49,50} cashew gum,⁵¹ konjac glucomannan,⁵² *Gleditsia sinensis* Lam. galactomannan,⁵³ *Mimosa scabrella* galactomannan⁵⁴ for the controlled delivery of various kinds of drugs. Chitosan and XG is an entrancing binary composition. The anionic gum electrostatically interacts with chitosan and forms polyelectrolyte complexes which serve as release-retardant for drugs.^{55,56} Even, the biocompatibility of chitosan–xanthan polyionic complex favors its pharmaceutical application.⁵⁷

XG is not digested in human stomach or small intestine, but degraded in presence of colon enzymes.⁵⁸ Thus, XG matrices can shield the drug from the environments of stomach and small intestine and deliver the drug to the colon. On reaching colon, they are assimilated by the anaerobic microflora of the colon, for example, bacteroides, bifidobacteria species and eubacteria, to smaller monosaccharides, which are then either used as energy source by the bacteria or degraded by enzyme. Keeping this in view, XG-based colon drug delivery systems are devised and tested for possible colon targeting of drugs. The systems are not limited to XG⁵⁹ but include other polymers such as konjac glucomannan,⁶⁰ chitosan/eudragit,⁶¹ guar gum⁶² as a component.

XG has an excellent biomimicking potential for tissue engineering applications such as bone, cartilage, and skin regeneration. XG-based hydrogels composed of other natural polymers and nanohydroxyapatite are fabricated and tested for tissue engineering applications^{63,64} and cellular uptake studies.⁶⁵ In addition, the use of XG with electroactive polypyrrole has shown significant improvement in fibroblast proliferation.⁶⁶ Kumar *et al.*⁶⁷ found that XG-based hybrid

scaffold system supported cell adhesion and proliferation of preosteoblast cells and thus seemed suitable for tissue engineering applications.

Its topical drug delivery application *via* intranasal, ocular, buccal and transdermal routes is also noteworthy. With large surface area, porous endothelial membrane and high total blood flow bypassing hepatic, gut wall metabolism and/or destruction in the gastrointestinal tract,⁶⁸ the intranasal route appears to be a reliable alternative to oral and parenteral route for drug delivery. For example, XG-based *in situ* nasal gel has shown its unique ability to enhance drug permeation, bioavailability, and deliver drug directly to brain *via* olfactory lobe pathway.^{69–72} In context of ocular therapy, the major problem lies in attaining optimal drug concentration at the site of action. This is largely due to precorneal loss as a consequence of blinking of eyes.⁷³ *In situ* gel forming system appears to be the solution to this problem. The ocular bioavailability can be enhanced by increasing pre-corneal retention time. Accordingly, *in situ* ocular gels of XG in combination with other polymers such as poloxamer-407–guar gum,⁷⁴ HPMC K4M,⁷⁵ alginate–carbopol 934,⁷⁶ gellan gum,^{77,78} alginate–ethylcellulose,⁷⁹ poloxamer 407/188 (ref. 80) have been tested. Ion-induced nanoemulsion-based *in situ* gelling system of XG with HPMC or carbopol is also examined.⁸¹ Reports on *in situ* vaginal gels for eradication of candidiasis⁸² and *in situ* gel forming tablets⁸³ are also evident. XG-based ophthalmic liquids,⁸⁴ vaginal gels,^{82,85} bioadhesive liquids laden with lipid nanoparticles,⁸⁶ microemulsion-based hydrogel⁸⁷ and additives for spray-drying and freeze-drying processes^{88,89} are reported as well.

There is an expanse of smooth muscle and immobile mucosa in buccal cavity, which sets a platform for delivering drugs directly to the systemic circulation through the internal jugular vein, thus avoiding hepatic metabolism and acid hydrolysis in the gastrointestinal tract.⁹⁰ Moreover, rapid mucosal cell recovery is attractive for opting buccal route for drug delivery purpose.⁹¹ These inspired researchers to develop nicotine patches for smoking cessation,⁹² desloratidine patches for treating allergic rhinitis⁹³ and zolmitriptan patches for migraine treatment.⁹⁴ Transdermal patches further provide benefits in terms of reduced first-pass metabolism, frequency of dosing, side effects secondary to gastrointestinal intolerance and fluctuations in drug levels.⁹⁵ Transdermal films/patches of XG with other polymers such as alginate,^{96,97} hydroxypropylmethyl cellulose–carboxymethyl cellulose⁹⁸ are reported for analgesic and antihypertensive drugs.

The beneficial effects after oral administration of XG on immune-surveillance against neoplasms are also reported.⁹⁹ Low-molecular-weight XG (4.07×10^4 Da) has shown antioxidant and excellent protective effect on H₂O₂-injured Caco-2 cells, suggesting its use in food/pharmaceuticals in order to alleviate/resist oxidative damage induced by overproduction of reactive oxygen species.¹⁰⁰ XG is a strong inhibitor of oil peroxidation¹⁰¹ and antioxidant for human corneal epithelial cells at 0.2% strength.¹⁰²

Despite numerous potential benefits, native XG poses some problems which include microbial contamination, unstable



viscosity, poor shear resistance, inadequate mechanical, thermal properties and uncontrolled rate of hydration. These limit its applications in food, pharmaceutical and biomedical fields. Further, XG dissolves at a slow rate particularly in cold water at high concentration. Upon dispersion, it forms lumps in water called fish eyes¹⁰³ due to its improper hydration. Consequently, a gelatinous outer layer is formed on surface of the particles, immediately after dispersion. This gelatinous layer prohibits water penetration and hinders complete dissolution of the particles.¹⁰⁴ Due to insufficient gelling, XG itself fails to afford self-sustainable, stable hydrogel beads in presence of metal ions. XG possesses a number of hydroxy and carboxy groups which are amenable for chemical modification with successive improvement in its physicochemical properties especially solubility, swelling and metal-induced gelling ability; mechanical and thermal stability. The chemical derivatives then become widely acceptable for diverse applications. As extracted from scientific literatures, the chemical modification of XG involves etherification, esterification, acetalation, oxidation, peptide linking, ionic and covalent crosslinking, and mechanical modification. However, there are no decent reviews that sum up the data on modification procedures of XG, resultant properties and the diverse application of XG derivatives.

Though some reviews cover industrial production,^{105–107} and drug delivery and biomedical applications of XG,^{108–111} these are not adequate. In 2003, Badwaik *et al.*¹¹² published a review on XG and its derivatives which discussed the synthesis of acetalated and carboxymethyl derivatives of XG, microwave and plasma assisted grafting of XG. The authors mostly focused on drug delivery application. However, this review lacks significant, in-depth and detailed up-to-date information on synthetic procedures, properties and diverse applications of modified xanthan polysaccharide. Till date, no review article describes XG derivatives in a comprehensive manner in terms of their detailed synthetic/crosslinking procedures, tailored properties beneficial for industrial applications in the field of pharmaceutical and biomedical science and technology. This review attempts to cover these aspects of XG derivatives to fulfill the unmet needs of students, researchers, and budding scientists in these fields.

2. Modified xanthan materials

2.1. Etherified XG

The most common and widely investigated ether derivatives of XG is the *O*-carboxymethyl XG. Carboxymethylation is a simple chemical etherification reaction which requires simple chemicals like monochloroacetic acid and sodium hydroxide (Fig. 1c). The degree of *O*-carboxymethyl substitution may vary proportionately with chloroacetic acid : XG ratio within 0.5 to 3.¹¹³ The viscosity, elastic modulus and molecular weight of XG are lessened after modification. The derivative becomes more hydrophilic than native XG with gradual increase in degree of substitution. However, the shear-thinning and weak gel behaviors of XG are not affected after carboxymethylation. In contrast, an aqueous solution of carboxymethyl hydroxypropyl XG (Fig. 1d) exhibits greater viscosity, elastic modulus, and

temperature- and shear-resistance than XG solution at the same strength. A better proppant carrying ability makes this derivative suitable for use as an additive of fracturing fluids.¹¹⁴

Similar viscoelastic behaviors are noticed for cationic and amphoteric XG derivatives, synthesized using *N*-(3-chloro-2-hydroxypropyl)trimethyl ammonium chloride^{115,116} and 3-chloro-2-hydroxypropyldimethylhexadecylammonium acetate,¹¹⁷ respectively. The amphoteric XG solution appears to be a better drag reducer than virgin XG solution. Oleamidopropyl dimethylamine- and triisopropanolamine-modified amphoteric derivatives of xanthan have been reported for their high viscosity (Fig. 2).¹¹⁸

The etherified derivatives of XG have been assessed for their quality attributes in the form of hydrogel films. Alupej *et al.*¹¹⁹ synthesized epichlorohydrin-crosslinked poly (vinyl alcohol) (PVA)/XG supersorbent hydrogel films (1 mm thickness) under basic medium and appraised their controlled swelling *in vitro*. Interestingly, higher polymer : crosslinker ratio (5 : 1) allowed swelling of the hydrogels up to ~2500%. Dhar and collaborators¹²⁰ highlighted the importance of pH on crosslinking reaction of epichlorohydrin with PVA/XG. They perceived that labile ester linkages predominated in the composite films at pH < 4; whereas both ether and ester bonds were registered over pH 7.0–7.5 and consequently, manifested different degrees of swelling in water. They argued that the films synthesized at pH > 9 were mechanically strong due to ether linkages and thus swelled progressively. Incorporation of XG in PVA films caused lesser swelling of the composite films and proved useful in controlling release of uric acid over 24 h at pH 7.4 (Fig. 3).

Zhang *et al.*¹²¹ spotted highly viscous, thixotropic, viscoelastic and thermoresistant characteristics of XG after crosslinking under alkaline condition at epichlorohydrin : XG weight ratio of 4 : 200, beyond which these additive effects nullified. The crosslinking effect of epichlorohydrin on theophylline loading and release from xanthan/chondroitin sulfate (1 : 1) hydrogels was assessed by Oprea *et al.*¹²² They noted that about 78% theophylline loading was possible by diffusion filling method. The hydrogels swelled within a minute but thereafter absorbed simulated gastric and intestinal fluids at a slower rate. The inclusion of chondroitin sulfate in the hydrogels suppressed drug release by 34% in acidic fluid (pH 2.2) in 7 h. However, they spotted a reverse phenomenon in intestinal fluid (pH 7.4). An enhanced oral bioavailability of 323% clearly provided an indication of preclinical success of this hydrogel system.

Maiti and groups¹²³ proposed hexadecyl etherification of XG as a means of enhancing solubility of glibenclamide through polymeric micellization process. The copolymer provided 122-fold higher aqueous solubility than pristine drug. The micelles had a command on the drug release in simulated biological fluids up to 8 h and on diabetes in alloxan-induced hyperglycemic rabbit model. The enmeshing of drug-loaded copolymer micelles into *O*-carboxymethyl aluminium xanthan hydrogel particles declined drug release rate in simulated fluids. They discerned better hypoglycemic activity of glibenclamide after trapping micelles into carboxymethyl XG hydrogels. The glucose-lowering action lasted up to 8 h in alloxan-induced



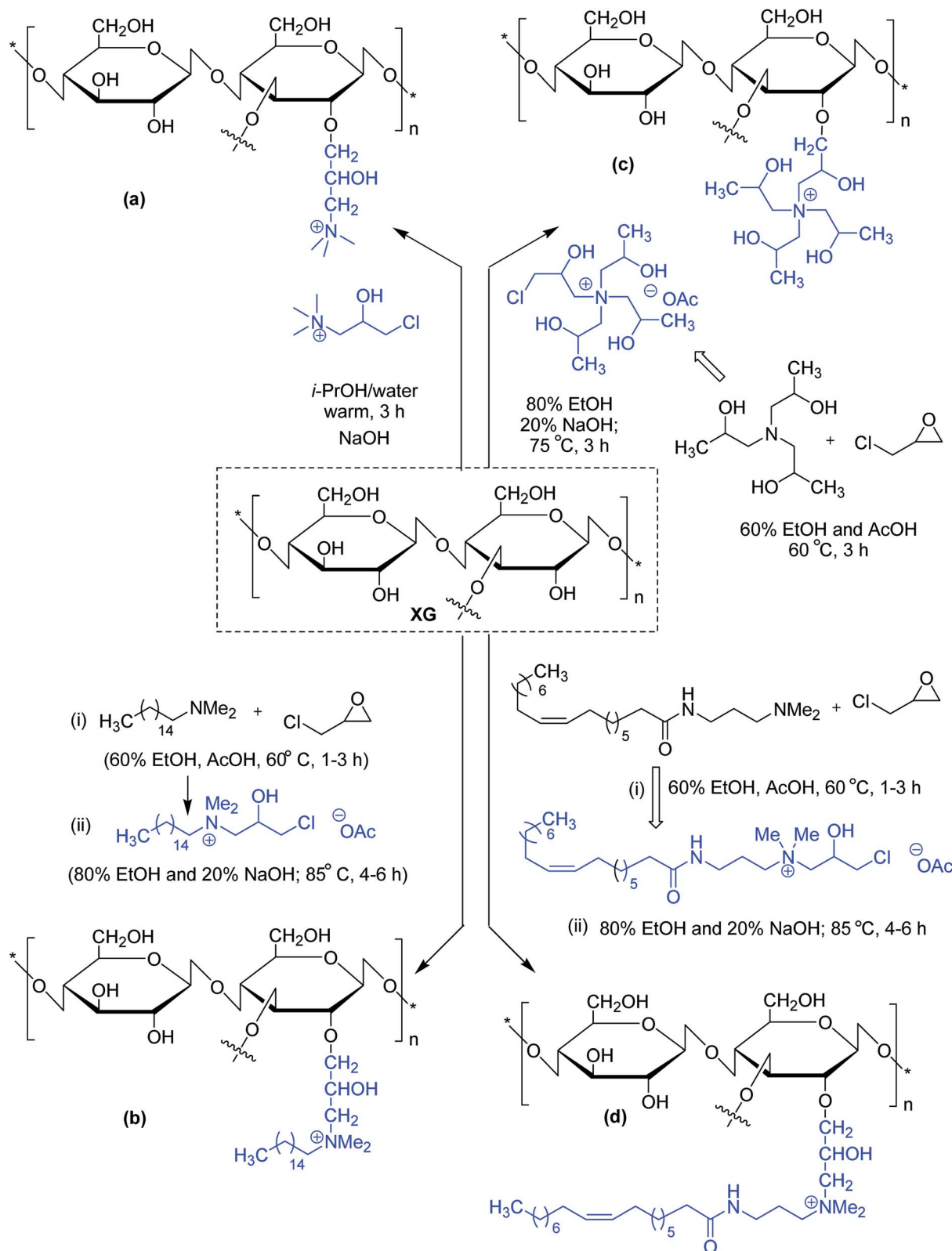


Fig. 2 Synthesis of cationic and amphoteric XG derivatives.

hyperglycemic rabbits.¹²⁴ Quan *et al.*¹²⁵ conjugated hexadecyl groups to XG under its ordered conformation, where its structure remains altered. Intriguing to note that, this derivative

associated at a very low concentration (0.16%) in water, and showed better thermal resistance, and 4-fold higher viscosity. Altogether the hydrophobic modification of XG under order



conformation contributed significantly to these changes in XG properties.¹²⁵

The substitution of 4- or 8-tetradecyl chains to XG assisted in denser association of the polymer network, resulting in higher solution viscosity. Greater hydrophobic interaction amongst the polymer chains was presumed for this close association at higher polymer concentration and alkyl substitution, though the temperature had a negative impact¹²⁶ (Fig. 4).

Bakshi *et al.*¹²⁷ found that solid dispersion prepared by hot melt process in combination with microwave irradiation resulted in 75-fold increase in glibenclamide solubility and better preclinical hypoglycemic activity at a drug : Soluplus® ratio of 1 : 7. This observation invigorated them to incorporate the solid dispersion into carboxymethyl XG mini-matrices in order to circumvent the fluctuations in dissolution rate associated with

pure drug under gastrointestinal fluids. Carboxymethyl XG matrices containing Soluplus gradually released the drug up to 70% till the end of 8 h without any burst effect. On contrary, XG matrices showed burst release of drug and corresponded to 100% in 6 h. It was noteworthy that the solubility enhancement caused by Soluplus dispersion was about 50-fold lower than that reported for hexadecyl xanthan copolymer.¹²⁷

Ahuja *et al.*¹²⁸ proceeded with synthesis and physical characterization of the carboxymethyl XG. They reported globular shape morphology and crystalline nature of carboxymethyl XG particles. The viscosity deteriorated after carboxymethylation. Carboxymethyl XG matrices revealed faster release of diclofenac sodium compared to unmodified XG. For brevity, the carboxymethyl XG matrices emptied its entire content in 3 h; while unmodified XG released half of its content in 16 h. It was

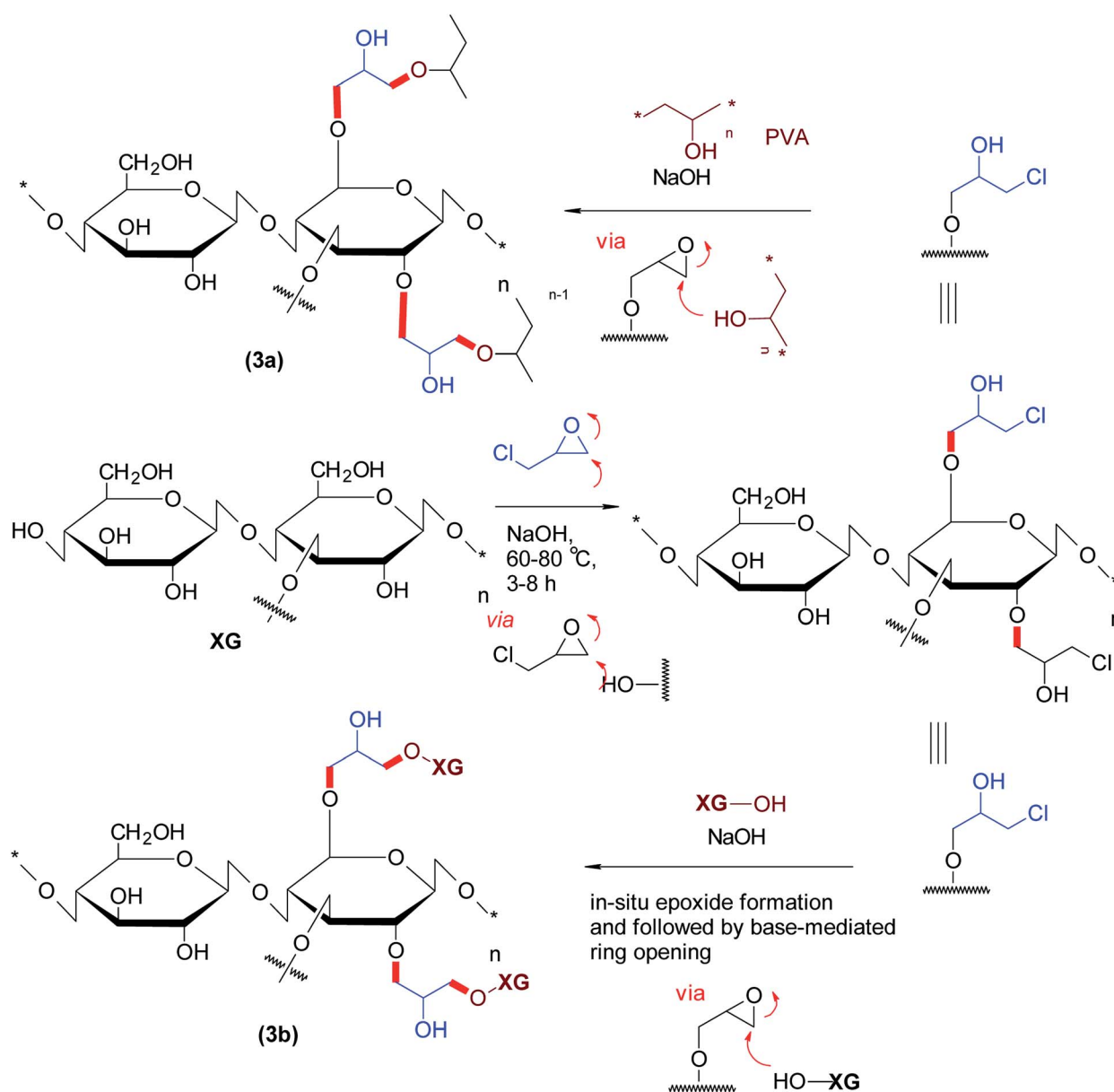


Fig. 3 Epichlorohydrin-crosslinked XG or PVA/XG.



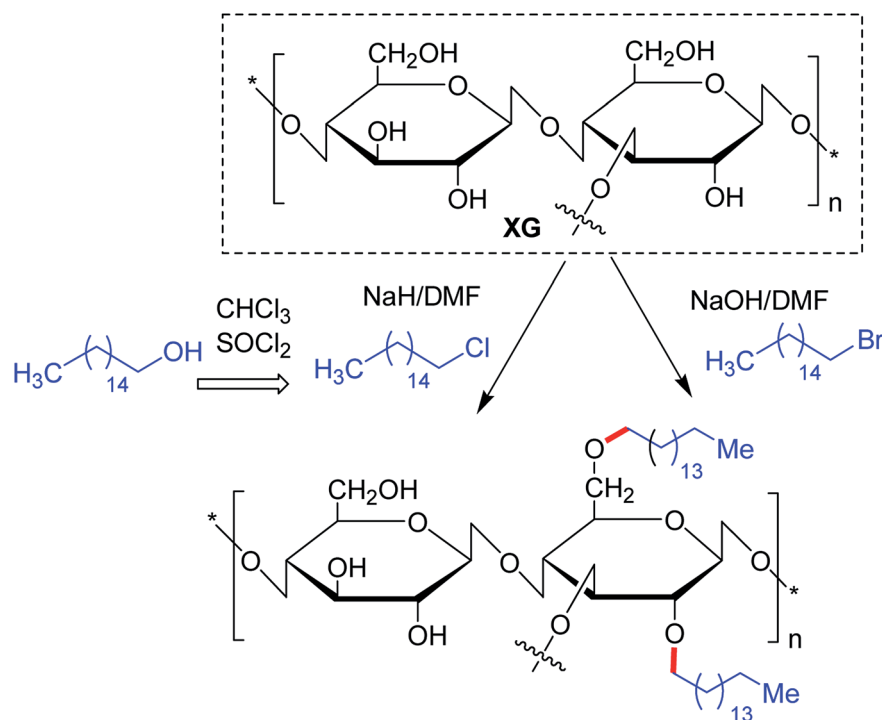


Fig. 4 Long alkyl chain conjugated XG derivatives.

postulated that less viscous carboxymethyl XG promoted penetration of water into the matrix, caused faster dissolution of the matrix resulting in faster drug release in phosphate buffer solution (pH 6.8). However, the release profiles of diclofenac-loaded carboxymethyl XG matrices were contradictory to that reported by Bakshi *et al.*¹²⁷

Munir *et al.*¹²⁹ reported better thermal stability and DPPH free radical scavenging (antioxidant) activity of XG after carboxymethylation. The antibacterial effects of gum became intensified after modification. In another study, carboxymethyl XG capped gold nanoparticles showed their unique excellence in cancer cell killing. Microwave synthesis of carboxymethyl XG-capped gold nanoparticles containing doxorubicin exhibited about 4.6-folds higher anticancer efficacy *in vitro* in presence of an ionophore (*i.e.* nigericin) than free doxorubicin.¹³⁰

The *O*-carboxymethyl ether derivative of XG was largely synthesized for the fabrication of ionically and/covalently crosslinked hydrogel particles and tested for controlled drug delivery. The outcomes of such investigations are summarized in Table 1.

