



Cite this: *Chem. Sci.*, 2023, **14**, 1742

All publication charges for this article have been paid for by the Royal Society of Chemistry

Rational design of mesoporous chiral MOFs as reactive pockets in nanochannels for enzyme-free identification of monosaccharide enantiomers†

Junli Guo,^a Xua Liu,^a Junjian Zhao,^a Huijie Xu,^a Zhida Gao,^a Zeng-Qiang Wu^{*b} and Yan-Yan Song   *

Monosaccharides play significant roles in daily metabolism in living organisms. Although various devices have been constructed for monosaccharide identification, most rely on the specificity of the natural enzyme. Herein, inspired by natural ionic channels, an asymmetrical MOF-in-nanochannel architecture is developed to discriminate monosaccharide enantiomers based on cascade reactions by combining oxidase-mimicking and Fenton-like catalysis in homochiral mesoporous CuMOF pockets. The identification performance is remarkably enhanced by the increased oxidase-mimicking activity of Au nanoparticles under a local surface plasmon resonance (LSPR) excitation. The apparent steady-state kinetic parameters and nano-fluidic simulation indicate that the different affinities induced by Au-LSPR excitation and the confinement effect from MOF pockets precipitate the high chiral sensitivity. This study offers a promising strategy for designing an enantiomer discrimination device and helps to gain insight into the origin of stereoselectivity in a natural enzyme.

Received 18th October 2022

Accepted 17th January 2023

DOI: 10.1039/d2sc05784k

rsc.li/chemical-science

Introduction

Chirality is essential in different fields, such as living matter, medical sciences, food chemistry, and drug manufacturing.¹ As a fundamental molecule, chiral monosaccharides play vital roles in the biological activity of all living organisms.^{2,3} Although monosaccharide enantiomers have similar taste quality,⁴ L-monosaccharides are non-transportable through living systems. D-Monosaccharides are the natural substrate for daily metabolism.⁴ Therefore, chiral recognition of monosaccharides has always been a hot topic in chemical and biological research. Compared with colorimetric⁵ and fluorescence methods,⁶ nanochannel-based enantiomer recognition techniques have shown great potential in discriminating monosaccharide enantiomers due to their distinct advantages of continuous operation, high sensitivity, and

selectivity.^{7–9} To date, many nanochannel-based discrimination systems have efficiently discriminated enantiomers by modifying nanochannels with natural enzymes as chiral receptors.^{10,11} Although these enzyme-modified nanochannels exhibit high selectivity for chiral monosaccharides, their fragile nature and the high cost of natural enzyme application limit their stability and practicability in harsh environments.

Metal-organic frameworks (MOFs) are widely recognized as promising building blocks for constructing catalytic systems due to their microporous structure and having plenty of functional groups.¹² In the past few years, several MOFs have demonstrated similar performance as biological enzymes.^{13–16} For example, Fe, Cu, Co, Ni, or Ce containing MOFs presented excellent enzyme-mimicking activity.¹⁷ In addition, due to the intrinsic frame structure and rich surface chemistry, MOFs are easily endowed with chiral properties, becoming one of the most promising hosts for selective adsorption and separation of racemic mixtures.¹⁸ However, the microporous structure of MOFs¹⁹ limits access to active sites, thus hindering their practical applications in this field. Many efforts have been made to prepare mesoporous MOFs to improve the accessibility and the catalytic efficiency of MOFs by introducing template molecules, adjusting the length of ligands, and treating MOFs with special solvents.^{20–22} However, due to complex signal transduction in chiral recognition using bulk MOF powders, the application of mesoporous MOFs for enantiomer discrimination has rarely been reported, despite the significant advantages of this approach.

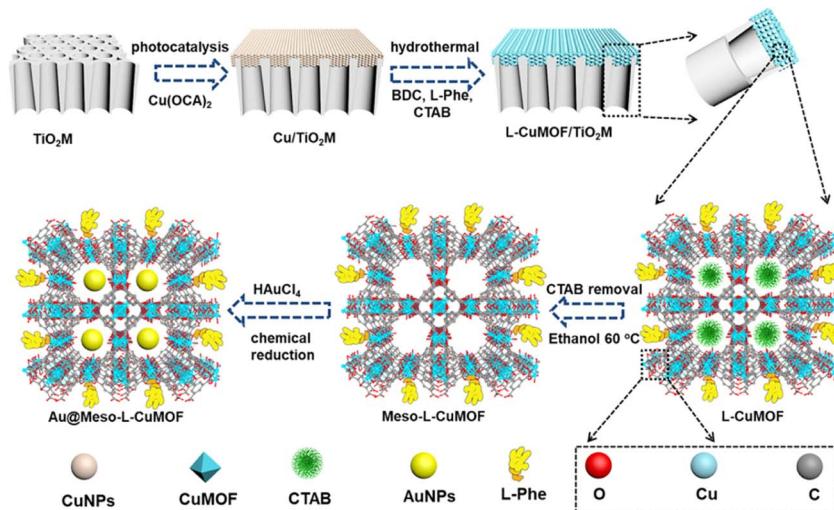
Inspired by mimicked biological channels, artificial ionic nanochannels have been applied in recent years in different fields, including energy conversion,²³ biosensing,²⁴ and others.²⁵

^aCollege of Sciences, Northeastern University, Shenyang 110819, China. E-mail: yysong@mail.neu.edu.cn

^bSchool of Public Health, Nantong University, Nantong, 226019, China. E-mail: zqwu@ntu.edu.cn

† Electronic supplementary information (ESI) available: The SEM images, FTIR, and XRD of TiO₂M, CuNPs/TiO₂M and Meso-L-CuMOF/TiO₂M; SEM images, TGA, N₂ sorption isotherms, UV-vis spectra and *I*-*V* properties of Au@Meso-L-CuMOF/TiO₂M; colorimetric assay of enzyme-mimicking; steady-state kinetic assay and kinetic parameters of Au@Meso-L-CuMOF/TiO₂M; infrared thermal imaging photographs; *I*-*V* curves and the corresponding calibration plots for different concentrations of L/D-glucose; *I*-*V* curves for sensing different chiral molecules and interfering species; *I*-*V* curves for sensing D-glucose in serum samples; zeta potentials of each step modification; comparison of various methods for L/D-glucose detection; computer domain and boundaries of FEM models; boundary conditions of FEM models; simulation concentration profiles of different ions and molecules. See DOI: <https://doi.org/10.1039/d2sc05784k>





Scheme 1 Schematic illustration of preparing Au@Meso-L-CuMOF/TiO₂M from TiO₂M.

Compared to individual material-based channels, asymmetric artificial nanochannels with different chemical compositions along the channels exhibit remarkably novel functions and properties.²⁶ Herein, asymmetrically grown MOF structures in a free-standing anodic TiO₂ nanochannel membrane (TiO₂M) featuring excellent photocatalytic activity and chemical stability (Scheme 1) are proposed for highly selective discrimination of monosaccharide enantiomers *via* mesoporous homochiral CuMOF (Meso-L-CuMOF) and artificial oxidase. In particular, an interfacial growth method to achieve asymmetrical MOF decoration by utilizing the catalytic activity of TiO₂ is reported for the first time. Gold nanoparticles (AuNPs), a kind of artificial oxidase, are filled in Meso-L-CuMOF pockets through an *in situ* reduction approach. This nanochannel-based monosaccharide sensing device eliminates the requirement for natural enzymes and is economical and stable. More importantly, experimental results and numerical simulations indicate that a better-localized surface plasmon resonance (LSPR) mediated ionic product conversion is achieved owing to the nanoconfinement effect of the Meso-L-CuMOF membrane, resulting in a stereoselective and substrate-selective robust sensing device. This study offers inspiration to design and prepare low-cost and stable asymmetrical nanochannels for monosaccharide enantiomer discrimination.

