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Lignin-derivable, thermoplastic, non-isocyanate polyurethanes with increased hydrogen-bonding content and toughness vs. petroleum-derived analogues†

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The functionality inherent in lignin-derivable bisguaiacols/bissyringols can improve the processability and performance of the resulting polymers. Herein, non-isocyanate polyurethanes (NIPUs) were synthesized from bisguaiacols/bissyringols with varying degrees of methoxy substitution and differing bridging groups. Notably, the presence of increasing numbers of methoxy groups (0, 2, and 4) in bisphenol F (BPF)-, bisguaiacol F (BGF)-, and bissyringol F (BSF)-NIPUs led to higher percentages of hydrogen-bonded –OH/–NH groups (*i.e.*, ~65%, ~85%, ~95%, respectively). Increased hydrogen bonding between chains improved the elongation-at-break (ϵ_{break}) and toughness of lignin-derivable NIPUs over their petroleum counterparts without a reduction in Young's moduli and tensile strengths. For example, BSF-NIPU exhibited the highest ϵ_{break} ~210% and toughness ~62 MJ m⁻³, followed by BGF-NIPU (ϵ_{break} ~185% and toughness ~58 MJ m⁻³), and then BPF-NIPU (ϵ_{break} ~140% and toughness ~42 MJ m⁻³). Similar trends were found in the dimethyl-substituted analogues, particularly for the bisphenol A-NIPU and bisguaiacol A-NIPU. Importantly, the melt rheology of the lignin-derivable NIPUs was comparable to that of the petroleum-derived analogues, with a slightly lower viscosity (*i.e.*, improved melt flow) for the bio-derivable NIPUs. These findings suggested that the added functionalities (methoxy groups) derived from lignin precursors improved thermomechanical stability while also offering increased processability. Altogether, the structure–property–processing relationships described in this work can help facilitate the development of sustainable, performance-advantaged polymers.

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1. Introduction

Thermoplastic polyurethanes (PUs) are highly desirable materials for numerous applications (*e.g.*, automotive parts, packaging films, medical materials) because they are readily processable and offer highly tunable chemical structures (*e.g.*, compositions of soft and hard segments) that yield a range of sought-after properties (*e.g.*, toughness, modulus, thermal stability, adhesion).^{1–4} Conventional thermoplastic PUs are synthesized by reacting diisocyanates with diols (and chain extenders in the case of segmented PUs),^{4,5}

however, this route has significant health and environmental challenges. For example, isocyanates are synthesized using highly toxic phosgene,^{6–8} and the most common aromatic isocyanates, such as methylene diphenyl diisocyanate (MDI) and toluene diisocyanate (TDI), have been classified as carcinogenic, mutagenic, and reprotoxic compounds.^{6,7} Long-term exposure to isocyanates also can cause respiratory disorders, and the European Union is considering restrictions to isocyanate usage.⁸

Non-isocyanate polyurethanes (NIPUs) are gaining attention in the research community as a means to address the concerns associated with traditional PUs,^{7–17} yet several synthetic routes for NIPUs reported in the literature use toxic precursors. For instance, NIPU syntheses have been reported that incorporate the rearrangement of acyl azides (*i.e.*, Curtius rearrangement)¹⁸ or employ hydroxamic azides (*i.e.*, Lossen rearrangement).¹⁹ Additionally, NIPUs have been synthesized through the polycondensation of polycarbamoyl chlorides and polyols.⁶ These compounds (acyl azides, hydroxamic azides, and polycarbamoyl chlorides) use phosgene or its derivatives as a key building block.

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Hence, an emerging pathway for NIPU synthesis is *via* the step-growth polyaddition of cyclic carbonates and diamines, which significantly reduces the use of toxic precursors.^{6,9–11,20–23} Moreover, CO₂ sequestration *via* the synthesis of cyclic carbonates may provide an environmental benefit.^{6,8}

Another consideration is that most precursors for PU/NIPU synthesis are derived primarily from petroleum-feedstocks, which are limited in quantity and unevenly distributed globally,²⁴ and there is significant opportunity for new building blocks with reduced toxicity and increased sustainability. Most bio-based/bio-derivable NIPU studies have focused on using aliphatic/cycloaliphatic building blocks derived from plant oils and fatty acids,⁷ but many polymers derived from such aliphatic/cycloaliphatic sources lack the robust structural features necessary to generate materials with properties competitive to those of similar polymers made from conventional aromatics (*e.g.*, TDI, MDI) in terms of glass transition temperatures [T_g s] and moduli.^{25,26} Petroleum-derived bisphenol A (BPA) has been used to make aromatic NIPUs,^{21,27} but BPA is a suspected endocrine disruptor, and the potential of BPA to leach out from bisphenol-derived plastics is implicated in numerous health hazards.^{28–30}

Lignin is an attractive feedstock for renewable polymer applications as it is the most abundant potential source of natural aromatics and can act as replacement feedstock for aromatic NIPUs.^{24–26,31–40} Bulk lignin is mainly comprised of *p*-hydroxyphenyl (H; no methoxy groups on the aromatic ring), guaiacyl (G; one methoxy group on the aromatic ring), and syringyl (S; two methoxy groups on the aromatic ring) units.^{31,34,41,42} The relative content of these units varies with the source of biomass; softwoods include a majority of G units, hardwoods contain a mixture of G and S units, and grasses contain a mixture of H, G, and S units.^{31,41,42} Unfortunately, materials made from bulk lignin can have inconsistent properties due to lignin's structural heterogeneity and limited reactivity.^{25,41} To overcome this hurdle, lignin can be broken down by catalytic, enzymatic, or pyrolytic pathways into smaller molecules with several inherent functional groups (*e.g.*, aliphatic/aromatic hydroxyl, methoxy, alkyl, allyl), and many of these functionalities can be used to develop renewable materials, such as polycarbonates, polyesters, epoxy resins, and PUs.^{25,31,34,41} Recently, renewable bisguaiacols/bissyringols have been reported as potential alternatives to commercial bisphenols because of their structural similarity. These renewable alternatives have methoxy groups on their aromatic rings^{30,43–46} that have the potential to mitigate the toxicity concerns associated with commercial aromatics (*e.g.*, estrogenic activity, genotoxicity, oxidative DNA damage).^{28,47,48}

Thermoplastic NIPUs have gained less attention in the literature in comparison to thermosetting NIPUs. The majority of research has been aimed at NIPU synthesis strategies, examining the role of reaction temperature, solvent, catalyst, *etc.*, on molar mass of the thermoplastic NIPU.^{6,7} Unique NIPU architectures (*i.e.*, segmented, phase-separated) also have been explored, in which the effect of different chain extenders and soft segments was investigated.^{49–51} Most studies in the lignin-derivable NIPU domain have focused on synthesizing NIPUs (especially thermosets) using bulk lignin or different

building blocks (limonene, vanillin, syringaresinol, *etc.*),^{7,12,52,53} and relatively few investigations have focused on understanding the rheological behavior of thermoplastic NIPUs.^{51,54} Importantly, the inherent pendent hydroxyl and methoxy groups in the lignin-derivable NIPUs offer additional hydrogen-bonding content not accessible in conventional PUs/NIPUs.⁴⁵ Together, a thorough understanding of these hydrogen-bonding functionalities (*i.e.*, the influence of methoxy moieties on lignin-aromatics with pendent hydroxyl groups) in the NIPU backbone, and the resulting impact of these groups on the thermal, mechanical, and rheological properties of bio-derivable thermoplastic NIPUs, remains under-explored.

Herein, NIPUs were synthesized from bisguaiacols/bissyringols with varying degrees of methoxy substitution and differing bridging groups (Scheme 1) to understand the effect of these structural features on thermal, mechanical, and rheological behavior. Bisphenol-based NIPUs with unsubstituted bridging carbons (bisphenol F [BPF]/bisguaiacol F [BGF]/bissyringol F [BSF]) and dimethyl-substituted bridging carbons (BPA/bisguaiacol A [BGA]/bissyringol A [BSA]) served as the library of materials for the investigation. First, the effect of key structural features (methoxy-group content and bridging-carbon substitutions) in the polymer repeat unit on the thermal (*i.e.*, T_g , degradation temperature [T_d]) and mechanical properties (*i.e.*, Young's modulus, tensile strength, toughness, elongation-at-break [ϵ_{break}]) was studied. Second, the impact of these monomer structures on the rheological response was explored to probe the melt behavior of these bio-derivable NIPUs. Importantly, the functionality of lignin-aromatics in NIPU (particularly methoxy groups) improved mechanical properties (particularly ϵ_{break} and toughness) in comparison to the petroleum-based counterparts without significant reduction in T_d , Young's modulus, and tensile strength. Additionally, the lignin-derivable NIPUs exhibited lower melt viscosities and had faster terminal relaxation times than the petroleum-derived NIPUs.

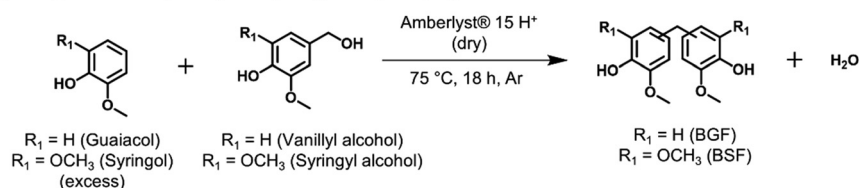
2. Experimental

2.1. Materials

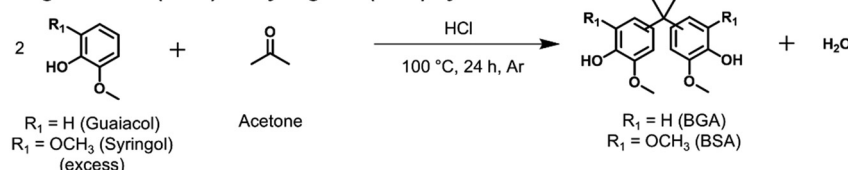
BPA diglycidyl ether (extent of oligomerization = ~ 0.02 , physical form = solid), guaiacol ($\geq 98\%$ food grade), syringol (99%), vanillyl alcohol ($\geq 98\%$), thioglycolic acid (98%), LiBr ($> 99\%$), LiCl ($> 99\%$), and Amberlyst[®] 15 hydrogen form (dry) were purchased from Sigma-Aldrich. DA-10 ($> 99\%$) was purchased from TCI. Syringyl alcohol (97%), deuterated dimethyl sulfoxide (DMSO- d_6 , $\geq 99.5\%$), *N,N*-dimethylacetamide (DMAc, 99.8+%, HPLC grade), DMSO (anhydrous, 99.7% extra dry), tetrabutylammonium bromide (TBAB, 99+%), epichlorohydrin (99%), and deuterated chloroform (CDCl₃, $\geq 99.75\%$) were purchased from acros organics. BPF diglycidyl ether (EPON[™] 862, extent of oligomerization = ~ 0.10 , physical form = viscous liquid), a product of Hexion, was purchased from Chemical Marketing Concepts Inc. Acetone ($\geq 99.5\%$), dichloromethane (DCM, $\geq 99.5\%$), *N,N*-dimethylformamide (DMF, $\geq 99.8\%$), HCl (36.5 to 38.0% [w/w]), NaOH ($\geq 97\%$), magnesium sulfate (anhydrous), sodium