2.2. Esterified XG

XG was also reshaped *via* esterification, and the derived properties were scrutinized for various applications. Qian *et al.*¹⁵⁴ esterified the carboxy groups of XG by 1-bromooctane to obtain water soluble XG. They attained an octyl substitution of 11 and 21 per 100 structural units. The esterification promoted hydrophobic association and consequently enhanced the viscosity of XG. The deacetylated XG (~1.3–1.4%) exhibited viscosifying effect¹⁵⁵ similar to that observed after octyl

substitution.¹⁵⁴ When esterified with poly(maleic anhydride/1-octadecene) (PMAO) in dimethyl sulfoxide (DMSO) solution (Fig. 5), the modified XG offered an outstanding resistance to shear force and displayed viscoelastic behaviors, attributable to hydrophobic association of the polar anhydride group and non-polar C16 side chain. The authors further acknowledged its excellent salt tolerance as well as temperature resistance properties which could be beneficial in oil recovery, pharmaceutical and food applications.¹⁵⁶

Tao *et al.*¹⁵⁷ endorsed sodium trimetaphosphate (STMP), a non-toxic and water soluble cyclic triphosphate for the crosslinking of hydroxy groups of XG chains under alkaline condition to design hydrogel disks. One phosphorous group intertwined two sugar rings of different XG chains (Fig. 6). STMP crosslinking endowed hydrogel disks more elasticity and mechanical stability than physical hydrogels. This esterification reaction induced porosity in the hydrogels, with a mean pore diameter of $114.5 \pm 22.1 \mu\text{m}$. However, the pore diameter dropped to $31.5 \pm 5.5 \mu\text{m}$ at 5% (w/v) XG due to sufficient crosslinking of the matrix. XG–STMP hydrogels swelled rapidly in the first hour and attained equilibrium in PBS (pH 7.4) in 28 h. However, the hydrogel disk swelled faster in water than that in PBS and remained undissolved for more than 11 days. Regarding protein diffusion, the hydrogels recorded a slow, sustained diffusion of bovine serum albumin in PBS medium over 50 h depending upon STMP concentration. The least diffusion (~28%) was evident at 5% STMP. STMP crosslinking perhaps introduced more anionic charges into XG, thus facilitating more water uptake by the hydrogels. Furthermore, the



Table 1 Properties of carriers systems made with ether derivatives via ionic crosslinking, polyelectrolyte complexation, covalent cross-linking or both

Type of devices	Polymers/cross linkers	Research outcomes	References
PEC hydrogels	Chitosan and XG/ Opadry	<ul style="list-style-type: none"> • Incorporation of metronidazole into pre-formed hydrogels in significant amount is possible by diffusion technique • Hydrogels swell less in simulated gastric medium • Enteric coating allowed 50% drug for release in colonic pH 	131
PEC hydrogels	<i>N</i> -Trimethyl chitosan (TMC)/carboxymethyl xanthan gum (CMXG)	<ul style="list-style-type: none"> • Encapsulation efficiency of ciprofloxacin reached to about 93.8% at higher drug load • Drug-loaded hydrogel was highly effective against the Gram positive and Gram negative bacterial strains • Highest diameter of inhibition zone against <i>Escherichia coli</i> as compared to gentamicin • Highest cell viability (97%) in lung human normal cell lines was noted at concentration up to 50 $\mu\text{g mL}^{-1}$ 	132
Ionic and polyelectrolyte complexation	Polyethyleneimine (PEI)/CMXG/ AlCl_3	<ul style="list-style-type: none"> • Increasing PEI (0.5–2%) and exposure time (5–30 min) decreased drug entrapment efficiency from 96.50 to 77.50% and from 92.25 to 70.37%, respectively • PEI treatment reduced swelling of the beads • Depending on formulation variables, 40% and 80% drug released in 2 h in pH 1.2 and in 5–6 h in pH 6.8, respectively • PEI treated diltiazem–resin complex beads released the drug following non-Fickian transport mechanism • Bioavailability was 1.59 times higher with PEI-treated formulation than pure drug solution in rabbit model • Data showed good <i>in vitro</i>–<i>in vivo</i> (IVIVC) correlation 	133
Magnetically responsive polyelectrolyte complex hydro-gels	XG and chitosan in the presence of iron oxide magnetic nanoparticles (MNPs) using D -(+)-glucuronic acid δ -lactone as a green acidifying agent	<ul style="list-style-type: none"> • Incorporation of Fe_3O_4 MNPs (8 nm size) into complex hydrogels induced porosity and greatly improved mechanical properties and storage modulus • Magnetically responsive hydrogels improved NIH3T3 fibroblasts cell proliferation and adhesion in an external magnetic field relative to pristine hydrogels without MNPs <i>in vitro</i> • Suitable for use as a magnetically stimulated system in tissue engineering applications 	134
Tablets	Polyethylene glycol/XG/ chitosan	<ul style="list-style-type: none"> • Drug release profiles of tested metronidazole tablets and commercial ER formulation were similar in 0.1 M HCl and phosphate buffer pH 6.8 • Bioadhesion of tested tablets was three times higher than commercial tablets to sheep duodenum • Absorption of metronidazole from test product was faster than that of commercial product with a maximum plasma level attained at 4.37 and 6.14 h, respectively 	135
Interpenetrating network (IPN) hydrogel beads	CMXG and carboxymethyl cellulose/ AlCl_3	<ul style="list-style-type: none"> • Methacrylic acid-based ion exchange resins (IER) were synthesized using ethylene glycol dimethacrylate, <i>N,N'</i>-methylene bis acrylamide, and divinyl benzene coded as ME, MB and MD respectively • IER : ofloxacin ratio of 1 : 2 provided highest drug loading \sim98% MD and MB; however, the same was \sim85% for ME • Taste masking studies at salivary pH 6.8 showed that MD : ofloxacin (1 : 4) showed lowest (1.22%) release of drug for a contact time of 30 s than others • Presence of bulky divinylbenzene imparted steric hindrance for the exchange of phosphate groups with the amine groups present in drug at salivary pH, ultimately resulting in slow release of the drug • CMXG beads containing MD : ofloxacin (1 : 4) (636 μm) had highest drug entrapment efficiency of 91.90% following treatment with 2% AlCl_3 • MD : ofloxacin (1 : 4)-carboxymethyl XG/carboxymethyl cellulose IPN beads (1122 μm) had 90.23% drug entrapment efficiency • Compared to gastric pH, drug release from MD : ofloxacin (1 : 4)-carboxymethyl XG beads and carboxymethyl XG/carboxymethyl cellulose IPN beads at intestinal pH 7.4 became prolonged and extended up to 10 h 	136



Table 1 (Contd.)

Type of devices	Polymers/cross linkers	Research outcomes	References
IPN hydrogel beads	Casein and CMXG/ aluminium chloride/ glutaraldehyde	<ul style="list-style-type: none"> • Glutaraldehyde treatment prohibited extent of degradation of hydrogels in pH 7.4 phosphate buffer containing 0.2% lysozyme • Theophylline release was slower in pH 1.2 buffer solution than in pH 6.8 buffer • Increasing carboxymethyl XG : casein ratio decreased the extent of drug release marginally both in acidic media and alkaline media in 2 h • Higher carboxymethyl XG : casein ratio and drug loading improved drug entrapment efficiency from 28.6 to 53.81% and from 53.81% to 83.59%, respectively • Increase in glutaraldehyde concentration caused lowering of drug entrapment efficiency from 83.01 to 40.03% upon 15 min exposure. Reduction of exposure time to 5 min increased DEE to 85.12% • Theophylline transformed into amorphous state after entrapment • Shifting of pH of dissolution medium from 1.2 to 6.8 caused significant swelling of beads • Increase in AlCl₃ concentration (2–8%) increased swelling of beads by 10.26% and 11.62% in pH 1.2 buffer and pH 6.8 buffer solutions, respectively 	137
IPN hydrogel beads	Pectin/CMXG/Al ³⁺ ions and covalent cross- linking with glutaraldehyde	<ul style="list-style-type: none"> • Both swelling and drug release were relatively low in acidic medium than in pH 6.8 buffer • <i>In vitro</i> delivery of diltiazem was dependent upon the extent of cross-linking and amount of drug used in the IPN hydrogel beads 	138
IPN hydrogel beads	CMXG and sodium alginate/AlCl ₃	<ul style="list-style-type: none"> • <i>C</i>_{max} was significantly less and <i>T</i>_{max} (2.91 ± 0.5) was relatively higher from the drug loaded IPN beads than those from the control and reference in rabbit model • Relative bioavailability of test formulation was 109.02% and 112.79% compared to control and reference, respectively 	139
Compression coated tablets	CMXG and sodium alginate/CaCl ₂	<ul style="list-style-type: none"> • Compression-coated tablets of Ca²⁺ ion crosslinked CMXG and sodium alginate could deliver prednisolone without the need of colonic bacterial intervention for degradation of the polysaccharide coat • Blend of CMXG and alginate (1.5 : 3.5) provided <i>T</i>_{lag} of 5.12 h and <i>T</i>_{rap} (time required for immediate release following <i>T</i>_{lag}) of 6.50 h 	140
Hydrogel beads	CMXG/AlCl ₃ / glutaraldehyde	<ul style="list-style-type: none"> • Diltiazem–cation exchange resin was loaded into carboxymethyl XG hydrogel beads • Sequential cross-linking involving glutaraldehyde treatment of ionically preformed hydrogel beads produced smaller beads with higher drug entrapment efficacy (86.52%) and prolonged release characteristics than ionic cross-linking, and simultaneous ionic/covalent cross-linking • Swelling of the beads was higher in acid solution of pH 1.2 than in buffer solution of pH 6.8 • Burst release (~50%) in acid solution, followed by extended release up to about 7 h 	141
Tablets	CMXG/CaCl ₂	<ul style="list-style-type: none"> • Increase in degree of calcium co-ordination/cross-linking reduced swelling of Ca–CMXG matrix tablets compared with carboxymethyl XG matrix • Erosion of Ca–CMXG matrices was higher than CMXG matrix • Release of prednisolone from Ca–CMXG matrices containing upto 33.33% (w/w) CaCl₂ was less than that from CMXG matrix • Release of drug from the matrix containing 50% (w/w) of CaCl₂ was rapid and approached almost to that from CMXG matrix 	142
IPN beads	Carboxymethyl cellulose and CMXG/ AlCl ₃	<ul style="list-style-type: none"> • Higher extent of cross-linking led to decreased particle size from 1080 to 1420 μm • Variation in cellulose to gum ratio from 1 : 1 to 2 : 1 dropped drug encapsulation efficiency of IPN beads from 96.96 to 77.45% • Drug entrapment efficiency of the beads decreased at higher salt strength • Increase in salt strength from 4 to 8% slowed drug release rate in acidic and alkaline media • Drug release continued up to 8 h in pH 7.4, indicating better intestinal drug release 	143



Table 1 (Contd.)

Type of devices	Polymers/cross linkers	Research outcomes	References
Homopolymeric and IPN beads	CMXG and sodium alginate/ AlCl_3	<ul style="list-style-type: none"> • Entire ibuprofen can be loaded into the beads with a maximum coefficient variation of 1.87% • IPN hydrogel beads provided more sustained release of ibuprofen than homopolymeric beads • Rapid drug release from Al–CMXG beads, accounting 42.5% release in 2 h • Incorporation of higher amounts of alginate in IPN beads decreased drug release • Release of drug from Al–alginate beads was the lowest, releasing 25.4% drug in 2 h in acidic medium • Release of drug was the fastest from Al–CMXG in same duration • Drug release from Al–alginate beads was faster than those from the IPN beads in pH 6.8 phosphate buffer 	144
IPN beads	CMXG and sodium alginate/aluminium chloride	<ul style="list-style-type: none"> • Ulcerogenicity decreased significantly with ibuprofen-loaded IPN beads in comparison to the pure drug in adult male albino Wistar rats 	145
Microparticles	CMXG/aluminium chloride	<ul style="list-style-type: none"> • CMXG or alginate-coated Al–CMXG microparticles were prepared • Higher salt concentration decreased BSA entrapment efficiency of the uncoated microparticles from 86–61% • BSA entrapment in coated microparticles was found lower (78–79%) than uncoated microparticles • Uncoated microparticles released almost half of its content in NaCl–HCl buffer solution (pH 1.2) in 2 h • Alginate and xanthan coated microparticles did not liberate a substantial amount of entrapped protein in acidic medium, rather prolonged protein release in PBS solution (pH 7.4) up to 10 and 12 h, respectively • Sodium dodecyl sulfate–polyacrylamide gel electrophoresis indicated retention of protein integrity in the microparticles 	146
Microparticles	CMXG/aluminium chloride	<ul style="list-style-type: none"> • Variation in pH of carboxymethyl XG solution did not affect protein entrapment and release significantly • Increase in initial protein loading tended to increase protein release in buffer solution of pH 1.2 and in PBS solution (pH 7.4) • Higher polymer concentration suppressed protein release substantially in both acidic and alkaline media 	147
Hydrogel beads	CMXG/aluminium chloride	<ul style="list-style-type: none"> • Maximum 86.39% protein entrapment efficiency was noted • Diltiazem–resin complex was loaded into CMXG beads • Higher gelation period (5–20 min) and AlCl_3 concentration (1–3%) decreased drug entrapment efficiency from 95 to 79% and 88.5 to 84.6%, respectively • Gum concentration up to 2.5% improved drug entrapment efficiency to 90.7% • Higher swelling was accounted for faster drug release in simulated gastric fluid than in intestinal fluid 	148
Hydrogel beads	CMXG/aluminium chloride	<ul style="list-style-type: none"> • Viscosity of CMXG was always lower than that of XG • Formation of discrete and spherical BSA-loaded microparticles were dependent on microenvironmental pH • BSA entrapment efficiency was 82% • Higher swelling contributed faster protein release in acidic medium than that in alkaline medium • pH of the gum solution influenced the swelling and protein release considerably 	149
Nanoparticles	XG/sorbitan monooleate/oleylamine	<ul style="list-style-type: none"> • XG-functionalised sorbitan monooleate/oleylamine nanoparticles had average diameter of 146.8 nm and high surface charge (–48 mV) • Core–shell morphology of enhanced green fluorescent protein plasmid (pEGFP) loaded nanoparticles was evident • Cytotoxicity and transfection capacity of nanoparticles were excellent in human umbilical vein endothelial cells (HUVECs) • Pre-clinical study confirmed the potential of XG-functionalized span nanoparticles for gene targeting to endothelial cells • Xanthan shell protected associated DNA from DNase degradation, a prerequisite for intact delivery of bioactives to its site of action 	150



Table 1 (Contd.)

Type of devices	Polymers/cross linkers	Research outcomes	References
Layered Fe-XG hydrogels	XG/FeCl ₃	<ul style="list-style-type: none"> • Fe³⁺-coordination enabled XG to form hydrogels under ambient temperature • Fe-XG hydrogel exhibited a regular laminated structure under scanning electron microscope • XG hydrogels possessed uniform layered structure, enhanced mechanical strength and excellent swelling behavior • Sol-gel conversion of XG-based hydrogel could be realized by UV light in the presence of sodium lactate • Sol-gel conversion ability of Fe-XG hydrogel could provide data for using as sensor to detect oxidizing or reducing agents, as an actuator under UV light with enough sodium lactate, or for the drug release in the future study 	151
Floating hydrogel beads	Chitosan/XG	<ul style="list-style-type: none"> • Variation in chitosan to XG ratio did not affect glipizide release behaviors of the beads in phosphate buffer pH 7.4 up to 24 h • Drug entrapment efficiency was 80–95% • Beads possessed comparable buoyancy in gastric fluids and satisfactory bioadhesive strength 	152
Hydrogel beads	CMXG/carboxymethyl cellulose/AlCl ₃	<ul style="list-style-type: none"> • Swelling kinetics differed significantly in pH 1.2 and 7.4 buffer • Beads released considerably less amount of aceclofenac in acid solution (maximum 14.2%) and provided controlled release in phosphate buffer solution (pH 6.8) • Aceclofenac was compatible with the matrix • Morphology, size and drug entrapment efficiency of beads, and <i>in vitro</i> drug release was affected by viscosity of polymer dispersion, initial drug load, and concentration of total polymer and AlCl₃ 	153

counter ions inside the XG-STMP hydrogels might increase the osmotic pressure and cause higher swelling.¹⁵⁸

Shalviri *et al.*¹⁵⁸ mingled gelatinized starch and XG in order to develop hydrogel films. STMP crosslinking produced continuous and homogeneous films with few micropores, even in absence of plasticizer. Higher STMP (20%) and XG (20%) levels speeded up the swelling of films in deionized water by 1.7-folds compared to that in phosphate buffer (pH 7.4). As usual, the gel mesh size became enlarged from 2.84 to 6.74 nm as swelling continues. The permeation of anionic drugs (ibuprofen sodium and sodium salicylate) through films was slower than neutral entities, thus demonstrating their potential for controlled release of ionizable drugs. Intermingling of a hyper-branched polysaccharide, extracted from the *Pleurotus tuber-regium* sclerotic, and XG followed by STMP-crosslinking ensued the formation of hydrogels that demonstrated shear-thinning behaviors, and improved mechanical stability. In addition, the hydrogels presented self-healing ability and controlled drug release property. The hybrid hydrogel had a storage modulus better than XG-STMP hydrogel.¹⁵⁹

Bejenariu *et al.*^{160,161} highlighted the importance of STMP : XG ratio on the swelling and release of cationic dyes from the hydrogels. The hydrogels obtained at STMP : XG ratio of 10 : 1 swelled 20 times in pH 7.0 and entrapped 87.5% methylene blue. The hydrogels liberated more amounts of dyes in NaCl solution than in acid solution (pH 3.0). Overall, STMP-crosslinking supported XG hydrogels for cationic drug release.

The dehydration of hydrogel-like human nucleus pulposus and imbalanced stress distribution in the intervertebral disc causes degenerative disc disease. Thus, the regeneration of nucleus pulposus is the key to success for treating this. However, the avascularity and low mitotic activity of chondrocytes limits their successful treatment.¹⁶² In view of this, Leone and coworkers¹⁶³ proposed STMP-crosslinked PVA : XG (4 : 1) hydrogels as a good substitute for nucleus pulposus.

Bueno *et al.*²¹ appraised citric acid as a natural crosslinker for the synthesis of XG hydrogel films (~0.03 mm thick) (Fig. 7). They observed that citric acid produced porous, homogeneous films without nanofibrils, indicating its crosslinking efficiency. However, it reduced the mechanical strength of the films. The properties of citric acid-crosslinked hydrogels were comparable to those reported for xanthan-starch films.¹⁶⁴ The swelling of hydrogels was significant at pH 10.0; however, the same was negligible at pH 2.0 and pH 6.5. The water absorption was initially controlled by quasi-Fickian diffusion mechanism; however switched to anomalous diffusion at later period. Perhaps, the disruption of ester bonds favored swelling of the films in basic medium. They further reported that XG chains could be intertwined by heating at 165 °C without the need of citric acid. In this case, the diffusion of water into hydrogels changed from quasi-Fickian to Fickian at later times. Heating brought about weaker crosslinking of XG chains. The nanofibrils (about 20–30 nm diameter) were evident on cryofractured hydrogel surface. Moreover, the hydrogels did not respond to



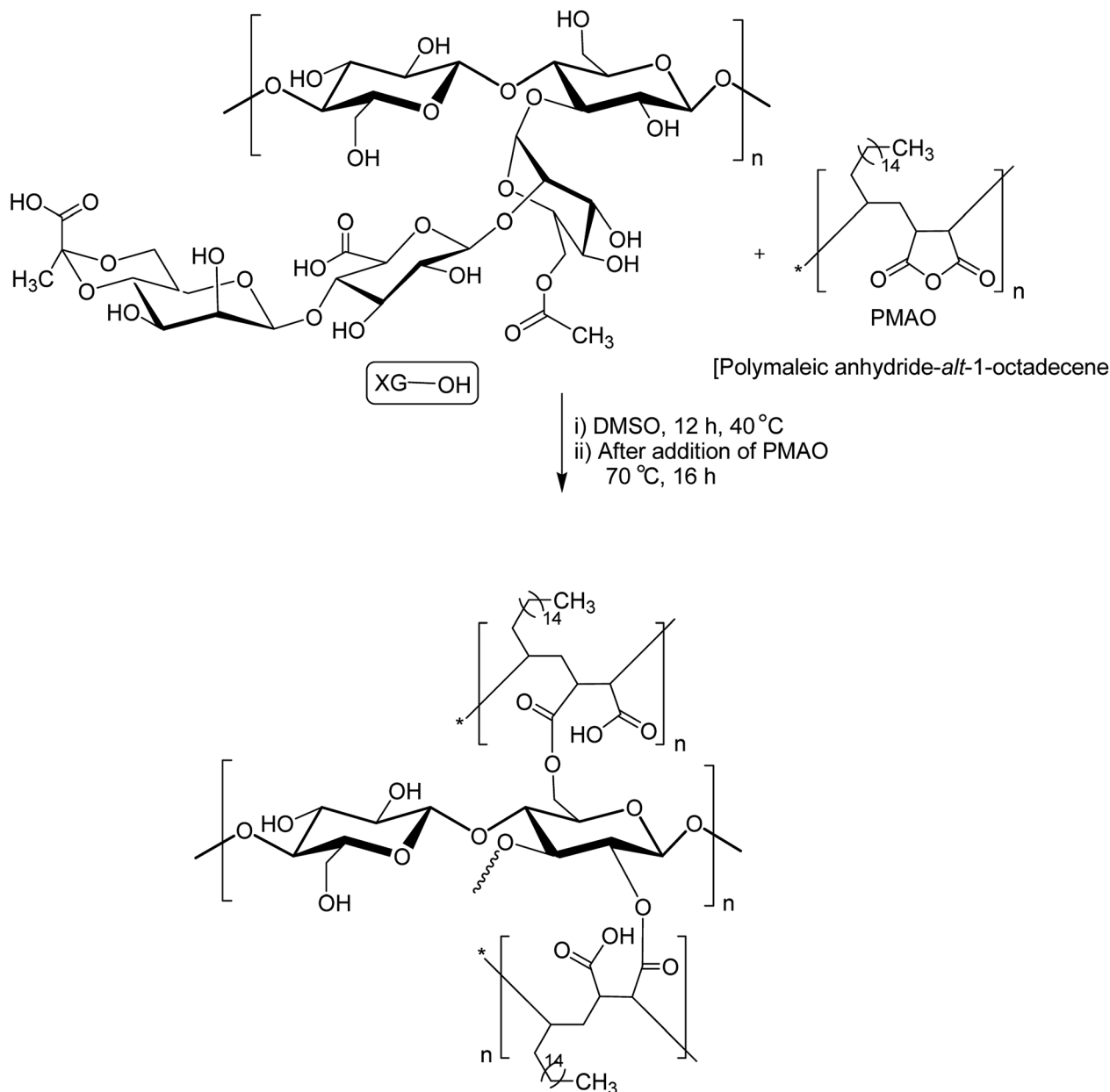


Fig. 5 Esterification of XG with PMAO.

pH variation promptly as was seen in case of citric acid-treated hydrogels, because they had more crosslinks to be hydrolyzed.