Results and discussion

Scheme 1 schematically illustrates the procedure for asymmetrical growth of Meso-L-CuMOF pockets and oxidase-mimicking Au NPs in TiO₂M. Benefiting from the homochiral environment of Meso-L-CuMOF, the constructed nanochannels show different affinities for glucose enantiomers. This distinct affinity enables one of the enantiomers to be transported through mesoporous MOFs and further oxidized by oxidase-mimicking AuNPs. The generated H₂O₂ is *in situ* monitored by the *I-V* properties of nanochannels utilizing the Fenton-like activity of a CuMOF. TiO₂M prepared by electrochemical anodization (details are presented in the Experimental section) is amorphous.²⁷ A two-hour annealing (450 °C in

air) burns the remaining electrolyte and synchronously transforms amorphous TiO₂M to anatase crystalline.²⁸ Fig. 1A shows the scanning electron microscopy (SEM) images of the as-formed TiO₂M, composed of ~48 μm thick parallel nanochannels (inset of Fig. 1A). Two entrances of nanochannels have different diameters, showing ~150 ± 20 nm (denoted as the base entrance, Fig. 1A) at one side and ~40 ± 10 nm at the other (denoted as the tip entrance, Fig. 1B). To achieve an asymmetrical decorated CuMOF for chiral sensing, CuNPs are first coated onto the tip entrance by utilizing the photocatalytic activity of TiO₂ nanochannels and interfacial growth strategy (details are presented in the Experimental section).²⁹ Fig. 1C shows that a high-density layer of CuNPs is distributed on the tip entrance and nanochannel walls. These CuNPs are then transferred to homochiral MOFs exhibiting a mesoporous structure (Meso-L-CuMOF) by a typical template-assisted synthesis method (using BDC as an organic ligand, L-Phe as a chiral environment, and CTAB as a template).³⁰ As shown in Fig. 1D and F, a layer of compact Meso-L-CuMOF nanocrystals appears on the tip entrance of TiO₂M with a thickness of ~1.5 μm (further morphology details are presented in Fig. S1†).

The sample's circular dichroism (CD) spectrum is recorded and compared with that of the model prepared without L-Phe (named Meso-CuMOF/TiO₂M) to identify the homochiral character of the resulting MOF layer. Fig. 2A shows an adsorption peak at 210 nm on the Meso-L-CuMOF/TiO₂M CD spectrum, indicating the homochirality feature. The liquid ¹H NMR spectrum of Meso-L-CuMOF/TiO₂M also demonstrates the successful loading of L-Phe into the Meso-CuMOF (Fig. 2B). The mass percentage (9.6 wt%) of L-Phe in the membrane is determined by thermogravimetric analysis (TGA, Fig. S2†). Compared with the sample containing the CTAB templates (named L-CuMOF/TiO₂M), the content of mesopores in Meso-L-CuMOF/TiO₂M increases as indicated by the enhanced intensity at a pore size of ~7.5 nm (Fig. 2C), confirming the presence of a mesoporous structure in MOF layers. Meanwhile, the presence of mesopores is also demonstrated by Brunauer–Emmett–Teller (BET) analysis (Fig. S3†). The obtained BET areas are 406 m² g⁻¹ and 268 m² g⁻¹ for L-CuMOF/TiO₂M and Meso-L-

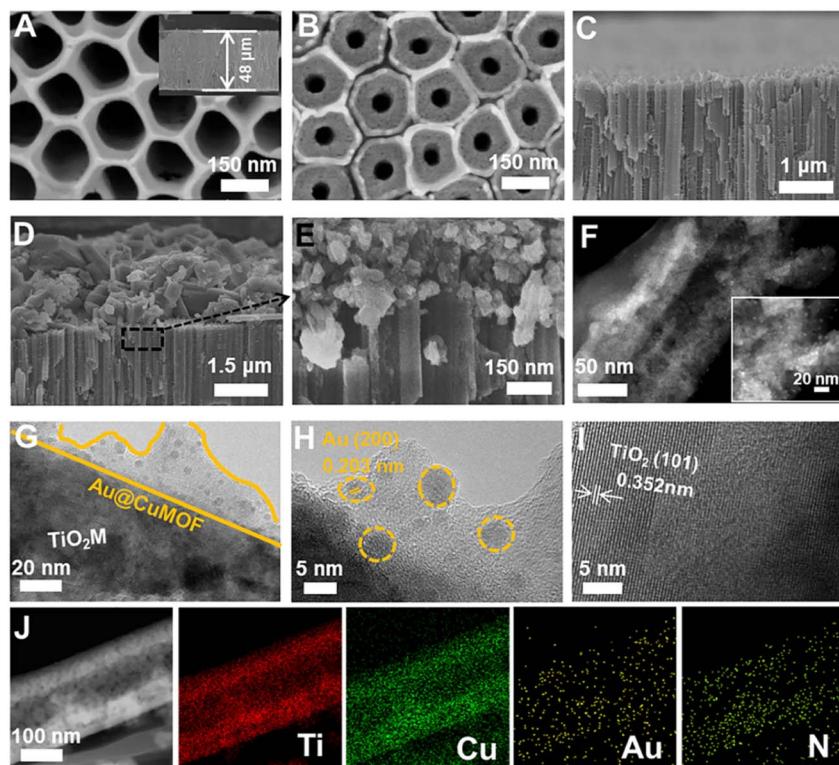


Fig. 1 Schematic illustration of preparing Au@Meso-L-CuMOF/TiO₂M. SEM images: (A) top view, (B) bottom view, and (C) side view of TiO₂M. (D and E) Side view of Meso-L-CuMOF/TiO₂M. (F) HAADF-STEM, (G) TEM and (H and I) HRTEM images of Au@Meso-L-CuMOF/TiO₂M. (J) Elemental mapping of Au@Meso-L-CuMOF/TiO₂M.

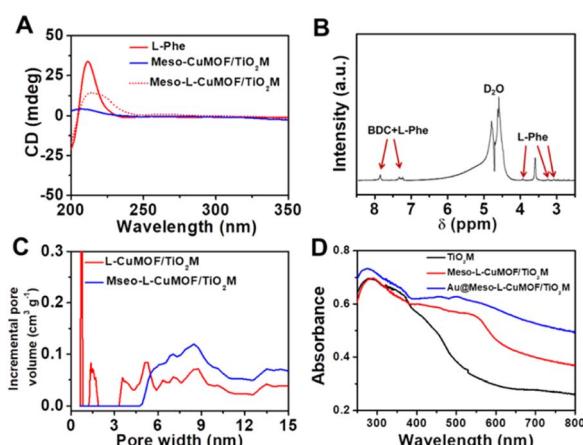


Fig. 2 (A) CD spectra of Meso-L-CuMOF/TiO₂M, L-Phe, and Meso-L-CuMOF/TiO₂M. (B) Liquid ¹H NMR spectrum of Meso-L-CuMOF/TiO₂M digested in D₂O. (C) Pore size distribution of Meso-L-CuMOF/TiO₂M before and after CTAB removal. (D) Absorption spectra of TiO₂M, Meso-L-CuMOF/TiO₂M, and Au@Meso-L-CuMOF/TiO₂M.