(a) Bisguaiaicol F (BGF)/bissyringol F (BSF) synthesis



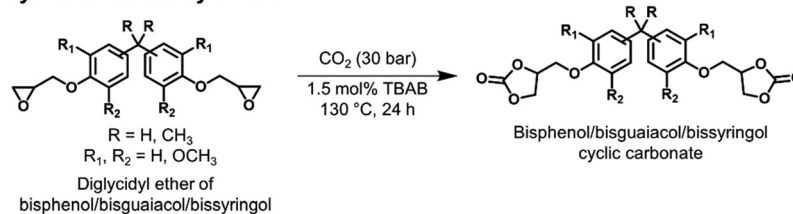
(b) Bisguaiaicol A (BGA)/bissyringol A (BSA) synthesis



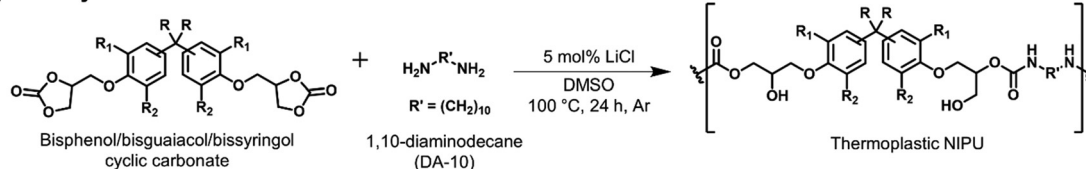
(c) Diglycidyl ether synthesis



(d) Cyclic carbonate synthesis



(e) NIPU synthesis



Scheme 1 Synthesis of (a) BGF/BSF using guaiacol/syringol and vanillyl/syringyl alcohol, (b) BGA/BSA using guaiacol/syringol and acetone, (c) diglycidyl ether of bisguaiaicol/bissyringol from respective bisguaiaicol/bissyringol and epichlorohydrin, (d) bisguaiaicol/bissyringol cyclic carbonate from respective diglycidyl ether and CO_2 , and (e) NIPU using respective bisguaiaicol/bissyringol cyclic carbonate and 1,10-diaminodecane (DA-10).

bicarbonate ($\geq 99.7\%$), NaCl ($\geq 99.5\%$), tetrahydrofuran (THF, $\geq 99\%$), hexanes (99%), ethyl acetate ($\geq 99.5\%$), and acetonitrile ($\geq 99.9\%$) were purchased from Thermo Fisher Scientific. Ar (grade 5), CO_2 (grade 5), and N_2 (grade 5) were purchased from Keen Compressed Gas. All chemicals were used as received except acetone, which was passed through a magnesium sulfate column before use.

2.2. Monomer synthesis and characterization

2.2.1. Synthesis of lignin-derivable bisphenols

BGF synthesis (Scheme 1a). BGF was synthesized as reported in the literature.⁴³ Briefly, guaiacol (40 g, 0.311 mol) and

vanillyl alcohol (12 g, 0.078 mol) were loaded into an oven-dried, single-neck, round-bottom flask (250 mL) equipped with a magnetic stir bar. The solution was heated to 70 °C in a silicone oil bath and sparged with Ar gas for 40 min. Then, under Ar flow, Amberlyst[®] 15 hydrogen form (dry) [3.6 g, 30 wt% relative to the vanillyl alcohol] was added to the reaction mixture, and the reaction was allowed to proceed for 18 h at 70 °C. Next, the reaction mixture was passively cooled to ~20 °C in air for ~1.5 h, dissolved in 15 mL of DCM, and the solid catalyst was separated from the crude mixture using a Buchner funnel. The liquid phase was then transferred to a separatory funnel and washed with deionized (DI) water



(3 times) and brine solution (2 times). The organic phase was collected, the solvent was removed under reduced pressure, and the product was purified by flash column chromatography (Biotage[®] Selekt Systems, Biotage[®] Sfär Silica columns – 60 μm , 100 g) with ethyl acetate (25 v/v%) and hexanes (75 v/v%). The product was dried at 50 $^{\circ}\text{C}$ under vacuum for 24 h, and the dried product was obtained as a white solid (yield \sim 50 mol%). Proton (^1H) nuclear magnetic resonance (^1H NMR) spectra (600 MHz, $\text{DMSO-}d_6$) δ 8.68 (s, 2H), 6.80–6.51 (m, 6H), 3.71 (d, $J = 4.7$ Hz, 8H). Fourier transform mass spectrometry (FTMS) (electrospray ionization [ESI]-FTMS, m/z): calculated for $\text{C}_{15}\text{H}_{16}\text{O}_4$ 260.1049; found 260.1041 [Fig. S1, ESI[†]].

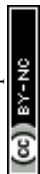
BSF synthesis (Scheme 1a). Syringol (40.1 g, 0.26 mol) and syringyl alcohol (12 g, 0.065 mol) were loaded into an oven-dried, single-neck, round-bottom flask (250 mL) equipped with a magnetic stir bar. The solution was heated to 85 $^{\circ}\text{C}$ in a silicone oil bath and sparged with Ar gas for 40 min. Then, Amberlyst[®] 15 hydrogen form (dry) [3.6 g, 30 wt% relative to the syringyl alcohol] was added to the reaction mixture, and the reaction was allowed to proceed for 18 h. After the designated time, the reaction mixture was allowed to passively cool to \sim 20 $^{\circ}\text{C}$ in air for \sim 1.5 h. The reaction mixture was subsequently dissolved in 15 mL of DCM, and the solid catalyst was separated from the crude mixture using a Buchner funnel. Like BGF purification, the liquid phase was washed with DI water (3 times) and brine solution (2 times) in a separatory funnel. The organic phase was collected, and the excess solvent was removed under reduced pressure. Further, the flash column chromatography (Biotage[®] Selekt Systems, Biotage[®] Sfär Silica columns – 60 μm , 100 g) with ethyl acetate (25 v/v%) and hexanes (75 v/v%) was used to purify BSF. The product was dried at 50 $^{\circ}\text{C}$ under vacuum for 24 h, and the dried product was obtained as a white solid (yield \sim 45 mol%). ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 8.50 (s, 1H), 8.04 (s, 1H), 6.69–6.28 (m, 4H), 3.83–3.54 (m, 14H). FTMS (ESI, m/z): calculated for $\text{C}_{17}\text{H}_{20}\text{O}_6$ 320.1260; found 320.1321 [Fig. S2, ESI[†]].

BGA synthesis (Scheme 1b). Guaiacol (40 g, 0.32 mol) and acetone (3.4 mL, 0.046 mol) were charged in an oven-dried, single-neck, round-bottom flask (250 mL) equipped with a condenser and magnetic stirrer. The reaction was conducted in a large excess of guaiacol to minimize the formation of by-products from the self-condensation of acetone. Next, 5 mL of concentrated HCl (6 g, 15 wt% relative to guaiacol) was slowly added to the above reactants, followed by thioglycolic acid (0.2 vol% with respect to the catalyst) as a promoter. The reaction mixture was sparged with Ar gas for 10 min, and the mixture was heated to 100 $^{\circ}\text{C}$ in a silicone oil bath for 24 h. After that, the reaction mixture was allowed to passively cool to \sim 20 $^{\circ}\text{C}$ in air for \sim 1.5 h. The crude mixture was dissolved in 15 mL of DCM, and the organic phase was washed with sodium bicarbonate until the aqueous layer reached neutral pH, then washed with DI water (3 times) and brine solution (2 times). The organic phase was collected, the solvent was removed under reduced pressure, and the product was purified by flash

column chromatography (Biotage[®] Selekt Systems, Biotage[®] Sfär Silica columns – 60 μm , 100 g) with ethyl acetate (25% v/v) and hexanes (75% v/v). The product was dried at 50 $^{\circ}\text{C}$ under vacuum for 24 h, and the dried product was obtained as a white solid (yield \sim 40 mol%). ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 8.69 (d, $J = 5.7$ Hz, 2H), 6.88–6.46 (m, 6H), 3.80–3.51 (m, 6H), 1.54 (d, $J = 18.2$ Hz, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{17}\text{H}_{20}\text{O}_4$ 288.1362; found 288.1354 [Fig. S3, ESI[†]].

BSA synthesis (Scheme 1b). Syringol (40 g, 0.26 mol) was weighed into an oven-dried, single-neck, round-bottom flask (250 mL) equipped with a condenser and magnetic stirrer. To melt syringol, the round-bottom flask was heated to 85 $^{\circ}\text{C}$ in a silicone oil bath under Ar flow for 30 min. Acetone (2.75 mL, 0.037 mol) and HCl (5 mL, \sim 15 wt% relative to syringol) were added dropwise to the reaction flask, and the mixture was heated to 100 $^{\circ}\text{C}$ for 24 h. After 24 h, the reaction mixture was allowed to passively cool to \sim 20 $^{\circ}\text{C}$ in air for \sim 1.5 h. The reaction mixture was dissolved in 15 mL of DCM and subsequently washed with sodium bicarbonate until the aqueous layer reached neutral pH. The organic phase was then washed with DI water (3 times) and brine solution (2 times). Next, the excess solvent was removed under reduced pressure from the organic phase, and the flash column chromatography (Biotage[®] Selekt Systems, Biotage[®] Sfär Silica columns – 60 μm , 100 g) with ethyl acetate (25 v/v%) and hexanes (75 v/v%) was employed to purify BSA. The product was dried at 50 $^{\circ}\text{C}$ under vacuum for 24 h, and the dried product was obtained as a viscous, orangish liquid (yield \sim 40 mol%). ^1H NMR (600 MHz, $\text{DMSO-}d_6$) δ 8.25–7.95 (m, 2H), 6.44 (d, $J = 0.9$ Hz, 3H), 3.68 (s, 12H), 1.56 (d, $J = 27.3$ Hz, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{19}\text{H}_{24}\text{O}_6$ 348.1573; found 348.1643 [Fig. S4, ESI[†]].

2.2.2. General procedure for the synthesis of diglycidyl ethers. Bisguaiacol/bissyringol diglycidyl ethers were synthesized per protocols reported in the literature (Scheme 1c).⁴³ In brief, bisguaiacol/bissyringol (0.02 mol), epichlorohydrin (0.20 mol), and TBAB (0.002 mol) were transferred into an oven-dried, single-neck, round-bottom flask (25 mL) equipped with a magnetic stir bar. The reaction mixture was sparged with Ar gas for 40 min at \sim 20 $^{\circ}\text{C}$ under constant stirring. Next, the mixture was heated at 50 $^{\circ}\text{C}$ for 2 h in a silicone oil bath. Subsequently, the reaction flask was immediately transferred into an ice bath maintained at \sim 0 $^{\circ}\text{C}$. After 15 min, 40 wt% of aqueous NaOH (0.08 mol) was added dropwise under constant stirring to the cooled reaction flask, and the reaction was continued at 20 $^{\circ}\text{C}$ for 16 h in a water bath. Then, the reaction mixture was dissolved in 10 mL of DCM, washed with 20 mL of DI water (3 times) in a separation funnel until the aqueous layer reached neutral pH, and then washed with brine solution (3 times). The organic layer was collected and then purified by automated flash column chromatography (Biotage[®] Selekt, Sfär Silica column – 60 μm particle size, 100 \AA pore size, 100 g silica gel) with a step gradient of ethyl acetate (60% v/v) and hexanes (40% v/v) for elution. The solvents were removed from the product using a rotary evaporator, and the concentrated product was dried in an oven under vacuum at 50 $^{\circ}\text{C}$ for 24 h.