Circular dichroism confirmed the disordered conformation (coils) of xanthan chains in case of citric-acid crosslinked hydrogels; whereas the crosslinking by heating left XG in an ordered conformation (helices) in the hydrogels.¹⁶⁵ Under coil conformation, usually a larger number of carboxylate groups are exposed, thereby providing citric acid an opportunity to give birth to more ester linkages. The protein release behaviors from hydrogels were quite amazing. Isoelectric pH of protein and pH of release medium played a crucial role in protein release. For instance, the heat-treated hydrogels completed the delivery of BSA after 1 h over the pH range of 2–10; except at pH 4.8 close to its isoelectric point. In contrast, the release of lysozyme became

very low at pH 7.0. This could be attributed to the electrostatic attraction between lysozyme and hydrogels. Both hydrogel films seemed efficient as carriers for proteins (lysozyme) having high isoelectric point. The hydrogels preserved native conformation of lysozyme after loading and thus could offer substantial bactericidal activity in wound dressings. Followed by this, Huang *et al.*¹⁶⁶ designed novel wound dressing material exploiting citric acid-crosslinked transparent, porous XG films laden with silver nanoparticles (AgNPs). The nanocomposite films were non-cytotoxic to fibroblasts (L929) at $<10 \mu\text{g mL}^{-1}$. The dressing prohibited the formation of biofilms, reduced the inflammatory reactions, and triggered angiogenesis of the granulation tissues in non-healing wounds, infected with methicillin-resistant *S. aureus*. The nanocomposites impeded



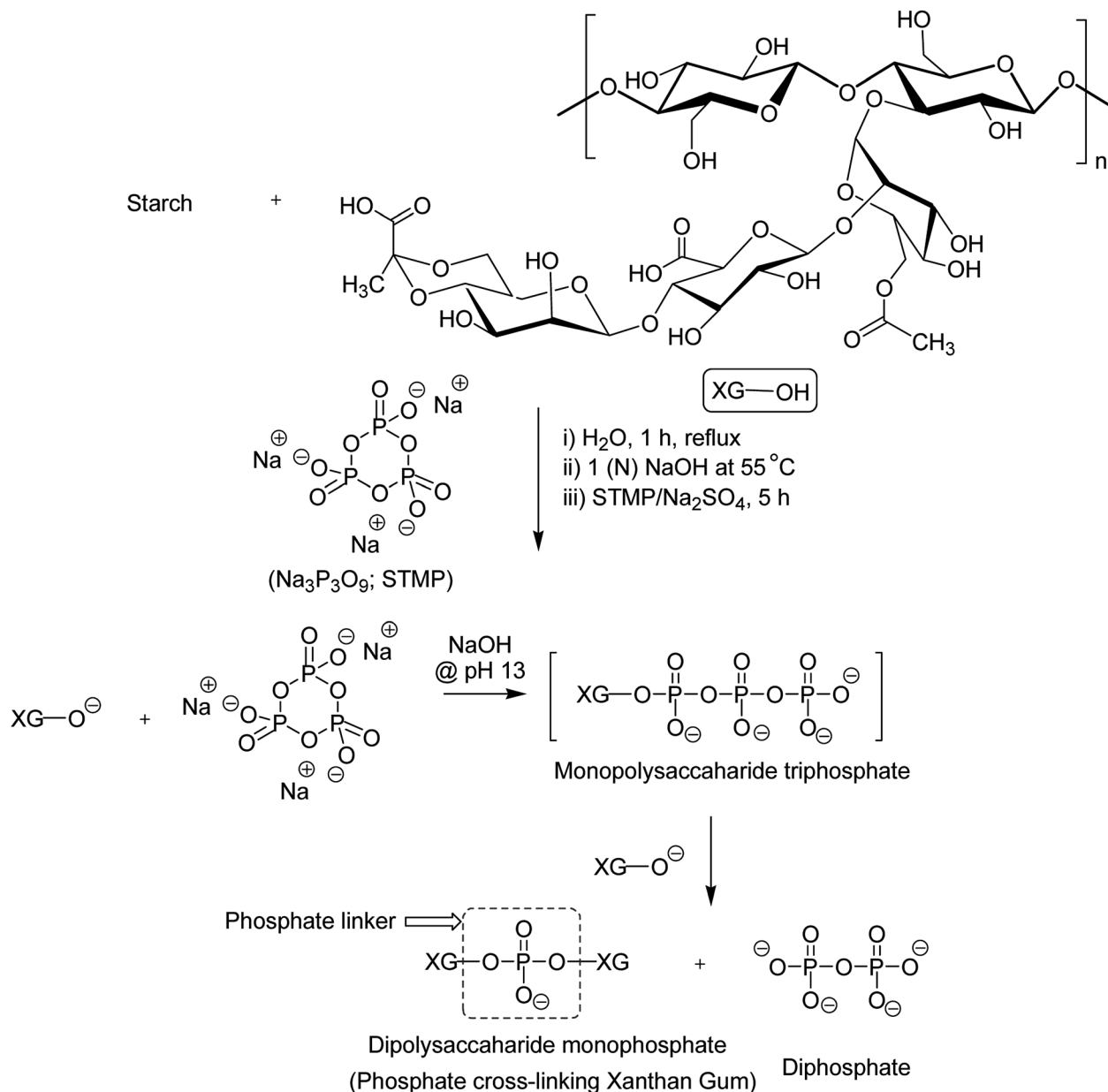


Fig. 6 STMP-crosslinked XG derivatives.

immature release of AgNP in PBS solution and hence exhibited the antibacterial effect for more than 24 h.

In a study,¹⁶⁷ glycerol was mentioned for chemical bonding of XG in a waterless or near-waterless environment at temperature $>105^\circ\text{C}$ (Fig. 8). It was hypothesized that water generated during cross-linking reaction could catalyze the unwinding of double-helix of XG and provide more access to its functional groups and backbone for cross-linking. The crosslinking occurred at glycerol : xanthan ratio of <27.6 . Higher xanthan : glycerol ratio imparted more hardness, swellability (>39 g water per g gel), dye sorptivity ($>85\%$) to the hydrogels. The hardness was almost 40 times greater with 50% xanthan gel than that made with 5%. The interdiction of red 40 and blue 1 (food dyes) release in water (60–70% in 24 h) unfolded the

possibilities for constructing slow-release matrices. Moreover, the crosslinked gels came up effectively with dye absorbent capacity; thus advocated its application in food, pharmaceutical and other industries.

Hamcerencu *et al.*¹⁶⁸ described esterification of XG with an unsaturated acrylic acid or acid reactive derivatives (acryloyl chloride, maleic anhydride) followed by grafting with *N*-isopropylacrylamide. This resulted in thermo- and pH-sensitive hydrogels. Maleic anhydride presented higher reactivity towards primary hydroxy groups (C6) as compared to others. Higher temperature and reaction time had positive impact on the degree of substitution.

Mendes *et al.*¹⁶⁹ evinced that palmitoylation of xanthan (xanthan : palmitoyl chloride = 1.7) expedited self-assembly of



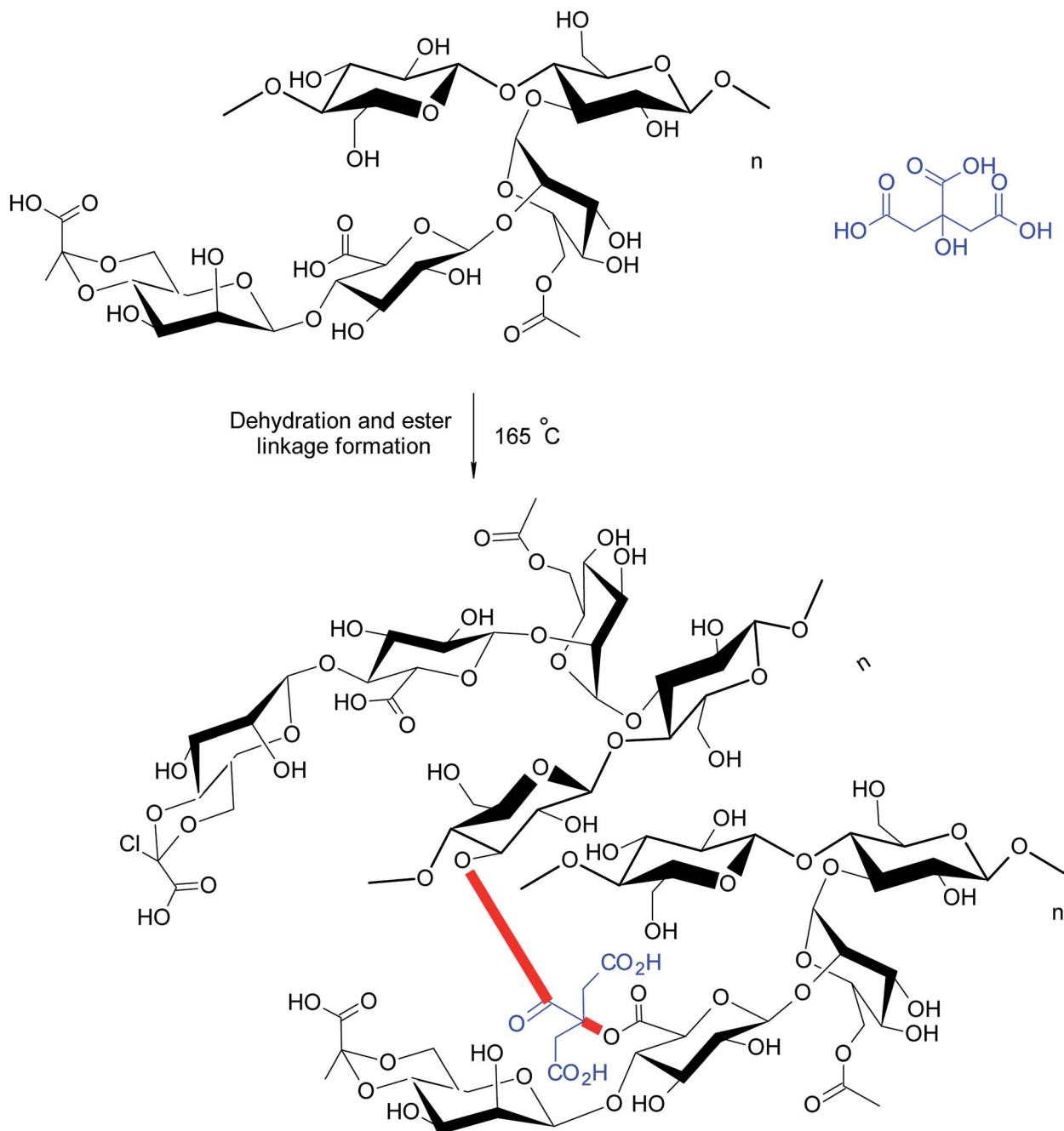


Fig. 7 Citric acid reinforced XG derivatives.

XG and furnished a compatible environment for cell microencapsulation. However, there was a dearth of physical stability to enclose cells and protect from immunotoxins. Subsequent membrane coating of the microcapsules with poly-L-lysine allowed permeation of sufficient oxygen and nutrients necessary for cell survival and proliferation (Fig. 9).

Bhatia *et al.*¹⁷⁰ pointed out that thio-esterification of XG with mercaptopropionic acid and thioglycolic acid evoked better *ex vivo* mucoadhesion (Fig. 10). The extent of mucosal retention of metronidazole-loaded buccal pellets of XG-mercaptopropionic acid was less than XG-thioglycolic acid. Improved

mucoadhesion of thiolated XG over native XG could be credited to the disulfide links between mucus and thiol groups. Nonetheless, the buccal pellets of XG-thioglycolic acid sustained the diffusion of metronidazole more effectively than XG-mercaptopropionic acid and native XG pellets. The degree of thiol substitution in XG-thioglycolic acid was 1.07 times higher than XG-mercaptopropionic acid. This could be responsible for variable *ex vivo* bioadhesion and *in vitro* release characteristics of the two conjugates.

In view of the potential toxicity of surfactants as suspending agents, Skender and groups¹⁷¹ synthesized three ester



derivatives of XG using diphenylmaleic anhydride, phthalic anhydride, and epichlorhydrin-phenol and assessed the worth of these derivatives as dispersing agent for double-wall carbon nanotubes in water. Xanthan itself is a very poor dispersing agent for carbon nanotubes. However, these derivatives proved useful as stabilizer for suspension of carbon nanotubes (0.5%, w/w). Xanthan esterified with diphenylmaleic-and phthalic-esters successfully immobilized carbon nanotubes in very acidic media compared to that esterified with epichlorhydrin-

phenol. Above pH 5.0, all the derivatives stabilized the suspensions effectively. Summarily, the presence of aromatic moiety in XG backbone was essential in dispersing carbon nanotubes in water *via* p-p stacking interaction.

A group of investigators synthesized succinoyl XG following reaction with succinic anhydride in presence of 4-dimethylaminopyridine catalyst at room temperature.¹⁷² Succinoyl XG turned into unflowable hydrogels at a very low strength (1.4% w/

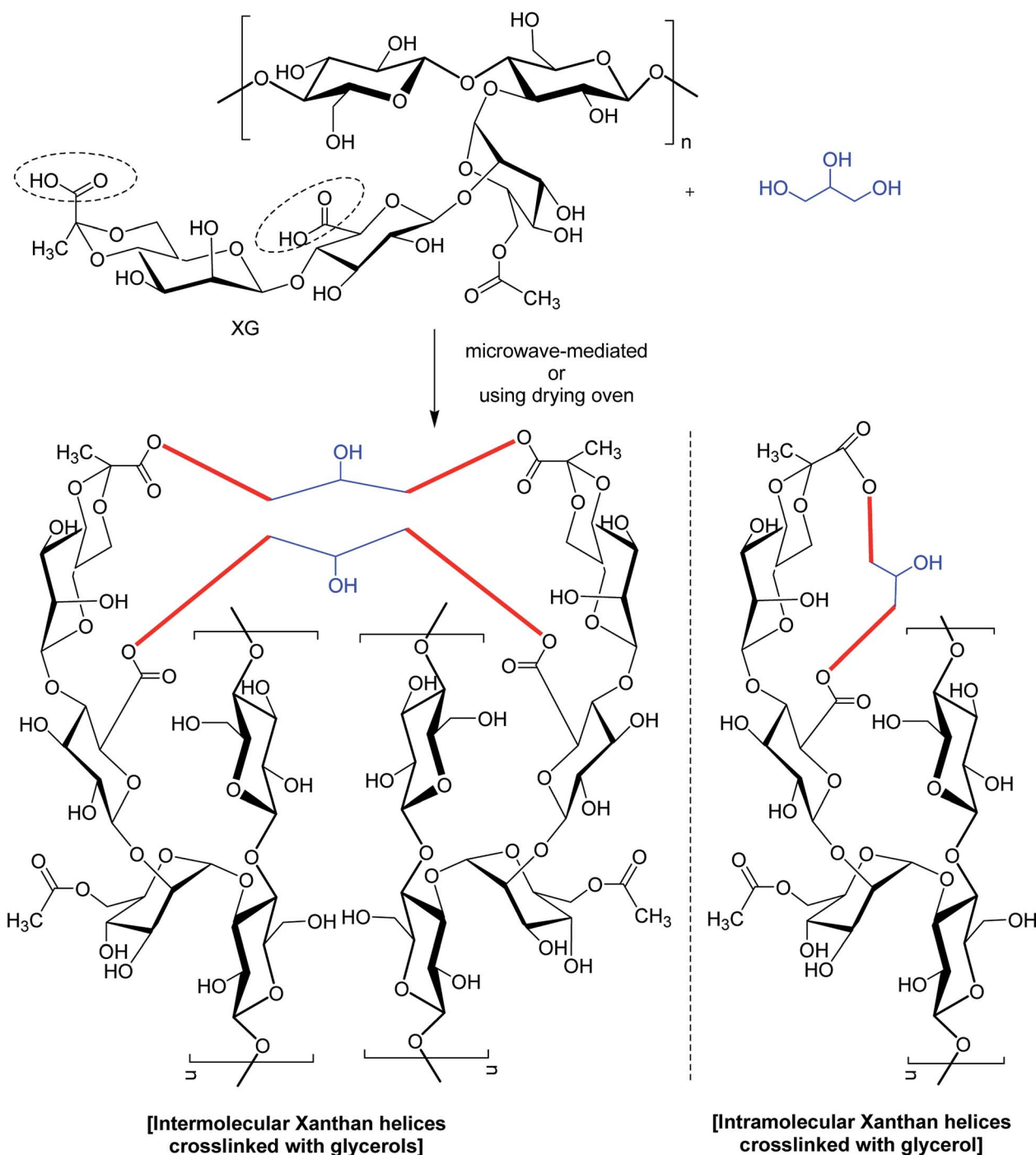


Fig. 8 Glycerol-crosslinked XG.



w), while the unmodified XG existed as flowable fluid at the same concentration.

In case of native XG, the hydrogel network is usually formed through physical entanglements. Succinylation promoted the formation of secondary bonds, rendering more elasticity to the hydrogels. Further, the hydrogels responded to ionic strength and sustained the release of gentamicin for 9 days in PBS (pH 7.4). The

hydrogels inhibited the formation of biofilm and demonstrated excellent bactericidal function in rabbit subcutaneous *S. aureus* infection model. As was evident from cytocompatibility study against human lens epithelial cells, the hydrogels supported cell adhesion; proliferation and migration at a dose of 1 mg g^{-1} . Overall succinoylated XG hydrogels were propitious as drug delivery materials for antibacterial applications.

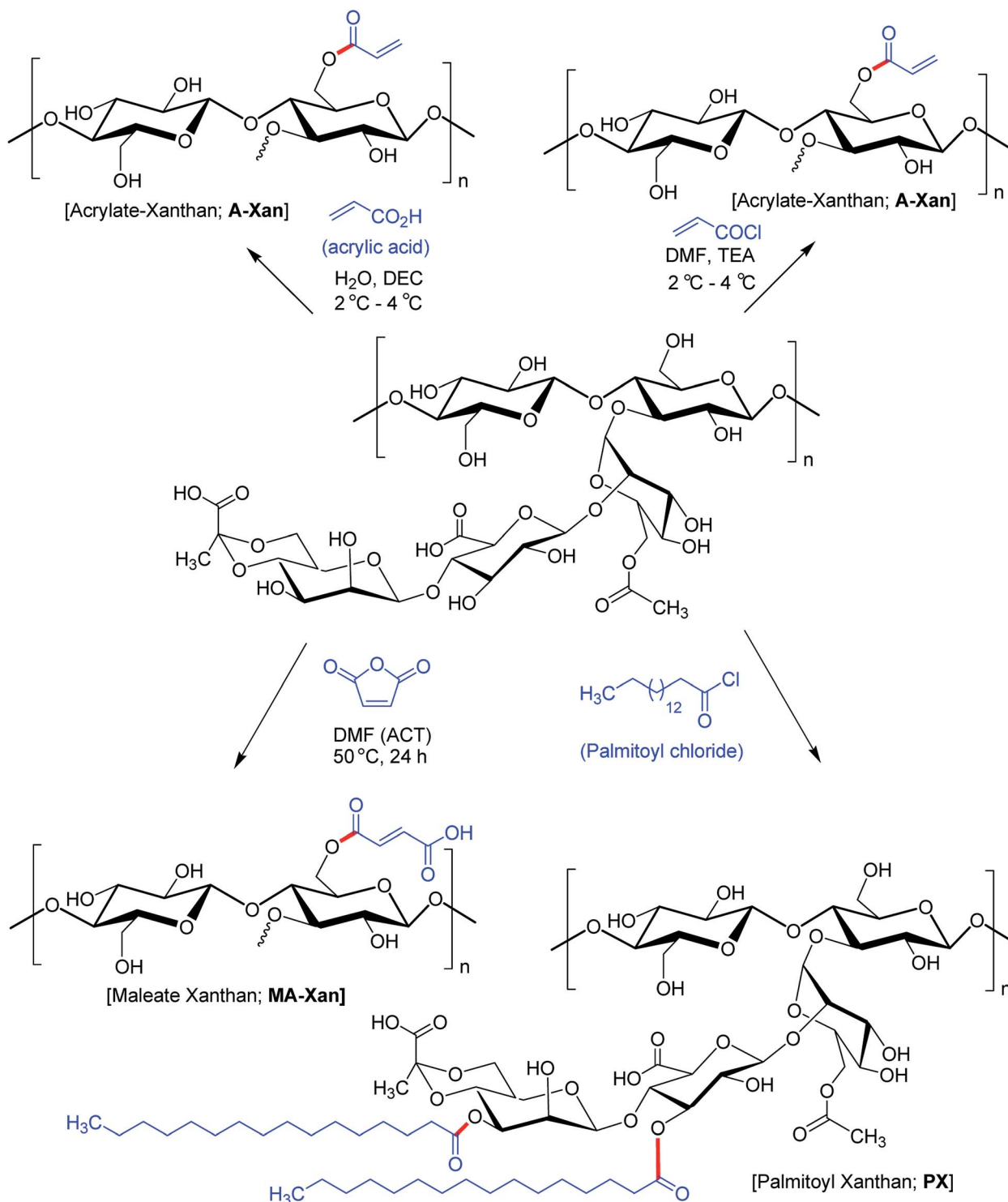


Fig. 9 Chemical structures of acrylate XG, maleate XG and palmitoyl XG.