CuMOF/TiO₂M, respectively.³¹ The reduced BET area can be ascribed to the lost micropores. The accommodation space of mesopores provides a favorable feature for further functionalization. To further confirm the successful removal of CTAB, the Raman and energy dispersive X-ray spectroscopy (EDS) analysis of L-CuMOF/TiO₂M and Meso-L-CuMOF/TiO₂M were performed. Fig. S4† shows the Raman spectra. The disappearance of CH₃ (from the

(CH₃)₃N⁺ group in CTAB) rocking bands at 758 and 1526 cm⁻¹ implies the successful removal of CTAB.³² Meanwhile, the removal of CTAB is also demonstrated by EDS analysis (Fig. S5†). The reduced N and C elements can be ascribed to the lost CTAB molecules. In this work, the well-known oxidase-mimicking AuNPs are integrated into Meso-L-CuMOF *via* a typical AuCl₄⁻ adsorption-NaBH₄ reduction method (Scheme 1).³³ As indicated by transmission electron microscopy (TEM) images, the nanochannel wall is coated with a MOF layer (Fig. S6†). As shown in Fig. 1F, the high-angle annular dark-field scanning TEM (HAADF-STEM) image shows that AuNPs are well dispersed on Meso-L-CuMOF/TiO₂M, and the diameter of AuNPs is 5 ± 2 nm (Fig. 1G). The high-resolution TEM (HRTEM) images in Fig. 1H and I exhibit the characteristic lattice spacing for the (200) plane of AuNPs (JCPDS no. 4-784) and the (101) plane of anatase TiO₂ (JCPDS no. 21-1272). The EDS mapping images reveal the well-dispersed Au, Cu, and N elements on channel walls (Fig. 1I). As the N resource only stems from the -NH₂ group in L-Phe, this result also verifies the successful fabrication of Au@Meso-L-CuMOF in TiO₂M. An increased absorption peak at ~400–700 nm is observed in the UV-vis absorption spectra (Fig. 2D), assigned to the optical absorption of scattered light by the LSPR effect of AuNPs,³⁴ indicating an improved light-harvesting ability in the visible-light region. To further demonstrate successful formation of the AuNPs with an according LSPR, we peeled off AuNPs from the Au@Meso-L-CuMOF/TiO₂M sample by dissolving the Meso-L-CuMOF/TiO₂M substrate in a HF solution. The resulting solution shows a pink



color with an absorption peak at ~ 530 nm typical for the SPR of AuNPs (Fig. S7†). In addition, the whole fabrication process is characterized by Fourier transform infrared (FTIR) spectra (Fig. S8†) and X-ray diffraction (XRD) patterns (Fig. S9†). The absorption bands at ~ 3065 , 1670, 1265, and 1050 cm^{-1} in the FTIR spectra coincide with those of bulk Meso-L-CuMOF powder. The adsorption band at 1265 cm^{-1} corresponds to the C–N stretching of L-Phe. In the XRD patterns, the appearance of Meso-L-CuMOF peaks at 7.5° , and the disappearance of CuNP peaks at 42.5° and 43.5° also implies the transfer of CuNPs to Meso-L-CuMOF. The characteristic diffraction peak at 44.6° is attributed to the Au (200) lattice place, confirming the successful introduction of Au.

A target-discrimination-induced signal-on sensing strategy is designed by integrating the cascade reactions catalyzed by oxidase-mimicking AuNPs and the subsequent Fenton-like CuMOF to achieve the recognition and quantification of glucose enantiomers in the as-proposed membrane. As illustrated in Fig. 3A, the glucose enantiomers selectively recognized on homochiral MOFs are transported into the mesopores and then oxidized to gluconic acid and H_2O_2 via oxidase-mimicking AuNPs.^{35,36} ABTS, one of the widely applied probes for evaluating $\cdot\text{OH}$ radicals, is employed as the signal reporter to provide a noticeable ionic current change. Owing to the Fenton-like performance of Cu(II),¹⁷ the CuMOF catalyzes the decomposition of H_2O_2 molecules into $\cdot\text{OH}$ radicals, further oxidizing ABTS to ABTS⁺. In addition, the Fenton-like activity of Cu(II) has been demonstrated to rely on the solution pH.¹⁷ The decrease in ambient pH caused by gluconic acid generated in the glucose oxidation reaction is thus favorable for activating the Fenton-like activity of the CuMOF. The generated ABTS⁺ cations change the ionic flux, as illustrated in Fig. 3B. The I – V properties of nanochannels thus offer sensitive signal transduction, indicating the corresponding details for enantiomer recognition and quantification. A traditional colorimetric assay studies the feasibility of the above hypothesis essay based on the characteristic blue color and the typical absorption peak of ABTS⁺

at 400–900 nm (Fig. S10†). By adding H_2O_2 , the apparent blue color and characteristic absorption peak of ABTS⁺ are observed in Meso-L-CuMOF/TiO₂M (Fig. S10A†), verifying the Fenton-like performance of the CuMOF. In addition, when replacing H_2O_2 with D-glucose, the formation of ABTS⁺ is only found in Au@Meso-L-CuMOF/TiO₂M (Fig. S10B†), verifying the oxidase-mimicking activity of the decorated Au NPs. Noticeably, the absorbance intensity of ABTS⁺ is discriminatory in the presence of glucose enantiomers (Fig. S11†). D-Glucose induces more ABTS⁺ generation than L-glucose at the same concentration, suggesting a higher affinity between Au@Meso-L-CuMOF/TiO₂M and D-glucose.

Encouraged by the difference in oxidation ability of nanochannels towards glucose enantiomers and their ionic transport feature, the enantiomer recognition and glucose oxidation kinetics of Au@Meso-L-CuMOF/TiO₂M are investigated by using the ionic transmembrane currents (I – V curves). The as-prepared membrane is in the middle of the “H-type” cell, as illustrated in Fig. 3B. Two Ag/AgCl electrodes are inserted into two-half cells containing the same electrolyte (1.0 μM KCl containing 1.0 mM ABTS). The ionic transmembrane currents are recorded on TiO₂M, Meso-L-CuMOF/TiO₂M, and Au@Meso-L-CuMOF/TiO₂M (Fig. 3C). The current enhancement is observed in Au@Meso-L-CuMOF/TiO₂M. The increased ionic flux indicates the formation of ABTS⁺ ions, and the ionic current results coincide with the results of the colorimetric assays (Fig. S10B†).

The Fenton or Fenton-like process decomposes H_2O_2 molecules and generates $\cdot\text{OH}$ radicals. In our design, as the yield of ABTS⁺ ions is directly decided by the ionic flux (sensing performance), a larger amount of $\cdot\text{OH}$ radicals is favorable. Recent reports show that the excited LSPR state of a plasmonic metal (mainly Au, Ag, and Pt) effectively facilitates the generation of hot electrons, subsequently transferred from the plasmonic metal to adsorbed reactants, thus elevating the enzyme-mimicking catalytic performance.^{37–39} The glucose oxidation in Au@Meso-L-CuMOF/TiO₂M nanochannels is investigated under visible-light irradiation (532 nm laser). When exposed to light, an apparent increase in ionic current appears (Fig. 3D, red line). The increased oxidase-mimicking catalytic activity of Au@Meso-L-CuMOF/TiO₂M under light irradiation is ascribed to the improved light-scattering ability of Au-LSPR (Fig. 3D and S12†). The colorimetric assays are further investigated at 25 and 60 °C to exclude the thermal effect on catalytic activity under laser irradiation (Fig. S13†). Although the ABTS⁺ products generated at 60 °C are slightly larger than that at 25 °C, higher ABTS⁺ yields are observed for both colorimetric assays when systems are exposed to a 532 nm laser. These results confirm that the enhanced ionic current mainly stems from the plasmon-enhanced oxidase-mimicking activity.