The yield of the respective bisguaiacol/bissyringol diglycidyl ether varied between ~75–80 mol% after purification (purity >99%). The ^1H NMR spectra of the diglycidyl ethers with peak assignments are in Fig. S5–S10 (ESI†).

BPF diglycidyl ether (purchased). Viscous, colorless liquid, ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 7.21–6.71 (m, 9H), 4.39–4.10 (m, 2H), 4.01–3.69 (m, 4H), 2.94–2.57 (m, 4H). FTMS (ESI, m/z): calculated for $\text{C}_{19}\text{H}_{20}\text{O}_4$ 312.1362; found 312.1444.

BGF diglycidyl ether. White solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.77 (dd, $J = 67.3, 8.0$ Hz, 6H), 4.22 (d, $J = 8.6$ Hz, 2H), 3.91–3.62 (m, 10H), 3.28 (d, $J = 4.0$ Hz, 2H), 2.88–2.77 (m, 2H), 2.67 (t, $J = 4.7$ Hz, 2H). FTMS (ESI, m/z): calculated for $\text{C}_{21}\text{H}_{24}\text{O}_6$ 372.1573; found 372.1574.

BSF diglycidyl ether. Viscous, light-greenish liquid (purity: >99%, yield: ~75 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.87 (d, $J = 8.5$ Hz, 1H), 6.74 (d, $J = 7.6$ Hz, 1H), 6.49 (s, 2H), 4.20 (d, $J = 14.1$ Hz, 1H), 4.03 (d, $J = 11.4$ Hz, 1H), 3.83–3.65 (m, 16H), 3.25 (d, $J = 32.8$ Hz, 2H), 2.77 (s, 2H), 2.55 (s, 2H). FTMS (ESI, m/z): calculated for $\text{C}_{23}\text{H}_{28}\text{O}_8$ 432.1784; found 432.1861.

BPA diglycidyl ether (purchased). White solid, ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 7.22–6.98 (m, 4H), 6.95–6.70 (m, 4H), 4.26 (dd, $J = 11.3, 2.8$ Hz, 2H), 3.79 (dd, $J = 11.3, 6.5$ Hz, 2H), 2.82 (dd, $J = 5.1, 4.3$ Hz, 2H), 2.69 (dd, $J = 5.1, 2.7$ Hz, 2H), 1.58 (s, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{21}\text{H}_{24}\text{O}_4$ 340.1675; found 340.1744.

BGA diglycidyl ether. Viscous, light-yellowish liquid (purity: >99%, yield: ~75 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.94–6.64 (m, 6H), 4.21 (ddd, $J = 24.1, 11.4, 2.8$ Hz, 2H), 3.84–3.61 (m, 8H), 2.80 (ddd, $J = 20.0, 5.1, 4.2$ Hz, 2H), 2.66 (ddd, $J = 13.9, 5.1, 2.6$ Hz, 2H), 1.60 (d, $J = 7.2$ Hz, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{23}\text{H}_{28}\text{O}_6$ 400.1886; found 400.1951.

BSA diglycidyl ether. Orangish solid (purity: >99%, yield: ~80 mol%), ^1H NMR (600 MHz, CDCl_3) δ 6.36 (s, 4H), 4.11–3.90 (m, 4H), 3.71 (s, 12H), 3.30 (dtd, $J = 5.8, 4.1, 2.6$ Hz, 2H), 2.74 (dd, $J = 5.0, 4.2$ Hz, 2H), 2.56 (dd, $J = 5.0, 2.7$ Hz, 2H), 1.58 (s, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{25}\text{H}_{32}\text{O}_8$ 460.2097; found 460.2172.

2.2.3. General procedure for the synthesis of cyclic carbonates. Cyclic carbonates were synthesized by catalytic carbonylation of diglycidyl ethers (Scheme 1d).⁴⁵ The protocol to synthesize BGA cyclic carbonate is provided as a representative example. BGA diglycidyl ether (0.0085 mol, 3.3 g) and TBAB (0.04 g, 1.5 mol% relative to diglycidyl ether) were added in a 25-mL Teflon™ cylindrical liner equipped with a magnetic stir bar. The liner was sealed in a 25-mL Parr reactor and flushed with CO_2 (3 times) at ~20 °C. Next, the reactor was pressurized with ~30 bar CO_2 and placed in a heating jacket (ceramic band heater) connected to a proportional-integral-derivative controller. The reaction continued for 24 h at 130 °C under constant stirring. After the designated time, the controller was turned off, and the reactor was allowed to passively cool to ~20 °C in air for ~1.5 h.

The reactor was subsequently depressurized and unsealed to collect the resulting product from the liner. The product was then dissolved in acetone (3 mL) and precipitated in DI water (30 mL) two times. The precipitated solid was dried in an oven under vacuum at 65 °C for 24 h, and the dried product was obtained as an off-white solid (yield ~85 mol% and purity >99%). The other cyclic carbonates were synthesized similarly with yields ~85 mol% and purities >99%. The ^1H NMR spectra of the cyclic carbonates with peak assignments are in Fig. S11–S16 (ESI†).

BPF cyclic carbonate. White solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 7.21–6.71 (m, 9H), 4.39–4.10 (m, 2H), 4.01–3.69 (m, 4H), 2.94–2.57 (m, 4H). FTMS (ESI, m/z): calculated for $\text{C}_{21}\text{H}_{20}\text{O}_8$ 400.1158; found 400.1243.

BGF cyclic carbonate. Off-white solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.95–6.66 (m, 6H), 5.10 (dddd, $J = 8.3, 6.2, 4.2, 2.0$ Hz, 2H), 4.61 (t, $J = 8.5$ Hz, 2H), 4.46–4.31 (m, 2H), 4.29–4.02 (m, 4H), 3.73 (d, $J = 6.0$ Hz, 8H). FTMS (ESI, m/z): calculated for $\text{C}_{23}\text{H}_{24}\text{O}_{10}$ 460.1369; found 460.1443.

BSF cyclic carbonate. Off-white solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.82 (dd, $J = 75.1, 8.6$ Hz, 2H), 6.50 (s, 2H), 5.11–4.84 (m, 2H), 4.73–4.38 (m, 4H), 4.26–3.87 (m, 4H), 3.82–3.66 (m, 14H). FTMS (ESI, m/z): calculated for $\text{C}_{25}\text{H}_{28}\text{O}_{12}$ 520.1581; found 520.1652.

BPA cyclic carbonate. White solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 7.21–7.02 (m, 4H), 6.92–6.75 (m, 4H), 5.13 (dtt, $J = 8.6, 6.0, 2.6$ Hz, 2H), 4.62 (t, $J = 8.5$ Hz, 2H), 4.37 (dd, $J = 8.4, 5.9$ Hz, 2H), 4.29–4.11 (m, 4H), 1.58 (s, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{23}\text{H}_{24}\text{O}_8$ 428.1471; found 428.1536.

BGA cyclic carbonate. Off-white solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.96–6.67 (m, 6H), 5.20–5.02 (m, 2H), 4.60 (dt, $J = 13.8, 8.5$ Hz, 2H), 4.50–4.31 (m, 2H), 4.31–4.06 (m, 4H), 3.71 (d, $J = 26.7$ Hz, 6H), 1.60 (d, $J = 6.6$ Hz, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{25}\text{H}_{28}\text{O}_{10}$ 488.1682; found 488.1771.

BSA cyclic carbonate. Light-orangish-yellow solid (purity: >99%, yield: ~85 mol%), ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 6.51 (s, 4H), 5.08–4.91 (m, 2H), 4.69–4.45 (m, 4H), 4.35–4.08 (m, 2H), 4.05–3.86 (m, 2H), 3.71 (s, 12H), 1.64 (s, 6H). FTMS (ESI, m/z): calculated for $\text{C}_{27}\text{H}_{32}\text{O}_{12}$ 548.1894; found 548.1967.

2.2.4. ^1H NMR spectroscopy. All NMR samples (except BSA diglycidyl ether) were prepared in $\text{DMSO}-d_6$ and analyzed on a Bruker AVIII 600 Hz spectrometer. The BSA diglycidyl ether sample was prepared in CDCl_3 due to its limited solubility in $\text{DMSO}-d_6$. ^1H NMR spectra were analyzed using the MestReNova v1.8 software package. Chemical shifts were reported with respect to the solvent peak at 2.50 ppm for $\text{DMSO}-d_6$ and at 7.26 ppm for CDCl_3 .

2.2.5. Mass spectrometry. The exact molar masses of the lignin-derivable bisphenols, diglycidyl ethers, and cyclic



carbonates were determined by ESI-FTMS using a Q-Exactive Orbitrap (Thermo Fisher Scientific) mass spectrometer. Samples were dissolved in acetonitrile (0.1 mg mL⁻¹), and the ESI-FTMS was conducted in positive-ion mode by direct syringe injection of samples in the mass spectrometer.

2.3. Polymer synthesis and characterization

2.3.1. General procedure for the synthesis of NIPUs. NIPUs were synthesized *via* the step-growth polyaddition of the cyclic carbonates and diamines (Scheme 1e). The protocol to synthesize BGA-NIPU is provided as a representative example. BGA cyclic carbonate (0.50 g, 1.02 mmol), DA-10 (0.176 g, 1.02 mmol), LiCl (2.1 mg, 0.05 mmol), and anhydrous DMSO (1 mL, 1 mmol L⁻¹) were all added to an oven-dried, single-neck, round-bottom flask (10 mL). The flask was sealed with a rubber septum, and the reaction mixture was sparged with Ar gas (~15 min). The reaction was continued in a silicone oil bath for 24 h at 100 °C. After the reaction, the mixture was passively cooled to ~20 °C, diluted with 4 mL of THF, and precipitated from DI water (40 mL). The polymer was redissolved in THF (4 mL) and again precipitated from DI water (40 mL). The polymer was dried under vacuum at 85 °C for 48 h. NMR spectra of all the NIPUs with peak assignments are in Fig. S17–S22 (ESI†).

BPF-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.31–6.58 (m, 11H), 5.20 (s, 1H), 5.03–4.80 (m, 1H), 4.09–3.70 (m, 10H), 3.56 (s, 1H), 2.93 (q, *J* = 6.5 Hz, 4H), 1.38–1.12 (m, 16H).

BGF-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.14 (d, *J* = 27.5 Hz, 2H), 6.77 (d, *J* = 59.6 Hz, 6H), 5.31–4.70 (m, 3H), 4.10–3.50 (m, 17H), 2.94 (s, 4H), 1.29 (d, *J* = 58.5 Hz, 16H).

BSF-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.06 (s, 2H), 6.77 (d, *J* = 53.8 Hz, 2H), 6.47 (d, *J* = 6.0 Hz, 2H), 5.12–4.66 (m, 3H), 4.17–3.53 (m, 23H), 2.93 (s, 4H), 1.29 (d, *J* = 58.6 Hz, 16H).