2.3. Oxidized XG

Oxidation of XG could partially convert glucopyranose units to aldehyde and create additional functional sites for covalent crosslinking. As reported with alginate polysaccharide, the sodium periodate oxidized the hydroxy groups on C2 and C3 of the repetitive unit of alginate and led to the formation of two aldehyde groups in each oxidized monomeric unit by the rupture of C2–C3 bond.¹⁷³

Guo and coworkers¹⁷⁴ brought into play xanthan aldehyde as a crosslinker for producing transparent, UV-protective edible gelatin films. Increased C=N groups by Schiff's base formation greatly improved the hydrophobicity, mechanical properties and thermal stability of gelatin films. Paiva and colleagues¹⁷⁵ exploited XG aldehyde as an adhesive for the natural cork stopper. The authors proclaimed that oxidized XG performed well due to the reaction of aldehyde groups with hydroxy groups on cork surface during high temperature drying. Ma *et al.*¹⁷⁶ disclosed a new hydrogel system based on XG aldehyde and phosphatidylethanolamine liposome. The hydrogel was sensitive to stimuli like heat, pH, and histidine and papain enzyme. The hydrogels manifested self-healing ability and provided an excellent environment for cell encapsulation for tissue engineering.

Intrinsic healing property of hydrogels is of utmost importance for *in vivo* drug delivery because this property could relieve hydrogels from destabilization under adverse mechanical or chemical exposure *in vivo*.¹⁷⁷ Salazar and associates¹⁷⁸ reported intrinsic self-healing ability of a composite hydrogel system of oxidized XG and chitosan. The hydrogels extended successful self-healing ability at room temperature and pressure while maintaining a good mechanical strength at equal chitosan : aldehyde XG mass ratio.

Injectable hydrogels have been abortive due to their instability to tissue exudates and delayed gelation under physiological conditions. Huang *et al.*¹⁷⁹ attempted to overcome these issues. They devised a stable and rapid gelation method for fabricating self-healing hydrogels based on hydrogen-bonding interaction and Schiff's base reaction between XG aldehyde and carboxymethyl chitosan (Fig. 11). The incorporation of vascular endothelial growth factor (VEGF) in hydrogels speeded up the reconstruction of abdominal wall in rats. The hydrogel managed the release of protein effectively.

The antioxidant activity of oxidized XG was also disclosed in the literature. Xiong *et al.*¹⁸⁰ exposed that XG when oxidized under alkaline medium exhibited better antioxidant activity than native XG. XG oxidized under acidic condition but having similar molecular weight had relatively poor antioxidant activity. The variable contents of pyruvate and reducing sugar could be responsible for this discrepancy. Nevertheless, dialdehyde XG could serve as a reducing agent for the synthesis of silver nanoparticles as well as designing biocompatible composite dressings suitable for burn healing.¹⁸¹

2.4. Amide functionalized XG

The carboxy functionalities of XG chains could be the focal points for bifunctional amine crosslinkers to establish covalent

amide bonds. Long hydrophobic alkyl chain can be substituted to XG backbone under ordered (helical) and disordered (coil) conformation *via* carbodiimide chemistry. Roy *et al.*¹⁸² conjugated octylamine onto the carboxy moieties of xanthan under its ordered (helical) conformation in water at room temperature (Fig. 12). The reaction involved complete protonation of the carboxy functions at pH 3.0 followed by its activation in presence of EDAC/NHS. The pH was adjusted afterward to 4.5 which promoted the protonation of EDAC's nitrogen atoms. Subsequent addition of an aqueous solution of octylamine, pH adjustment (pH 10.0) led to the formation of amide bonds through the nucleophilic attack of octylamine onto the activated carboxy functions of xanthan. The grafting density varied between 8 and 29% depending upon the stoichiometry of octylamine and EDAC with respect to carboxy functions. In spite of additional intermolecular interactions, hydrophobic chain grafting did not alter the viscoelasticity, and chain conformation of XG. Even being amphiphilic, the modified XG did not exhibit any additional self-assembling properties. Perhaps, the high stiffness of xanthan helices precluded self-assembly of modified XG, usually observed for flexible amphiphilic polymers. However, the hydrophobic interactions strengthened the suspending ability of xanthan considerably at rest but its shear-thinning behavior remained untouched at high shear rate. This property could be tuned by changing the grafting density. Thus, ordered modification of XG could enrich the list of potential stabilizers and thickeners in pharmaceutical formulation. As observed earlier by Roy *et al.*,¹⁸² the modification of XG under ordered conformation exhibited similar behavior to native XG; albeit the chain relaxation occurred slowly. Conversely, XG when modified under its disordered conformation displayed a chemical gel-like behavior without any relaxation of the chain.¹⁸³ The gel formation took place as low as 0.05 g L⁻¹ concentration. This owed to the presence of flexible backbone and squealed to intermolecular hydrophobic interactions. Circular dichroism study unveiled the fact that acidic form of xanthan adopted disordered conformation in DMSO. Further, they altered the structure of XG under its disordered conformation using *O*-(benzotriazol-1-yl)-*N,N,N'*-tetramethyluronium hexafluorophosphate (HBTU) as reagent for acid-amine coupling. Briefly, acidified XG was solubilized into DMSO at 80 °C and was then cooled to room temperature. Then, HBTU, triethylamine (deprotonating agent for carboxylic acid) and octylamine were introduced. Fantou *et al.*¹⁸⁴ synthesized octyl derivative of XG under its ordered conformation. The octyl XG had an exceptional capacity to stabilize oil-in-water emulsion at 0.2% w/w concentration, without the need of additional surfactant. The grafting density (8–32%) had negligible effect on emulsion stability. This finding opened up the possibilities of formulating emulsion that requires little or no surfactant in order to limit their toxicological and environmental impact.

However, the study findings on emulsion containing octyl xanthan (disorder conformation) thwarted emulsion stability through creaming, regardless of its concentration. The instability problem became more pronounced at higher grafting density. However, this phenomenon reversed for *n*-hexadecylamine XG (disorder conformation).¹⁸⁵



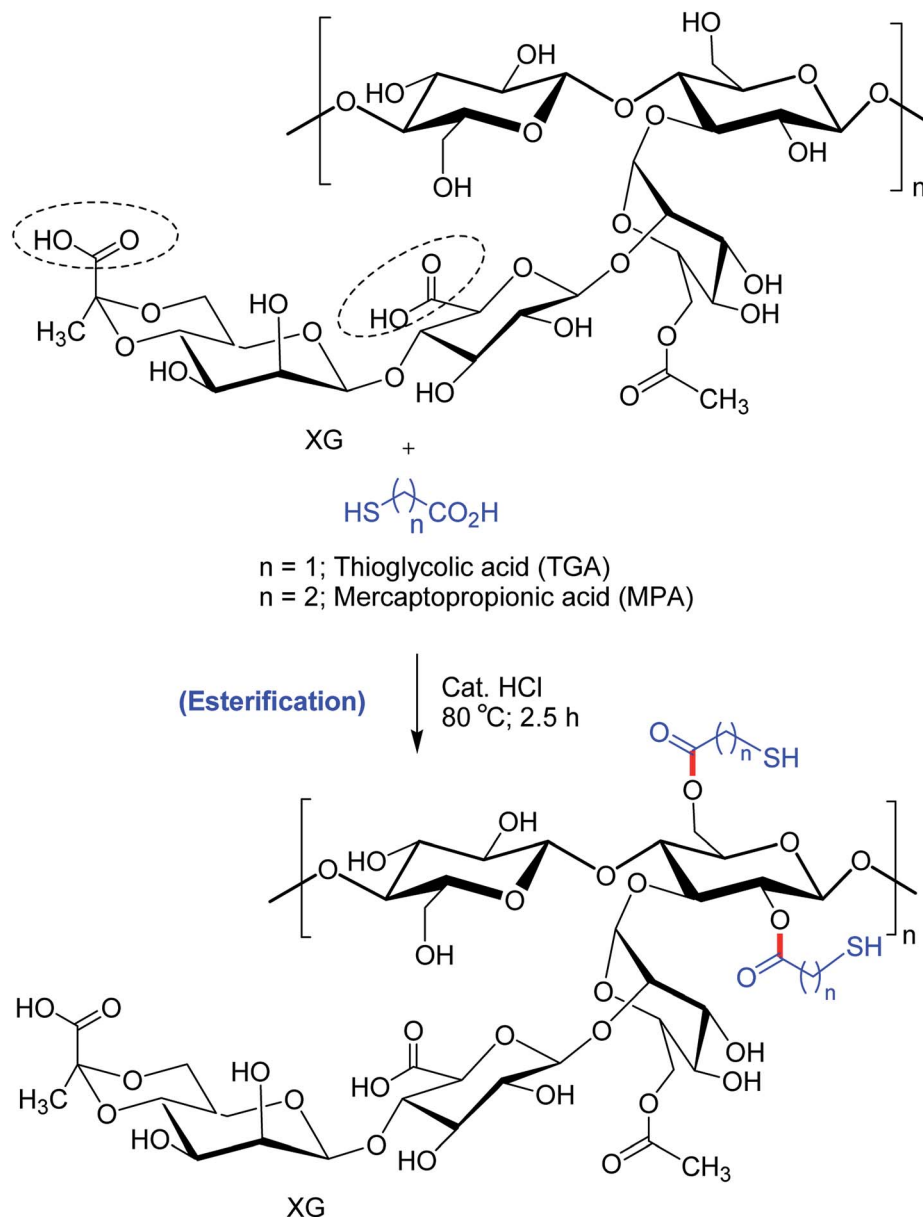


Fig. 10 TGA- and MPA-derived modified XG.

Bejenariu *et al.*^{161,186} reported a new XG hydrogels based on adipic acid dihydrazide (ADH) treatment. The crosslinking of XG with ADH is displayed in Fig. 13a. In the crosslinking reaction, LiNO_3 was used for the stabilization of the helicoidal structure of XG at pH 3.0. The hydrogels exhibited pH-dependent swelling but the pH-effect was not so drastic. The least degree of swelling was noted at pH 3.0 because the helical conformation of XG predominated in crosslinked networks. Despite the prevalence of coil conformation, the weak amide bonds restricted the networks from exhibiting similar swelling behaviors at pH 13.0 to that noted under acidic condition. Notably, the highest degree of swelling was recorded at neutral pH. The hydrogels absorbed $\sim 98\%$ of methylene blue in 24 h and released the dye in salt solution at a rate faster than in acidic solution (pH 1.5). Laffleur and groups¹⁸⁷ showed that

thiolated XG disc worked better in terms of stability (1.62-fold), *ex vivo* buccal mucoadhesion (8.35-fold) and tensile strength (2.65-fold) compared to unmodified xanthan. These findings coupled with its non-cytotoxicity towards Caco-2 cells inspired the authors in developing buccal patch to treat sialorrhea, a buccal disease associated with an involuntary loss of saliva. To fight against enhanced psychosocial and mental problems of sialorrhea patients,¹⁸⁸ tannic acid was found suitable for inhibition of excessive flow of saliva. Laffleur *et al.*¹⁸⁹ developed L-cysteine-conjugated XG buccal patch for tannic acid. The constant swelling in simulated saliva solution (pH 6.75) and non-cytotoxicity to Carey 24 cell lines embellished the buccal patch. Thiolation prohibited matrix erosion about five times and lessened the saliva flow about 33% more than the unmodified XG in excised porcine buccal mucosa.



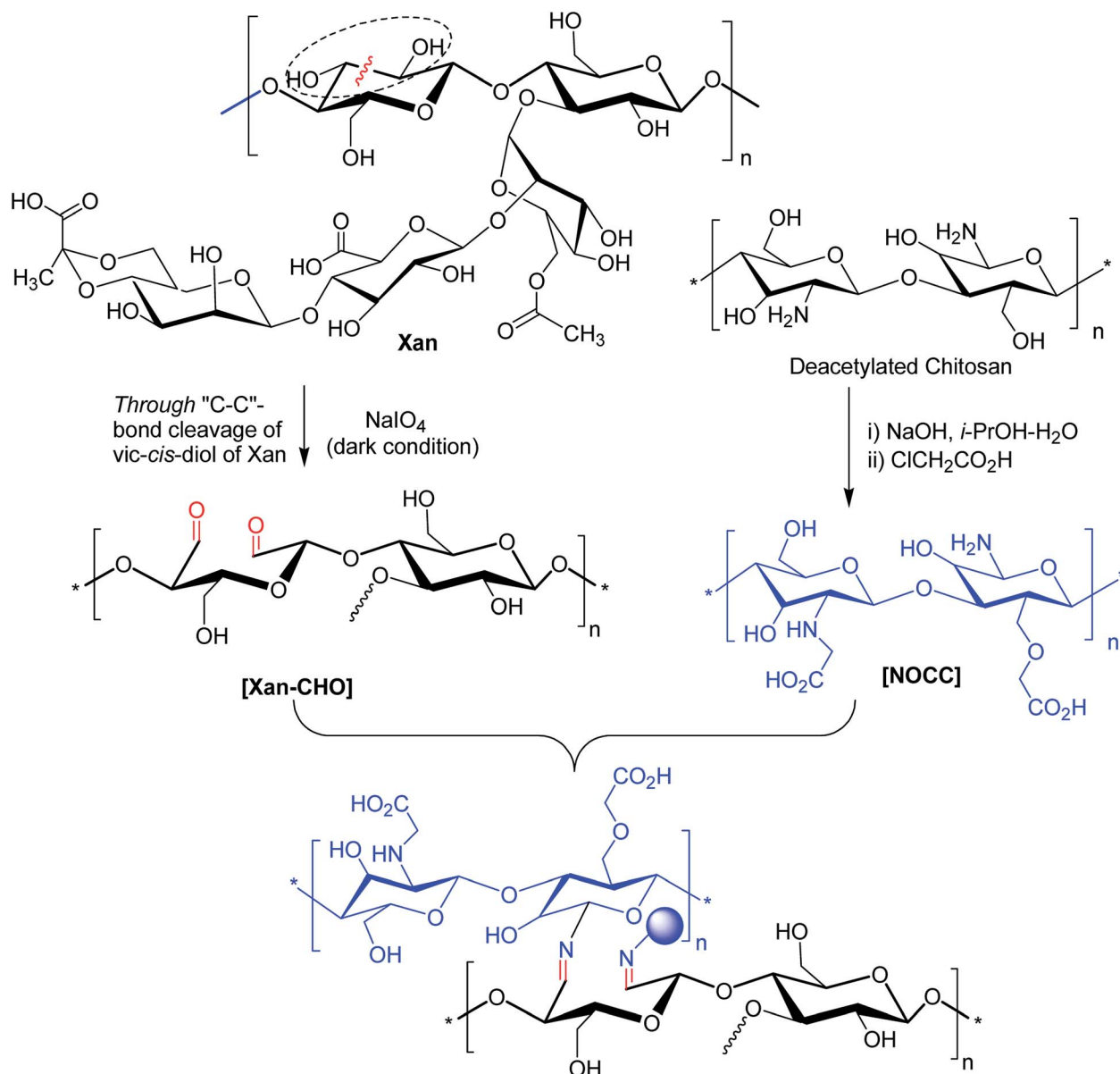


Fig. 11 Formation of XG-CHO/NOCC hydrogel via imine formation.

Mucoadhesive and water vapor uptake properties of XG became more pronounced after thiolation of XG. The patch released tannic acid half as fast as XG in 6 h in simulated saliva solution and thus opened an avenue for sialorrhoea research. Menzel *et al.*¹⁹⁰ adopted a distinct strategy for thiolation of XG. The process comprised of coupling reaction of 2-mercaptopicolinic acid (MNA) to *L*-cysteine by disulfide exchange and direct attachment to XG via carbodiimide mediated amide bond formation (Fig. 13b). XG-cysteine-MNA concurred about 1.7-fold and 2.5-fold higher mucoadhesion than cysteine-XG (XG-Cys) and XG *ex vivo*, respectively. In the presence of H_2O_2 oxidizing agent, XG-cysteine-MNA showed high stability towards oxidation. Reversible ciliary beat frequency in porcine nasal epithelial cell culture indicated low toxicity of all xanthan derivatives. The *in situ* gelling behavior of XG-Cys-MNA was

independent of polyvalent cations, thus hinted safe application of aqueous solutions.

Mendes *et al.*¹⁹¹ synthesized an amphiphilic XG following conjugation of phospholipid (1,2-dioleoyl-*sn*-glycerophosphoethylamine, DOPE) to the anionic XG (Fig. 14). Considering two carboxy groups in xanthan repeating unit, the degree of substitution was estimated to about 1.16 ± 0.024 . The investigators applied microfluidic approach for the production of microcapsules for cell entrapment. Coating with 0.1% poly (*L*-lysine) improved the mechanical strength of the capsules. The viability of ATDC5 cells remained unaltered after encapsulation. The droplets generated through microfluidic approach and the self-assembly of xanthan-DOPE produced microcapsules with an environment compatible for cell survival and proliferation.



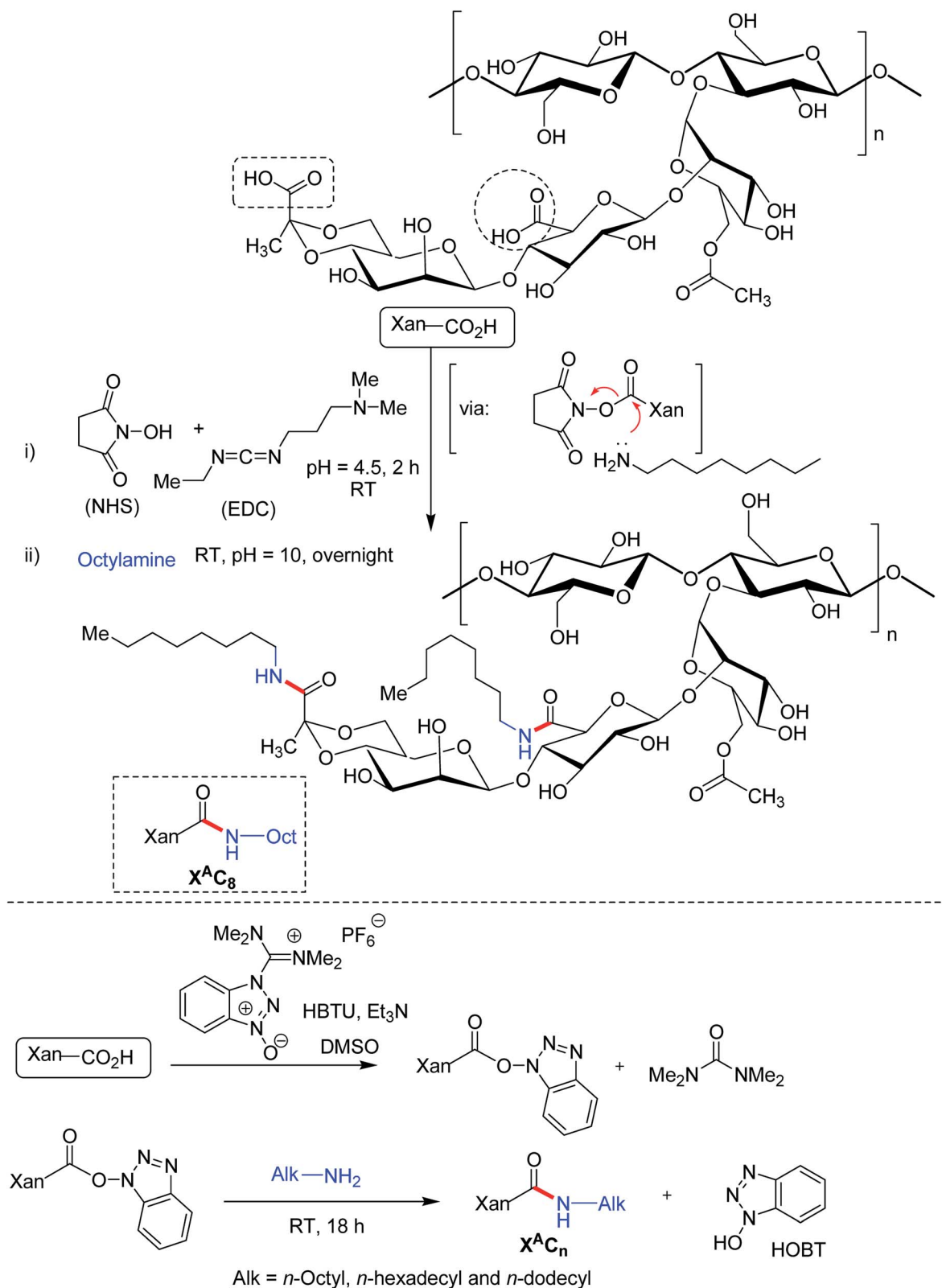


Fig. 12 Long alkyl chain conjugation to XG under its ordered and disordered conformation.