Considering the ionic current variation in the oxidase-mimicking activity of AuNPs and the Fenton-like activity of CuMOFs, the LSPR-enhanced catalytic activity of Au@Meso-L-CuMOF/TiO₂M is explored from these two aspects. The reaction kinetics of glucose oxidation is further determined in the presence and absence of a 532 nm laser (Fig. S14A and B†) to gain insight into the LSPR effect on the oxidase-mimicking activity of AuNPs. Fig. 4A and S14C† depict the corresponding Lineweaver–Burk plots towards different concentrations of D-glucose and ABTS, respectively. Upon LSPR excitation, the catalytic efficiency ($K_{\text{cat}}/K_{\text{m}}$) is

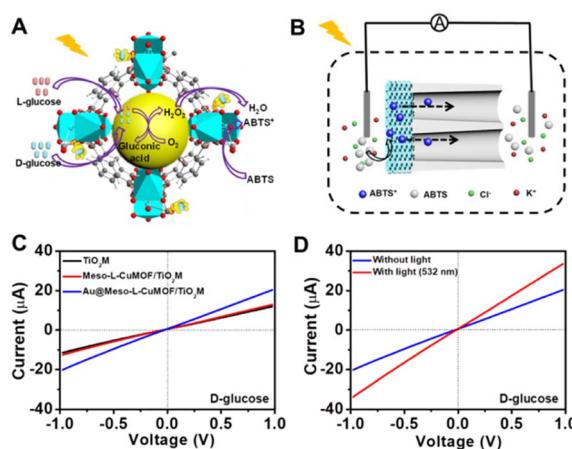


Fig. 3 (A) Schematic illustration of enantioselective glucose detection based on cascade reactions. (B) Schematic setup for enantioselective glucose detection based on cascade reactions. (C) I – V curves of TiO₂M, Meso-L-CuMOF/TiO₂M, and Au@Meso-L-CuMOF/TiO₂M in 10 μM D-glucose. (D) I – V curves of Au@Meso-L-CuMOF/TiO₂M for 10 μM D-glucose with or without 532 nm laser irradiation.



increased 1.82 and 3.07 times for D-glucose and ABTS, respectively (Table S1†), confirming the LSPR-enhanced oxidase-mimicking activity. The plasmon-enhanced Fenton-like activity of Meso-L-CuMOF is also investigated. The reaction kinetics for H₂O₂ decomposition by Fenton-like catalysis is also evaluated (Fig. S15†). The steady-state kinetic parameters of the Fenton-like activity of Meso-L-CuMOF/TiO₂M for H₂O₂ are shown in Fig. 4B, suggesting an effective reaction rate under irradiation. Upon LSPR excitation, the catalytic efficiency increases by 1.74 and 1.55 for ABTS and H₂O₂, respectively (Table S2†), demonstrating the LSPR-enhanced Fenton-like activity. These enhancements are attributed to the oscillating energetic hot spots (including “hot holes” and “hot electrons”) generated from LSPR-induced charge separation on a plasmonic metal surface. The hot holes are driven to the AuNP surface by the local electromagnetic fields and then participate in the glucose oxidation,^{38–40} thus leading to the enhanced production of H₂O₂, as shown in Fig. 4C.

Due to energy matching, the energetic hot electrons are injected into the conduction band (CB) of CuMOFs, to convert Cu(II) nodes to Cu(I). The outstanding Fenton-like activity of Cu(I) also accelerates H₂O₂ cleavage into ·OH radicals,⁴¹ thus achieving an improved ABTS yield. In addition, the Fenton-like activity at different temperatures is also carried out to show that the increased surface temperature does not induce enhanced activity under laser irradiation (the sample temperature exhibits a ~7 °C enhancement after 30 min of 532 nm laser irradiation, Fig. S16†). Comparing the colorimetric assays (Fig. S17†) and I–V properties (Fig. S18†) at 25 °C and 60 °C, the heating influence on Fenton-like performance of Meso-L-CuMOF/TiO₂M is still limited, implying that the increased ABTS⁺ yield is mainly attributed to the formation of Cu(I). To further confirm the increased ionic transport

performance result after hot injection, the I–V properties of Au@Meso-L-CuMOF/TiO₂M at 32 °C (increased by 7 °C from 25 °C) and 67 °C (increased by 7 °C from 60 °C) were tested and compared with the I–V curves measured after 532 nm-laser irradiation. As shown in Fig. S19A,† a slight current enhancement is observed when the temperature increased from 25 °C to 32 °C. When exposed to light (523 nm laser), a more apparent enhancement in ionic current is observed (Fig. S19A†). As shown in Fig. S19B,† the ionic current enhancement is ignored as the temperature increased from 60 °C to 67 °C, which can be attributed to the stable GOD-like activity of AuNPs at higher temperature.⁴² In contrast, an apparent increase in ionic current appears when exposed to light (Fig. S19B†). The above results demonstrate that the increased ionic transmembrane current is related to the hot injection and increased temperature at a lower temperature, and is mainly caused by hot injection at a higher temperature. These results indicate that the formed energetic charge carriers improve oxidase-mimicking and Fenton-like activity upon LSPR excitation.

Besides the dielectric constant of the plasmonic metal, the LSPR intensity of plasmonic nanoparticles is related to many factors, such as the particles' size, shape, and density, as well as the dielectric environment.^{43,44} To gain additional evidence of energetic hot spot generation by LSPR of AuNPs in the Meso-L-CuMOF, the finite difference time domain (FDTD) method is employed to investigate the field strength distribution in the Au@Meso-L-CuMOF structure, based on the TEM images in Fig. S6† (the permittivity of gold used in the simulation is obtained from Palik's data⁴⁵). Besides the plasmonic enhancement in the nanogap between two Au nanoparticles, LSPR intensity shows an apparent increase at the interface between AuNPs and the Meso-L-CuMOF substrate (Fig. 4D). Moreover, according to a recent report by Beckham⁴⁶ and Tiefenbacher,⁴⁷ the nanocapacitance effect of supramolecular hosts also facilitates the dual activation of both a nucleophilic (*i.e.*, glucose oxidation) and an electrophilic (*i.e.*, H₂O₂ cleavage) reaction in isolated binding pockets (*i.e.*, the well-aligned mesoporous MOFs). These results indicate that the dielectric CuMOF substrate and the mesoporous frame structure favor the energetic charge carrier generation process.