BPA-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.09 (d, *J* = 8.5 Hz, 6H), 6.81 (d, *J* = 8.6 Hz, 4H), 5.20 (d, *J* = 5.1 Hz, 1H), 4.95–4.78 (m, 1H), 4.11–3.79 (m, 8H), 3.56 (s, 1H), 2.94 (s, 4H), 1.57 (s, 6H), 1.29 (d, *J* = 59.7 Hz, 16H).

BGA-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.24–6.99 (m, 2H), 6.92–6.64 (m, 6H), 5.18 (s, 1H), 4.87 (d, *J* = 10.6 Hz, 1H), 4.18–3.48 (m, 15H), 2.94 (s, 4H), 1.59 (s, 6H), 1.29 (d, *J* = 59.0 Hz, 16H).

BSA-NIPU. ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.07 (s, 2H), 6.59 (d, *J* = 77.9 Hz, 4H), 5.05–4.53 (m, 2H), 4.31–3.45 (m, 21H), 2.93 (s, 4H), 1.61 (d, *J* = 19.7 Hz, 6H), 1.24 (d, *J* = 17.3 Hz, 16H).

2.3.2. ¹H NMR spectroscopy. All NIPU NMR samples were prepared in DMSO-*d*₆ and analyzed on a Bruker AVIII 600 Hz spectrometer, and ¹H NMR spectra were analyzed using the MestReNova v1.8 software package. Chemical shifts were reported with respect to the solvent peak at 2.50 ppm for DMSO-*d*₆.

2.3.3. Attenuated total reflectance-Fourier transform infrared (ATR-FTIR) spectroscopy. Hydrogen bonding and NIPU

formation were assessed through ATR-FTIR spectroscopy using a Thermo Nicolet NEXUS 870 FTIR (Thermo Fisher Scientific) with a deuterated triglycine sulfate/potassium bromide (DTGS/KBr) detector. All cyclic carbonates and NIPUs were recorded at a resolution of 4 cm⁻¹ with 256 scans in the 4000–500 cm⁻¹ range. NIPU formation was assessed by examination of the cyclic carbonate carbonyl stretching band at ~1790 cm⁻¹, the urethane carbonyl stretching band at ~1700 cm⁻¹, and the broad hydroxyl stretching band at ~3100–3700 cm⁻¹ (Fig. S23 and S24, ESI†).

2.3.4. Gel permeation chromatography (GPC). The molar masses of the polymers were determined using an HLC-8420 EcoSEC Elite GPC instrument from TOSOH with a refractive index (RI) detector and four columns in series (1x TSKgel SuperAW-L guard column, 2x TSKgel SuperAWM-H columns, and 1x TSKgel SuperAW2500 column). The mobile phase was DMAc with 0.5 wt.% of LiBr. The flow rate of the mobile phase was 0.4 mL min⁻¹ with an injection volume of 80 μL. A calibration curve was made using narrow-dispersity poly(methyl methacrylate) [PMMA] standards from Agilent Technologies. The polymer solutions were prepared by dissolving samples in DMAc at ~1.0 mg mL⁻¹ (at ~20 °C for 24 h). The samples were then filtered using 0.1-μm PTFE filters. The differential weight fraction distributions as a function of molar mass for the NIPUs are shown in Fig. S25 (ESI†).

2.3.5. NIPU film fabrication. Solvent casting was employed for NIPU film preparation. The respective polymer was dissolved in DMF (~10 w/v%) and stirred at ~20 °C for 24 h before pouring the solution into the mold (Teflon™ Petri dish of diameter ~7 cm). The mold was covered with perforated aluminum foil and placed on a hot plate maintained at 60 °C for 2 h. Then, the mold was placed into an oven at 85 °C for 24 h at ambient pressure, followed by 85 °C for 48 h under vacuum. Next, thermogravimetric analysis (TGA) [see Section 2.3.8 for additional details] was employed to ensure the complete removal of solvent from the NIPU films. Images of free-standing films are shown in Fig. S26 (ESI†).

2.3.6. Tensile tests. The tensile properties, *i.e.*, Young's modulus, ϵ_{break} , ultimate tensile strength, toughness (area under the tensile stress-strain curve), were measured using a Zwick/Roell Z 0.5 (500 N zwicki-Line) tensile tester with a 100 N-capacity load cell and a clamp force of 200 N. The specimens with dimensions (Fig. S27, ESI†) of 9.5 mm × 2.5 mm × 0.17–0.24 mm (gauge length × width × thickness) were subjected to an extension rate of 10 mm min⁻¹ until film breakage. Five replicates were analyzed, and the average values were reported herein. All tests were performed at ~20 °C.

2.3.7. Differential scanning calorimetry (DSC). The *T*_gs of synthesized NIPUs were determined using a Discovery Series DSC 972000.901 instrument (TA Instruments). Samples were loaded into hermetically sealed aluminum pans. The pans were heated from -20 °C to 170 °C at a rate of 10 °C min⁻¹ under continuous N₂ flow (50 mL min⁻¹) and held isothermally for 2 min at 170 °C. Then, the pans were cooled to -20 °C at a rate of 10 °C min⁻¹ under N₂ flow. The data from the third heating trace were used for analysis. The *T*_g was reported as the



midpoint of the inflection in the thermogram from the third heating.

2.3.8. Thermogravimetric analysis (TGA). The thermal stability of the polymers and the solvent content of the films were investigated using a discovery series thermogravimetric analyzer TA Q500 (TA Instruments). ~5 mg of the polymer was placed into a 100 μL platinum pan. Under a continuous flow of N_2 (50 mL min^{-1}) gas, the sample was heated to 200 $^\circ\text{C}$ at a rate of 10 $^\circ\text{C min}^{-1}$ and held at this temperature for 2 min to evaporate any water or solvents that may have been present. Then, the sample was cooled to 30 $^\circ\text{C}$ and heated to 700 $^\circ\text{C}$ at a rate of 10 $^\circ\text{C min}^{-1}$ in the N_2 atmosphere. The thermal stability ($T_{\text{d}5\%}$) was reported as the temperature at which a 5 wt% loss was noted.

2.3.9. Rheological analysis. The rheological response of the polymers was quantified using a strain-controlled, torsional Ares G2 rheometer (TA Instruments). Vacuum-dried films were stacked and loaded into the test geometry (8-mm-diameter parallel plates) that had been preheated to 160 $^\circ\text{C}$. The samples were slowly pressed and trimmed, such that the final thickness was 1 mm. A pre-shear step was then applied at 1 rad s^{-1} and 0.2% strain for 60 s to relax any stresses arising from sample loading. A series of frequency sweeps (between 0.1 and 100 rad s^{-1}) and temperature ramps were performed to obtain linear viscoelastic responses between 150 $^\circ\text{C}$ and 40 $^\circ\text{C}$ in 10 $^\circ\text{C}$ increments. The strain amplitude remained within the linear viscoelastic regime (as determined by amplitude sweeps) and was selected to be 0.5% and 0.2% for temperatures above and below 90 $^\circ\text{C}$, respectively. All experiments were conducted in an N_2 atmosphere.

Linear viscoelastic spectra at a reference temperature of $T_{\text{ref}} = 150$ $^\circ\text{C}$ were constructed using the frequency sweep data *via* time-temperature superposition. Vertical shift factors of

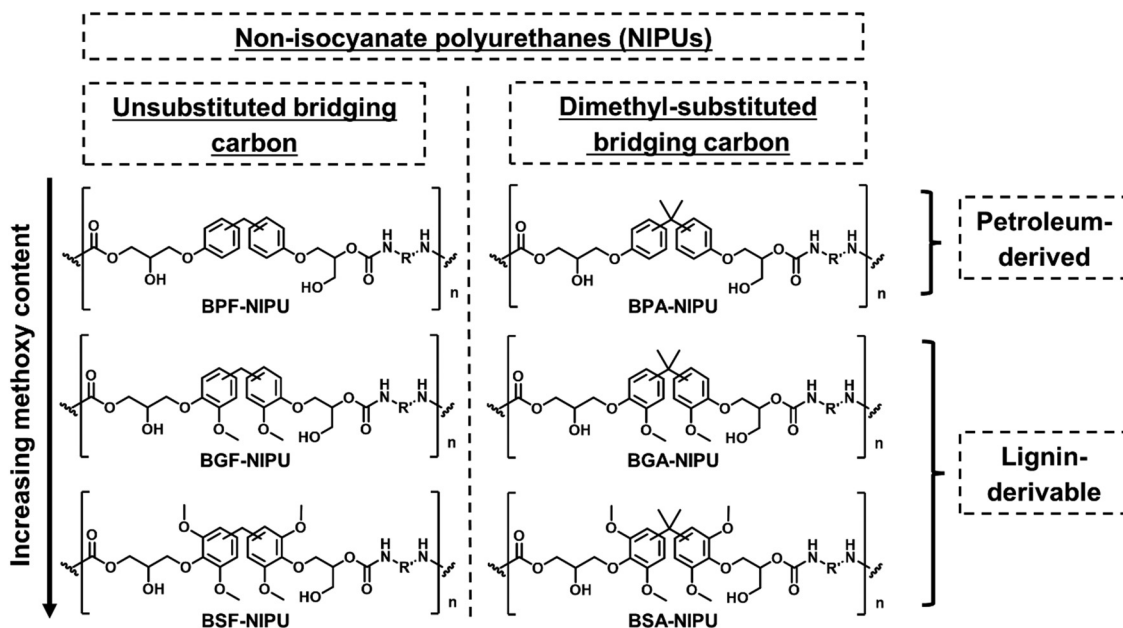
T_{ref}/T were applied, and, subsequently, horizontal shift factors were determined using the algorithm in the TRIOS software (TA Instruments).

3. Results and discussion

For this work, NIPUs, synthesized from monomers with varying methoxy-group content on the aromatic rings and the substituents to the bridging carbons, were studied to understand the impact of these structural features on the thermal, mechanical, and rheological behavior of the resulting polymers. The molecular structures of petroleum-based NIPUs and their lignin-derivable analogues are presented in Scheme 2. In the following sections, the synthesis of lignin-derivable precursors and NIPUs is discussed. Additionally, the impact of bridging carbon substituents and methoxy content on the hydrogen-bonding and mechanical properties is presented, and the influence of these structural features on the rheological response is examined.

3.1. Synthesis of lignin-derivable building blocks and NIPUs

Several lignin-derivable alternatives to commercial bisphenols (BPA/BPF) were synthesized, as shown in Schemes 1a and b. BGF and BSF were synthesized by an acid-catalyzed electrophilic aromatic substitution reaction between guaiacol and vanillyl alcohol (for BGF) or syringol and syringyl alcohol (for BSF) (Scheme 1a).^{43,46} The direct condensation approach eliminates the use of hazardous formaldehyde inherent in BPF production.⁴¹ BGF and BSF do not have a substituted bridging carbon between the two aromatic rings, which makes them more structurally similar to BPF. BGA and BSA were synthesized *via* an electrophilic aromatic condensation between acetone and guaiacol (for BGA) or acetone and syringol (for BSA)



Scheme 2 The library of synthesized NIPUs highlighting key features of their chemical structure and source.