The antimicrobial activity of lysozyme is limited to Gram positive bacteria, but its antibacterial spectrum can be extended towards Gram negative bacteria after carbohydrate conjugation through the Maillard reaction. Hashemi *et al.*¹⁹² synthesized

xanthan gum-lysozyme (1 : 1) conjugate under mild Maillard reaction condition (sodium phosphate buffer, pH 8.5 and 60 °C for 10 days). It was found that approximately 1.9 mmol XG could be attached to one mole lysozyme. Lysozyme-XG conjugates

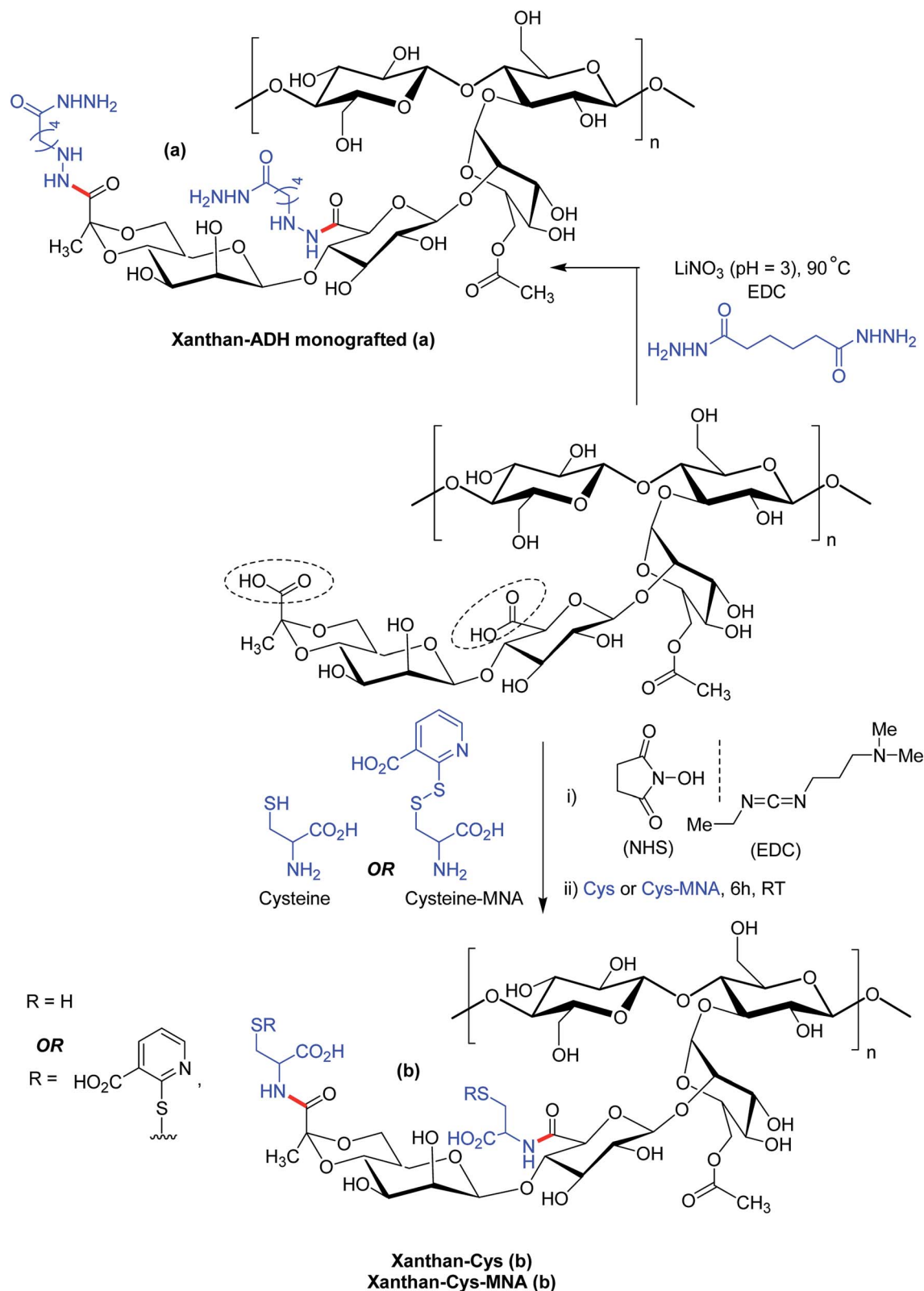


Fig. 13 Synthesis of (a) ADH-reinforced XG and (b) XG-cysteine or XG-cysteine-MNA.

showed pH- and temperature-dependent solubility, and heat stability with improved emulsion and foaming properties. The lysozyme conjugation significantly impeded the growth of

Staphylococcus aureus and *Escherichia coli* in a dose dependent manner. These findings broaden the application of this enzyme in food or pharmaceuticals.



2.5. Acetalated XG

The alcoholic hydroxy groups could react to aldehydes under acidic condition. Thus, the properties of XG could be modified through the formation of acetal linkages in presence of aldehydes. Su *et al.*¹⁰⁴ proclaimed that formaldehyde treatment gave birth to acetal linkages in XG. Despite a loss in crystallinity, the acetalation reaction caused enhancement of viscosity and solubility of XG. Ray *et al.*¹⁹³ described a microparticulate system where they used glutaraldehyde for acetalation of XG and PVA. Regardless of gum : PVA ratio and crosslinker concentration, the microparticles showed 84% drug entrapment efficiency. The crosslinker concentration declined the rate of delivery of entrapped diclofenac molecules considerably *in vitro*. On contrary, XG promoted the drug release rate in biological fluids. Preclinical trials demonstrated higher relative bioavailability (1.69 times) having a good *in vitro-in vivo* correlation. The pioneering work of Ray *et al.*¹⁹³ enlightened us on the potential utility of glutaraldehyde as a crosslinker for XG for controlled drug delivery applications. However, the toxicity profile of glutaraldehyde poses a major concern which perhaps precludes its use as chemical crosslinker for drug delivery matrices (Fig. 15).

2.6. Physically modified XG

XG could be physically modified using various treatments like heat, moisture, annealing, dry heating to change its physico-chemical properties. Sereno *et al.*¹⁹⁴ extruded XG under water flow in a twin screw extruder with co-rotating screws. It was then dried in a vacuum oven at 65 °C for 72 h under a pressure of 100 Pa to retain final water to <8%. The viscosity and dispersibility of 0.75% XG in distilled water was appreciably improved after extrusion processing. Foster and Mitchell¹⁹⁵ noted that the extruded XG behaved like polyelectrolyte particles, demonstrating excellent dispersibility and a strong salt dependence on the degree of swelling. Heating of the extruded XG dispersion resulted in viscosity development identical to starch, citing end use advantages for XG. In another study,¹⁹⁶ it was found that high pressure homogenization (HPH) (276 MPa) precipitated viscosity of XG solution, possibly due to disruption of structural network.¹⁹⁷ Further, this mechanical treatment reduced molecular weight and caused an increase in the hydrodynamic volume and polydispersity of the polymer remarkably. The treatment further led to irreversible change in the rheological and structural characteristics of XG. The HPH-modified XG responded differently to temperature, compared to that observed with XG after low pressure homogenization (LPH, 69 MPa). LPH caused sharp lowering of storage modulus (elasticity) around 50 °C; whereas HPH-treated XG exhibited similar trend around 40 °C. Zhang *et al.*¹⁹⁸ elucidated the swelling, mechanical, rheological and adsorption properties of PVA-XG hydrogel synthesized over three freezing (−15 °C for 24 h) and thawing (room temperature for 1 h) cycles. They postulated that during freezing and thawing of PVA-XG solution, the hydroxy and carboxy functional groups of the polymers assumed a three-dimensional network through hydrogen bonding interactions, and formed a physical hydrogel. The wrinkles were evident on

the hydrogel surface, especially at high XG concentration. Increase in specific surface area of the network, in turn made the hydrogel more elastic in terms of absorption or release of water, anions or cations through a large number of contact points established between the hydrophilic chain and the solvent. A distinct fluid characteristic was noticed. At low shear rate, hydrogels behaved like a Newtonian fluid; whereas the same turned into pseudoplastic non-Newtonian fluid at high shear rate. The shear stress of the composite hydrogel increased with increasing concentration of XG and reached maxima at PVA : XG ratio of 12 : 8. Additionally, the apparent viscosity and shear thinning characteristics of XG progressively increased at higher strength. At higher XG content, more hydrophilic groups entangled into regular network structure by hydrogen bonding and van der Waals forces, which enabled wrapping of more free water. Consequently, the viscosity of the composite solution became larger and pseudoplasticity came into existence.¹⁹⁸ The PVA-XG (12 : 8) hydrogels swelled abruptly at high pH and attained a maximum equilibrium swelling index of 1.3 at pH 8.0. This indicated that alkaline pH favored the adsorption of methylene blue by composite hydrogels. XG contributed to the improved swelling of PVA-XG hydrogels; however reduced the compression properties owing to the formation of hydrogen bonds between polymers. Ultimately, the mechanical properties of the hydrogel diminished. The maximum adsorption of methylene blue dye was noted at PVA : XG ratio of 12 : 8. A strong electrostatic attraction between XG and dye could contribute to this effect. The recycling ability of hydrogels strongly supported its application in dye removal from waste water. Shimizu *et al.*¹⁹⁹ observed that the addition of 0.2% XG in 17% PVA solution minimized liquid viscosity while maintaining mechanical properties and pourability into injection molds during the fabrication of 3D models. The addition of XG facilitated the fabrication of biomodels through gelation of PVA/XG solution in water : DMSO (20 : 80) at −30 °C for 24 h. Bhunia and coworkers²⁰⁰ prepared PVA (10%) : XG (1–5% w/w) hydrogel films following irradiation at 5 kGy dose under high energy electron beam and investigated the influence of low molecular weight (1.4 kDa) PVA (LPVA) and high molecular weight (11.5 kDa) PVA (HPVA) on the physical and mechanical properties of the film. The physical properties of the films under dry state remained indistinguishable but HPVA-XG hybrids were superior in terms of wet-mechanical properties and diltiazem release. Tensile strength of HPVA-XG films was excellent at 1% XG and the same was much less in LPVA-XG films. The degree of swelling became intensified with increasing XG content in HPVA hydrogels.

In LPVA-XG hydrogels, the swelling dropped drastically on addition of 1% XG, but eventually increased at 3% and 5% XG concentration. Thus, the most physico-mechanically balanced hydrogel membranes were obtained at 1% and 5% XG for HPVA and LPVA, respectively. A sharp loss in crystallinity in presence of 1% XG indicated huge loss of cohesive interaction and establishment of higher HPVA-XG interaction. On contrary, gradual increase in XG content up to 5% produced greater crystallinity in LPVA hydrogels. LPVA was surprisingly more viscous than HPVA²⁰¹ and hence, LPVA-XG produced better



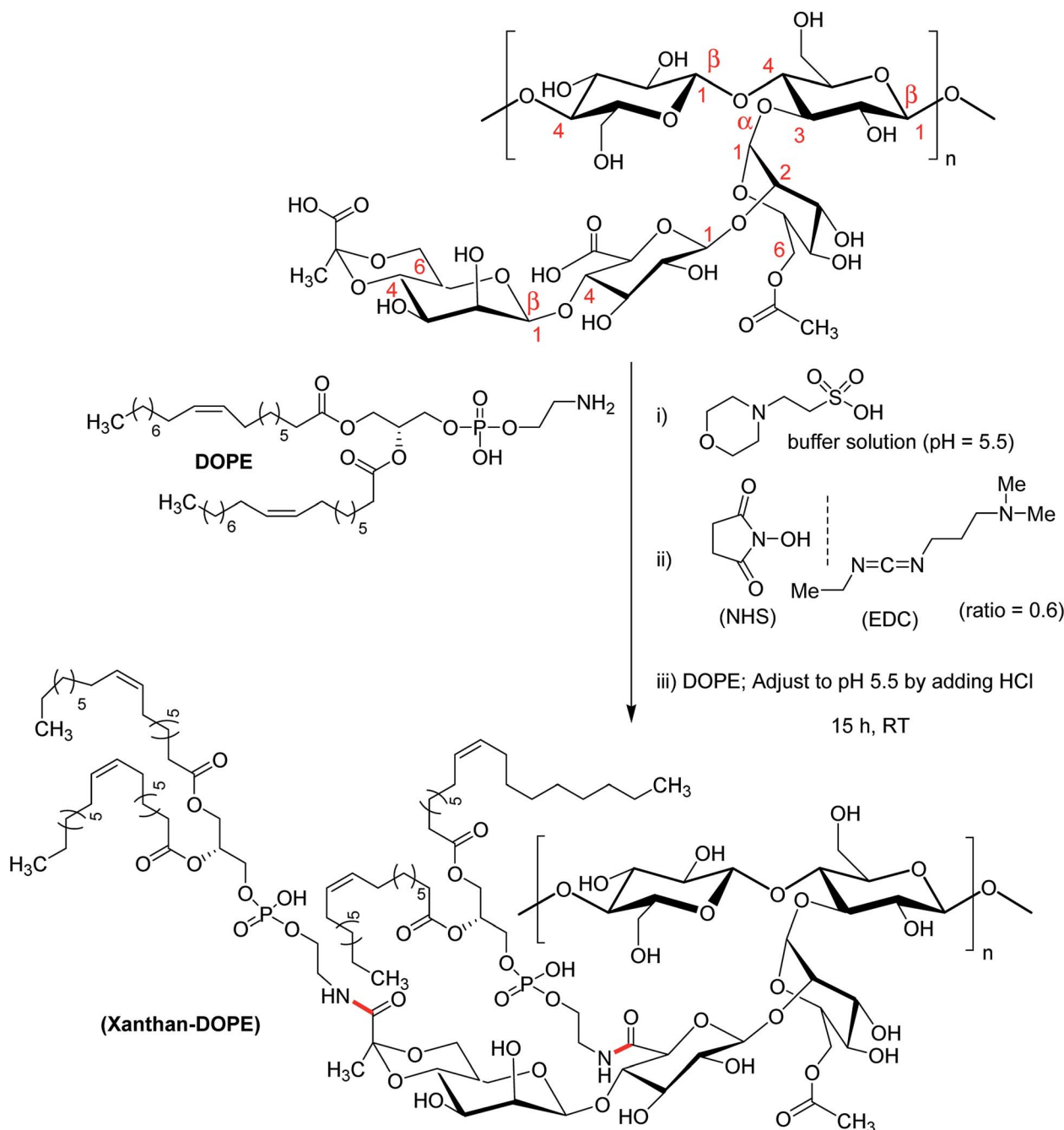


Fig. 14 Synthesis of amphiphilic DOPE-XG.

phase compatibility, particularly at higher XG content. On contrary, intermingling of HPVA and XG generated better phase mixing at low XG content but led to huge phase separation at elevated XG level due to viscosity mismatches. LPVA showed better phase mixing with 5% XG. In HPVA : XG (1%) films, the diltiazem molecules were not uniformly distributed because of their different phase viscosity, thus failed to retain drug appreciably alike LPVA : XG (5%) films. The drug-PVA-XG hydrogen bonding interaction dictated the entrapment of diltiazem within PVA-XG hybrid. High flexibility in LPVA : XG (5%) thus allowed more diltiazem to elute in swelled state despite

their better drug entrapment efficiency than HPVA : XG (1%) films. Regardless of XG concentration, the drug release was insignificant (2.5–4.6% in 12 h). The investigators achieved success in suppressing the burst release of drug from PVA hydrogels following exposure to irradiation. Disha *et al.*²⁰² prepared hydrogels by heating XG, sodium benzoate, and potassium sorbate and glycerin mixture at 85 °C. The biodegradability characteristics of XG persisted over the concentration range of 2–3%. The hydrogels were devoid of antimicrobial activity against *E. coli* and *Klebsiella* spp. The physical cross-linking (H-bonding interactions) of XG with poly (*N*-vinyl



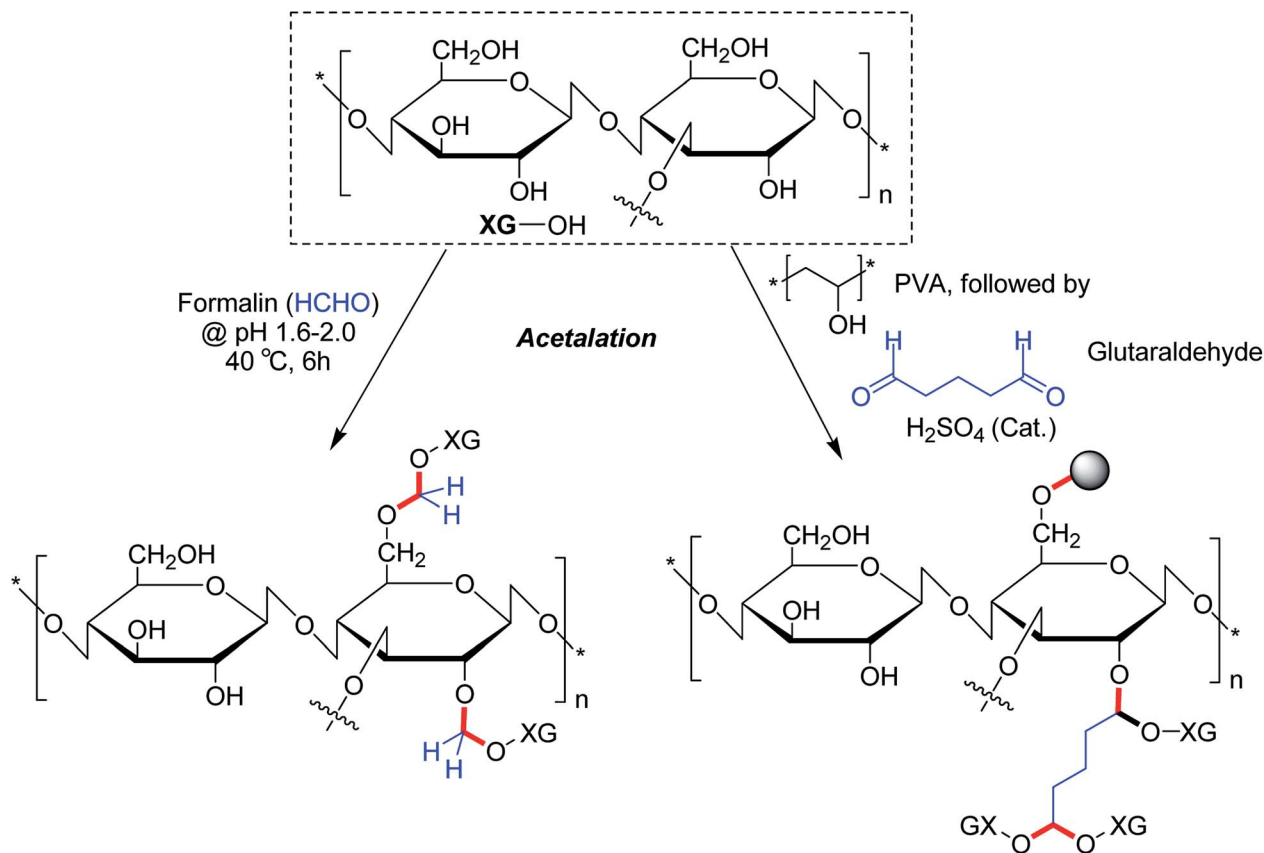


Fig. 15 Acetalation of XG using formaldehyde and glutaraldehyde.

imidazole) enabled Sabaa and coworkers²⁰³ to fabricate hydrogels containing bovine serum albumin (BSA). Depending on variation in gelation time, BSA loading and polymer concentration, the protein entrapment efficiency escalated to 99.17%. Higher polymer concentration promoted BSA release in PBS. The biocompatibility of hydrogel was ascertained on VERO cell line. SDS-PAGE analysis confirmed the structural integrity of protein after encapsulation or release.

Intra-articular injection of hyaluronic acid could be effective in treating osteoarthritis due to its lubricating and cushioning properties.^{204,205} However, hyaluronic acid rapidly degrades *in vivo* by hydrolytic or enzymatic reactions.²⁰⁶ Henceforth, a polymer which is similar to hyaluronic acid in structure and function, but with a longer effect in the joint is needed. XG is similar to hyaluronic acid in rheology and viscosity.²⁰⁷ Additionally, XG solution is very stable in a wide range of pH, ionic strength and temperature.²⁰⁸ XGlyase is mainly produced by bacterium;²⁰⁹ therefore XG is not easily degraded *in vivo*. Intra-articular injection of XG could be an effective therapy for osteoarthritis. Han *et al.*²¹⁰ extensively purified XG, dissolved in buffered physiological saline and sterilized at 121 °C for 15 min and evaluated the protective effect of XG injection on articular cartilage. Results indicated that injection of XG could protect the joint cartilage and reduce papain-induced osteoarthritis progression in rabbit model. The same result was obtained at an injection frequency lower than HA. This finding seemed promising in the

development of a new therapeutic method for osteoarthritis. Bueno *et al.*⁶⁵ synthesized XG-hydroxyapatite particles following homogenization of XG solution with Ca(OH)₂ and H₃PO₄ solution under pH 7.5. Similarly, xanthan strontium-substituted hydroxyapatite particles were obtained using Ca(OH)₂, Sr(CH₃-COO)₂, xanthan solution and H₃PO₄ solution at pH 7.5. Initially Ca²⁺ ions were dissolved at pH 10.0 in presence of xanthan chains. Calcium ions were chelated by xanthan chains, forming nucleation sites for hydroxyapatite crystal growth. As phosphoric acid was added to the solution, the medium pH decreased to ~7.5, promoting the precipitation of Ca²⁺ ions as hydroxyapatite. XG-hydroxyapatite and xanthan strontium-substituted hydroxyapatite nanoparticles presented mean size of 56 ± 18 nm and 75 ± 18 nm, respectively. Xanthan aqueous solution containing nanoparticles was casted into xanthan-based nanocomposite hydrogels in presence of citric acid. The dehydration crosslinking was accomplished by heating the films at 165 °C for 7 min. The nanocomposite films were apparently homogeneous. Hence, the presence of xanthan chains on the surface of particles provided an indication of compatibility between particles and xanthan chains. Nanocomposites presented porous structure and the incorporation of Sr or xanthan-hydroxyapatites did not affect the surface energy and consequently, posed no substantial influence on osteoblast proliferation. On the other hand, the chemical composition of nanocomposites influenced ALP activity. ALP is a cell membrane glycoprotein that catalyzes the hydrolysis of



phosphate esters in alkaline pH and thus plays an important role in the mineralization process of bone matrix. Its activity in human serum is used as indication for diseases related to liver and/or bone. It was noted that the osteoblast ALP activity increased as the filler content was increased from 10% to 30% in both the nanocomposites, but this enhancement was much more pronounced in case of citric acid crosslinked XG–strontium hydroxyapatite. In fact, the presence of strontium in hydroxyapatite structure supported osteoblast and osteoclast growth. Nanocomposites made of crosslinked xanthan chains and xanthan hydroxyapatite (10%) was homogeneous and mechanically stronger than bare xanthan networks or nanocomposites with hydroxyapatites. Mixing of ZnO nanoparticles (size 15–25 nm) with XG–PVA mixture followed by irradiation at 30 kGy led to the generation of biocompatible XG–PVA/ZnO nanocomposite dressing hydrogels.²¹¹ Homogeneous porous hydrogel network and the presence of ZnO nanoparticles provided an excellent fluid uptake capacity in pseudo-extracellular fluid (554–664%) and water (1281–1603%). The water retention capacity (about 50–65% after air exposure for 6 h) and water vapor transmission rate (167–184 g m⁻² h⁻¹) was sufficient to keep wound's surface moist. ZnO dressings exhibited profound antimicrobial activity against *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*. To avoid lumps and accelerate the dissolution of XG in water, Zhao *et al.*²¹² modified the surfaces of XG particles in dimethylbenzene with organometallic complex COMe at pH 4.5 adjusted by citric acid. The modification didn't change the molecular structure of XG; however weakened the hydrogen bonding interaction between XG molecules. Dispersible XG powders were completely dissolved in simulated seawater in 35 min without lumps. It was reported that sequential control of wetting and drying of XG produced sticky samples, thus posed difficulty in handling.²¹³ Co-grounding of this treated XG with mannitol (1 : 1) improved the flowability and crystallinity marginally. Co-grounding afforded excellent water loading capacity (60.93%) and thus appeared to be an effective disintegrant for roxithromycin orodispersible tablets. The modified XG (10% w/w) and microcrystalline cellulose (MCC) (15%) allowed the disintegration of tablets in less than 60 s. At 1 : 3 ratio of co-grounded XG : microcrystalline cellulose, the tablets even disintegrated in about 14 s. The tablets reduced the lag time 9-fold and remained stable over a period of 12 months. Highly porous structure of modified XG and higher level of MCC promoted faster wetting (6.52 ± 0.42 s) and water absorption (150%). A marginal difference in drug release was noted at salivary pH (pH 6.4) and physiological pH (pH 7.4) (~91–93% in 30 min). Higher percentage of drug release could provoke drug absorption through buccal cavity. Above all, the modified XG behaved like a directly compressible excipient and exhibited swelling dynamics suitable for use in rapidly disintegrating tablets.