Encouraged by the remarkable catalytic activity of Au@Meso-L-CuMOF/TiO₂M for glucose oxidation and the homochiral character of nanochannels, the I–V properties of the resulting membrane are applied to discriminate between glucose enantiomers. The transmembrane ionic conductances are investigated at different KCl concentrations to evaluate the ionic transport performance of Au@Meso-L-CuMOF/TiO₂M (Fig. S20†). The ionic conductance is mainly attributed to the bulk when the concentrations exceed 1.0 mM, as presented in Fig. 5A. When the concentrations are lower than 1.0 mM, the ionic conductance deviates from the bulk value, attributed to the electric double layer (EDL)-governed ionic transport in mesopores at low concentrations.^{48,49} To enable the ion current which mainly stems from ABTS⁺ ions, a low-concentrated KCl (1.0 μM) solution is chosen for L/D-glucose recognition and detection. For high detection sensitivity, the operating parameters, such as the concentration of ABTS, reaction temperature, and pH, are optimized through I–V curves of Au@Meso-L-CuMOF/TiO₂M under light irradiation. The

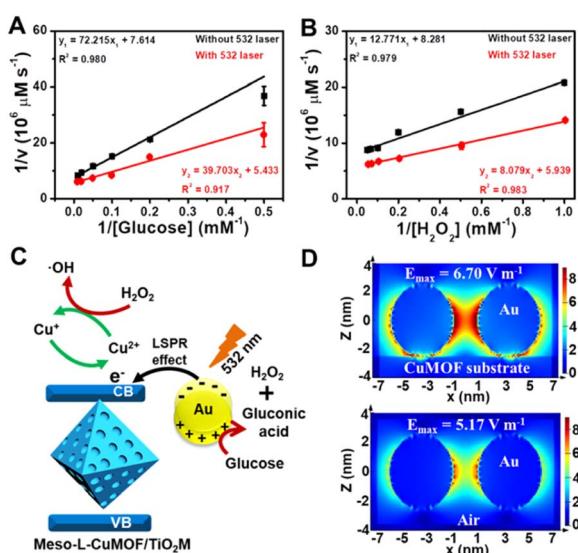


Fig. 4 Lineweaver–Burk plot of Au@Meso-L-CuMOF/TiO₂M for the changes in (A) glucose and (B) H₂O₂ in the presence and absence of 532 nm irradiation. (C) Schematic illustration of surface plasmon-enhanced nanoenzyme activity under 532 nm irradiation. (D) The intensity of electric field distribution in the monitor window around the hotspot on Au/CuMOF and Au NPs.



ABTS concentration is optimized as 1.0 mM based on the change in the *I*-*V* curves (Fig. S21†). The reaction temperature is studied from 25–80 °C, and the highest catalytic activity is obtained at 60 °C (Fig. S22†). The pH effect on cascade reactions is also evaluated based on the *I*-*V* curves (Fig. S23†). A burst ABTS production is found at pH 5.0. Accordingly, *I*-*V* measurements in the following study are performed in KCl solution (1.0 μM, pH 5) containing 1.0 mM ABTS at 60 °C. However, considering high temperatures largely influence the hydrogen bonds formed in the enantiomeric identification process,⁵⁰ recognition is achieved at 4 °C for 4 h to gain a saturated adsorption state (Fig. S24†). The nanochannels exhibit negligible catalytic activity at 4 °C, and recognition at 4 °C reaches a saturated state after 4 h. Therefore, before triggering the glucose oxidation reaction at 60 °C, enantiomer recognition is performed at 4 °C for 4 h. The discrimination abilities of Au@Meso-L-CuMOF/TiO₂M and Au@Meso-CuMOF/TiO₂M for 10 μM of glucose enantiomers are evaluated through *I*-*V* curves under optimal conditions. Fig. S25A† shows the *I*-*V* curves of Au@Meso-L-CuMOF/TiO₂M after cascade reactions catalyzed by enzymatic-mimicking for 10 μM L-glucose and 10 μM D-glucose. Apparently, the ionic current elevation induced by 10 μM D-glucose is larger than that of 10 μM L-glucose. For comparison, the *I*-*V* curves of Au@Meso-CuMOF/TiO₂M were also recorded (Fig. S25B†), which showed a similar ionic current elevation for the recognition of L/D-glucose. The above results demonstrate that the enantioselectivity stems from the chiral environment induced by L-CuMOF. To further confirm this conclusion, we also employed colorimetric assay to investigate the effect of Au NPs on selectivity. The oxidation ability of Au@Meso-CuMOF/TiO₂M to L/D-glucose can be indicated by the color change of ABTS solution (the generated H₂O₂ would oxidize ABTS to form blue colored ABTS⁺). As shown in Fig. S25C and D,† the results are consistent with the *I*-*V* curves. The L-Phe molecules act as enantioselective identifiers to interact with D-glucose than with L-glucose *via* hydrogen-bonding interactions.⁵¹ The discrimination abilities of Au@Meso-L-CuMOF/TiO₂M for different concentration of glucose enantiomers are evaluated

through *I*-*V* curves under optimal conditions (Fig. S26 and S27†). Fig. 5B and C plot the *I*-*V* curves of different concentrations of D-glucose and L-glucose at a 30 min-reaction period at 60 °C, respectively. The ionic currents gradually increase with glucose concentration from 0.1 to 10 μM. The ionic current elevation induced by D-glucose is larger than that of L-glucose from the ionic current change (ΔI) at different glucose concentrations (Fig. S28†). The calibration curves show that the resulting Au@Meso-L-CuMOF/TiO₂M has an excellent linear response to D-glucose sensing over two ranges: from 0.1 μM to 1 μM and from 1.0 μM to 10 μM. The limit of detection (LOD) is 0.089 μM, based on a 3 SD/L method. Compared with other reported recognition methods (Table S3†), this strategy exhibits a better LOD value, comparable with the recently reported colorimetric assay,⁵² CD spectra,⁵³ and electrochemical methods.⁵⁴

Application versatility and enantioselectivity are essential for evaluating a chiral recognition platform. The chiral sensing possibility of Au@Meso-L-CuMOF/TiO₂M for monosaccharide enantiomers is estimated (Fig. S29†). Fig. 5D plots the ionic current changes of the membrane for L/D-mannose, L/D-xylose, L/D-tagatose, L/D-ribose and L/D-galactose discrimination. The glucose enantiomers show a more significant relative ionic current variation, attributed to the substrate selectivity of oxidase-mimicking AuNPs, also discovered in a previous report.⁵⁵ Fig. 5E further presents the normalized ionic current changes. The chiral sensing device exhibits an excellent stereochemically controlled reaction process; D-monosaccharides show a more significant affinity for chiral MOFs.

Considering the promising applications in biological samples, the main interfering species co-existing with D-glucose in human blood serum are investigated (Fig. S30†). The ionic current changes induced by these interferences are ignored under the same experimental conditions as those of D-glucose sensing. Encouraged by the above results, the detection of D-glucose is further investigated in serum samples (Fig. 5F and S31†). The glucose concentrations determined from ΔI values (Table S4†)

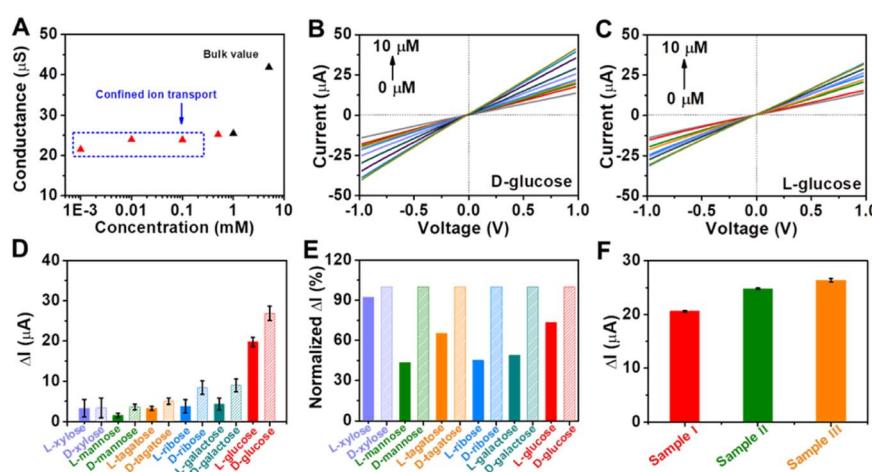


Fig. 5 (A) Ionic conductance of the membrane as a function of KCl concentrations. *I*-*V* curves for sensing different concentrations of (B) D-glucose and (C) L-glucose. (D) Ionic and (E) normalized ionic current changes at +1.0 V for monosaccharide enantiomers (10 μM). (F) Detection of D-glucose in serum samples. The electrolyte contains 1.0 mM ABTS and 1.0 μM KCl (pH 5.0).