(Scheme 1b).⁴⁴ The availability of bio-based acetone enables a potentially greener process.⁴⁴ Both BGA and BSA have dimethyl substituents on the bridging carbon between the two aromatic moieties and more closely resemble the structure of BPA. As mentioned above, the methoxy groups on these lignin-derivable compounds are believed to reduce the toxicity concerns of BSA, BGA, BSF, and BGF vs. BPA and BPF.⁴⁷

The respective bisphenols were converted to diglycidyl ethers and then to cyclic carbonates (Schemes 1c and d). The exact molar masses of the precursors at each step were determined using liquid chromatography–mass spectroscopy, and the chemical structures of the compounds were confirmed using ¹H NMR spectroscopy. Both mass spectrometry and ¹H NMR spectroscopy results showed excellent agreement with the expected chemical formulae and chemical structures of the compounds, respectively.

These cyclic carbonates were reacted with a bio-derivable diamine (DA-10)^{53,55,56} to obtain NIPUs (Scheme 1e). All NIPUs exhibited comparable molar masses of ~15–20 kg mol⁻¹, as shown in Table 1. Although traditional PUs can be obtained with molar masses >20 kg mol⁻¹, most of the NIPUs reported in the literature possess a number-average molecular weight (M_n) <10 kg mol⁻¹ due to the lower reactivity of cyclic carbonates vs. isocyanates,^{9,53,57} along with numerous possible side reactions (*e.g.*, urea formation, amidification reaction, oxazolidinone formation) that can limit molar mass.^{6–8} Additionally, the five-membered cyclic carbonates are relatively less reactive than six-, seven-, or eight-membered cyclic carbonates because of the ring strain.^{6–8} Notably, in this work, higher-than-average M_n values were achieved, and free-standing films were fabricated (Fig. S26, ESI[†]), which could suggest that the hydrogen bonding in these polymers improves mechanical robustness even in lower-molar-mass NIPUs. As shown in Table 1, most of the NIPUs had dispersities (D_s) ~2.5–3, except for BPF-NIPU ($D \sim 4$). This exception was likely a result of the higher extent of oligomerization (~0.10) in the commercial BPF diglycidyl ether vs. the other diglycidyl ethers (~0.01–0.03).

3.2. Estimation of hydrogen-bonding in NIPUs

Hydrogen bonding plays a crucial role in the properties (mechanical/rheological) of PUs/NIPUs.^{58,59} Herein, the lignin-derivable NIPUs displayed higher percentages of hydrogen-bonded acceptors (attributable to methoxy moieties) than their petroleum-based analogues. The total number of hydrogen-bond

acceptors and donors was calculated from the theoretical structure of the repeat unit. In particular, –OH and –NH were considered the hydrogen-bond donors, and urethane carbonyls (–C=O), ether oxygens (–O–), and methoxy groups (–OCH₃) were considered the hydrogen-bond acceptors. As shown in Table 2, the hydrogen-bond acceptor count per repeat unit in the NIPUs increased with the number of methoxy groups (0, 2, and 4) in BPF-/BGF-/BSF-NIPUs.

Semi-quantitative analysis using ATR-FTIR spectroscopy was employed to estimate the amount of hydrogen bonding in these NIPUs. The –OH/–NH absorption bands (3100–3700 cm⁻¹) of the NIPUs samples were deconvoluted using Gaussian distributions.^{58,60} The –OH/–NH absorption bands were fit to two subpeaks because the stretching vibrations of hydrogen-bonded –OH/–NH were at a slightly lower wavenumber (~3300–3350 cm⁻¹) than the free –OH/–NH vibrations (~3450–3550 cm⁻¹), and the relative areas of these two peaks were calculated.^{58,60} Hydrogen bonding can cause the vibrational frequencies to red-shift (*i.e.*, migrate towards lower energy) for the hydrogen-bonding motifs, with stronger bonds precipitating shifts to lower energies.^{61,62} The hydrogen-bonded region includes all self- and trans-associations, and the broadness of this region depends upon the number/strength of associations. Herein, the added methoxy groups (hydrogen-bond acceptors) increased the interactions in the hydrogen-bonded region, and therefore, the broadness of this region. FTIR analysis suggested that increasing the number of methoxy groups from 0 to 2 to 4 in BPF-/BGF-/BSF-NIPUs led to higher percentages of hydrogen-bonded –OH/–NH groups (~65%, ~85%, ~95%, respectively). Similar trends were found in the dimethyl-substituted analogues, BPA-/BGA-/BSA-NIPUs (Table 2 and Fig. 1). This finding demonstrated that the added hydrogen-bonding capability of the monomers directly translated to greater secondary interactions between NIPU chains, which was expected to influence mechanical and rheological properties.^{58,60,63}

3.3. Mechanical behavior of NIPUs

The impacts of both hydrogen bonding and substitutive group on the solid-state performance of the NIPUs were quantified using tensile testing. The lignin-derivable NIPUs exhibited improved mechanical properties (ϵ_{break} and toughness) over their petroleum-based counterparts without a reduction in Young's modulus or tensile strength (Fig. 2 and Fig. S28 (ESI[†]) and Table 3). The ϵ_{break} and toughness of NIPUs increased with an increase in the methoxy-group content. For example, in the unsubstituted bridging-group case, BSF-NIPU exhibited the highest ϵ_{break} (~210%) and toughness (~62 MJ m⁻³), followed by BGF-NIPU ($\epsilon_{\text{break}} \sim 185\%$, toughness ~58 MJ m⁻³), and then BPF-NIPU ($\epsilon_{\text{break}} \sim 140\%$, toughness ~42 MJ m⁻³). In the dimethyl-substituted cases, both lignin-derivable versions, BGA-NIPU ($\epsilon_{\text{break}} \sim 145\%$, toughness ~51 MJ m⁻³) and BSA-NIPU ($\epsilon_{\text{break}} \sim 120\%$, toughness ~44 MJ m⁻³) exhibited higher ϵ_{break} and toughness than petroleum-derived BPA-NIPU ($\epsilon_{\text{break}} \sim 95\%$, toughness ~36 MJ m⁻³). The increased toughness was attributed to the hydrogen bonds between chains that acted as physical crosslinks. As the petroleum-derived NIPUs had a fixed

Table 1 Molecular weight characterization data for the NIPUs

	Polymer	Methoxy groups	M_n^a (kg mol ⁻¹)	D
Unsubstituted bridging carbon	BPF-NIPU	0	17	4.1
	BGF-NIPU	2	19	2.3
	BSF-NIPU	4	14	2.7
Dimethyl-substituted bridging carbon	BPA-NIPU	0	19	2.6
	BGA-NIPU	2	17	2.6
	BSA-NIPU	4	15	3.0

^a Determined relative to PMMA standards using data from GPC with RI detectors [GPC solvent: DMAc + 0.5 wt% LiBr].



Table 2 Calculated hydrogen-bond donors, acceptors, and hydrogen-bonded -OH/-NH percentage for NIPUs

	Polymer	Methoxy groups	Hydrogen-bond donor count (per repeat unit)	Hydrogen-bond acceptor count (per repeat unit)	Hydrogen-bonded -OH/-NH ^a (%)
Unsubstituted bridging carbon	BPF-NIPU	0	4	6	~ 65
	BGF-NIPU	2	4	8	~ 85
	BSF-NIPU	4	4	10	~ 95
Dimethyl-substituted bridging carbon	BPA-NIPU	0	4	6	~ 60
	BGA-NIPU	2	4	8	~ 90
	BSA-NIPU	4	4	10	~ 95

^a Estimated from ATR-FTIR spectroscopy data.



Fig. 1 Deconvoluted hydroxyl region in the ATR-FTIR spectra of the NIPUs. The -OH/-NH absorption bands were fit using two Gaussian distributions: one for hydrogen-bonded -OH/-NH groups and another for free -OH/-NH groups.

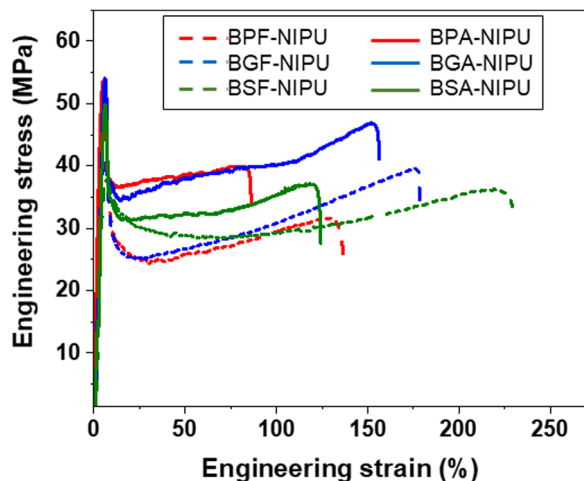


Fig. 2 Representative tensile engineering stress vs. engineering strain curves for NIPUs acquired from uniaxial tensile testing using a load cell of 100 N and a strain rate of 10% min⁻¹ at ~20 °C.

number of donors for potential hydrogen-bonding interactions, the methoxy moieties in the lignin-derivable systems could provide additional sites for intra- and inter-molecular hydrogen bonding in the NIPU matrix.^{58,64} The density of these physical crosslinks possibly increased with an increase in methoxy content, which likely led to higher extensibility; however, the increased in the extent of hydrogen bonding in BSA-NIPU did not equate to a higher ϵ_{break} and toughness than that found for BGA-NIPU. Thus, more in-depth mechanical investigations of the chain dynamics under stretching might help to understand the phenomenon behind this anomaly.

The unsubstituted analogues exhibited a higher ϵ_{break} than dimethyl-substituted NIPUs. The ϵ_{break} trend was BPF-NIPU > BPA-NIPU; BGF-NIPU > BGA-NIPU; and BSF-NIPU > BSA-NIPU (see Table 3). This trend was probably a result of the absence of substituents on the bridging carbon, allowing free rotation (increasing chain mobility) of the polymer backbone.^{30,65} Additionally, the Young's moduli for all the unsubstituted bridging



Table 3 Mechanical property characterization summary of the NIPUs. Values were calculated for five individual samples and averaged. The reported errors were standard deviations

	Polymer	Methoxy groups	Ultimate tensile strength (MPa)	Young's modulus (GPa)	ϵ_{break} (%)	Toughness (MJ m ⁻³)
Unsubstituted bridging carbon	BPF-NIPU	0	47 ± 3	1.2 ± 0.1	140 ± 21	42 ± 10
	BGF-NIPU	2	44 ± 3	1.2 ± 0.1	185 ± 17	58 ± 5
	BSF-NIPU	4	39 ± 4	1.2 ± 0.2	210 ± 18	62 ± 8
Dimethyl-substituted bridging carbon	BPA-NIPU	0	55 ± 1	1.4 ± 0.2	95 ± 13	36 ± 6
	BGA-NIPU	2	52 ± 2	1.4 ± 0.1	145 ± 20	51 ± 13
	BSA-NIPU	4	55 ± 6	1.4 ± 0.1	120 ± 16	44 ± 8

carbon-based NIPUs (BPF-/BGF-/BSF-NIPUs) were comparable (~ 1.2 GPa) within experimental error. A similar trend was noted for all the dimethyl-substituted bridging carbon-based NIPUs (BPA-/BGA-/BSA-NIPUs) with Young's moduli of ~ 1.4 GPa. This phenomenon was likely due to the modulus of the glassy state being dominated by localized motion of the polymer backbone, and therefore, the substituents, had little effect on the modulus. This relationship was expected because of the rigid, aromatic units present in the polymeric backbones could contribute much more to the modulus than inter-chain friction from small side groups.