3. Conclusion

This review inferred that the physicochemical and mechanical properties of virgin XG could be tailored *via* etherification, esterification, oxidation, acetalation and amidation for specific

end use. The ionic and/covalent crosslinking of XG allowed the fabrication of hydrogel films, hydrogel beads, matrix tablets for oral and transdermal drug delivery applications. The conjugation of long alkyl chain to XG backbone triggered self-assembly of XG and facilitated incorporation of substantial amount of drug molecules in the lipophilic confines of nanomicelles. The octyl XG unveiled its potential use as emulsifying agent. L-Cysteine conjugation to XG further assisted in developing buccal patch for sialorrhoea treatment. Even, the phospholipid conjugation to anionic XG followed by poly(L-lysine) cross-linking proffered an environment adequate for cell survival and proliferation. Thiolation markedly improved mucoadhesion of XG. The esterification of XG further conferred self-healing as well as sustained drug release properties. The esterified hydrogels seemed suitable as a substitute for human nucleus pulposus. The self-healing feature of aldehyde XG–carboxymethyl chitosan hydrogels was also found promising.

The acetalated XG hydrogels improved the pharmacokinetics of entrapped drug remarkably. The physical treatments such as extrusion, freezing–thawing and high shear homogenization greatly improved the mechanical properties of XG. In addition, the physically crosslinked XG hydrogels had immense potential in encapsulating proteins without alteration of its structural integrity. The intra-articular injection of purified XG on articular cartilage was found effective as well against osteoarthritis. The XG-based nanocomposites had shown excellent mechanical stability, osteoblast ALP activity as well as wound dressing capability. Even, simple co-grounding of XG with mannitol caused reasonable improvement in flow characteristics of XG. Overall, the findings suggested that the functionalized XG had great potential for pharmaceutical and biomedical applications. However, the *in vivo* potential of these novel XG derivatives are yet to be assessed explicitly in order to unveil their safety and efficacy towards drug delivery and biomedical applications. This review would certainly encourage the pharmaceutical and biomedical researchers to explore other polysaccharides for developing newer materials for commercial use.

Conflicts of interest

There are no conflicts to declare.

References

- 1 Y. Luo and Q. Wang, Recent development of chitosan-based polyelectrolyte complexes with natural polysaccharides for drug delivery, *Int. J. Biol. Macromol.*, 2014, **64**, 353–367, DOI: 10.1016/j.ijbiomac.2013.12.017.
- 2 S. Dumitriu, *Polysaccharides: Structural Diversity and Functional Versatility*, Marcel Dekker, New York, 2nd edn, 2005.
- 3 F. García-Ochoa, V. E. Santos, J. A. Casas and E. Gómez, Xanthan gum: production, recovery, and properties, *Biotechnol. Adv.*, 2000, **18**, 549–579, DOI: 10.1016/S0734-9750(00)00050-1.
- 4 D. Bergmann, G. Furth and C. Mayer, Binding of bivalent cations by xanthan in aqueous solution, *Int. J. Biol.*



- Macromol.*, 2008, **43**, 245–251, DOI: 10.1016/j.ijbiomac.2008.06.001.
- 5 A. F. Dário, L. M. A. Hortêncio, M. R. Sierakowski, J. C. Queiroz Neto and D. F. S. Petri, The effect of calcium salts on the viscosity and adsorption behavior of xanthan, *Carbohydr. Polym.*, 2011, **84**, 669–676, DOI: 10.1016/j.carbpol.2010.12.047.
 - 6 T. A. Camesano and K. J. Wilkinson, Single molecule study of xanthan conformation using atomic force microscopy, *Biomacromolecules*, 2001, **2**, 1184–1191, DOI: 10.1021/bm015555g.
 - 7 H. Li, W. Hou and X. Li, Interaction between xanthan gum and cationic cellulose JR400 in aqueous solution, *Carbohydr. Polym.*, 2012, **89**, 24–30, DOI: 10.1016/j.carbpol.2012.02.022.
 - 8 J. A. Carmona, A. Lucas, P. Ramírez, N. Calero and J. Muñoz, Nonlinear and linear viscoelastic properties of a novel type of xanthan gum with industrial applications, *Rheol. Acta*, 2015, **54**, 993–1001, DOI: 10.1007/s00397-015-0888-1.
 - 9 G. Bumphrey, Extremely useful new suspending agent, *Pharm. J.*, 1986, **237**, 665–671.
 - 10 V. B. Junyaprasert and G. Manwiwattanakul, Release profile comparison and stability of diltiazem-resin microcapsules in sustained release suspensions, *Int. J. Pharm.*, 2008, **352**, 81–91, DOI: 10.1016/j.ijpharm.2007.10.018.
 - 11 B. Katzbauer, Properties and applications of xanthan gum, *Polym. Degrad. Stab.*, 1998, **59**, 81–84, DOI: 10.1016/S0141-3910(97)00180-8.
 - 12 R. Geremia and M. Rinaudo, Biosynthesis, structure, and physical properties of some bacterial polysaccharides, in *Polysaccharides: Structural Diversity and Functional Versatility*, ed. S. Dumitriu, Marcel Dekker, New York, 2005, pp. 411–430.
 - 13 S. K. Psomas, M. Liakopoulou-Kyriakides and D. A. Kyriakidis, Optimization study of xanthan gum production using response surface methodology, *Biochem. Eng. J.*, 2007, **35**, 273–280, DOI: 10.1016/j.bej.2007.01.036.
 - 14 A. Palaniraj and V. Jayaraman, Production, recovery and applications of xanthan gum by *Xanthomonas campestris*, *J. Food Eng.*, 2011, **106**, 1–12, DOI: 10.1016/j.jfoodeng.2011.03.035.
 - 15 Y. L. Yang, L. Ding, J. Zhang, Y. L. Zhang, J. Yao and D. F. Cui, The study on salt-resistant stability of sophora bean gum and mixed gum, *J. Northwest Norm. Univ.*, 2001, **37**, 70–72.
 - 16 B. Tinland and M. Rinaudo, Dependence of the stiffness of the xanthan chain on the external salt concentration, *Macromolecules*, 1989, **22**, 1863–1865, DOI: 10.1021/ma00194a058.
 - 17 Z. Liu and P. Yao, Injectable thermo-responsive hydrogel composed of xanthan gum and methylcellulose double networks with shear-thinning property, *Carbohydr. Polym.*, 2015, **132**, 490–498, DOI: 10.1016/j.carbpol.2015.06.013.
 - 18 Z. Liu and P. Yao, Injectable shear-thinning xanthan gum hydrogel reinforced by mussel-inspired secondary crosslinking, *RSC Adv.*, 2015, **5**, 103292–103301, DOI: 10.1039/c5ra17246b.
 - 19 T. M. Bresolin, M. Milas, M. Rinaudo and J. L. Ganter, Xanthan-galactomannan interactions as related to xanthan conformations, *Int. J. Biol. Macromol.*, 1998, **23**, 263–275, DOI: 10.1016/S0141-8130(98)00061-0.
 - 20 A. S. Hoffman, Hydrogels for biomedical applications, *Adv. Drug Delivery Rev.*, 2012, **64**, 18–23.
 - 21 V. B. Bueno, R. Bentini, L. H. Catalani and D. F. Petri, Synthesis and swelling behavior of xanthan-based hydrogels, *Carbohydr. Polym.*, 2013, **92**, 1091–1099, DOI: 10.1016/j.carbpol.2012.10.062.
 - 22 P. Giannouli and E. R. Morris, Cryogelation of xanthan, *Food Hydrocolloids*, 2003, **17**, 495–5016, DOI: 10.1016/S0268-005X(03)00019-5.
 - 23 M. Carafa, C. Marianecchi, L. Di Marzio, F. Rinaldi, C. Meo, P. Matricardi, F. Alhaique and T. Coviello, A new vesicle-loaded hydrogel system suitable for topical applications: preparation and characterization, *J. Pharm. Pharm. Sci.*, 2011, **14**, 336–346, DOI: 10.18433/j3160b.
 - 24 F. Shi, X. Wang, R. Guo, M. Zhong and X. M. Xie, Highly stretchable and super tough nanocomposite physical hydrogels facilitated by coupling of intermolecular hydrogen bond and analogous chemical crosslinking of nanoparticles, *J. Mater. Chem. B*, 2015, **3**, 1187–1192, DOI: 10.1039/c4tb01654h.
 - 25 L. Gao, W. Dai, J. Chen, Z. Xie and X. Yue, Enhanced electro-responsive behaviors of agar/xanthan gum interpenetrating compound hydrogel, *Soft Mater.*, 2017, **15**, 163–172, DOI: 10.1080/1539445X.2017.1282872.
 - 26 F. Li, Y. Zhu, B. You, D. Zhao, Q. Ruan, Y. Zeng and C. Ding, Smart hydrogels co-switched by hydrogen bonds and π - π stacking for continuously regulated controlled-release system, *Adv. Funct. Mater.*, 2010, **20**, 669–676, DOI: 10.1002/adfm.200901245.
 - 27 G. Li, J. Wu, B. Wang, S. Yan, K. Zhang, J. Ding and J. Yin, Self-healing supramolecular self-assembled hydrogels based on poly(L-glutamic acid), *Biomacromolecules*, 2015, **16**, 3508–3518, DOI: 10.1021/acs.biomac.5b01287.
 - 28 W. J. Malfait and C. Sanchez-Valle, Effect of water and network connectivity on glass elasticity and melt fragility, *Chem. Geol.*, 2013, **346**, 72–80, DOI: 10.1016/j.chemgeo.2012.04.034.
 - 29 B. T. Stokke, B. E. Christensen and O. Smidsrød, in *Polysaccharides: structural, diversity and functional versatility*, ed. S. Dumitriu, Marcel Dekker, New York, 2nd edn, 1998, p. 436.
 - 30 H. Nolte, S. John, O. Smidsrød and B. T. Stokke, Gelation of xanthan with trivalent metal ions, *Carbohydr. Polym.*, 1992, **18**, 243–251, DOI: 10.1016/0144-8617(92)90089-9.
 - 31 A. B. Rodd, D. E. Dunstan, D. V. Boger, J. Schmidt and W. Burchard, Heterodyne and nonergodic approach to dynamic light scattering of polymer gels: aqueous Xanthan in the presence of metal ions (aluminum(III)), *Macromolecules*, 2001, **34**, 3339–3352, DOI: 10.1021/ma001706g.
 - 32 T. Pongjanyakul and S. Puttipipatkachorn, Xanthan-alginate composite gel beads: molecular interaction and



- in vitro characterization, *Int. J. Pharm.*, 2007, **331**, 61–71, DOI: 10.1016/j.ijpharm.2006.09.011.
- 33 N. Sharma, R. D. Deshpande, D. Sharma and R. K. Sharma, Development of locust bean gum and xanthan gum based biodegradable microparticles of celecoxib using a central compositedesign and its evaluation, *Ind. Crops Prod.*, 2016, **82**, 161–170, DOI: 10.1016/j.indcrop.2015.11.046.
- 34 R. M. Mounica, V. Shanmugam and K. Rajesh, Design and characterization of insulin nanoparticles for oral delivery, *Int. J. Innovative Pharm. Res.*, 2012, **3**, 238–243.
- 35 D. Pooja, S. Panyaram, H. Kulhari, S. S. Rachamalla and R. Sistla, Xanthan gum stabilized gold nanoparticles: characterization, biocompatibility, stability and cytotoxicity, *Carbohydr. Polym.*, 2014, **110**, 1–9, DOI: 10.1016/j.carbpol.2014.03.041.
- 36 G. Copetti, M. Grassi, R. Lapasin and S. Pricl, Synergistic gelation of xanthan gum with locust bean gum: a rheological investigation, *Glycoconjugate J.*, 1997, **14**, 951–961, DOI: 10.1023/a:1018523029030.
- 37 M. P. Venkataraju, D. V. Gowda, K. S. Rajesh and H. G. Shivakumar, Xanthan and locust bean gum (from *Ceratonia siliqua*) matrix tablets for oral controlled delivery of propranolol hydrochloride, *Asian J. Pharm. Sci.*, 2007, **2**, 239–248, DOI: 10.2174/157488508783331216.
- 38 W. Xu, W. Jin, L. Lin, C. Zhang, Z. Li, Y. Li, R. Song and B. Li, Green synthesis of xanthan conformation-based silver nanoparticles: antibacterial and catalytic application, *Carbohydr. Polym.*, 2014, **101**, 961–967, DOI: 10.1016/j.carbpol.2013.10.032.
- 39 A. Ebrahiminezhad, M. Zare, S. Kiyampour, A. Berenjian, S. V. Niknezhad and Y. Ghasemi, Biosynthesis of xanthan gum-coated INPs by using *Xanthomonas campestris*, *IET Nanobiotechnol.*, 2018, **12**, 254–258, DOI: 10.1049/iet-nbt.2017.0199.
- 40 M. Manconi, S. Mura, M. L. Manca, A. M. Fadda, M. Dolz, M. J. Hernandez, A. Casanova and O. Diez-Sales, Chitosomes as drug delivery systems for C-phycocyanin: preparation and characterization, *Int. J. Pharm.*, 2010, **392**, 92–100, DOI: 10.1016/j.ijpharm.2010.03.038.
- 41 M. L. Manca, M. Manconi, D. Valenti, F. Lai, G. Loy, P. Matricardi and A. M. Fadda, Liposomes coated with chitosan–xanthan gum (chitosomes) as potential carriers for pulmonary delivery of rifampicin, *J. Pharm. Sci.*, 2012, **101**, 566–575, DOI: 10.1002/jps.22775.
- 42 V. Dhopeshwarkar and J. L. Zatz, Evaluation of xanthan gum in the preparation of sustained release matrix tablets, *Drug Dev. Ind. Pharm.*, 1993, **19**, 999–1017, DOI: 10.3109/03639049309062997.
- 43 M. M. Talukdar and R. Kinget, Swelling and drug release behavior of xanthangum matrix tablets, *Int. J. Pharm.*, 1995, **120**, 63–72, DOI: 10.1016/0378-5173(94)00410-7.
- 44 M. M. Talukdar, G. Van den Mooter, P. Augustijns, T. Tjandra-Maga, N. Verbeke and R. Kinget, In vivo evaluation of xanthan gum as a potential excipient for oral controlled-release matrix tablet formulation, *Int. J. Pharm.*, 1998, **169**, 105–113, DOI: 10.1016/s0378-5173(98)00112-4.
- 45 N. Billa, Y. Kah-Hay, A. K. Mohamed Ali and O. Alias, Gamma scintigraphic study of the gastrointestinal transit and in vivo dissolution of a controlled release diclofenac sodium formulation in xanthan gum matrices, *Int. J. Pharm.*, 2000, **201**, 109–120, DOI: 10.1016/s0378-5173(00)00399-9.
- 46 O. N. El-Gazayerly, Release of pentoxifylline from xanthan gum matrix tablets, *Drug Dev. Ind. Pharm.*, 2003, **29**, 241–246, DOI: 10.1081/ddc-120016732.
- 47 R. Kar, S. Mohapatra, S. Bhanja, D. Das and B. Barik, Formulation and in vitro characterization of xanthan gum-based sustained release matrix tables of isosorbide-5-mononitrate, *Iran. J. Pharm. Res.*, 2010, **9**, 13–19.
- 48 V. F. Patel and N. M. Patel, Statistical evaluation of influence of xanthan gum and guar gum blends on dipyridamole release from floating matrix tablets, *Drug Dev. Ind. Pharm.*, 2007, **33**, 327–334, DOI: 10.1080/03639040601050155.
- 49 M. F. Lu, L. Woodward and S. Borodkin, Xanthan Gum and alginate based controlled release theophylline formulations, *Drug Dev. Ind. Pharm.*, 1991, **17**, 1987–2004, DOI: 10.3109/03639049109048063.
- 50 W. M. Zeng, Oral controlled release formulation for highly water-soluble drugs: drug–sodium alginate–xanthan gum–zinc acetate matrix, *Drug Dev. Ind. Pharm.*, 2004, **30**, 491–495, DOI: 10.1081/ddc-120037479.
- 51 K. Ofori-Kwakye, K. A. Mfofo, S. L. Kipo, N. Kuntworbe and M. E. Boakye-Gyasi, Development and evaluation of natural gum-based extended release matrix tablets of two model drugs of different water solubilities by direct compression, *Saudi Pharm. J.*, 2016, **24**, 82–91, DOI: 10.1016/j.jsps.2015.03.005.
- 52 J. Fan, K. Wang, M. Liu and Z. He, In vitro evaluations of konjac glucomannan and xanthan gum mixture as the sustained release material of matrix tablet, *Carbohydr. Polym.*, 2008, **73**, 241–247, DOI: 10.1016/j.carbpol.2007.11.027.
- 53 H. Jian, L. Zhu, W. Zhang, D. Sun and J. Jiang, Galactomannan (from *Gleditsia sinensis* Lam.) and xanthan gum matrix tablets for controlled delivery of theophylline: in vitro drug release and swelling behavior, *Carbohydr. Polymer*, 2012, **87**, 2176–2182, DOI: 10.1016/j.carbpol.2011.10.043.
- 54 C. W. Vendruscolo, I. F. Andrezza, J. L. M. S. Ganter, C. Ferrero and T. M. B. Bresolin, Xanthan and galactomannan (from *M. scabrella*) matrix tablets for oral controlled delivery of theophylline, *Int. J. Pharm.*, 2005, **296**, 1–11, DOI: 10.1016/j.ijpharm.2005.02.007.
- 55 T. Phaechamud and G. C. Ritthidej, Sustained-release from layered matrix system comprising chitosan and xanthan gum, *Drug Dev. Ind. Pharm.*, 2007, **33**, 595–605, DOI: 10.1080/03639040601015521.
- 56 M. Fukuda, N. A. Peppas and J. W. McGinity, Properties of sustained release hot-melt extruded tablets containing chitosan and xanthan gum, *Int. J. Pharm.*, 2006, **310**, 90–100, DOI: 10.1016/j.ijpharm.2005.11.042.