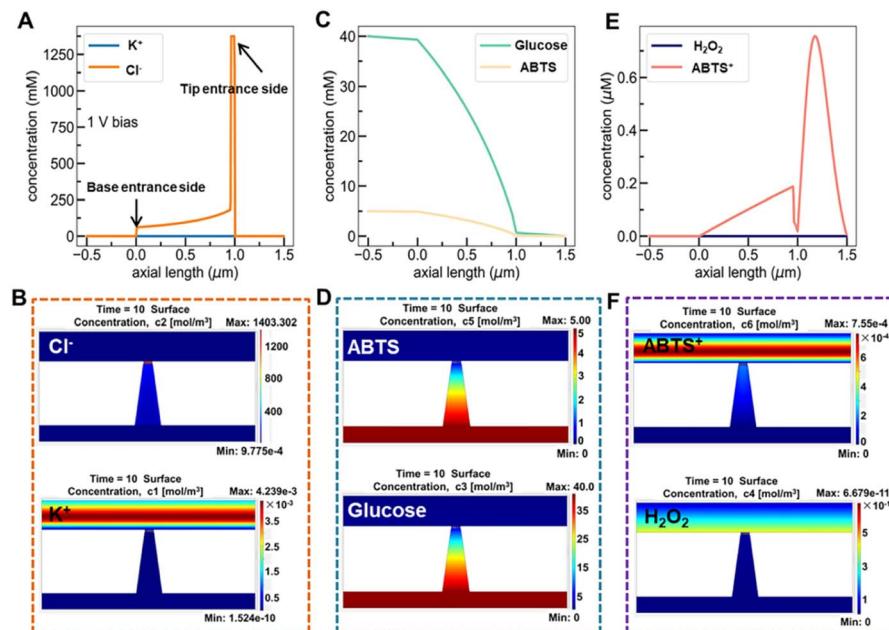


Fig. 6 (A) Simulation concentration profiles of KCl along a nanochannel without an enzyme catalytic reaction. (B) Simulated ionic concentration images of Cl⁻ and K⁺ ions in a nanochannel without an enzyme catalytic reaction. (C) Simulation concentration profiles of glucose and ABTS along the nanochannel. (D) Simulated ionic concentration images of glucose and ABTS in a nanochannel. (E) Simulation concentration profiles of H₂O₂ and ABTS⁺ along the nanochannel. (F) Simulated ionic concentration images of H₂O₂ and ABTS⁺ in a nanochannel. The transmembrane voltage is set as +1.0 V.

show satisfactory recovery with a low relative standard deviation (RSD), consistent with the hospital results obtained by a classic glucose oxidase-peroxidase method.

The finite element method (FEM) simulation is further applied to gain insight into the reactant and ionic product distribution in nanochannels to understand the mechanism of the high sensitivity of the proposed sensing system. A theoretical model (Fig. S32 and Table S5†) based on FEM combined with Poisson and Nernst–Planck (PNP) equations^{56,57} (details are provided in the ESI†) is employed to simulate the glucose sensing process in Au@Meso-L-CuMOF. Upon LSPR excitation, AuNPs catalyze glucose oxidation to H₂O₂ and gluconic acid. The reaction kinetics, eqn (1) and (2), can be formulated according to the Michaelis–Menten equation.

$$\frac{[\text{Glucose}]}{dt} = -\frac{V_{m,\text{glucose}}[\text{glucose}]}{(k_{m,\text{glucose}} + [\text{glucose}])} \quad (1)$$

$$\frac{[\text{H}_2\text{O}_2]}{dt} = \frac{V_{m,\text{glucose}}[\text{glucose}]}{(k_{m,\text{glucose}} + [\text{glucose}])} \quad (2)$$

The reaction intermediate H₂O₂ is subsequently cleaved to ·OH radicals by CuMOF with the aid of hot electrons. The kinetics are expressed as follows:

$$\frac{[\text{ABTS}^+]}{dt} = -\frac{V_{m,\text{H}_2\text{O}_2}[\text{ABTS}] + V_{m,\text{ABTS}}[\text{H}_2\text{O}_2]}{(k_{m,\text{H}_2\text{O}_2}[\text{ABTS}] + k_{m,\text{ABTS}}[\text{H}_2\text{O}_2] + k_{m,\text{H}_2\text{O}_2}k_{m,\text{ABTS}})} \quad (3)$$

Fig. 6A and B show the simulated distribution of K⁺ and Cl⁻ concentration profiles and the corresponding ionic flux images (at +1.0 V) on the Au@Meso-L-CuMOF/TiO₂M surface in the absence of glucose. Since the Au@Meso-L-CuMOF/TiO₂M zone carries positive surface charges (Fig. S33†), the strong electrostatic interactions attract more Cl⁻ ions into the nanochannels, thus accumulating Cl⁻ ions in Au@Meso-L-CuMOF/TiO₂M. The concentration profiles (Fig. 6C) and corresponding ionic flux images (Fig. 6D) of glucose and ABTS along the nanochannel are also simulated before the reaction. The electroneutral feature of both substrates and the electrical field does not influence their diffusion behaviors. The concentration profiles and corresponding ionic flux images of H₂O₂ and ABTS⁺ along the nanochannel after glucose oxidation are also simulated (Fig. 6E and F). Due to the low affinity between glucose and CuMOF pockets (high K_m constant, compared to other K_m), the generated H₂O₂ molecule oxidizes ABTS to ABTS⁺ using a CuMOF catalyst. Since ABTS⁺ ions carry a positive charge, they are depleted from mesoporous MOF pockets under solid electrostatic repulsion, increasing ABTS⁺ ion diffusion current. The ion/molecule concentration distribution on the Au@Meso-L-CuMOF/TiO₂M surface is also investigated at -1.0 V (Fig. S34†), showing similar results as the distribution at +1.0 V.

Conclusions

In summary, a highly enantioselective and sensitive platform for monosaccharide sensing is designed by the asymmetric assembly of Au@Meso-L-CuMOFs on TiO₂M. Benefiting from the dual catalytic activity and homochiral environment, the ionic



transmembrane currents of nanochannels provide excellent discrimination for monosaccharide enantiomers. Owing to the confinement effect, the primarily generated energetic “hot holes” and “hot electrons” under LSPR excitation in the nanoscaled MOF pocket significantly accelerated the sensing performance. The photocatalysis-triggered asymmetrical decoration approach can be easily extended to preparing many MOF membranes on porous TiO_2 substrates. This work provides a strategy to acquire and improve the signal for enantioselective recognition.

Experimental

Preparation of CuNPs/ TiO_2 M

Cu/TiO_2 M was prepared in a self-made H-type cell. The membrane was placed between two cells of a homemade electrolyte cell, and a quartz window was set on one side of the cell (top side of TiO_2 M) to allow UV light to pass through and reach the TiO_2 M surface. One half of the cell (bottom side of the TiO_2 M membrane) was filled with 5.0 mM $\text{Cu}(\text{CH}_3\text{COO})_2 \cdot \text{H}_2\text{O}$. Another half of the cell (top side of TiO_2 M) was filled with pure water. At +1.0 V for 120 min, the CuNPs migrated and were then deposited on the top side of the nanochannel, which was exposed to UV light (3 W LED, 365 nm).