Next, π - π stacking interactions are expected to be comparable across all NIPUs as it has been reported that lignin-derivable G and S units can form π - π stacking interactions,^{66,67} and these interactions are almost equivalent in softwood and hardwood lignin samples.³⁸ Thus, the methoxy substituents may not significantly interrupt π - π interactions in lignin-derivable-NIPUs [BGF-/BGA-NIPUs (softwood derivable) and BSF-/BSA-NIPUs (hardwood derivable)] vs. petroleum-derived-NIPUs [BPF-/BPA-NIPUs].

3.4. Thermal properties of NIPUs

The thermal properties of these lignin-derivable NIPUs were benchmarked against their petroleum-derived analogues (Fig. 3). The T_g s of the NIPUs increased with an increase in the methoxy-group content (Fig. 3a). For example, BPF-NIPU with zero methoxy groups had a T_g of ~ 36 °C, BGF-NIPU with two methoxy groups had a T_g of ~ 39 °C, and BSF-NIPU with four methoxy groups had a T_g of ~ 42 °C. Similarly, BPA-NIPU with zero methoxy groups had a T_g of ~ 46 °C, BGA-NIPU with two methoxy groups had a T_g of ~ 50 °C, and BSA-NIPU with four methoxy groups had a T_g of ~ 55 °C. These results were consistent with the literature, as the presence of side groups (*i.e.*, methoxy moieties) on the polymer backbone could increase T_g by limiting the chain mobility.⁶⁸ Additionally, secondary forces, such as hydrogen-bonding and dipole-dipole interactions,⁶⁹⁻⁷¹ between the chains also might be responsible for an increase in T_g due to greater constraints on segmental mobility imposed by associations.⁷²

The NIPUs based on monomers with dimethyl-substituted bridging carbons exhibited higher T_g s than their unsubstituted analogues (Fig. 3b). The T_g values were BPA-NIPU > BPF-NIPU; BGA-NIPU > BGF-NIPU; and BSA-NIPU > BSF-NIPU (see Fig. 3b). The trend was most likely related to the rotational freedom of the aromatic ring around the central carbon, which was lower for the dimethyl group⁶⁸ as the dimethyl bridge



Fig. 3 (a) DSC thermograms of NIPUs with a heating rate of 10 °C min⁻¹ in an N₂ atmosphere (data from the third heating trace and curves were shifted vertically for clarity with T_g values marked on the respective curves), (b) T_g as a function of the methoxy-group content from 0 to 4: for unsubstituted bridging carbon- and dimethyl-substituted bridging carbon-based NIPUs. [Note: T_g values from the second and third heating traces were identical, and the error bars were determined using T_g values from the third heating traces of two separate samples for each NIPU.]

restricted the free rotation of the polymer backbone (BPA-/BGA-/BSA-NIPUs) and led to a comparably higher T_g (BPF-/BGF-/BSF-NIPUs). This finding was important because it had been reported previously that the T_g s of the bisguaiacol/



bissyringol-based polymers were lower than BPA-based counterparts.⁴¹ In combination with the mechanical responses of the NIPUs, these DSC results demonstrate that hydrogen bonding contributes to the ability to obtain glassy polymers with tunable extensibility at room temperature.

The thermal stabilities of lignin-derivable NIPUs were equivalent to those of the BPA-/BPF-based NIPUs ($T_{d5\%} \sim 280$ °C) (Fig. 4a). The $T_{d5\%}$ values for BPF-/BGF-/BSF-NIPU were ~ 283 °C, 276 °C, and 288 °C, respectively. Similarly, BPA-/BGA-/BSA-NIPUs exhibited $T_{d5\%}$ values of ~ 275 °C, 285 °C, and 283 °C, respectively. The char contents were nearly identical across all NIPUs. Additionally, the most significant degradation event occurred at similar temperatures (~ 350 °C) across all NIPUs (Fig. 4b). The comparable thermal stabilities of all NIPUs ($T_{d5\%} \sim 280$ °C) were mainly attributed to the initial breakage of the hydroxyurethane linkages, which occurred at ~ 250 – 270 °C.^{25,45} These results were consistent with our previous findings for lignin-derivable BGF-/BGA-NIPU vs. BPF-/BPA-NIPU thermosets.⁴⁵ Interestingly, the increased methoxy content in these lignin-derivable NIPUs did not reduce thermal stability. This observation was noteworthy because it had been reported

previously that the increased oxygen content in lignin-derivable thermoplastic polymers, such as polycarbonates/polyesters,^{30,73} reduced thermal stability. Together, the methoxy moieties did not strongly impact the thermal decomposition temperature across the different lignin-derivable NIPUs.

3.5. Rheological behavior of NIPUs

Thermoplastic PUs are typically melt processed for fabrication of commercial products, and thus, their rheological behavior is key to macromolecular design. It has been well-established that the addition of hydrogen-bonding groups strongly affects melt flow in a wide range of polymers (*e.g.*, polyacrylates, polyolefins, polyethers) and in several complex ways (*e.g.*, increasing terminal relaxation time, altering slope in the terminal regime).^{69,74–77} Therefore, it is critical to gain an understanding of the contribution of additional hydrogen-bonding capabilities in lignin-derivable NIPUs. For unsubstituted NIPUs, the rheological behavior in the rubbery and glassy states (*i.e.*, frequencies $> 10^5$ rad s^{-1}) was similar for all polymers (Fig. 5a). A slight trend in the glassy modulus (*i.e.*, the plateau value of storage modulus, G' , at higher frequencies) was noticed whereby the modulus increased with increasing numbers of methoxy groups. The critical relaxation timescale (τ_c) (*i.e.*, the inverse of the frequency at which a crossover of G' and G'' occurs) when the transitional and rubbery regimes meet is related to localized, fast relaxations.⁷⁸ Interestingly, τ_c appeared to be non-monotonic with the number of methoxy groups (Table 4). The value of τ_c is typically sensitive to inter-monomer friction,⁷⁹ yet this trend was unexpected given the probable increase in friction with increasing methoxy moieties. This discrepancy might be due to the higher dispersity of BPF-NIPU as result of the extent of oligomerization in comparison to the other NIPUs or relative differences in the frictional contribution of methoxies subject to their position on the aromatic rings.

Whereas the high-frequency region of the viscoelastic response provides information regarding the motion of monomer units, the intermediate frequencies reveal long-range interactions between chains. Given the hypothesis that hydrogen bonding was improved for the NIPUs studied in this work, the rubbery response at moderate frequencies could be important for understanding both viscous and elastic behavior. With an increasing number of methoxy groups, a steady increase in the values of the plateau (*i.e.*, rubbery) modulus ($G_{N,0}$) occurred (Table 4). The $G_{N,0}$ values for BSF- and BGF-NIPUs were higher ($\sim 8 \times 10^5$ Pa) than that of BPF-NIPU ($\sim 7 \times 10^5$ Pa). This behavior was directly related to the increase in hydrogen bonding for BSF-NIPU ($\sim 90\%$) and BGF-NIPU ($\sim 85\%$) in comparison to BPF-NIPU ($\sim 60\%$) that was shown by ATR-FTIR spectroscopy. Additionally, as the difference in the hydrogen bonding between BSF- and BGF-NIPUs was not significant, the $G_{N,0}$ values for both the polymers were equivalent. The most prominent differences in the NIPU rheology were seen in the terminal regime. Between the petroleum-derived and lignin-derivable polymers, there was an order of magnitude decrease in the long relaxation time (τ_x). Although it would be expected that an increase in hydrogen bonding would lead to an increase in this timescale,⁶⁹ differences in molar mass and



Fig. 4 (a) TGA thermograms of NIPUs in N_2 atmosphere; sample weight (%) as a function of temperature with a heating rate of 10 °C min^{-1} , and (b) the first derivative of the weight remaining (%/°C) as a function of temperature for all NIPUs.



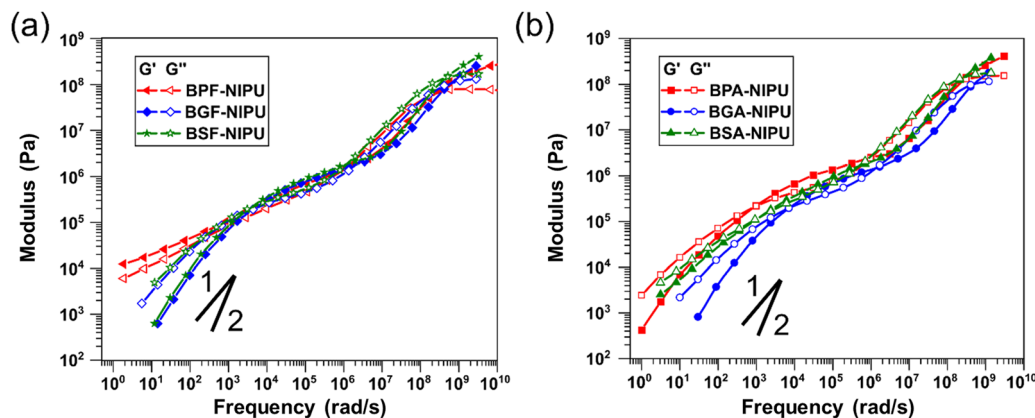


Fig. 5 Frequency-dependent storage (G') and loss (G'') moduli for (a) unsubstituted bridging carbon- and (b) dimethyl-substituted bridging carbon-based NIPUs. The master curves were constructed using time-temperature superposition and reported at 150 °C. Lines indicate typical power-law scalings of G' (slope of 2) and G'' (slope of 1).

Table 4 Rheological properties of NIPUs

	Polymer	Methoxy groups	τ_x^{ab} (s)	τ_c^{ac} (s)	$G_{N,0}^{ad}$ (Pa)	C_1	C_2 (K)
Unsubstituted bridging carbon	BPF-NIPU	0	1.6×10^{-3e}	7.6×10^{-7}	7.2×10^5	3.1	137
	BGF-NIPU	2	2.3×10^{-4}	4.7×10^{-7}	8.7×10^5	3.1	141
	BSF-NIPU	4	2.7×10^{-4}	9.6×10^{-7}	8.8×10^5	3.2	142
Dimethyl-substituted bridging carbon	BPA-NIPU	0	1.1×10^{-3}	1.0×10^{-6}	1.3×10^6	3.1	128
	BGA-NIPU	2	1.3×10^{-4}	7.1×10^{-7}	8.2×10^5	3.7	141
	BSA-NIPU	4	8.0×10^{-4}	2.5×10^{-6}	7.8×10^5	3.3	131

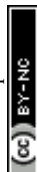
^a Given at the reference temperature of 150 °C. ^b Calculated as the inverse of the low-frequency crossover. ^c Calculated as the inverse of the crossover at high frequencies bounded by the rubbery and transitional regimes. ^d The value of G' at the minimum of phase angle vs. complex modulus (Fig. S29, ESI). ^e Approximated as the frequency in the terminal regime at which phase angle vs. complex modulus went through a maximum.

dispersity also would impact this relaxation behavior.⁷⁸ The BPF-NIPU exhibited a stronger deviation from the classical terminal power-law slopes of G' (slope of 2) and G'' (slope of 1) than the BGF- and BSF-NIPUs, indicating a greater strength of association between chains for the former.⁸⁰ This behavior was unsurprising considering the associations in BPF-NIPU were primarily between hydroxyls, which typically had a stronger hydrogen-bond energy than between methoxy and hydroxyl groups.^{81,82} Thus, even with a greater number of potential hydrogen-bonding groups, the lignin-derivable NIPUs flowed more easily because these interactions were more easily interrupted.