- 57 F. Chellat, M. Tabrizian, S. Dumitriu, E. Chornet, P. Magny, C. H. Rivard and L. Yahia, In vitro and in vivo biocompatibility of chitosan-xanthan polyionic complex, *J. Biomed. Mater. Res.*, 2000, **51**, 107–116, DOI: 10.1002/(sici)1097-4636(200007)51:1<107::aid-jbm14>3.0.co;2-f.
- 58 A. A. Salyers, J. R. Vercellotti, S. H. E. West and T. D. Wilkins, Fermentation of mucin and plant polysaccharides by strains of bacteroides from the human colon, *Appl. Environ. Microbiol.*, 1977, **33**, 319–322, PMID:PMC170684.
- 59 V. R. Sinha and R. Kumria, Binders for colon specific drug delivery: an in vitro evaluation, *Int. J. Pharm.*, 2002, **249**, 23–31, DOI: 10.1016/S0378-5173(02)00398-8.
- 60 F. Alvarez-Manceño, M. Landin and R. Martínez-Pacheco, Konjac glucomannan/xanthan gum enzyme sensitive binary mixtures for colonic drug delivery, *Eur. J. Pharm. Biopharm.*, 2008, **69**, 573–581, DOI: 10.1016/j.ejpb.2008.01.004.
- 61 C. Caddeo, A. Nácher, O. Díez-Sales, M. Merino-Sanjuán, A. M. Fadda and M. Manconi, Chitosan-xanthan gum microparticle-based oral tablet for colon-targeted and sustained delivery of quercetin, *J. Microencapsulation*, 2014, **31**, 694–699, DOI: 10.3109/02652048.2014.913726.
- 62 K. Niranjan, A. Shivapooja, J. Muthyala and P. Pinakin, Effect of guar gum and xanthan gum compression coating on release studies of metronidazole in human fecal media for colon targeted drug delivery systems, *Asian J. Pharm. Clin. Res.*, 2013, **6**, 315–318.
- 63 N. Almeida, A. Mueller, S. Hirschi and L. Rakesh, Rheological studies of polysaccharides for skin scaffolds, *J. Biomed. Mater. Res.*, 2014, **102**, 1510–1517, DOI: 10.1002/jbm.a.34805.
- 64 M. Z. Bellini, C. Caliar-Oliveira, A. Mizukami, K. Swiech, D. T. Covas, E. A. Donadi, P. Oliva-Neto and Â. M. Moraes, Combining xanthan and chitosan membranes to multipotent mesenchymal stromal cells as bioactive dressings for dermo-epidermal wounds, *J. Biomater. Appl.*, 2015, **29**, 1155–1166, DOI: 10.1177/0885328214553959.
- 65 V. B. Bueno, R. Bentini, L. H. Catalani, L. R. Barbosa and D. F. Petri, Synthesis and characterization of xanthan-hydroxyapatite nanocomposites for cellular uptake, *Mater. Sci. Eng., C*, 2014, **37**, 195–203, DOI: 10.1016/j.msec.2014.01.002.
- 66 V. B. Bueno, S. H. Takahashi, L. H. Catalani, S. Torresi and D. F. Siqueira Petri, Biocompatible xanthan/polypyrrole scaffolds for tissue engineering, *Mater. Sci. Eng., C*, 2015, **52**, 121–128, DOI: 10.1016/j.msec.2015.03.023.
- 67 A. Kumar, K. M. Rao, S. E. Kwon, Y. N. Lee and S. S. Han, Xanthan gum/bioactive silica glass hybrid scaffolds reinforced with cellulose nanocrystals: morphological, mechanical and in vitro cytocompatibility study, *Mater. Lett.*, 2017, **193**, 274–278, DOI: 10.1016/j.matlet.2017.01.143.
- 68 S. Türker, E. Onur and Y. Ozer, Nasal route and drug delivery systems, *Pharm. World Sci.*, 2004, **26**, 137–142, DOI: 10.1023/b:phar.0000026823.82950.ff.
- 69 R. Bommer, Drug delivery-nasal route, in *Encyclopedia of Pharmaceutical Technology*, ed. J. Swarbrick and J. C. Boylan, Marcel Dekker, New York, 2nd edn, 2011, p. 854.
- 70 S. A. Pagar, D. M. Shinkar and R. B. Saudagar, Development and evaluation of in situ nasal mucoadhesive gel of metoprolol succinate by using 3² full factorial design, *Int. J. Pharm. Pharm. Sci.*, 2014, **6**, 218–223.
- 71 N. S. Malekar, S. B. Gondkar, B. A. Bhairav, P. S. Paralkar and R. B. Saudagar, Development of naratriptan hydrochloride in-situ nasal gel, *Res. J. Pharm. Technol.*, 2017, **10**, 979–985.
- 72 S. D. Manohar, P. B. Anil and S. R. Bhanudas, Formulation and evaluation of in-situ mucoadhesive nasal gel of venlafaxine hydrochloride, *International Journal of Institutional Pharmacy and Life Sciences*, 2015, **5**, 185–198.
- 73 A. K. Mitra, *Ophthalmic Drug Delivery System*, Marcel Dekker, New York, 1993, vol. 58, pp. 105–110.
- 74 M. Bhowmik, P. Kumari, G. Sarkar, M. K. Bain, B. Bhowmick, M. M. R. Mollick, D. Mondal, D. Maity, D. Rana, D. Bhattacharjee and D. Chattopadhyay, Effect of xanthan gum and guar gum on in situ gelling ophthalmic drug delivery system based on poloxamer-407, *Int. J. Biol. Macromol.*, 2013, **62**, 117–123, DOI: 10.1016/j.ijbiomac.2013.08.024.
- 75 A. L. Deulker, A. Sancoaltar, S. Vaidya and R. Gude, Formulation development and evaluation of long acting ophthalmic in-situ gelling system of dorzolamide hydrochloride, *Int. J. Drug Dev. Res.*, 2013, **5**, 156–163.
- 76 S. S. P. Hiremath, F. S. Dasankoppa, A. Nadaf, V. G. Jamakandi, J. S. Mulla, S. A. Sreenivas, H. N. Sholapur, Aezazahmed and N. G. Nanjundaswamy, Formulation and evaluation of a novel in situ gum based ophthalmic drug delivery system of linezolid, *Sci. Pharm.*, 2008, **76**, 515–532, DOI: 10.3797/scipharm.0803-17.
- 77 P. Viram and A. N. Lumbhani, Development and evaluation of ion-dependent in-situ nasal gelling systems of metoclopramide hydrochloride as an antimigraine model drug, *International Journal of Latest Research in Science and Technology*, 2012, **1**, 80–89.
- 78 V. A. Ahmed and D. Goli, Development and characterization of in situ gel of xanthan gum for ophthalmic formulation containing brimonidine tartrate, *Asian J. Pharm. Clin. Res.*, 2018, **11**, 277–284, DOI: 10.22159/ajpcr.2018.v11i7.25221.
- 79 S. Kala, P. Gurudiwan and D. Juyal, Formulation and evaluation of besifloxacin loaded in situ gel for ophthalmic delivery, *Pharm. Biosci. J.*, 2018, **6**, 36–40, DOI: 10.20510/ukjpb/6/i2/175583.
- 80 D. H. Shastri, S. T. Prajapati and L. D. Patel, Design and development of thermoreversible ophthalmic in situ hydrogel of moxifloxacin HCl, *Curr. Drug Delivery*, 2010, **7**, 238–243, DOI: 10.2174/156720110791560928.
- 81 N. Morsia, M. Ibrahim, H. Refaia and H. E. Sorogy, Nanoemulsion-based electrolyte triggered in situ gel for ocular delivery of acetazolamide, *Eur. J. Pharm. Sci.*, 2017, **104**, 302–314, DOI: 10.1016/j.ejps.2017.04.013.
- 82 K. J. R. Kumar, E. Jayachandran, S. Muralidharan and S. A. Dhanaraj, Development and in-vitro evaluation of



- Xanthan gum based ion induced solution to gel systems containing Ciclopirox Olamine for vaginal thrush, *J. Pharm. Res.*, 2012, 5, 1673–1678.
- 83 R. Nagendra, R. S. Pai and G. Singh, Design and optimization of novel in situ gel of mercaptopurine for sustained drug delivery, *Braz. J. Pharm. Sci.*, 2014, 50, 107–119, DOI: 10.1590/s1984-82502011000100011.
- 84 J. Ceulemans, I. Vinckier and A. Ludwig, The use of xanthan gum in an ophthalmic liquid dosage form: rheological characterization of the interaction with mucin, *J. Pharm. Sci.*, 2002, 91, 1117–1127, DOI: 10.1002/jps.10106.
- 85 K. Vermani, S. Garg and L. J. Zaneveld, Assemblies for in vitro measurement of bioadhesive strength and retention characteristics in simulated vaginal environment, *Drug Dev. Ind. Pharm.*, 2002, 28, 1133–1146, DOI: 10.1081/ddc-120014580.
- 86 P. V. Pople and K. K. Singh, Development and evaluation of topical formulation containing solid lipid nanoparticles of vitamin A, *AAPS PharmSciTech*, 2006, 7, 91, DOI: 10.1208/pt070491.
- 87 H. Chen, X. Chang, D. Du, J. Li, H. Xu and X. Yang, Microemulsion-based hydrogel formulation of ibuprofen for topical delivery, *Int. J. Pharm.*, 2006, 315, 52–58, DOI: 10.1016/j.ijpharm.2006.02.015.
- 88 S. Corveleyn and J. P. Remon, Stability of freeze-dried tablets at different relative humidities, *Drug Dev. Ind. Pharm.*, 1999, 25, 1005–1013, DOI: 10.1081/ddc-100102263.
- 89 N. Patel, B. L. Craddock, J. N. Staniforth, M. Tobyn and M. Welham, Spray-dried insulin particles retain biological activity in rapid in-vitro assay, *J. Pharm. Pharmacol.*, 2001, 53, 1415–1418, DOI: 10.1211/0022357011777774.
- 90 P. C. Reddy, K. S. C. Chaitanya and Y. M. Rao, A review on bioadhesive buccal drug delivery systems: current status of formulation and evaluation methods, *Daru*, 2011, 19, 385–403.
- 91 A. H. Shojaei, R. K. Chang and X. Guo, Systemic drug delivery via the buccal mucosal route, *Pharm. Technol.*, 2001, 25, 70–81.
- 92 R. Abu-Huwajj, R. M. Obaidat, K. Sweidan and Y. Al-Hiari, Formulation and in vitro evaluation of xanthan gum or carbopol 934-based mucoadhesive patches, loaded with nicotine, *AAPS PharmSciTech*, 2011, 12, 21–27, DOI: 10.1208/s12249-010-9534-5.
- 93 M. Jyostna, B. Reddy, E. Mohanambal, S. Narendiran, M. Murugan and M. Nishanthi, Formulation and in vitro evaluation of buccal patches of Desloratidine, *Int. J. Novel Trends Pharm. Sci.*, 2012, 2, 158–165.
- 94 R. R. Shiledar, A. A. Tagalpallearwar and C. R. Kokare, Formulation and in vitro evaluation of xanthan gum-based bilayeredmucoadhesive buccal patches of zolmitriptan, *Carbohydr. Polym.*, 2014, 101, 1234–1242, DOI: 10.1016/j.carbpol.2013.10.072.
- 95 M. Isaac and C. Holvey, Transdermal patches: the emerging mode of drug delivery system in psychiatry, *Ther. Adv. Psychopharmacol.*, 2012, 2, 255–263, DOI: 10.1177/2045125312458311.
- 96 N. Rajesh, S. D. V. Gowda and C. N. Somashekar, Formulation and evaluation of biopolymer based transdermal drug delivery, *Int. J. Pharm. Pharm. Sci.*, 2010, 2, 142–147.
- 97 A. R. Dezfuli, A. S. Aravindram, M. Manjunath, N. S. Ganesh and T. Shailesh, Development and evaluation of transdermal films loaded with antihypertensive drug, *Int. J. Pharm. Biol. Sci.*, 2012, 3, 559–569.
- 98 K. Mounika, B. V. Reddy and K. N. Reddy, Formulation and evaluation of carvedilol transdermal patches with hydrophilic polymers, *World J. Pharm. Res.*, 2014, 3, 815–826.
- 99 A. Takeuchi, Y. Kamiryu, H. Yamada, M. Eto, K. Shibata, K. Haruna, S. Naito and Y. Yoshikai, Oral administration of xanthan gum enhances antitumor activity through Toll-like receptor 4, *Int. Immunopharmacol.*, 2009, 9, 1562–1567, DOI: 10.1016/j.intimp.2009.09.012.
- 100 X. Hu, K. Wang, M. Yu, P. He, H. Qiao, H. Zhang and Z. Wang, Characterization and antioxidant activity of a low-molecular-weight xanthan gum, *Biomolecules*, 2019, 9, 730, DOI: 10.3390/biom9110730.
- 101 K. Shimada, K. Fujikawa, K. Yahara and T. Nakamura, Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion, *J. Agric. Food Chem.*, 1992, 40, 945–948, DOI: 10.1021/jf00018a005.
- 102 C. Amico, T. Tornetta, C. Scifo and A. R. Blanco, Antioxidant effect of 0.2% xanthan gum in ocular surface corneal epithelial cells, *Curr. Eye Res.*, 2015, 40, 72–76, DOI: 10.3109/02713683.2014.914542.
- 103 P. A. Sandford, J. Baird and I. W. Cottrell, *Xanthan gum with improved dispersibility. Solution properties of polysaccharides*, American Chemical Society, Washington, DC, 1981, pp. 31–38.
- 104 L. Su, W. K. Ji, W. Z. Lan and X. Q. Dong, Chemical modification of xanthan gum to increase dissolution rate, *Carbohydr. Polym.*, 2003, 53, 497–499, DOI: 10.1016/s0144-8617(02)00287-4.
- 105 A. Becker, F. Katzen, A. Pühler and L. Ielpi, Xanthan gum biosynthesis and application: a biochemical/genetic perspective, *Appl. Microbiol. Biotechnol.*, 1998, 50, 145–152, DOI: 10.1007/s002530051269.
- 106 S. Rosalam and R. England, Review of xanthan gum production from unmodified starches by *Xanthomonas compestris* sp, *Enzyme Microb. Technol.*, 2006, 39, 197–207, DOI: 10.1016/j.enzymictec.2005.10.019.
- 107 B. D. M. Lopes, V. L. Lessa, B. M. Silva, M. A. D. S. C. Filho, E. Schnitzler and L. G. Lacerda, Xanthan gum: properties, production conditions, quality and economic perspective, *J. Food Nutr. Res.*, 2015, 54, 185–194.
- 108 I. S. Benny, G. Varadarajan and V. Ponnusami, Review on application of Xanthan gum in Drug delivery, *Int. J. PharmTech Res.*, 2004, 6, 1322–1326.
- 109 V. S. Verma, K. Sakure and H. R. Badwaik, Xanthan gum a versatile biopolymer: current status and future prospectus in hydro gel drug delivery, *Curr. Chem. Biol.*, 2017, 11, 10–20, DOI: 10.2174/2212796810666161110152815.



- 110 D. F. S. Petri, Xanthan gum: a versatile biopolymer for biomedical and technological applications, *J. Appl. Polym. Sci.*, 2015, 1–13, DOI: 10.1002/app.42035.
- 111 A. Kumar, K. M. Rao and S. S. Han, Application of xanthan gum as polysaccharide in tissue engineering: a review, *Carbohydr. Polym.*, 2018, **180**, 128–144, DOI: 10.1016/j.carbpol.2017.10.009.
- 112 H. R. Badwaik, T. K. Giri, K. T. Nakhate, P. Kashyap and D. K. Tripathi, Xanthan gum and its derivatives as a potential bio-polymeric carrier for drug delivery system, *Curr. Drug Delivery*, 2013, **10**, 587–600, DOI: 10.2174/1567201811310050010.
- 113 M. M. Yahoum, N. Moulai-Mostefa and D. L. Cerf, Synthesis, physicochemical, structural and rheological characterizations of carboxymethyl xanthan derivatives, *Carbohydr. Polym.*, 2016, **154**, 267–275, DOI: 10.1016/j.carbpol.2016.06.080.
- 114 L. Shuang, Z. Hong, F. Bo, L. Yongjun, Q. Xiaohui and Z. Wen, Carboxymethylhydroxypropylxanthan gum and its rheological properties, *Drilling and Completion Fluids*, 2017, **34**, 107–116, DOI: 10.3969/j.issn.1001-5620.2017.05.020.
- 115 Z. Hong, L. U. Yongjun, F. Bo, Q. Xiaohui, W. Liwei, L. Yuting, T. Meng and L. Kejing, Synthesis and performance evaluation of cationic xanthan gum, *Oilfield Chemistry*, 2017, **34**, 34–37, DOI: 10.19346/j.cnki.1000-4092.2017.01.008.
- 116 L. Shi, Y. Wei, N. Luo, T. Tan and H. Cao, The rheological and thickening properties of cationic xanthan gum, *J. Dispersion Sci. Technol.*, 2018, **39**, 55–61, DOI: 10.1080/01932691.2017.1293547.
- 117 L. Shuang, Z. Hong, F. Bo, L. Yongjun, Q. Xiaohui, Z. Wen and W. Liwei, Preparation and rheology evaluation of hydrophobic amphoteric xanthan gum, *Oilfield Chemistry*, 2018, **35**, 224–230, DOI: 10.19346/j.cnki.1000-4092.2018.02.007.
- 118 L. Chengcheng, F. Bo, L. Yongjun, Q. Xiaohui, Z. Wen and W. Liwei, Rheological properties of oleamidopropyl dimethylamine modified xanthan gum solution, *Oilfield Chemistry*, 2018, **35**, 628–633, DOI: 10.19346/j.cnki.1000-4092.2018.04.012.
- 119 I. C. Alupeii, M. Popa, M. Hamcerencu and M. J. M. Abadie, Superabsorbent hydrogels based on xanthan and poly(vinyl alcohol) 1. The study of the swelling properties, *Eur. Polym. J.*, 2002, **38**, 2313–2320, DOI: 10.1016/s0014-3057(02)00106-4.
- 120 N. Dhar, H. Setia, A. Thakur and R. K. Wanchoo, Poly (vinyl alcohol) and xanthan gum composite films for sustained release, *Res. Rev. Polym.*, 2012, **3**, 121–126.
- 121 H. Zhang, B. Fang, Y. Lu, X. Qiu, H. Jin, Y. Liu, L. Wang, M. Tian and K. Li, Rheological properties of water-soluble crosslinked xanthan gum, *J. Dispersion Sci. Technol.*, 2017, **38**, 361–366, DOI: 10.1080/01932691.2016.1169929.
- 122 A.-M. Oprea, M.-T. Nistor, L. Profire, M. I. Popa, C. E. Lupusoru and C. Vasile, Evaluation of the controlled release ability of theophylline from xanthan/chondroitin sulfate hydrogels, *J. Biomater. Nanobiotechnol.*, 2013, **4**, 123–131, DOI: 10.4236/jbnt.2013.42017.
- 123 S. Maiti, S. Mukherjee and R. Datta, Core-shell nanobiomaterials for controlled oral delivery and pharmacodynamic activity of glibenclamide, *Int. J. Biol. Macromol.*, 2014, **70**, 20–25, DOI: 10.1016/j.ijbiomac.2014.06.031.
- 124 S. Maiti and S. Mukherjee, Controlled drug delivery attributes of co-polymer micelles and xanthan-O-carboxymethyl hydrogel particles, *Int. J. Biol. Macromol.*, 2014, **70**, 37–43, DOI: 10.1016/j.ijbiomac.2014.06.015.
- 125 H. Quan, Y. Hu, Z. Huang and D. Wenmeng, Preparation and property evaluation of a hydrophobically modified xanthan gum XG-C16, *J. Dispersion Sci. Technol.*, 2020, **41**, 656–666, DOI: 10.1080/01932691.2019.1610425.
- 126 X.-L. Qian, W.-H. Wu, P.-Z. Yu and J.-Q. Wang, Synthesis and aqueous solution viscosity of hydrophobically modified xanthan gum, *J. Beijing Inst. Technol.*, 2007, **16**, 346–351.
- 127 P. Bakshi, S. Sadhukhan and S. Maiti, Design of modified xanthan mini-matrices for monitoring oral discharge of highly soluble Soluplus®-glibenclamide dispersion, *Mater. Sci. Eng., C*, 2015, **54**, 169–175, DOI: 10.1016/j.msec.2015.05.014.
- 128 M. Ahuja, A. Kumar and K. Singh, Synthesis, characterization and in vitro release behavior of carboxymethyl Xanthan, *Int. J. Biol. Macromol.*, 2012, **51**, 1086–1090, DOI: 10.1016/j.ijbiomac.2012.08.023.
- 129 M. Munir, M. Shahid, H. Munir, F. Anjum, S. Javaid and E.-G. Ahmed, Xanthan gum biochemical profiling, antioxidant, antibacterial, biofilm inhibition and mutagenic potential, *Curr. Sci.*, 2017, **113**, 1904–1913.
- 130 M. Alle, R. G. Bhagavanth, T. H. Kim, S. H. Park, S.-H. Lee and J.-C. Kim, Doxorubicin-carboxymethyl xanthan gum capped gold nanoparticles: microwave synthesis, characterization, and anti-cancer activity, *Carbohydr. Polym.*, 2020, **229**, 115511, DOI: 10.1016/j.carbpol.2019.115511.
- 131 B. W. Soltovski and P. C. Ferrari, Swelling capability and drug release evaluation of chitosan and xanthan gum hydrogels for potential colonic drug delivery, *Lat. Am. J. Pharm.*, 2017, **36**, 1331–1337.
- 132 D. H. Hanna and G. R. Saad, Encapsulation of ciprofloxacin within modified xanthan gum-chitosan based hydrogel for drug delivery, *Bioorg. Chem.*, 2019, **84**, 115–124, DOI: 10.1016/j.bioorg.2018.11.036.
- 133 S. Ray, S. Maiti and B. Sa, Polyethyleneimine-treated xanthan beads for prolonged release of diltiazem: *in vitro* and *in vivo* evaluation, *Arch. Pharmacol. Res.*, 2010, **33**, 575–583, DOI: 10.1007/s12272-010-0412-1.
- 134 K. M. Rao, A. Kumar and S. S. Han, Polysaccharide-based magnetically responsive polyelectrolyte hydrogels for tissue engineering applications, *J. Mater. Sci. Technol.*, 2018, **34**, 1371–1377, DOI: 10.1016/j.jmst.2017.10.003.
- 135 A. F. Eftaiha, N. Qinna, I. S. Rashid, M. M. Al Remawi, M. R. AlShami, T. A. Arafat and A. A. Badwan, Bioadhesive controlled metronidazole release matrix