Preparation of Au@Meso-L-CuMOF/ TiO_2 M

First, 11.6 mg BDC was dissolved in a 10 mL mixture of DMF and CH_3OH ($V_{\text{DMF}}:V_{\text{CH}_3\text{OH}} = 1:1$). Known amounts of surfactant CTAB (20 mg) and the chelating agent L-Phe (4.3 mg) were added, and the mixture was sonicated for 30 min to form a homogeneous solution. The as-formed CuNPs/ TiO_2 M (diameter 10 nm) was added to the solution. The sealed vessel was held at 80 °C for 24 h for MOF growth in TiO_2 M. The resulting chiral CuMOF/ TiO_2 M sample was carefully washed with DMF and CH_3OH to remove the unreacted ligands and then dried at 80 °C in a vacuum oven to remove the remaining solvent. The as-synthesized sample was extracted twice with an ethanol solution to remove the template (CTAB) from the framework ($t = 4$ h at 60 °C). The Meso-L-CuMOF/ TiO_2 M was incubated in a solution of HAuCl_4 (0.5 mM) at 70 °C for 2 h. After cleaning with DI water, the sample was dipped in a 0.1 mM NaBH_4 solution for 15 min to produce Au NPs.

Electrochemical measurements

$I-V$ curves of channels were measured using a homemade electrochemical cell. The effective exposed area of samples was 0.8 mm^2 , clamped between two PDMS films and placed between two Teflon cells. Two Ag/AgCl electrodes were used to apply the transmembrane potential and measure the ionic current. The electrochemical detection of molecules was performed in an aqueous solution (60 °C) containing different concentrations of L-D-glucose, 1.0 mM ABTS, and 1.0 μM KCl (pH 5). The ionic current transport through hybrid channels was measured in the voltage range of -1.0 to +1.0 V at a scan rate of 50 mV s⁻¹.

Numerical simulations for ionic transport in a Au@Meso-L-CuMOF/ TiO_2 M nanochannel array

The nanofluidic sensor mechanism for glucose was proposed by using a theoretical model and was evaluated by the finite

element method (FEM), combined with Poisson and Nernst–Planck (PNP) equations. The equations are shown below:^{56,57}

$$\nabla^2 \varphi = -\frac{F}{\varepsilon} \sum z_i c_i \quad (4)$$

$$j_i = D_i \left(\nabla c_i + \frac{z_i F c_i}{RT} \nabla \varphi \right) \quad (5)$$

$$\nabla j_i = 0 \quad (6)$$

where j_i , D_i , C_i , φ , R , F , T , and ε are the ionic flux, diffusion coefficient, ion concentration, electrical potential, universal gas, Faraday constant, absolute temperature, and dielectric constant of electrolyte solution, respectively. Eqn (4), called the Poisson equation, characterized the electric potential and ionic concentration. Eqn (5) is the Nernst–Planck equation, indicating the charged nanochannel transport behavior. Besides, when the system reaches a steady state, the flux should satisfy eqn (6). The coupled eqn (4)–(6) was used to calculate the ion concentration distribution and solved by assuming appropriate boundary conditions.

$$\vec{n} \nabla \varphi = -\frac{\sigma}{\varepsilon} \quad (7)$$

$$\vec{n} j = 0 \quad (8)$$

The boundary condition for the potential φ on the channel wall was eqn (7), where the ionic flux exhibited zero normal components at boundaries eqn (8).

Data availability

The data supporting the findings of this study are available within the article and in the ESI.†

Author contributions

Y.-Y. Song conceived the concept and directed the project. J. L. Guo, X. J. Xu, and J. J. Zhao performed the experiments. Z.-Q. Wu carried out the theoretical study. Z. D. Gao collected and analyzed the data. J. L. Guo prepared the first draft of this manuscript, and all the authors modified the manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (No.22074013, 21775066, and 21974058), and the Fundamental Research Funds for the Central Universities (No. N2105018). Special thanks are due to the instrumental or data analysis from the Analytical and Testing Center, Northeastern University.



Notes and references

1 J. M. Brown and S. G. Davies, *Nature*, 1989, **342**, 631–636.

2 R. A. Laine, *Glycobiology*, 1994, **4**, 759–767.

3 B. G. Davis, *Chem. Rev.*, 2002, **102**, 579–602.

4 K. K. Kalsi, E. H. Baker, O. Fraser, Y. L. Chung, O. J. Mace, E. Tarelli, B. J. Philips and D. L. Baines, *Pflügers Archiv: European Journal of Physiology*, 2009, **457**, 1061–1070.

5 G. T. Williams, J. L. Kedge and J. S. Fossey, *ACS Sens.*, 2021, **6**, 1508–1528.

6 L. Chen, Y. Chen, Y. Zhang and Y. Liu, *Angew. Chem., Int. Ed.*, 2021, **60**, 7654–7658.

7 S. S. Ding, S. M. Cao, A. W. Zhu and G. Y. Shi, *Anal. Chem.*, 2016, **88**, 12219–12226.

8 Y. Sun, F. Zhang, J. Quan, F. Zhu, W. Hong, J. Ma, H. Pang, Y. Sun, D. Tian and H. B. Li, *Nat. Commun.*, 2018, **9**, 2617.

9 S. Y. Zhang, Z. Y. Cao, P. P. Fan, Y. Q. Wang, W. D. Jia, L. Y. Wang, K. F. Wang, Y. Liu, X. Y. Du, C. Z. Hu, P. K. Zhang, H.-Y. Chen and S. Huang, *Angew. Chem., Int. Ed.*, 2022, **61**, e202203769.

10 G. L. Hou, H. C. Zhang, G. H. Xie, K. Xiao, L. P. Wen, S. H. Li, Y. Tian and L. Jiang, *J. Mater. Chem. A*, 2014, **2**, 19131–19135.

11 Z. Q. Dai, J. L. Guo, T. T. Su, J. F. Wang, Z. D. Gao and Y.-Y. Song, *Green Chem.*, 2021, **23**, 8685–8693.

12 H.-C. J. Zhou and S. Kitagawa, *Chem. Soc. Rev.*, 2014, **43**, 5415–5418.

13 J. Liu, L. Chen, H. Cui, J. Zhang, L. Zhang and C.-Y. Su, *Chem. Soc. Rev.*, 2014, **43**, 6011–6061.

14 F. Luo, Y. Lin, L. Zheng, X. Lin and Y. Chi, *ACS Appl. Mater. Interfaces*, 2015, **7**, 11322–11329.

15 K. Manna, T. Zhang and W. Lin, *J. Am. Chem. Soc.*, 2014, **136**, 6566–6569.

16 D. Farrusseng, S. Aguado and C. Pinel, *Angew. Chem., Int. Ed.*, 2009, **48**, 7502–7513.

17 S.-S. Ding, L. He, X.-W. Bian and G. Tian, *Nano Today*, 2020, **35**, 100920.

18 J. Navarro-Sánchez, A. I. Argente-García, Y. Moliner-Martínez, D. Roca-Sanjuan, D. Antypov, P. Campíns-Falco, M. J. Rosseinsky and C. Martí-Gastaldo, *J. Am. Chem. Soc.*, 2017, **139**, 4294–4297.

19 P. Li, J. A. Modica, A. J. Howarth, E. Vargas L, P. Z. Moghadam, R. Q. Snurr, M. Mrksich, J. T. Hupp and O. K. Farha, *Chem.*, 2016, **1**, 154–169.