The transitional and glassy behavior also were similar in the case of the dimethyl-substituted NIPUs (Fig. 5b); however, there was a lesser dependence of the glassy moduli on methoxy content than for unsubstituted NIPUs. This finding might be related to the presence of the bridging group substituents, suggesting these substituents more prominently control solid viscoelasticity. τ_c followed the same trend as for unsubstituted NIPUs yet was slightly higher in magnitude for BPA-NIPU in comparison to BPF-NIPU, which might be because of the added friction associated with the bridging substituents that was insignificant in the presence of bulkier methoxy substituents on the ring. Although the value of $G_{N,0}$ for the dimethyl-substituted, lignin-derivable NIPUs was comparable to the

unsubstituted counterparts, BPA-NIPU had a higher value than BPF-NIPU. These results suggested that the rigidity of dimethyl-substituted NIPUs increases physical crosslinking; however, this effect had little impact on lignin-derivable NIPUs potentially because the methoxy groups interfere with the hydroxyl-hydroxyl hydrogen bonding, yielding a transient network with more, but weaker, junctions.

Dimethyl-substituted NIPUs exhibited additional features related to hydrogen bonding in the terminal regime.^{69,80} The expected terminal power-law scaling was followed for all the NIPUs (except BSA-NIPU). This finding demonstrated the strength of hydrogen bonding in BSA-NIPU. Although the impact was small on the plateau modulus, there was a more pronounced influence in the melt state because of the accessibility of hydrogen bonds once backbone features (*e.g.*, chain segments, entanglements) had relaxed. The trend in τ_x was similar to that seen for unsubstituted NIPUs, except for BSA-NIPU. This relationship could be related to the chain flexibility (*i.e.*, persistence length), as well as the hydrogen-bond strength.^{69,80} More extensive studies, such as computational investigations into the dynamic conformations of NIPUs under flow and systematic rheological experiments on NIPUs with varying molecular weights, will be required to fully understand the relaxation phenomena in these polymers.



Finally, the thermo-rheological responses of the NIPUs were examined. Time-temperature superposition was performed to construct master curves; however, the quality of the superposition varied. This phenomenon was best visualized using van Gorp-Palmen plots (Fig. S29, ESI†). For all unsubstituted NIPUs, the data did not superimpose to a single curve around the transitional regime, although only BPF-NIPU was poorly superimposed in the terminal regime, which was likely a consequence of a higher molecular-weight dispersity in comparison to the other NIPUs.⁷⁶ The dimethyl-substituted NIPUs showed good superposition across all temperatures, with only a noticeable spread in the data in the terminal regime caused by the noise related to measuring the low moduli. Vertical shift factors were nearly identical between all of the NIPUs (Fig. S30, ESI†), as demonstrated by the general equivalence of the Williams–Landel–Ferry parameters C_1 and C_2 (Table 4). The greatest deviation between the NIPUs was noticed at temperatures approaching the glass transition.

Lignin-derivable NIPUs exhibited a similar behavior to their petroleum-derived counterparts in the rubbery and glassy states, yet unexpectedly, the lignin-derivable NIPUs flowed more easily (*i.e.*, exhibited overall lower moduli in the terminal regime) and had faster relaxation times than the petroleum-derived NIPUs. Overall, these findings suggested that the added functionalities derived from lignin precursors were not a hindrance to processability but rather a benefit to modulating the strength of hydrogen bonding from sustainably manufactured NIPUs.

4. Conclusions

This work studied the effect of key structural handles in lignin-derivable bisphenols, such as methoxy-group content and bridging-carbon substitutions, on the thermal, mechanical, and rheological properties on the thermoplastic NIPUs. Notably, the methoxy groups in lignin-derivable thermoplastic NIPUs increased the extent of hydrogen bonding in such polymers, which led to increased ϵ_{break} and toughness *vs.* petroleum-based versions. For example, lignin-derivable, BGF-NIPU ($\epsilon_{\text{break}} \sim 185\%$, toughness $\sim 58 \text{ MJ m}^{-3}$) and BSF-NIPU ($\epsilon_{\text{break}} \sim 210\%$, toughness $\sim 62 \text{ MJ m}^{-3}$) exhibited higher ϵ_{break} and toughness than BPF-NIPU ($\epsilon_{\text{break}} \sim 140\%$, toughness $\sim 42 \text{ MJ m}^{-3}$). Furthermore, the T_g of NIPUs increased with an increase in the methoxy-group content as the side groups and secondary interactions arising due to methoxy substituents reduced the chain mobility. These increases in ϵ_{break} , toughness, and T_g s were obtained without any adverse effect on other polymer properties, such as modulus ($\sim 1.2\text{--}1.4 \text{ GPa}$), tensile strength ($\sim 40\text{--}55 \text{ MPa}$), and thermal stability ($\sim 280 \text{ }^\circ\text{C}$). Importantly, the melt rheology of the lignin-derivable NIPUs remained favorable, and these methoxy moieties offered improved processability *vs.* their non-methoxy-containing, petroleum-derived counterparts. Altogether, the thermomechanical properties of lignin-derivable NIPUs can be readily tuned with careful choice of lignin-derivable building blocks, and the structure–property–processing relationships established in this work can aid the development of sustainable polymeric materials.

Abbreviations

ATR-FTIR	Attenuated total reflectance-Fourier transform infrared
BGA	Bisguaiacol A
BGF	Bisguaiacol F
BPA	Bisphenol A
BPF	Bisphenol F
BSA	Bissyringol A
BSF	Bissyringol F
C_1	First Williams–Landel–Ferry equation parameter
C_2	Second Williams–Landel–Ferry equation parameter
\mathcal{D}	Dispersity
ϵ_{break}	Elongation-at-break
DA-10	1,10-diaminodecane
DCM	Dichloromethane
DI	Deionized
DMAC	<i>N,N</i> -dimethylacetamide
DMF	<i>N,N</i> -dimethylformamide
DMSO	Dimethyl sulfoxide
DMSO- d_6	Deuterated dimethyl sulfoxide
DSC	Differential scanning calorimetry
DTGS	Deuterated triglycine sulfate
ESI	Electrospray ionization
FTMS	Fourier transform mass spectrometry
G	Guaiacyl
G'	Storage modulus
G''	Loss modulus
$G_{N,0}$	Plateau modulus
GPC	Gel permeation chromatography
H	<i>p</i> -hydroxyphenyl
MDI	Methylene diphenyl diisocyanate
M_n	Number-average molecular weight
NIPU	Non-isocyanate polyurethane
NMR	Nuclear magnetic resonance
PU	Polyurethane
RI	Refractive index
S	Syringyl
TBAB	Tetrabutylammonium bromide
τ_c	Fast relaxation/critical timescale
$T_{d5\%}$	Temperature of 5% weight loss
TDI	Toluene diisocyanate
T_g	Glass transition temperature
TGA	Thermogravimetric analysis
THF	Tetrahydrofuran
T_{ref}	Rheology reference temperature
τ_x	Longest relaxation time

Author contributions

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acquisition, supervision, and project administration: T. H. E. and L. T. J. K.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 BASF Polyurethane, <https://polyurethanes.basf.us/>, (accessed January 10, 2024).
- 2 Lubrizol, Thermoplastic Polyurethane (TPU), <https://www.lubrizol.com/Engineered-Polymers/About/What-is-TPU>, (accessed January 10, 2024).
- 3 J. O. Akindoyo, M. D. H. Beg, S. Ghazali, M. R. Islam, N. Jeyaratnam and A. R. Yuvaraj, *RSC Adv.*, 2016, **6**, 114453–114482.
- 4 O. Kreye, H. Mutlu and M. A. R. Meier, *Green Chem.*, 2013, **15**, 1431–1455.
- 5 L. T. J. Korley, B. D. Pate, E. L. Thomas and P. T. Hammond, *Polymer*, 2006, **47**, 3073–3082.
- 6 A. Cornille, R. Auvergne, O. Figovsky, B. Boutevin and S. Caillol, *Eur. Polym. J.*, 2017, **87**, 535–552.
- 7 C. Carré, Y. Ecochard, S. Caillol and L. Avérous, *ChemSusChem*, 2019, **12**, 3410–3430.
- 8 A. Gomez-Lopez, F. Elizalde, I. Calvo and H. Sardon, *Chem. Commun.*, 2021, **57**, 12254–12265.
- 9 L. Maisonneuve, O. Lamazelle, E. Rix, E. Grau and H. Cramail, *Chem. Rev.*, 2015, **115**, 12407–12439.
- 10 M. S. Kathalewar, P. B. Joshi, A. S. Sabnis and V. C. Malshe, *RSC Adv.*, 2013, **3**, 4110–4129.
- 11 G. Rokicki, P. G. Parzuchowski and M. Mazurek, *Polym. Adv. Technol.*, 2015, **26**, 707–761.
- 12 N. Fanjul-Mosteirín, L. P. Fonseca, A. P. Dove and H. Sardon, *Mater. Adv.*, 2023, **4**, 2437–2448.
- 13 G. Seychal, C. Ocando, L. Bonnaud, J. De Winter, B. Grignard, C. Detrembleur, H. Sardon, N. Aramburu and J.-M. Raquez, *ACS Appl. Polym. Mater.*, 2023, **5**, 5567–5581.
- 14 C. Pronoitis, M. Hakkarainen and K. Odellius, *ACS Sustainable Chem. Eng.*, 2022, **10**, 2522–2531.
- 15 G. Hua and K. Odellius, *Macromol. Biosci.*, 2017, **17**, 1–9.
- 16 D. C. Aduba Jr., K. Zhang, A. Kanitkar, J. M. Sirrine, S. S. Verbridge and T. E. Long, *J. Appl. Polym. Sci.*, 2018, **135**, 1–11.
- 17 P. B. V. Scholten, J. Cai and R. T. Mathers, *Macromol. Rapid Commun.*, 2021, **42**, 1–3.
- 18 D. V. Palaskar, A. Boyer, E. Cloutet, C. Alfos and H. Cramail, *Biomacromolecules*, 2010, **11**, 1202–1211.
- 19 O. Kreye, S. Wald and M. A. R. Meier, *Adv. Synth. Catal.*, 2013, **355**, 81–86.
- 20 K. Zhang, A. M. Nelson, S. J. Talley, M. Chen, E. Margareta, A. G. Hudson, R. B. Moore and T. E. Long, *Green Chem.*, 2016, **18**, 4667–4681.
- 21 C. Wulf, M. Reckers, A. Perechodjuk and T. Werner, *ACS Sustainable Chem. Eng.*, 2020, **8**, 1651–1658.
- 22 R. H. Lambeth, *Polym. Int.*, 2021, **70**, 696–700.
- 23 J. Guan, Y. Song, Y. Lin, X. Yin, M. Zuo, Y. Zhao, X. Tao and Q. Zheng, *Ind. Eng. Chem. Res.*, 2011, **50**, 6517–6527.
- 24 G. F. Bass and T. H. Epps, III, *Polym. Chem.*, 2021, **12**, 4130–4158.
- 25 J. S. Mahajan, R. M. O'Dea, J. B. Norris, L. T. J. Korley and T. H. Epps, III, *ACS Sustainable Chem. Eng.*, 2020, **8**, 15072–15096.
- 26 R. M. Cywar, N. A. Rorrer, C. B. Hoyt, G. T. Beckham and E. Y. X. Chen, *Nat. Rev. Mater.*, 2022, **7**, 83–103.
- 27 R. H. Lambeth and T. J. Henderson, *Polymer*, 2013, **54**, 5568–5573.
- 28 Y. Peng, K. H. Nicastro, T. H. Epps, III and C. Wu, *Food Chem.*, 2021, **338**, 1–9.
- 29 Y. Peng, K. H. Nicastro, T. H. Epps, III and C. Wu, *J. Agric. Food Chem.*, 2018, **66**, 11775–11783.
- 30 S.-F. Koelewijn, D. Ruijten, L. Trullemans, T. Renders, P. Van Puyvelde, H. Witters and B. F. Sels, *Green Chem.*, 2019, **21**, 6622–6633.
- 31 W. Schutyser, T. Renders, S. Van den Bosch, S. F. Koelewijn, G. T. Beckham and B. F. Sels, *Chem. Soc. Rev.*, 2018, **47**, 852–908.
- 32 A. J. Ragauskas, G. T. Beckham, M. J. Bidy, R. Chandra, F. Chen, M. F. Davis, B. H. Davison, R. A. Dixon, P. Gilna, M. Keller, P. Langan, A. K. Naskar, J. N. Saddler, T. J. Tschaplinski, G. A. Tuskan and C. E. Wyman, *Science*, 2014, **344**, 1246843.
- 33 A. J. Shapiro, R. M. O'Dea, S. C. Li, J. C. Ajah, G. F. Bass and T. H. Epps, III, *Annu. Rev. Chem. Biomol. Eng.*, 2023, **14**, 109–140.
- 34 Z. Sun, B. Fridrich, A. de Santi, S. Elangovan and K. Barta, *Chem. Rev.*, 2018, **118**, 614–678.