- based on chitosan and xanthan gum, *Mar. Drugs*, 2010, **8**, 1716–1730, DOI: 10.3390/md8051716.
- 136 A. M. Rajesh and K. M. Papat, Taste masking of ofloxacin and formation of interpenetrating polymer network beads for sustained release, *J. Pharm. Anal.*, 2017, **7**, 244–251, DOI: 10.1016/j.jpha.2016.11.001.
- 137 T. K. Giri, A. Thakur and D. K. Tripathi, Biodegradable hydrogel bead of casein and modified xanthan gum for controlled delivery of theophylline, *Curr. Drug Ther.*, 2016, **11**, 150–162, DOI: 10.2174/1574885511666160830123807.
- 138 T. K. Giri, C. Choudhary, A. Alexander, Ajazuddin, H. Badwaik, M. Tripathy and D. K. Tripathi, Sustained release of diltiazem hydrochloride from cross-linked biodegradable IPN hydrogel beads of pectin and modified xanthan gum, *Indian J. Pharm. Sci.*, 2013, **75**, 619–627.
- 139 S. Maity and B. Sa, Compression-coated tablet for colon targeting: impact of coating and core materials on drug release, *AAPS PharmSciTech*, 2016, **17**, 504–515, DOI: 10.1208/s12249-015-0359-0.
- 140 S. Karan, R. Pal, B. Ruhidas, S. Banerjee and T. K. Chatterjee, Comparative pharmacokinetic study and quantification of ibuprofen released from interpenetrating polymer network beads of sodium carboxymethyl xanthan and sodium alginate, *Indian J. Pharm. Educ. Res.*, 2016, **50**, 442–450, DOI: 10.5530/ijper.50.3.18.
- 141 S. Mitra, S. Maity and B. Sa, Effect of different cross-linking methods and processing parameter on drug release from hydrogel beads, *Int. J. Biol. Macromol.*, 2015, **74**, 489–497, DOI: 10.1016/j.ijbiomac.2014.12.008.
- 142 S. Maity and B. Sa, Ca-carboxymethyl xanthan gum mini-matrices: swelling, erosion and their impact on drug release mechanism, *Int. J. Biol. Macromol.*, 2014, **68**, 78–85, DOI: 10.1016/j.ijbiomac.2014.04.036.
- 143 S. S. Bhattacharya, S. Shukla, S. Banerjee, P. Chowdhury, P. Chakraborty and A. Ghosh, Tailored IPN hydrogel bead of sodium carboxymethyl cellulose and sodium carboxymethyl xanthan gum for controlled delivery of diclofenac sodium, *Polym.-Plast. Technol. Eng.*, 2013, **52**, 795–805, DOI: 10.1080/03602559.2013.763361.
- 144 R. Ray, S. Maity, S. Mandal, T. K. Chatterjee and B. Sa, Studies on the release of ibuprofen from al^{3+} ion cross-linked homopolymeric and interpenetrating network hydrogel beads of carboxymethyl xanthan and sodium alginate, *Adv. Polym. Technol.*, 2011, **30**, 1–11, DOI: 10.1002/adv.20199.
- 145 R. Ray, R. Pal, S. Karan, B. Sa and T. K. Chatterjee, Evaluation of pharmacological activities of ibuprofen loaded interpenetrating polymer network (IPN) beads from sodium carboxymethyl xanthan and sodium alginate on rats, *Pharmacologyonline*, 2010, **2**, 719–736.
- 146 S. Maiti, S. Ray and B. Sa, Controlled delivery of bovine serum albumin from carboxymethyl xanthan microparticles, *Pharm. Dev. Technol.*, 2009, **14**, 165–172, DOI: 10.1080/10837450802498878.
- 147 S. Maiti, S. Ray and B. Sa, Effect of formulation variables on entrapment efficiency and release characteristics of bovine serum albumin from carboxymethyl xanthan microparticles, *Polym. Adv. Technol.*, 2008, **19**, 922–927, DOI: 10.1002/pat.1061.
- 148 S. Ray, S. Maiti and B. Sa, Preliminary investigation on the development of diltiazem resin complex loaded carboxymethyl xanthan beads, *AAPS PharmSciTech*, 2008, **9**, 295–301, DOI: 10.1208/s12249-007-9012-x.
- 149 S. Maiti, S. Ray, B. Mandal, S. Sarkar and B. Sa, Carboxymethyl xanthan microparticles as a carrier for protein delivery, *J. Microencapsulation*, 2007, **24**, 743–756, DOI: 10.1080/02652040701647300.
- 150 I. Fernandez-Piñeiro, J. Alvarez-Trabado, J. Márquez, I. Badiola and A. Sanchez, Xanthan gum-functionalised span nanoparticles for gene targeting to endothelial cells, *Colloids Surf., B*, 2018, **170**, 411–420, DOI: 10.1016/j.colsurfb.2018.06.048.
- 151 M. Kang, O. Oderinde, S. Liu, Q. Huang, W. Ma, F. Yao and G. Fu, Characterization of Xanthan gum-based hydrogel with Fe^{3+} ions coordination and its reversible sol-gel conversion, *Carbohydr. Polym.*, 2019, **203**, 139–147, DOI: 10.1016/j.carbpol.2018.09.044.
- 152 T. Kundu, K. Mukherjee and B. Sa, Hydrogel beads composed of sodium carboxymethyl xanthan and sodium carboxymethyl cellulose for controlled release of aceclofenac: effect of formulation variables, *Res. J. Pharm. Technol.*, 2012, **5**, 103–113.
- 153 N. Kulkarni, P. Wakte and J. Naik, Development of floating chitosan-xanthan beads for oral controlled release of glipizide, *Int. J. Pharm. Invest.*, 2015, **5**, 73–80, DOI: 10.4103/2230-973x.153381.
- 154 X.-L. Qian, J.-Z. Su, W.-H. Wu and C.-M. Niu, Aqueous solution viscosity properties of hydrophobically modified xanthan gum HMXG-C8, *Oilfield Chemistry*, 2007, **24**, 154–157.
- 155 E. P. Pinto, L. Furlan and C. T. Vendruscolo, Chemical deacetylation natural xanthan (Jungbunzlauer®), *Polímeros*, 2011, **21**, 47–52, DOI: 10.1590/s0104-14282011005000005.
- 156 X. Wang, H. Xin, Y. Zhu, W. Chen, E. Tang, J. Zhang and Y. Tan, Synthesis and characterization of modified xanthan gum using poly(maleic anhydride/1-octadecene), *Colloid Polym. Sci.*, 2016, **294**, 1333–1341, DOI: 10.1007/s00396-016-3898-3.
- 157 Y. Tao, R. Zhang, W. Xu, Z. Bai, Y. Zhou, S. Zhao, Y. Xu and D. Q. Yu, Rheological behavior and microstructure of release-controlled hydrogels based on xanthan gum crosslinked with sodium trimetaphosphate, *Food Hydrocolloids*, 2016, **52**, 923–933, DOI: 10.1016/j.foodhyd.2015.09.006.
- 158 A. Shalviri, Q. Liu, M. J. Abdekhodaie and X. Wu, Novel modified starch-xanthan gum hydrogels for controlled drug delivery: synthesis and characterization, *Carbohydr. Polym.*, 2010, **79**, 898–907, DOI: 10.1016/j.carbpol.2009.10.016.
- 159 R. Zhang, Y. Tao, W. Xu, S. Xiao, S. Du, Y. Zhou and A. Hasan, Rheological and controlled release properties of hydrogels based on mushroom hyperbranched



- polysaccharide and xanthan gum, *Int. J. Biol. Macromol.*, 2018, **120**, 2399–2409, DOI: 10.1016/j.ijbiomac.2018.09.008.
- 160 A. Bejenariu, M. Popa, V. Dulong, L. Picton and D. L. Cerf, Trisodium, trimetaphosphate crosslinked xanthan networks: synthesis, swelling, loading and releasing behavior, *Polym. Bull.*, 2009, **62**, 525–538, DOI: 10.1007/s00289-008-0033-8.
- 161 A. Bejenariu, M. Popa, L. Picton and D. Le Cerf, Synthesis of xanthan based hydrogels. Influence of the synthesis parameters on hydrogels behavior, *Rev. Roum. Chim.*, 2009, **54**, 565–569.
- 162 J. Lam, E. C. Clark, E. L. S. Fong, E. J. Lee, S. Lu, Y. Tabata and A. G. Mikos, Evaluation of cell-laden polyelectrolyte hydrogels incorporating poly(L-lysine) for applications in cartilage tissue engineering, *Biomaterials*, 2016, **83**, 332–346, DOI: 10.1016/j.biomaterials.2016.01.020.
- 163 G. Leone, M. Consumi, S. Lamponi, C. Bonechi, G. Tamasi, A. Donati, C. Rossi and A. Magnani, Hybrid PVA-xanthan gum hydrogels as nucleus pulposus substitutes, *Int. J. Polym. Mater. Polym. Biomater.*, 2019, **68**, 681–690, DOI: 10.1080/00914037.2018.1482468.
- 164 P. Veiga-Santos, L. M. Oliveira, M. P. Cereda, A. J. Alves and A. R. P. Scamparini, Mechanical properties, hydrophilicity and water activity of starch–gum films: effect of additives and deacetylated xanthan gum, *Food Hydrocolloids*, 2005, **19**, 341–349, DOI: 10.1016/j.foodhyd.2004.07.006.
- 165 V. B. Bueno and D. F. S. Petri, Xanthan hydrogel films: molecular conformation, charge density and protein carriers, *Carbohydr. Polym.*, 2014, **101**, 897–904, DOI: 10.1016/j.carbpol.2013.10.039.
- 166 J. Huang, J. Ren, G. Chen, Y. Deng, G. Wang and X. Wu, Evaluation of the xanthan-based film incorporated with silver nanoparticles for potential application in the nonhealing infectious wound, *J. Nanomater.*, 2017, **2017**, 6802397, DOI: 10.1155/2017/6802397.
- 167 D. Bilanovic, J. Starosvetsky and R. H. Armon, Cross-linking xanthan and other compounds with glycerol, *Food Hydrocolloids*, 2015, **44**, 129–135, DOI: 10.1016/j.foodhyd.2014.09.024.
- 168 M. Hamcerencu, J. Desbrieres, M. Popa, A. Khoukh and G. Riess, New unsaturated derivatives of Xanthan gum: synthesis and characterization, *Polymer*, 2007, **48**, 1921–1929, DOI: 10.1016/j.polymer.2007.01.048.
- 169 A. C. Mendes, E. T. Baran, C. Nunes, M. A. Coimbra, H. S. Azevedo and R. L. Reis, Palmitoylation of xanthan polysaccharide for self-assembly microcapsule formation and encapsulation of cells in physiological conditions, *Soft Matter*, 2011, **7**, 9647–9658, DOI: 10.1039/c1sm05594a.
- 170 M. Bhatia, M. Ahuja and H. Mehta, Thiol derivatization of xanthan gum and its evaluation as amucoadhesive polymer, *Carbohydr. Polym.*, 2015, **131**, 119–124, DOI: 10.1016/j.carbpol.2015.05.049.
- 171 A. Skender, A. Hadj-Ziane-Zafour and E. Flahaut, Chemical functionalization of Xanthan gum for the dispersion of double-walled carbon nanotubes in water, *Carbon*, 2013, **62**, 149–156, DOI: 10.1016/j.carbon.2013.06.006.
- 172 B. Wang, Y. Han, Q. Lin, H. Liub, C. Shen, K. Nan and H. Chen, In vitro and in vivo evaluation of xanthan gumsuccinic anhydride hydrogels for ionic strengthsensitive release of antibacterial agents, *J. Mater. Chem. B*, 2016, **4**, 1853–1861, DOI: 10.1039/c5tb02046h.
- 173 C. G. Gomez, M. Rinaudo and M. A. Villar, Oxidation of sodium alginate and characterization of the oxidized derivatives, *Carbohydr. Polym.*, 2007, **67**, 296–304, DOI: 10.1016/j.carbpol.2006.05.025.
- 174 J. Guo, L. Ge, X. Li, C. Mu and D. Li, Periodate oxidation of xanthan gum and its crosslinking effects on gelatin-based edible films, *Food Hydrocolloids*, 2014, **39**, 243–250, DOI: 10.1016/j.foodhyd.2014.01.026.
- 175 D. Paiva, C. Gonçalves, I. Vale, M. M. S. M. Bastos and F. D. Magalhães, Oxidized xanthan gum and chitosan as natural adhesives for cork, *Polymers*, 2016, **8**, 259–271, DOI: 10.3390/polym8070259.
- 176 Y.-H. Ma, J. Yang, B. Li, Y.-W. Jiang, X. Lu and Z. Chen, Biodegradable and injectable polymer–liposome hydrogel: a promising cell carrier, *Polym. Chem.*, 2016, **7**, 2037–2044, DOI: 10.1039/c5py01773d.
- 177 N. Li, W. Chen, G. Chen and J. Tian, Rapid shape memory TEMPO-oxidized cellulose nanofibers/polyacrylamide/gelatin hydrogels with enhanced mechanical strength, *Carbohydr. Polym.*, 2017, **171**, 77–84, DOI: 10.1016/j.carbpol.2017.04.035.
- 178 G. M. Salazar, N. C. Sanoh and D. P. Penaloza Jr, Synthesis and characterization of a novel polysaccharide-based self-healing hydrogel, *Kimika*, 2018, **29**, 44–48, DOI: 10.26534/kimika.v29i2.44-48.
- 179 J. Huang, Y. Deng, J. Ren, G. Chen, G. Wang, F. Wang and X. Wu, Novel in situ forming hydrogel based on xanthan and chitosan re-gelifying in liquids for local drug delivery, *Carbohydr. Polym.*, 2018, **186**, 54–63, DOI: 10.1016/j.carbpol.2018.01.025.
- 180 X. Xiong, M. Li, J. Xie, Q. Jin, B. Xue and T. Sun, Antioxidant activity of xanthan oligosaccharides prepared by different degradation methods, *Carbohydr. Polymer*, 2013, **92**, 1166–1171, DOI: 10.1016/j.carbpol.2012.10.069.
- 181 L. Ge, Y. Xu, X. Li, L. Yuan, H. Tan, D. Li and C. Mu, Fabrication of antibacterial collagen-based composite wound dressing, *ACS Sustainable Chem. Eng.*, 2018, **6**, 9153–9166, DOI: 10.1021/acssuschemeng.8b01482.
- 182 A. Roy, S. Comesse, M. Grisel, N. Hucher, Z. Souguir and F. Renou, Hydrophobically modified xanthan: an amphiphilic but not associative polymer, *Biomacromolecules*, 2014, **15**, 1160–1170, DOI: 10.1021/bm4017034.
- 183 F. Céline, A. N. Roy, E. Dé, S. Comesse, M. Grisel and F. Renou, Chemical modification of xanthan in the ordered and disordered states: an open route for tuning the physico-chemical properties, *carbohydr., Polymer*, 2017, **178**, 115–122, DOI: 10.1016/j.carbpol.2017.09.039.
- 184 C. Fantou, S. Comesse, F. Renou and M. Grisel, Hydrophobically modified xanthan: thickening and surface active agent for highly stable oil in water



- emulsions, *Carbohydr. Polym.*, 2019, **205**, 362–370, DOI: 10.1016/j.carbpol.2018.10.052.
- 185 C. Fantou, S. Comesse, F. Renou and M. Grisel, Impact of backbone stiffness and hydrophobic chain length of modified xanthan on oil in water emulsion stabilization, *Carbohydr. Polym.*, 2019, **216**, 352–359, DOI: 10.1016/j.carbpol.2019.03.079.
- 186 A. Bejenariu, M. Popa, D. Le Cerf and L. Picton, Stiffness xanthan hydrogels: synthesis, swelling characteristics and controlled release properties, *Polym. Bull.*, 2008, **61**, 631–641, DOI: 10.1007/s00289-008-0987-6.
- 187 F. Laffleur, M. Michalek, W. Suchaoin and M. Ijaz, Design and evaluation of buccal-adhesive system made of modified xanthan, *Ther. Delivery*, 2016, **7**, 423–429, DOI: 10.4155/tde-2016-0021.
- 188 A. Mato, J. Limeres, I. Tomás, M. Muñoz, C. Abuín, J. F. Feijoo and P. Diz, Management of drooling in disabled patients with scopolamine patches, *Br. J. Clin. Pharmacol.*, 2010, **69**, 684–688, DOI: 10.1111/j.1365-2125.2010.03659.x.
- 189 F. Laffleur and M. Michalek, Modified xanthan gum for buccal delivery—A promising approach intreating sialorrhea, *Int. J. Biol. Macromol.*, 2017, **102**, 1250–1256, DOI: 10.1016/j.ijbiomac.2017.04.123.
- 190 C. Menzel, M. Jelkmann, F. Laffleur and A. Bernkop-Schnürch, Nasal drug delivery: design of a novel mucoadhesive and in situ gelling polymer, *Int. J. Pharm.*, 2017, **517**, 196–202, DOI: 10.1016/j.ijpharm.2016.11.055.
- 191 A. C. Mendes, E. T. Baran, R. L. Reis and H. S. Azevedo, Fabrication of phospholipid–xanthan microcapsules by combining microfluidics with self-assembly, *Acta Biomater.*, 2013, **9**, 6675–6685, DOI: 10.1016/j.actbio.2013.01.035.
- 192 M. M. Hashemi, A. Mahmoud and M. Moosavinasab, Preparation of and studies on the functional properties and bactericidal activity of the lysozyme-xanthan gum conjugate, *LWT—Food Sci. Technol.*, 2014, **57**, 594–602, DOI: 10.1016/j.lwt.2014.01.040.
- 193 S. Ray, S. Banerjee, S. Maiti, B. Laha, S. Barik, B. Sa and U. K. Bhattacharyya, Novel interpenetrating network microspheres of xanthan gum–poly(vinyl alcohol) for the delivery of diclofenac sodium to the intestine—in vitro and in vivo evaluation, *Drug Delivery*, 2010, **17**, 508–519, DOI: 10.3109/10717544.2010.483256.
- 194 N. M. Sereno, S. E. Hill and J. R. Mitchell, Impact of the extrusion process on xanthan gum behavior, *Carbohydr. Res.*, 2007, **342**, 1333–1342, DOI: 10.1016/j.carres.2007.03.023.
- 195 T. J. Foster and J. R. Mitchell, Physical Modification of Xanthan gum, in *Gums and Stabilisers for the Food Industry*, ed. P. A. Williams and G. O. Phillips, RSC Publishing, 2012, pp. 77–88. DOI: 10.1039/9781849734554.
- 196 N. M. Eren, P. H. S. Santos and O. Campanella, Mechanically modified xanthan gum: rheology and polydispersity aspects, *Carbohydr. Polym.*, 2015, **134**, 475–484, DOI: 10.1016/j.carbpol.2015.07.092.
- 197 M. A. Zirnsak, D. V. Boger and V. Tirtaatmadja, Steady shear and dynamic rheological properties of xanthan gum solutions in viscous solvents, *J. Rheol.*, 1999, **43**, 627–650, DOI: 10.1122/1.551007.
- 198 Q. Zhang, X. M. Hu, M. Y. Wu, M. M. Wang, Y. Y. Zhao and T. T. Li, Synthesis and performance characterization of poly(vinyl alcohol)-xanthan gum composite hydrogel, *React. Funct. Polym.*, 2019, **136**, 34–43, DOI: 10.1016/j.reactfunctpolym.2019.01.002.
- 199 Y. Shimizu, T. Tanabe, H. Yoshida, M. Kasuya, T. Matsunaga, Y. Haga, K. Kurihara and M. Ohta, Viscosity measurement of xanthan–poly(vinyl alcohol) mixture and its effect on the mechanical properties of the hydrogel for 3D modeling, *Sci. Rep.*, 2018, **8**, 16538, DOI: 10.1038/s41598-018-34986-4.
- 200 T. Bhunia, D. Chattopadhyay and A. Bandyopadhyay, Gel viscosity influenced by nanosilica phase morphology in high and low molecular weights PVA-ex-situ silica hybrids, *J. Sol-Gel Sci. Technol.*, 2011, **59**, 260–268, DOI: 10.1007/s10971-011-2494-8.
- 201 T. Bhunia, A. Giri, T. Nasim, D. Chattopadhyay and A. Bandyopadhyay, Uniquely different PVA-xanthan gum irradiated membranes as transdermal diltiazem delivery device, *Carbohydr. Polym.*, 2013, **95**, 252–261, DOI: 10.1007/s10971-011-2494-8.
- 202 J. S. Disha, M. H. A. Begum, M. M. A. K. Shawan, N. Khatun, S. Ahmed, M. S. Islam, M. R. Karim, M. R. L. Islam, M. M. Hossain and M. A. Hasan, Preparation and characterization of xanthan gum-based biodegradable polysaccharide hydrogels, *Res. J. Mater. Sci.*, 2016, **4**, 13–18.
- 203 M. W. Sabaa, M. H. Abu Elella, D. H. Hanna and R. R. Mohamed, Encapsulation of bovine serum albumin within novel xanthan gum based hydrogel for protein delivery, *Mater. Sci. Eng., C*, 2019, **94**, 1044–1055, DOI: 10.1016/j.msec.2018.10.040.
- 204 M. Abate, D. Pulcini, A. D. Iorio and C. Schiavone, Viscosupplementation with intra-articular hyaluronic acid for treatment of osteoarthritis in the elderly, *Curr. Pharm. Des.*, 2010, **16**, 631–640, DOI: 10.2174/138161210790883859.
- 205 N. Kumahashi, K. Naitou, H. Nishi, K. Oae, Y. Watanabe, S. Kuwata, M. Ochi, M. Ikeda and Y. Uchio, Correlation of changes in pain intensity with synovial fluid adenosine triphosphate levels after treatment of patients with osteoarthritis of the knee with high-molecular-weight hyaluronic acid, *Knee*, 2011, **18**, 160–164, DOI: 10.1016/j.knee.2010.04.013.
- 206 R. Barbucci, S. Lamponi, A. Borzacchiello, L. Ambrosio, M. Fini, P. Torricelli and R. Giardino, Hyaluronic acid hydrogel in the treatment of osteoarthritis, *Biomaterials*, 2002, **23**, 4503–4513, DOI: 10.1016/s0142-9612(02)00194-1.
- 207 A. Garcia-Abuín, D. Gómez-Díaz, J. M. Navaza, L. Regueiro and I. Vidal-Tato, Viscosimetric behaviour of hyaluronic acid in different aqueous solutions, *Carbohydr. Polym.*, 2011, **85**, 500–505, DOI: 10.1016/j.carbpol.2011.02.028.
- 208 A. F. Dário, L. M. A. Hortêncio, M. R. Sierakowski, J. C. Queiroz Neto and D. F. S. Petri, The effect of calcium



- salts on the viscosity and adsorption behavior of xanthan, *Carbohydr. Polym.*, 2011, **84**, 669–676, DOI: 10.1016/j.carbpol.2010.12.047.
- 209 H. Nankai, W. Hashimoto, H. Miki, S. Kawai and K. Murata, Microbial system for polysaccharide depolymerization: enzymatic route for xanthan depolymerization by *Bacillus* sp. strain GL1, *Appl. Environ. Microbiol.*, 1999, **65**, 2520–2526.
- 210 G. Han, G. Wang, X. Zhu, H. Shao, F. Liu, P. Yang, Y. Ying, F. Wang and P. Ling, Preparation of xanthan gum injection and its protective effect on articular cartilage in the development of osteoarthritis, *Carbohydr. Polym.*, 2012, **87**, 1837–1842, DOI: 10.1016/j.carbpol.2011.10.016.
- 211 A. I. Raafat, N. M. El-Sawy, N. A. Badawy, E. A. Mousa and A. M. Mohamed, Radiation fabrication of xanthan-based wound dressing hydrogels embedded ZnO nanoparticles: in vitro evaluation, *Int. J. Biol. Macromol.*, 2018, **118**, 1892–1902, DOI: 10.1016/j.ijbiomac.2018.07.031.
- 212 Z. Zhao, T. Geng and H. Miao, Interface modification and property of dispersible xanthan gum, *Oilfield Chemistry*, 2016, **33**, 1–4.
- 213 V. Sharma and K. Pathak, Modified xanthan gum as hydrophilic disintegrating excipient for rapidly disintegrating tablets of roxithromycin, *Indian J. Pharm. Educ. Res.*, 2013, **47**, 79–87.