20 L. G. Qiu, T. Xu, Z. Q. Li, W. Wang, Y. Wu, X. Jiang, X. Y. Tian and L. D. Zhang, *Angew. Chem., Int. Ed.*, 2008, **120**, 9629–9633.

21 K. Shen, L. Zhang, X. Chen, L. Liu, D. Zhang, Y. Han, J. Chen, J. Long, R. Luque, Y. Li and B. Chen, *Science*, 2018, **359**, 206–210.

22 P. Wang, S. Wang, W. Zhang, X. Li, Z. Gu, W. Li, S. Zhao and Y. Fu, *New J. Chem.*, 2020, **44**, 9611–9615.

23 M. Wang, Y. Hou, L. Yu and X. Hou, *Nano Lett.*, 2020, **20**, 6937–6946.

24 K. Zhan, Z. Li, J. Chen, Y. Hou, J. Zhang, R. Sun, Z. Bu, L. Wang, M. Wang, X. Chen and X. Hou, *Nano Today*, 2020, **33**, 100868.

25 M. Cheng, F. Zhu, S. Zhang, X. Zhang, M. K. Dhinakaran and H. Li, *Nano Lett.*, 2021, **21**, 4086–4091.

26 Y. C. Hao, L. W. Chen, J. Li, Y. Guo, X. Su, M. Shu, Q. Zhang, W. Y. Gao, S. Li, Z. L. Yu, L. Gu, X. Feng, A. X. Yin, R. Si, Y. W. Zhang, B. Wang and C. H. Yan, *Nat. Commun.*, 2021, **12**, 2682.

27 J. L. Guo, L. L. Yang, H. J. Xu, C. X. Zhao, Z. Q. Dai, Z. D. Gao and Y. Y. Song, *Anal. Chem.*, 2019, **91**, 13746–13751.

28 F. Riboni, N. T. Nguyen, S. Soa and P. Schmuki, *Nanoscale Horiz.*, 2016, **1**, 445–466.

29 J. Lu, H. C. Zhang, J. Hou, X. Y. Li, X. Y. Hu, Y. X. Hu, C. D. Easton, Q. Y. Li, C. H. Sun, A. W. Thornton, M. R. Hill, X. W. Zhang, G. P. Jiang, J. Z. Liu, A. J. Hill, B. D. Freeman, L. Jiang and H. T. Wang, *Nat. Mater.*, 2020, **19**, 767–774.

30 L. B. Sun, J. R. Li, J. Park and H. C. Zhou, *J. Am. Chem. Soc.*, 2012, **134**, 126–129.

31 J. L. Guo, L. L. Yang, C. X. Zhao, Z. D. Gao, Y.-Y. Song and P. Schmuki, *J. Mater. Chem. A*, 2021, **9**, 14911–14919.

32 B. J. Hao, K. G. Wang, Y. K. Zhou, C. F. Sui, L. Wang, R. Bai and Z. J. Yang, *ACS Omega*, 2020, **5**, 1109–1119.

33 Y. H. Hu, H. J. Cheng, X. Z. Zhao, J. J. X. Wu, F. Muhammad, S. C. Lin, J. He, L. Q. Zhou, C. P. Zhang, Y. Deng, P. Wang, Z. Y. Zhou, S. M. Nie and H. Wei, *ACS Nano*, 2017, **11**, 5558–5566.

34 C. X. Zhao, L. Q. Zhou, Z. Q. Zhang, Z. D. Gao, H. M. Weng, W. Zhang, L. W. Li and Y.-Y. Song, *J. Phys. Chem. Lett.*, 2020, **11**, 9931–9937.

35 J. X. Chen, Q. Ma, M. H. Li, D. Y. Chao, L. Huang, W. W. Wu, Y. X. Fang and S. J. Dong, *Nat. Commun.*, 2021, **12**, 3375.

36 Y. Y. Huang, J. S. Ren and X. G. Qu, *Chem. Rev.*, 2019, **119**, 4357–4412.

37 C. Wang, X.-G. Nie, Y. Shi, Y. Zhou, J.-J. Xu, X.-H. Xia and H.-Y. Chen, *ACS Nano*, 2017, **11**, 5897–5905.

38 Q. Zhang, S. Chen and H. Wang, *Green Chem.*, 2018, **20**, 4067–4074.

39 X. Q. Zhao, T. T. Yang, D. Q. Wang, N. Zhang, H. B. Yang, X. Jing, R. X. Niu, Z. W. Yang, Y. C. Xie and L. J. Meng, *Anal. Chem.*, 2022, **94**, 4484–4494.

40 X. X. Jian, J. Xu, J. L. Guo, J. J. Zhao, T. Shen, Z. D. Gao and Y.-Y. Song, *Small*, 2023, DOI: [10.1002/smll.202205995](https://doi.org/10.1002/smll.202205995).

41 W. C. Wu, L. D. Yu, Q. Z. Jiang, M. F. Huo, H. Lin, L. Y. Wang, Y. Chen and J. L. Shi, *J. Am. Chem. Soc.*, 2019, **141**, 11531–11539.

42 W. J. Luo, C. F. Zhu, S. Su, D. Li, Y. He, Q. Huang and C. H. Fan, *ACS Nano*, 2010, **4**, 7451–7458.

43 P. Anger, P. Bharadwaj and L. Novotny, *Phys. Rev. Lett.*, 2006, **96**, 113002.

44 L. Zhou, Y. Tan, D. Ji, B. Zhu, P. Zhang, J. Xu, Q. Gan, Z. Yu and J. Zhu, *Sci. Adv.*, 2016, **2**, e1501227.

45 E. D. Palik, *Handbook of Optical Constants of Solids*, Academic Press, 1985.

46 R. M. Cywar and G. T. Beckham, *Nat. Chem.*, 2022, **14**, 967–969.

47 T.-R. Li, F. Huck, G. M. Piccini and K. Tiefenbacher, *Nat. Chem.*, 2022, **14**, 985–994.

48 R. B. Schoch, J. Han and P. Renaud, *Rev. Mod. Phys.*, 2008, **80**, 839.

49 D. Stein, M. Kruithof and C. Dekker, *Phys. Rev. Lett.*, 2004, **93**, 035901.

50 K. Z. Qu, J. Xu, Y. F. Xue, J. L. Guo, Z. D. Gao, Y.-Y. Song and Y. Mei, *Anal. Chem.*, 2022, **94**, 588–592.

51 P. Cheng, H. Wang and X. H. Shi, *Nanoscale*, 2020, **12**, 3050–3057.

52 L. Chen, Y. Chen, Y. Zhang and Y. Liu, *Angew. Chem., Int. Ed.*, 2021, **60**, 7654–7658.

53 R. A. Tromans, S. K. Samanta, A. M. Chapman and A. P. Davis, *Chem. Sci.*, 2020, **11**, 3223–3227.

54 Y. Sun, F. Zhang, J. Quan, F. Zhu, W. Hong, J. Ma, H. Pang, Y. Sun, D. Tian and H. Li, *Nat. Commun.*, 2018, **9**, 2617.

55 L. Chen, Y. Chen, Y. Zhang and Y. Liu, *Angew. Chem., Int. Ed.*, 2021, **60**, 7654–7658.

56 D. Momotenko and H. H. Girault, *J. Am. Chem. Soc.*, 2011, **133**, 14496–14499.

57 H. Daiguji, *Chem. Soc. Rev.*, 2010, **39**, 901–911.