- 35 F. M. Haque, J. S. A. Ishibashi, C. A. L. Lidston, H. Shao, F. S. Bates, A. B. Chang, G. W. Coates, C. J. Cramer, P. J. Dauenhauer, W. R. Dichtel, C. J. Ellison, E. A. Gormong, L. S. Hamachi, T. R. Hoye, M. Jin, J. A. Kalow, H. J. Kim, G. Kumar, C. J. LaSalle, S. Liffland, B. M. Lipinski, Y. Pang, R. Parveen, X. Peng, Y. Popowski, E. A. Prebihalo, Y. Reddi, T. M. Reineke, D. T. Sheppard, J. L. Swartz, W. B. Tolman, B. Vlasisavljevich, J. Wissinger, S. Xu and M. A. Hillmyer, *Chem. Rev.*, 2022, **122**, 6322–6373.
- 36 S. Zhao and M. M. Abu-Omar, *ACS Sustainable Chem. Eng.*, 2021, **9**, 1477–1493.
- 37 M. M. Abu-Omar, K. Barta, G. T. Beckham, J. S. Luterbacher, J. Ralph, R. Rinaldi, Y. Román-Leshkov, J. S. M. Samec, B. F. Sels and F. Wang, *Energy Environ. Sci.*, 2021, **14**, 262–292.
- 38 I. Ribca, B. Sochor, M. Betker, S. V. Roth, M. Lawoko, O. Sevastyanova, M. A. R. Meier and M. Johansson, *Eur. Polym. J.*, 2023, **194**, 112141.
- 39 M. Lawoko, L. Berglund and M. Johansson, *ACS Sustainable Chem. Eng.*, 2021, **9**, 5481–5485.
- 40 E. M. Maines, M. K. Porwal, C. J. Ellison and T. M. Reineke, *Green Chem.*, 2021, **23**, 6863–6897.
- 41 R. M. O'Dea, J. A. Willie and T. H. Epps, III, *ACS Macro Lett.*, 2020, **9**, 476–493.
- 42 C. Gioia, M. Colonna, A. Tagami, L. Medina, O. Sevastyanova, L. A. Berglund and M. Lawoko, *Biomacromolecules*, 2020, **21**, 1920–1928.
- 43 K. H. Nicastro, C. J. Kloxin and T. H. Epps, III, *ACS Sustainable Chem. Eng.*, 2018, **6**, 14812–14819.
- 44 T. H. Epps, III, L. T. J. Korley, M. D. Green, J. S. Mahajan and H. Behbahani, *U.S. Patent*, App. No. 17/616,245, 2022.
- 45 S. V. Mhatre, J. S. Mahajan, T. H. Epps, III and L. T. J. Korley, *Mater. Adv.*, 2023, **4**, 110–121.
- 46 K. H. S. Reno, J. F. Stanzione, III, R. P. Wool, J. M. Sadler, J. J. LaScala and E. D. Hernandez; *U.S. Pat.*, No. 10273684, 2020.
- 47 A. Amitrano, J. S. Mahajan, L. T. J. Korley and T. H. Epps, III, *RSC Adv.*, 2021, **11**, 22149–22158.
- 48 X. Zhang, J. S. Mahajan, L. T. J. Korley, T. H. Epps, III and C. Wu, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2023, **885**, 1–9.
- 49 E. K. Leitsch, G. Beniah, K. Liu, T. Lan, W. H. Heath, K. A. Scheidt and J. M. Torkelson, *ACS Macro Lett.*, 2016, **5**, 424–429.
- 50 S. Hu, X. Chen and J. M. Torkelson, *Polymer*, 2022, **256**, 1–10.
- 51 G. Beniah, D. J. Fortman, W. H. Heath, W. R. Dichtel and J. M. Torkelson, *Macromolecules*, 2017, **50**, 4425–4434.
- 52 V. Schimpf, B. S. Ritter, P. Weis, K. Parison and R. Mühlaupt, *Macromolecules*, 2017, **50**, 944–955.
- 53 M. Janvier, P.-H. Ducrot and F. Allais, *ACS Sustainable Chem. Eng.*, 2017, **5**, 8648–8656.
- 54 S. H. Lee and D. S. Lee, *Macromol. Res.*, 2019, **27**, 460–469.
- 55 M. A. R. Meier, *Macromol. Rapid Commun.*, 2019, **40**, 1–10.
- 56 V. Froidevaux, C. Negrell, S. Caillol, J.-P. Pascault and B. Boutevin, *Chem. Rev.*, 2016, **116**, 14181–14224.
- 57 M. Blain, A. Cornille, B. Boutevin, R. Auvergne, D. Benazet, B. Andrioletti and S. Caillol, *J. Appl. Polym. Sci.*, 2017, **134**, 1–13.
- 58 T. Wang, H. Deng, N. Li, F. Xie, H. Shi, M. Wu and C. Zhang, *Green Chem.*, 2022, **24**, 8355–8366.
- 59 R. S. Waletzko, L. T. J. Korley, B. D. Pate, E. L. Thomas and P. T. Hammond, *Macromolecules*, 2009, **42**, 2041–2053.
- 60 Q. Li, H. Zhou, D. A. Wicks, C. E. Hoyle, D. H. Magers and H. R. McAlexander, *Macromolecules*, 2009, **42**, 1824–1833.
- 61 I. S. Ryu, X. Liu, Y. Jin, J. Sun and Y. J. Lee, *RSC Adv.*, 2018, **8**, 23481–23488.
- 62 J. Mattia and P. Painter, *Macromolecules*, 2007, **40**, 1546–1554.
- 63 D. Wołosz and P. G. Parzuchowski, *Polymer*, 2022, **254**, 1–10.
- 64 P. Saha, L. Goswami and B. S. Kim, *ACS Sustainable Chem. Eng.*, 2022, **10**, 4623–4633.
- 65 L. Trullemans, S. F. Koelewijn, I. Scodeller, T. Hendrickx, P. Van Puyvelde and B. F. Sels, *Polym. Chem.*, 2021, **12**, 5870–5901.
- 66 I. Ribca, M. E. Jawerth, C. J. Brett, M. Lawoko, M. Schwartzkopf, A. Chumakov, S. V. Roth and M. Johansson, *ACS Sustainable Chem. Eng.*, 2021, **9**, 1692–1702.
- 67 X. Chen, X. Ouyang, J. Li and Y.-L. Zhao, *ACS Omega*, 2021, **6**, 22578–22588.
- 68 R. J. Young and P. A. Lovell, *Introduction to polymers*, CRC press, 3rd edn, 2011, pp. 1–688.
- 69 A. Shabbir, H. Goldansaz, O. Hassager, E. van Ruymbeke and N. J. Alvarez, *Macromolecules*, 2015, **48**, 5988–5996.
- 70 L. F. Wang, E. M. Pearce and T. K. Kwei, *J. Polym. Sci., Part B: Polym. Phys.*, 1991, **29**, 619–626.
- 71 A. L. Agapov, Y. Wang, K. Kunal, C. G. Robertson and A. P. Sokolov, *Macromolecules*, 2012, **45**, 8430–8437.
- 72 Q.-H. Zhou, M. Li, P. Yang and Y. Gu, *Macromol. Theory Simul.*, 2013, **22**, 107–114.
- 73 S. F. Koelewijn, C. Cooreman, T. Renders, C. Andecochea Saiz, S. Van den Bosch, W. Schutyser, W. De Leger, M. Smet, P. Van Puyvelde, H. Witters, B. Van der Bruggen and B. F. Sels, *Green Chem.*, 2018, **20**, 1050–1058.
- 74 S. Gupta, X. Yuan, T. C. Mike Chung, M. Cakmak and R. A. Weiss, *Polymer*, 2016, **107**, 223–232.
- 75 C. L. Lewis, K. Stewart and M. Anthamatten, *Macromolecules*, 2014, **47**, 729–740.
- 76 C. Osterwinter, C. Schubert, C. Tonhauser, D. Wilms, H. Frey and C. Friedrich, *Macromolecules*, 2015, **48**, 119–130.
- 77 R. Stadler and L. de Lucca Freitas, *Colloid Polym. Sci.*, 1986, **264**, 773–778.
- 78 M. Rubinstein and R. H. Colby, *Polymer Physics*, Oxford University, New-York, 1st edn, 2003, vol. 100, pp. 1–433.
- 79 X. Callies, C. Véchambre, C. Fonteneau, S. Pensec, J. M. Chenal, L. Chazeau, L. Bouteiller, G. Ducouret and C. Creton, *Macromolecules*, 2015, **48**, 7320–7326.
- 80 A. Jangizehi, M. Ahmadi and S. Seiffert, *J. Polym. Sci., Part B: Polym. Phys.*, 2019, **57**, 1209–1223.
- 81 M. Palusiak and S. J. Grabowski, *J. Mol. Struct.*, 2002, **642**, 97–104.
- 82 K. M. Hambleton and J. F. Stanzione, III, *ACS Omega*, 2021, **6**, 23855–23861.

